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Different effects of pectin and  $\kappa$ -carrageenan on the multiscale structures and *in vitro* digestibility of extruded rice starch

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PII: S0268-005X(20)30622-6

DOI: https://doi.org/10.1016/j.foodhyd.2020.106216

Reference: FOOHYD 106216

To appear in: Food Hydrocolloids

Received Date: 5 March 2020

Revised Date: 22 June 2020

Accepted Date: 26 July 2020

Please cite this article as: He, H., Bian, H., Xie, F., Chen, L., Different effects of pectin and  $\kappa$ -carrageenan on the multiscale structures and *in vitro* digestibility of extruded rice starch, *Food Hydrocolloids* (2020), doi: https://doi.org/10.1016/j.foodhyd.2020.106216.

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built all the proof



2	structures and in vitro digestibility of extruded rice starch
3	
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17	

Different effects of pectin and  $\kappa$ -carrageenan on the multiscale

## 18 Abstract

This study compared the different effects of pectin (PE) and  $\kappa$ -carrageenan (CG) on the multilevel 19 20 structures (i.e. granule, crystallites, helices, fractals, and short-range order) and in vitro digestibility 21 of extruded rice starch (MERS). The addition of either PE or CG altered the multiscale structures, 22 increased the resistant component (RC) content, and reduced predicted glycemic index (pGI), with CG being more effective especially at a low content. CG led to the growth of molecular aggregates 23 ( $\alpha$ ), single helices, and V-type crystals ( $X_V$ ). PE resulted in higher contents of double helices, 24 25 A+B-type crystals ( $X_{A+B}$ ) and enhanced the short-range order ( $R_{1045/1022}$ ) and particle compactness. 26 Pearson correlation analysis indicates that, for MERS/PE, the slowly-digestible component (SDC) and RC contents were influenced by different factors in the sequence of total crystallinity  $(X_{Total}) >$ 27  $X_{A+B} > X_V >$  single-helix content >  $\alpha > R_{1045/1022} >$  double-helix content. For MERS/CG, the RC 28 29 content was affected by  $X_{\text{Total}} > \text{single-helix content} > X_{\text{V}}$ . Keywords: Pectin; Carrageenan; Extrusion; Rice starch; Multiscale structure; In vitro digestibility 30

## 32 1 Introduction

33	Rice is the typical staple food and most critical cereal crop in Asian (Marshall & Wadsworth,
34	1994). Starch is the main ingredient of rice, which plays an essential role in human nutrition (Czech,
35	Pastuszak, & Kusior, 2014). Unfortunately, gelatinized rice starch has more than 95% rapidly
36	digestible starch (RDS) (Zheng et al., 2018), which can cause negative physiological effects such as
37	high glycemic response.

The enzymatic digestion behavior of cooked rice may be changed with addition of some 38 non-starch polysaccharides (NSPs) (Chung, Liu, & Lim, 2007). The underlying mechanism for this 39 40 change could be different depending on NSP structure, with the most typical explanation being an 41 increased viscosity of intestinal contents (Brennan, Suter, Luethi, Matiamerino, & Qvortrup, 2008). 42 Furthermore, NSPs have been reported to reduce starch digestion by forming a "barrier" on the 43 starch granule surface against amylase (Dartois, Singh, Kaur, & Singh, 2010; Tomoko Sasaki & Kohyama, 2012; Sasaki, Sotome, & Okadome, 2015). Moreover, NSPs may also inhibit α-amylase in 44 45 a direct non-competitive matter, which restrain starch digestion and, hence, inhibit postprandial glycemia (Slaughter, Ellis, Jackson, & Butterworth, 2002). 46 47 Pectin (PE) is an important NSP. It is composed of the galacturonic acid main chain and neutral sugar side chains, which contains approximately 65% homogalacturonan (HG), 20-35% 48

49 rhamnogalacturonan I (RGI), 10% rhamnogalacturonan II (RGII), and small amounts of

- 50 xylogalacturonan (XG) (Guo et al., 2016; Naqash, Masoodi, Rather, Wani, & Gani, 2017). Based on
- 51 the degree of methoxylation (DM), PE can be divided into high-methoxyl PE (HMP) and
- 52 low-methoxyl PE (LMP). In addition, κ-Carrageenan (CG) is an NSP consisting of sulfureted or

53	non-sulfated galactose and 3,6-anhydrogalactose that are alternately connected by $\alpha$ -1,3 glycosidic
54	bonds and $\beta$ -1,4 bonds, while the 1,3-linked D-galactose unit C <sub>4</sub> carries a sulfate group (Ueda, Itoh,
55	Matsuzaki, Ochiai, & Imamura, 2001; Viebke, Borgström, Carlsson, Piculell, & Williams, 1998).
56	How the structural difference between PE (branched chain) and CG (linear chain) can lead to
57	different effects on the digestion of rice starch is interesting but has rarely been reported.
58	In most studies, simple physical blending of starch with NSPs with abundant water was used to
59	investigate the effects of NSPs on the digestibility of starch (Anynda, Kelvin Kim Tha, Allan Keith,
60	& Lara, 2019; Gularte & Cristina, 2011; Tomoko Sasaki et al., 2012; Tester & Sommerville, 2003).
61	Besides, the digestibility of starch-NSP composites subjected to physical modification such as
62	heat-moisture treatment and extrusion has also been reported (Adamu, 2001; Chen, Xiong, & Gao,
63	2017). Our previous work (He et al., 2020) has shown that the complexation between rice starch and
64	guar gum could be assisted by thermomechanical treatment. However, the enzymatic digestibility of
65	rice starch-NSP composites thermomechanically processed with limited moisture content have still
66	been limited.
67	Extrusion has promising application prospects in the food manufacturing industry owing to its
68	benefits of simple operation, environmental friendliness, and high safety (Tran, Hendriks, & van der
69	Poel, 2008). Many studies have proved that the paste properties (e.g. rheology) of starch will change
70	greatly during the extrusion process (Bhattacharya & Hanna, 1987; Cai & Diosady, 1993; Chuang &
71	Yeh, 2004; Fishman, Coffin, Konstance, & Onwulata, 2000). Nonetheless, extrusion is normally a
72	complex process, during which the precise control of starch structural changes is challenging due to
73	the difficulties in manipulating the residence time and thermomechanical history of each small

