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Number 9

MANUAL OF RESEARCH METHODS FOR MARINE INVERTEBRATE REPRODUCTION

Issued on the occasion of the Workshop on
MARINE INVERTEBRATE REPRODUCTION
jointly organised by
the Department of Zoology, University of Madras and
the Centre of Advanced Studies in Mariculture,
Central Marine Fisheries Research Institute, Cochin
held at the University of Madras
from 25th October to 10th November 1982

The Centre of Advanced Studies in Mariculture was started in 1979 at the Central Marine Fisheries Research Institute, Cochin. This is one of the Sub-projects of the ICAR/UNDP project on 'Post-graduate agricultural education and research'. The main objective of the CAS in Mariculture is to catalyse research and education in mariculture which forms a definite means and prospective sector to augment fish production of the country. The main functions of the Centre are to :

—provide adequate facilities to carry out research of excellence in mariculture/coastal aquaculture ;

—improve the quality of post-graduate education in mariculture ;

—make available the modern facilities, equipments and the literature ;

—enhance the competance of professional staff ;

—develop linkages between the Centre and other Institutions in the country and overseas ;

—undertake collaboration programmes ; and

—organise seminars and workshops.

Under the programmes of the Centre, post-graduate courses leading to M.Sc. (Mariculture) and Ph.D. are offered in collaboration with the University of Cochin since 1980.

Front cover : SEM picture showing surface topography of *Streptocephalus dichotomus* egg.

Manual of Research Methods for Marine Invertebrate Reproduction

EDITED BY

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ISSUED ON THE OCCASION OF THE WORKSHOP ON MARINE INVERTEBRATE REPRODUCTION JOINTLY ORGANISED BY THE DEPARTMENT OF ZOOLOGY, UNIVERSITY OF MADRAS AND THE CENTRE OF ADVANCED STUDIES IN MARICULTURE, CENTRAL MARINE FISHERIES RESEARCH INSTITUTE HELD AT THE UNIVERSITY OF MADRAS FROM 25TH OCTOBER TO 10TH NOVEMBER, 1982.

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PREFACE

The technologies of controlled reproduction, induction of spawning, sex reversal, artificial fertilisation, sterilisation and preservation of gametes are increasingly applied in aquaculture to obtain quality seed, quality fish stock and better yield. In this context, researches on different aspects of reproduction, developmental biology and physiology have assumed considerable importance besides their values in understanding of the ontogeny of the organisms. Extensive researches carried out in recent years from several laboratories in the world have not only accumulated a body of information, but also broughtforth several new concepts to our understanding of the development and reproductive behaviour of finfishes and shellfishes.

In India, directed research on reproductive physiology and biology is taken up only recently and the field is still in an infant stage. In view of its emerging importance, it is identified as an area for priority research and for expertise development in the programmes of the Centre of Advanced Studies in Mariculture at the Central Marine Fisheries Research Institute, and several programmes of research are being taken up in this field with particular reference to the reproductive behaviour of the cultivable finfishes and shellfishes.

Advances made on the frontiers of invertebrate reproduction in recent years have been significant enough to organise a national workshop and to prepare a manual on research methodologies for the study of the subject. Several histological, histochemical and biochemical methods and sophisticated instruments have been introduced in these studies making it essential that the scholars who desire to work and specialise in the field are given adequate basic information on the research methods so as to enable them to appreciate and advance research to understand the problems confronted in the field.

The present manual, the third in the series, is prepared and compiled by Dr. T. Subramoniam, Leader of the ' Unit of

Invertebrate Reproduction' of the Zoology Department of the University of Madras, Tamil Nadu. During the past decade, a team of research scholars are working on different aspects of marine invertebrate reproduction including the cultivable crustaceans such as *Scylla serrata*, *Panulirus homarus* and *Macrobrachium* spp. under his leadership. Contributing to our knowledge on the subject, the research results achieved so far in these aspects by the Unit have unfolded several new concepts in oogenesis, spermatogenesis, sperm transfer strategy, fertilization and endocrine control of reproduction and gamete formation.

