






Review

# Barley's Second Spring as a Model Organism for Chloroplast Research

Lisa Rotasperti , Francesca Sansoni , Chiara Mizzotti , Luca Tadini and Paolo Pesaresi \* 

Dipartimento di Bioscienze, Università Degli Studi di Milano, 20133 Milano, Italy; lisa.rotasperti@unimi.it (L.R.); francesca.sansoni1@studenti.unimi.it (F.S.); chiara.mizzotti@unimi.it (C.M.); luca.tadini@unimi.it (L.T.)

\* Correspondence: paolo.pesaresi@unimi.it

Received: 10 June 2020; Accepted: 24 June 2020; Published: 27 June 2020



**Abstract:** Barley (*Hordeum vulgare*) has been widely used as a model crop for studying molecular and physiological processes such as chloroplast development and photosynthesis. During the second half of the 20th century, mutants such as *albostrians* led to the discovery of the nuclear-encoded, plastid-localized RNA polymerase and the retrograde (chloroplast-to-nucleus) signalling communication pathway, while *chlorina-f2* and *xantha* mutants helped to shed light on the chlorophyll biosynthetic pathway, on the light-harvesting proteins and on the organization of the photosynthetic apparatus. However, during the last 30 years, a large fraction of chloroplast research has switched to the more “user-friendly” model species *Arabidopsis thaliana*, the first plant species whose genome was sequenced and published at the end of 2000. Despite its many advantages, *Arabidopsis* has some important limitations compared to barley, including the lack of a real canopy and the absence of the proplastid-to-chloroplast developmental gradient across the leaf blade. These features, together with the availability of large collections of natural genetic diversity and mutant populations for barley, a complete genome assembly and protocols for genetic transformation and gene editing, have relaunched barley as an ideal model species for chloroplast research. In this review, we provide an update on the genomics tools now available for barley, and review the biotechnological strategies reported to increase photosynthesis efficiency in model species, which deserve to be validated in barley.

**Keywords:** Barley; genome; functional genomics; chloroplast biogenesis; photosynthesis improvement

## 1. barley, the Crop and the Model Species

Barley (*Hordeum vulgare*) is a self-pollinating monocotyledonous plant species that belongs to the *Poaceae*, a grass family that includes several major crops exploited in modern agriculture. Its domestication dates back to 10,000 BC, took place in the Fertile Crescent and began with the wild species *Hordeum vulgare* ssp. *spontaneum* [1]. Barley ranks fourth in terms of annual grain tonnage after maize, wheat and rice, with a worldwide production level (2018/2019) of 141 million tons. The primary role of cultivated barley (*Hordeum vulgare* ssp. *vulgare*) is as a source of animal feed (about 75% of the global production), with subsidiary uses in alcoholic and non-alcoholic beverages (20%), and in human nutrition (5%)—partly due to its high content of beta-glucan, a beneficial fibre that can reduce levels of cholesterol in the blood. During the 20th century, barley was widely exploited as a model species for crop studies. As a self-pollinating species with a diploid (2n) genome and a haploid complement of only seven chromosomes, barley proved to be an excellent model organism for both basic and applied research. Furthermore, due to the fact that wild barley (*Hordeum vulgare* ssp. *spontaneum*) can grow in a wide range of environments and climates, from the Arctic Circle to the equatorial highlands, barley is cultivated more widely than any other major crop. This resilience relies on a wealth of natural genetic

diversity which enables the plant to adapt effectively to various environmental challenges such as cold temperatures, drought, alkalinity and salinity, and makes it a perfect model species for investigating crop adaptation to abiotic stresses [2].

### 1.1. A Brief History of Genome Manipulation in Barley

*Hordeum vulgare* was one of the very first crops used in cereal improvement programs based on different induced mutation strategies. In 1930, Stadler studied the mutagenic effects of different types of radiation on maize and barley, describing chlorophyll-deficient and virescent phenotypes in seedlings [3]. In 1938, Nilsson-Ehle and Gustafsson tested X-rays and UV light on the barley cultivar (cv.) ‘Gull’ and isolated several mutants, which were named *albina*, *xantha*, *alboviridis*, *viridis*, *tigrina*, *striata* and *maculata*, categorizing them by their carotenoid and chlorophyll contents and distribution within the leaf blade [4]. The characteristics of several mutated lines were recognized as being very valuable for potential use in agriculture, since they exhibited alterations in grain yield, straw stiffness, straw length and tillering capacity, as well as changes in spike firmness, kernel maturation and pigmentation [5]. Later on, two varieties of barley ‘Trebil’ and ‘Moister’ were exposed to the radiation generated by the first aerial atomic explosion at Bikini atoll in 1946 [6]. Meanwhile, Gustafsson and Mackey applied mustard gas to barley to observe the effect of chemical mutagenesis [7], whereas Ehrenberg and collaborators tested various mutagenic compounds on barley and evaluated their impact on chlorophyll accumulation [8]. After these pioneering experiments, a broad range of chemical and physical mutagens were tested systematically. During this phase, alkylating agents able to generate G/C to A/T transitions in DNA, such as EMS (Ethyl Methane Sulfonate), ENU (N-nitroso-N-ethylurea), MNU (N-nitroso-N-methylurea), DES (diEthyl Sulfate) and sodium azide ( $\text{NaN}_3$ ), were widely used for the mutagenesis of barley. The first chemically induced barley variety, ‘Luther’, was released in the US in 1966. ‘Luther’ was obtained by exposing the variety ‘Alpine’ to DES. In 1965, in Czechoslovakia, the variety ‘Diamant’ was obtained after gamma-ray irradiation. This new variety was ~ 15 cm shorter than the parental ‘Valticky’ and displayed an increase in grain yield amounting to about 12% [9]. At around the same time, in the UK, ‘Golden Promise’ was registered. This semi-dwarf cultivar originated from exposure of the salt-sensitive variety ‘Maythorpe’ to gamma rays [10]. The generation of ‘Golden Promise’ represented an important step towards the development of tissue culture and barley transformation techniques (see below).

### 1.2. Early Studies and Milestones in Understanding of Chloroplast Biogenesis and Physiology in Barley

Genetic studies of barley have not been restricted to breeding programs. The plant has also been used as a model species to dissect the molecular mechanisms that underlie plant development and physiology and, for a large part of the 20th century, it served as a major experimental system for the investigation of chloroplast biogenesis and photosynthesis. In particular, several studies during the 1970s characterized different aspects of plastid structure and development, such as plastid growth, replication and differentiation. Dark-grown barley seedlings were used to determine the protochlorophyll content and structure of the etioplasts. Exposure to different light conditions allowed chloroplast development to be characterised from both structural and biochemical points of view [11–14].

The organization of chloroplast membranes was analysed in chloroplast preparations solubilised with digitonin and fractionated by electrophoresis, proving the existence of distinct sets of membranes [15]. The functionality and structural organization of thylakoids were also studied in barley mutants altered in chlorophyll biosynthesis [16] and revealed the impact of such changes on thylakoid membrane organization. For instance, the *chlorina-f2* mutant, which is impaired in chlorophyll b accumulation, led to the discovery of light-harvesting chlorophyll-binding proteins [17–20]. *Chlorina-f2* was also used to assess the impact of protein-chlorophyll complexes on the ultrastructure of thylakoid membranes, shedding light on the organisation of the photosynthetic apparatus [17,21]. In addition, the *tigrina-d* mutant [22], originally suggested to be involved in the early steps of tetrapyrrole

biosynthesis prior to ALA formation, was recently identified as the barley orthologue [23] of the *FLU* gene of *Arabidopsis thaliana*, a nuclear-encoded, plastid-localized protein that plays a key role in the negative-feedback control of chlorophyll biosynthesis, with an essential role during the dark-to-light switch [24]. Moreover, the barley *xantha* mutants helped to elucidate key steps in chlorophyll biosynthesis [25]: *xantha-l* was shown to code for a mutated form of Mg-protoporphyrin IX monomethyl ester cyclase, while *xantha-f*, *-g*, and *-h* carry genetic lesions at three distinct loci encoding the three Mg-chelatase subunits [26,27].

From a physiological point of view, Smith et al. [28] documented changes in chloroplast activity during de-etiolation of barley seedlings by measuring the Hill reaction in relation to chlorophyll accumulation. The correlation between plastid ultrastructure, chlorophyll synthesis and development of photosynthetic activity was also evaluated by measuring O<sub>2</sub> evolution [29].

Besides the characterization of the photosynthetic apparatus, barley played an important part in the dissection of the chloroplast's gene expression machinery. Indeed, evidence for a fully nuclear-encoded transcriptional activity in plastids, later named the Nuclear-Encoded RNA Polymerase (NEP; [30]), was first reported in barley, based on analysis of the *albostrians* mutant. In particular, the synthesis of RNA was reported in the white sectors of *albostrians* leaves, which harbor plastids that are devoid of ribosomes. These data provided initial evidence for the existence of a nuclear-encoded and plastid-localized RNA polymerase [31]. In addition, ribosome-free plastids of *albostrians* were helpful in distinguishing between the set of plastid genes preferentially transcribed by NEP, such as *rRNA*, *rpo* and *rps15*, and the set transcribed by the Plastid-Encoded RNA polymerase (PEP), which is enriched in photosynthesis-related genes such as *psbA*, *rbcL*, *atpI-H* [31,32]. Furthermore, the barley *albostrians* mutant was essential to the initial detection of communication between organellar and nuclear genomes. By analyzing *albostrians*, which is characterized by reduced amounts and/or activities of nucleus-encoded chloroplast proteins including the small subunit of ribulose-1,5-bisphosphate carboxylase/oxygenase (Rubisco), ferredoxin NADP<sup>+</sup> reductase, and enzymes of the Calvin cycle, Börner provided the first evidence for plastid signals that control nuclear gene expression, leading to the discovery of chloroplast-to-nucleus retrograde communication [33–35].

## 2. *Arabidopsis thaliana* as the Model Plant of Modern Times

In the 1990s, crop models lost their dominant position in basic research on plants to *Arabidopsis thaliana*, which has now reigned supreme for three decades. Its small size, short life cycle, ability to produce thousands of seeds from a single plant and simple growth requirements were perfectly compatible with lab facilities and research workflows. Moreover, its small diploid nuclear genome (~135 Mb on 5 chromosomes) and the *Agrobacterium tumefaciens*-based transformation protocol made *Arabidopsis* ideal for use in basic research [36]. The *Arabidopsis* Genome Project was initiated in 1990, and led to the publication of the first sequenced plant genome in 2000 [37]. This, together with large collections of insertional mutants (SIGnAL, <http://signal.salk.edu/cgi-bin/tdnaexpress>), permitted the functional characterization of large numbers of genes and biological processes, thus laying the foundations for the modern age of plant biology.

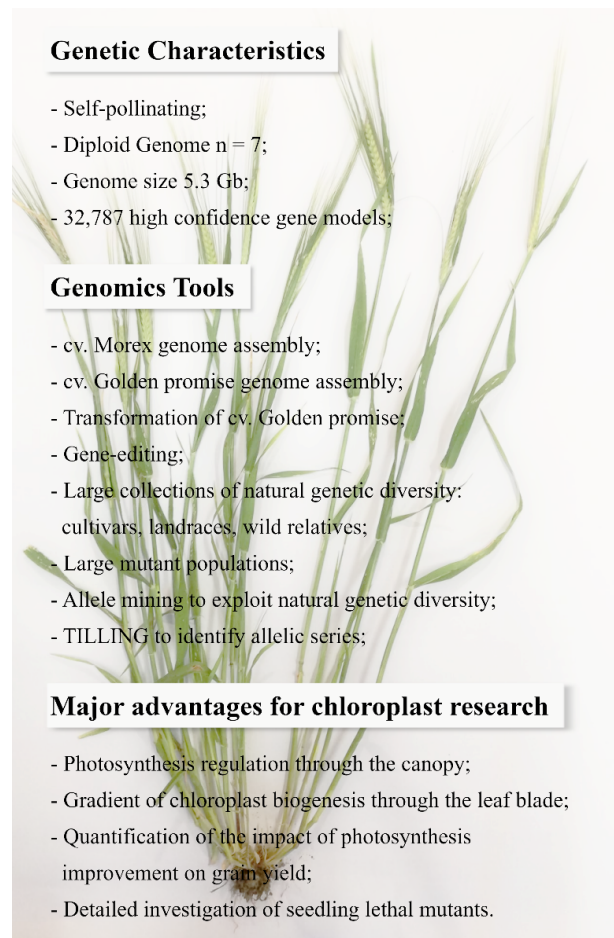
Although *Arabidopsis* has been considered the “golden” model species in plant science, it does have some limitations in terms of the extent to which lessons on development and physiology learnt from this model species can be extrapolated to monocots, including barley. This is particularly true for processes such as chloroplast biogenesis and photosynthesis. For instance, *Arabidopsis* does not produce an overhead canopy, and therefore cannot be employed in studies of plant architecture and optimization of photosynthesis under field conditions [38,39]. Moreover, the biogenesis of the multisubunit photosynthetic complexes, and indeed the chloroplast more generally, appear to differ significantly between monocotyledonous and dicotyledonous plants [40]. In monocots, the process of chloroplast development from the proplastid to functional chloroplasts can be observed as a gradient along the leaf blade, since leaves have a basal meristem and, as a consequence, the youngest cells carrying proplastids are found at the leaf base, while the leaf tip consists of the oldest cells with

mature chloroplasts [29,41,42]. In contrast, in dicots like *Arabidopsis thaliana* chloroplast development varies between plant organs—i.e., between cotyledons and leaves—and with respect to the leaves, most of the events take place inside the shoot apex, which constitutes a major limitation for functional studies [43,44]. In light of these limitations, the widely accepted knowledge transfer route from *Arabidopsis* to crops is not always the most convenient and effective strategy, especially in the era of next-generation sequencing technologies and gene-editing approaches that make functional genomics studies feasible in species with complex genomes.

