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The body-size dependence of mutual interference

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Abstract

The parameters that drive population dynamics typically show a relationship with body size. By contrast, there is no theoretical or empirical support for a body-size dependence of mutual interference, which links foraging rates to consumer density. Here, I develop a model to predict that interference may be positively or negatively related to body size depending on how resource body size scales with consumer body size. Over a wide range of body sizes, however, the model predicts that interference will be body-size independent. This prediction was supported by a new data set on interference and consumer body size. The stabilizing effect of intermediate interference therefore appears to be roughly constant across size, while the effect of body size on population dynamics is mediated through other parameters.

Keywords: interference, population dynamics, interaction strength, allometry

1. Introduction

The abundance and dynamics of populations depend on the parameters that set species interactions, growth rates and death rates [1–4]. Documenting patterns in these parameters is therefore crucial to understanding ecological communities and predicting changes in their structure in space and time. One common pattern is that the parameters are strongly tied to body size. For example, intrinsic rates of growth and mortality rates both show $-\frac{1}{4}$ power scalings with body size [5,6].

Foraging interactions between consumers (*C*) and their resources (*R*) are also body-size dependent [7]. These interactions are generally modelled with a functional response that relates prey density to per capita foraging rate (f) [8]. A typical functional response is

$$f = \frac{aR}{1+ahR} \tag{1.1}$$

where *a* is the area of capture, which sets how fast a consumer clears its environment of resources, and *h* is the pause in searching upon prey capture during which organisms "handle" their prey. Both *a* and *h* have power-law-like relationships with body size for a wide array of taxonomic groups [2–4, 9].

To account for the negative effect of increasing consumer density on foraging rates (mutual interference), Equation (1.1) has been modified in several ways [10, 11]. One common way to account for interference is with the Hassel–Varley–Holling (HVH) model, which reduces the a parameter by linking it to consumer density with a power-law function [12, 13]:

$$f = \frac{\alpha C^m R}{1 + \alpha C^m h R'} \tag{1.2}$$

where *m* is "mutual" interference, and α is the value of *a* when C = 1 or *m* = 0. Because interference has a strong effect on population stability [14, 15], any body-size dependence of this parameter would indicate systematic dependence of stability on body size [16,17]. Here, I assess the body-size dependence of mutual interference using a new mechanistic model and an empirical analysis of a new dataset assembled from the literature.

2. Model

Although originally phenomenological, the HVH model can be mechanistically generated by incorporating the effect of predator density on the average predator velocity [15]. Area of capture (a) can be decomposed into an area of detection A_d and the encounters between consumer and resource individuals: $a = A_d \sqrt{V_c^2 + V_r^2}$, where V_c and V_r are the velocities of the consumer and the resource, respectively [18]. By rescaling the consumer velocity by C^2 , to represent mass-action encounters among consumers, the rescaled area of capture \tilde{a} declines as *C* increases:

$$\tilde{a} = A_{\rm d} \sqrt{\frac{V_{\rm c}^2}{C^4} + V_{\rm r}^2}$$
(2:1)

This change causes the effective searching velocity of the consumer to decline as consumer density increases, lowering encounters and thus foraging rates. Equation (2.1) does not, however, produce *m* analytically. Instead, the value of *m* must be determined from a linear regression of \tilde{a} against C [15]. Nonetheless, equation (2.1) clearly demarcates the typical range of mutual interference values found in the literature (0 to -2) [11, 15]. It produces a dependence of \tilde{a} on *C* that ranges from the power of -2 when the prey are stationary (when $V_r^2 = 0$, \tilde{a} is a function of C^2) to the power of 0 when predators are sit-andwait (when $V_c^2 = 0$, \tilde{a} is independent of *C*). It also collapses to the original expression when C = 1, when there are no other individuals with which to interfere.

