



Sirtuins in tumorigenesis

ANA KULIĆ¹
MAJA SIROTKOVIĆ-SKERLEV¹
NATALIJA DEDIĆ PLAVETIĆ²
BORISLAV BELEV²
SAŠA KRALIK-OGUIĆ³
MARIJA IVIĆ⁴
DAMIR VRBANEC²

¹ Department of Oncology, Division of Pathophysiology and Experimental Oncology, University Hospital Center, Zagreb, Croatia,

² Department of Oncology, Division of Medical Oncology, University Hospital Center and Zagreb Medical School, Zagreb, Croatia

³ Department of Laboratory Diagnostic, University Hospital Center, Zagreb, Croatia,

⁴ University Hospital Center and Zagreb Medical School, Zagreb, Croatia

Correspondence:

Ana Kulić, PhD
Department of Oncology
Division of Pathophysiology and Experimental Oncology
University Hospital Center Zagreb
Kišpatićeva 12, HR- 10 000 Zagreb, Croatia
E-mail: ana.kulic1@zg.t-com.hr

Key words: sirtuins, cancer

Abstract

Sirtuins (SIRT) are group of enzymes that require nicotinamide adenine dinucleotide (NAD⁺) to catalyze their reactions. These chemical compounds have mono (ADP-ribosyl) transferase or deacetylases activities, and they can be found in nearly all species. The mammalian sirtuin family is described by seven proteins, namely. Every group of sirtuins can be found in the different regions of the cells; SIRT1 is predominantly nuclear, SIRT2 is located mainly in the cytoplasm (but it can shuttle between the nucleus and the cytoplasm), SIRT3, SIRT4, and SIRT5 are mitochondrial proteins, (SIRT3 can move from the nucleus to mitochondria during cellular stress), SIRT6 and SIRT7 are nuclear sirtuins. Sirtuins have a lot of functions in different physiological processes such as gene repression, metabolic control, apoptosis and cell survival, DNA repair, development, inflammation, neuroprotection, and healthy aging. Because of so many roles in physiological processes there is a huge interest not just in their functions but also in the different compounds which can modify their functions. In this article we will focus on the role of sirtuins in tumorigenesis.

Abbreviations:

SIRT – sirtuins
NAD – nicotinamide
ADP – adenosine diphosphate
SNPs – single nucleotide polymorphisms
HDACs – histone deacetylases
BMI – body mass index
IDH2 – isocitrate dehydrogenase 2
GDH – glutamate dehydrogenase
MnSOD – manganese superoxide dismutase
VNTR – variable number tandem repeat
TNF- α – tumor necrosis factor – alfa
HIC1 – hypermethylated in cancer 1
FOXO 1 – Forkhead Box 1
FOXO 3 – Forkhead Box 3
Cdh1 – cadherin-1
Cdc20 – cell-division cycle protein 20
NEDD4 – neural precursor cell expressed developmentally down-regulated protein 4
AML – acute myeloid leukemia
ROS – reactive oxygen species
SOD2 – superoxide dismutase
ETC – electron transport chain
Skp2-S – phase kinase associated protein 2

INTRODUCTION

Each sirtuin is characterized by approximately 275 amino acid conserved catalytic core region and by unique additional N-terminal and/or C-terminal sequences of variable length (1). Sirtuins are also known as class III histone deacetylases (HDACs), but their NAD⁺-dependency distinguishes them from other HDACs classes (2, 3).

The main sirtuin structure is characterized by a large Rossmann-fold domain (small part of compounds that is typical for NAD⁺ binding proteins), a small zinc-binding domain, and a number of flexible loops (4). The large Rossmann-fold domain is characterized by six parallel β-strands and a different number of β-helices, depending on the type of sirtuin. Zinc-binding domain is characterized by the three antiparallel β-sheets, a variable β-helical region and a Zn²⁺ cation (5). One of the most flexible regions of sirtuins is cofactor binding loop that connects the large and the small domain of enzyme (6, 7). This loop conformation is dependent of the NAD⁺ or other reaction intermediates presence (the most important is 2-O-acetyl-ADP-ribose) (8).

OVERVIEW OF SIRTUINS FUNCTIONS

Different roles of sirtuins have been described in a great number of physiological or pathological conditions.

Some of them are metabolism, aging, circadian clock regulation, pathophysiology of cancer, different inflammatory conditions, nutritional behavior and obesity (5, 9). However, different types of sirtuins have different physiological function in the human body (Figure 1).

A recent study has reported associations of SIRT1 single nucleotide polymorphisms (SNPs) to the both obesity and body mass index (BMI) (10). SIRT1 is linked to the mitochondrial biogenesis in some tissues. It also stimulates fat and cholesterol catabolism in liver, skeletal muscle, and adipose tissue.

