



EuroBlight tracking of evolving European populations of *Phytophthora infestans* as an aid to late blight management

David Cooke

Jens G Hansen, Poul Lassen, Alison Lees, Geert Kessel+ many partners

Whatman® Lot No. FT6196205

3rd AsiaBlight International Meeting, Beijing 25-27 Oct 2019



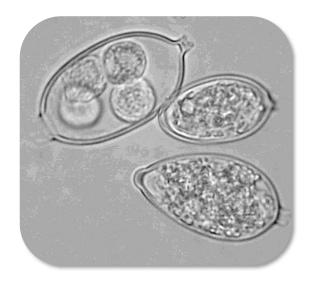






Objectives

- How EuroBlight database benefits growers/industry?
- How it works mechanisms?
- Share our experiences
- Continue to support AsiaBlight
- Learn from AsiaBlight network experiences
- Key to network success
 - TRUST
 - CO-OPERATION
 - COMMITMENT
 - STANDARDISATION
 - MUTUAL GAIN



Need to study population change



- Environmental response (RH and °C) DSS
- Fungicide insensitivity rapid evolution
- Fitness, Aggressiveness, Virulence



- Survival
- Rate of evolution



My links with research in Asia

- Ying Li 12-plex SSR development
- Sanjoy Guha Roy broad collaboration
- Buddhi Sharma Nepal SSR analysis
- Mei Guo China SSR analysis
- Kwangsoo Cho South Korea SSR analysis
- Louise Cooke and many collaborators FTA analysis
- Dolf DeBoer & Birte Komolong PNG & Australasia
- Mohammad Rashidul Islam Bangladesh SSR analysis
- Huub Schepers/Geert Kessel- WUR projects

European sponsors/contributors/collaborators

Commercial

ACVNPT * **BASF SE** Belchim Crop Prot. Certis Cheminova CropSolutions **CUConsulting Emsland Group** Germicopa SAS* HZPC Holland B.V. Neiker Nordisk Alkali PCA Profytodsd Staphyt Syngenta/ ChinaChem SynTech Research UPL

ADAMA Agrifirm Agriphar Bayer CropScience AG Corteva



+Many more global teams

AFBI

Agricultural Institute of Slovenia AHDB Potatoes and FAB Scouts Aarhus University* **ARVALIS-Institut du Végétal* Bar-Ilan University** Bayerische Landesanstalt für Landwirtschaft Centre Wallon de Recherches Agronomigues Estonian University of Life Sciences* INRA* Inst. Plant Protection & Environment, Serbia The James Hutton Institute* NIBIO, Norway* Wageningen University Plant Breeding & Acclimatization Inst. (IHAR) Swedish University of Agricultural Sciences TEAGASC * Partners in the IPMBlight2.0 project

Academic/state research





EucaBlight themes

- Tracking the potato blight pathogens in a global context Tools, network infrastructures and activities
- Stewardship of host resistance genes and active ingredients - develop and adopt innovative and sustainable control strategies on regional scales
- Education, advocacy and communication taking into account different scales and stakeholders



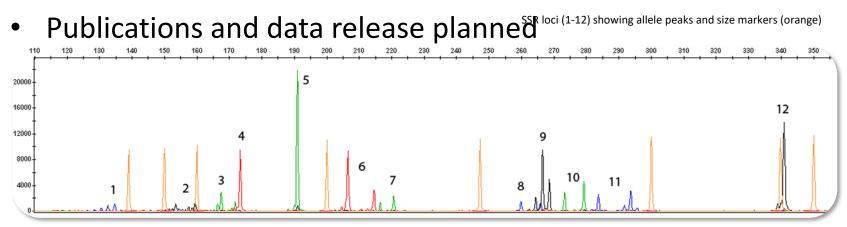


Monitoring methods





- Scouts monitoring crops provide samples
- Outbreak data recorded
- Active lesions sampled
- DNA fingerprinted using 12-plex SSRs (Li *et al.* 2013)
 - Hutton, INRA, IHAR, WUR strong co-operation
- Genotypes defined QC and to EuroBlight database
- Data mapped on <u>www.euroblight.net</u> + annual summary







- Scouts issued with sample forms and FTA cards
 - Costs €25 per sample (covers costs not profit-making)
 - Cards pre-labelled with organisation and year. e.g. Syngenta19001
 - Some groups collect isolates needed for phenotyping
- Outbreak data (e.g. location, crop type, cultivar) recorded
 - Sampling not prescriptive aim for broad geographical spread
 - AHDB GB 4-8 lesions per crop across outbreak
 - Companies sample efficacy trials at lower frequency
 - Avoid excess sampling in one location creates bias



Use of FTA cards



Protocol will be made available via web page



- Fresh, sporulating lesion onto FTA card
- Air dry at room temp. store & post
- 12 year old cards OK
- Handle carefully stop cross-contamination



Database

- Custom-built & managed by Jens Hansen and Poul Lassen (Aarhus University, Denmark)
- Synergy with their Wheat Rust database
- David Cooke population data manager
 - Co-ordinate FTA card provision
 - Collate cards at end of each season
 - Run SSR analysis (& receive other group's data)
 - Quality control data 2 stages
 - Step 1 Sample data
 - Step 2 SSR data
- Raw data never shared without owner permission



PLB Toolbox

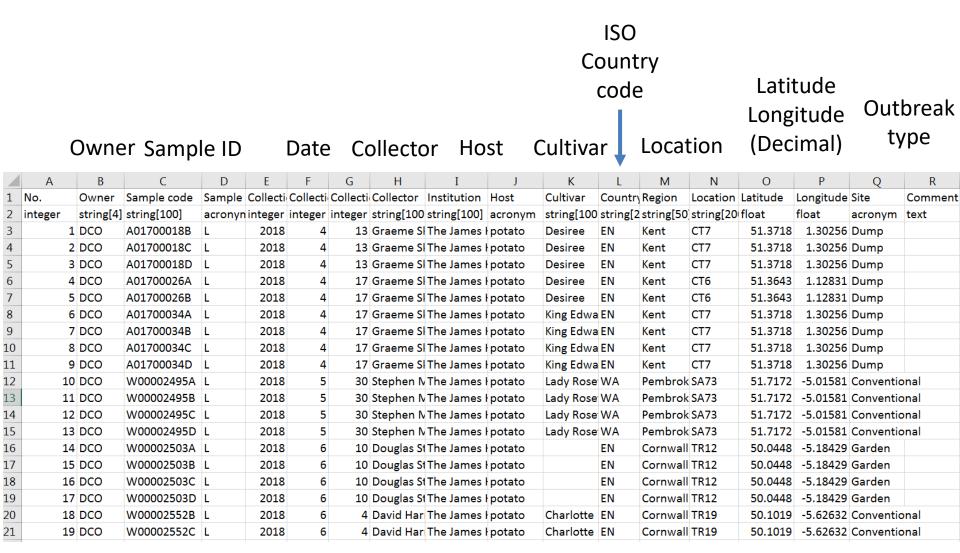
16K samples with genotype data in EU database

