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29. FEEDING BEHAVIOUR OF THE GREEN MUSSEL, *PERNA VIRIDIS* (LINN.) IN LABORATORY

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ABSTRACT

The rate of filtration and feeding on six species of diatoms by the green mussel *Perna viridis* Linn in the laboratory has been studied. The number of cells removed per hour depended upon the size and suspension density of the diatom cultures. Generally the mussel was found to eliminate more than 50% of the filtered cells as pseudofaeces. The rate of ingestion was enhanced when the suspension density and cell size were less. Even though large quantities of pseudofaeces were produced when suspension density was increased, the actual ingestion of food was not affected by cell concentration. The maximum filtration rate [341.43] ml hour⁻¹ gm⁻¹ was noted in *Thalassiosira fluviatilis* suspension. The relation between the rate of filtration and cell size and density of the suspension was studied and discussed.

INTRODUCTION

Efficient production of culturable bivalve molluscs in controlled environments require exact information about the rate of feeding and particle transport. Clams and oysters subsist mainly on particles filtered from the surrounding water, which they pump through the gills. An extensive literature deals with the results of microscopical examination of the content of the digestive tract, to estimate the relative significance as food of the various types of organic matter present in bivalves (Allen 1921, Coe 1948 Coe and Fox 1944, Savage 1925, Vervey 1952).

The volume of water from which particles are removed in a given period is termed the filtration rate [Fox et al 1937]. Generally the filtration rate and pumping rate are not same, because the retention of particles by bivalves is rarely 100% efficient [Jorgensen 1955], the pumping rate is usually greater than filtering rate. The ingestion of food by bivalves mainly depend on the filtration rate, but various other factors also control the ingestion rate. Some of the particles filtered by the gills are eaten, while others are rejected as pseudofaeces. The ejection of feces and pseudofaeces are known as biodeposition.

Although there is considerable information concerning the filtration and pumping rates of bivalves, little consensus regarding rates of

feeding or ingestion is available. Under experimental conditions different types of organisms and different mixtures of organisms of various size result in different rates of filtration and possibly of feeding (Loosanoff and Engle 1947; Rice and Smith 1958; Smith 1958; Allen 1962; Davids 1964). Rice and Smith (1958) conducted the major study of *Mercentaria mercenaria*. They supplied the clams with the four species of algae tagged with ¹⁴C in un-replenished suspensions, Allen (1962) studied the filtration rate, ingestion rate and biodeposition of ¹⁴C labelled *Pheodactylum* by four species of bivalves. The total biodeposition of three species of bivalves in relation to suspension density was studied by Haven and Morales-Alamo (1966), and found that the density of suspension determines the rate of filtration, pseudofaeces formation and rate of food ingestion. Walne (1970) investigated the food value of different species of algae in culture. Allen [1970] investigated the filtration rate and biodeposition of *Hiatella arctica* by using two species of micro algae in different cell densities. Tenore and Dunstan [1973] studied the rates of feeding and biodeposition of American oyster by using four species of phytoplankton, viz., *Thalassiosira pseudonana*, *Skeletonema costatum*, *Dunaliella tertiolecta* and *Nitzschia closterium*. Tenore et al [1973] studied the food chain dynamics of the oyster, clam and mussel in an aquaculture food chain.

Epifanio [1976] studied the nutritional requirements of juvenile and adult bivalves. Foster-Smith [1975] investigated the assimilation efficiency of three species of bivalves by using of *Phaeodactylum* sp and found that the assimilation mainly depend upon the rate of ingestion. The rate of removal of four species of algae from suspension by the oyster *Crassostrea virginica* was determined by Epifanio and Ewart [1977] and found that the filtration rate was influenced by the suspension density and cell size of the algae. Malouf and Breese [1977] measured the effects of algal concentration and larval density on the growth of larval oysters in a flow through feeding system.

Salzwedel [1979] quantitatively estimated the production of pseudofeces and feces by *Tellina jabula*. For long term bioaccumulation studies with suspension feeding mussels, Boetter-Jensen and Dahlgard [1981] designed an apparatus to maintain phytoplankton cell concentrations at a constant level. Wilson [1983] studied the retention efficiency and pumping rate of *Ostrea edulis* in suspension of *Isochrysis galbana* in relation to cell concentration. Urban and Pruder [1933] compared the growth of *Crassostrea virginica* at five algal ration levels for a period of 3 weeks. Berry and Schleyer [1983] estimated the assimilation efficiency of *DeA7a* on a natural diet of particles <100µm diameter. Colwell et al (1984) has developed microencapsulation techniques for artificial food to rear bivalve molluscs in recirculating system.

