BDNF Val66Met polymorphism moderates the association between sleep spindles and overnight visual recognition

Risto Halonen, Liisa Kuula, Jari Lahti, Tommi Makkonen, Katri Räikkönen, Anu-Katriina Pesonen

S0166-4328(19)30462-0
https://doi.org/10.1016/j.bbr.2019.112157
112157
BBR 112157
Behavioural Brain Research
22 March 2019
16 August 2019
17 August 2019

Please cite this article as: Halonen R, Kuula L, Lahti J, Makkonen T, Räikkönen K, Pesonen A-Katriina, *BDNF* Val66Met polymorphism moderates the association between sleep spindles and overnight visual recognition, *Behavioural Brain Research* (2019), doi: https://doi.org/10.1016/j.bbr.2019.112157

This is a PDF file of an article that has undergone enhancements after acceptance, such as the addition of a cover page and metadata, and formatting for readability, but it is not yet the definitive version of record. This version will undergo additional copyediting, typesetting and review before it is published in its final form, but we are providing this version to give early visibility of the article. Please note that, during the production process, errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

© 2019 Published by Elsevier.



BDNF Val66Met polymorphism moderates the association between sleep spindles and overnight visual recognition

Authors: MA Risto Halonen^a, PhD Liisa Kuula^a, PhD Jari Lahti^a, MSc Tommi Makkonen^a, Prof. Katri Räikkönen^a, Prof. Anu-Katriina Pesonen^a

^aDepartment of Psychology and Logopedics, Faculty of Medicine, University of Helsinki, Finland,

P.O. Box 9, University of Helsinki 00014, Helsinki, Finland

Corresponding author: Risto Halonen, Department of Psychology and Logopedics, Faculty of Medicine, University of Helsinki, P.O. Box 9, University of Helsinki 00014, Helsinki, Finland; Tel: +358 40 847 91 34; e-mail: risto.halonen@helsinki.fi

Co-author e-mails (in order):

liisa.kuula-paavola@helsinki.fi jari.lahti@helsinki.fi tommi.makkonen@helsinki.fi katri.raikkonen@helsinki.fi anukatriina.pesonen@helsinki.fi

Highlights

- Interaction of BDNF genotype and sleep spindles largely unstudied
- Val66Met polymorphism did not impact picture recognition accuracy
- Frontal spindles related with better recognition accuracy in Val homozygotes only

- Val66Met moderated significantly the association regarding frontal fast spindles
- Sleep spindles may not associate with learning equally across individuals

Abstract

A common single nucleotide polymorphism (SNP) of the brain-derived neurotrophic factor (*BDNF*) gene, Val66Met, has been reported to impair BDNF secretion and memory function. However, few studies have investigated the interaction of *BDNF* genotype and sleep characteristics, such as sleep spindles, that promote long-term potentiation during sleep. In this study we compared overnight visual memory between the carriers of *BDNF* Met and non-carriers (Val homozygotes), and examined how sleep spindle density associated with memory performance.

The sample constituted of 151 adolescents (mean age 16.9 years; 69% Val homozygotes, 31% Met carriers). The learning task contained high and low arousal pictures from Interactive Affective Picture System. The learning task and all-night polysomnography were conducted at the homes of the adolescents. Slow (10–13 Hz) and fast (13–16 Hz) spindles were detected with automated algorithm.

Neither post-sleep recognition accuracy nor spindle density differed between Val homozygotes and Met carriers. While frontal slow and fast spindle densities associated with better recognition accuracy in the entire sample, examining the allelic groups separately indicated paralleling associations in Val homozygotes only. Interaction analyses revealed a significant genotype-moderated difference in the associations between frontal fast sleep spindles and high arousal pictures.

In sum, sleep spindles promote or indicate visual learning in Val homozygote adolescents but not in Met carriers. The result suggests that the role of sleep spindles in visual recognition memory is not equal across individuals but moderated by a common gene variant.

Keywords: LTP, plasticity, sleep spindle, Val66Met, visual memory

1. Introduction

Brain-derived neurotrophic factor (BDNF) is a growth hormone mediating neuronal survival and differentiation [1, 2]. BDNF also promotes synaptic plasticity [3] such as hippocampal longterm potentiation (LTP) [1, 2, 4-7], important in memory and learning [8]. Regarding memory function specifically, a single nucleotide polymorphism (SNP) of the human *BDNF* gene is rs6265, or Val66Met, has drawn interest. This methionine (Met) substitution for valine (Val) at codon 66 has been observed to alter intracellular packaging of pro-BDNF, its axonal transport while also reducing the activity-dependent dendritic secretion of BDNF [9, 10]. Evidence in rodents suggests that Val66Met polymorphism impairs hippocampal synaptic plasticity [11]. However, human studies examining memory function between Met carriers and non-carriers (Val homozygotes) have provided contrasting observations of hippocampal activation [12-14] and memory performance [15-19], leaving the impact of Val66Met polymorphism on human memory performance unestablished [20].

Sleep is a powerful memory enhancer [21-23]. Given that BDNF facilitates late-phase LTP (i.e. memory lasting longer than 1-2 hours) [24, 25], studies involving *BDNF* variants in postencoding sleep deserve focus. In some studies differences in memory performance emerged only after night's sleep, even when reporting equal short-delay recall between the genotypes [26, 27]. In overnight studies the memory performance of Val homozygotes has been reported equal [15] or better [19, 26, 27] compared to Met carriers. A study associating sleep characteristics with overnight learning found that the improved face picture recognition of Val homozygotes related to the increase of slow oscillation (SO) power between baseline and test night, suggesting a more profound impact of pre-sleep learning on sleep power dynamics [26]. Sleep spindles, bursts of thalamocortical sigma-band oscillations (~10–16 Hz) mostly seen in stage 2 (N2) sleep [28], did not affect the learning outcome in the study. However, sleep was only analyzed in the first quartile of

the first non-rapid eye movement sleep (NREM) episode [26], hardly entirely representing the role of spindles in overnight learning.

Comprehensive evidence links sleep spindles with enhanced memory performance [29-37]. Yet, there is a lack of studies investigating sleep spindles together with BDNF, despite existing basis to assume interaction in memory function. Sleep spindles are involved in memory replay during sleep [21, 38-42] and considered a mechanism of LTP [43]. Triggered by N-Methyl-Daspartate receptor (NMDAR) activation, strong Ca²⁺ influx during sleep spindles activates postsynaptic signaling cascades underlying LTP [44]. BDNF, on the other hand, promotes NMDAR function [45, 46]. In addition, recognizing the role of sleep spindles on hippocampal memory formation and neocortical information transfer [47-49], observations of Val66Met-related alterations in the hippocampal activation [12, 50], connectivity [13, 51, 52] and synchronization with neocortical processing [50] gain interest.