Some recent studies have shown impaired regulation of SIRT2 in glioma. SIRT2 deletion can cause tumor occurrence; to the contrary its repletion may be used as a tumor suppressive therapy (11). SIRT2-deficient mice develop gender-specific tumorigenesis; mammary tumor in females and hepatocellular carcinoma in males (12). SIRT3 is presented in both mitochondria and nucleus. It has main role in cellular energy metabolism and redox regulation by deacetylating some mitochondrial proteins such as acetyl-coenzyme A synthetase 2, isocitrate dehydrogenase 2 (IDH2), glutamate dehydrogenase (GDH), manganese superoxide dismutase (MnSOD) (13). This sirtuin is the only one for which the correlation between a polymorphism and prolonged human life has been proven (12).

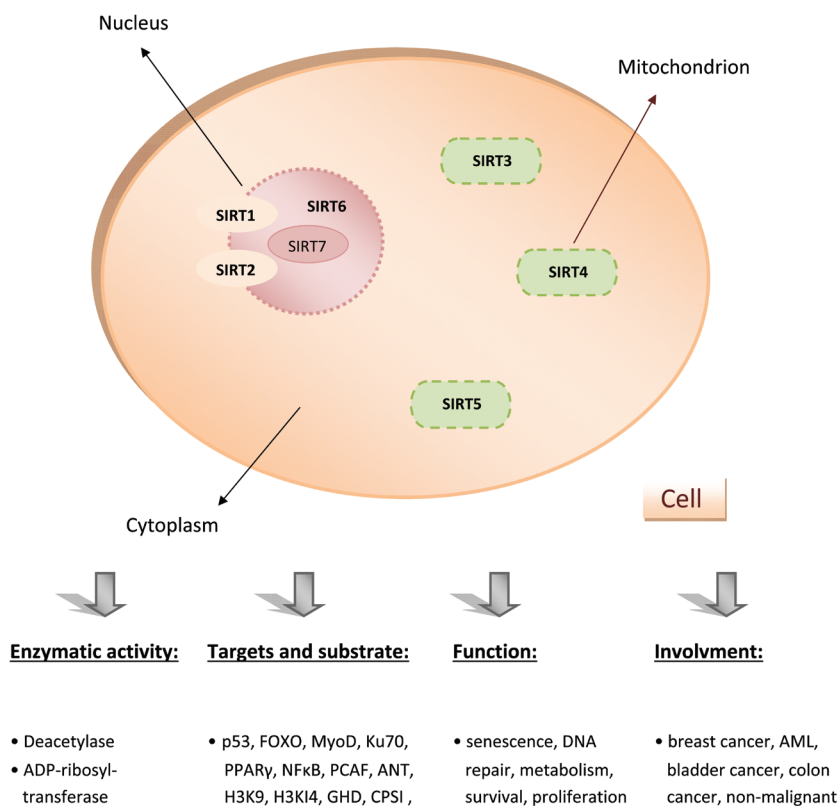


Figure 1. Location of the sirtuins in the cell. Their enzymatic activity, targets and substrates, function and involvement in various tumors are listed.

It is known that a VNTR polymorphism in intron 5 of the SIRT3 gene has an allele-specific enhancer activity. The allele which has not enhancer activity is almost absent in males older than 90 years (14).

SIRT4 is localized in mitochondria and has a role in the regulation of cellular metabolic functions like insulin secretion and fatty acid oxidation (15, 16). Some research has shown that SIRT4-depleted mice develop hyperinsulinemia and lung carcinoma (15, 17). SIRT5 is also mitochondrial sirtuin just like SIRT3 and SIRT 4.

Study of Nakagawa et al has shown that SIRT5-null mice can develop urea cycle defect and hyperammonemia after fasting (18).

SIRT6 is localized to the nucleus and has both deacetylase and ADP-ribosyltransferase activity. SIRT6 has been shown to have a function in the regulation of transcription, genome stability, TNF- α secretion, metabolism and life expectancy (19). SIRT6 deficient mice die around 1 month after birth, showing premature aging phenotypes, hypoglycemia, cardiac hypertrophy, hypersensitivity to DNA damage, and genomic instability (19).

SIRT7 is the only sirtuin localized to the nucleolus and it is linked to the ribosome biogenesis by the regulation of transcription by positive regulation of RNA polymerase I transcription (20). SIRT7 deficiency induces apoptosis in human cells what indicates SIRT7 importance in cell survival. The results of some recent studies have shown that SIRT7-deficient mice die around 1 year after birth, and develop inflammatory cardiomyopathy (21).

