



Editorial Basic and Translational Models of Cooperative Oncogenesis

Helena E. Richardson ^{1,*}, Julia B. Cordero ² and Daniela Grifoni ³

- ¹ Department of Biochemistry and Genetics, La Trobe Institute for Molecular Science, La Trobe University, Melbourne, VIC 3086, Australia
- ² Wolfson Wohl Cancer Research Centre, Institute of Cancer Science, University of Glasgow, Glasgow G61 1QH, UK; julia.cordero@glasgow.ac.uk
- ³ Department of Life, Health and Environmental Sciences, University of L'Aquila, I-67100 L'Aquila, Italy; daniela.grifoni@univaq.it
- * Correspondence: H.Richardson@latrobe.edu.au

Received: 12 August 2020; Accepted: 14 August 2020; Published: 18 August 2020



Cancer is a complex set of diseases involving genetic or epigenetic changes within cells, as well as interactions between the developing tumour cells and their microenvironment, which leads to uncontrolled tumour growth, altered differentiation, local invasion and metastasis to distant sites [1,2]. Molecular changes in oncogenes or tumour suppressor genes promote various cancer hallmarks, including the continued proliferation, inhibition of differentiation, inhibition of apoptosis, changes to metabolism, evasion of the immune system and the promotion of invasion/metastasis [3,4]. Although many oncogenes and tumour suppressor gene mutations promote more than one hallmark of cancer, several mutations are required to generate malignant cancers, a process referred to as cooperative oncogenesis/tumourigenesis [5,6]. Through decades of research, we have gained much knowledge on key molecular events and processes involved in the formation of cancer, and much of this knowledge has stemmed from investigations using model organisms, such as the mouse and the vinegar fly, Drosophila melanogaster, in addition to in vitro cell line studies. In this Special Issue, we present a collection of original research papers on various aspects of cancer research utilising human cell lines [7,8], or in vivo using *Drosophila* as a model system [9,10], as well as reviews highlighting the Drosophila model organism in cancer research [11–15]. Drosophila is a particularly useful model organism for the study of cancer mechanisms, because it has a rapid life cycle and is genetically manipulatable and since cancer genes and signalling pathways are highly conserved between humans and *Drosophila*, and interactions between tumour cells and surrounding normal cells can be readily examined in *Drosophila* tissues [6,16–20].

In a research paper pertaining to in vitro models of cancer, Di Giorgio et al. [8] analyse the transcriptional response to the expression of three key oncogenes (*RAS*, *MYC*, and *HDAC4*) in human fibroblasts, revealing common signalling pathways that are deregulated by these genes, and suggesting potential therapeutic avenues for the treatment of cancers driven by these oncogenes. In the second research paper on this topic, Mayer et al. [7] focus on human pancreatic cancer, where they observe in tissue sections the infiltration of Th17-like T cells expressing IL21 and IL26, and the expression of receptors for IL21 and IL26 in the pancreatic epithelial cells. They show in human pancreatic cell lines that IL21 and IL26 signal through ERK1/2 and STAT, which leads to increased tumour cell growth in colony forming assays.

In a research paper utilising the *Drosophila* model, Parniewska and Stocker [9] identify the novel splicing factor, SF2, which is essential for the survival and hyperproliferation of tissues that upregulate the target of rapamycin complex 1 (TORC1), a protein kinase involved in cellular growth that is upregulated in many cancers [21,22]. The identification of SF2 as a key conserved target of TORC1,

2 of 4

which is required for early tumour growth in *Drosophila*, provides a potential new approach to develop anti-cancer therapies for tumours with upregulated TORC1 activity, such as those carrying loss of function of *PTEN* (phosphatase and tensin homolog deleted on chromosome ten) or constitutively active mutations in *PI3K* (phospho inositol 3 kinase) [23–25]. In a second research paper utilising the *Drosophila* model, Hamaratoglu and Atkins [10] undertook an analysis of published transcriptional data from various *Drosophila* imaginal disc epithelial cell models of cancer, and found that the JNK stress response pathway [26,27], and JAK/STAT [28,29], Hippo [30,31] and Notch [32,33] tissue growth signalling pathways are commonly deregulated. This important meta-analysis has opened-up new potential avenues of research to examine the cooperative interactions between these signalling pathways using model organisms, as well as to assess the co-dependency of these conserved pathways in human cancers.