74	portion of the processed material. To overcome these difficulties, a miniature twin-screw extruder
75	providing simplified, well-controlled, and more uniform processing conditions was employed in this
76	current study to investigate the functions of PE or CG on the digestibility of extruded rice starch
77	(MERS) in a more accurate way. Based on this, we have established the relationship between the
78	multiscale structures ( <i>i.e.</i> granule, crystallites, helices, fractals, and short-range order) and the <i>in</i>
79	<i>vitro</i> digestibility of the thermomechanically-processed composites.

80 2 Materials and Methods

#### 81 2.1 Materials

Rice starch (GABIOSTA-F) was supplied by Jinnong Biotechnology Co., Ltd (China); Pectin from Aladdin Biochemical Technology Co., Ltd. (China);  $\kappa$ -Carrageenan from CP Kelco U.S., Inc.; Pancreatic  $\alpha$ -amylase (A3306) from Sigma-Aldrich (USA). The specifications of the pectin include galacturonic acid (GA) = 65.59±0.57%, degree of methoxylation (DM) = 28.28±0.77%, degree of amidation (DA) = 20.63±0.51%, molecular mass ( $M_w$ ) = 786±76 kDa. The glucose oxidation kit (GOPOD) was purchased from Megazyme (Ireland). Other chemical reagents were of analytical reagent.

## 89 2.2 Processing of extruded rice starch (MERS) with pectin (PE) or κ-carrageenan

90 (CG)

MERS/PE or MERS/CG was prepared following our previous work (He et al., 2020) but with
addition of PE or CG (2.5%, 5%, 7.5%, 10%, wt/wt, based on rice starch) instead. The moisture
content of the samples was adjusted to 40% moisture content. Extrusion processing was undertaken
using a Haake MiniLab II 40-mm co-rotating twin-screw micro-extruder (Thermo Fisher, USA), as

95	shown in Fig. 1. The reliability of this equipment in wide polymer processing research has been well
96	demonstrated (Wang et al., 2014). The screw speed was set to be 150 rpm and the barrel temperature
97	(only one temperature zone) was 85 $^{\circ}$ C. 5 g of the sample was fed into the extruder at the feeding
98	port and the extrudate was collected at the exit opening without reflux. The residence time was about
99	5 min. (Fig. 1). The two types of composite prepared were named MERS/PE-X and MERS/CG-X
100	respectively, where the letter "X" stands for the addition of PE or CG (wt%). The MERS sample
101	without PE or CG (control) was named MERS/0, which was characterized previously (He et al.,
102	2020). The term "MERS" is used to describe extruded rice starch (without or with an NSP) in
103	general.



**Fig. 1.** Pictures of HAAKE MiniLab Micro-extruder and the inside configuration. The yellow line

107 indicates the flow direction used in this work (the backflow channel was not used).

## 109 **2.3** Characterization and statistical analysis

110 The samples were characterized and statistically analyzed using the same methods as reported111 before (He et al., 2020).

## 112 **3 Results and discussion**

#### **3.1** Effects of PE/CG on the digestibility and predicted glycemic index (pGI) of MERS

114 **Table 1** shows that inclusion of PE or CG remarkably inhibited the digestion of MERS.

115 Specifically, with a higher content of PE or CG in the formulation, the RDC content became lower,

116 the resistant component (RC) content higher, and pGI was also reduced. CG seems to be more

117 effective than PE to reduce the digestibility of MERS. Only 2.5% addition of CG could significantly

118 reduce the rapidly-digestible component (RDC) and slowly-digestible component (SDC) contents

and predicted glycemic index (pGI) and increase the RC content of MERS. Compared with

120 MERS/PE, MERS/CG exhibited a higher RC content and lower pGI at the same addition amounts

121 (2.5–10%). However, MERS/PE-10% presented even a lower RDC content than MERS/CG-10%.

122 Moreover, the addition of PE increased the SDC content whereas CG had a reverse effect. Due to the

123 difference in chemical structure, the ways of interaction between PE or CG and starch molecules

124 could be quite different, which, in turn, may largely affect the composite structure (Mahmood et al.,

125 2017). A branched biopolymer generally presents a lower viscosity than a linearly structured

126 biopolymer (Xie et al., 2009). The lower viscosity of PE could reduce the overall viscosity of the

127 starch/PE blend, so that starch could suffer from less shear-induced degradation (Hasjim, Xie, Halley,

128 & Gilbert, 2014). Moreover, PE with lower viscosity could be easier to encapsulate starch domains

129 in the blends, restricting starch gelatinization (Tester et al., 2003) and, thus, reduce starch

130	digestibility. In contrast, CG with a linear chain structure had a higher viscosity so it is less likely to
131	encapsulate starch domains (Marcotte, Hoshahili, & Ramaswamy, 2001). However, the linear
132	structure could make CG chains to interact with starch chains more effectively, leading to a higher
133	RC content of the complex. The above results showed that PE or CG effectively regulated the
134	anti-digestion performance and pGI of rice starch during extrusion, but the regulation effect was not
135	the same. More specifically, the different structural features between PE and CG could lead to
136	different ways of molecular interaction with rice starch.