SIRTUINS AND TUMORIGENESIS

Sirtuins require special attention in cancer research because of their biological role in metabolism, cell death, regulation of genomic stability, inflammation and cellular proliferation. Various studies have shown that studying sirtuins can contribute to answering the question about the connection between tumor development and aging.

SIRTUIN 1 AND TUMORIGENESIS

Sirtuin 1 (SIRT1) has a dual role in the development of tumors as a tumor suppressor or promoter depending on the type of tumor and the spatial distribution of SIRT1 upstream and downstream factors (Figure 1).

SIRT1 participates in the development and progression of tumors through deacetylation causing inhibition of the function of the tumor suppressor p53, p73, and gene hypermethylated in cancer 1 (HIC1) (17-19). During aging reduction of HIC1 promoter hypermethylation occurs as well as an increase of SIRT1 content. Both processes increase cell survival and tumor development (19). SIRT1 promotes cell migration by direct interaction with cactactin (22) and promotes the expression of multidrug

resistance – associated protein 2(23). Additionally, SIRT1 promotes chemoresistance of tamoxifen – resistant breast cancer cells by deacetylating FOXO1 (23). Increased expression of SIRT1 in advanced prostate cancer promotes cell invasion, migration and metastasis through matrix metalloproteinase -2 (24). A study of SIRT 1 in colorectal cancer showed a correlation with increased expression of c – Myc (25). SIRT1 is increased in breast carcinoma (26), colon cancer (27), lung cancer (28), prostate cancer (29), thyroid cancer (30), gastric cancer (31), liver cancer (32), pancreatic cancer (33), ovarian and cervical cancers (34).

On the other hand SIRT1 can prevent tumor growth through mechanisms of genomic protection by enabling protection from stress, DNA repair mechanism and regulation of metabolism. In addition, tumor cells themselves can prevent and control innate and adaptive immune responses through inactivation of NF – κ B transcription factor and reduction of surviving (35, 36). SIRT1 expression is reduced in human head and neck squamous cell carcinoma and is associated with poor prognosis in patients with this type of carcinoma (37). It has been shown that inhibition of SIRT1 blocks p53-dependent apoptosis and DNA damage signaling which favors the growth of tumors. Further studies are needed in order to clarify the different roles of SIRT1 in tumor development.

SIRTUIN 2 AND TUMORIGENESIS

In tumorigenesis, sirtuin 2 (SIRT2) can have the function of a promoter or a suppressor according to the type of tumor. SIRT2 functions as a tumor suppressor through the deacetylation of its substrates, such as FOXO1 (Forkhead Box 1)(38), FOXO 3a (Forkhead Box 3) (39), Cdh1 (12), Cdc20 (12), H3K56 (40), or H4K16 (41). These are important molecules that maintain cell cycles, replication, and DNA damage response. Several studies have shown that SIRT2 may function as a tumor suppressor by maintaining mitotic integrity in a cell (11). It was shown that the expression of SIRT2 is down-regulated in breast cancer (41), head and neck squamous cell carcinoma(37), gliomas (11) and esophageal adenocarcinoma. The deficiency of SIRT2 increases the levels of mitotic regulators such as Aurora A and Aurora B (12).

There are also some opposite data suggesting that SIRT2 may have tumor-promoter characteristics. It was observed that the expression of SIRT2 was increased in acute myeloid leukemia (42), neuroblastoma cells, pancreatic cancer cells (43), and hepatocellular carcinoma (44). It is upregulated in pancreatic cancer cells by c-MYC and in neuroblastoma cells by N-MYC. SIRT2 stabilizes N-MYC and c-MYC protein by downregulation of ubiquitin-protein ligase NEDD4 expression. In AML cells SIRT2 and NAD⁺ salvage enzyme nicotinamide phosphoribosyl transferase are upregulated and included in the abnormal proliferation and survival of leukemic cells.

SIRTIUIN 3 AND TUMORIGENESIS

Metabolic transformation is one of the major characteristics of tumor.

It has been shown that knock-out mitochondrial-derived Sirtuin 3 (SIRT3) increases spontaneous tumorigenesis in mammary glands, indicating a role of SIRT3 as a tumor suppressor (45). SIRT3 participates in mitochondrial metabolism, counteracts oxidative stress, defends cells against apoptosis and prevents cell ageing and tumor formation.

Several studies have shown that under conditions of stress SIRT3 regulates ROS homeostasis through deacetylating and activating superoxide dismutase (SOD2) (46, 47). Reduced expression of SIRT3 leads to increased production of ROS by reducing activity of SOD2 or increasing leakage of electrons in the electron transport chain (ETC), which promotes genomic instability and the development of tumors (45). Some studies have shown that lack of SIRT3 increased glycolytic metabolism by enhancing the stability of HIF1 α (48).