### 3. The Genomes of Barley

#### 3.1. The Nuclear Genome

For a long time, the absence of a reference genome has been the major obstacle to the exploitation of barley genomic resources in both research and breeding programs. The relatively large size of the barley genome (5.3 Gb), together with its high proportion of repetitive DNA (more than 80%), has severely compromised the assembly of the whole-genome shotgun sequence and the generation of a reference genome (Figure 1). However, in 2012 the International Barley Sequencing Consortium circumvented these problems by integrating several different strategies. This involved coupling a detailed physical map of the barley cv. 'Morex' (a US spring six-row malting barley) with high-density genetic maps, superimposing deep short-read whole-genome shotgun assemblies, and annotating the resulting linear genomic sequence with dense-coverage RNA-derived, i.e., full-length cDNA and RNA-seq, data. This strategy allowed approximately 4 Gb (80%) of the genome to be delineated, including more than 90% of the expressed genes, together with their physical distribution and patterns of expression [45]. This partially ordered sequence assembly has since been substantially improved by Mascher and collaborators [46] through the release of a map-based reference sequence of the barley cv. 'Morex' genome that included the first comprehensively ordered assembly of the pericentromeric regions. The final genome sequence covered 4.79 Gb (approximately 95% of the total genome size), of which 4.54 Gb were assigned to precise chromosomal locations. Mapping of transcriptome data and reference protein sequences from other plant species to the assembly identified 39,734 high- and 41,949 low-confidence genes, representing 98% of the Morex gene complement. Furthermore, homology-guided repeat annotation identified 3.7 Gb (80.8%) of the assembled sequence as derived from transposable elements, most of which were present as truncated and degenerated copies, with only 10% of mobile elements being intact and potentially active. A second improved version of the barley cv. 'Morex' reference genome has recently been released [47]. This is based on the use of TRITEX, an open-source computational workflow, whose output is available on the IPK Barley BLAST server ([https://webblast.ipk-gatersleben.de/barley\\_ibsc/](https://webblast.ipk-gatersleben.de/barley_ibsc/), see Table 1). The need for an improved assembly arose from large sequence gaps and local mis-assemblies present in the first reference sequence. A total of 32,787 high- and 30,871 low-confidence gene models were annotated in the second version of the barley genome. The smaller number of high-confidence gene models described in the second version of the genome (32,787 vs 39,734) is certainly due to the more precise annotation process, making the TRITEX-based assembly a greatly improved version of the reference genome. More recently, a reference genome assembly for the barley cv. Golden Promise has been reported [48]. The assembled genome of seven chromosomes comprising 4.13 Gb contains 95.2% of complete and single-copy genes and will prove particularly useful for functional genomics studies, given that Golden Promise is the most readily transformable barley genotype.



**Figure 1.** Overview of the genetic characteristics and genomics tools available for barley. These features, together with its canopy architecture and developmental properties, make barley an optimal model for chloroplast research.

**Table 1.** List of databases, genome browsers and bioinformatics tools available for barley genome analyses.

Tool	Description/Application	URL	Reference
BARLEX	The Barley Genome Explorer permits visual inspection of BAC overlaps, and comparisons of BACs and provides useful information on genes and markers	<a href="http://barlex.barleysequence.org">http://barlex.barleysequence.org</a>	[49]
EnsemblPlants	A genome browser that incorporates genomic data from diverse organisms, including numerous plant species. It enables users to compare genome-scale datasets with the aid of a single collection of interfaces	<a href="http://plants.ensembl.org">http://plants.ensembl.org</a>	[50]
IPK Barley BLAST Server	Barley BLAST server for genome-scale homology-based searches	<a href="http://webblast.ipk-gatersleben.de/barley">http://webblast.ipk-gatersleben.de/barley</a>	[51]



Table 1. Cont.

Tool	Description/Application	URL	Reference
Golden Promise Genome	GMAP and BLAST server for barley (cv. Golden Promise) genome comparisons, including mapping of transcripts	<a href="https://ics.hutton.ac.uk/gmapper/">https://ics.hutton.ac.uk/gmapper/</a>	[48]
Gramene	Integrated data resource for comparative functional genomics in crops and model plant species	<a href="http://www.gramene.org">http://www.gramene.org</a>	[52]
PlantsDB	Provides data and information resources for individual plant species and a platform for integrative and comparative plant genome research.	<a href="http://pgsb.helmholtz-muenchen.de/plant/">http://pgsb.helmholtz-muenchen.de/plant/</a>	[53]
BaRTv1.0	Barley Reference Transcript Dataset provides access to 177,240 barley-expressed transcripts covering 60,444 genes	<a href="https://ics.hutton.ac.uk/barleyrtd/">https://ics.hutton.ac.uk/barleyrtd/</a>	[54]

### 3.2. The Exomes

A broader knowledge of the genetic diversity of barley is an essential prerequisite for the development of new varieties with increased yields and greater environmental robustness. A comprehensive genotyping of germplasm based on exome sequencing currently offers the best route to this goal. Sequencing of the coding DNA alone dramatically decreases the complexity of the task, and reduces the computational effort and associated costs compared with whole-genome approaches. This makes it highly suitable for crops like barley, which contain very high proportions of transposon DNA [46]. The application of the exome approach to barley was initially reported in Mascher et al. [55], and was first applied to examine the crop's adaptive responses in an analysis of 267 geo-referenced landraces and wild accessions [56]. This analysis combined exome capture and sequencing with field trials, bioclimatic data and various statistical approaches to investigate the genomic signatures that underlie barley's adaptive responses to various environmental stresses. A total of 1,688,807 SNPs and 143,872 short InDels were identified in 59.5 Mb of genomic sequence. The study yielded a large pool of genetic variation to be exploited in future breeding programs, as many of the SNPs identified were rare, showing an overall allele frequency below 5% and being more highly represented in wild accessions. A similar strategy based on exome capture sequencing [57] explored the genetics of barley adaptation to multiple contrasting environments in 371 domesticated lines, comprising cultivars and landraces of both two- and six-rowed types. The identification of 435,431 SNPs uncovered significant genetic diversity—including a well-defined subset of spring-growth-habit barleys, made up of 111 cultivars and 63 landraces—as well as revealing strong differentiation at specific chromosome positions between two- and six-row barley lines, and high adaptation and heritability of phenotypes such as days to heading, plant height, 1000-grain weight and awn length.

## 4. Barley Genetic Resources: Natural and Induced Genetic Diversity

### 4.1. Natural Genetic Diversity

Crop improvement through crossing of high-performance cultivars has resulted in the loss of genetic diversity across cultivated genomes, a phenomenon known as the “domestication bottleneck” [58]. Therefore, landraces and wild accessions of barley are a precious pristine source of natural allelic variability that can be exploited in barley breeding programs, as has now been verified by exome sequencing assays (see above). Over the years, several research institutes around the world have collected barley accessions with the aim of preserving this genetic variability and making it accessible to breeders through the adoption of advanced methods that are better able to discover, dissect and deploy useful variations [46,59]. The major collections are maintained in institutions around the

world, and the most representative are listed in Table 2. Among them, the ICARDA Collection hosts 222,704 barley accessions. Most of these are advanced materials, such as released cultivars and research lines, while 22% are geo-referenced wild barley relatives and landraces. The International Barley Core Collection (BCC) is a research collection that aims to represent the fullest possible range of the extant diversity of wild and cultivated barley. About 1300 accessions are currently available. Of these, some 300 varieties and landraces are held in the IPK Gatersleben Genebank. Of special relevance is the WHEALBI collection (<http://www.whealbi.eu/>), which comprises 511 accessions. This source of material represents a worldwide selection of barley's genetic diversity, including landraces, cultivars and progenitors. In particular, the WHEALBI panel includes accessions originating from a wide range of locations covering key crop production regions in Europe, Africa, the Middle East and Asia. A subset of 371 domesticated lines chosen from the entire WHEALBI germplasm has been subjected to exome sequencing in order to correlate genomic and phenotypic data [57]. Various online platforms have been developed to facilitate searches of germplasm collections and provide detailed information on their origins and characteristics. Some of these are listed in Table 2.

**Table 2.** List of representative collections of natural variants of barley available at different institutions worldwide, together with online platforms that provide information about barley genetic resources.

Collections of natural genetic diversity		
Gene Bank	Country	URL
<b>PGRC</b> Plant Gene Resources of Canada, Saskatoon Research Centre, Agriculture and Agri-Food Canada)	Canada	<a href="https://pgrc.agr.gc.ca/index_e.html">https://pgrc.agr.gc.ca/index_e.html</a>
<b>NSGC</b> The National Small Grains Collection is part of the National Plant Germplasm System (NPGS) of the United States Department of Agriculture - Agricultural Research Service (USDA-ARS)	USA	<a href="https://www.ars.usda.gov/pacific-west-area/aberdeen-id/small-grains-and-potato-germplasm-research/docs/barley-wheat-genetic-stocks-collections-1/">https://www.ars.usda.gov/pacific-west-area/aberdeen-id/small-grains-and-potato-germplasm-research/docs/barley-wheat-genetic-stocks-collections-1/</a>
<b>ICARDA</b> International Centre for Agricultural Research in the Dry Areas	Global	<a href="https://grs.icarda.org/">https://grs.icarda.org/</a>
<b>IPK</b> Leibniz Institute of Plant Genetics and Crop Plant Research	Germany	<a href="http://gbis.ipk-gatersleben.de">http://gbis.ipk-gatersleben.de</a>
<b>WHEALBI</b> WHEAt and barley Legacy for Breeding Improvement	France	<a href="http://wheat-urgi.versailles.inra.fr/Projects/Achieved-projects/Whealbi">http://wheat-urgi.versailles.inra.fr/Projects/Achieved-projects/Whealbi</a>
<b>NORDGEN</b> Nordic Genetic Resources Centre	Sweden	<a href="https://www.nordgen.org/bgs/">https://www.nordgen.org/bgs/</a>
<b>GRU</b> Germplasm Resource Unit, John Innes Centre	UK	<a href="https://www.seedstor.ac.uk/">https://www.seedstor.ac.uk/</a>
<b>NARO</b> NIAS, National Institute of Agrobiological Sciences	Japan	<a href="https://www.gene.affrc.go.jp/databases_en.php">https://www.gene.affrc.go.jp/databases_en.php</a>
Online platforms for barley germplasm searches		
Name	Description	URL
<b>GENESIS</b>	An online platform containing information about plant genetic resources for food and agriculture, conserved in gene banks worldwide	<a href="https://www.genesys-pgr.org/">https://www.genesys-pgr.org/</a>
<b>SINGER (The system-wide Information Network for Genetic Resources)</b>	An online catalogue of crop collections together with their locations	<a href="https://www.gbif.org/dataset/85818aea-f762-11e1-a439-00145eb45e9a">https://www.gbif.org/dataset/85818aea-f762-11e1-a439-00145eb45e9a</a>
<b>EURISCO (The European Search Catalogue for Plant Genetic Resources)</b>	Information on more than 2 million crop plant accessions and their wild relatives, preserved ex situ by almost 400 institutes in Europe and beyond	<a href="https://www.ecpgr.cgiar.org/resources/germplasm-databases/eurisco-catalogue/">https://www.ecpgr.cgiar.org/resources/germplasm-databases/eurisco-catalogue/</a>

#### 4.2. Induced Genetic Diversity: Random Chemical and Physical Mutagenesis

Besides natural genetic diversity, the availability of barley mutants is very important for understanding gene functions and their links with phenotypical traits (Figure 1). As described above (see Section 1.1), chemical and physical agents have been used to generate random mutagenized barley populations by several research groups. A few of these populations, derived from diverse genetic backgrounds, are listed in Table 3. Two large-scale EMS mutant populations from the cv. Optic, for instance, have been developed [60] and comprise approximately 20,000 M<sub>2</sub> plants. TILLMore, a sodium azide-mutagenized population of cv. Morex, has also been created [61] and consists of 4906 M<sub>3</sub> families. More recently, the *HorTILLUS* (*Hordeum*—TILLING—University of Silesia) population, derived from the spring barley cultivar Sebastian following treatment of seeds with two chemical mutagens (NaN<sub>3</sub> and MNU) and consisting of more than 9600 M<sub>2</sub> plants, was reported [62]. However, one limitation of the available resources is that the parental cultivars used for mutant population development are all recalcitrant to genetic transformation. Consequently, gene-specific complementation assays, which are essential for phenotype-to-genotype association, are generally not possible. To mitigate this drawback, a heavily mutagenized EMS population of cv. Golden Promise (the reference variety used across the barley research community for genetic transformation and functional genomics) has been developed [63]. This population permits direct complementation of candidate mutations, opening up new possibilities for efficient functional genomics studies.

**Table 3.** List of representative barley mutant populations obtained by either chemical or physical mutagenesis in different genetic backgrounds, i.e., cultivars.