To illuminate the effect of individual genotype on memory functioning, we investigated how Val homozygotes and Met carriers differ in visual overnight learning outcome, and how sleep spindles moderate the associations in a community-based sample of 151 late adolescents in a natural overnight in-home setting. We hypothesize that Val homozygotes would display better overnight picture recognition performance and positive association between recognition performance and spindle density during N2 and N3 sleep.

2. Experimental Procedure

2.1. Participants

The participants comprised an urban community-based cohort composed of 1049 healthy singletons born between March and November 1998 in Helsinki, Finland [53]. Detailed descriptions of the cohort and follow-up participation are found elsewhere [54, 55]. In the current study, the adolescents who lived within a 30 kilometer radius of Helsinki and whom had participated in the previous follow-up and given consent for further contact, were recruited by phone and were offered a monetary compensation (50 \in) for their effort. In total 196 adolescents participated of which 173 had been genotyped at an earlier follow-up. 22 participants had to be excluded from the sleep spindle analyses due to poor impedance levels or other measurement problems, and visual memory task data was missing from three participants due to technical problems. The final analytical sample consisted of 151 Caucasian adolescents (56 % girls; mean age 16.9 y, SD=0.1, range 16.6–17.2). We did not exclude any participants, as there were no current neurodevelopmental disorders reported. Two cases reported having had learning difficulties during elementary school.

The Ethics Committee of the Children's Hospital in Helsinki University Central Hospital approved the study protocol (177/13/03/03/2014). Informed written consent was obtained from the participants. All parts of the study were conducted in accordance with the Declaration of Helsinki.

2.2. Experiment flow

The in-home assessment started between 6–7 p.m. with a short questionnaire about possible factors affecting testing, e.g. handedness, native language and possible sensory or motor handicaps. After that a trained research nurse administered a cognitive assessment and the encoding phase of the recognition accuracy task. The polysomnography (PSG) device was then attached, and the subjects were instructed to follow their own sleep schedule. The next morning the research nurse detached the PSG wiring and administered the recognition phase of the recognition accuracy task.

2.3. Picture Recognition Task

The stimuli consisted of two sets of 100 pictures from the International Affective Picture System [56]. The sets were differentiated by their arousal (calm-exciting dimension) ratings: the mean normative arousal of the high and low arousal picture sets were 5.68 (5.00-7.35) and 3.47 (2.28–3.99), respectively (statistically significant difference, p<.001). The mean valence ratings were parallelized between the sets (mean valence of high and low arousal pictures were 5.75 and 5.84, respectively; p = .63). In the learning phase the participants were instructed to memorize 100 target pictures (50 low and 50 high arousal), viewed on a 14" laptop screen. The following morning, in the recognition phase, the 100 target pictures were mixed with 100 unseen sham pictures and displayed to the participants in random order. If they recognized the picture, the participants were instructed to press a key (space bar) while the picture was visible. In both learning and recognition phases the pictures lasted for 1000 ms on the screen, followed by blank black screen, lasting 1500 ms. The research nurse monitored that participants focused on the task. Recognition accuracy scores (d', separately for high and low arousal) were calculated as the difference between the hit rate (standardized proportion of correctly recognized target pictures of all target pictures) and the false alarm rate (standardized proportion of incorrectly recognized sham pictures of all sham pictures) to correct for response bias. Due to false alarm rates of 0, we applied loglinear approach [57].

2.4. PSG protocol and spindle detection

All recordings were done using SOMNOscreen plus (SOMNOmedics GmbH, Germany). A trained research nurse attached gold cup electrodes at 6 electroencephalography (EEG) locations (frontal (F) hemispheres: F3, F4; central (C) hemispheres: C3, C4; occipital (O) hemispheres: O1, O2) and two for the mastoids (A1, A2) accordingly. The electro-oculogram (EOG) and the electromyogram (EMG) were measured by using disposable adhesive electrodes (Ambu Neuroline 715, Ambu A/S, Denmark), two locations for EOG and three locations for EMG. In addition, an

online reference Cz and a ground electrode in the middle of forehead were used. The sampling rate was 256 Hz (the hardware filters for SOMNOscreen plus are 0.2-35 Hz).

PSG data were scored manually using the DOMINO program (v2.7; SOMNOmedics GmbH, Germany) in 30-sec epochs into N1, N2, N3 and REM according to AASM guidelines (The AASM Manual for the Scoring of Sleep and Associated Events). Percentages of each stage were calculated based on total sleep time. All signals were digitally offline filtered with pass band of 0.5-35 Hz (Hamming windowed sinc zero-phase FIR filter, cut-off (-6dB) 0.25 Hz and 39.3 Hz respectively) and re-referenced to the average signal of A1 and A2 electrodes.

The manually scored PSG signals were converted to EDF format in DOMINO software and then further analyzed by using functions of EEGlab 14.1.2b (Delorme and Makeig 2004) running on Matlab R2018a (The Mathworks Inc., USA). All signals were digitally offline filtered with pass band of 0.5–35 Hz (Hamming windowed sinc zero-phase FIR filter, cut-off (-6dB) 0.25 Hz and 39.3 Hz respectively) and re-referenced to the average signal of A1 and A2 electrodes. Electrodes located at F3, F4, C3, C4 were included in the analysis. Only epochs with with electrode-scalp and both mastoids impedance equal or lower than 10 k Ω were included in the analyses.