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## **Table 1.** RDC, SDC, RC and pGI of MERS/0, MERS/PE and MERS/CG samples<sup>\*</sup>

Samples	RDC (%)	SDC (%)	RC (%)	pGI
MERS/0 <sup>#</sup>	$75.2{\pm}0.6^{g}$	$10.3 \pm 0.6^{d}$	$14.5 \pm 0.5^{a}$	$82.3 \pm 1.5^{g}$
MERS/PE-2.5%	$73.2 \pm 1.0^{g}$	$11.8 \pm 1.2^{e}$	$15.0 \pm 1.2^{b}$	79.1 $\pm$ 1.3 <sup><i>f</i></sup>
MERS/PE-5%	55.9±0.9 <sup>f</sup>	$14.2 \pm 0.4^{f}$	27.9±1.6 <sup>c</sup>	$75.7 \pm 0.7^{e}$
MERS/PE-7.5%	37.0±0.7 <sup>e</sup>	$16.2 \pm 0.4^{g}$	$46.8 \pm 1.2^{d}$	$71.8 \pm 2.1^{d}$
MERS/PE-10%	$29.9 \pm 0.8^{a}$	$17.5 \pm 0.9^{h}$	$52.6 \pm 1.1^{e}$	$70.3 \pm 1.5^{d}$
MERS/CG-2.5%	$40.2 \pm 0.6^{d}$	$4.2 \pm 0.2^{c}$	$55.6 \pm 1.1^{e}$	$67.9 \pm 2.1^{c}$
MERS/CG-5%	$37.9 \pm 1.0^{c}$	$2.7 \pm 0.9^{a}$	$59.4 \pm 0.7^{e}$	$65.6 \pm 0.9^{b}$
MERS/CG-7.5%	$36.5 \pm 0.7^{b}$	$2.3{\pm}1.1^{a}$	$61.2 \pm 1.4^{f}$	$65.0\pm0.8^b$
MERS/CG-10%	30.2±1.3 <sup>a</sup>	$2.1 \pm 0.6^{a}$	$67.7 \pm 1.3^{f}$	63.1±1.6 <sup>a</sup>

<sup>\*</sup>All data were repeated in triplicates and expressed as mean with standard deviation (SD). Different values of

140 different letters in the same column indicate statistical significance (p < 0.05).

141 <sup>#</sup> Data from reference (He et al., 2020).

142

## 143 **3.2 First-order kinetics analysis**

144 The digestion behaviors of MERS/PE and MERS/CG samples could be well fitted to

145	first-order and log of slope (LoS) plots ( $R^2 > 0.9$ , Fig. 2). The associated parameters such as the first
146	order rate of digestion ( <i>k</i> ) and the digested components ratio at the end of the reaction ( $C_{\infty}$ ) are
147	shown in <b>Table 2.</b> The MERS/PE composites digested more rapidly within the first 1 h. However, a
148	shorter time was used to reach a plateau for the MERS/CG samples (about 40 min) than for the
149	MERS/PE samples (about 60 min). Moreover, the digestion of MERS/PE and MERS/CG samples
150	was a single-phase process (Butterworth, Frederick, Terri, Hamung, & Peter, 2012; He et al., 2020).
151	Compared with those for MERS/PE and MERS/CG samples, <i>k</i> for MERS/0 was significantly higher.
152	However, the MERS/CG samples had lower $k$ than the MERS/PE samples. $k$ is controlled by the
153	catalytic properties of the amylase itself, namely the catalytic rate constant (Roder et al., 2009;
154	Slaughter, Ellis, & Butterworth, 2001). It has been suggested that as digestion proceeds, a low $k$
155	value reflects slow diffusion of amylase into the starch granules (Dhital, Shrestha, & Gidley, 2010).
156	Moreover, the difference in the digestion rate of starch mainly depends on $C_{\infty}$ , namely, the total
157	amount of available/digestible starch. The addition of PE or CG led to notably decreased $C_{\infty}$ . The
158	MERS/CG samples had much lower $C_{\infty}$ than the MERS/PE samples. This result corresponds to the
159	digestion properties (Table 1).



Fig. 2. First-order plots (A) and LOS plots (B) for MERS/PE and MERS/CG samples.