Induced Mutant Populations		
Cultivar	Mutagen	Reference
Optic	EMS	[60]
Barke	EMS	[64]
Morex	NaN <sub>3</sub>	[61]
Lux	NaN <sub>3</sub>	[65]
DH-930-36	MNU	[66]
DH-930-36	Gamma rays	[66]
Sebastian	NaN <sub>3</sub> +MNU	[62]
Golden Promise	EMS	[63]

During the last 15 years, mutagenized populations have emerged as a key resource for gene discovery [67]. Using forward genetics approaches, many genes, especially those that confer morphological or developmental phenotypes, have now been isolated [68–71]. In addition, Targeting Induced Local Lesions in Genomes (TILLING; [72]) has become particularly powerful for gene validation studies, and for exploring the roles of genes for which no obvious visual phenotype can be predicted, i.e., the reverse genetics approach [73]. Moreover, the TILLING approach produces allelic series, which are important for genes whose knock-out would be lethal, but also in cases where proteins/enzymes with novel properties are needed. Identification of the DNA sequence changes responsible for mutant phenotypes has been performed, so far, through heteroduplex analysis [74]. This involves amplification of the gene of interest from a DNA pool, enzymatic cleavage of heteroduplexes formed by allelic mismatches, and detection of the cleaved strands in polyacrylamide gels, followed by sequencing for confirmation of the variation. However, with the advent of Next Generation Sequencing (NGS) technologies and the availability of reference genomes, the use of exome capture sequencing and/or pooled amplicon sequencing of multiple target genes appears to be more effective in the case of barley [63,75]. Moreover, it is worth mentioning that these mutagenized barley plants are not



considered as Genetically Modified (GM), and can be used in field trials to evaluate their performance, even in countries in which the cultivation of GM organisms is forbidden.

#### 4.3. Induced Genetic Diversity: Genome Transformation and Insertional Mutagenesis

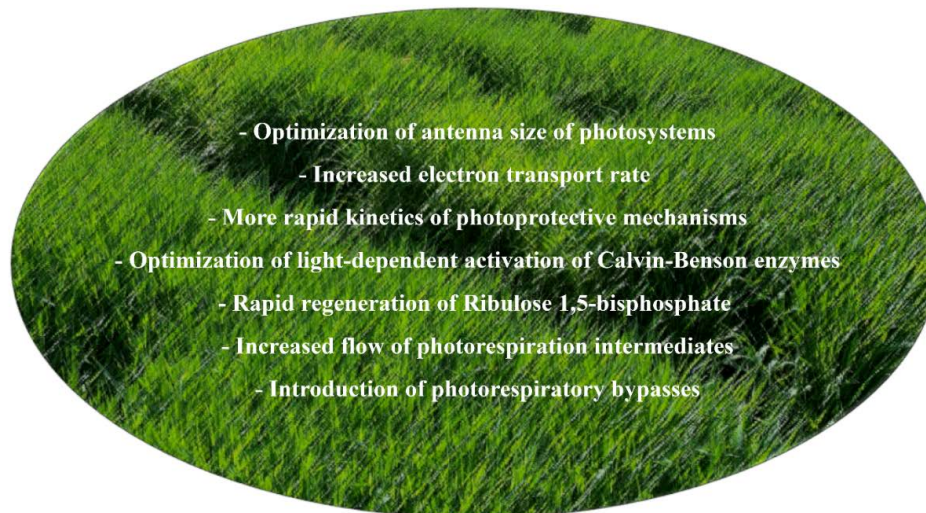
The need for complete sequencing of the barley genome, the range of genetic diversity and the numerous mutant populations all underline the importance of developing an efficient and versatile transformation protocol for functional genomics studies. In barley, many types of explant tissues have been used for tissue culture and plant regeneration, and immature embryos have proven to be the most suitable for barley transformation. Immature embryos were first used as explants for barley transformation in 1986 [76], and gradually became the most popular system. However, plant regeneration from immature embryo-derived callus is influenced by genotype, with the highest rates of success having been obtained in the cv. Golden Promise [77]. Up to now, a variety of DNA delivery methods, which involve biological, chemical, mechanical and/or physical treatment, have proven effective in barley. However, *Agrobacterium*-mediated transformation seems to be the best strategy, since it is characterised by low cost, high efficiency, simple integration, stable inheritance and expression of the transgene over generations. Indeed, the current transformation protocol integrates *Agrobacterium* and immature embryos, yielding an average transformation efficiency of 25% [78]. This protocol is widely used for overexpression, RNAi (RNA interference) applications and, more recently, CRISPR/Cas 9-mediated gene editing, as discussed below. Transgenic approaches have been employed to control pathogens such as Barley Yellow Dwarf Virus (BYDV; [79]), *Fusarium graminearum* [80], leaf stripe disease (*Pyrenophora graminea*; [81]), powdery mildew (*Blumeria graminis* f. sp. *Hordei*; [82]) and stem rust (*Puccinia graminis* f. sp. *Tritici*; [83]). Moreover, the transgenic technology has also been used to increase tolerance to environmental stresses, such as drought [84,85] and frost [86–88], and to modify enzymes, such as  $\alpha$ -amylase,  $\beta$ -amylase,  $\beta$ -glucanase and (1,3;1,4)- $\beta$ -D-glucan endohydrolase [89–93], which have an influence on the brewing process. The transgenic technology is also essential for mutagenesis. In barley, insertional mutagenesis has been used to produce loss-of-function mutations based on transposable elements such as the Ac/Ds-based tagging system [94–96], and gain-of-function mutations using the activation tagging strategy that promotes or enhances, through random genomic insertion, the expression of neighbouring regions [95]. However, unlike the case in *Arabidopsis*, no large-scale T-DNA insertional populations are currently available for barley.

#### 4.4. Induced Genetic Diversity: Gene Editing

The availability of the whole genome sequence, together with the recently developed gene-editing strategies, makes targeted mutagenesis possible in barley [97]. Among the various customized endonucleases used in plant research, the type II Clustered Regularly Interspaced Short Palindromic repeat (CRISPR)/CRISPR-associated protein 9 (Cas9) system has proven to be the best tool for gene editing [98]. The CRISPR/Cas9 gene-editing technology is easy to design and very precise. It requires the synthesis of oligonucleotides which, once transcribed into RNA, guide the Cas9 enzyme to the desired target. Being based on RNA–DNA interaction, the method is quite specific. The CRISPR/Cas9 technology can be used to create null alleles (i.e., gene knock-outs), and by adding a designed DNA template to the CRISPR/Cas9 system, it is possible to replace the target sequence via the error-free homology-directed repair pathway [99]. Furthermore, the system can be used to modulate gene expression [100]. In the past few years, the CRISPR/Cas9 technique has been increasingly applied to barley. In particular, a simple and efficient CRISPR/Cas9 platform for the induction of single and multiple, heritable mutations has been introduced [101]. The CRISPR/Cas9 technique has also been utilised to study genes involved in responses to pathogens, such as *HvMORC1*, whose protein product is one of the seven MORC members encoded by the barley genome [102]. Knock-out alleles of *HvCKX1* or *HvCKX3*, which are involved in the regulation of cytokinin metabolism and root morphology [103], as well as mutants in the *D-hordein* gene, which participates in the control of grain size and grain composition in barley [104], were also generated with the aid of CRISPR/Cas9.

## 5. Barley is Ready for a New Age of Functional Genomics Studies and Genetic Improvements

With the aid of the recently acquired collection of functional genomics tools, in a large part described above, the unique potential of barley as an ideal system for functional genomics studies can now be fully exploited. These new methods can elucidate, for instance, the molecular mechanisms behind chloroplast-to-nucleus communication, which is essential for chloroplast biogenesis and leaf emergence, leaf senescence and adaptation to environmental stresses. They can also be used to test—in an established crop plant—strategies intended to increase photosynthesis efficiency and biomass production, which have been shown to work in model species (Figure 2).



**Figure 2.** Biotechnological strategies that have been shown to enhance photosynthetic efficiency in model species. All of them can be applied to barley using the available genetic tools, and can potentially be improved by exploiting the genetic diversity of barley.

### 5.1. Plastid-to-Nucleus Retrograde Signalling

The chloroplast genome in barley encodes only 78 of the 3000 proteins that compose the plastid proteome [105,106]. The rest now reside in the nuclear DNA. Hence, signalling pathways that allow plastid and nuclear genomes to communicate with each other are essential for proper chloroplast development and functionality. The plastid-to-nucleus component of this circuit is often referred as “retrograde signalling” and it was first discovered in the barley mutant *albostrians* (see Section 1.2), which lacks plastid ribosomes and, concomitantly, shows reduced amounts and/or activities of nuclear-encoded plastidic proteins [33–35]. This channel is used to keep the nucleus informed of the developmental state of plastids (known as biogenic control), but it also signals changes in the functional status of fully developed chloroplasts in response to environmental factors, a process termed operational control [107]. Thus, chloroplast-to-nucleus communication is vital for chloroplast biogenesis and leaf emergence, as well as for the transition from chloroplast to gerontoplast during leaf senescence. Since the modulation of early leaf emergence and leaf senescence extends the proportion of the photosynthetically active radiation (PAR) that is intercepted by the crop over the growing season (the interception efficiency), both aspects of this communication are likely to be important determinants of crop yield. In addition, leaf senescence is a central process in maximising the efficiency of nutrient use, i.e., the ability of the plant to mobilise and translocate nutrients from leaves to grains. This is particularly true for small-grained cereals like barley, where up to 90% of the nitrogen is mobilised to the grains, mainly from the photosynthetic apparatus present in the leaves, and including Rubisco.

During the last 15 years, studies performed mainly in *Arabidopsis thaliana* have revealed a complex network of signals that allows chloroplasts to communicate their functional status to the nucleus. Singlet oxygen [108], H<sub>2</sub>O<sub>2</sub> [109], the redox state of the photosynthetic electron transport

chain [110], 3'-phosphoadenosine 5'-phosphate [111], the isoprenoid precursor methylerythritol cyclodiphosphate [112],  $\beta$ -cyclocitral [113,114] and other potential candidates [115–119] have been added to the list of operational control signals [107]. Understanding the degree to which these pathways are operative in monocot species like barley, together with a deeper knowledge of the regulatory, biochemical and redox networks that control the stability, functionality and disassembly of the photosynthetic apparatus under stress conditions and during induced senescence is pivotal for the identification of novel genes and favourable allelic variants for use in breeding programs.

Chloroplast biogenesis during leaf emergence is initiated upon light perception and is also dependent on plastid retrograde signals. Over the past two decades, many publications have explored the role of plastid gene expression and tetrapyrrole biosynthesis as sources of biogenic signals. This system is disrupted in 'genomes uncoupled' mutants (*gun*; [120]). Five of the six GUN proteins (GUN2-6) are enzymes of the tetrapyrrole biosynthesis pathway and control the branched pathways downstream of Protoporphyrin IX (for a review see [121]). GUN1, however, does not take part in tetrapyrrole biosynthesis, but is required for the generation of retrograde signals triggered by the accumulation of tetrapyrrole precursors and inhibition of plastid gene expression (PGE) [122]. More recently, GUN1 has been reported to play a prominent role in the maintenance of chloroplast protein homeostasis by modulating plastid protein synthesis through its interaction with the plastid ribosomal protein S1 [123], and to control the activity of the plastid protein import machinery [124,125], suggesting that unimported preproteins in the cytosol could act as messenger molecules. Although most of the information on biogenic retrograde signalling has been obtained in *Arabidopsis*, the barley mutant *albostrians* has proved to be a valuable system for studying the regulation of tetrapyrrole biosynthesis and the involvement of these compounds in communication between plastids and the nucleus. Due to the lack of plastid-encoded proteins in this mutant, low levels of tRNA<sup>Glu</sup>, which serves as a substrate activator in tetrapyrrole biosynthesis, are observed in bleached *albostrians* leaves, and this might be one reason for the much lower chlorophyll content in *albostrians* plastids [126–128]. In the mutant, the common precursors of all tetrapyrroles are channelled in the direction of heme synthesis, while the formation of chlorophylls is repressed [127]. This, in turn, suggests that excess heme might leave the chloroplast and act as a signalling molecule, as has been observed in *Arabidopsis* [129]. Recently, the mutation responsible for the *albostrians* phenotype has been identified. It lies in the barley gene *HvCMF7*, which codes for a putative plastid protein that belongs to the CCT motif family (CMF), which includes CONSTANS, CO-like and TOC1 [130]. This gene is likely to play a crucial role in plastid ribosome formation during early embryo development and hence for chloroplast development. The identification of the gene defect that causes the *albostrians* phenotype represents a major step forward in the understanding of the molecular mechanisms that mediate chloroplast biogenesis in barley. In this context, it would be interesting to determine whether a GUN1-like protein exists in barley. Furthermore, the gradient in chloroplast biogenesis observed in the barley leaf blade provides access to leaf sectors that contain cells of the same developmental stage, which facilitates the use of RNA-seq and proteomics approaches to investigate the molecular network at the basis of proplastid-to-chloroplast differentiation.

## 5.2. Photosynthesis and Yield

Doubling agricultural production by 2050 is essential if the demands of a constantly growing population for food and biomass are to be satisfied. Among cereals, barley straw is characterised by the highest content of carbohydrates [131]. Barley is therefore an ideal feedstock for the bio-based economy, since it can be used for the production of food/feed/spirits from grains, and renewable resources, including biofuel, from straw. It is worth mentioning here that none of the huge improvements in agricultural production made during the 'Green Revolution' were directly related to manipulations of photosynthesis. Hence, the process remains an unexplored target with a high potential for crop improvement. Indeed, the theoretical maximum efficiency of the conversion of solar energy into biomass in a C3 crop like barley is around 4.6% and this value decreases to 2.4% under field conditions

across the entire growing season. Therefore, the conversion efficiency of visible solar energy is considerably below its theoretical maximum, and several promising targets for its improvement have been identified in model species, some of which are described below (Figure 2 and Table 4).