To make *m* dependent on body size, I first define the scaling of velocity *V* with body mass *M* as $V = v_0 M^{\gamma}$, where γ is a scaling exponent, and n_0 is the value of *V* when M = 1. Assuming that γ does not vary between consumer and resource, because it usually falls in a narrow range of about 0.1–0.25 [19], I substitute to get

$$\tilde{a} = A_{\rm d} v_{0r} \sqrt{\frac{\Delta v_0^2 M_c^{2\gamma} + M_r^{2\gamma}}{C^4}}$$
(2:2)

Here, I have specified the mass for the consumer, M_c , and the resource, M_r , separately. For convenience, I have defined the relative velocity as $\Delta v_0 = v_{0c}/v_{0r}$ (following the notation of [20], again with subscripts *c* and *r* for consumer and resource, respectively) which allows us to have only one parameter in the radical indicating mass-specific velocity differences. When predators travel much faster than their prey, Δv_0 is large, but $\Delta v_0 \approx 0$ for sit-and-wait predators. Finally, I define the consumer-resource body size scaling as $M_r = s_0 M_c^{\psi}$, where s_0 is the value of M_r when $M_c = 1$, and ψ is a scaling exponent, and substituting this yields

$$\tilde{a} = A_{\rm d} v_{0\rm r} \sqrt{\frac{\Delta v_0^2 M_{\rm c}^{2\gamma}}{C^4} + {s_0}^{2\gamma} M_{\rm c}^{\gamma\psi}}$$
(2.3)

Because interference is more severe when V_c^2 is large relative to V_r^2 (see equation (2.1)), equation (2.3) shows that the magnitude of interference depends on the relative velocity (Δv_0) and the scaling of resource body size with consumer body size (s_0 and ψ). In other words, a relatively fast-moving consumer makes the $\Delta v_0 M_c^{2\gamma}$ term relatively large, magnifying the effect of *C* on \tilde{a} and making interference stronger (closer to m = -2). An important special case in equation (2.3) is when the scaling of resource to consumer body size (ψ) is one, which is approximately true across a large body-size range [21]. In this case, $M_c^{2\gamma}$ can be factored out, showing that interference is independent of body size. Otherwise, for any given set of parameters corresponding to specific consumer-resource groups, ψ will determine whether interference increases (when $\psi < 1$) or decreases (when $\psi > 1$) with body size.

3. Material and methods

I assessed the model's predictions for the body-size dependence of interference in two ways. In both cases, I estimated *m* by regressing \tilde{a} on C across a broad range of body sizes (10⁻⁷) to 10^5 g). In the first case, I varied only the level of ψ to show its effect. In the second case, I randomly sampled all parameters in Equation (2.3) from an empirically observed range and again estimated m across the same range of body sizes. This time I drew 500 sets of parameter values from a uniform distribution set by the typical ranges for each parameter reported in the literature: γ (0.1–0.3 [19]), ψ (0.5–1.5 [22]) and s_0 (0.001–1000 [21]). The value of Δv_0 could range from that for a sit-and-wait predator that never moves ($\Delta v_0 = 0$) to that for a consumer that moves considerably faster than its prey, such as predatory birds eating small mammals. From the velocitymass relationships in [19], this could be as much as 10-fold, so I varied Δv_0 from 0 to 10. These parameter sets reflect a behaviorally and taxonomically diverse range of possible consumerresource interactions across body sizes. For the 500 parameter sets, I plotted the resulting *m* against body size with a grey line in Figure 1b.

I then assembled a dataset on body size and mutual interference from the literature (see the electronic supplementary material and data in [23]). These data came from studies where foraging or parasitism rates were measured under a range of resource and consumer densities, as required by equation (1.2). The estimates of *m* came from either the original source or were recalculated from data presented in the figures following the approaches in [11, 13]. For this study, I added new data from [17, 24–28] to the datasets in [11, 15] and then searched the original papers and the literature for estimates of body size for each of the focal consumers. I averaged multiple observations for the same consumer. The final dataset included observations for 33 consumers of a variety of taxa including insects (20), arachnids (2), crustaceans (4), birds (2), protists (2), mammal (1), flatworm (1) and rotifer (1).