The reviews in this Special Issue highlight the power of using *Drosophila* as an in vivo model system to study various aspects of cancer research, from its application in the study of the function of specific genes/pathways in cancer [11,15], to understanding particular cellular processes in cancer [13,14], and for functional analyses of cancer Omics data [12]. Sechi et al. [15] review the mechanisms of the Golgi phosphoprotein 3 (GOLPH3) oncogene in cancer, covering research on human cancer samples, in vitro cell line analyses and Drosophila in vivo analyses. Carmena [11] reviews the involvement of the Scribble cell polarity module [34] in asymmetric cell division (ACD) of Drosophila neural stem cells in tumourigenesis, highlighting the importance of correct ACD for appropriate differentiation and exit from the cell cycle. Casas-Tintó and Portela [14] review the involvement of specialised cellular extensions, termed cytonemes, in cell-cell communication and in tumourigenesis. Cytonemes have recently been shown to be highly important in the development of glioblastoma and the associated neural degeneration that occurs in *Drosophila* models and also in the human disease [35], and there is accumulating evidence for their role in tumourigenesis in other cancer types. Newman and Gregory [13] review the connection between chromosomal aberrations (aneuploidy) and metabolic changes, highlighting the role of oxidative stress and particularly reactive oxygen species (ROS) in this process. Finally, Bangi [12] reviews how *Drosophila* can be utilised to functionally analyse the vast amount of human cancer Omics data that is currently being generated, in order to validate key genes/pathways that contribute to cancer, to build new models to interrogate cancer mechanisms, and to screen for novel cancer therapeutics. Drosophila has already proven its worth in identifying novel drugs that target particular types of human cancers, such as multiple endocrine neoplasia type 2 (MEN2), colorectal and non-small cell lung cancers [36–42], and undoubtably, the development of more sophisticated Drosophila models that incorporate additional genetic lesions will enable better modelling of human cancers and new anti-cancer drug discovery.

This first iteration of this Special Issue on basic and translational models of cooperative oncogenesis presents only a snapshot of the vast amount of research into cancer currently being conducted worldwide, yet it highlights the important contribution of the simple multicellular model organism, *Drosophila*, to our current understanding of cancer. Undoubtably, further primary research papers and literature reviews for future iterations of this Special Issue will highlight new cancer genes/pathways and processes involved in cancer, additional in vivo models of cancer (such as worms, zebra fish, and mice), and novel approaches for the understanding of cancer mechanisms and for developing new cancer therapies.

Funding: This research received no external funding.

Conflicts of Interest: The authors declare no conflict of interest.

References

- Radisky, D.; Hagios, C.; Bissell, M.J. Tumors are unique organs defined by abnormal signaling and context. Semin. Cancer Biol. 2001, 11, 87–95. [CrossRef] [PubMed]
- 2. Bissell, M.J.; Radisky, D. Putting tumours in context. Nat. Rev. Cancer 2001, 1, 46–54. [CrossRef] [PubMed]