Moreover SIRT3 inhibits tumorigenesis by deacetylating and inactivating S – phase kinase associated protein 2 (Skp2), a subunit of the E3 ubiquitin kinases that are important in the S phase of the cell cycle (49). Accordingly SIRT3 is reduced in human breast cancers, hepatocellular carcinoma, and head and neck squamous cell carcinoma (50). Further studies are needed in order to explain the possible role of SIRT3 in therapeutic procedures.

SIRTIUIN 4 AND TUMORIGENESIS

The role of Sirtuin 4 (SIRT4) has not yet been elucidated, but recent studies have shown that it may have a role as a tumor suppressor (17, 51).

Expression of protein SIRT4 is decreased in human gastric, ovarian, bladder, and breast carcinoma compared to normal tissue. Increased content of SIRT4 reduces transformation and cell proliferation (51). Some other studies have shown that a lack of SIRT4 leads to increased glutamine – dependent cell proliferation and genome instability caused by the stress resulting in tumorigenesis (17). SIRT4 knock-out mice were shown to have increased spontaneous lung tumors compared to wild-type mice (17). Future research should show the significance of SIRT4 expression in various cancers, and the significance of its interaction with glutamine.

SIRTIUIN 5 AND TUMORIGENESIS

The role of Sirtuin 5 (SIRT5) in the development of cancer is not known.

SIRTIUIN 6 AND TUMORIGENESIS

Some studies have shown that Sirtuin 6 (SIRT6) may have a tumor suppressor function through reduction of Myc and HIF1 α transcriptional activity (52).

Expression of SIRT6 is reduced in human pancreatic cancer and colon carcinoma. Some studies have shown that SIRT6 can suppress the initiation of liver cancer through reduction of survivin expression by deacetylation of H3K9 (52).

Additionally, SIRT 6 reduces the activity of NF–kB (53). SIRT6 also participates as a tumor suppressor in breast cancer by regulating DNA repair and metabolism.

Some studies have shown that SIRT6 can increase the secretion of cytokines in cancer and produce an increase in angiogenesis through regulation of Ca²⁺ responses or deacetylation of TNF α (54).

Increased expression of SIRT6 can increase resistance to chemotherapy such as paclitaxel and epirubicin by inhibiting FOXO 3a activity (55).

These data suggest multiple mechanisms by which SIRT6 exerts its effects in cancer cells. Future studies should investigate the biological effects and molecular mechanisms of SIRT6 in individual tumors.

SIRTIUIN 7 AND TUMORIGENESIS

Biological role of Sirtuin 7 (SIRT7) in tumors is not fully explained.

Previous results have shown that SIRT7 can be oncogene, activating oncogenes capacity of cancer cells such as loss of contact inhibition and anchorage -independent growth. Lack of SIRT 7 reduces tumor potential of cancer cells (56). Recent research has shown an increase of SIRT7 in hepatocellular carcinoma and that deletion of SIRT7 may cause suppression of cell growth (57).

Further studies should investigate the molecular mechanisms of SIRT7 activities in cancer.

CONCLUSIONS

Sirtuins play a significant role in inflammation, metabolism, cellular proliferation, regulation of genomic stability, and cell death. Based on their involvement in these processes sirtuins are involved in cancer pathophysiology. It became clear that sirtuins can help to address the question on the molecular relationship between the tumor and aging. Future research is needed in order to determine the molecular mechanism of action of each sirtuin in cancer and the possible role of sirtuins in cancer therapy.

REFERENCES

1. SANDERS B D, JACKSON B, MARMORSTEIN R 2010. Structural basis for sirtuin function: What we know and what we don't. *Biochim Biophys Acta* 1804: 1604–1616
2. FRYE R A 1999 Characterization of five human cDNAs with homology to the yeast SIR2 gene: Sir2-like proteins (sirtuins) metabolize NAD and may have protein ADP-ribosyltransferase activity. *Biochem Biophys Res Commun* 260: 273-289