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**Table 2.**  $k \,(\min^{-1})$  and  $C_{\infty}$  (%) for MERS/0, MERS/PE and MERS/CG samples<sup>\*</sup>

Samples	$k (\mathrm{min}^{-1})$	$\mathcal{C}_{\infty}$ (%)	
MERS/0 <sup>#</sup>	$0.122 \pm 0.003^{e}$	83.169±1.104 <sup>g</sup>	
MERS/PE-2.5%	$0.092{\pm}0.008^{d}$	82.961±1.379 <sup>g</sup>	
MERS/PE-5%	$0.078 \pm 0.011^{c}$	73.826±0.926 <sup>f</sup>	
MERS/PE-7.5%	$0.077 \pm 0.002^{c}$	57.155±1.308 <sup>e</sup>	
MERS/PE-10%	$0.042 \pm 0.003^{a}$	$53.729 \pm 0.586^{e}$	
MERS/CG-2.5%	$0.081 \pm 0.004^d$	$46.331 \pm 0.863^{d}$	
MERS/CG-5%	$0.079 \pm 0.002^{c}$	40.476±0.709 <sup>c</sup>	
MERS/CG-7.5%	$0.073 \pm 0.005^{c}$	$38.035 \pm 0.412^{b}$	
MERS/CG-10%	$0.067 \pm 0.002^{b}$	31.686±0.542 <sup>a</sup>	

<sup>\*</sup> All data were repeated in triplicates and expressed as mean with standard deviation (SD). Different values of

166 different letters in the same column indicate statistical significance (p < 0.05).

<sup>#</sup> Data from reference (He et al., 2020).

## 169 **3.3** Effect of PE/CG on the morphology of MERS

170 The morphology was investigated by scanning electron microscopy (SEM) as shown in Fig. 3. Our previous study (He et al., 2020) showed that the granules of MERS presented irregular shapes 171 172 and a rough surface, with pores on the surface. This indicates that thermomechanical treatment 173 significantly destroyed the granule structure of native rice starch (NRS). The size of MERS became 174 more homogeneous, ranging from 80 µm to 150 µm. In contrast, MERS/PE or MERS/CG particles had a smooth surface without obvious pores, suggesting that the interactions between PE/CG and 175 176 rice starch during extrusion led to a homogenous phase on the macroscale and a more compact 177 structure. This structure may inhibit the diffusion of amylase into the interior of the particles. The MERS/PE and MERS/CG samples exhibited similar morphologies. 178

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- 181
- **Fig. 3.** SEM images of MERS/PE and MERS/CG samples (Scale bar: 100 µm; magnification:
- 182

500×).

3.4 Effect of PE/CG on the aggregated structure of MERS

185	The double-logarithmic small-angle X-ray scattering (SAXS) patterns for the different samples
186	are shown in <b>Fig. 4</b> . NRS displayed a typical scattering peak at $q$ of 0.614 nm <sup>-1</sup> , which disappeared
187	for all the MERS samples including MERS/0 (He et al., 2020). The MERS/PE and MERS/CG
188	samples exhibited a higher scattering intensity than MERS/0 in a small $q$ range, indicating the
189	formation of complexes through interaction between rice starch and PE/CG during extrusion. The
190	fractal dimensions for the MERS/PE and MERS/CG samples with different PE or CG contents were
191	calculated from the slopes of the linear curves in the power-law relationship $I(q) \sim q^{\alpha}$ (Fan et al., 2014;
192	Li, Senesi, & Lee, 2016). The $ \alpha $ value of the two types of composite ( <b>Table 3</b> ) was in the range from
193	1 to 3, suggesting the formation of a mass fractal dimension characteristics of fractal structure ( $D_{\rm m}$ )
194	(Suzuki, Chiba, & Yarno, 1997; Zhang et al., 2014). A higher content of PE or CG caused a
195	significant increase in $\alpha$ for MERS, meaning that a denser aggregated structure resulted from the
196	molecular interaction between PE/CG and rice starch. At the same content of addition, the
197	MERS/CG composites exhibited a higher $\alpha$ value than the MERS/PE samples. Given this, linear CG
198	chains were more likely to interact with starch chains leading to a higher degree of molecular
199	aggregation.

200



Fig. 4. Double-logarithmic SAXS patterns for MERS/PE and MERS/CG samples.

**Table 3.**  $\alpha$ , total crystallinity ( $X_{Total}$ ), A+B-type crystallinity ( $X_{A+B}$ ), V-type crystallinity ( $X_V$ ), and

D = - f MEDC/0 MEDC/DE and MEDC/CC as multiple of the second se	
$K_{10} = K_{10} = K$	20
$\Lambda_{1045/1077}$ OI MILING/U, MILING/I L and MILING/UO sample	~o.

Samples	α	$X_{\mathrm{Total}}$ (%)	$X_{A+B}$ (%)	$X_{\mathrm{V}}\left(\% ight)$
NRS <sup>#</sup>	$1.66 \pm 0.02^{a}$	$32.4 \pm 0.5^{f}$	$32.0\pm0.4^{f}$	$0.4{\pm}0.1^{a}$
MERS/0	$2.10 \pm 0.03^{b}$	16.6±0.13 <sup>a</sup>	$15.4 \pm 0.14^{b}$	$1.2 \pm 0.12^{b}$
MERS/PE-2.5%	2.12±0.01 <sup>c</sup>	$18.2 \pm 0.17^{b}$	$16.8 \pm 0.18^{c}$	$1.4 \pm 0.12^{c}$
MERS/PE-5%	$2.15 \pm 0.02^{d}$	18.8±0.13 <sup>c</sup>	$17.2 \pm 0.14^{d}$	$1.6 \pm 0.12^{d}$
MERS/PE-7.5%	$2.22 \pm 0.03^{e}$	$20.1 \pm 0.19^{d}$	18.3±0.16 <sup>e</sup>	$1.8 \pm 0.17^{e}$
MERS/PE-10%	$2.23 \pm 0.02^{f}$	$20.9 \pm 0.18^{d}$	$18.7 \pm 0.16^{e}$	2.1±0.16 <sup>f</sup>
MERS/CG-2.5%	$2.21 \pm 0.03^{e}$	$20.0 \pm 0.17^{d}$	$15.0 \pm 0.18^{b}$	$5.0\pm0.12^{g}$
MERS/CG-5%	$2.24 \pm 0.01^{g}$	$20.8 \pm 0.13^{d}$	$14.5 \pm 0.14^{a}$	$6.3 \pm 0.12^{h}$
MERS/CG-7.5%	$2.25 \pm 0.03^{g}$	21.2±0.19 <sup>e</sup>	14.6±0.16 <sup>a</sup>	$6.8 \pm 0.17^{i}$
MERS/CG-10%	$2.26 \pm 0.02^{h}$	21.9±0.18 <sup>e</sup>	$14.2 \pm 0.16^{a}$	$7.7 \pm 0.16^{j}$