- 3. Hanahan, D.; Weinberg, R.A. The hallmarks of cancer. Cell 2000, 100, 57–70. [CrossRef]
- 4. Hanahan, D.; Weinberg, R.A. Hallmarks of cancer: The next generation. Cell 2011, 144, 646–674. [CrossRef]
- 5. Delaval, B.; Birnbaum, D. A cell cycle hypothesis of cooperative oncogenesis (Review). *Int. J. Oncol.* 2007, *30*, 1051–1058. [CrossRef]
- Richardson, H.E.; Portela, M. Modelling Cooperative Tumorigenesis in Drosophila. *Biomed. Res. Int.* 2018, 2018, 4258387. [CrossRef]
- Mayer, P.; Linnebacher, A.; Glennemeier-Marke, H.; Marnet, N.; Bergmann, F.; Hackert, T.; Klauss, M.; Poth, T.; Gaida, M.M. The Microarchitecture of Pancreatic Cancer as Measured by Diffusion-Weighted Magnetic Resonance Imaging is Altered by T Cells with a Tumor Promoting Th17 Phenotype. *Int. J. Mol. Sci.* 2020, 21, 346. [CrossRef]
- 8. Di Giorgio, E.; Paluvai, H.; Picco, R.; Brancolini, C. Genetic Programs Driving Oncogenic Transformation: Lessons from in Vitro Models. *Int. J. Mol. Sci.* **2020**, *20*, 6283. [CrossRef]
- 9. Parniewska, M.M.; Stocker, H. The Splicing Factor SF2 is Critical for Hyperproliferation and Survival in a TORC1-Dependent Model of Early Tumorigenesis in Drosophila. *Int. J. Mol. Sci.* **2019**, *21*, 4465. [CrossRef]
- 10. Hamaratoglu, F.; Atkins, M. Rounding up the Usual Suspects: Assessing Yorkie, AP-1, and Stat Coactivation in Tumorigenesis. *Int. J. Mol. Sci.* **2020**, *21*, 4580. [CrossRef]
- 11. Carmena, A. The Case of the Scribble Polarity Module in Asymmetric Neuroblast Division in Development and Tumorigenesis. *Int. J. Mol. Sci.* **2020**, *21*, 2865. [CrossRef]
- 12. Bangi, E. Strategies for Functional Interrogation of Big Cancer Data Using Drosophila Cancer Models. *Int. J. Mol. Sci.* **2020**, *21*, 3754. [CrossRef]
- 13. Newman, D.L.; Gregory, S.L. Co-Operation between Aneuploidy and Metabolic Changes in Driving Tumorigenesis. *Int. J. Mol. Sci.* 2020, 20, 4611. [CrossRef]
- 14. Casas-Tintó, S.; Portela, M. Cytonemes, Their Formation, Regulation, and Roles in Signaling and Communication in Tumorigenesis. *Int. J. Mol. Sci.* **2019**, *20*, 5641. [CrossRef]
- 15. Sechi, S.; Frappaolo, A.; Karimpour-Ghahnavieh, A.; Piergentili, R.; Giansanti, M.G. Oncogenic Roles of GOLPH3 in the Physiopathology of Cancer. *Int. J. Mol. Sci.* **2019**, *21*, 933. [CrossRef]
- 16. Rudrapatna, V.A.; Cagan, R.L.; Das, T.K. Drosophila cancer models. Dev. Dyn. 2012, 241, 107–118. [CrossRef]
- 17. Sonoshita, M.; Cagan, R.L. Modeling Human Cancers in Drosophila. Curr. Top. Dev. Biol. 2017, 121, 287–309.
- 18. Gonzalez, C. Drosophila melanogaster: A model and a tool to investigate malignancy and identify new therapeutics. *Nat. Rev. Cancer* **2013**, *13*, 172–183. [CrossRef]
- 19. Tipping, M.; Perrimon, N. Drosophila as a model for context-dependent tumorigenesis. *J. Cell. Physiol.* **2014**, 229, 27–33.
- 20. Brumby, A.M.; Richardson, H.E. Using Drosophila melanogaster to map human cancer pathways. *Nat. Rev. Cancer* **2005**, *5*, 626–639. [CrossRef]
- 21. Zou, Z.; Tao, T.; Li, H.; Zhu, X. mTOR signaling pathway and mTOR inhibitors in cancer: Progress and challenges. *Cell Biosci.* **2020**, *10*, 31. [CrossRef]
- 22. Liu, G.Y.; Sabatini, D.M. mTOR at the nexus of nutrition, growth, ageing and disease. *Nat. Rev. Mol. Cell Biol.* **2020**, *21*, 183–203. [CrossRef]
- 23. Parsons, R. Discovery of the PTEN Tumor Suppressor and Its Connection to the PI3K and AKT Oncogenes. *Cold Spring Harb. Perspect. Med.* **2020**, *10*, a036129. [CrossRef]
- 24. Ngeow, J.; Eng, C. PTEN in Hereditary and Sporadic Cancer. *Cold Spring Harb. Perspect. Med.* 2020, 10, a036087. [CrossRef]
- 25. Noorolyai, S.; Shajari, N.; Baghbani, E.; Sadreddini, S.; Baradaran, B. The relation between PI3K/AKT signalling pathway and cancer. *Gene* **2019**, *698*, 120–128. [CrossRef]
- 26. La Marca, J.E.; Richardson, H.E. Two-Faced: Roles of JNK Signalling During Tumourigenesis in the Drosophila Model. *Front. Cell Dev. Biol.* **2020**, *8*, 42. [CrossRef]
- 27. Wu, Q.; Wu, W.; Fu, B.; Shi, L.; Wang, X.; Kuca, K. JNK signaling in cancer cell survival. *Med. Res. Rev.* 2019, 39, 2082–2104. [CrossRef]
- 28. Trivedi, S.; Starz-Gaiano, M. Drosophila Jak/STAT Signaling: Regulation and Relevance in Human Cancer and Metastasis. *Int. J. Mol. Sci.* **2018**, *19*, 4056. [CrossRef]
- 29. Jin, W. Role of JAK/STAT3 Signaling in the Regulation of Metastasis, the Transition of Cancer Stem Cells, and Chemoresistance of Cancer by Epithelial-Mesenchymal Transition. *Cells* **2020**, *9*, 217. [CrossRef]