### 5.3. Optimization of Antenna Size in Crop Canopies

One of the main reasons for the lower conversion efficiency of solar radiation is the saturation of the photosynthetic machinery with light. Indeed, it has been demonstrated that the photosynthetic apparatus operates with near maximum efficiency when light levels are low. For instance, the photosynthetic process in a C3 plant is already saturated at approximately 25% of maximum sunlight [153]. As light absorption increases, photosynthetic efficiency declines. In fact, the antennal apparatus of photosystems is larger than optimal, since under competitive natural conditions, shading of neighbouring plants confers an important selective advantage on the upper storey [154]. However, this behaviour is clearly disadvantageous for cultivated crops. Reducing the size of the antenna systems in the leaves of the upper canopy can offer important advantages, saving the metabolic resources required for the production of the antenna complex and the activity of photoprotective mechanisms, while increasing the amount and quality of light able to reach the lower leaves [155]. Several studies have provided evidence that the reduction of antenna size can improve photosynthetic efficiency. For instance, cell suspensions of *Synechocystis* PCC6714 and *Chlorella pyrenoidosa* with reduced contents of light-harvesting pigments showed a photosynthetic activity 20–30% higher than the wild type [156]. An engineered *Chlamydomonas reinhardtii* strain with a small PSII antenna size exhibited about a 50% increase in photosynthetic efficiency under saturating levels of light [157]. In the same alga, a partial reduction in chlorophyll b levels resulted in a two-fold increase in photosynthetic rate at high light intensities [158]. The hypothesis that a constitutively smaller antenna size should improve canopy photosynthetic efficiency by minimizing the over-absorption of the incident sunlight, and improving canopy light penetration, has also been tested in higher plants. A decrease in antenna size in tobacco, for instance, led to an increase of about 25% in plant–canopy biomass accumulation under high-density cultivation conditions [39]. Similarly, beneficial effects were observed in a rice genotype with pale green leaves cultivated under high light conditions [132].

### 5.4. Increased Photosynthetic Electron Transport

The modification of the thylakoid electron transport chain has also been reported to contribute to the improvement of photosynthetic performance and biomass accumulation. For instance, Chida et al. [133] showed that the expression of the algal *Porphyra yezoensis* cytochrome  $c_6$  in the chloroplasts of *Arabidopsis* led to an increase of CO<sub>2</sub> assimilation, and biomass production [133]. The overexpression of cytochrome  $c_6$  from *Ulva fasciata* in tobacco gave similar results [134]. Moreover, overexpression of plastocyanin in *Arabidopsis* resulted in 1.6-fold increase in leaf area [135]. A large increase in biomass and seed yield was also obtained in *Arabidopsis* upon overexpression of its endogenous Rieske FeS protein, a subunit of the cytochrome  $b_6/f$  [136].

**Table 4.** Brief summary of biotechnological strategies that are being employed for photosynthesis improvement in barley.

Target	Efficiency Gain	Strategy	Outcome	References
Retrograde signalling				
1. Investigating the existence of a GUN1-dependent retrograde signalling pathway in barley	Not expected	Knock-out of the HORVU.MOREX.r2.5HG0366860.1 gene through gene editing	Molecular details of the retrograde signalling pathway involved in chloroplast biogenesis with possible repercussions for the control of leaf life-cycle	[122]
Light phase of photosynthesis				
1. Optimisation of the antenna size	20–50%	Reduction of photosystem antenna size obtained by either reducing chlorophyll production or decreasing levels of the photosystem antenna proteins by gene editing or introgression of induced mutations. Identification of allelic variants by allele mining and TILLING.	More uniform photosynthetic performance throughout the crop canopy and prevention of photo-oxidative damage in the upper layers of the canopy. Increases in land–surface reflectivity to offset greenhouse gas warming.	[39,132]
2. Increased photosynthetic electron transport	30–70%	Increased accumulation of electron carriers, such as cytochrome <i>c</i> <sub>6</sub> , plastocyanin and Rieske proteins, by transgenic approaches. Identification of allelic variants by allele mining and TILLING.	Increased electron transport rate through the thylakoid membranes	[133–136]
3. Fine-Tuning of NPQ	30%	Increased accumulation of VDE, ZEP and PsbS by transgenic approaches. Identification of allelic variants by allele mining and TILLING.	More rapid induction and relaxation of heat dissipation at PSII.	[137]
Dark phase of photosynthesis				
1. Increasing the abundance of different enzymes of the Calvin–Benson cycle	>30%	Increased accumulation of SBPase and FBPA enzymes by transgenic approaches. Identification of allelic variants by allele mining and TILLING.	Optimization of ribulose 1,5-bisphosphate (RuBP) regeneration.	[138–142]
2. Increasing the efficiency of light activation of Calvin–Benson enzymes	>20%	Increased accumulation of Rubisco activase, TRX <i>f</i> and NTRC by transgenic approaches. Identification of allelic variants by allele mining and TILLING.	More efficient light-dependent activation of Calvin–Benson enzyme optimises CO <sub>2</sub> fixation.	[143–147]
Photorespiration				
1. Increasing the photorespiration flow of intermediates	>15%	Increased accumulation of H- and L-proteins by transgenic approaches.	Reduced accumulation of photorespiration intermediates and increased CO <sub>2</sub> assimilation rate	[148–150]
2. Synthetic bypasses to photorespiration	>20%	Introduction of natural and synthetic glycolate catabolic pathways in the chloroplast	Increased CO <sub>2</sub> assimilation rates	[151,152]



### 5.5. Improving the Adaptation to Fluctuating Light: Dissipation of Excess Energy through Non-Photochemical Quenching

Non-Photochemical Quenching (NPQ) serves as photoprotective mechanism in leaves, and is responsible for the dissipation of excess absorbed light energy as heat, thus preventing oxidative damage. Dissipation takes place in photosystem II (PSII) and involves the enzymes violaxanthin de-epoxidase (VDE) and zeaxanthin epoxidase (ZEP) [159], together with PsbS, a PSII protein subunit [160]. Activation and relaxation of NPQ take place over timescales of seconds to minutes, which are rather slow with respect to the instantaneous changes in light intensities observable within plant canopies in field settings. This leads to loss of photosynthetic efficiency, as heat dissipation continues even when light does not exceed the photosynthetic capacity [161]. Recently, the overexpression of *PsbS*, *ZEP* and *VDE* genes was reported in tobacco plants. These plants displayed an improved kinetics of NPQ, resulting in about 20% increase in biomass accumulation under both greenhouse and field conditions [137].

### 5.6. Transgenic Manipulation of the Calvin–Benson Cycle

Attempts to improve photosynthetic efficiency through transgenic manipulations have also focused on the overexpression of single enzymes of the Calvin–Benson cycle (Table 4). For example, overexpression of sedoheptulose-1,7-bisphosphatase (SBPase) in *Arabidopsis* [139], tobacco [138,140] and tomato [141] has shown that an increased SBPase activity results in a 30–40% increase in biomass yield, depending on the species. More recently it was shown that significant increases in photosynthetic rates, biomass and grain yield can be achieved by augmenting SBPase activity in wheat [162]. In 2012, the overexpression of the fructose 1,6-bisphosphate aldolase (FBPA) enzyme in tobacco also resulted in an increase in biomass production of 10–30% [142]. Overall, these findings demonstrated that SBPase and FBPA are enzymes that can exert control over the flow of carbon in the Calvin–Benson cycle in a number of different species, and proved that their manipulation also benefits grain yield. Efforts to increase the light activation rate of the Calvin–Benson cycle have also yielded very promising results. For instance, the overexpression of maize Rubisco activase in rice increased the rate of Rubisco activation by light and at high temperature (40 °C; [143]). Increased levels of thioredoxin *f* (TRX *f*), which is known to reductively activate enzymes of the Calvin–Benson cycle, have also increased leaf weight and sugar content under both ambient and increased CO<sub>2</sub> conditions [144,145]. Overexpression of the chloroplast NADPH-dependent thioredoxin reductase (NTRC), also reported to interact with several Calvin–Benson enzymes, has also been shown to be beneficial for productivity in *Arabidopsis*. Indeed, the biomass increases in the NTRC-overexpressing *Arabidopsis* plants were between 2- and 2.5-fold in plants grown under long- and short-day conditions, respectively, at fluence levels of 600 μmol m<sup>-2</sup> s<sup>-1</sup> light [146,147]. Additionally, overexpression of NTRC has been reported to enhance tolerance to oxidative and drought stresses. These are traits of great significance for improvement of crop productivity under field conditions [163].

### 5.7. Photorespiration and Photorespiratory Bypasses

Photorespiration is also an important target to improve photosynthesis. For instance, the reversible conversion of glycine into serine that takes place in mitochondria is crucial for plants [164–169]. These reactions involve the pyridoxal phosphate-dependent enzyme glycine decarboxylase (P-protein), the THF-dependent enzyme aminomethyltransferase (T-protein), the NAD<sup>+</sup>-dependent enzyme dihydrolipoyl dehydrogenase (L-protein) and the lipoic acid-containing H-protein. In *Arabidopsis*, overexpression of the H- or L-protein resulted in an improvement in photosynthetic efficiency and larger biomass accumulation [148,149]. Similar results were also obtained with the mesophyll-specific overexpression of the H-protein in tobacco [150]. Besides increasing photorespiration flow and reducing accumulation of photorespiratory intermediates, the most promising strategies for enhancing productivity are based on photorespiratory bypasses, i.e., the introduction of alternative pathways to metabolize 2PG, thus liberating CO<sub>2</sub> in the chloroplast stroma for Rubisco fixation [151,152].

In particular, Kebeish et al. introduced the *Escherichia coli* glycolate catabolic pathway into Arabidopsis. In these transgenic plants, the glycolate derived from the dephosphorylation of 2-phosphoglycolate was converted into glycerate in the chloroplast without the release of ammonia, which can make nitrogen use more efficient. Moreover, CO<sub>2</sub> release was shifted from mitochondria to chloroplasts, based on the idea that CO<sub>2</sub> should have a better chance to be re-fixed by Rubisco if it is released in the chloroplast rather than in the mitochondria. As a result of the increased concentration of CO<sub>2</sub> in the chloroplasts and the reduced energy demand for photorespiration, transgenic plants grew faster and produced more biomass, indicating that the bypass effectively reduced photorespiration and enhanced photosynthesis. Inspired by this work, another group [152] introduced synthetic glycolate metabolic pathways that are more efficient than the endogenous pathway into tobacco chloroplasts. Flux through the synthetic pathways was maximized by inhibiting glycolate export from the chloroplast. These synthetic pathways were able to improve photosynthetic quantum yield by 20% and biomass productivity by >40% in replicated field trials.

It is reasonable to expect that the various transgenic approaches described above will result in increased photosynthetic quantum yield, biomass and, eventually, grain yield also in barley, although species-dependent effects were observed in the multigene manipulation of the Calvin–Benson cycle [138,139]. In addition to that, the large genetic diversity readily available in barley also allows the exploitation of natural and/or induced allelic variants of enzymes involved in defining the antenna size of photosystems, in thylakoid electron transport, NPQ, the Calvin–Benson cycle, and photorespiration, which could ameliorate barley yield (Table 4). These allelic variants can be identified either by allele mining of exome sequences of barley cultivars, landraces and wild varieties (see Table 2), or through TILLING of mutant populations (Table 3). As mentioned above, the latter approaches enable one to obtain new barley varieties by using the classical breeding approach based on crosses. Thus, the performance of the new varieties can be verified under field conditions and they could be grown even in countries that have banned the cultivation of genetically modified plants.

## 6. Conclusions

Its genetic diversity and the availability of a large collection of molecular tools make barley an ideal model crop for functional genomics studies related to chloroplast biogenesis and retrograde communication. Such studies will reveal to what extent retrograde signalling mechanisms are conserved between Arabidopsis and barley, and permit us to learn more about aspects of chloroplast biogenesis that are specific to monocots. The recent identification of the genetic factor responsible for the *albostrians* phenotype demonstrates that this type of analysis can now be effectively conducted in barley. The fact that the gene concerned, *HvCMF7*, encodes a protein that is apparently located exclusively in plastids highlights the need for systematic investigation of barley mutants with defects in chloroplast biogenesis. Furthermore, novel approaches to the screening of barley mutant populations are required to elucidate the molecular details of the chloroplast-to-nucleus communication. The genes and allelic variants identified in future studies could have an important impact in breeding programs, since retrograde communication controls the leaf life cycle.

Barley can also make a significant contribution to the testing of novel biotechnological strategies for improving photosynthesis, and the validation of their effects on biomass accumulation and grain yield. In recent decades, our knowledge of the photosynthetic process has increased substantially, and improvements in its efficiency have been demonstrated in different model species. The high level of conservation of the photosynthetic process strongly argues that similar enhancements can be achieved in barley. Thanks to the high content of sugars in the straw, barley could be transformed into a dual-purpose crop suitable for the production of biofuel from the straw, and food, feed and spirits from the grain. Furthermore, the use of barley varieties characterised by high photosynthetic efficiency and reduced antenna size of photosystems is a promising strategy for boosting productivity and water use efficiency, while increasing land–surface reflectivity to offset greenhouse gas warming. In light of the foreseeable rise in the demand for food by the middle of this century, and the fact that

the development and commercialization of a new plant variety with improved quality takes 10 to 15 years, concerted efforts to increase agricultural yields through manipulation of photosynthesis must be initiated immediately. The “redesign” of photosynthesis must represent one of the main pillars of the next “Green Revolution”.

**Author Contributions:** Conceptualization, P.P., L.R., F.S., and L.T.; methodology, P.P., L.R., F.S., and L.T.; validation, P.P., L.R., and F.S.; resources, P.P.; data curation, P.P., L.R., and F.S.; writing—original draft preparation, P.P., L.R., F.S., and L.T.; writing—review and editing, P.P. and L.R.; visualization, C.M.; supervision, P.P.; project administration, P.P.; funding acquisition, P.P. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research received funding from ERA-NET Cofund FACCE SURPLUS (BarPLUS grant id. 93).

**Acknowledgments:** We apologize to the authors who did not get their work discussed in this review due to space limitation. The authors thank Paul Hardy for critical reading of the manuscript.