Рот	Potato Late Blight Toolbox																										
Home	Samples	Trials	Partne	ers																			M	/elcome	e David	Cooke	logout
SAMPLE	OVERVIEW																										
Data	Samples	s 🔍 Geno	types b	y SSR 🤇	Matin	g type	O Virule	ence 🔍	Withou	it result	s																
Continent	Europe			-								ia															
Country		Total	2019	2018	2017	2016	2015	2014	2013	2012	2011	2010	2009	2008	2007	2006	2005	2004	2003	2002	2001	2000	1999	1998	1997	1996	Older
Austria		106		2					4											100							
Belarus		45		1		5		29	10																		
Belgium		1571		98	74	169	97	187	75	193	58	72	35	24	54	26	13	16	15	32	105	97	82	49			
Bulgaria		17				2	3	6	6																		
Croatia		5		2				3																			
Cypres		1							1																		
Czech R	epublic	380		3	6	7		7	5							80	177	64	31								
Denmar	k	1136	228	121	174	96	64	37	126	114	87				23	1			65								
England		5967		115	397	363	97	403	188	451	61	101	306	590	901	801	56	98	47					336	630		26
Estonia		1026			24	107	1	68	54	49	51	41				103	103	144	84	89	83	13	12				
Finland		2981			6	7	5	4	4							87			234		210	675	457	538	602	16	136
France		4762		236	260	230	116	222	185				12	125	331	382	303	444	72	136	213	160	332	233	123	156	491
German	у	439		3	94	86	87	62	40				3		53				3	1		7					
Greece		44		3	3	6	14	4			14																
Hungary	/	309		12	3	4										56	67			93	27	3		36	5	1	2
Ireland		150		1				1								1										147	
Italy		43		5	4	2		2	13				6													10	1
Jersey		1								1																	
Latvia		183			1	5			52	44	48	33															
Lithuani		172			5	5		40	53	25	27	17															
Netherla	ands	2579		125	249	195	79	149	84				12		15		1	25	109			481				353	702

Data upload steps

Potato Late Blight Toolbox													
Home Samples Trials Partners			Welcor	me David Cooke <mark>logout</mark>									
GENOTYPES Unpublished Published for project partners New samples Change previous created sam CREATE SAMPLE INPUT TEMPLATE FILE Select data owner Alison Lees UPLOAD AND IMPORT SAMPLE INPUT FILE CSV sample file (file size limit is 1 MB)	nples New genotype results Cha	olished for all Genotype map Impo nge previous created genotype results Select year 2019 Select number of sam	Date format dd.mm.yyyy	File format Comma ▼									
Enter new Sample data (makes db key)	Change Sample data	Link in new SSR data	Change SSR data										

Sample data entry (csv)



Genotypic marker – <u>S</u>imple <u>S</u>equence <u>R</u>epeats



ABI 3730 capillary sequencer

Journal of Microbiological Methods 92 (2013) 316-322 Contents lists available at SciVerse ScienceDirect

5-52.20	
Eille	
ELSEVIER	

Journal of Microbiological Methods

journal homepage: www.elsevier.com/locate/jmicmeth

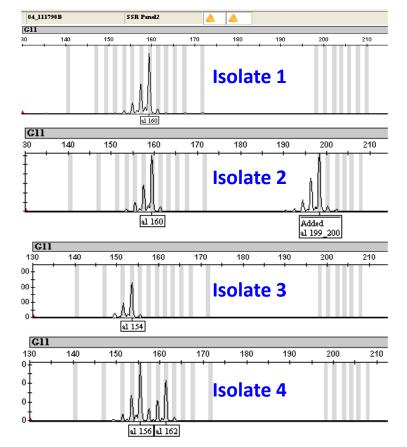
Efficient multiplex simple sequence repeat genotyping of the oomycete plant pathogen *Phytophthora infestans*



Ying Li ^{a,c,d}, David E.L. Cooke ^{b,*}, Evert Jacobsen ^c, Theo van der Lee ^{d,*}

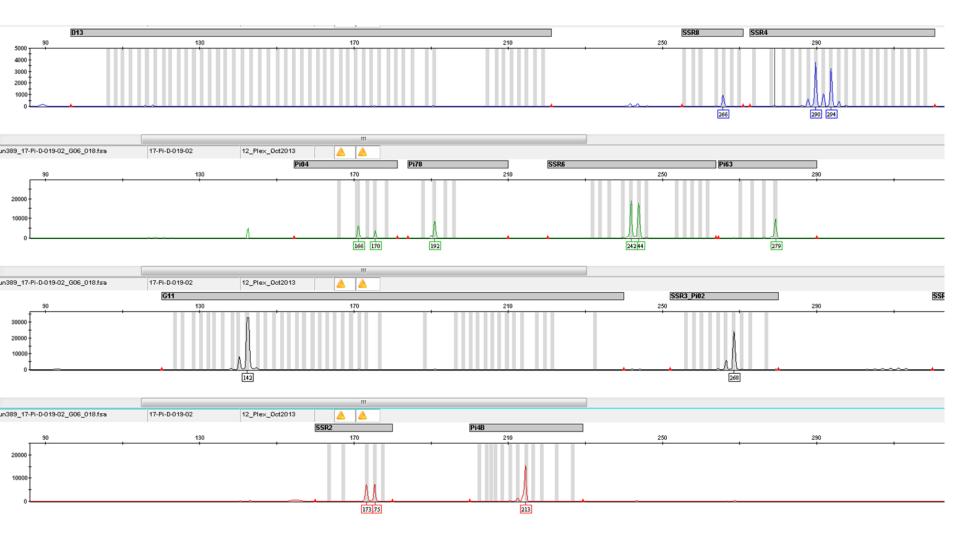
- * Institute of Vegetables and Howers, Chinese Academy of Agricultural Sciences, Beijing, 100081, China
- ^b The James Hutton Institute, Invergowrie, Dundee, DD2 SDA, UK
- ¹ Plant Breeding, Wageningen UR, P.O. Box 386, 6708 PB Wageningen, The Netherlands
- ^d Bio-interaction and Plant Health, Plant Research International B.V., Wageningen UR, P.O. Box 16, 6700 AA Wageningen, The Netherlands