- <sup>\*</sup> All data were repeated in triplicates and expressed as mean with standard deviation (SD). Different values of different letters in the same column indicate statistical significance (p < 0.05).
- <sup>#</sup> Data from reference (He et al., 2020).
- 209

## **3.5** Effect of PE/CG on the crystalline structure of MERS

211 Fig. 5 shows the X-ray diffraction (XRD) patterns for the different samples. The peaks of MERS/0 appeared at 15°, 17°, 20°, and 23.3° (2 $\theta$ ), suggesting an A+B+V hybrid crystalline structure 212 (He et al., 2020). Compared with NRS (He et al., 2020), all the MERS samples had low crystallinity. 213 214 Given this, the extrusion process had largely destroyed the original crystalline structure and caused some extents of gelatinization of these samples. Compared with MERS/0, the samples added with PE 215 216 or CG presented the same type of crystalline structure but slightly higher crystallinity (Table 3). 217 These results indicate that addition of PE or CG probably inhibited gelatinization during the 218 thermomechanical processing. CG dramatically increased the V-type crystallinity  $(X_V)$  of MERS whereas there were no apparent changes to the A+B-type crystallinity  $(X_{A+B})$ . In comparison, PE 219 220 gradually increased the  $X_{A+B}$  of MERS with increasing content but only significantly moved  $X_V$  to a much higher value when the addition amount was 10%. As a result, the MERS/CG samples only 221 showed slightly higher  $X_{Total}$  than the MERS/PE samples. During extrusion, starch undergoes a phase 222 223 transition at a low moisture content, which disrupts its crystalline structure and semi-crystalline 224 lamellae (Chen, Zhu, & Liu, 2017; Liu et al., 2017; Zhang et al., 2014). Previous studies reported that extrusion led to the destruction of the original A-type crystal structure of sweet potato and its 225 226 reorganization to form a weak B-type crystal structure (Van Soest, Hulleman, De Wit, & Vliegenthart, 227 1996; Waramboi, Gidley, & Sopade, 2014). NSPs such as PE and β-glucan may adhere to the surface

228 of starch particles through van der Waals forces or hydrogen bonding under extrusion treatment,

- 229 which inhibits the ordered structure destruction, resulting in higher  $X_{A+B}$  (Brennan, Derbyshire,
- 230 Tiwari, & Brennan, 2013; Robin, Schuchmann, & Palzer, 2012; Wang, Jin, & Yuan, 2007). CG might
- 231 not have such a strong effect due to its linear chain structure and higher viscosity, although it could
- 232 promote the growth of V-type crystallites.





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Fig. 5. X-ray diffractograms for MERS/PE and MERS/CG samples.

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## 237 **3.6 Effect of PE/CG on the helical structure of MERS**



242	Gidley, & Karkalas, 1993). Accordingly, the contents of double helices, single helices and
243	amorphous structure can be calculated (Gidley et al., 2002; Morrison et al., 1993), which are listed in
244	Table 4. Compared with NRS, the MERS samples exhibited higher contents of single helices and
245	amorphous starch but a lower content of double helices. Compared with MERS/0 (He et al., 2020),
246	both MERS/PE and MERS/CG samples showed a remarkably reduced content of amorphous starch.
247	Nonetheless, the MERS/PE samples presented both significantly higher contents of single helices
248	and double helices whereas CG notably increased the single helices content but reduced the double
249	helices content in MERS. Likely, the branched chains of PE can interact with rice starch to form new
250	double helices through hydrogen bonding (Naqash et al., 2017). In contrast, CG with a linear
251	molecular structure tends to interact with starch by hydrogen bonding between their main chains. As
252	both CG and amylose tend to form a helical structure, one may form single helices incorporating the
253	other (Lascombes et al., 2017). Moreover, the thermomechanical treatment broke $\alpha$ -1,4 and $\alpha$ -1,6
254	glycosidic bonds, resulted in more amylose content and the production of more single helical
255	structure (Liu et al., 2017; Van Soest et al., 1996). The susceptibility of polymer chains to shear
256	degradation is affected by the branch structure; a shorter branch length and higher branch density are
257	related to higher sensitivity to shear degradation (Liu, Halley, & Gilbert, 2010). PE with a branch
258	structure tends to undergo chain scission under shear treatment. This could reduce the shear effect on
259	the starch, leading to a higher double-helix content. However, this protection effect was much less
260	apparent when linearly structured CG was used.