3. TANNY J C, DOWD G J, HUANG J, HILZ H, MOAZED D 1999 An enzymatic activity in the yeast Sir2 protein that is essential for gene silencing. *Cell* 99: 735-745
4. BRUZZONE S, PARENTI M D, GROZIO A, BALLESTRERO A, BAUER I, DEL RIO A, NENCIONI A 2013 Rejuvenating Sirtuins: The Rise of a New Family of Cancer Drug Targets. *Curr Pharm Des* 19: 614-623
5. FINKEL T, DENG C X, MOSTOSLAVSKY R 2009 Recent progress in the biology and physiology of sirtuins. *Nature* 460: 587-591
6. VAZIRI H, DESSAIN S K, NG EATON E, IMAI S I, FRYE R A, PANDITA T K, GUARENTE L, WEINBERG R A 2001 hSIR2 (SIRT1) functions as an NAD-dependent p53 deacetylase. *Cell* 107: 149-159
7. MICHISHITA E, PARK J Y, BURNESKIS J M, BARRETT J C, HORIKAWA I 2005 Evolutionarily conserved and nonconserved cellular localizations and functions of human SIRT proteins. *Mol Biol Cell* 16: 4623-4635
8. MAHLKNECHT U, HO A D, VOELTER-MAHLKNECHT S 2006 Chromosomal organization and fluorescence in situ hybridization of the human Sirtuin 6 gene. *Int J Oncol* 28: 447-456
9. LIBERT S, POINTER K, BELL E L, DAS A, COHEN D E, ASARA J M, KAPUR K, BERGMANN S, PREISIG M, OTOWA T, KENDLER K S, CHEN X, HETTEMA J M, VAN DEN OORD E J, RUBIO J P, GUARENTE L 2011 SIRT1 activates MAO-A in the brain to mediate anxiety and exploratory drive. *Cell* 147: 1459-1472
10. FORD E, VOIT R, LISZT G, MAGIN C, GRUMMT I, GUARENTE L 2007 Mammalian Sir2 homolog SIRT7 is an activator of RNA polymerase I transcription. *Genes Dev* 20: 1075-1080
11. HIRATSUKA M I, INOUE T, TODA T, KIMURA N, SHIRAYOSHI Y, KAMITANI H, WATANABE T, OHAMA E, TAHIMIC C G, KURIMASA A, OSHIMURA M 2003 Proteomics-based identification of differentially expressed genes in human gliomas: down-regulation of SIRT2 gene. *Biochem Biophys Res Commun* 309: 558-566
12. KIM H S, VASSILOPOULOS A, WANG R H, LAHUSEN T, XIAO Z, XU X, LI C, VEENSTRAT D, LI B, YU H, JIJ, WANG X W, PARK S H, CHA Y I, GIUS D, DENG C X 2011 SIRT2 maintains genome integrity and suppresses tumorigenesis through regulating APC/C activity. *Cancer Cell* 20: 487-499
13. HONGFENG Y, LEILA S, WENYONH C 2013 The emerging and diverse roles of sirtuins in cancer: a clinical perspective. *Oncotargets and Therapy* 10: 1399-1416
14. BELLIZZI D, ROSE G, CAVALCANTE P, COVELLO G, DATO S, DE RANGO F, GRECO V, MAGGIOLINI M, FERACO E, MARI V, FRANCESCHI C, PASSARINO G, DE BENEDICTIS G 2005 A novel VNTR enhancer within the SIRT3 gene, a human homologue of SIR2, is associated with survival at oldest ages. *Genomics* 85: 258-263
15. HAIGIS M C, MOSTOSLAVSKY R, HAIGIS K M, FAHIE K, CHRISTODOULOU D C, MURPHY A J, VALENZUELA D M, YANCOPOULOS G D, KAROW M, BLANDER G, WOLBERGER C, PROLLA T A, WEINDRUCH R, ALT F W, GUARENTE L 2006 SIRT4 inhibits glutamate dehydrogenase and opposes the effects of calorie restriction in pancreatic beta cells. *Cell* 126: 941-954
16. NASRIN N, WUX, FORTIER E, FENG Y, BARE' O C, CHEN S, REN X, WU Z, STREEPER R S, BORDONE 2010 SIRT4 regulates fatty acid oxidation and mitochondrial gene expression in liver and muscle cells. *J Biol Chem* 285: 31995-32002
17. JEONG S M, XIAO C, FINLEY L W, LAHUSEN T, SOUZA A L, PIERCE K, LI Y H, WANG X, LAURENT G, GERMAN N J, XU X, LI C, WANG R H, LEE J, CSIBI A, CERIONE R, BLENIS J, CLISH C B, KIMMELMAN A, DENG C X, HAIGIS M C 2013 SIRT4 has tumor-suppressive activity and regulates the cellular metabolic response to DNA damage by inhibiting mitochondrial glutamine metabolism. *Cancer Cell* 23: 450-463