3. Results

The value of ψ strongly influenced the relationship between interference and body size and controlled whether there was a positive or negative nonlinear relationship or no relationship at all (Figure 1a). Randomly sampling parameters indicates that the model does not predict a systematic variation of interference across a wide range of body sizes (Figure 1b). In other words, any level of interference is possible for any body size given the underlying parameters. The empirical data support this observation. A linear regression of *m* on body size has a non-significant slope of -0.02 (95% CI: -0.04 to 0.01), and the running mean of *m* (window length of 10; heavy dashed line in Figure 1b) followed the overall mean (thin horizontal line) very closely.

4. Discussion

By setting growth and mortality, body-size-dependent parameters determine the abundance, stability and dynamical properties of populations [3, 29, 30]. The results presented here indicate that unlike all other population parameters, mutual



Figure 1. (a) The body-size dependence of mutual interference driven by the scaling exponent relating resource body size to consumer body size (ψ , see text). The other parameters in this simulation were $\gamma = 0.25$, $s_0 = 10$ and $\Delta v_0 = 10$. Similar results were obtained with other parameter combinations.

(b) The body-size dependence of mutual interference across a wide range of taxa, with parameters for Equation (2.3) drawn randomly from typical ranges (grey lines, see Material and methods). Both model and data indicate no systematic effect of mass on interference. The overall (thin horizontal line) and running mean (heavy dashed line) of the observed levels of interference are shown.

interference appears broadly independent of body size (figure 1*b*). Under certain conditions, there may be a small effect; for example, four orders of magnitude variation in terrestrial mammalian predator body size would change interference from about –0.75 to –1.25, assuming ψ = 1.5 [3]. By contrast, this change produces three orders of magnitude of change in area of capture [3].

Interference is generally a stabilizing force in populations because it decreases interaction strengths [14]. If interference were tied to body size, then population stability would be as well. Instead, most species show intermediate levels of interference, with a mean of approximately –0.7. Thus, the effect of body size on populations is more pronounced through parameters other than interference, while interference applies to about the same degree across a wide size range. There appears to be a benefit to these intermediate levels, as no interference allows large swings in population sizes, while severe interference, because of its association with high levels of consumerresource engagement, tends to push populations deterministically towards extinction [15].

Equation (2.2) can make testable predictions about interference from knowledge of consumer and resource velocities. Such predictions may apply to any given system as well as the broad effects of environmental factors such as temperature [17, 20]. For example, if warming accelerates the velocities of the consumers more than their resources, interference should increase, and vice versa. Indeed, in the case of two ground beetles, interference levels went up for one species and down for another species with temperature [17]. Such effects could be predicted from Equation (2.2), although in the case of the beetles it is not known whether velocity changes could account for the observations.

With a mechanistic model linking body mass and temperature to a functional response with interference in hand, we can more thoroughly investigate how population properties respond to environmental change. Yet the vast majority of functional response studies have measured foraging rates of only one individual consumer, and as a result levels of interference are mostly unknown. More work is needed to understand how factors like body mass, predation mode and temperature influence interference, as independent of body mass, it has potent effects on populations and the communities in which they reside.

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Supplementary Material

DeLong, J.P., "The body size dependence of mutual interference"

Data

This ESM describes briefly how I found data and estimated values of interference and body mass. See Dryad entry for more details and raw data. The data set was built upon a previous data set on mutual interference [1]. I conducted searches on Google scholar for additional works on mutual interference. I searched each available source for an estimate of the body size of the focal consumer or parasitoid. Body sizes were generally not available, so other sources were sought. These often included websites reporting on biocontrol agents or the natural history of certain organisms, as well as publications that had previously compiled body size estimates for a wide range of species (e.g. [2,3]), or related publications by the same authors [4]. In one case a personal communication was used [5]. Body sizes were given in lengths, widths, dry masses, wet masses, or volumes, and all were converted to wet mass (g). Body sizes given in length for insects or arachnids were converted to dry mass using the length-weight relationship from [6] for insects (dry mass (mg) = 0.0266 length (mm) ^2.494) and then converted to wet mass assuming water content of 62% [7]. Body widths given for crabs was converted to wet mass given the carapace-weight relationship in [8].