Marker G11



SSR amplification





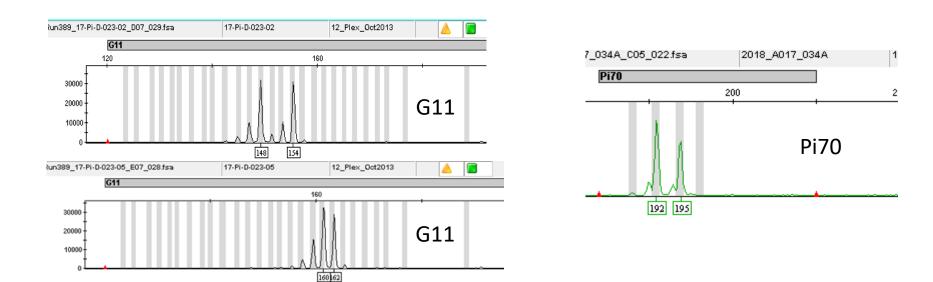
• 12 SSR loci amplified in single PCR





SSR scoring

- Key labs trained Hutton, WUR, INRA, IHAR
- 12 SSR loci (2 tri-& 10 dinucleotide rpts)
- Allele numbers per loci 4-40
- Allele name standardisation is critical bins help this sharing of bin profiles
- Allele-specific pattern of stutter peaks







SSR output

- Quality control very important
- Some variation within clones observed

						_																														
Sample Name	Country	Genotype	Allele 1	Allele 2	Allele 3	Allele 1	Allele 2	Allele 3	Allele 1	Allele 2	Allele 3	Allele 1	Allele 2	Allele 3	Allele 1	Allele 2	Allele 3	Allele 1	Allele 2	Allele 3	Allele 1	Allele 2	Allele 3	Allele 1	Allele 2	Allele 3	Allele 1A	llele 2Allel	e 3 Allele	1Allele	2Allele 3	Allele 1	Allele 2/	Allele 3 A	Allele 1Al	llele 2Alle
			Pi02	Pi02	Pi02	Pi4B	Pi4B	Pi4B	G11	G11	G11	Pi04	Pi04	Pi04	Pi63	Pi63	Pi63	Pi70	Pi70	Pi70	D13	D13	D13	SSR11	SSR11	SSR11	SSR2 S	SR2 SSF	R2 SSR	4 SSR4	4 SSR4	SSR6	SSR6	SSR6 S	3SR8 S	SR8SS
2019_W00003831E	GB	36_A2	268	268	0	205	213	217	150	154	160	166	170	0	270	279	0	192	195	0	136	152	156	341	341	0	173	175 0	288	294	0	240	242	244	260	266
2019_W00003831F	GB	36_A2	268	268	0	205	213	217	150	154	160	166	170	0	270	279	0	192	195	0	136	152	156	341	341	0	173	175 0	288	294	0	240	242	244	260	266
2019_W00003831G	GB	36_A2	268	268	0	205	213	217	150	154	160	166	170	0	270	279	0	192	195	0	136	152	156	341	341	0	173	175 0	288	294	0	240	242	244	260	266
2019_W00003831H		36_A2	268	268	0	205	213	217	150	154	160	166	170	0	270	279	0	192	195	0	136	152	156	341	341	0	173	175 0	288	294	0	240	242	244	260	266
2019_W00003872E	GB	13_A2	266	268	0	205	213	0	154	160	164	166	170	0	273	279	0	192	192	0	132	136	154	341	341	0	173	173 0	284	294	0	240	244	0	260	266
2019_W00003872F	GB	13_A2	266	268	0	205	213	0	154	160	164	166	170	0	273	279	0	192	192	0	132	136	154	341	341	0	173	173 0	284	294	0	240	244	0	260	266
2019_W00003872G	GB	13_A2	266	268	0	205	213	0	154	160	164	166	170	0	273	279	0	192	192	0	132	136	154	341	341	0	173	173 0	284	294	0	240	244	0	260	266
2019_W00003872H	GB	13_A2	266	268	0	205	213	0	154	160	164	166	170	0	273	279	0	192	192	0	132	136	154	341	341	0	173	173 0	284	294	0	240	244	0	260	266
2019_W00003930A	GB	36_A2	268	268	0	205	213	217	150	154	160	166	170	0	270	279	0	192	195	0	136	152	154	341	341	0	173	175 0	288	294	0	240	242	244	260	266
2019_W00003930C	GB	36_A2	268	268	0	205	213	217	150	154	160	166	170	0	270	279	0	192	195	0	136	152	154	341	341	0	173	175 0	288	294	0	240	242	244	260	266
2019_W00003930F	GB	36_A2	268	268	0	205	213	217	150	154	160	166	170	0	270	279	0	192	195	0	136	152	154	341	341	0	173	175 0	288	294	0	240	242	244	260	266
2019_W00003930H	GB	36_A2	268	268	0	205	213	217	150	154	160	166	170	0	270	279	0	192	195	0	136	152	154	341	341	0	173	175 0	288	294	0	240	242	244	260	266
2019_W00003948A	GB	6_A1	258	266	268	213	217	0		158	0	166	170	0	273	279	0	192	195	0	null	null	0	341	355	0	173	175 0	284	292	0	244	244	0	260	266
2019_W00003948B	GB	6_A1	258	266	268	213	217	0	158	158	0	166	170	0	273	279	0	192	195	0	null	null	0	341	355	0	173	175 0	284	292	296	244	244	0	260	266
AGRI1_1	DK	41_A2	266	268	0	213	213	0	156	206	0	168	168	0	279	279	0	192	192	0	136	136	0	341	341	0	173	175 0	284	288	294	242	244	0	260	266
AGRI1_2	DK	41_A2	266	268	0	213	213	0	156	206	0	168	168	0	279	279	0	192	192	0	136	136	0	341	341	0	173	175 0	284	288	294	242	244	0	264	266
AGRI3_1	DK	Other	266	268	0	213	217	0	148	154	0	166	170	0	273	279	0	192	192	0	null	null	0	341	341	0	173	175 0	284	294	0	242	244	0	260	266
AGRI3_2	DK	Other	266	268	0	213	217	0	148	154	0	166	170	0	273	279	0	192	192	0	null	null	0	341	341	0	173	175 0	284	294	0	242	244	0	260	266
AGRI6_1	DK	41_A2	266	268	0	213	213	0	156	206	0	166	166	0	279	279	0	192	192	0	136	136	0	341	341	0	173	175 0	284	288	294	242	244	0	264	266
AGRI6_2	DK	41_A2	266	268	0	213	213	0	156	206	0	168	168	0	279	279	0	192	192	0	136	136	0	341	341	0	173	175 0	284	288	294	242	244	0	260	266
AKV_2019_1_A	DK	Other	268	268	0	213	213	0	156	162	0	168	168	0	279	279	0	192	192	0	118	118	0	341	341	0	173	175 0	284	284	0	242	242	0	260 3	260
AKV_2019_2_A	DK	Other	268	268	0	213	213	0	154	154	0	168	168	0	270	279	0	192	192	0	null	null	0	341	341	0	173	173 0	284	294	0	242	242	0	266	266
AKV_2019_2_B	DK	Other	258	268	0	205	217	0	154	156	0	166	170	0	279	279	0	192	192	0	118	118	0	341	341	0	173	175 0	284	288	0	242	244	0	260	260
AKV 2019 3 A	DK	Other	258	268	0	205	217	0	154	156	0	166	170	0	279	279	0	192	192	0	118	118	0	341	341	0	173	175 0	284	288	0	242	244	0	260	260