- 30. Kulkarni, A.; Chang, M.T.; Vissers, J.H.A.; Dey, A.; Harvey, K.F. The Hippo Pathway as a Driver of Select Human Cancers. *Trends Cancer* **2020**. [CrossRef]
- 31. Zygulska, A.L.; Krzemieniecki, K.; Pierzchalski, P. Hippo pathway—Brief overview of its relevance in cancer. *J. Physiol. Pharmacol.* **2017**, *68*, 311–335.
- 32. McIntyre, B.; Asahara, T.; Alev, C. Overview of Basic Mechanisms of Notch Signaling in Development and Disease. *Adv. Exp. Med. Biol.* **2020**, 1227, 9–27.
- 33. Previs, R.A.; Coleman, R.L.; Harris, A.L.; Sood, A.K. Molecular pathways: Translational and therapeutic implications of the Notch signaling pathway in cancer. *Clin. Cancer Res.* **2015**, *21*, 955–961. [CrossRef]
- Stephens, R.; Lim, K.; Portela, M.; Kvansakul, M.; Humbert, P.O.; Richardson, H.E. The Scribble Cell Polarity Module in the Regulation of Cell Signaling in Tissue Development and Tumorigenesis. *J. Mol. Biol.* 2018, 430, 3585–3612. [CrossRef]
- 35. Portela, M.; Venkataramani, V.; Fahey-Lozano, N.; Seco, E.; Losada-Perez, M.; Winkler, F.; Casas-Tinto, S. Glioblastoma cells vampirize WNT from neurons and trigger a JNK/MMP signaling loop that enhances glioblastoma progression and neurodegeneration. *PLoS Biol.* **2019**, *17*, e3000545. [CrossRef]
- 36. Bangi, E.; Ang, C.; Smibert, P.; Uzilov, A.V.; Teague, A.G.; Antipin, Y.; Chen, R.; Hecht, C.; Gruszczynski, N.; Yon, W.J.; et al. A personalized platform identifies trametinib plus zoledronate for a patient with KRAS-mutant metastatic colorectal cancer. *Sci. Adv.* **2019**, *5*, eaav6528. [CrossRef]
- Sonoshita, M.; Scopton, A.P.; Ung, P.M.U.; Murray, M.A.; Silber, L.; Maldonado, A.Y.; Real, A.; Schlessinger, A.; Cagan, R.L.; Dar, A.C. A whole-animal platform to advance a clinical kinase inhibitor into new disease space. *Nat. Chem. Biol.* 2018, 14, 291–298. [CrossRef]
- Das, T.K.; Esernio, J.; Cagan, R.L. Restraining Network Response to Targeted Cancer Therapies Improves Efficacy and Reduces Cellular Resistance. *Cancer Res.* 2018, 78, 4344–4359. [CrossRef]
- Das, T.K.; Cagan, R.L. Non-mammalian models of multiple endocrine neoplasia type 2. *Endocr. Relat. Cancer* 2018, 25, T91–T104. [CrossRef]
- 40. Levine, B.D.; Cagan, R.L. Drosophila Lung Cancer Models Identify Trametinib plus Statin as Candidate Therapeutic. *Cell Rep.* **2016**, *14*, 1477–1487. [CrossRef]
- 41. Bangi, E.; Murgia, C.; Teague, A.G.; Sansom, O.J.; Cagan, R.L. Functional exploration of colorectal cancer genomes using Drosophila. *Nat. Commun.* **2016**, *7*, 13615. [CrossRef] [PubMed]
- 42. Dar, A.C.; Das, T.K.; Shokat, K.M.; Cagan, R.L. Chemical genetic discovery of targets and anti-targets for cancer polypharmacology. *Nature* **2012**, *486*, 80–84. [CrossRef] [PubMed]



© 2020 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).