Spindles were computationally extracted separately in N2 and N3 sleep with a method based on an automated detection algorithm described by Ferrarelli [58]. The spindle analysis was conducted in valid N2 and N3 epochs in two different frequency bands (slow: 10–13 Hz, and fast: 13–16 Hz) in order to differentiate between slow and fast spindles, which are likely to serve different functions in overnight learning [49, 59, 60]. Before applying the spindle thresholding method, the pre-processed EEG data were further filtered using the above-mentioned frequency bands separately, using high-order filter (13,200) to minimize overlap between the frequency bands [61]. The threshold values for finding spindle peak amplitude in each channel were defined by the mean of the channel amplitude (μ V) multiplied with 5 (higher) including all valid epochs (impedance in the target channel and both mastoids $\leq 10 \text{ k}\Omega$). The higher threshold (5) was iterated

by visual inspection of EEG data to provide best detection of spindle events. The spindle's amplitude was required to stay over the mean channel amplitude multiplied by 2 (lower) for 250 ms in both directions from the peak maximum, resulting in minimum spindle duration of 0.5 seconds [62]. Thus, we used channel-wise threshold definitions, taking into account that signals may vary across the channels. The maximum cut-off for spindle length was set 3.0 seconds [28] and maximum peak amplitude was set to 200 μ V. Also, between spindles the signal amplitude was required to stay under the lower threshold for 78.1 ms which is approximately the duration of one period of sine at 13 Hz, in order to prevent false alarms. Spindle-like bursts detected during arousals were excluded. Fast and slow spindle densities (number of spindles per minute) in each EEG locations were used as measures of spindle activity. Spindle densities were calculated only if the amount of valid minutes in N2 or N3 equaled or exceeded 10.

2.5. Genotyping

DNA was extracted from blood (22%) and saliva samples (78%) collected at the 2009–2011 follow-up. Genotyping was performed with the Illumina OmniExpress Exome 1.2 bead chip at the Tartu University, Estonia, in September 2014 according to the standard protocols. We assessed the frequencies of GG (Val/Val), GA (Val/Met) and AA (Met/Met) genotypes. For data analysis Val/Met and Met/Met were grouped as Met carriers.

2.6. Statistics

All statistical analyses were done using IBM SPSS Statistics version 25.0 (IBM Corp. Released 2016. IBM SPSS Statistics for Windows, Version 25.0. Armonk, NY: IBM Corp.). Significance was set at p<0.05. Baseline differences between the genotype groups were analyzed using one-way analysis of variance (ANOVA).

Mixed ANOVAs were used in assessing recognition accuracy and the interaction of arousal level and genotype, with arousal level (low, high) as within-subjects factor and genotype (Val homozygotes, Met carriers) as between-subjects factor.

To examine the effect of sleep spindles on learning we first averaged the densities of frontal (F3, F4) and central (C3, C4) slow and fast spindles, resulting in four variables: central slow density, central fast density, frontal slow density and frontal fast density. Linear regression analysis was used to test the significance of each spindle variable (independent) on learning task score (dependent) for the whole sample and separately for the allelic groups. To test if the associations between each spindle variable and recognition task score differed between the genotypes, General Linear Model (GLM) two-way ANOVA was used to compare the regression slopes with an interaction term of 'spindle density variable x genotype'.

The analyses were run with two models including different covariates. As covariates in Model 1 we used only sex. In model 2 we used added full-scale intelligence quotient (FSIQ), total sleep time (TST) and the time awake between the encoding and recognition phase of the memory task (before and after night's sleep). To control for the impact of general cognitive ability on learning performance, we assessed intellectual ability with a shortened version of the Wechsler Adult Intelligence Scale III (WAIS-III) [63]. The assessment included five WAIS-III subtests in the following order: Vocabulary, Block Design, Similarities, Matrix Reasoning and Digit Span. Full-Scale IQ (FSIQ) was calculated by averaging the Z scores of the subtests. TST was included in the covariates to control for differences in sleep duration, which may affect overnight learning performance [64]. As we chose not to directly affect the bedtime or awakening of the participants, the time the participants spent awake between the encoding phase and sleep onset and between awakening and recognition phase were used as a single covariate. Timing of the encoding and recollection phases in overnight learning paradigms may affect results [65].

3. Results

3.1. Genotyping

rs6265 showed genotyping success rate \geq 95%, minor allele frequency of 0.16, and was in Hardy–Weinberg equilibrium (p-value > 0.05). In the analytic sample, there were 103 (68 %), 42 (27 %), and 6 (4 %) of GG (Val/Val), GA (Val/Met), AA (Met/Met) genotypes. For analyses, Val/Met and Met/Met groups were combined (31 % in any Met carrier group). Possible differences in sample characteristics variables between Val/Met and Met/Met groups were examined with oneway ANOVA, but no significant differences were detected (p values > .08, data not shown).

3.2. Sample Characteristics

3.3. Recognition accuracy

A two x two mixed ANOVA analyzed the influence of *BDNF* genotype group and image arousal on overnight recognition performance. We run analyses separately for Model 1 and Model 2. With recognition performance (*d'*) as the dependent variable (Fig. 1), the analyses showed no significant main effects of picture arousal (Model 1: $F_{1, 148} = 0.101$, p = 0.751, $\eta_p^2 = 0.001$; Model 2: $F_{1, 145} = 0.240$, p = 0.625, $\eta_p^2 = 0.002$) nor of the *BDNF* genotype ($F_{1, 148} = 1.820$, p = 0.179, $\eta_p^2 =$ 0.012; Model 2: $F_{1, 145} = 0.903$, p = 0.344, $\eta_p^2 = 0.006$). The interaction of genotype and arousal was not significant ($F_{1, 148} = 0.014$, p = 0.906, $\eta_p^2 < 0.001$; Model 2: $F_{1, 145} = 0.029$, p = 0.865, $\eta_p^2 <$ 0.001).

3.4. Sleep Spindles in Post-Sleep Recognition Accuracy

To assess how N2 slow and fast spindle densities associated with post-sleep recognition accuracy in high and low arousal pictures, the whole sample underwent regression analyses. In all subjects we observed recognition accuracy for high arousal (high d') pictures to associate with N2 frontal slow spindles when controlling for sex only (Model 1: B = 0.092, t = 2.205, p = .029. Also,

high *d*' associated significantly with frontal fast spindles in both models in the entire sample (Model 1: B = 0.079, t = 2.258, p = .025; Model 2: B = 0.091, t = 2.386, p = .018). In addition, recognition performance of low arousal pictures (low *d*') related significantly with frontal slow spindles (Model 1: B = 0.124, t = 2.626, p = .010); Model 2: B = 0.097, t = 2.106, p = .037). No associations were found regarding N3 spindles (see Supplementary Material 2).