SSR data entry

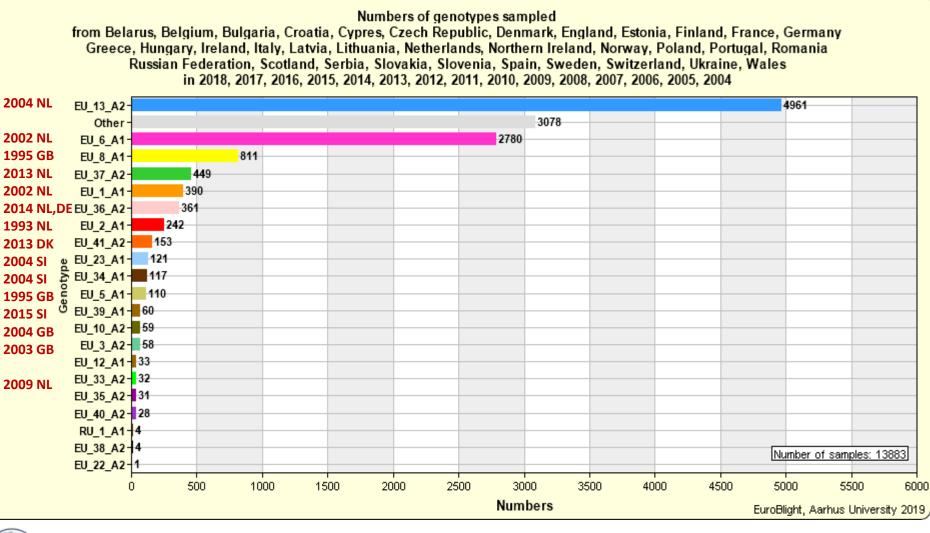
- Excel sheet generated by database
- Data added in standard format

	А	В	С	D	E	F	G	Н	Ι	J	K	L	Μ	Ν	0	Р	Q	R
1	Collection	SampleID	ResultID	ResultGer	SSRLocus	SampleCode	Owner	Year	Cultivar	Country	Location	Collection	Laborat	SampleFile	Genotype	Locus PiO2	Locus Pi02	Locus Pi02
2	NO EDIT	NO EDIT	NO EDIT	NO EDIT	NO EDIT	NO EDIT	NO EDI	NO EDI	NO EDIT	NO EDIT	NO EDIT	NO EDIT	acronyn	rstring[200	acronym	Allele 1	Allele 2	Allele 3
3	47197	1	1	1	2	A01700018B	DCO	2018	Desiree	EN	CT7	13.4.2018	JHI	Run380_2	EU_37_A2	268		
4	47197	2	1	1	2	A01700018C	DCO	2018	Desiree	EN	CT7	13.4.2018	JHI	Run380_2	EU_6_A1	258	266	268
5	47197	3	1	1	2	A01700018D	DCO	2018	Desiree	EN	CT7	13.4.2018	JHI	Run380_2	EU_6_A1	258	266	268
6	47198	1	1	1	2	A01700026A	DCO	2018	Desiree	EN	CT6	17.4.2018	JHI	Run380_2	EU_36_A2	268		
7	47198	2	1	1	2	A01700026B	DCO	2018	Desiree	EN	CT6	17.4.2018	JHI	Run380_2	EU_36_A2	268		
8	47199	1	1	1	2	A01700034A	DCO	2018	King Edwa	EN	CT7	17.4.2018	JHI	Run380_2	EU_36_A2	268		
9	47199	2	1	1	2	A01700034B	DCO	2018	King Edwa	EN	CT7	17.4.2018	JHI	Run380_2	EU_36_A2	268		
10	47199	3	1	1	2	A01700034C	DCO	2018	King Edwa	EN	CT7	17.4.2018	JHI	Run380_2	EU_36_A2	268		
11	47199	4	1	1	2	A01700034D	DCO	2018	King Edwa	EN	CT7	17.4.2018	JHI	Run380_2	EU_36_A2	268		
12	47212	1	1	1	2	A01700067B	DCO	2018	Desiree	EN	CT7	29.6.2018	JHI	Run384_A	EU_36_A2	268		
13	47212	2	1	1	2	A01700067C	DCO	2018	Desiree	EN	CT7	29.6.2018	JHI	Run384_A	EU_36_A2	268		
14	47212	3	1	1	2	A01700067D	DCO	2018	Desiree	EN	CT7	29.6.2018	JHI	Run384_A	EU_36_A2	268		
15	47200	1	1	1	2	W00002495A	DCO	2018	Lady Rose	WA	SA73	30.5.2018	JHI	Run383_2	EU_6_A1	258	266	268
16	47200	2	1	1	2	W00002495B	DCO	2018	Ladv Rose	WA	SA73	30.5.2018	.IHI	Run383 2	FU 6 A1	258	266	268