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

- Zohary, D.; Hopf, M.; Weiss, E. Domestication of Plants in the Old World: The origin and spread of domesticated plants in Southwest Asia, Europe, and the Mediterranean Basin. In *Domestication of Plants in the Old World: The Origin and Spread of Domesticated Plants in Southwest Asia, Europe, and the Mediterranean Basin*; Oxford University Press: Oxford, UK, 2013; ISBN 9780191810046.
- Dawson, I.K.; Russell, J.; Powell, W.; Steffenson, B.; Thomas, W.T.B.; Waugh, R. Barley: A translational model for adaptation to climate change. *New Phytol.* **2015**, *206*, 913–931. [[CrossRef](#)] [[PubMed](#)]
- Stadler, L.J. Some genetic effect of X-rays in plants. *J. Hered.* **1930**, *21*, 3–20. [[CrossRef](#)]
- Afsson, Å.K.E.G. Studies on the genetic basis of chlorophyll formation and the mechanism of induced mutating. *Hereditas* **1938**, *24*, 33–93. [[CrossRef](#)]
- Afsson, A.K.E.G. Mutation experiments in barley. *Hereditas* **1941**, *27*, 225–242. [[CrossRef](#)]
- Smith, L. Effects of atomic bomb radiations and x-rays on seeds of cereals: A comparison of the effects of ionizing radiations from the “ test able” atomic bomb and from x-rays on seeds of barley, wheat and oats. *J. Hered.* **1950**, *41*, 125–130. [[CrossRef](#)] [[PubMed](#)]
- Gustafsson, Å.; Key, J.M. The genetical effects of musterd gas substances and neutrons. *Hereditas* **1948**, *34*, 371–386. [[CrossRef](#)]
- Ehrenberg, L.; Gustafsson, Å.; Lundqvist, U.; Stenhagen, E.; Thorell, B. Chemically induced mutation and sterility in Barley. *Acta Chem. Scand.* **1956**, *10*, 492–494. [[CrossRef](#)]
- Bouma, J.; Ohnoutka, Z. Importance and application of the mutant “Diamant” in spring barley breeding. *Plant Mutat. Breed. Crop Improv.* **1991**, *1*, 127–134.
- Forster, B.P. Mutation genetics of salt tolerance in barley: An assessment of Golden Promise and other semi-dwarf mutants. *Euphytica* **2001**, *120*, 317–328. [[CrossRef](#)]
- Henningsen, K.W.; Boynton, J.E. Macromolecular physiology of plastids. VII. The effect of a brief illumination on plastids of dark-grown barley leaves. *J. Cell Sci.* **1969**, *5*, 757–793.
- Kannanga, G.; Gamini, C. The formation of Ribulose Diphosphate Carboxylase Protein during chloroplast development in Barley. *Plant Physiol.* **1969**, *44*, 1533–1537. [[CrossRef](#)] [[PubMed](#)]
- Wellburn, A.R.; Robinson, D.C.; Wellburn, F.A.M. Chloroplast development in low light-grown barley seedlings. *Planta* **1982**, *154*, 259–265. [[CrossRef](#)] [[PubMed](#)]
- Apel, K.; Gollmer, I.; Batschauer, A. The light-dependent control of chloroplast development in barley (*Hordeum vulgare* L.). *J. Cell. Biochem.* **1983**, *23*, 181–189. [[CrossRef](#)] [[PubMed](#)]
- Fradkin, L.I.; Kolyago, V.M.; Nisenbaum, G.D.; Domanskaya, I.N. Disintegration and fractionation of Barley chloroplast membranes at different concentrations of digitonin and chloroplasts. *Biokhimiya* **1978**, *43*, 723–733.
- Nielsen, N.C.; Smillie, R.M.; Henningsen, K.W.; Von Wettstein, D.; French, C.S. Composition and function of thylakoid membranes from grana-rich and grana-deficient chloroplast mutants of barley. *Plant Physiol.* **1979**, *63*, 174–182. [[CrossRef](#)]
- Thornber, J.P.; Highkin, H.R. Composition of the photosynthetic apparatus of normal barley leaves and a mutant lacking chlorophyll b. *Eur. J. Biochem.* **1974**, *41*, 109–116. [[CrossRef](#)]

18. Król, M.; Spangfort, M.D.; Huner, N.P.; Oquist, G.; Gustafsson, P.; Jansson, S. Chlorophyll a/b-binding proteins, pigment conversions, and early light-induced proteins in a chlorophyll b-less barley mutant. *Plant Physiol.* **1995**, *107*, 873–883. [[CrossRef](#)]
19. Król, M.; Ivanov, A.G.; Jansson, S.; Kloppstech, K.; Huner, N.P.A. Greening under high light or cold temperature affects the level of xanthophyll-cycle pigments, early light-inducible proteins, and light-harvesting polypeptides in wild-type barley and the chlorina f2 mutant. *Plant Physiol.* **1999**, *120*, 193–203. [[CrossRef](#)]
20. Bossmann, B.; Knoetzel, J.; Jansson, S. Screening of chlorina mutants of barley (*Hordeum vulgare* L.) with antibodies against light-harvesting proteins of PS I and PS II: Absence of specific antenna proteins. *Photosynth. Res.* **1997**, *52*, 127–136. [[CrossRef](#)]
21. Goodchild, D.J.; Highkin, H.R.; Boardman, N.K. The fine structure of chloroplasts in a barley mutant lacking chlorophyll B. *Exp. Cell Res.* **1966**, *43*, 684–688. [[CrossRef](#)]
22. Von Wettstein, D.; Kahn, A.; Nielsen, O.F.; Gough, S. Genetic Regulation of Chlorophyll Synthesis Analyzed with Mutants in Barley. *Science* **1974**, *184*, 800–802. [[CrossRef](#)] [[PubMed](#)]
23. Lee, K.P.; Kim, C.; Lee, D.W.; Apel, K. TIGRINA d, required for regulating the biosynthesis of tetrapyrroles in barley, is an ortholog of the FLU gene of Arabidopsis thaliana. *FEBS Lett.* **2003**, *553*, 119–124. [[CrossRef](#)]
24. Meskauskiene, R.; Nater, M.; Goslings, D.; Kessler, F.; Op den Camp, R.; Apel, K. FLU: A negative regulator of chlorophyll biosynthesis in Arabidopsis thaliana. *Proc. Natl. Acad. Sci. USA* **2001**, *98*, 12826–12831. [[CrossRef](#)]
25. Gustafsson, Å. Drastic morphological mutation in Barley. *Hereditas* **1946**, *32*, 120–122. [[CrossRef](#)]
26. Rzeznicka, K.; Walker, C.J.; Westergren, T.; Kannangara, C.G.; Von Wettstein, D.; Merchant, S.; Gough, S.P.; Hansson, M. Xantha-I encodes a membrane subunit of the aerobic Mg-protoporphyrin IX monomethyl ester cyclase involved in chlorophyll biosynthesis. *Proc. Natl. Acad. Sci. USA* **2005**, *102*, 5886–5891. [[CrossRef](#)] [[PubMed](#)]
27. Jensen, P.E.; Willows, R.D.; Petersen, B.L.; Vothknecht, U.C.; Stummann, B.M.; Kannangara, C.G.; Von Wettstein, D.; Henningsen, K.W. Structural genes for Mg-chelatase subunits in barley: Xantha-f, -g and -h. *Mol. Gen. Genet.* **1996**, *250*, 383–394. [[CrossRef](#)] [[PubMed](#)]
28. Smith, J.H.C.; French, C.S.; Koski, V.M. The Hill Reaction: Development of Chloroplast Activity During Greening of Etiolated Barley Leaves. *Plant Physiol.* **1952**, *27*, 212–213. [[CrossRef](#)] [[PubMed](#)]
29. Robertson, D.; Laetsch, W.M. Structure and Function of Developing Barley Plastids. *Plant Physiol.* **1974**, *54*, 148–159. [[CrossRef](#)]
30. Hajdukiewicz, P.T.J.; Allison, L.A.; Maliga, P. The two RNA polymerases encoded by the nuclear and the plastid compartments transcribe distinct groups of genes in tobacco plastids. *EMBO J.* **1997**, *16*, 4041–4048. [[CrossRef](#)]
31. Siemenroth, A.; Wollgiehn, R.; Neumann, D.; Börner, T. Synthesis of ribosomal RNA in ribosome-deficient plastids of the mutant “albostrians” of *Hordeum vulgare* L. *Planta* **1981**, *153*, 547–555. [[CrossRef](#)]
32. Hess, W.R.; Prombona, A.; Fieder, B.; Subramanian, A.R.; Börner, T. Chloroplast rps15 and the rpoB/C1/C2 gene cluster are strongly transcribed in ribosome-deficient plastids: Evidence for a functioning non-chloroplast-encoded RNA polymerase. *EMBO J.* **1993**, *12*, 563–571. [[CrossRef](#)] [[PubMed](#)]
33. Hagemann, R.; Börner, T.; Knoth, R. Plastid ribosome deficiency in plastid mutants of Pelargonium and Hordeum. *Genetics* **1973**, *74*, 103–104.
34. Börner, T.; Schumann, B.; Hagemann, R. Biochemical studies on a plastid ribosome-deficient mutant of *Hordeum vulgare*. In *Genetics Biogenesis of Chloroplasts Mitochondria*; Bücher, T., Neupert, W., Sebald, W., Werner, S., Eds.; Elsevier/North-Hill Medical Press: Amsterdam, The Netherlands, 1976; pp. 41–48.
35. Börner, T. The discovery of plastid-to-nucleus retrograde signaling—a personal perspective. *Protoplasma* **2017**, *254*, 1845–1855. [[CrossRef](#)] [[PubMed](#)]
36. Chilton, M.D.; Drummond, M.H.; Merlo, D.J.; Sciaky, D.; Montoya, A.L.; Gordon, M.P.; Nester, E.W. Stable incorporation of plasmid DNA into higher plant cells: The molecular basis of crown gall tumorigenesis. *Cell* **1977**, *11*, 263–271. [[CrossRef](#)]
37. Kaul, S.; Koo, H.L.; Jenkins, J.; Rizzo, M.; Rooney, T.; Tallon, L.J.; Feldblyum, T.; Nierman, W.; Benito, M.I.; Lin, X.; et al. Analysis of the genome sequence of the flowering plant Arabidopsis thaliana. *Nature* **2000**, *408*, 796–815. [[CrossRef](#)]



38. Song, W.; Li, C.; Sun, X.; Wang, P.; Zhao, S. Effects of ridge direction on growth and yield of tomato in solar greenhouse with diffuse film. *Nongye Gongcheng Xuebao/Trans. Chinese Soc. Agric. Eng.* **2017**, *33*, 242–248. [[CrossRef](#)]
39. Kirst, H.; Gabilly, S.T.; Niyogi, K.K.; Lemaux, P.G.; Melis, A. Photosynthetic antenna engineering to improve crop yields. *Planta* **2017**, *245*, 1009–1020. [[CrossRef](#)]
40. Pogson, B.J.; Ganguly, D.; Albrecht-Borth, V. Insights into chloroplast biogenesis and development. *Biochim. Biophys. Acta Bioenerg.* **2015**, *1847*, 1017–1024. [[CrossRef](#)]
41. Boffey, S.A.; Selldén, G.; Leech, R.M. Influence of Cell Age on Chlorophyll Formation in Light-grown and Etiolated Wheat Seedlings. *Plant Physiol.* **1980**, *65*, 680–684. [[CrossRef](#)]
42. Mullet, J.E. Chloroplast Development and Gene Expression. *Annu. Rev. Plant Physiol. Plant Mol. Biol.* **1988**, *39*, 475–502. [[CrossRef](#)]
43. Pogson, B.J.; Albrecht, V. Genetic dissection of chloroplast biogenesis and development: An overview. *Plant Physiol.* **2011**, *155*, 1545–1551. [[CrossRef](#)] [[PubMed](#)]
44. Jarvis, P.; López-Juez, E. Biogenesis and homeostasis of chloroplasts and other plastids. *Nat. Rev. Mol. Cell Biol.* **2013**, *14*, 787–802. [[CrossRef](#)] [[PubMed](#)]
45. Mayer, K.F.X.; Waugh, R.; Langridge, P.; Close, T.J.; Wise, R.P.; Graner, A.; Matsumoto, T.; Sato, K.; Schulman, A.; Ariyadasa, R.; et al. A physical, genetic and functional sequence assembly of the barley genome. *Nature* **2012**, *491*, 711–771. [[CrossRef](#)] [[PubMed](#)]
46. Mascher, M.; Gundlach, H.; Himmelbach, A.; Beier, S.; Twardziok, S.O.; Wicker, T.; Radchuk, V.; Dockter, C.; Hedley, P.E.; Russell, J.; et al. A chromosome conformation capture ordered sequence of the barley genome. *Nature* **2017**, *544*, 427–433. [[CrossRef](#)] [[PubMed](#)]
47. Monat, C.; Padmarasu, S.; Lux, T.; Wicker, T.; Gundlach, H.; Himmelbach, A.; Ens, J.; Li, C.; Muehlbauer, G.J.; Schulman, A.H.; et al. TRITEX: Chromosome-scale sequence assembly of Triticeae genomes with open-source tools. *Genome Biol.* **2019**, *20*, 284. [[CrossRef](#)] [[PubMed](#)]
48. Schreiber, M.; Mascher, M.; Wright, J.; Padmarasu, S.; Himmelbach, A.; Heavens, D.; Milne, L.; Clavijo, B.; Stein, N.; Waugh, R. A Genome Assembly of the Barley “Transformation Reference” Cultivar Golden Promise. *G3 Genes Genomes Genet.* **2020**, *10*, 1823–1827. [[CrossRef](#)]
49. Colmsee, C.; Beier, S.; Himmelbach, A.; Schmutzer, T.; Stein, N.; Scholz, U.; Mascher, M. BARLEX—The barley draft genome explorer. *Mol. Plant* **2015**, *8*, 964–966. [[CrossRef](#)]
50. Bolser, D.; Staines, D.M.; Pritchard, E.; Kersey, P. Ensembl plants: Integrating tools for visualizing, mining, and analyzing plant genomics data. *Methods Mol. Biol.* **2016**, *1533*, 1–31. [[CrossRef](#)]
51. Deng, W.; Nickle, D.C.; Learn, G.H.; Maust, B.; Mullins, J.I. ViroBLAST: A stand-alone BLAST web server for flexible queries of multiple databases and user’s datasets. *Bioinformatics* **2007**, *23*, 2334–2336. [[CrossRef](#)]
52. Tello-Ruiz, M.K.; Naithani, S.; Stein, J.C.; Gupta, P.; Campbell, M.; Olson, A.; Wei, S.; Preece, J.; Geniza, M.J.; Jiao, Y.; et al. Gramene 2018: Unifying comparative genomics and pathway resources for plant research. *Nucleic Acids Res.* **2018**, *46*, D1181–D1189. [[CrossRef](#)]
53. Spannagl, M.; Nussbaumer, T.; Bader, K.C.; Martis, M.M.; Seidel, M.; Kugler, K.G.; Gundlach, H.; Mayer, K.F.X. PGSB plantsDB: Updates to the database framework for comparative plant genome research. *Nucleic Acids Res.* **2016**, *44*, D1141–D1147. [[CrossRef](#)] [[PubMed](#)]
54. Rapazote-Flores, P.; Bayer, M.; Milne, L.; Mayer, C.D.; Fuller, J.; Guo, W.; Hedley, P.E.; Morris, J.; Halpin, C.; Kam, J.; et al. BaRTv1.0: An improved barley reference transcript dataset to determine accurate changes in the barley transcriptome using RNA-seq. *BMC Genom.* **2019**, *20*, 968. [[CrossRef](#)] [[PubMed](#)]
55. Mascher, M.; Richmond, T.A.; Gerhardt, D.J.; Himmelbach, A.; Clissold, L.; Sampath, D.; Ayling, S.; Steuernagel, B.; Pfeifer, M.; D’Ascenzo, M.; et al. Barley whole exome capture: A tool for genomic research in the genus *Hordeum* and beyond. *Plant J.* **2013**, *76*, 494–505. [[CrossRef](#)] [[PubMed](#)]
56. Russell, J.; Mascher, M.; Dawson, I.K.; Kyriakidis, S.; Calixto, C.; Freund, F.; Bayer, M.; Milne, I.; Marshall-Griffiths, T.; Heinen, S.; et al. Exome sequencing of geographically diverse barley landraces and wild relatives gives insights into environmental adaptation. *Nat. Genet.* **2016**, *48*, 1024–1030. [[CrossRef](#)]
57. Bustos-Korts, D.; Dawson, I.K.; Russell, J.; Tondelli, A.; Guerra, D.; Ferrandi, C.; Strozzi, F.; Nicolazzi, E.L.; Molnar-Lang, M.; Ozkan, H.; et al. Exome sequences and multi-environment field trials elucidate the genetic basis of adaptation in barley. *Plant J.* **2019**, *99*, 1172–1191. [[CrossRef](#)]