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Table 4. Percentages of single helices, double helices and amorphous starch in MERS samples

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Table 4. Ferenages of single hences, double hences and anotphous staten in WERS samples					
	and NRS me	easured by <sup>13</sup> C Cl	P/MAS NMR.*		
Samples	Amorphous	Single helix	Double helix	$R_{1045/1022}$	
NRS	$27.7 \pm 1.3^{a}$	$1.8{\pm}0.9^{a}$	$70.5 \pm 0.4^{g}$	$0.773 {\pm} 0.004^{a}$	
MERS/0 <sup>#</sup>	79.2 $\pm$ 0.5 <sup>e</sup>	$2.5 \pm 0.3^{b}$	$18.3 \pm 0.8^{d}$	$0.815 {\pm} 0.002^{b}$	
MERS/PE-2.5%	78.7 $\pm$ 0.4 <sup>e</sup>	$2.9{\pm}0.1^{b}$	$18.4 \pm 0.8^{c}$	$0.878 \pm 0.004^{f}$	
MERS/PE-5%	$77.7 \pm 0.1^{c}$	$3.4 \pm 0.0^{c}$	$18.9 \pm 0.5^{d}$	$0.882 \pm 0.012^{g}$	

 $4.6 \pm 0.3^{d}$ 

 $4.9 \pm 0.0^{d}$ 

5.6±0.1<sup>e</sup>

 $6.4\pm0.3^{f}$ 

 $19.4 \pm 0.1^{e}$ 

 $20.6 \pm 0.0^{f}$ 

 $17.6 \pm 0.5^{b}$ 

 $16.6 \pm 0.8^{a}$ 

 $0.887 \pm 0.003^{g}$ 

 $0.894 \pm 0.016^{h}$ 

 $0.825 \pm 0.014^{c}$ 

 $0.832 \pm 0.012^{d}$ 

 $76.0\pm0.5^{c}$ 

 $74.5 \pm 0.0^{b}$ 

 $76.8 \pm 0.4^{c}$ 

 $77.0\pm0.5^{d}$ 

MERS/PE-7.5%

MERS/PE-10%

MERS/CG-2.5%

MERS/CG-5%

	J	ournal Pre-pro	oof	
MERS/CG-7.5%	$76.8 \pm 0.0^{c}$	7.8±0.0 <sup>f</sup>	$15.4{\pm}0.0^{a}$	$0.859 \pm 0.003^{e}$
MERS/CG-10%	$76.6 \pm 0.4^{c}$	$8.1 \pm 0.1^{f}$	$14.3 \pm 0.5^{a}$	$0.877 {\pm} 0.016^{e}$

<sup>\*</sup> All data were repeated in triplicates and expressed as mean with standard deviation (SD). Different values of different letters in the same column indicate statistical significance (p < 0.05).

<sup>#</sup> Data from reference (He et al., 2020).

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## 271 3.7 Effect of PE/CG on short-range order of MERS

272 The changes of short-range order in starch was investigated by Fourier-transform infrared (FTIR) spectroscopy (Van Soest, Tournois, De Wit, & Vliegenthart, 1995; Warren, Gidley, & 273 274 Flanagan, 2016). The ATR-FTIR spectra for NRS and the MERS/PE and MERS/CG samples are 275 shown in Fig. 7. The calculated  $R_{1045/1022}$  values are listed in Table 4. A higher  $R_{1045/1022}$  value 276 indicates higher amounts of short-range ordered structure (Soest, Tournois, Wit, & Vliegenthart, 1995; Warren et al., 2016). Compared with NRS, the MERS samples resulted in significant higher 277  $R_{1045/1022}$ . The data shows that the inclusion of PE or CG enhanced short-range order in both types of 278 279 composite, with a higher amount of addition providing a better effect as compare to MERS/0 (He et al., 2020). The effect of PE was shown to be more pronounced, which may be due to its lower 280 281 viscosity (Marcotte et al., 2001) and, thus, greater effect to assist short-range structural ordering in 282 starch. This result corresponds to the NMR data (Table 4).





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Fig. 7. FTIR spectra for MERS/PE, MERS/CG samples and NRS.

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## 287 3.8 Correlation between PE/CG addition and the structure and digestibility of MERS

288**Table 5** shows the Pearson correlation coefficients calculated for the relationship among the

289 PE/CG content, MERS structure and digestibility.

290 For the MERS/PE samples, the RDC content significantly negatively correlates with the RC

291 content and SDC content; the SDC and RC contents significantly negatively correlate with

amorphous starch content and pGI, and significantly positively correlate with  $\alpha$ ,  $X_{\text{Total}}$ ,  $X_{\text{A+B}}$ ,  $X_{\text{V}}$ ,

- 293 single-helix content, double-helix content, and  $R_{1045/1022}$ . This indicates that the enhancement of these
- structural features increases the SDC and RC contents, following the sequence of  $X_{\text{Total}} > X_{\text{A+B}} > X_{\text{V}} >$
- single-helix content >  $\alpha$  >  $R_{1045/1022}$  > double-helix content. The PE addition amount shows a

296	significant negative correlation with RDC content, amorphous starch content, and pGI; and a
297	significant positive correlation with RC, $\alpha$ , $X_{\text{Total}}$ , $X_{\text{A+B}}$ , $X_{\text{V}}$ , $R_{1045/1022}$ , single-helix content, and
298	double-helix content. This indicates an increasing amount of PE addition would promote the
299	structural ordering and, thus, digestion resistance. The PE addition influences these structural
300	features in the sequence of $X_{\text{Total}} > R_{1045/1022} > X_{\text{A+B}} > \text{single-helix content} > X_{\text{V}} > \text{double-helix}$
301	content.

