

1 Drought effects in dock

2 **Drought Stress alters Solute Allocation in Broadleaf Dock (*Rumex obtusifolius*)**

3

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5 According to climate models, drier summers must be expected more frequently in
6 Central Europe during the next decades which may influence plant performance and
7 competition in grassland. The overall source-sink relations in plants, especially allocation of
8 solutes to above- and below-ground parts, may be affected by drought. To investigate solute
9 export from a given leaf of broadleaf dock, a solution containing ^{57}Co and ^{65}Zn was
10 introduced through a leaf flap. The export from this leaf was detected by analysing
11 radionuclide contents in various plant parts. Less label was allocated to new leaves and more
12 to roots under drought. The observed alterations of source-sink relations in broadleaf dock
13 were reversible during a subsequent short period of re-watering. These findings suggest an
14 increased resource allocation to roots under drought improving the functionality of the plants.

15 **Nomenclature:** Broadleaf dock, *Rumex obtusifolius* L. RUMOB.

16 **Key words:** Grassland, climate change, water limitation, recovery, phloem transport.

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17 The current increase in atmospheric CO₂ concentration leads to a number of changes
18 in climate (Meehl et al. 2007). One of the changes projected by climate models is a decrease
19 in summer precipitation and, in general, an increasing frequency of summer droughts in
20 Central Europe (Christensen et al. 2007). Reduced water availability can decrease plant
21 biomass production considerably (Ciais et al. 2005; Peñuelas et al. 2007). As a consequence,
22 the agricultural sector will be affected by increasing drought frequencies in the future (Brown
23 et al. 2011; Fuhrer et al. 2006). Along with yield reductions, changes in climate often lead to
24 changes in the competition between species, e.g. between crops and weeds (McDonald et al.
25 2009; Patterson 1995a, 1995b).

26 One of the most troublesome weeds in Europe (Doyle et al. 1984; Gebhardt et al.
27 2006; Zaller 2004b), broadleaf dock (*Rumex obtusifolius* L.), was found to be less sensitive to
28 drought than the other species (i.e. fodder plants) in intensively managed temperate grassland
29 (Gilgen et al. 2010). Although this phenomenon might be limited to the more humid regions
30 of Europe (i.e. western Central Europe), any increase in competitive ability of broadleaf dock
31 due to climate change will be problematic for farmers. Broadleaf dock is a strong competitor
32 for light and space (fast growth of big leaves) as well as nutrients and water. Roots may grow
33 as deep as 2.5 m (Kutschera et al. 1992). This weed reduces both the quantity (Iijima and
34 Kurokawa 1999; Oswald and Haggart 1983) and the quality (Nashiki et al. 1991) of yield.
35 Since the control of broadleaf dock is very difficult and laborious (see Strnad et al. (2010) for
36 a summary of available methods) an increase in the abundance would cause additional costs
37 for weed management.

38 Better understanding of the physiological mechanisms behind the observed increase in
39 competitive ability of broadleaf dock against surrounding grassland species under drought
40 would be a prerequisite to an adaptation of management or mitigation of drought effects. It
41 was suggested that broadleaf dock benefits from its deep roots under drought (Gilgen et al.

42 2010). In competition with grassland species, broadleaf dock was found to invest into root
43 biomass, thereby building the base for the species' success in temperate grassland (Zaller
44 2004a). An efficient supply of carbohydrates to roots of broadleaf dock especially before
45 flowering has repeatedly been detected (Imhoff and Voigtländer 1979; Lang et al. 1975;
46 Voigtländer et al. 1976). However, potential effects of drought stress on these allocation
47 patterns have not yet been studied. In a study of drought and heat effects on temperate
48 grassland forbs it was shown that root growth shapes the community's response (Dreesen et
49 al. 2012). The allocation of resources to above- and below-ground could thus be a key to
50 understand plant responses to drought. A change in source-sink relations and as a
51 consequence the reallocation of leaf-borne solutes via the phloem to the roots could improve
52 the performance of broadleaf dock under drought.