58. Hasan, M.; Hasibuzzaman, A.S.M.; Abdullah, H.M.; Kallol, M.M.H. Rediscovery of Genetic and Genomic Resources for Future Food Security. In *Genetic and Genomic Resources and their Exploitation for Unlocking Genetic Potential from the Wild Relatives*; Springer: Singapore, 2020; pp. 193–210.
59. Dempewolf, H.; Baute, G.; Anderson, J.; Kilian, B.; Smith, C.; Guarino, L. Past and future use of wild relatives in crop breeding. *Crop Sci.* **2017**, *57*, 1070–1082. [[CrossRef](#)]
60. Caldwell, D.G.; McCallum, N.; Shaw, P.; Muehlbauer, G.J.; Marshall, D.F.; Waugh, R. A structured mutant population for forward and reverse genetics in Barley (*Hordeum vulgare* L.). *Plant J.* **2004**, *40*, 143–150. [[CrossRef](#)]
61. Talamè, V.; Bovina, R.; Sanguineti, M.C.; Tuberosa, R.; Lundqvist, U.; Salvi, S. TILLMore, a resource for the discovery of chemically induced mutants in barley. *Plant Biotechnol. J.* **2008**, *6*, 477–485. [[CrossRef](#)]
62. Szurman-Zubrzycka, M.E.; Zbieszczczyk, J.; Marzec, M.; Jelonek, J.; Chmielewska, B.; Kurowska, M.M.; Krok, M.; Daszkowska-Golec, A.; Guzy-Wrobelska, J.; Gruszka, D.; et al. HorTILLUS—a rich and renewable source of induced mutations for forward/reverse genetics and pre-breeding programs in barley (*Hordeum vulgare* L.). *Front. Plant Sci.* **2018**, *9*, 216. [[CrossRef](#)]
63. Schreiber, M.; Barakate, A.; Uzrek, N.; Macaulay, M.; Sourdille, A.; Morris, J.; Hedley, P.E.; Ramsay, L.; Waugh, R. A highly mutagenised barley (cv. Golden Promise) TILLING population coupled with strategies for screening-by-sequencing. *Plant Methods* **2019**, *15*, 99. [[CrossRef](#)]
64. Gottwald, S.; Bauer, P.; Komatsuda, T.; Lundqvist, U.; Stein, N. TILLING in the two-rowed barley cultivar “Barke” reveals preferred sites of functional diversity in the gene HvHox1. *BMC Res. Notes* **2009**, *2*, 258. [[CrossRef](#)] [[PubMed](#)]
65. Lababidi, S.; Mejlhede, N.; Rasmussen, S.K.; Backes, G.; Al-Said, W.; Baum, M.; Jahoor, A. Identification of barley mutants in the cultivar “lux” at the dhn loci through tilling. *Plant Breed.* **2009**, *128*, 332–336. [[CrossRef](#)]
66. Kurowska, M.; Labocha-Pawłowska, A.; Gnizda, D.; Maluszynski, M.; Szarejko, I. Molecular analysis of point mutations in a barley genome exposed to MNU and Gamma rays. *Mutat. Res. Fundam. Mol. Mech. Mutagen.* **2012**, *738–739*, 52–70. [[CrossRef](#)] [[PubMed](#)]
67. Druka, A.; Franckowiak, J.; Lundqvist, U.; Bonar, N.; Alexander, J.; Houston, K.; Radovic, S.; Shahinnia, F.; Vendramin, V.; Morgante, M.; et al. Genetic dissection of barley morphology and development. *Plant Physiol.* **2011**, *155*, 617–627. [[CrossRef](#)] [[PubMed](#)]
68. Dockter, C.; Gruszka, D.; Braumann, I.; Druka, A.; Druka, I.; Franckowiak, J.; Gough, S.P.; Janeczko, A.; Kurowska, M.; Lundqvist, J.; et al. Induced variations in brassinosteroid genes define barley height and sturdiness, and expand the green revolution genetic toolkit. *Plant Physiol.* **2014**, *166*, 1912–1927. [[CrossRef](#)]
69. Jost, M.; Taketa, S.; Mascher, M.; Himmelbach, A.; Yuo, T.; Shahinnia, F.; Rutten, T.; Druka, A.; Schmutzer, T.; Steuernagel, B.; et al. A homolog of blade-on-petiole 1 and 2 (BOP1/2) controls internode length and homeotic changes of the barley inflorescence. *Plant Physiol.* **2016**, *171*, 1113–1127. [[CrossRef](#)]
70. Komatsuda, T.; Pourkheirandish, M.; He, C.; Azhaguvel, P.; Kanamori, K.; Perovic, D.; Stein, N.; Graner, A.; Wicker, T.; Tagiri, A.; et al. Six-rowed barley originated from a mutation in a homeodomain-leucine zipper I-class homeobox gene. *Proc. Natl. Acad. Sci. USA* **2007**, *104*, 1424–1429. [[CrossRef](#)]
71. Ramsay, L.; Comadran, J.; Druka, A.; Marshall, D.F.; Thomas, W.T.B.; MacAulay, M.; MacKenzie, K.; Simpson, C.; Fuller, J.; Bonar, N.; et al. INTERMEDIUM-C, a modifier of lateral spikelet fertility in barley, is an ortholog of the maize domestication gene TEOSINTE BRANCHED 1. *Nat. Genet.* **2011**, *43*, 169–172. [[CrossRef](#)]
72. Henikoff, S.; Till, B.J.; Comai, L. TILLING. Traditional mutagenesis meets functional genomics. *Plant Physiol.* **2004**, *135*, 630–636. [[CrossRef](#)]
73. Waugh, R.; Leader, D.J.; McCallum, N.; Caldwell, D. Harvesting the potential of induced biological diversity. *Trends Plant Sci.* **2006**, *11*, 71–79. [[CrossRef](#)]
74. Till, B.J.; Reynolds, S.H.; Greene, E.A.; Codomo, C.A.; Enns, L.C.; Johnson, J.E.; Burtler, C.; Odden, A.R.; Young, K.; Taylor, N.E.; et al. Large-scale discovery of induced point mutations with high-throughput TILLING. *Genome Res.* **2003**, *13*, 524–530. [[CrossRef](#)] [[PubMed](#)]
75. Henry, I.M.; Nagalakshmi, U.; Lieberman, M.C.; Ngo, K.J.; Krasileva, K.V.; Vasquez-Gross, H.; Akhunova, A.; Akhunov, E.; Dubcovsky, J.; Tai, T.H.; et al. Efficient genome-wide detection and cataloging of EMS-induced mutations using Exome capture and next-generation sequencing. *Plant Cell* **2014**, *26*, 1382–1397. [[CrossRef](#)] [[PubMed](#)]

76. Goldstein, C.S.; Kronstad, W.E. Tissue culture and plant regeneration from immature embryo explants of Barley, *Hordeum vulgare*. *Theor. Appl. Genet.* **1986**, *71*, 631–636. [[CrossRef](#)] [[PubMed](#)]
77. Dahleen, L.S.; Bregitzer, P. An improved media system for high regeneration rates from barley immature embryo-derived callus cultures of commercial cultivars. *Crop Sci.* **2002**, *42*, 934–938. [[CrossRef](#)]
78. Harwood, W.A. A protocol for high-throughput agrobacterium-mediated barley transformation. *Methods Mol. Biol.* **2014**, *1099*, 251–260. [[CrossRef](#)] [[PubMed](#)]
79. McGrath, P.F.; Vincent, J.R.; Lei, C.H.; Pawlowski, W.P.; Torbert, K.A.; Gu, W.; Kaeppler, H.F.; Wan, Y.; Lemaux, P.G.; Rines, H.R.; et al. Coat protein-mediated resistance to isolates of barley yellow dwarf in oats and barley. *Eur. J. Plant Pathol.* **1997**, *103*, 695–710. [[CrossRef](#)]
80. Hüchelhoven, R. BAX Inhibitor-1, an ancient cell death suppressor in animals and plants with prokaryotic relatives. *Apoptosis* **2004**, *9*, 299–307. [[CrossRef](#)]
81. Bulgarelli, D.; Biselli, C.; Collins, N.C.; Consonni, G.; Stanca, A.M.; Schulze-Lefert, P.; Valè, G. The CC-NB-LRR-Type RDG2a resistance gene confers immunity to the seed-borne barley leaf stripe pathogen in the absence of hypersensitive cell death. *PLoS ONE* **2010**, *5*, e12599. [[CrossRef](#)]
82. Eichmann, R.; Bischof, M.; Weis, C.; Shaw, J.; Lacomme, C.; Schweizer, P.; Duchkov, D.; Hensel, G.; Kumlehn, J.; Hüchelhoven, R. Bax inhibitor-1 is required for full susceptibility of barley to powdery mildew. *Mol. Plant-Microbe Interact.* **2010**, *23*, 1217–1227. [[CrossRef](#)]
83. Horvath, H.; Rostoks, N.; Brueggeman, R.; Steffenson, B.; Von Wettstein, D.; Kleinhofs, A. Genetically engineered stem rust resistance in barley using the Rpg1 gene. *Proc. Natl. Acad. Sci. USA* **2003**, *100*, 364–369. [[CrossRef](#)]
84. Morran, S.; Eini, O.; Pyvovarenko, T.; Parent, B.; Singh, R.; Ismagul, A.; Eliby, S.; Shirley, N.; Langridge, P.; Lopato, S. Improvement of stress tolerance of wheat and barley by modulation of expression of DREB/CBF factors. *Plant Biotechnol. J.* **2011**, *9*, 230–249. [[CrossRef](#)]
85. Seiler, C.; Harshavardhan, V.T.; Reddy, P.S.; Hensel, G.; Kumlehn, J.; Eschen-Lippold, L.; Rajesh, K.; Korzun, V.; Wobus, U.; Lee, J.; et al. Abscisic acid flux alterations result in differential abscisic acid signaling responses and impact assimilation efficiency in barley under terminal drought stress. *Plant Physiol.* **2014**, *164*, 1677–1696. [[CrossRef](#)] [[PubMed](#)]
86. Soltész, A.; Vágújfalvi, A.; Rizza, F.; Kerepesi, I.; Galiba, G.; Cattivelli, L.; Coraggio, I.; Crosatti, C. The rice Osmyb4 gene enhances tolerance to frost and improves germination under unfavourable conditions in transgenic barley plants. *J. Appl. Genet.* **2012**, *53*, 133–143. [[CrossRef](#)]
87. Soltész, A.; Smedley, M.; Vashegyi, I.; Galiba, G.; Harwood, W.; Vágújfalvi, A. Transgenic barley lines prove the involvement of TaCBF14 and TaCBF15 in the cold acclimation process and in frost tolerance. *J. Exp. Bot.* **2013**, *64*, 1849–1862. [[CrossRef](#)] [[PubMed](#)]
88. Kovalchuk, N.; Jia, W.; Eini, O.; Morran, S.; Pyvovarenko, T.; Fletcher, S.; Bazanova, N.; Harris, J.; Beck-Oldach, K.; Shavrukov, Y.; et al. Optimization of TaDREB3 gene expression in transgenic barley using cold-inducible promoters. *Plant Biotechnol. J.* **2013**, *11*, 659–670. [[CrossRef](#)] [[PubMed](#)]
89. Murray, F.; Matthews, P.; Jacobsen, J.; Gubler, F. Increased expression of HvGAMYB in transgenic barley increases hydrolytic enzyme production by aleurone cells in response to gibberellin. *J. Cereal Sci.* **2006**, *44*, 317–322. [[CrossRef](#)]
90. Kihara, M.; Okada, Y.; Kuroda, H.; Saeki, K.; Yoshigi, N.; Ito, K. Improvement of  $\beta$ -amylase thermostability in transgenic barley seeds and transgene stability in progeny. *Mol. Breed.* **2000**, *6*, 511–517. [[CrossRef](#)]
91. Tull, D.; Phillipson, B.A.; Kramhøft, B.; Knudsen, S.; Olsen, O.; Svensson, B. Enhanced amyolytic activity in germinating barley through synthesis of a bacterial Alpha-amylase. *J. Cereal Sci.* **2003**, *37*, 71–80. [[CrossRef](#)]
92. Jensen, L.G.; Olsen, O.; Kops, O.; Wolf, N.; Thomsen, K.K.; Von Wettstein, D. Transgenic barley expressing a protein-engineered, thermostable (1,3-1,4)- $\beta$ -glucanase during germination. *Proc. Natl. Acad. Sci. USA* **1996**, *93*, 3487–3491. [[CrossRef](#)]
93. Nuutila, A.M.; Ritala, A.; Skadsen, R.W.; Mannonen, L.; Kauppinen, V. Expression of fungal thermotolerant endo-1,4- $\beta$ -glucanase in transgenic barley seeds during germination. *Plant Mol. Biol.* **1999**, *41*, 777–783. [[CrossRef](#)]
94. Koprek, T.; McElroy, D.; Louwerse, J.; Williams-Carrier, R.; Lemaux, P.G. An efficient method for dispersing Ds elements in the barley genome as a tool for determining gene function. *Plant J.* **2000**, *24*, 253–263. [[CrossRef](#)] [[PubMed](#)]