Examining the allelic groups separately revealed significant associations in the Val homozygote group. Better recognition of high arousal pictures associated with frontal slow spindle density (Model 1: B = 0.114, t = 2.569, p = .012; Model 2: B = 0.098, t = 2.193, p = .031) and frontal fast spindle density (Model 1: B = 0.122, t = 2.977, p = .004; Model 2: B = 0.124, t = 3.080, p = .003). Regarding low arousal pictures, analogous relationships were found between low *d*' and frontal slow spindle density (Model 1: B = 0.147, t = 2.695, p = .008; Model 2: B = 0.114, t = 2.160, p = .033) and frontal fast spindle density (Model 1: B = 0.147, t = 2.695, p = .008; Model 2: B = 0.114, t = 2.160, p = .033) and frontal fast spindle density (Model 2: B = 0.104, t = 2.155, p = .034). No associations were found regarding N3 spindles (see Supplementary Material 2).

To examine if the relationship between N2 sleep spindle density and recognition accuracy differed according to genotype, we ran two-way ANOVA interaction tests with the interaction term 'genotype x spindle density' (Table 2). After controlling for the covariate(s) and main effects, the interaction of N2 frontal fast spindles and genotype showed significance in high *d*' (Model 1: $F_{1,146}$ = 3.891, p = 0.050, ηp^2 = 0.026; Model 2: $F_{1,143}$ = 4.662, p = 0.033, ηp^2 = 0.032), indicating divergence in the associations between the genotypes. No differences were found regarding N3 spindles (see Supplementary Material 2).

Residual plots for the extracted coefficients of determination (\mathbb{R}^2) in Fig. 2 illustrate the genotype-moderated associations between frontal fast spindle density and high *d*' separately for Val homozygotes and Met carriers. In the Val/Val group frontal fast spindle density explains 8.9 % of the variability of high *d*' scores and 0.1 % in Met carriers.

4. Discussion

Our study found that carriers of common *BDNF* alleles (Val/Val, ie. Val homozygotes vs. Val/Met and Met/Met, ie. Met carriers) showed no diverging performance in overnight learning of pictures of low and high arousal in a large adolescent sample. Val homozygotes represented 69 % and Met carriers 31 % of the cohort, corresponding to allelic distribution in European populations [67, 68]. In the entire sample frontal spindle density associated with better post-sleep picture recognition, and when examining the allelic groups separately, this pattern recurred in Val homozygotes only. The genotype-moderated difference between spindle-recognition-relationship was significant regarding frontal fast spindle density and high arousal picture recognition.

The null finding in the difference in visual recognition performance between the genotypes contradicts an earlier overnight study reporting improved performance in Val homozygotes [26]. However, it aligns with another study's findings where the overall recall performance was similar between Val homozygotes and Met carriers [15]. Interestingly, the authors found that emotionally high picture valence (positive and negative) improved the performance in Met carriers compared to

neutral pictures. Similar impact was not detected regarding picture arousal (calm–exciting) in our study. According to neuroimaging studies, image valence, but not arousal, correlates with amygdala activity [69, 70]. Amygdala activity during encoding associates with enhanced memory consolidation [71], and Met carriers have been reported to show higher amygdala activity towards emotional stimuli than Val homozygotes [72, 73]. Based on our results, it appears that the perception and processing of arousal does not separate *BDNF* polymorphisms to the extent of valence. However, we could not divide recognition performance into 'recollection' and 'familiarity' responses [74, 75] as a recent study (not involving post-encoding sleep), showing that only 'familiarity' response accuracy differed between Val homozygotes and Met carriers [76].

In our study, we also focused on the associations between sleep spindles and recognition accuracy. In the entire sample, frontal slow and fast spindle density in N2 sleep associated with better post-sleep recognition of pictures. Considering that numerous reports associate (also) central spindle density with declarative learning [29, 34, 77, 78] the topographical dichotomy urges further scrutiny. Previously frontal spindle activity has been linked with the learning of word-pairs [29, 79, 80], with associating faces and names [30, 81, 82] and with contextual memory [83]. One study found both frontal and central spindle power to correlate with better neutral picture memory [84]. Considering that recognition memory is deemed to depend upon prefrontal cortex [85-89], medial temporal lobe structures and interconnecting white matter projections [76, 90], our results contribute to relatively scarcely studied matter.

Arguably slow and fast spindles serve diverging functions in learning. Especially fast spindles are implicated with offline memory consolidation as they associate with greater encodingrelated hippocampal activation [82] and higher hippocampal-neocortical functional connectivity [91]. Fast spindles coincide with slow oscillations and hippocampal sharp wave ripples, consequently promoting memory transfer between hippocampus and neocortex [49, 92] Moreover, Mander et al. [82] found frontal fast spindles to restore next-day learning capacity. Slow spindles,

on the other hand, have been proposed to follow fast spindles and be involved in cortico-cortical information processing within prefrontal cortex [60], although rather limited evidence associates slow spindles with better learning in adults [33]. Notably, sleep spindles of varying frequency ranges [93-95] have been attributed with pre-sleep memory performance, a measure not examinable in our study. Such 'learning aptitude' does not equal, but overlaps with [96, 97], the construct of general cognitive ability. Introducing full-scale intelligence quotient – a contested correlate with sleep spindle characteristics [98] – as a covariate did not dispel the associations between recognition accuracy and slow or fast frontal spindle density in our data. This implies a learning component beyond general cognitive ability regarding frontal spindles. However, the exact contribution of each spindle type on the learning performance would require further investigation of e.g. inter-spindle dynamics and synchronization with other sleep oscillations.

Not all studies report associations between spindles and declarative learning (for example, see [99]). Given that sleep spindles reflect synchronized activity of inter-individually variable neuroanatomical structures [100], few studies have considered the influence of subject-specific factors beyond age and sex on the obtained results. Hence, we examined the interaction between Val66Met polymorphism and sleep spindles on overnight memory performance. Analyses run separately for Val homozygotes and Met carriers revealed that each of the significant association in the entire sample involved a strong, paralleling association confined to Val homozygotes only, whereas spindle density in recognition accuracy showed no significance in Met carrier group. The associations were significantly divergent regarding frontal fast spindles in high arousal picture recognition. The result proposes that frontal fast sleep spindles promote or indicate visual recognition memory formation differently between Val homozygotes and Met carriers.