The food value of unialgal diet and mixed algal diets to juvenile oysters were determined, and mixed algal diet proved to be good for juvenile oysters (Romberger and Epifanio 1981-)

MATERIAL AND METHODS

Six species of diatoms (Table 1) were cultured in the lab. condition by using, f/2 medium (Guillard and Ryther 1962) made up in aged and filtered sea water with the salinity adjusted to 28‰ and sterilized by autoclaving. All the six species of diatoms came from clones isolated from Vellar estuary (11°29'N; 79°49'E) and maintained in laboratory. For

the feeding experiments, mass cultures of each species were developed. The cultures were shaken periodically to keep the cells in suspension. Continuous illumination at 4000 lux was supplied by 40 W 'cool white' florescent lamps for 16 h in a day and temperature ranged from 28-30°C. Cell counts were made with Haemocytometer (Neubauer improved double ruling' Fein optic. Made in GDR). Algae used in the experiment were from cultures that had reached the stationary phase to avoid the multiplication during the experimental period.

TABLE - 1. *The algal species used for the experiment and their size in laboratory culture.*

Sl. No.	Species of diatoms used	Cell size (A m)	Suspension density cells/ml (x10*)
1.	<i>Chaetoceros gracilis</i> Schutt non pant	3-5	45 to 500
2.	<i>The lass iosira fluviatilis</i> Hust	8-10	7 to 100
3.	<i>Skeletonema costatum</i> (Grev.) Cl	7-9	10 to 100
4.	<i>Thalassionema nitzschooides</i> Grun	3-5 dia. 15-25 leng,	6 to 55
5.	<i>Chaetoceros didymus</i> Ehr	10-18	3.8 to 25
6.	<i>Streptotheca tamesis</i> Shrubs	22-30	1.8 to 10

The green mussel *Perna viridis* individuals of same age groups [the wet weight of the whole animals ranged from 2.0 g to 3.1 g] were collected from the field, washed free of epifauna and flora and acclimated in the laboratory aquarium tanks. During the acclimation period the animals were fed with cultured diatoms-mixed form. Prior to the experiment, the animals were allowed to starve for about 12 h. On the day of experiment they were removed from the acclimation tank, washed free of adhering debris, blotted free of excess water and weighed. Then the animals were placed in the experimental tank and the algal suspension of known concentration was added. Table 2 shows the cell concentrations of the suspensions used for the experiments. The algal suspension was continuously agitated by mild air bubbles to

TABLE 2. Rate of filtration and feeding of *P. viridis* in different suspension density of algae.

6 Z	« a o o a o o »	ex 0) »r • • 1 =	c o 9 4 — (A ^ ^ — c c E a u « o o ® ,2	c S o - 01 S i 5 E	c o o c o 'A Z c a g j o o 2	M O o a "A c » — S i ^ S £ £	(0 O U £ i i » £ 5 £	c T3 u « O c c » S o < o																
									2.5	3.1	2.2	2.4	2.5	2.8	2.1	2.0	2.0	2.7	2.5	2.0	2.7	2.4	2.0	2.3
1.	<i>C. gracilis</i>	2.5	45	232.2	59.00	202.00	104.50	22.60																
		3.1	75	241.0	64.40	483.90	180.80	11.68																
		2.2	200	250.0	60.00	1040.00	500 00	5.50																
		2.4	500	128.0	58.80	1451.50	640.00	3 66																
2.	<i>T. fluviatilis</i>	2.5	7	278.5	22.50	26,24	19.50	46.16																
		2.8	15	341.4	44.00	101.00	51.10	30.10																
		2.1	40	252.0	53.10	141.80	92.80	27,24																
		2.0	50	181.40	62.60	118.80	90.70	34.51																
		2.0	100	157.50	6500	250.00	157.50	20.63																
3.	<i>S. costatum</i>	2.7	10	155.5	36.30	5.60	15.55	86.54																
		2.5	35	172 0	82.00	68.00	60.20	54.49																
		2.0	60	149 2	81.00	98 80	89 50	45.25																
		2.7	100	148.0	136.00	256 20	148.00	34.68																
4.	<i>T. nitzsctioides</i>	2.4	6	108 0	10 08	5.00	6.48	6702																
		2.0	15	122.5	16.60	7.90	12 25	67.76																
		2.3	25	128,0	26.50	47.10	32.00	36.01																
		2.5	55	92.2	25 00	102.OJ	50.72	1976																
5.	<i>C. didymus</i>	2.1	3.8	1179	6.40	3.01	4 48	68.00																
		2.3	8	129 9	10 90	12 00	10.39	45.61																
		2.5	10	106.4	9.50	17 00	10.64	35.71																
		2.5	25	83.5	14.50	37.70	20.83	27.78																
6.	<i>S. tamesis</i>	2.0	1.8	131.7	3.80	0.94	2.37	80.16																
		2.5	5	176.7	10.06	12.00	8.82	45.60																
		2.6	7.5	182.0	11.85	24.32	13.65	32.76																
		2.1	10	113.3	9.10	14.70	11.33	38.23																