53 A suitable technique is needed to investigate the export of solutes from mature leaves
54 to sinks (e.g. roots and growing shoot parts). Radiolabelled heavy metals which are not
55 metabolised may be helpful in this context. Such isotopes were originally used to study the
56 phloem and xylem mobility of heavy metals (Page and Feller 2005; Riesen and Feller 2005;
57 Zeller and Feller 1998). Now that the mobility of the different heavy metals is known, their
58 radionuclides can be used to track phloem and xylem transport of plants. In contrast to
59 organic compounds, heavy metals are not metabolised and not released from plants as gaseous
60 compounds. The radionuclides ^{57}Co and ^{65}Zn can be detected simultaneously in a sensitive
61 manner and are therefore suitable for long-distance translocation studies (Page and Feller
62 2005; Riesen and Feller 2005). Due to source-sink dynamics we know that a radioactive label
63 fed to a fully expanded leaf can be transported to younger leaves or roots via the phloem (as
64 they both are phloem sinks and need resources like photosynthates or other solutes). From the
65 roots, the solutes (including the radioactive labels) can then be transported to other plant parts
66 with the transpiration stream in the xylem. All label found in older leaves has to be

67 transported there via xylem as older leaves are phloem sources and not sinks. On the other
68 hand, the redistribution of solutes from fully expanded leaves to growing shoot parts or roots
69 depends on the symplastic transport via the phloem. Good mobility in the phloem was
70 reported for Co and Zn in gramineae (Riesen and Feller 2005) as well as in dicots (Page et al.
71 2006). Thus, the distribution of radioactive Co and Zn in plants offers an insight into the
72 allocation of resources to the different plant parts. Changes in long-distance transport and in
73 solute allocation caused by drought are reflected in an altered distribution of Co and Zn after
74 labelling a defined leaf with the radionuclides.

75 To better understand the mechanisms involved in the previously observed rather high
76 biomass of broadleaf dock in grassland under drier conditions (Gilgen et al. 2010), a labelling
77 experiment to track solute transport (i.e. allocation of solutes via the phloem) in this weed was
78 designed. The aim of this study was to understand how the transport of solutes is affected by
79 drought. We hypothesised that the amount of label transported to the roots would increase
80 under drought, as plants would invest more resources into roots to maintain their basic
81 functions.

82

83 **Materials and Methods**

84 Seeds of broadleaf dock (*Rumex obtusifolius* ssp. *obtusifolius*; originating from the
85 region of Bern, Switzerland) were germinated on coarse quartz sand and grown on deionised
86 water first and later on a standard nutrient solution (according to Page et al. (2012)). At the
87 age of two months, 24 plants were each transferred to a 0.8 L pot with a soil mixture
88 containing 45% Landerde (nutrient rich soil washed off sugar beet grown on the Swiss
89 Plateau), 36% turf, 18% sand and some Seramis clay granules. The pots were randomly
90 assigned to the control or drought treatment before the start of the experiment. Soil water

91 potential sensors (Watermark soil moisture sensor, Irrrometer Company, Inc., Riverside, CA,
92 USA) were placed at the bottom of each drought pot and four of the 12 control pots. The pots
93 were arranged on two shelves in a climate cabinet and positions were randomly rotated every
94 week. The cabinet was set to a 14 h day at 24°C and a 10 h night at 16°C. Light was supplied
95 with 55 W lamps and adjusted to a level of around 100 to 120 $\mu\text{mol m}^{-2} \text{s}^{-1}$ at leaf level.
96 Deionised water was supplied regularly to keep the pots well saturated. The
97 evapotranspiration (i.e. the water loss) of every pot was assessed gravimetrically by weighing
98 the pots before and after watering (every second or third day).