Neural underpinnings behind the observed difference between Val homozygotes and Met carriers remain a matter of speculation. Volumetric analyses indicate Val homozygosity to associate with larger prefrontal [101-103] and hippocampal [9, 50, 101, 104, 105] gray matter volume in

comparison to Met carriers. Based on altered activation patterns and higher error rate in verbal learning task, Schofield et al. [50] suggested Met-allele-associated dysregulated activation of hippocampus and its prefrontal projections. Concordantly, functional connectivity between hippocampal and neocortical areas is reportedly higher in Val homozygotes than in Met carriers [13, 51, 52], which proposedly [106] derives from more efficient pruning of silent axons, a process modulated by BDNF [107, 108]. Indeed, whereas white matter (WM) integrity underlies the propagation [100, 109] and memory benefit [110] of sleep spindles, the relation between WM characteristics and cognition may be affected by Val66Met polymorphism, and appears discernible in Val homozygotes only [111, 112]. Acknowledging the methodological distance, this evidence encourages us to suggest that the addressed genotypic differences in fronto-hippocampal network and in connectivity dynamics contribute to the frontal emphasis in spindle-learning-relation in Val homozygotes in our sample. Furthermore, it is well-established that specifically fast spindles promote memory consolidation via inter-oscillation synchronization [113, 114], orchestrated by prefrontal cortex [92]. Higher connectivity enhances synchrony [115-117] and BDNF is involved in stabilizing even complex patterns of potential fluctuations [118]. While this theoretically parallels the accentuated genotypic moderation regarding fast spindles in our data, further studies investigating the exact impact of Val66Met polymorphism on phase-amplitude coupling characteristics are warranted.

Some issues should be underscored here. Met carriers fared equally with Val homozygotes in recognition accuracy in our study, although no other correlate with learning aside spindles was identified. Val66Met polymorphism affects the dynamics between sleep and learning in a complex manner, affecting also next-day cognition [119, 120], which necessitates more research with varying settings. In addition, our sample consisted of closely-aged adolescents, with ongoing neural reorganization of brain [121], increasing thalamocortical functional connectivity [122] and altering spindle characteristics [78, 123]. The narrow age-range may highlight subtle inter-group differences

in neural activity and cognitive functioning. Hence, these results can only be cautiously generalized to other age groups.

4.1. Strengths and Limitations

A key strength of our study was the large, community-based longitudinal sample with a high age coherence. This study adds to the increasing research literature on adolescents' spindles. Uniquely, our study is the first to assess how *BDNF* gene moderates the association between sleep spindles and overnight picture recognition.

There are also major shortcomings requiring attention. Firstly, the study setting enables only correlative scrutiny of the associations. That is, the singular morning picture recognition without a pre-sleep measurement makes the overnight change in recognition accuracy unexaminable. This undermines the deductibility of causal role of sleep spindles in learning. Further obscuring causation, having only one night with PSG recording prevented us from examining how learning affected sleep spindle characteristics, as well as from considering previous night's impact on learning [82, 119, 120]. Also, due to the lack of a waking control group we could not properly assess how, or whether, sleep affected the recognition accuracy scores. Within these limitations it is not inferable if the associations between frontal spindles and recognition accuracy derives from offline consolidation or from general learning capability. Controlling for FSIQ diminishes the effect of intelligence on learning performance, but hardly equals immediate learning ability. Secondly, 'recollection' and 'familiarity' responses, evidently representing divergent neural processes [124], were undifferentiated. Thus, we were unable to investigate these subtypes separately although their relevance regarding Val66Met polymorphism has recently been reported [76]. Thirdly, while we parallelized the mean valence ratings of the applied picture sets, the variance in valence was rather high, possibly introducing unassessed impact on recognition performance [15]. Finally, due to less

Met carriers than Val homozygotes, the analyses run separately on the groups are not fully comparable due to difference in statistical power. This calls for balanced groups in further studies.

4.2. Conclusions

The relation between sleep microstructure and memory formation may not be equal across all individuals. This study is the first to show that sleep spindle density associates with better visual post-sleep recognition accuracy only in Val homozygotes. In Met carriers, sleep spindles did not associate with learning. We suggest that the results reflect genotype-moderated functional differences in fronto-hippocampal network. The exact nature of the divergence requires further studies.

Funding

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

Declarations of interest: None

Acknowledgements

The authors would like to thank Helena Alfthan for her effort in participant recruitment and data collection and Rachel Robinson for improving the language.

References

- Zagrebelsky, M.,M. Korte. Form follows function: BDNF and its involvement in sculpting the function and structure of synapses. *Neuropharmacology*, 76 (2014): pp. 628-638, DOI: https://doi.org/10.1016/j.neuropharm.2013.05.029.
- Park, H.,M.M. Poo. Neurotrophin regulation of neural circuit development and function. *Nat Rev Neurosci*, 14 (2013): pp. 7-23, DOI: 10.1038/nrn3379.
- Gottmann, K., T. Mittmann, V. Lessmann. BDNF signaling in the formation, maturation and plasticity of glutamatergic and GABAergic synapses. *Experimental Brain Research*, 199 (2009): pp. 203-234, DOI: 10.1007/s00221-009-1994-z.
- Bramham, C.R.,E. Messaoudi. BDNF function in adult synaptic plasticity: The synaptic consolidation hypothesis. *Progress in Neurobiology*, 76 (2005): pp. 99-125, DOI: https://doi.org/10.1016/j.pneurobio.2005.06.003.
- Kovalchuk, Y., E. Hanse, K.W. Kafitz, A. Konnerth. Postsynaptic Induction of BDNF-Mediated Long-Term Potentiation. *Science*, 295 (2002): pp. 1729.
- Novkovic, T., T. Mittmann, D. Manahan-Vaughan. BDNF contributes to the facilitation of hippocampal synaptic plasticity and learning enabled by environmental enrichment. *Hippocampus*, 25 (2015): pp. 1-15, DOI: 10.1002/hipo.22342.
- Leal, G., P.M. Afonso, I.L. Salazar, C.B. Duarte. Regulation of hippocampal synaptic plasticity by BDNF. *Brain Res*, 1621 (2015): pp. 82-101, DOI: 10.1016/j.brainres.2014.10.019.
- 8. Bliss, T.V.,G.L. Collingridge. A synaptic model of memory: long-term potentiation in the hippocampus. *Nature*, 361 (1993): pp. 31-9, DOI: 10.1038/361031a0.
- Egan, M.F., M. Kojima, J.H. Callicott, T.E. Goldberg, B.S. Kolachana, A. Bertolino, E.
 Zaitsev, B. Gold, D. Goldman, M. Dean, B. Lu, D.R. Weinberger. The BDNF val66met

polymorphism affects activity-dependent secretion of BDNF and human memory and hippocampal function. *Cell*, 112 (2003): pp. 257-69.