303 Table 5. Pearson correlation coefficients for the relationship among PE/CG content, MERS structure
 304 and digestibility.\*

	MERS/PE			30	MERS/CG	ł		
	content	RDC	SDC	RC	content	RDC	SDC	RC
PE or CG content	1	_	-	_		_	_	_
RDC	-0.994**	1	-0.	_	-0.947	1	_	_
SDC	0.992**	-0.985*	1	-	-0.910	0.777	1	_
RC	0.983*	-0.961*	0.992**	1	0.973*	-0.993*	-0.846	1
α	0.956*	-0.966*	0.981*	$0.954^{*}$	0.964*	-0.854	-0.991*	0.910
$X_{ m Total}$	0.996**	$-0.982^{*}$	0.995**	0.995**	0.993**	$-0.950^{*}$	-0.932	$0.980^{*}$
$X_{\mathrm{A+B}}$	$0.979^{*}$	-0.951*	0.974 <sup>*</sup>	0.991**	-0.899	0.903	0.905	-0.935
$X_{ m V}$	0.973*	$-0.984^{*}$	0.986*	$0.958^{*}$	0.985*	-0.929	-0.954*	0.966*
<i>R</i> <sub>1045/1022</sub>	0.992**	$-0.987^{*}$	0.967*	$0.957^{*}$	0.979*	-0.935	-0.820	0.946
Amorphous starch	-0.995**	0.981*	$-0.978^{*}$	$-0.977^{*}$	-0.632	0.734	0.258	-0.670
Single helices	$0.974^{*}$	-0.944*	$0.977^{*}$	0.996**	$0.980^{*}$	-0.858	-0.913	0.968*
Double helices	$0.972^{*}$	$-0.975^{*}$	0.935*	0.916*	-0.999**	0.947	0.899	-0.971*
pGI	-0.986*	$0.970^{*}$	$-0.977^{*}$	-0.999**	$-0.979^{*}$	0.945	0.938	$-0.977^{*}$

p < 0.05 and p < 0.01 indicate statistical significance.

307	For the MERS/CG samples, the content of RDC significantly negatively correlates with RC
308	content. The RC content shows a significant positive correlation with $X_{Total}$ , $X_V$ , and single-helix
309	content and a significantly negative correlation with the content of double helix and pGI. The
310	influence of these structural features on RC content follows the sequence $X_{\text{Total}}$ > single-helix
311	content > $X_V$ . The content of CG addition demonstrates a significant positive correlation with RC
312	content, $\alpha$ , $X_{\text{Total}}$ , $X_{\text{V}}$ , $R_{1045/1022}$ and single-helix content, and an obvious negative correlation with the
313	content of double helix and pGI. This indicates that a higher content of CG added increases the
314	digestion resistance by promoting the formation of the crystalline and short-range ordered structures.
315	The CG addition positively influences the different structural features in the sequence of $X_{\text{Total}} > X_{\text{V}} >$
316	single-helix content > $R_{1045/1022}$ > $\alpha$ and reduces the double-helix content.
217	3.9 Discussion on mechanism regarding the digestion resistance of MERS/PE and
517	5.5 Discussion on meenanism regarding the digestion resistance of merton e and
317	MERS/CG
<ul><li>317</li><li>318</li><li>319</li></ul>	MERS/CG Fig. 8 is a schematic representing the evolution of the multiscale structural of
<ul><li>317</li><li>318</li><li>319</li><li>320</li></ul>	MERS/CG Fig. 8 is a schematic representing the evolution of the multiscale structural of thermomechanically-processed rice starch with PE or CG.
<ul> <li>317</li> <li>318</li> <li>319</li> <li>320</li> <li>321</li> </ul>	MERS/CG Fig. 8 is a schematic representing the evolution of the multiscale structural of thermomechanically-processed rice starch with PE or CG. The different molecular structures of PE and CG could make their interactions with starch
<ul> <li>317</li> <li>318</li> <li>319</li> <li>320</li> <li>321</li> <li>322</li> </ul>	MERS/CG Fig. 8 is a schematic representing the evolution of the multiscale structural of thermomechanically-processed rice starch with PE or CG. The different molecular structures of PE and CG could make their interactions with starch different. The PE has a branched-chain structure and may inhibit the proximity between the main
<ul> <li>317</li> <li>318</li> <li>319</li> <li>320</li> <li>321</li> <li>322</li> <li>323</li> </ul>	MERS/CG Fig. 8 is a schematic representing the evolution of the multiscale structural of thermomechanically-processed rice starch with PE or CG. The different molecular structures of PE and CG could make their interactions with starch different. The PE has a branched-chain structure and may inhibit the proximity between the main chains of PE and starch (Luo, Chen, Li, Liang, & Chen, 2017). The interaction between starch and
<ul> <li>317</li> <li>318</li> <li>319</li> <li>320</li> <li>321</li> <li>322</li> <li>323</li> <li>324</li> </ul>	MERS/CG Fig. 8 is a schematic representing the evolution of the multiscale structural of thermomechanically-processed rice starch with PE or CG. The different molecular structures of PE and CG could make their interactions with starch different. The PE has a branched-chain structure and may inhibit the proximity between the main chains of PE and starch (Luo, Chen, Li, Liang, & Chen, 2017). The interaction between starch and PE mainly occurs via their side chains by hydrogen bonding and entanglement. As a result, new
<ul> <li>317</li> <li>318</li> <li>319</li> <li>320</li> <li>321</li> <li>322</li> <li>323</li> <li>324</li> <li>325</li> </ul>	<b>MERS/CG</b> <b>Fig. 8</b> is a schematic representing the evolution of the multiscale structural of thermomechanically-processed rice starch with PE or CG. The different molecular structures of PE and CG could make their interactions with starch different. The PE has a branched-chain structure and may inhibit the proximity between the main chains of PE and starch (Luo, Chen, Li, Liang, & Chen, 2017). The interaction between starch and PE mainly occurs via their side chains by hydrogen bonding and entanglement. As a result, new double helices and A+B-type crystallites (higher $X_{A+B}$ ) are formed. Owing to the steric hindrance
<ul> <li>317</li> <li>318</li> <li>319</li> <li>320</li> <li>321</li> <li>322</li> <li>323</li> <li>324</li> <li>325</li> <li>326</li> </ul>	<b>MERS/CGFig. 8</b> is a schematic representing the evolution of the multiscale structural ofthermomechanically-processed rice starch with PE or CG.The different molecular structures of PE and CG could make their interactions with starchdifferent. The PE has a branched-chain structure and may inhibit the proximity between the mainchains of PE and starch (Luo, Chen, Li, Liang, & Chen, 2017). The interaction between starch andPE mainly occurs via their side chains by hydrogen bonding and entanglement. As a result, newdouble helices and A+B-type crystallites (higher $X_{A+B}$ ) are formed. Owing to the steric hindranceeffect, it is difficult for PE chains to be wrapped by amylose to form single-helices or V-type