99 The labelling solution containing the radionuclides ^{57}Co and ^{65}Zn was introduced into
100 the leaf via a flap in the petiole. The method described by Schenk and Feller (1990) was
101 adapted to the different leaf morphology of dicots. A test prior to the experiment had shown
102 that cutting the petiole longitudinally in the middle through the symmetry axis and using one
103 of the two equal parts as the flap resulted in the best uptake of liquids. After 46 days of
104 growth on soil, an approximately 4 cm long flap was cut into the petiole of the youngest fully
105 expanded leaf (in general the 10th or 11th leaf). This flap was positioned in a tube containing
106 0.8 ml of the radionuclides ^{57}Co and ^{65}Zn dissolved in 10 mM RbCl and 10 mM SrCl₂. If
107 necessary, the flap was repositioned in the remaining liquid after 48 h. All except two (one
108 drought and one control plant) of the 24 plants took the solution up completely. After
109 approximately 96 h, the tubes were recovered for later verification of label uptake (see
110 below). All plants were watered before the label was applied to make sure that the uptake of
111 label was not confounded by the treatment. Following that, the 12 drought plants did not
112 receive water any longer while the 12 control plants were still watered as before.

113 Seven days after the application of the label and the last watering of the drought
114 plants, four randomly chosen plants from both treatments (well watered control vs. drought)
115 were harvested. Of the remaining eight drought plants, four were kept at drought conditions

116 while the other four were re-watered. The watering of the eight control plants remained
117 unchanged. Four plants from each of these three groups (control, drought, re-watered) were
118 harvested after another seven days. Four control plants were used for evapotranspiration
119 measurements only but were not analysed further.

120 The labelled petiole and leaf blade were sampled separately while the other leaves
121 were sampled as a whole. All dead leaves were pooled in one sample as were the side shoots.
122 Roots were washed from the soil and also sampled. For practical reasons, the base (lowest
123 part of leaves and uppermost part of roots) was sampled separately. Dry weight (after drying
124 at 60°C for 24 h) of the different samples was measured.

125 The dried plant samples as well as the recovered labelling tubes and a tube containing
126 0.8 ml of the labelling solution (i.e. a reference tube) were analysed with an automatic gamma
127 counter (1480 Wizard 3'', Wallac, Turku, Finland) recording gamma radiation emitted by
128 ⁵⁷Co and ⁶⁵Zn at the same time. Counting duration was set to 60 min and results are expressed
129 as counts per minute (cpm) per sample.

130 To ensure that no contamination with the label had occurred, Sr content in the
131 different plant parts (as described above) was also assessed. Sr is immobile in the phloem and
132 should therefore only be found in the labelled leaf. Sr content was measured by atomic
133 absorption spectrometry (SpectrAA 220FS, Varian Techtron, Mulgrave, Australia). Once
134 gamma counting was finished, samples were ashed at 550°C for several hours. After cooling
135 0.2 ml 10 N HCl and subsequently 2 ml deionised water were added to the ash. An adequate
136 dilution with 5000 ppm LaCl₃ in 0.1 N HCl was used to assure all samples fit the
137 measurement range of the instrument (0-8 ppm). To quantify the background content of Sr
138 originating from the soil, an additional set of six control plants was grown under the same
139 conditions and analysed for Sr content as well.

140 The effect of the treatment on evapotranspiration and soil water content was tested
141 using one-way ANOVA. For statistical analysis, leaf samples were pooled in groups: labelled
142 plant parts (labelled petiole and leaf blade), leaves that were older than the labelled leaf
143 (including dead leaves; in the following called “old leaves”), leaves that were younger than
144 the labelled leaf but present and not fully expanded at the time of labelling (“young leaves”),
145 leaves that emerged after labelling (“new leaves”), leaves from additional shoots (“side
146 shoots”) and below-ground parts (roots and base, in the following referred to as “roots”). In
147 general, four independent replicates in a fully randomised design were analysed for each
148 treatment and each harvest date. However, one of the drought replicates was excluded from
149 all analyses on harvested plants as the plant only took up approximately 75% of the labelling
150 solution. The effect of the treatment was tested for the two harvests separately using ANOVA.
151 The different treatments within the respective harvest were compared using a LSD test. All
152 statistical analyses were performed with R 2.14.2 (R Development Core Team 2012).