- Chen, Z.-Y., P.D. Patel, G. Sant, C.-X. Meng, K.K. Teng, B.L. Hempstead, F.S. Lee.
 Variant Brain-Derived Neurotrophic Factor (BDNF) (Met66) Alters the Intracellular
 Trafficking and Activity-Dependent Secretion of Wild-Type BDNF in Neurosecretory Cells
 and Cortical Neurons. *The Journal of Neuroscience*, 24 (2004): pp. 4401.
- Ninan, I., K.G. Bath, K. Dagar, R. Perez-Castro, M.R. Plummer, F.S. Lee, M.V. Chao. The BDNF Val66Met polymorphism impairs NMDA receptor-dependent synaptic plasticity in the hippocampus. *The Journal of neuroscience : the official journal of the Society for Neuroscience*, 30 (2010): pp. 8866-8870, DOI: 10.1523/JNEUROSCI.1405-10.2010.
- Hariri, A.R., T.E. Goldberg, V.S. Mattay, B.S. Kolachana, J.H. Callicott, M.F. Egan, D.R. Weinberger. Brain-Derived Neurotrophic Factor val<sup>66</sup>met Polymorphism Affects Human Memory-Related Hippocampal Activity and Predicts Memory Performance. *The Journal of Neuroscience*, 23 (2003): pp. 6690.
- Fera, F., L. Passamonti, A. Cerasa, M.C. Gioia, M. Liguori, I. Manna, P. Valentino, A. Quattrone. The BDNF Val66Met Polymorphism Has Opposite Effects on Memory Circuits of Multiple Sclerosis Patients and Controls. *PLOS ONE*, 8 (2013): pp. e61063, DOI: 10.1371/journal.pone.0061063.
- Dennis, N.A., R. Cabeza, A.C. Need, S. Waters-Metenier, D.B. Goldstein, K.S. LaBar. Brain-derived neurotrophic factor val66met polymorphism and hippocampal activation during episodic encoding and retrieval tasks. *Hippocampus*, 21 (2011): pp. 980-989, DOI: 10.1002/hipo.20809.
- 15. Harrington, M.O., K. Klaus, M. Vaht, J. Harro, K. Pennington, S.J. Durrant. Overnight retention of emotional memories is influenced by BDNF Val66Met but not 5-HTTLPR.

Behavioural Brain Research, 359 (2019): pp. 17-27, DOI: https://doi.org/10.1016/j.bbr.2018.10.015.

- Mandelman, S.D.,E.L. Grigorenko. BDNF Val66Met and Cognition: All, None, or Some? A Meta-Analysis of the Genetic Association. *Genes, Brain, and Behavior*, 11 (2012): pp. 127-136, DOI: 10.1111/j.1601-183X.2011.00738.x.
- Yogeetha, B.S., L.M. Haupt, K. McKenzie, H.G. Sutherland, R.K. Okolicsyani, R.A. Lea,
 B.H. Maher, R.C.K. Chan, D.H.K. Shum, L.R. Griffiths. BDNF and TNF-α polymorphisms in memory. *Molecular Biology Reports*, 40 (2013): pp. 5483-5490, DOI: 10.1007/s11033-013-2648-6.
- Avgan, N., H.G. Sutherland, L.K. Spriggens, C. Yu, O. Ibrahim, C. Bellis, L.M. Haupt,
 D.H.K. Shum, L.R. Griffiths. BDNF Variants May Modulate Long-Term Visual Memory
 Performance in a Healthy Cohort. *International journal of molecular sciences*, 18 (2017):
 pp. 655, DOI: 10.3390/ijms18030655.
- Goldberg, T.E., J. Iudicello, C. Russo, B. Elvevåg, R. Straub, M.F. Egan, D.R. Weinberger. BDNF Val66Met polymorphism significantly affects d' in verbal recognition memory at short and long delays. *Biological Psychology*, 77 (2008): pp. 20-24, DOI: https://doi.org/10.1016/j.biopsycho.2007.08.009.
- Toh, Y.L., T. Ng, M. Tan, A. Tan, A. Chan. Impact of brain-derived neurotrophic factor genetic polymorphism on cognition: A systematic review. *Brain and behavior*, 8 (2018): pp. e01009-e01009, DOI: 10.1002/brb3.1009.
- Rasch, B.,J. Born. About Sleep's Role in Memory. 93 (2013): pp. 681-766, DOI: 10.1152/physrev.00032.2012.
- Stickgold, R.,M.P. Walker. Sleep-dependent memory consolidation and reconsolidation.
 Sleep medicine, 8 (2007): pp. 331-343, DOI: 10.1016/j.sleep.2007.03.011.

- 23. Diekelmann, S.,J. Born. The memory function of sleep. *Nat Rev Neurosci*, 11 (2010): pp. 114-26, DOI: 10.1038/nrn2762.
- Lu, Y., K. Christian, B. Lu. BDNF: a key regulator for protein synthesis-dependent LTP and long-term memory? *Neurobiology of learning and memory*, 89 (2008): pp. 312-323, DOI: 10.1016/j.nlm.2007.08.018.
- 25. Panja, D.,C.R. Bramham. BDNF mechanisms in late LTP formation: A synthesis and breakdown. *Neuropharmacology*, 76 Pt C (2014): pp. 664-76, DOI: 10.1016/j.neuropharm.2013.06.024.
- Mascetti, L., A. Foret, J. Schrouff, V. Muto, V. Dideberg, E. Balteau, C. Degueldre, C. Phillips, A. Luxen, F. Collette, V. Bours, P. Maquet. Concurrent Synaptic and Systems Memory Consolidation during Sleep. *The Journal of Neuroscience*, 33 (2013): pp. 10182.
- Cathomas, F., C. Vogler, J.C. Euler-Sigmund, D.J. de Quervain, A. Papassotiropoulos. Finemapping of the brain-derived neurotrophic factor (BDNF) gene supports an association of the Val66Met polymorphism with episodic memory. *Int J Neuropsychopharmacol*, 13 (2010): pp. 975-80, DOI: 10.1017/s1461145710000519.
- Purcell, S.M., D.S. Manoach, C. Demanuele, B.E. Cade, S. Mariani, R. Cox, G. Panagiotaropoulou, R. Saxena, J.Q. Pan, J.W. Smoller, S. Redline, R. Stickgold. Characterizing sleep spindles in 11,630 individuals from the National Sleep Research Resource. *Nat Commun*, 8 (2017): pp. 15930, DOI: 10.1038/ncomms15930.
- Gais, S., M. Mölle, K. Helms, J. Born. Learning-Dependent Increases in Sleep Spindle Density. *The Journal of Neuroscience*, 22 (2002): pp. 6830.
- Clemens, Z., D. Fabó, P. Halász. Overnight verbal memory retention correlates with the number of sleep spindles. *Neuroscience*, 132 (2005): pp. 529-535, DOI: https://doi.org/10.1016/j.neuroscience.2005.01.011.