avoid diatom settling. The resulting concentration of the diatom suspension was determined by using haemocytometer for each 15 minutes interval of feeding. In each 15 minutes after sampling, a new suspension of diatom was added to maintain the original concentration.

During the experiment the production of pseudofeces and feces were removed from the tank and collected in a beaker to minimize the chances of reingestion by the mussels. All the feeding in the laboratory at the room temperature of 28°C ± 2°C. The salinity of the feeding suspension was 28±1‰ and the pH was 7.9 ± 0.2. The control cultures of

diatoms of the same age kept in the experimental condition showed no growth in the experimental duration.

The collected pseudofeces were resuspended in known volume of sterile water and the cell numbers were counted. Then the total number of cells cleared from the suspension was calculated. The rate of filtration was calculated by using the formula $F = R/C$ (Epifanio and Ewart 1977) where 'F' is the filtration rate in ml/h/g whole weight, 'R' is the mean number of cells removed from suspension per gram whole weight per hour and 'C' is the number of algal cells per ml in suspension.

RESULTS

Rate of Filtration

The total number of cells removed from the suspension appeared to be related to the size of the algal cells and the suspension concentration (Fig 1) The filtration rate was high in small celled diatom suspensions and decreased towards the increase of diatom cell size. The maximum number of cells filtered in unit time was clearly less for the larger algae, *Streptotheca tamesis*, *Chaetoceros didymus* and

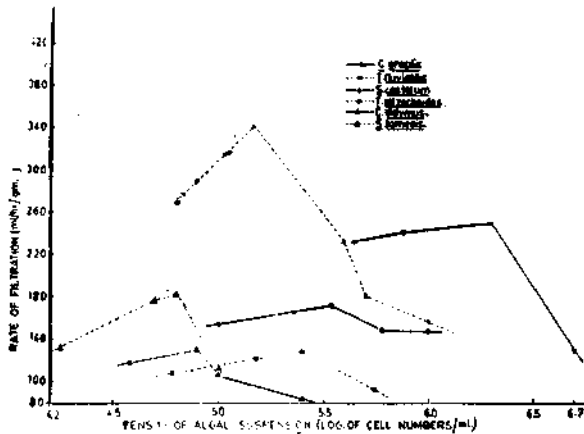


Fig. 1. Mean filtration rate as a function of density of algal suspension in *P. viridis*.

Thalassionema nitzschoides than the other three species (Table 2). The highest value of filtration density of 1.5×10^6 cells/ml of *Thalassiosira fluviatilis*. Rate of filtration was the lowest (83.5 ml/h/gm) when fed on *C. didymus* at 2.5×10^4 cells/ml. In all the algal suspensions, the filtration rate was relatively low in low cell densities and gradually increased up to a particular concentration and then declined with the further increase of suspension concentration. A sudden decline of filtration rate was observed in *Chaetoceros gracilis* and *S. tamesis* suspensions due to high suspension density [Table 2]. Eventhough the filtration rate was reduced with the increase of suspension density in all the experiments the number of cells removed per unit time increased with the increase of suspension density [Fig 2]. The following is the order of filtration rate observed for the six species employed: *T. fluviatilis* > *C. gracilis* > *S. tamesis* > *Skeletonema costatum* > *C. didymus* > *T. nitzschooides*. It is interesting to note that

the rate of filtration of *S. costatum* was slightly higher than that of *S. tamesis* eventhough the cell size of the former species is larger than the latter. But the mean number of cells of *S. tamesis* removed by the mussel was lower than that of all the other forms.