Approaches 2, 3, and 4 described in [1] were used. Wherever possible, original estimates of the mutual interference parameter *m* were used from the original source. In several cases, new values of *m* were calculated in an earlier compendium and used here [9]. When necessary, data were digitized and values of *m* were estimated using non-linear least squares regression for the equation $f = \frac{\alpha C^m R}{1 + \alpha C^m h R}$, following methods reported in [1]. Confidence intervals were available for some estimates, including the recalculations, but authors variably reported standard errors, ranges across replicates, or no error.

Original Publication	Source of estimate	m	Error (SE, CI, range)	Species	Туре	Wet mass (g)	Source for mass
[10]	Calculated in original source	-0.76	-	Aphidius matricariae	Insect	6.88E-04	www.evergreengrowers.com
[11]	Recalculated by [9]	-1.05	± 0.36 (SE)	Daphnia pulex	Crustacean	4.62E-04	[3]
[12]	Recalculated by [9]	-0.50	± 0.09 (SE)	Amblyseius degenerans	Insect	2.88E-05	www.Biotech-system.com

[13]	Recalculated by [9]	-1.14	± 0.15 (SE)	Nasonia vitripennis	Insect	3.94E-04	http://www.bios.niu.edu/bking/nasonia.htm
[14]	Recalculated by [9]	-0.83	± 0.09 (SE)	Tribolium castaneum	Insect	1.08E-03	http://entnemdept.ifas.ufl.edu/creatures/
[13]	Recalculated by [9]	-0.89	± 0.07 (SE)	Trichogramma evanescens	Insect	1.24E-05	http://usagardener.com/disease_pests_and weeds/garden_pests_and_control.php
[15]	Recalculated by [9]	-0.33	± 0.14 (SE)	Trichogramma pretiosum	Insect	1.24E-05	http://usagardener.com/disease_pests_and weeds/garden_pests_and_control.php
[12]	Recalculated by [9]	-0.92	± 0.16 (SE)	Phytoseiulus persimilis	Arachnid	1.24E-05	http://www.biocontrol.entomology.cornell.e du/index.php
[16]	Calculated by Skalski and Gilliam 2001	-0.33	-0.20 to - 0.43 (Cl)	Back swimmer	Insect	1.20E-02	Wikipedia entry for 'Notonectidae'
[17]	Calculated in original source	-0.63	-0.59 to - 0.66	Bracon hebetor	Insect	2.50E-03	[18]
[17]	Calculated in original source	-0.45	-0.41 to - 0.49	Bracon hebetor	Insect	2.50E-03	[18]
[5]	Calculated in original source	-0.67	± 0.11 (SE)	Stenostomum virginianum	Flatworm	2.21E-05	P. Kratina, pers. communication
[19]	Calculated in original source	-0.50	-0.40 to - 0.61 (Cl)	Polistes dominulus	Insect	1.23E-01	http://www.cirrusimage.com/Bees_wasp_p olistes.htm
[20]	Calculated in original source	-0.32	-0.26 to - 0.38 (Cl)	Polistes dominulus	Insect	1.23E-01	http://www.cirrusimage.com/Bees_wasp_p olistes.htm
[21]	Calculated in original source	-1.00	-0.58 to - 1.43 (Cl)	Thanasimus dubius	Insect	3.00E-01	http://bugguide.net/node/view/33027
[22]	Calculated in original source	-0.35	-0.06 to - 0.78	Anisops bouvieri	Insect	6.68E-03	Reported in original paper
[22]	Calculated in original source	-0.22	-0.09 to - 0.35	Diplonychus annulatus	Insect	1.61E-01	Reported in original paper
[22]	Calculated in original source	-0.24	-0.01 to - 0.35	Diplonychus rusticus	Insect	6.33E-02	Reported in original paper
[23]	Calculated in original source	-1.06	-2.1 to - 0.02	Didinium nasutum	Protist	7.35E-07	Reported in original paper
[24]	Calculated in	-1.85	-2.17 to -	Canis lupus	Mammal	4.60E+04	[2]