SSR management

Potat Home S	SSR Locus	GHT TOOLI Partners	вох		SSR Alleles			C	ienoty list	pe				w	/elcome Dav	id Coo	ke logout
GENOTYPES										Y							
Unpublishe	ed Publisł	ned for project part			for data suppliers	Published	for all		notype map	Import		ci, alleles and	d types \				
SSR Locus	_			SSR Allele					types	-			-				
Name	Repeat type		Active	No	Repeat number	2	ŵ	ID	Name	Pattern	Sort	Color	Image	Color old	-	Edit	Delete
Pi02	2	1		0 256	0 9	2		1	EU_1_A1		1	FF9900	•	FF9900	•	2	
Pi04	2	2	Image: A state of the state	258	10	2	<u></u>	7	EU_2_A1		2	FF0000	•	FF0000	•	2	
Pi16	2	3		260	11	2	Ē	13	EU_5_A1		3	CCCC66	•	CCCC66	•	2	Ē
Pi33	3	4	1	262	12	2	Ē	2	EU_6_A1		4	FF33CC	•	FF33CC	•	2	Ē
Pi56	2	5	Image: A start of the start	264	13	Z	Ē	8	EU_8_A1		5	FFFF00	•	FFFF00	•	2	
Pi63	3	6	1	266	14	Z		12	EU_12_A1		6	996600	•	996600	•	Z	
Pi70	3	7	1	268	15	2		3	EU_13_A2		7	3399FF	•	3399FF	٠	Z	Ē
Pi89	2	8	1	270	16	2		9	EU_23_A1		8	99CCFF	•	99CCFF	•	Z	1
Pi4B	2	9	1	272	17	2		4	EU_33_A2		9	00FF00		00FF00		2	Ē
D13	2	10	1	274	18	2		10	EU_34_A1		10	663300		663300	•	Z	
G11	2	11	1	276	19		ш	11	EU_35_A2		11	990099		990099		2	ŵ
SSR1	2	12	1					16	EU_36_A2		12	FFCCCC		FFCCCC		2	Ē
SSR2	2	13	1					19	EU_37_A2		13	33CC33		33CC33	•	2	
SSR4	2	14	1												•	2	
SSR6	2	15	1					14	KR_1_A1		15	00FF00	•	00FF00	•	2	
SSR7	2	16						15	SIB-1_A1		16	663300	•	663300	•		
SSR8	2	17	1					17	US_1_A1		17	990099	•	000001	•	2	Ť
SSR11	2	18	1					18	US_7_like		18	FFCCCC		FFCCCC		2	Ť
N = 18								20	EU_39_A1		20	996600	•	996600	•	Z	Ē
								21	EU_38_A2		21	000001	•	000001	•	Z	Ē
				N = 12				22	AR 1 A2		22	CCFFFF		9CEAF1		2	1 1 1