- Clemens, Z., D. Fabo, P. Halasz. Twenty-four hours retention of visuospatial memory correlates with the number of parietal sleep spindles. *Neurosci Lett*, 403 (2006): pp. 52-6, DOI: 10.1016/j.neulet.2006.04.035.
- Schabus, M., G. Gruber, S. Parapatics, C. Sauter, G. Klösch, P. Anderer, W. Klimesch, B. Saletu, J. Zeitlhofer. Sleep Spindles and Their Significance for Declarative Memory Consolidation. *Sleep*, 27 (2004): pp. 1479-1485, DOI: 10.1093/sleep/27.7.1479.
- Schmidt, C., P. Peigneux, V. Muto, M. Schenkel, V. Knoblauch, M. Munch, D.J. de Quervain, A. Wirz-Justice, C. Cajochen. Encoding difficulty promotes postlearning changes in sleep spindle activity during napping. *J Neurosci*, 26 (2006): pp. 8976-82, DOI: 10.1523/jneurosci.2464-06.2006.
- 34. Göder, R., A. Graf, F. Ballhausen, S. Weinhold, P.C. Baier, K. Junghanns, A. Prehn-Kristensen. Impairment of sleep-related memory consolidation in schizophrenia: relevance of sleep spindles? *Sleep Medicine*, 16 (2015): pp. 564-569, DOI: https://doi.org/10.1016/j.sleep.2014.12.022.
- Peters, K.R., L. Ray, V. Smith, C. Smith. Changes in the density of stage 2 sleep spindles following motor learning in young and older adults. *Journal of Sleep Research*, 17 (2008): pp. 23-33, DOI: 10.1111/j.1365-2869.2008.00634.x.
- Fogel, S.M.,C.T. Smith. Learning-dependent changes in sleep spindles and Stage 2 sleep.
 Journal of Sleep Research, 15 (2006): pp. 250-255, DOI: 10.1111/j.1365-2869.2006.00522.x.
- 37. Nishida, M.,M.P. Walker. Daytime naps, motor memory consolidation and regionally specific sleep spindles. *PloS one*, 2 (2007): pp. e341-e341, DOI: 10.1371/journal.pone.0000341.

- Wei, Y., G.P. Krishnan, M. Komarov, M. Bazhenov. Differential roles of sleep spindles and sleep slow oscillations in memory consolidation. *PLOS Computational Biology*, 14 (2018): pp. e1006322, DOI: 10.1371/journal.pcbi.1006322.
- Cox, R., W.F. Hofman, M. de Boer, L.M. Talamini. Local sleep spindle modulations in relation to specific memory cues. *NeuroImage*, 99 (2014): pp. 103-110, DOI: https://doi.org/10.1016/j.neuroimage.2014.05.028.
- 40. Antony, J.W., M. Schonauer, B.P. Staresina, S.A. Cairney. Sleep Spindles and Memory Reprocessing. *Trends Neurosci* (2018), DOI: 10.1016/j.tins.2018.09.012.
- Jegou, A., M. Schabus, O. Gosseries, B. Dahmen, G. Albouy, M. Desseilles, V. Sterpenich, C. Phillips, P. Maquet, C. Grova, T.T. Dang-Vu. Cortical reactivations during sleep spindles following declarative learning. *Neuroimage*, 195 (2019): pp. 104-112, DOI: 10.1016/j.neuroimage.2019.03.051.
- Schabus, M., T.T. Dang-Vu, G. Albouy, E. Balteau, M. Boly, J. Carrier, A. Darsaud, C. Degueldre, M. Desseilles, S. Gais, C. Phillips, G. Rauchs, C. Schnakers, V. Sterpenich, G. Vandewalle, A. Luxen, P. Maquet. Hemodynamic cerebral correlates of sleep spindles during human non-rapid eye movement sleep. 104 (2007): pp. 13164-13169, DOI: 10.1073/pnas.0703084104 %J Proceedings of the National Academy of Sciences.
- 43. Rosanova, M.,D. Ulrich. Pattern-specific associative long-term potentiation induced by a sleep spindle-related spike train. *J Neurosci*, 25 (2005): pp. 9398-405, DOI: 10.1523/jneurosci.2149-05.2005.
- Lindemann, C., J. Ahlbeck, S.H. Bitzenhofer, I.L. Hanganu-Opatz. Spindle Activity Orchestrates Plasticity during Development and Sleep. *Neural Plast*, 2016 (2016): pp. 5787423, DOI: 10.1155/2016/5787423.

- 45. Levine, E.S., R.A. Crozier, I.B. Black, M.R. Plummer. Brain-derived neurotrophic factor modulates hippocampal synaptic transmission by increasing N-methyl-D-aspartic acid receptor activity. *Proc Natl Acad Sci U S A*, 95 (1998): pp. 10235-9.
- Caldeira, M.V., C.V. Melo, D.B. Pereira, R.F. Carvalho, A.L. Carvalho, C.B. Duarte. BDNF regulates the expression and traffic of NMDA receptors in cultured hippocampal neurons.
 Mol Cell Neurosci, 35 (2007): pp. 208-19, DOI: 10.1016/j.mcn.2007.02.019.
- Mednick, S.C., E.A. McDevitt, J.K. Walsh, E. Wamsley, M. Paulus, J.C. Kanady, S.P.A. Drummond. The critical role of sleep spindles in hippocampal-dependent memory: a pharmacology study. *The Journal of neuroscience : the official journal of the Society for Neuroscience*, 33 (2013): pp. 4494-4504, DOI: 10.1523/JNEUROSCI.3127-12.2013.
- Anderer, P., G. Klösch, G. Gruber, E. Trenker, R.D. Pascual-Marqui, J. Zeitlhofer, M.J. Barbanoj, P. Rappelsberger, B. Saletu. Low-resolution brain electromagnetic tomography revealed simultaneously active frontal and parietal sleep spindle sources in the human cortex. *Neuroscience*, 103 (2001): pp. 581-592, DOI: https://doi.org/10.1016/S0306-4522(01)00028-8.
- 49. Staresina, B.P., T.O. Bergmann, M. Bonnefond, R. van der Meij, O. Jensen, L. Deuker, C.E. Elger, N. Axmacher, J. Fell. Hierarchical nesting of slow oscillations, spindles and ripples in the human hippocampus during sleep. *Nature Neuroscience*, 18 (2015): pp. 1679, DOI: 10.1038/nn.4119
- Schofield, P.R., L.M. Williams, R.H. Paul, J.M. Gatt, K. Brown, A. Luty, N. Cooper, S. Grieve, C. Dobson-Stone, C. Morris, S.A. Kuan, E. Gordon. Disturbances in selective information processing associated with the BDNF Val66Met polymorphism: evidence from cognition, the P300 and fronto-hippocampal systems. *Biol Psychol*, 80 (2009): pp. 176-88, DOI: 10.1016/j.biopsycho.2008.09.001.