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154

Results and Discussion

155 Withholding water significantly reduced evapotranspiration within four days (Fig. 1).
156 While control plants lost $65.03 \text{ g H}_2\text{O d}^{-1}$ ($\pm 3.65 \text{ g H}_2\text{O d}^{-1}$; mean \pm SE, n=12), drought stressed
157 plants lost seven times less water ($9.31 \pm 0.72 \text{ g H}_2\text{O d}^{-1}$) at the end of phase 1.
158 Evapotranspiration significantly increased in response to re-watering. However,
159 evapotranspiration of re-watered pots did not reach the level of control pots after one week.
160 Evapotranspiration of drought pots further decreased during phase 2 and reached almost zero
161 at the end of the experiment. Soil water potential progressively declined during phase 1. In
162 contrast to evapotranspiration, soil water potential immediately recovered once pots were re-
163 watered (Fig. 1).

164 Total dry weight was not significantly affected by one week of drought (3.45 ± 0.23 g
165 in drought plants compared to 3.73 ± 0.44 g in control plants, mean \pm SE, n=4; p=0.60). Only
166 the dry weight of new leaves was significantly reduced by drought at the first harvest. This
167 was also reflected in the relative contributions of the different plant parts to the total plant
168 biomass (Fig. 2).

169 At the second harvest, total dry weight of drought stressed and re-watered plants was
170 similar (3.55 ± 0.45 g and 3.35 ± 0.13 g, respectively; mean \pm SE, n=3-4) but significantly lower
171 than total dry weight of control plants (4.84 ± 0.24 g; mean \pm SE, n=4). Control plants had been
172 able to increase their biomass (i.e. their total dry weight) during phase 2, while total dry
173 weight of drought stressed and re-watered plants remained constant. The lower total dry
174 weight of drought stressed and re-watered plants was mainly caused by a decrease in root dry
175 weight and a significantly decreased dry weight of new leaves in drought plants and old
176 leaves in re-watered plants, respectively (Fig. 2). Thus, while one week of re-watering was
177 not enough for root biomass to recover, it allowed growth of new leaves and the accumulation
178 of similar dry weights in new leaves as in control plants. The production of new leaves
179 indicates that one week of drought stress did not irreversibly damage the plants. This
180 enormous potential of broadleaf dock to withstand unfavourable conditions and recover
181 quickly is clearly underlined by the relative contributions of the different plant parts to total
182 dry weight (Fig. 2). There was no difference between control and re-watered plants in the
183 relative biomass invested in any of the plant parts. In contrast, drought stressed plants
184 attributed significantly less biomass to new leaves while significantly more biomass was
185 concentrated in old leaves (that were mostly dead) compared to plants from the other two
186 groups. However, it is remarkable that the proportion of root dry weight was unaffected by
187 the treatment (Fig. 2).

188 The labelling solution (with 855 cpm ^{57}Co and 257 cpm ^{65}Zn fed to each plant) was
189 almost completely taken up (largest rest remaining in the tube <10% for both ^{57}Co and ^{65}Zn).
190 The phloem-immobile Sr was detected in the labelled leaf, but was not above the background
191 in the other plant parts, indicating that no contaminations had occurred (data not shown). An
192 average label of 840 ± 9 cpm ^{57}Co and 265 ± 4 cpm ^{65}Zn (mean \pm SE, n=19) was recorded in the
193 plants. A significant treatment effect was only observed in phase 2 when significantly lower
194 activities of ^{57}Co and ^{65}Zn were detected in drought stressed plants. Most of the label was
195 retained in the labelled leaf and only between 5.5% and 9.4% of the ^{57}Co and between 5.5%
196 and 14.1% of the more mobile ^{65}Zn were exported to other plant parts. However, the activity
197 transported (on average 54 ± 6 cpm and 21 ± 3 cpm for ^{57}Co and ^{65}Zn , respectively; mean \pm SE,
198 n=19) was still high enough to analyse the allocation to roots and other shoot parts. There
199 were no significant treatment effects on absolute or relative amounts of activity transported,
200 except for ^{65}Zn in the second harvest. Significantly more ^{65}Zn was transported out of the
201 labelled leaf of re-watered plants compared to drought stressed plants. A similar, but not
202 significant, trend was observed for ^{57}Co (data not shown).