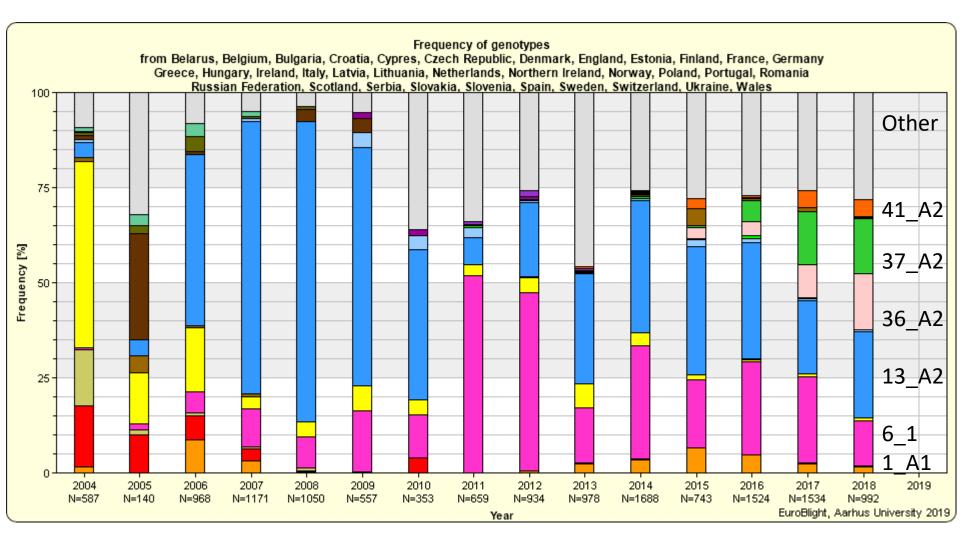
Introducing the clones 2004-2018







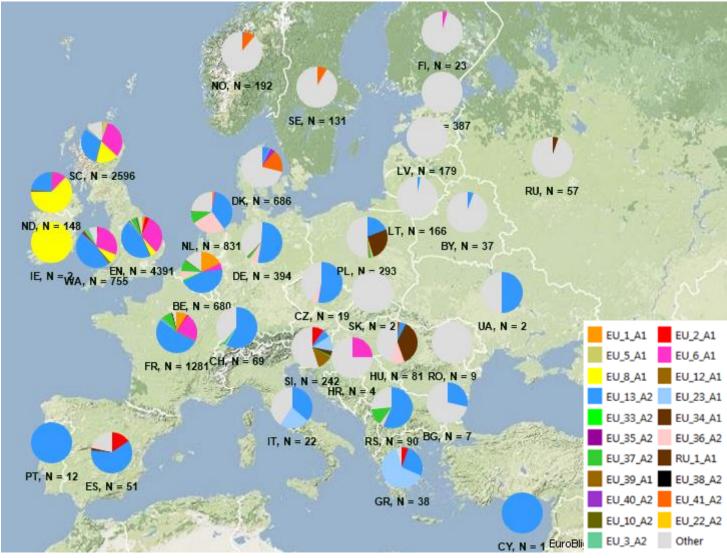
EU P. infestans genotype change







2013-2018 summary

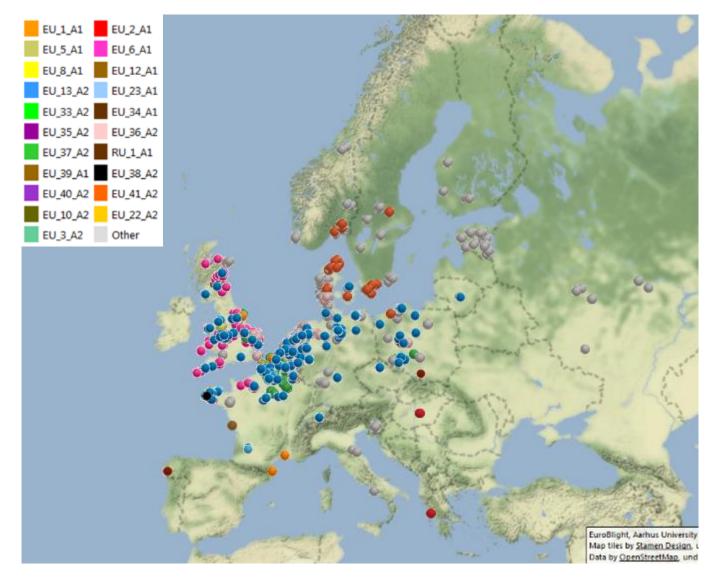






• >8000 samples from 34 countries

2017 samples

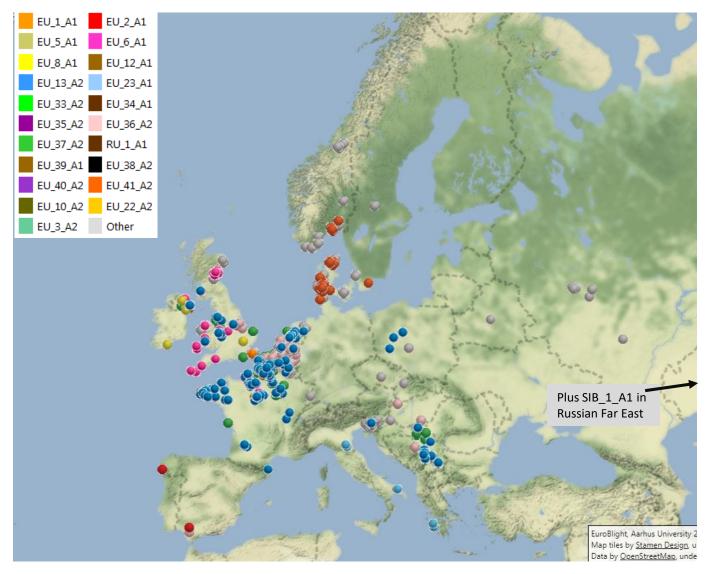






• 1534 samples from 22 countries

2018 samples

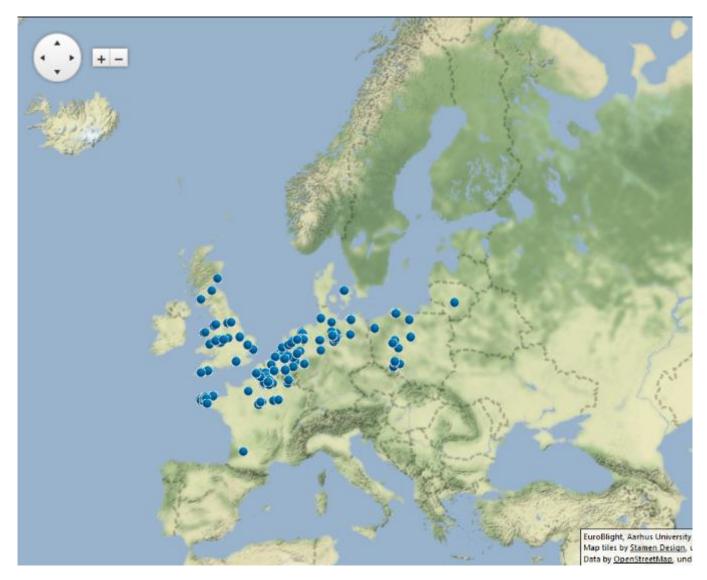






• 979 samples from 23 countries

2017 samples – EU_13_A2

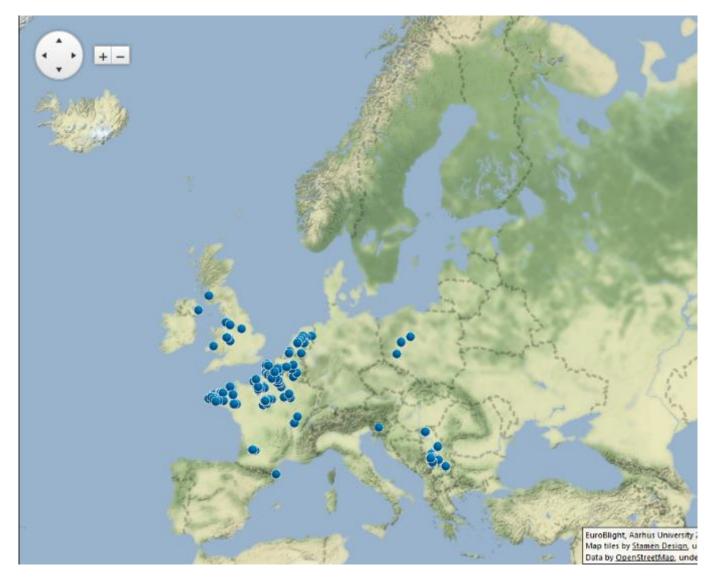






Live map at <u>https://agro.au.dk/forskning/internationale-platforme/euroblight/pathogen-characteristics-and-host-resistance/genotype-map/</u>

2018 samples – EU_13_A2

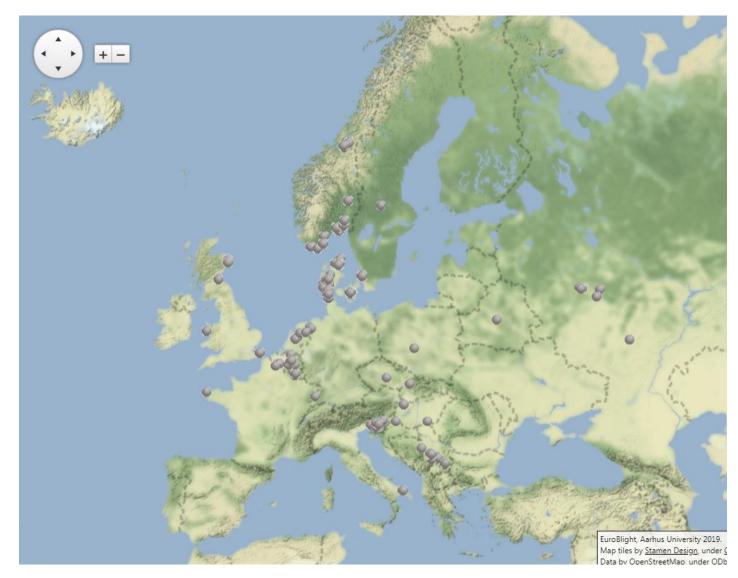






Live map at <u>https://agro.au.dk/forskning/internationale-platforme/euroblight/pathogen-characteristics-and-host-resistance/genotype-map/</u>

2018 samples – 'Other'