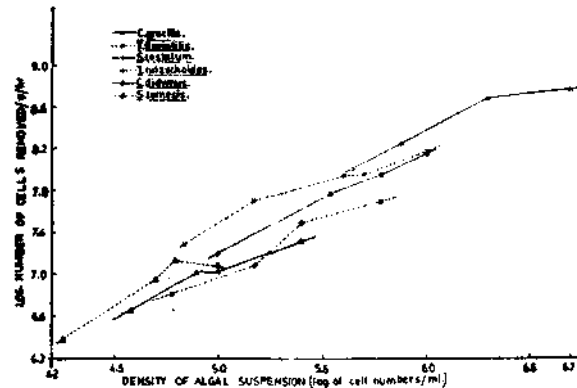


Fig. 2. Rate of removal as a function of algal suspension in *P. viridis*.

Food Ingestion and Pseudofeces Production

The rate of food ingestion also varied with respect to the algal cell size and suspension density. In low Suspension density, most of the filtered cells were ingested and the vice versa was observed in high concentrations, The filtered cells rejected were eliminated as strings of pseudofeces. But in most of the cases the actual amount of cells ingested was not affected, by concentration. The least amount of filtered cells were ingested when fed on *C. gracilis*. More than 75% of the filtered cells were eliminated out as pseudofeces even in the lowest suspension density [4.5×10^4] and only 22.6% was ingested. The ingestion rate gradually decreased and only 3.66% was ingested at the highest suspension density [5×10^6 cells/ml]. The mean number of cells ingested by the mussel was more or less same ($60.55 \pm 2.6 \times 10^4$ cells/h) in all the suspension densities. More than 50% of the filtered cells of *T. fluviatilis* were ejected as pseudofeces and 46.16% was ingested at the lowest suspension density used. The pseudofecal production increased with the increase of suspension density and 20.63% was ingested at the cell density of 1×10^4 cells/ml. The actual number of cells ingested by the mussel

was 22.5×10^8 /h and 55×10^8 /h per animal at the lower and higher cell densities respectively.

The highest ingestion rate was observed when fed on *S. costatum*. In the lowest suspension density [1×10^6 cells/ml] 86.54% of the filtered cells were ingested. The remaining was [13.46%] ejected out as pseudofeces. But the percentage of ingestion was decreased to 34.68% in the highest concentration of alga, cells (1×10^8 cells/ml.) The actual number of cells ingested was more or less same in the concentrations 3.5×10^6 and 6×10^6 cells/ml. But the values were slightly lower and higher in the extreme suspension densities.

The cells of the other 3 diatoms, *T. nitzschoides*, *C. didymus* and *S. tamesis* were effectively ingested at the lowest concentrations. Eventhough the cell sizes of these diatoms are larger than the other three, the ingestion rates were relatively high. As in other cases, the pseudofecal production was enhanced when the suspension density increased. Eventhough the number of cells removed from the suspension was increased with the increase of suspension density, the percentage of filtered cells ingested decreased. The mean number of cells ingested shows a complicated pattern with respect to the suspension density. The number of cells ingested by the mussel ranged from 10.08×10^6 to 26.5×10^6 cells/h per animal in *T. nitzschoides*, 6.4×10^6 to 14.5×10^6 cells/h per animal in *C. didymus* and 3.8×10^8 to 11.85×10^8 cells/h per animal in *S. tamesis*.

DISCUSSION

Rate of Filtration

As many workers have emphasized, the pumping rate in bivalves is a function of the concentration of particles present in the suspension. The present study reveals that the filtration was very much decreased due to the increase of suspension density and cell size. Mostly the diatoms of small cells were efficiently cleared by the mussel. When the cell concentration was low, the efficiency of retention was high. The retention efficiency mainly depend upon the cell size, suspension density and the relative

effect of the bivalve. Loosanoff and Engle (1947) observed that the cells of flagellate *Euglena* of 60M. could easily pass through the gills, sometimes only 15% were strained from the suspension, and maximum retention was 80%. But the cells of *Chlorella* [5 (*)] were retained by the gills, varyingly from 0 to 85%. Tammes and Oral [1955] observed widely varying retention of particles less than 30-40 μ by mussels. The retention efficiency of the mussel *Mytilus edulis* on blood corpuscles of 7-8 μ varied from 0 to 98%. Jorgensen (1966) reported that very high concentrations can evoke suppression or deterring responses in suspension feeders. Davids [1964] found a reduction in particle retention of mussels at concentration of *Nitzschia* above 1×10^6 cells/ml. Epifanio and Ewart [1977] reported that the total number of cells removed from suspension was related to the size of the algal cells. They observed that the maximum number of cells filtered was clearly less for the larger algae than for the smaller ones. Further, they found that the rates of filtration was inversely related to the concentration of particles in suspension. However Ballantine and Morton [1956] claimed that *Lasaea rubra* cleared suspensions of *Chromulina pusilli* (1 to 2μ) and *Prorocentrum micans* (30-40 μ) with equal rates independent of the size. The results of the present study closely agrees with the results of Epifanio and Ewart [1977].