203 A high fraction of the transported label was detected in new leaves and roots, where
204 also the main treatment effects were observed (Fig. 3). The fraction of label transported to
205 new leaves tended to decrease in response to drought, while higher levels of ^{57}Co and ^{65}Zn
206 were detected in the roots of these plants. In re-watered plants, the fraction of label in new
207 leaves and roots became again very similar to control plants indicating a reorganisation of the
208 source-sink relations during the recovery phase. The fraction of label transported to the roots
209 was higher in drought stressed compared to control and re-watered plants even though the
210 root biomass of drought stressed plants was lower. No difference between treatments at any of
211 the two harvests was found in the other three plant parts (young leaves, old leaves and side
212 shoots; data not shown). Zaller (2004a) suggested that the success of docks under competitive

213 conditions is caused by their ability to invest into roots. Our observation that in broadleaf
214 dock more phloem-mobile solutes were directed to the roots under drought are consistent with
215 this concept. The additional resources directed to roots were obviously not used to grow new
216 root biomass (root dry matter decreased under drought) but for maintenance (relative
217 contribution of root dry matter to total biomass was constant) and as storage for later regrowth
218 (resources allocated from roots to new leaves under re-watering).

219 From the experiment reported here it became evident that source-sink relations, and as
220 a consequence solute allocations, in broadleaf dock are strongly affected by a drought period.
221 However, the changes can be reversed quite rapidly during a subsequent recovery phase.
222 Modifications in the redistribution pattern during a drought period as well as during a
223 subsequent recovery phase may improve the overall performance of a species. It must be
224 borne in mind that the persistence of broadleaf dock is controlled by many different factors
225 like water but also nutrient availability (Hann et al. 2012; Hejzman et al. 2012; Humphreys et
226 al. 1999; Křišťálová et al. 2011) or management (Hopkins and Johnson 2002; Martinkova et
227 al. 2009). Possible interactions of these factors remain to be investigated in the future. In a
228 community, the persistence of broadleaf dock will also depend on the presence and identity of
229 neighbours and their behaviour in competition. To date, no other species were studied using
230 the method presented here. Results from experiments solely focusing on above- and below-
231 ground biomass responses to drought are contradictory, showing either a decrease (e.g.
232 Weisshuhn et al. 2011) or an increase (e.g. Dreesen et al. 2012) in the share of biomass
233 invested into roots under drought. Nevertheless, the ability to recover quickly after drought
234 might improve the performance of broadleaf dock under future drier conditions, at least in the
235 more humid regions of Central Europe.

236 From a more general point of view, it can be concluded from the findings reported
237 here that analysing the transport of radioisotopes of phloem-mobile heavy metals (e.g. ⁵⁷Co

238 and ^{65}Zn) after their introduction into a defined leaf is a sensitive and suitable technique to
239 detect changes in the source-sink network. Questions related to impacts of abiotic stresses on
240 the whole plant level (e.g. in the context of climate change) can be addressed with this
241 method.

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243

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246

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359

Figures Legends

360 Figure 1. Average daily evapotranspiration and soil water potential of pots with a broadleaf
361 dock plant. In phase 1 water was withheld from pots of the drought and re-watered treatments.
362 In phase 2 only drought plants were not irrigated. Averages and standard errors are shown
363 (n=4-12).

364 Figure 2. Absolute and relative dry weights of plant parts (excluding the labelled leaf) at the
365 two harvests (i.e. at the end of phase 1 and phase 2). Old leaves were older than the labelled
366 leaf, young leaves were younger than the labelled leaf but already present at the time of
367 labelling, new leaves were formed after labelling and side leaves are leaves from additional
368 shoots (all age classes). Averages and standard errors are presented (n=3-4). Within each
369 harvest significant treatment differences are shown by different letters.