18. NAKAGAWA T, LOMB D J, HAIGIS M C, GUARENTE L 2009 SIRT5 Deacetylates carbamoyl phosphate synthetase 1 and regulates the urea cycle. *Cell* 137: 560-570
19. MAO Z, TIAN X, VAN METER M, KE Z, GORBUNOVA V, SELUANOV A 2012 Sirtuin 6 (SIRT6) rescues the decline of homologous recombination repair during replicative senescence. *Proc Natl Acad Sci USA* 109: 11800-11805
20. VILLALBA J, ALCAIN F J 2012 Sirtuins activators and inhibitors. *Bio Factors* 10: 349-359
21. VAKHRUSHEVA O I, SMOLKA C, GAJAWADA P, KOSTIN S, BOETTGER T, KUBIN T, BRAUN T, BOBER E 2008 Sir7 increases stress resistance of cardiomyocytes and prevents apoptosis and inflammatory cardiomyopathy in mice. *Circ Res* 102: 703-710
22. ZHANG Y, ZHANG M, DONG H, YONG S, LI X, OLASHAW N, KRUK P A, CHENG J Q, BAI W, CHEN J, NICOSIA S V, ZHANG X 2009. Deacetylation of cortactin by SIR T1 promotes cell migration. *Oncogene* 28: 445-460
23. CHOI H K, CHO K B, PHUONG N T, HAN C Y, HAN H K, HIEN T T, CHOI H S, KANG K W 2013 SIRT1-Mediated FoxO1. Deacetylation Is Essential for Multidrug Resistance-Associated Protein 2 Expression in Tamoxifen-Resistant Breast Cancer Cells. *Mol Pharm* 10: 2517-27
24. LOVAAS J D, ZHU L, CHIAO C Y, BYLES V, FALLER D V, DAI Y 2013 SIRT1 enhances matrix metalloproteinase-2 expression and tumor cell invasion in prostate cancer cells. *Prostate* 73: 522-530
25. MENSSEN A I, HYDBRING P, KAPPELLE K, VERVOORTS J, DIEBOLD J, LÜSCHER B, LARSSON L G, HERMEKING H 2012 The c-MYC oncoprotein, the NAMPT enzyme, the SIRT1-inhibitor DBC1, and the SIRT1 deacetylase form a positive feedback loop. *Proc Natl Acad Sci USA* 109: E187-E196
26. WANG F, CHAN C H, CHEN K, GUAN X, LIN H K, TONG Q 2012 Deacetylation of FOXO3 by SIRT1 or SIRT2 leads to Skp2-mediated FOXO3 ubiquitination and degradation. *Oncogene* 31: 1546-1557
27. JUNG W, HONG K D, JUNG W Y, LEE E, SHIN B K, KIM H K, KIM A, KIM B H 2013 SIRT1 Expression is Associated with Good Prognosis in Colorectal Cancer. *Korean J Pathol* 47: 332-339
28. NOH S J, BAEK H A, PARK H S, JANG K Y, MOON W S, KANG M J, LEE D G, KIM M H, LEE J H, CHUNG M J 2013 Expression of SIRT1 and cortactin is associated with progression of non-small cell lung cancer. *Pathol Res Pract* 209: 365-370
29. HIRAIKE H, WADA-HIRAIKE O, NAKAGAWA S, SAJI S, MAEDA D, MIYAMOTO Y, SONE K, TANIKAWA M, ODA K, NAKAGAWA K, YANO T, FUKAYAMA M, TAKETANI Y 2011 Expression of DBC1 is associated with nuclear grade and HER2 expression in breast cancer. *Exp Ther Med* 2: 1105-1109
30. HERRANZ D, MARAVER A, CAÑAMERO M, GÓMEZ-LÓPEZ G, INGLADA-PÉREZ L, ROBLEDO M, CASTELBLANCO E, MATIAS-GUIU X, SERRANO M 2013 SIRT1 promotes thyroid carcinogenesis driven by PTEN deficiency. *Oncogene* 32: 4052-4056
31. FENG A N, ZHANG L H, FAN X S, HUANG Q, YE Q, WU H Y, YANG J 2011 Expression of SIRT1 in gastric cardiac cancer and its clinicopathologic significance. *Int J Surg Pathol* 19: 743-750
32. CHEN H C, JENG Y M, YUAN R H, HSU H C, CHEN Y L 2012 SIRT1 promotes tumorigenesis and resistance to chemotherapy in hepatocellular carcinoma and its expression predicts poor prognosis. *Ann Surg Oncol* 19: 2011-2019
33. ZHAO G, CUI J, ZHANG J G, QIN Q, CHEN Q, YIN T, DENG S C, LIU Y, LIU L, WANG B, TIAN K, WANG G B,