lower viscosity (Marcotte et al., 2001; Sudhakar et al., 1996), PE tends to increase the mobility of starch chains (especially side chains of amylopectin), resulting in a higher degree of molecular order  $(R_{1045/1022})$  and enhanced aggregated structure ( $\alpha$ ) of MERS/PE composites. Moreover, the lower viscosity of PE may also allow it to encapsulate starch domains in the blends. All these structural changes could provide a shielding effect on the binding site of starch chains against amylase. In this way, the hydrolysis rate and degree of hydrolysis of rice starch could be reduced with higher contents of SDC and RC.



**Fig. 8.** Schematic representation of the alterations in the multiscale structure and the digestibility of

337

#### MERS/PE and MERS/CG.

338

339	CG has a linear molecular structure so it can interact with starch through hydrogen bonding
340	more effectively via its main chain. This interaction during extrusion may also disrupt the original
341	double-helical structure and destroy the original A+B-type crystallites (lower $X_{A+B}$ ). The interaction
342	also contributes to a greater structural ordering including the formation of new molecular aggregates,
343	single helices, and V-type crystallites (with amylose), which can effectively shield the binding sites
344	of starch chains for amylase and restrict starch hydrolysis. Even CG with low addition could
345	effectively interact with starch to increase the RC content of MERS significantly, with a reduction in
346	SDC content.

## 347 **4 Conclusion**

This study has demonstrated the improved digestion resistance (higher RC and lower pGI) of 348 349 MERS/PE and MERS/CG composites prepared by thermomechanical treatment. A relevant model is 350 established and the mechanism is elucidated. PE has a branched-chain structure, could only form 351 hydrogen bonding with starch via its side chains, leading to higher contents of double helices and 352 A+B-type crystallites (higher  $X_{A+B}$ ), and higher degrees of molecular aggregation ( $\alpha$ ), and 353 short-range order ( $R_{1045/1022}$ ). All these structural changes caused increases in SDC and RC contents. 354 On the other hand, CG, which has a linear chain structure, can interact with starch via its main chains 355 by hydrogen bonding. Therefore, CG is more effective at increasing the structural ordering of MERS, which has higher contents of single helices, V-type crystallites (higher  $X_V$ ), and leading to higher RC 356 content. The multiscale structural changes of starch due to the addition of NSPs such as PE and CG 357

- during extrusion could effectively shield the action sites on starch chains for amylase and restrict the
- interaction of amylase with starch, leading to a higher RC content and lower pGI.

## 360 Acknowledgements

- 361 The authors thank for the financial support provided by the National Natural Science Foundation of
- 362 China (NSFC)–Guangdong Joint Fund under a Key Project (No. U1501214), the National Natural
- 363 Science Foundation of China (NSFC) under a General Project (No. 31871751) and the Guangzhou
- 364 Science and Technology Program under a Key Project (No. 201804020036). F. Xie acknowledges
- 365 the support from the European Union's Horizon 2020 research and innovation programme under the
- 366 Marie Skłodowska-Curie grant agreement No. 798225.

## 367 **Conflicts of interest**

368 There are no conflicts of interest to declare.

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## **Highlights**

- $\checkmark$  Extruded rice starch (MERS) was prepared with pectin (PE) or  $\kappa$ -carrageenan (CG)
- ✓ Both MERS/PE and CG showed higher resistant components (RC) content
- ✓ Both MERS/PE and CG presented lower predicted glycemic index (pGI)
- ✓ Crystallinity and helical structures contributed to SDC and RC in MERS/PE
- ✓ More single helices and V-type crystallites  $(X_V)$  led to higher RC in MERS/CG

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## Declaration of Interest –

# Different effects of pectin and κ-carrageenan on the multiscale structures and *in vitro* digestibility of extruded rice starch

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The authors declare that there is no conflict of interest regarding the publication of this article.