	original source,		1.53 (CI)				
	раск-scale						
[25]	Recalculated from	-0.65	-1.1/to -	Callinectes sapidus	Crustacean	1.42E+02	Carapace width in original paper
	data in Figure 1		0.12 (CI)				
[26]	Recalculated from	-0.85	-1.04 to -	Daphnia pulex	Crustacean	4.62E-04	[3]
[20]	data in Figure 1		0.66 (CI)				[3]
[27]	Recalculated from	-1.25	-1.48 to -	Trichogramma	Insect	1.24E-05	http://usagardener.com/disease_pests_and
	data in Figure 1a,b		1.02 (CI)	minutum			<u>_weeds/garden_pests_and_control.php</u>
[20]	Recalculated from	4 27	-1.04 to -	Tetragoneuria	luce est	1.32E-03	Dry mass approximate given ~ 4x size
[28]	data in Figure 2	-1.27	1.50 (CI)	<i>cynosura</i> larvae	insect		difference between tc1 and tc2
[20]	Recalculated from	0.00	-0.19 to	Brachionus	D 111	4 5 4 5 9 6	[0]
[29]	data in Figure 2	-0.02	0.15 (CI)	calyciflorus	Rotifer	1.54E-06	[3]
	Recalculated from		-0.22 to -		Bird		
[30]	data in Figure 4	-0.63	1.04 (CI)	Arenaria interpres		1.37E+02	http://www.allaboutbirds.org
	Recalculated from	-0.55	-0.43 to -	Calidris canutus	Bird	1.35E+02	
[30]	data in Figure 4		0.66 (CI)				http://www.allaboutbirds.org
	Recalculated from	-0.63	-0.28 to -	Ananteles (Cotesia)	Insect	8.97E-03	http://www.biocontrol.entomology.cornell.e
[31]	data in Table 1		0.20 (0)	alomeratus			du/index.nbn
	Recalculated from		-1.99 to -	Pteromalus			http://www.entomology.wisc.edu/mbcn/kyf
[31]	data in Table 1	-1.99	-1.99 (0 -	Pleiomulus	Insect	1.59E-03	212 html
			2.00 (CI)	μαρατατη			<u>512.II(III)</u>
[32]		-1.18	-1.2 10 -	Pardosa milvina	Arachnid	1.13E-02	[4]
	original source		1.16				
[33]	Recalculated from	-0.70	-0.47 to -	Canis lupus	Mammal	4.60E+04	[2]
	data in Figure 1		0.94 (CI)	,			
[34]	Recalculated from	-0.79	-1.14 to -	Poecilus versicolor	Insect	6.10E-02	Reported in original paper
[34]	data in Figure 1		0.45 (CI)				
[34]	Recalculated from	-0.78	-1.10 to -	Poecilus versicolor	Insect	6.10E-02	Reported in original paper
	data in Figure 1		0.45 (CI)				
[34]	Recalculated from	-0.71	-1.04 to -	Pterostichus melanarius	Insect	1.43E-01	Reported in original paper
	data in Figure 1		0.39 (CI)				
[34]	Recalculated from	-0.93	-1.21 to -	Pterostichus	Incost	1.43E-01	Departed in original paper
	data in Figure 1		0.65 (CI)	melanarius	insect		Reported in original paper
[05]	Recalculated from	0.50	-0.85 to -		Crustacean 2.34		
[35]	data in Figure 2	-0.52	0.18 (CI)	Mysis mixta		2.34E-02	Reported in original paper

[36]	Recalculated from data in Figure 3	-2.83	-7.54 to 1.88 (CI)	Calidris canutus	Bird	1.35E+02	http://www.allaboutbirds.org
[37]	Recalculated from data in Figure 3b	-0.42	-2.84 to 2.00 (CI)	Anagrus delicatus	Insect	2.88E-05	Reported in original paper
[38]	Recalculated from data in Figure 7	0.00	-0.16 to 0.15	Woodruffia metabolica	Protist	1.18E-07	Assume similar in cell volume to <i>Woodruffia</i> rostrata, size from EOL (<u>http://eol.org/</u>)
[39]	Recalculated from data in Figure 7b	-1.60	-4.06 to 0.86 (CI)	Anagrus delicatus	Insect	2.88E-05	[37]
[40]	Recalculated from data in Figure 2	-1.00	-1.23 to - 0.80 (Cl)	Carcinus aestuarii	Crustacean	1.32E+01	Carapace width in original paper

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