I wish to express my great appreciation to Dr. T. Subramoniam and his team of Scholars, who by their dedication and interest evolved a series of tested research methods and set a theme of investigation through insight and skill on marine invertebrate reproduction. I am sure that this manual will be of immense use to the research scholars and scientists who would like to specialise in the subject and cognate fields.

This is the second workshop we are organising in close collaboration with the University of Madras. I wish to express my gratitude to Dr. M. Santappa, Vice-Chancellor, University of Madras for the keen interest evinced in such collaborative programmes and for the advice. I am also indebted to Dr. K. Ramalingam, Professor and Head of the Department of Zoology, University of Madras for productive discussions, continuous support and suggestions. I wish to thank Shri P. T. Meenakshisundaram and Shri K. Rengarajan, Scientists of the Central Marine Fisheries Research Institute for their help in the preparation of this manual.

E. G. SILAS,
Director, C.M.F.R.I.

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EXPERIMENTS ON YOLK PROTEIN UPTAKE IN CRUSTACEAN OVARY*

5.1. INTRODUCTION

Recent electron microscopic and biochemical investigations have revealed the pinocytotic uptake of extra ovarian proteins into the vitellogenic oocytes (Hinsch and Cone, 1969 ; Wolin *et al.*, 1973). The uptake of these macromolecular yolk precursor substances could be demonstrated using trypan blue as well as horse radish peroxidase.

5.2. MATERIALS

Ovaries of *Scylla serrata* and *Emerita asiatica* in different stages of maturation.

5.3. TRYPAN BLUE METHOD

5.3.1. Principle

Trypan blue dye used as the indicator in the present experiment mimics the physical properties of proteins and the dye molecules are too big to enter living cells except by pinocytosis. Hence, incorporation of trypan blue into the oocyte indicates its ability to sequester extra ovarian macromolecular proteins.

5.3.2. Procedure

1. Dialyse trypan blue against water.
2. Prepare 1% trypan blue solution in saline.
3. Dissect out ovareis of *S. serrata* and *E. asiatica* in different stages of ovarian maturation under sterile conditions.

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* Prepared and verified by S. Ezhilarasi,Unit of Invertebrate Reproduction, Department of Zoology, University of Madras, Madras-600 005.

4. Wash in saline to remove the adhering haemolymph.
5. Incubate the ovaries in 1% trypan blue solution for 1 hour.
6. Wash the ovaries in four changes of saline for 1 hour.
7. Prepare the ovaries for cryocut sectioning.
8. Examine the sections of the ovaries under microscope.
9. Observe and differentiate the rate of micropinocytosis of trypan blue in different stages of ovarian maturation.

5.4. Horse Radish Peroxidase Method

5.4.1. Principle

Horse radish peroxidase is an enzyme whose presence can be detected histochemically using hydrogen peroxide and benzidine. When a solution of peroxidase is injected into the haemolymph of crustaceans these macromolecules would be sequestered by a tissue or cell involved in pinocytosis. The tissue, after appropriate incubation, can be processed for benzidine reaction. Since the vitellogenic oocytes are known to sequester the yolk precursors during vitellogenesis the peroxidase method could be used to demonstrate the uptake of protein molecules into the oocytes. This method has been previously used to demonstrate the protein sequestration into the fat body of insects and millipedes (Lock and Collins, 1968 ; Subramoniam, 1971).

5.4.2. Procedure

1. Prepare 1% solution of horse radish peroxidase in normal saline.
2. Inject 1 ml of horse radish peroxidase solution into the crab.
3. After 4 hours dissect out the ovaries and fix in cold 4% neutral buffered formalin for 4 hours.
4. Wash in 10% sucrose solution 3 times at an interval of 30 minutes.
5. Wash in phosphate buffer (pH 7).

6. With gentle shaking, add 9 ml of benzidine reagent (dissolve 300 mg of benzidine in phosphate buffer at pH 7).
7. After 2 minutes add 1 ml of hydrogen peroxide and shake vigorously at room temperature for 10-30 minutes.
8. Prepare the tissues for cryocut sections.
9. Observe the dark brown granules in the ooplasm for the presence of peroxidase.

5.5. References

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