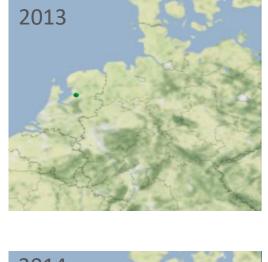






Timeline of two new clones







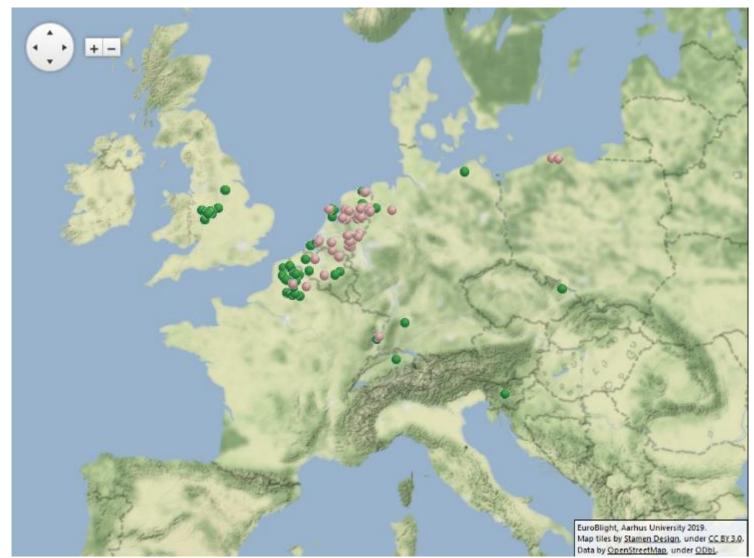






EU_36_A2 EU_37_A2

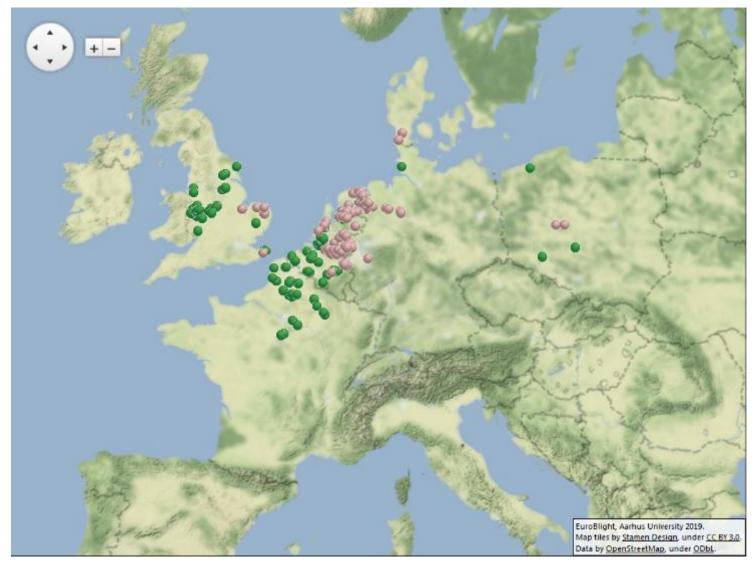
2016 36_A2 & 37_A2







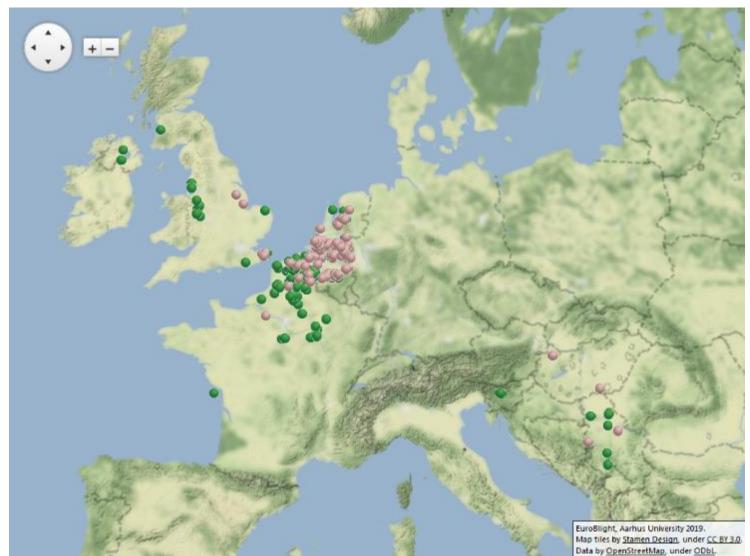
2017 36 & 37_A2







2018 36_A2 & 37_A2





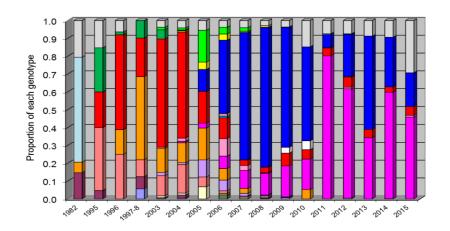


• What does it all mean to growers with crops to protect?

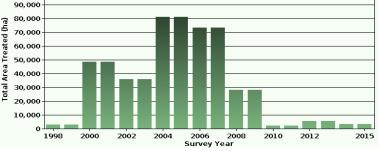
Population change and blight management



- Fungicide resistance management important
- EU_13_A2 emerged 2004, resistant to metalaxyl
- Metalaxyl use fell dramatically in UK







UK Pesticide Usage Surveys – Fera Science Ltd, UK

13_A2 global distribution



13_A2 also in;

- Egypt (Sherif el Ganainy)
- China (Ying Li)
- Bangladesh (Geert Kessel)
- Nepal (Buddhi Sharma)
- India (Pallem Chowdappa and Sanjoy Guhar Roy)
- Myanmar (WUR)
- Israel (Yigal Cohen)
- Advice to growers not always given rapidly enough



Fluazinam insensitivity



CrossMark

EU_33_A2 and EU_37_A2 show insensitivity to fluazinam

Eur J Plant Pathol https://doi.org/10.1007/s10658-018-1430-y



H. T. A. M. Schepers • G. J. T. Kessel • F. Lucca • M. G. Förch • G. B. M. van den Bosch • C. G. Topper • A. Evenhuis

UK industry awareness campaign



Richard Allison



Dark Green 37: Coming to a field near you

BY JOHN SWIRE ON OCTOBER 26, 2017

CROPS, NEWS, POTATOES

The emergence of a new strain of potato late blight (Phytophthora infestans) with resistance to fluazinam, one of the most commonly used blight fungicides, is raining concern among agronomists.

Fungicide resistance warning for new potato blight strain

Friday 30 June 2017 14:58

A reduced sensitivity to a key blight fungicide is being partly blamed for the spread of a new strain of the potato disease across Europe, with UK farmers urged to alternate their fungicide actives this season. The Dark Green 37 (EU-37) strain of blight was first detected in the Netherlands in 2013 and it has now spread to England, German, Belgium and north-west France. See also: How spud growers will benefit from blight forecasts Worryingly, this strain of the most important [...]



© Tim Scrivener

Blight actives feel the strain



Late blight pressure and a flurry of activity from blight scouts gives an early indication that new blight strain 37_A2 is on the rise. *CPM* reports.

By Lucy de la Pasture

For the first time, blight genotyping has been carried out in 'real time' this season by David Cooke at James Hutton Institute (JHI). The monitoring has been carried out following the spread of the blight strain 37_A2 (dark green) in the Netherlands (where it was first found in 2013), Germany, Belgium, and NW France where isolates of have shown a reduced sensitivity to fluazinam.