370 Figure 3. Relative amounts of ^{57}Co and ^{65}Zn transported into new leaves and roots expressed
371 in % of the transported label. New leaves were formed after labelling. Averages and standard
372 errors are presented (n=3-4). Within each harvest significant treatment differences are shown
373 by different letters.

Figure 1

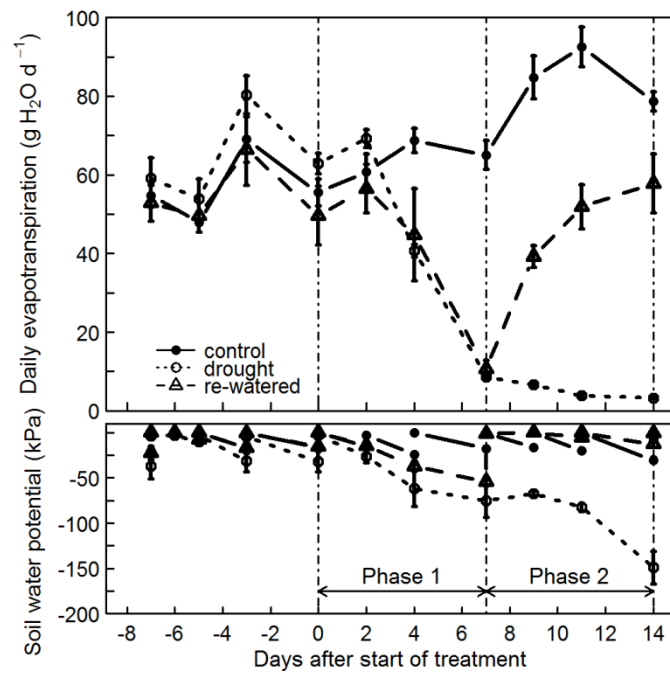


Figure 2

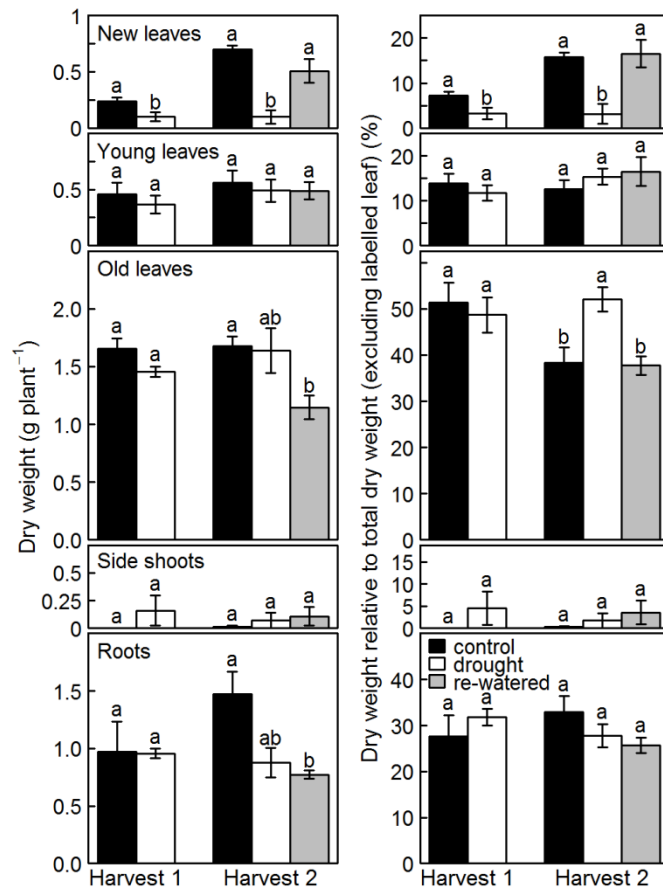


Figure 3