95. Ayliffe, M.A.; Pallotta, M.; Langridge, P.; Pryor, A.J. A barley activation tagging system. *Plant Mol. Biol.* **2007**, *64*, 329–347. [[CrossRef](#)] [[PubMed](#)]
96. Lazarow, K.; Lütticke, S. An Ac/Ds-mediated gene trap system for functional genomics in barley. *BMC Genom.* **2009**, *10*, 55. [[CrossRef](#)] [[PubMed](#)]
97. Ryder, P.; McHale, M.; Fort, A.; Spillane, C. Generation of stable nulliplex autopolyploid lines of *Arabidopsis thaliana* using CRISPR/Cas9 genome editing. *Plant Cell Rep.* **2017**, *36*, 1005–1008. [[CrossRef](#)] [[PubMed](#)]
98. Jinek, M.; Chylinski, K.; Fonfara, I.; Hauer, M.; Doudna, J.A.; Charpentier, E. A Programmable Dual-RNA – Guided DNA Endonuclease S figs. *Science* **2012**, *337*, 816–821. [[CrossRef](#)]
99. Lawrenson, T.; Shorinola, O.; Stacey, N.; Li, C.; Østergaard, L.; Patron, N.; Uauy, C.; Harwood, W. Induction of targeted, heritable mutations in barley and *Brassica oleracea* using RNA-guided Cas9 nuclease. *Genome Biol.* **2015**, *16*, 258. [[CrossRef](#)]
100. Holme, I.B.; Wendt, T.; Gil-Humanes, J.; Deleuran, L.C.; Starker, C.G.; Voytas, D.F.; Brinch-Pedersen, H. Evaluation of the mature grain phytase candidate HvPAPhy\_a gene in barley (*Hordeum vulgare* L.) using CRISPR/Cas9 and TALENs. *Plant Mol. Biol.* **2017**, *95*, 111–121. [[CrossRef](#)]
101. Gasparis, S.; Kała, M.; Przyborowski, M.; Łyżnik, L.A.; Orczyk, W.; Nadolska-Orczyk, A. A simple and efficient CRISPR/Cas9 platform for induction of single and multiple, heritable mutations in barley (*Hordeum vulgare* L.). *Plant Methods* **2018**, *14*, 111. [[CrossRef](#)]
102. Kumar, N.; Galli, M.; Ordon, J.; Stuttmann, J.; Kogel, K.H.; Imani, J. Further analysis of barley MORC1 using a highly efficient RNA-guided Cas9 gene-editing system. *Plant Biotechnol. J.* **2018**, *16*, 1892–1903. [[CrossRef](#)]
103. Gasparis, S.; Przyborowski, M.; Kała, M.; Nadolska-Orczyk, A. Knockout of the HvCKX1 or HvCKX3 Gene in Barley (*Hordeum vulgare* L.) by RNA-Guided Cas9 Nuclease Affects the Regulation of Cytokinin Metabolism and Root Morphology. *Cells* **2019**, *8*, 782. [[CrossRef](#)]
104. Yang, Q.; Zhong, X.; Li, Q.; Lan, J.; Tang, H.; Qi, P.; Ma, J.; Wang, J.; Chen, G.; Pu, Z.; et al. Mutation of the D-hordein gene by RNA-guided Cas9 targeted editing reducing the grain size and changing grain compositions in barley. *Food Chem.* **2020**, *311*, 125892. [[CrossRef](#)] [[PubMed](#)]
105. Saski, C.; Tomkins, J.; Lee, S.-B.; Daniell, H.; Fjellheim, S.; Rognli, O.A.; Guda, C.; Jansen, R.K.; Luo, H.; Clarke, J.L. Complete chloroplast genome sequences of *Hordeum vulgare*, *Sorghum bicolor* and *Agrostis stolonifera*, and comparative analyses with other grass genomes. *Theor. Appl. Genet.* **2007**, *115*, 571–590. [[CrossRef](#)] [[PubMed](#)]
106. Petersen, J.; Rogowska-Wrzesinska, A.; Jensen, O.N. Functional proteomics of barley and barley chloroplasts-strategies, methods and perspectives. *Front. Plant Sci.* **2013**, *4*, 52. [[CrossRef](#)] [[PubMed](#)]
107. Pogson, B.J.; Woo, N.S.; Förster, B.; Small, I.D. Plastid signalling to the nucleus and beyond. *Trends Plant Sci.* **2008**, *3*, 602–609. [[CrossRef](#)]
108. Wagner, D.; Przybyla, D.; Op Den Camp, R.; Kim, C.; Landgraf, F.; Keun, P.L.; Würsch, M.; Laloi, C.; Nater, M.; Hideg, E.; et al. The genetic basis of singlet oxygen-induced stress response of *Arabidopsis thaliana*. *Science* **2004**, *306*, 1183–1185. [[CrossRef](#)]
109. Maruta, T.; Noshi, M.; Tanouchi, A.; Tamoi, M.; Yabuta, Y.; Yoshimura, K.; Ishikawa, T.; Shigeoka, S. H<sub>2</sub>O<sub>2</sub>-triggered retrograde signaling from chloroplasts to nucleus plays specific role in response to stress. *J. Biol. Chem.* **2012**, *287*, 11717–11729. [[CrossRef](#)]
110. Pfalz, J.; Liebers, M.; Hirth, M.; Grübler, B.; Holtzegel, U.; Schröter, Y.; Dietzel, L.; Pfannschmidt, T. Environmental control of plant nuclear gene expression by chloroplast redox signals. *Front. Plant Sci.* **2012**, *3*, 257. [[CrossRef](#)]
111. Estavillo, G.M.; Crisp, P.A.; Pornsiriwong, W.; Wirtz, M.; Collinge, D.; Carrie, C.; Giraud, E.; Whelan, J.; David, P.; Javot, H.; et al. Evidence for a SAL1-PAP chloroplast retrograde pathway that functions in drought and high light signaling in *Arabidopsis*. *Plant Cell* **2011**, *23*, 3992–4012. [[CrossRef](#)]
112. Xiao, Y.; Savchenko, T.; Baidoo, E.E.K.; Chehab, W.E.; Hayden, D.M.; Tolstikov, V.; Corwin, J.A.; Kliebenstein, D.J.; Keasling, J.D.; Dehesh, K. Retrograde signaling by the plastidial metabolite MEcPP regulates expression of nuclear stress-response genes. *Cell* **2012**, *149*, 1525–1535. [[CrossRef](#)]
113. Ramel, F.; Birtic, S.; Ginies, C.; Soubigou-Taconnat, L.; Triantaphylidès, C.; Havaux, M. Carotenoid oxidation products are stress signals that mediate gene responses to singlet oxygen in plants. *Proc. Natl. Acad. Sci. USA* **2012**, *109*, 5535–5540. [[CrossRef](#)]
114. D’Alessandro, S.; Havaux, M. Sensing  $\beta$ -carotene oxidation in photosystem II to master plant stress tolerance. *New Phytol.* **2019**, *223*, 1776–1783. [[CrossRef](#)] [[PubMed](#)]

115. Chi, W.; Feng, P.; Ma, J.; Zhang, L. Metabolites and chloroplast retrograde signaling. *Curr. Opin. Plant Biol.* **2015**, *25*, 32–38. [[CrossRef](#)] [[PubMed](#)]
116. Tian, L. Recent advances in understanding carotenoid-derived signaling molecules in regulating plant growth and development. *Front. Plant Sci.* **2015**, *6*, 790. [[CrossRef](#)] [[PubMed](#)]
117. Chan, K.X.; Phua, S.Y.; Crisp, P.; McQuinn, R.; Pogson, B.J. Learning the Languages of the Chloroplast: Retrograde Signaling and Beyond. *Annu. Rev. Plant Biol.* **2016**, *67*, 25–53. [[CrossRef](#)] [[PubMed](#)]
118. Kleine, T.; Leister, D. Retrograde signaling: Organelles go networking. *Biochim. Biophys. Acta Bioenerg.* **2016**, *1857*, 1313–1325. [[CrossRef](#)] [[PubMed](#)]
119. de Souza, A.; Wang, J.-Z.; Dehesh, K. Retrograde Signals: Integrators of Interorganellar Communication and Orchestrators of Plant Development. *Annu. Rev. Plant Biol.* **2017**, *68*, 85–108. [[CrossRef](#)]
120. Susek, R.E.; Ausubel, F.M.; Chory, J. Signal transduction mutants of arabidopsis uncouple nuclear CAB and RBCS gene expression from chloroplast development. *Cell* **1993**, *74*, 787–799. [[CrossRef](#)]
121. Larkin, R.M.; Stefano, G.; Ruckl, M.E.; Stavoe, A.K.; Sinkler, C.A.; Brandizzi, F.; Malmstrom, C.M.; Osteryoung, K.W.; Chory, J. REDUCED CHLOROPLAST COVERAGE genes from Arabidopsis thaliana help to establish the size of the chloroplast compartment. *Proc. Natl. Acad. Sci. USA* **2016**, *113*, E1116–E1125. [[CrossRef](#)]
122. Koussevitzky, S.; Nott, A.; Mockler, T.C.; Hong, F.; Sachetto-Martins, G.; Surpin, M.; Lim, J.; Mittler, R.; Chory, J. Signals from Chloroplasts Converge to Regulate Nuclear Gene Expression. *Science* **2007**, *316*, 715–719. [[CrossRef](#)]
123. Tadini, L.; Pesaresi, P.; Kleine, T.; Rossi, F.; Guljamow, A.; Sommer, F.; Mühlhaus, T.; Schroda, M.; Masiero, S.; Pribil, M.; et al. Gun1 controls accumulation of the plastid ribosomal protein S1 at the protein level and interacts with proteins involved in plastid protein homeostasis. *Plant Physiol.* **2016**, *170*, 1817–1830. [[CrossRef](#)]
124. Tadini, L.; Peracchio, C.; Trotta, A.; Colombo, M.; Mancini, I.; Jeran, N.; Costa, A.; Faoro, F.; Marsoni, M.; Vannini, C.; et al. GUN1 influences the accumulation of NEP-dependent transcripts and chloroplast protein import in Arabidopsis cotyledons upon perturbation of chloroplast protein homeostasis. *Plant J.* **2020**, *101*, 1198–1220. [[CrossRef](#)]
125. Wu, G.Z.; Meyer, E.H.; Richter, A.S.; Schuster, M.; Ling, Q.; Schöttler, M.A.; Walther, D.; Zoschke, R.; Grimm, B.; Jarvis, R.P.; et al. Control of retrograde signalling by protein import and cytosolic folding stress. *Nat. Plants* **2019**, *5*, 525–538. [[CrossRef](#)] [[PubMed](#)]
126. Börner, T.; Meister, A. Chlorophyll and carotenoid content of ribosome-deficient plastids. *Photosynthetica* **1980**, *14*, 589–593.
127. Yaronskaya, E.; Ziemann, V.; Walter, G.; Averina, N.; Börner, T.; Grimm, B. Metabolic control of the tetrapyrrole biosynthetic pathway for porphyrin distribution in the barley mutant albostrians. *Plant J.* **2003**, *35*, 512–522. [[CrossRef](#)] [[PubMed](#)]
128. Feierabend, J.; Mikus, M. Occurrence of a High Temperature Sensitivity of Chloroplast Ribosome Formation in Several Higher Plants. *Plant Physiol.* **1977**, *59*, 863–867. [[CrossRef](#)]
129. Woodson, J.D.; Perez-Ruiz, J.M.; Chory, J. Heme synthesis by plastid ferrochelatase i regulates nuclear gene expression in plants. *Curr. Biol.* **2011**, *21*, 897–903. [[CrossRef](#)]
130. Li, M.; Hensel, G.; Mascher, M.; Melzer, M.; Budhagatapalli, N.; Rutten, T.; Himmelbach, A.; Beier, S.; Korzun, V.; Kumlehn, J.; et al. Leaf variegation and impaired chloroplast development caused by a truncated CCT domain gene in albostrians barley. *Plant Cell* **2019**, *31*, 1430–1445. [[CrossRef](#)]
131. Kim, S.; Dale, B.E. Global potential bioethanol production from wasted crops and crop residues. *Biomass Bioenergy* **2004**, *26*, 361–375. [[CrossRef](#)]
132. Gu, J.; Zhou, Z.; Li, Z.; Chen, Y.; Wang, Z.; Zhang, H.; Yang, J. Photosynthetic properties and potentials for improvement of photosynthesis in pale green leaf rice under high light conditions. *Front. Plant Sci.* **2017**, *8*, 1082. [[CrossRef](#)]
133. Chida, H.; Nakazawa, A.; Akazaki, H.; Hirano, T.; Suruga, K.; Ogawa, M.; Satoh, T.; Kadokura, K.; Yamada, S.; Hakamata, W.; et al. Expression of the algal cytochrome c6 gene in Arabidopsis enhances photosynthesis and growth. *Plant Cell Physiol.* **2007**, *48*, 948–957. [[CrossRef](#)]
134. Yadav, S.K.; Khatri, K.; Rathore, M.S.; Jha, B. Introgression of UfCyt c 6, a thylakoid lumen protein from a green seaweed *Ulva fasciata* Delile enhanced photosynthesis and growth in tobacco. *Mol. Biol. Rep.* **2018**, *45*, 1745–1758. [[CrossRef](#)] [[PubMed](#)]