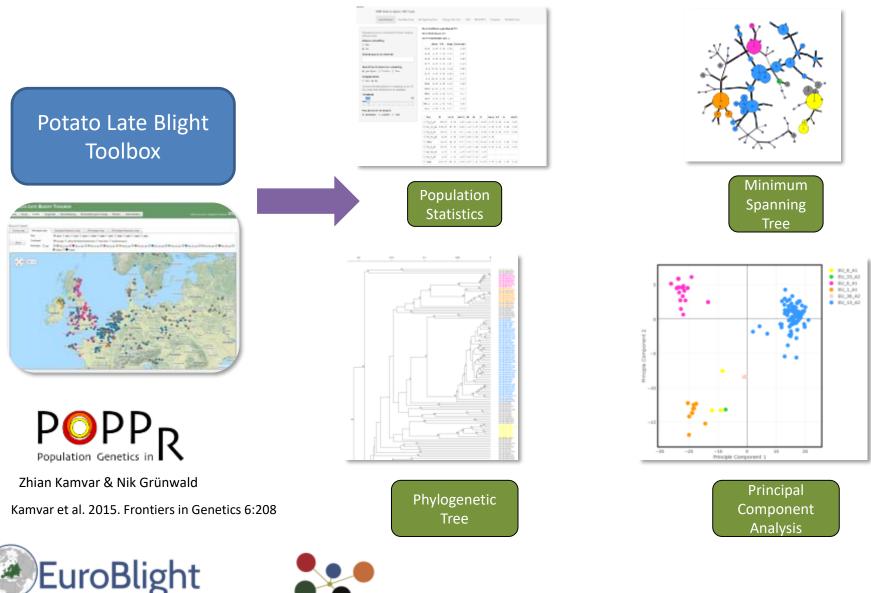
Over the summer, there've been 15 findings (up to 25 Sept) of the new blight strain, reported for the first time in the UK in 2016 in a very small number of samples.

Implications for tuber blight management





EuroBlight Population Genetics Platform

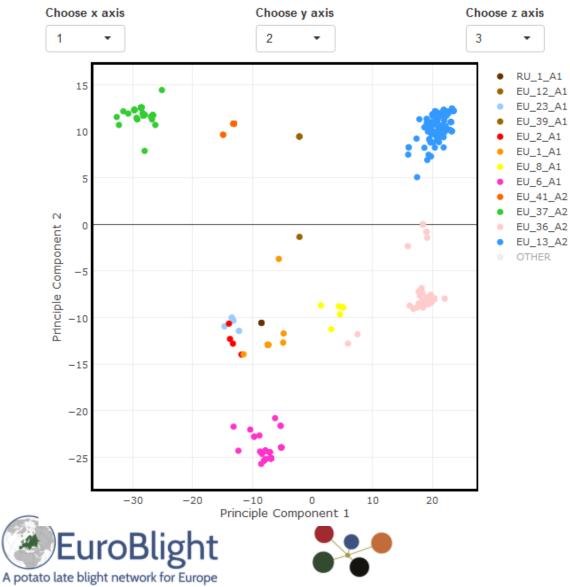


A potato late blight network for Europe



2018 Genetic diversity - PCA

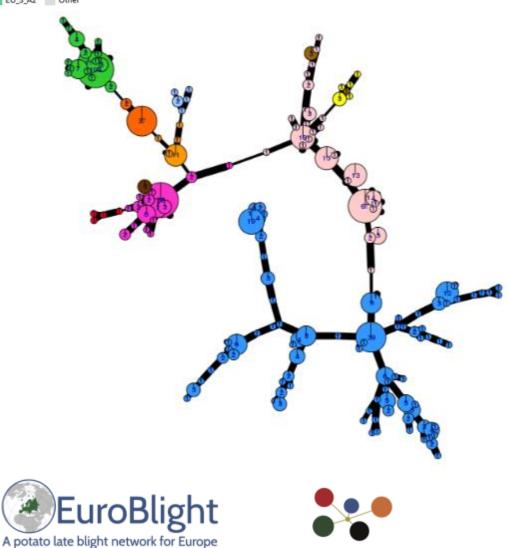
PCA with Bruvo distance as input



- 2018 Principle Coordinate Analysis (minus 'Other' samples)
- Distinct genetic clusters representing the major clones
- Minor variation within clones explains spread of points
- PCA an imperfect tool as some overlap of points inevitable

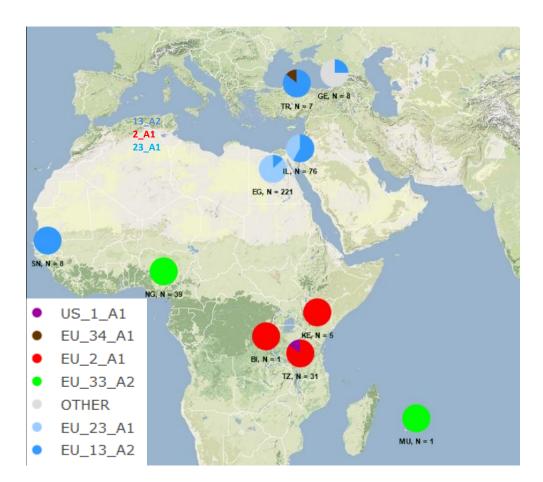


2018 Genetic diversity - MSN



- Minimum Spanning Network of 2018 samples (clones only)
- Node size proportional to the number of samples & coloured by clone
- Edge thickness proportional to genetic distance. Dark thick line genetically close - thinner lines between clones genetically distant
- 37_A2 diploid and null locus so less diverse
- Age of EU_13_A2 apparent with 84 variant MLGs in 224 samples

Global context



- Senegal 13_A2 present in 2019 (Chris Ursell CUConsulting)
- 33_A2 present in Nigeria (Emmanuel Nnadi, Plateau State University, Bokkos)
- East Africa remains free of contemporary EU lineages
- Anne Njoroge hoping to run AfricaBlight
- 13_A2 also present in Turkey, Egypt and Georgia)





Conclusions

- New insights on pathogen diversity
- Dominance of few clones across large areas of European crops means EU growers/industry share management challenges
- New clones (EU_36_A2, EU_37_A2 and EU_41_A2) continued to spread in 2018 - displacing older genotypes
- Phenotypic data is helping industry manage blight
- Primary inoculum is locally generated and spread. Better management of inoculum sources would aid management
- 'Other' populations highly diverse, ephemeral and most likely the result of sexual oospore germination. Drivers?
- High genetic diversity increases the risk of blight management problems - evolving virulence against novel host resistance genes and reduced sensitivity to fungicide active ingredients
- Collaboration with partner networks is strengthening