- WANG CY 2011 SIRT1 RNAi knockdown induces apoptosis and senescence, inhibits invasion and enhances chemosensitivity in pancreatic cancer cells. *Gene Ther* 18: 920–928
34. JANG K Y, KIM K S, HWANG S H, KWON K S, KIM K R, PARK H S, PARK B H, CHUNG M J, KANG M J, LEE D G, MOON W S 2009 Expression and prognostic significance of SIRT1 in ovarian epithelial tumours. *Pathology* 41: 366–371
 35. KONG S, KIM S J, SANDAL B, LEE S M, GAO B, ZHANG D D, FANG D 2011 The type III histone deacetylase Sirt1 protein suppresses p300-mediated histone H3 lysine 56 acetylation at Bclaf1 promoter to inhibit T cell activation. *J Biol Chem* 286: 16967–16975
 36. YEUNG F, HOBERG J E, RAMSEY C S, KELLER M D, JONES D R, FRYE R A, MAYO M W 2004 Modulation of NF-kappaB-dependent transcription and cell survival by the SIRT1 deacetylase. *EMBO J* 23: 2369–2380
 37. LAI C C, LIN P M, LIN S F, HSU C H, LIN H C, HU M L, HSU C M, YANG M Y 2013 Altered expression of SIRT gene family in head and neck squamous cell carcinoma. *Tumour Biol* 34: 1847–1854
 38. JING E, GESTA S, KAHN C R 2007 SIRT2 regulates adipocyte differentiation through FoxO1 acetylation/deacetylation. *Cell Metab* 6: 105–114
 39. WANG F, NGUYEN M, QIN F X, TONG Q 2007 SIRT2 deacetylates FOXO3a in response to oxidative stress and caloric restriction. *Aging Cell* 6: 505–514
 40. DAS C, LUCIA M S, HANSEN K C, TYLER J K 2009 CBP/p300-mediated acetylation of histone H3 on lysine 56. *Nature* 459: 113–117
 41. SERRANO L, MARTÍNEZ-REDONDO P, MARAZUELA-DUQUE A, VAZQUEZ B N, DOOLEY S J, VOIGT P, BECK D B, KANE-GOLDSMITH N, TONG Q, RABANAL R M, FONDEVILA D, MUÑOZ P, KRÜGER M, TISCHFIELD J A, VAQUERO A 2013 The tumor suppressor Sirt2 regulates cell cycle progression and genome stability by modulating the mitotic deposition of H4K20 methylation. *Genes Dev* 27: 639–653
 42. DAN L, KLIMENKOVA O, KLIMIANKOU M, KLUSMAN J H, VAN DEN HEUVEL-EIBRINK M M, REINHARDT D, WELTE K, SKOKOWA J 2012 The role of sirtuin 2 activation by nicotinamide phosphoribosyl transferase in the aberrant proliferation and survival of myeloid leukemia cells. *Haematologica* 97: 551–559
 43. LIU P Y, XU N, MALYUKOVA A, SCARLETT C J, SUN Y T, ZHANG X D, LING D, SU S P, NELSON C, CHANG D K, KOACH J, TEE A E, HABER M, NORRIS M D, TOON C, ROOMAN I, XUE C, CHEUNG B B, KUMAR S, MARSHALL G M, BIANKIN A V, LIU T 2013 The histone deacetylase SIRT2 stabilizes Myc oncoproteins. *Cell Death Differ* 20: 503–514
 44. CHEN J, CHAN A W, TO K F, CHEN W, ZHANG Z, REN J, SONG C, CHEUNG Y S, LAI P B, CHENG S H, NG M H, HUANG A, KO B C 2013 SIRT2 overexpression in hepatocellular carcinoma mediates epithelial to mesenchymal transition by protein kinase B/glycogen synthase kinase-3 β / β -catenin signaling. *Hepatology* 57: 2287–2298
 45. KIM H S, PATEL K, MULDOON-JACOBS K, BISHT K S, AYKIN-BURNS N, PENNINGTON J D, VAN DER MEER R, NGUYEN P, SAVAGE J, OWENS K M, VASSILOPOULOS A, OZDEN O, PARK S H, SINGH K K, ABDULKADIR S A, SPITZ D R, DENG C X, GIUS D 2010 SIRT3 is a mitochondria-localized tumor suppressor required for maintenance of mitochondrial integrity and metabolism during stress. *Cancer Cell* 17: 41–52