135. Pesaresi, P.; Hertle, A.; Pribil, M.; Kleine, T.; Wagner, R.; Strissel, H.; Lhnatowicz, A.; Bonardi, V.; Scharfenberg, M.; Schneider, A.; et al. Arabidopsis STN7 kinase provides a link between short- and long-term photosynthetic acclimation. *Plant Cell* **2009**, *21*, 2402–2423. [[CrossRef](#)]
136. Simkin, A.J.; McAusland, L.; Lawson, T.; Raines, C.A. Overexpression of the rieskeFeS protein increases electron transport rates and biomass yield. *Plant Physiol.* **2017**, *175*, 134–145. [[CrossRef](#)]
137. Kromdijk, J.; Glowacka, K.; Leonelli, L.; Gabilly, S.T.; Iwai, M.; Niyogi, K.K.; Long, S.P. Improving photosynthesis and crop productivity by accelerating recovery from photoprotection. *Science* **2016**, *354*, 857–861. [[CrossRef](#)] [[PubMed](#)]
138. Simkin, A.J.; McAusland, L.; Headland, L.R.; Lawson, T.; Raines, C.A. Multigene manipulation of photosynthetic carbon assimilation increases CO<sub>2</sub> fixation and biomass yield in tobacco. *J. Exp. Bot.* **2015**, *66*, 4075–4090. [[CrossRef](#)] [[PubMed](#)]
139. Simkin, A.J.; Lopez-Calcagno, P.E.; Davey, P.A.; Headland, L.R.; Lawson, T.; Timm, S.; Bauwe, H.; Raines, C.A. Simultaneous stimulation of sedoheptulose 1,7-bisphosphatase, fructose 1,6-bisphosphate aldolase and the photorespiratory glycine decarboxylase-H protein increases CO<sub>2</sub> assimilation, vegetative biomass and seed yield in Arabidopsis. *Plant Biotechnol. J.* **2017**, *15*, 805–816. [[CrossRef](#)]
140. Lefebvre, S.; Lawson, T.; Zakhleniuk, O.V.; Lloyd, J.C.; Raines, C.A. Increased sedoheptulose-1,7-bisphosphatase activity in transgenic tobacco plants stimulates photosynthesis and growth from an early stage in development. *Plant Physiol.* **2005**, *138*, 451–460. [[CrossRef](#)]
141. Ding, F.; Wang, M.; Zhang, S.; Ai, X. Changes in SBPase activity influence photosynthetic capacity, growth, and tolerance to chilling stress in transgenic tomato plants. *Sci. Rep.* **2016**, *6*, 32741. [[CrossRef](#)] [[PubMed](#)]
142. Uematsu, K.; Suzuki, N.; Iwamae, T.; Inui, M.; Yukawa, H. Increased fructose 1,6-bisphosphate aldolase in plastids enhances growth and photosynthesis of tobacco plants. *J. Exp. Bot.* **2012**, *63*, 3001–3009. [[CrossRef](#)]
143. Yamori, W.; Masumoto, C.; Fukayama, H.; Makino, A. Rubisco activase is a key regulator of non-steady-state photosynthesis at any leaf temperature and, to a lesser extent, of steady-state photosynthesis at high temperature. *Plant J.* **2012**, *71*, 871–880. [[CrossRef](#)]
144. Sanz-Barrio, R.; Corral-Martinez, P.; Ancin, M.; Segui-Simarro, J.M.; Farran, I. Overexpression of plastidial thioredoxin f leads to enhanced starch accumulation in tobacco leaves. *Plant Biotechnol. J.* **2013**, *11*, 618–627. [[CrossRef](#)]
145. Farran, I.; Fernandez-San Millan, A.; Ancin, M.; Larraya, L.; Veramendi, J. Increased bioethanol production from commercial tobacco cultivars overexpressing thioredoxin f grown under field conditions. *Mol. Breed.* **2014**, *34*, 457–469. [[CrossRef](#)]
146. Toivola, J.; Nikkanen, L.; Dahlström, K.M.; Salminen, T.A.; Lepistö, A.; Vignols, F.; Rintamäki, E. Overexpression of chloroplast NADPH-dependent thioredoxin reductase in Arabidopsis enhances leaf growth and elucidates in vivo function of reductase and thioredoxin domains. *Front. Plant Sci.* **2013**, *4*, 389. [[CrossRef](#)] [[PubMed](#)]
147. Nikkanen, L.; Toivola, J.; Rintamäki, E. Crosstalk between chloroplast thioredoxin systems in regulation of photosynthesis. *Plant Cell Environ.* **2016**, *39*, 1691–1705. [[CrossRef](#)] [[PubMed](#)]
148. Timm, S.; Florian, A.; Arrivault, S.; Stitt, M.; Fernie, A.R.; Bauwe, H. Glycine decarboxylase controls photosynthesis and plant growth. *FEBS Lett.* **2012**, *586*, 3692–3697. [[CrossRef](#)] [[PubMed](#)]
149. Timm, S.; Wittmisch, M.; Gamlien, S.; Ewald, R.; Florian, A.; Frank, M.; Wirtz, M.; Hell, R.; Fernie, A.R.; Bauwe, H. Mitochondrial dihydrolipoyl dehydrogenase activity shapes photosynthesis and photorespiration of Arabidopsis Thaliana. *Plant Cell* **2015**, *27*, 1968–1984. [[CrossRef](#)] [[PubMed](#)]
150. López-Calcagno, P.E.; Fisk, S.; Brown, K.L.; Bull, S.E.; South, P.F.; Raines, C.A. Overexpressing the H-protein of the glycine cleavage system increases biomass yield in glasshouse and field-grown transgenic tobacco plants. *Plant Biotechnol. J.* **2019**, *7*, 141–151. [[CrossRef](#)]
151. Kebeish, R.; Niessen, M.; Thiruveedhi, K.; Bari, R.; Hirsch, H.J.; Rosenkranz, R.; Stäbler, N.; Schönfeld, B.; Kreuzaler, F.; Peterhänsel, C. Chloroplastic photorespiratory bypass increases photosynthesis and biomass production in Arabidopsis thaliana. *Nat. Biotechnol.* **2007**, *25*, 593–599. [[CrossRef](#)]
152. South, P.F.; Cavanagh, A.P.; Liu, H.W.; Ort, D.R. Synthetic glycolate metabolism pathways stimulate crop growth and productivity in the field. *Science* **2019**, *367*, 45. [[CrossRef](#)]
153. Jansson, C.; Wullschleger, S.D.; Kalluri, U.C.; Tuskan, G.A. Phytosequestration: Carbon Biosequestration by Plants and the Prospects of Genetic Engineering. *Bioscience* **2010**, *60*, 685–696. [[CrossRef](#)]



154. Zhu, X.-G.; Long, S.P.; Ort, D.R. Improving Photosynthetic Efficiency for Greater Yield. *Annu. Rev. Plant Biol.* **2010**, *61*, 235–261. [[CrossRef](#)] [[PubMed](#)]
155. Blankenship, R.E.; Chen, M. Spectral expansion and antenna reduction can enhance photosynthesis for energy production. *Curr. Opin. Chem. Biol.* **2013**, *17*, 457–461. [[CrossRef](#)]
156. Nakajima, Y.; Ueda, R. Improvement of photosynthesis in dense microalgal suspension by reduction of light harvesting pigments. *J. Appl. Phycol.* **1997**, *9*, 503–510. [[CrossRef](#)]
157. Beckmann, J.; Lehr, F.; Finazzi, G.; Hankamer, B.; Posten, C.; Wobbe, L.; Kruse, O. Improvement of light to biomass conversion by de-regulation of light-harvesting protein translation in *Chlamydomonas reinhardtii*. *J. Biotechnol.* **2009**, *142*, 70–77. [[CrossRef](#)] [[PubMed](#)]
158. Perrine, Z.; Negi, S.; Sayre, R.T. Optimization of photosynthetic light energy utilization by microalgae. *Algal Res.* **2012**, *1*, 134–142. [[CrossRef](#)]
159. Demmig-Adams, B.; Adams, W.W. Chlorophyll and carotenoid composition in leaves of *Euonymus kiautschovicus* acclimated to different degrees of light stress in the field. *Aust. J. Plant Physiol.* **1996**, *23*, 649–659. [[CrossRef](#)]
160. Li, X.P.; Müller-Moulé, P.; Gilmore, A.M.; Niyogi, K.K. PsbS-dependent enhancement of feedback de-excitation protects photosystem II from photoinhibition. *Proc. Natl. Acad. Sci. USA* **2002**, *99*, 15222–15227. [[CrossRef](#)]
161. Pérez-Bueno, M.L.; Johnson, M.P.; Zia, A.; Ruban, A.V.; Horton, P. The Lhcb protein and xanthophyll composition of the light harvesting antenna controls the  $\Delta$ pH-dependency of non-photochemical quenching in *Arabidopsis thaliana*. *FEBS Lett.* **2008**, *582*, 1477–1482. [[CrossRef](#)]
162. Driever, S.M.; Simkin, A.J.; Alotaibi, S.; Fisk, S.J.; Madgwick, P.J.; Sparks, C.A.; Jones, H.D.; Lawson, T.; Parry, M.A.J.; Raines, C.A. Increased sbpase activity improves photosynthesis and grain yield in wheat grown in greenhouse conditions. *Philos. Trans. R. Soc. B Biol. Sci.* **2017**, *372*, 20160384. [[CrossRef](#)]
163. Kim, M.R.; Khaleda, L.; Jung, I.J.; Kim, J.Y.; Lee, S.Y.; Cha, J.Y.; Kim, W.Y. Overexpression of chloroplast-localized NADPH-dependent thioredoxin reductase C (NTRC) enhances tolerance to photo-oxidative and drought stresses in *Arabidopsis thaliana*. *J. Plant Biol.* **2017**, *60*, 175–180. [[CrossRef](#)]
164. Kisaki, T.; Tolbert, N.E. Glycine as a substrate for photorespiration. *Plant Cell Physiol.* **1970**, *11*, 247–258. [[CrossRef](#)]
165. Kisaki, T.; Imai, A.; Tolbert, N.E. Intracellular localization of enzymes related to photorespiration in green leaves. *Plant Cell Physiol.* **1971**, *12*, 267–273. [[CrossRef](#)]
166. Eisenhut, M.; Ruth, W.; Haimovich, M.; Bauwe, H.; Kaplan, A.; Hagemann, M. The photorespiratory glycolate metabolism is essential for cyanobacteria and might have been conveyed endosymbiotically to plants. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 17199–17204. [[CrossRef](#)] [[PubMed](#)]
167. Kikuchi, G.; Motokawa, Y.; Yoshida, T.; Hiraga, K. Glycine cleavage system: Reaction mechanism, physiological significance, and hyperglycinemia. *Proc. Japan Acad. Ser. B Phys. Biol. Sci.* **2008**, *84*, 246–263. [[CrossRef](#)] [[PubMed](#)]
168. Zelitch, I.; Schultes, N.P.; Peterson, R.B.; Brown, P.; Brutnell, T.P. High glycolate oxidase activity is required for survival of maize in normal air. *Plant Physiol.* **2009**, *149*, 195–204. [[CrossRef](#)]
169. Hackenberg, C.; Kern, R.; Hüge, J.; Stal, L.J.; Tsuji, Y.; Kopka, J.; Shiraiwa, Y.; Bauwe, H.; Hagemann, M. Cyanobacterial lactate oxidases serve as essential partners in N<sub>2</sub> fixation and evolved into photorespiratory glycolate oxidases in plants. *Plant Cell* **2011**, *23*, 2978–2990. [[CrossRef](#)]