Food Ingestion

Although there is considerable information concerning the filtration rates of bivalves, less is known about their actual feeding rates. In the present study, the results presented in the Table 2 shows that considerable amounts of filtered cells were ingested in low suspensions concentrations [Fig 3]. The ingestion rate was found to be high in *S. costatum* and low in *C. gracilis*. Considerable amounts of *S. tamesis* also was ingested. Matthiessen and Toner [1966] calculated that adult oysters growing near Martha's vineyard, Massachusetts could not possibly eat more than 1.1×10^7 algal cells/animal/day. Tenore and Dunstan (1973) showed that both clams and oysters fed most efficiently at food concentrations of 2×10^6 cells/ml.

Eventhough the filtration rate was affected by high suspension concentration, the actual



Fig 3 Number of cells ingested in relation to percentage of total cells filtered in *P. viridis*.

number of cells ingested was not affected (winter, 1973). The findings of the present study closely agrees with Winter's (1973) concept. The table 2 shows that the percentage of filtered cells ingested decreased with the increase of suspension, concentration, since copious amounts of pseudofeces were produced when the suspension density increased. It is evident that *Perna viridis* apparently have a maximum rate of ingestion in low cell concentration.

Some other factors also govern the ingestion rate of bivalves. Loosanoff and Engle (1917) and Floyd (1953) reported that the depressed feeding rate of oyster on *Chlorella* was due to the external metabolites present in the filtrate. The absence of silt with the algal diet reduced the clearance rate and ingestion rate in *NyctHus edulis* (Kiorbe et al 1980) Walne (1970) found that the higher the concentration of *Dunaliella*, the lower the growth rate of oysters i.e. the food ingestion was reduced. The results of the present study shows that the number of cells ingested was slightly increased when the suspension concentration increased. In *T. nitzschoides*, the number of cells ingested was found to be doubled in high concentration. Navarro and Winter (1982) also observed similar results in *Aytilus chilensis*. In these cases, the digestion would be partial, and the high ingestion rate however counter balanced by a significant decrease in assimilation efficiency (Navarro and Winter 1982).

The differences in ingestion rate of various diatoms may be due to the nutritive value of the species and sorting mechanism of the bivalve. Loosanoff (1949) and Menzel (1955) described ciliary mechanisms which they

believed may be responsible for sorting certain particles containing perhaps the more nutritious material in the food strained off the gill. Jorgensen (1949) found that when the rate of uptake of graphite suspensions of 4 to 5 μ particle size was compared with that of flagellate cultures of *D. inornata* and *I. galbana* of similar particle size, the filtration rate of flagellate cell was greater. However, the ingestion of palatable or nutritious particles may stimulate filtration. Epifanio (1979 b), and Romberger and Epifanio (1981) reported that the difference in growth rate in oysters when fed with various species of algae was due to the relative digestability of the algae. So it is believed that the mussel *P. viridis* is having some selective mechanism to avoid less nutritive materials. The least ingestion rate of *C. gracilis* might be due to the poor palatable or nutritive quality of the species

Generally *S. costatum* and *T. fluviatilis* are considered to be good source of food to bivalves. The high feeding rate of *Perna viridis* on *S. costatum* proves the same. Tenor and Dunstan (1973) argued that the oysters fed on *Tthalassiosira pseudonana* and *Isoctirysis galbana* have grown well. This was due to the high rate of ingestion and assimilation. Ukeles and Wikfors (1982) reported that the growth of oysters was rapid when fed with *Thalassiosira*. Dean (1957) compared the food value of *S. costatum* which field observations had indicated to be a good food to bivalves. The present investigation also proved *S. costatum* as a good source of food to *P. viridis*.

The three species of diatoms *T. nitzschoides*, *C. didymus* and *S. tamesis* also seem to be equally good food source to *P. viridis*, since the cells of these forms also ingested considerably. Walne (1970) stated that with some variations, that algae which were good or bad foods for one species of bivalve were of similar value to the other species of bivalves also.

Pseudofeces Production

All the particles filtered on the gill surface are carried by ciliary currents to the labial palps and so to the mouth or to the mantle edge to be rejected as pseudofeces, Table 2 shows that

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the production of pseudofeces increased with the increase of suspension concentration. Loosanoff and Engle (1947) using a continuous flow system, added *Chlorella* suspension to seawater and estimated that the pseudofeces

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