 46. TAO R, COLEMAN M C, PENNINGTON J D, OZDEN O, PARK S H, JIANG H, KIM H S, FLYNN C R, HILL S, HAYES MCDONALD W, OLIVIER A K, SPITZ D R, GIUS D 2010 Sirt3-mediated deacetylation of evolutionarily conserved lysine 122 regulates MnSOD activity in response to stress. *Mol Cell* 40: 893–904
 47. CHEN Y, ZHANG J, LIN Y, LEI Q, GUAN K L, ZHAO S, XIONG Y 2011 Tumour suppressor SIRT3 deacetylates and activates manganese superoxide dismutase to scavenge ROS. *EMBO Rep* 2: 534–541
 48. FINLEY L W, CARRACEDO A, LEE J, SOUZA A, EGIA A, ZHANG J, TERUYA-FELDSTEIN J, MOREIRA P I, CARDOSO S M, CLISH C B, PANDOLFI P P, HAIGIS M C 2011 SIRT3 opposes reprogramming of cancer cell metabolism through HIF1 α destabilization. *Cancer Cell* 19: 416–428
 49. INUZUKA H, GAO D, FINLEY L W, YANG W, WAN L, FUKUSHIMA H, CHIN Y R, ZHAI B, SHAIK S, LAU A W, WANG Z, GYGI S P, NAKAYAMA K, TERUYA-FELDSTEIN J, TOKER A, HAIGIS M C, PANDOLFI P P, WEI W 2012 Acetylation-dependent regulation of Skp2 function. *Cell* 50: 179–193
 50. FINLEY L W, HAIGIS M C 2012 Metabolic regulation by SIRT3: implications for tumorigenesis. *Trends Mol Med* 18: 516–523
 51. CSIBI A, FENDT S M, LI C, POULOGIANNIS G, CHOO A Y, CHAPSKI D J, JEONG S M, DEMPSEY J M, PARKHITKO A, MORRISON T, HENSKE E P, HAIGIS M C, CANTLEY L C, STEPHANOPOULOS G, YU J, BLENIS J 2013 The mTORC1 pathway stimulates glutamine metabolism and cell proliferation by repressing SIRT4. *Cell* 153: 840–854
 52. SEBASTIÁN C, ZWAANS B M, SILBERMAN D M, GYMREK M, GOREN A, ZHONG L, RAM O, TRUELOVE J, GUIMARAES A R, TOIBER D, COSENTINO C, GREENSON J K, MACDONALD A I, MCGLYNN L, MAXWELL F, EDWARDS J, GIACOSA S, GUCCIONE E, WEISSLEDER R, BERNSTEIN B E, REGEV A, SHIELS P G, LOMBARD D B, MOSTOSLAVSKY R 2012 The histone deacetylase SIRT6 is a tumor suppressor that controls cancer metabolism. *Cell* 151: 1185–1199
 53. MIN L, JI Y, BAKIRI L, QIU Z, CEN J, CHEN X, CHEN L, SCHEUCH H, ZHENG H, QIN L, ZATLOUKAL K, HUI L, WAGNER E F 2012 Liver cancer initiation is controlled by AP-1 through SIRT6-dependent inhibition of survivin. *Nat Cell Biol* 14: 1203–1211
 54. BAUER I, GROZIO A, LASIGLIE D, BASILE G, STURLA L, MAGNONE M, SOCIALI G, SONCINI D, CAFFA I, POGGI A, ZOPPOLI G, CEA M, FELDMANN G, MOSTOSLAVSKY R, BALLESTRERO A, PATRONE F, BRUZZONE S, NENCIONI A 2012 The NAD⁺-dependent histone deacetylase SIRT6 promotes cytokine production and migration in pancreatic cancer cells by regulating Ca²⁺ responses. *J Biol Chem* 287: 40924–40937
 55. KHONGKOW M, OLMOS Y, GONG C, GOMES A, MONTEIRO L J, YAGUE E, CAVACO T B, KHONGKOW P, MAN E P, LAOHASINNARONG S, KOO C Y, HARADA-SHOJI N, TSANG J W, COOMBES R C, SCHWER B, KHOO U S, LAM E W 2013 SIRT6 modulates paclitaxel and epirubicin resistance and survival in breast cancer. *Carcinogenesis* 34: 1476–1486
 56. BARBER M F, MICHISHITA-KIOIE, XI Y, TASSELLI L, KIOI M, MOQTADERI Z, TENNEN R I, PAREDES S, YOUNG N L, CHEN K, STRUHL K, GARCIA B A, GOZANI O, LI W, CHUA K F 2012 SIRT7 links H3K18 deacetylation to maintenance of oncogenic transformation. *Nature* 487: 114–118
 57. KIM J K, NOH J H, JUNG K H, EUN J W, BAE H J, KIM M G, CHANG Y G, SHEN Q, PARK W S, LEE J Y, BORLAK J, NAM S W 2013 Sirtuin7 oncogenic potential in human hepatocellular carcinoma and its regulation by the tumor suppressors MiR-125a-5p and MiR-125b. *Hepatology* 57: 1055–1067