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Stabilin-Mediated Cellular Internalization of Phosphorothioate-Modified Antisense Oligonucleotides (ASOs)

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Abstract

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Introduction: Antisense oligonucleotides (ASOs) are short chemically modified oligonucleotides (5-7.4 kDa) that can produce a pharmacological effect by binding to RNA and affecting intermediary metabolism. Over 35 phosphorothioate (PS) ASOs are at various stages of clinical development for use as therapeutic agents and pharmacological tools. Antisense therapy is a progressing area of research, as these small strands of nucleotide oligomers can be produced to silence genes that aggravate chronic disorders or infections. An important distinction for ASOs compared to DNA is the substitution of the phosphodiester (PO) backbone with the PS modification. This sulfur substitution allows for these polar polyanionic molecules to have high stability in biological fluids and selective binding to cell surfaces. Although there is a premium for clinical development of these short chain molecules, the current understanding of the pathways for their ability to traverse plasma membranes remains unresolved. Injected gymnotic ASOs have been shown to accumulate in the liver via the organ's functional scavenging mechanism in highly endocytically active sinusoidal endothelial cells (SECs) and Kupffer cells (KC) compared to hepatocytes. Our work outlines how a non-DNA binding class of scavenger receptors known as Stabilins binds to, and internalizes these small PS ASOs. **Methods:** Primary cells from rat and mouse were isolated and cultured with ¹²⁵I-ASOs. Stable cell lines expressing Stabilin-1 and two Stabilin-2 isoforms (315-HARE and 190-HARE), were used to analyze binding affinity, endocytosis and degradation of ¹²⁵I-ASOs in the cell. Colocalization was also done to analyze trafficking of ASOs once internalized within the cell by use of fluorescent ASO and lysotracker. **Results:** It was determined that PS ASOs bind with high affinity to the Stabilin class receptors with the majority of internalization performed by clathrin-mediated endocytosis. Binding was determined to be dependent on proper folding of the receptor, along with relying on salt-bridge formation. Once inside the cell via the Stabilin receptors, co-localization analysis showed ASOs being trafficked for degradation in the lysosome. Increased internalization rates of an ASO targeting the noncoding RNA of malat-1, in the Stabilin-expressing cell lines reduced malat-1 expression more efficiently, indicating not all ASOs are trafficked to the lysosome after internalization. **Conclusion:** Our work shows that ASOs are internalized into the cells of the liver through clathrin-mediated endocytosis. The understanding of the pathway(s) for chemically modified ASO internalization and trafficking with cell surface receptors will aid in future clinical design of ASOs as therapeutic agents.

Introduction: Stabilin receptors



FIG. 1: The Stabilin receptors. Stab1 and Stab2 are expressed in endothelium of liver, lymph node, spleen and bone marrow. Both receptors are comprised of the same domain organization, although Stab1 does not have a functional LINK domain and cannot be categorized as a hyalectin. Both receptors bind to heparin, GDF-15, phosphatidylserine, advanced glycation end-products (AGE), and modified LDL. Individually, Stab1 binds to lactogen and SPARC and Stab2 binds with HA, CS, and collagen pro-peptides. Neither receptor binds natural DNA or RNA with phosphodiester backbones. Both receptors undergo proteolytic cleavage/maturation with Stab1 expressed as isomers of nearly the same size in a 1:1 ratio and Stab2 expressed as 315and 190-kDa receptors in a 5:1 to 2:1 ratio depending on the tissue. \bullet = EGF domain, \triangleright = Fas-1 domain, **I** = transmembrane domain, **I** = Link domain

Introduction: Antisense Oligonucleotides (ASO)



OMe F fmfmfmfmfmfmfmfmee <u>afnfmfmfmfmfmfmee</u> <u>fofofofofofofofof</u> OMe F F-OM

 Table. 1: ASO polymers used in this study.



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