- 51. Thomason, M., D. Yoo, G. Glover, I. Gotlib. BDNF genotype modulates resting functional connectivity in children. 3 (2009), DOI: 10.3389/neuro.09.055.2009.
- 52. Wei, S.-M., D.P. Eisenberg, P.D. Kohn, J.S. Kippenhan, B.S. Kolachana, D.R. Weinberger, K.F. Berman. Brain-derived neurotrophic factor Val⁶ ⁶ Met polymorphism affects resting regional cerebral blood flow and functional connectivity differentially in women versus men. *The Journal of neuroscience : the official journal of the Society for Neuroscience*, 32 (2012): pp. 7074-7081, DOI: 10.1523/JNEUROSCI.5375-11.2012.
- 53. Strandberg, T.E., A.L. Jarvenpaa, H. Vanhanen, P.M. McKeigue. Birth outcome in relation to licorice consumption during pregnancy. *Am J Epidemiol*, 153 (2001): pp. 1085-8.
- 54. Kuula, L., A.K. Pesonen, I. Merikanto, M. Gradisar, J. Lahti, K. Heinonen, E. Kajantie, K. Raikkonen. Development of Late Circadian Preference: Sleep Timing From Childhood to Late Adolescence. *J Pediatr*, 194 (2018): pp. 182-189.e1, DOI: 10.1016/j.jpeds.2017.10.068.
- 55. Pesonen, A.K., S. Martikainen, K. Heinonen, K. Wehkalampi, J. Lahti, E. Kajantie, K. Raikkonen. Continuity and change in poor sleep from childhood to early adolescence. *Sleep*, 37 (2014): pp. 289-97, DOI: 10.5665/sleep.3400.
- 56. Lang, P.J., M.M. Bradley, B.N. Cuthbert, International Affective Picture System (IAPS): Instruction Manual and Affective Ratings. Technical Report A-6. 2005: University of Florida, Gainesvile, FL.
- Stanislaw, H.,N. Todorov. Calculation of signal detection theory measures. *Behav Res Methods Instrum Comput*, 31 (1999): pp. 137-49.
- 58. Ferrarelli, F., M.J. Peterson, S. Sarasso, B.A. Riedner, M.J. Murphy, R.M. Benca, P. Bria, N.H. Kalin, G. Tononi. Thalamic dysfunction in schizophrenia suggested by whole-night deficits in slow and fast spindles. *The American journal of psychiatry*, 167 (2010): pp. 1339-1348, DOI: 10.1176/appi.ajp.2010.09121731.

- 59. Hahn, M., A.-K. Joechner, J. Roell, M. Schabus, D.P.J. Heib, G. Gruber, P. Peigneux, K. Hoedlmoser. Developmental changes of sleep spindles and their impact on sleep-dependent memory consolidation and general cognitive abilities: A longitudinal approach. *Developmental Science*, 0 (2018): pp. e12706, DOI: 10.1111/desc.12706.
- Mölle, M., T.O. Bergmann, L. Marshall, J. Born. Fast and Slow Spindles during the Sleep Slow Oscillation: Disparate Coalescence and Engagement in Memory Processing. *Sleep*, 34 (2011): pp. 1411-1421, DOI: 10.5665/SLEEP.1290.
- Cox, R., A.C. Schapiro, D.S. Manoach, R. Stickgold. Individual Differences in Frequency and Topography of Slow and Fast Sleep Spindles. *Frontiers in human neuroscience*, 11 (2017): pp. 433-433, DOI: 10.3389/fnhum.2017.00433.
- 62. Iber, C., S. Ancoli-Israel, A.L. Chesson, S. Quan, The AASM Manual for the Scoring of Sleep and Associated Events: Rules, Terminology and Technical Specifications. 2007.
- 63. Wechsler, D., WAIS-III administration and scoring manual. 1997: The Psychological Corporation, San Antonio, TX.
- 64. Wagner, U., N. Kashyap, S. Diekelmann, J. Born. The impact of post-learning sleep vs. wakefulness on recognition memory for faces with different facial expressions. *Neurobiology of Learning and Memory*, 87 (2007): pp. 679-687, DOI: https://doi.org/10.1016/j.nlm.2007.01.004.
- Holz, J., H. Piosczyk, N. Landmann, B. Feige, K. Spiegelhalder, D. Riemann, C. Nissen, U. Voderholzer. The Timing of Learning before Night-Time Sleep Differentially Affects Declarative and Procedural Long-Term Memory Consolidation in Adolescents. *PLoS ONE*, 7 (2012): pp. e40963, DOI: 10.1371/journal.pone.0040963.
- 66. Petersen, A.C., L. Crockett, M. Richards, A. Boxer. A self-report measure of pubertal status: Reliability, validity, and initial norms. *Journal of Youth and Adolescence*, 17 (1988): pp. 117-133, DOI: 10.1007/BF01537962.

- 67. Petryshen, T.L., P.C. Sabeti, K.A. Aldinger, B. Fry, J.B. Fan, S.F. Schaffner, S.G.
 Waggoner, A.R. Tahl, P. Sklar. Population genetic study of the brain-derived neurotrophic factor (BDNF) gene. *Molecular psychiatry*, 15 (2010): pp. 810-815, DOI: 10.1038/mp.2009.24.
- Vulturar, R., A. Chiş, M. Hambrich, B. Kelemen, L. Ungureanu, A.C. Miu. Allelic distribution of BDNF Val66Met polymorphism in healthy Romanian volunteers. *Translational neuroscience*, 7 (2016): pp. 31-34, DOI: 10.1515/tnsci-2016-0006.
- 69. Anders, S., F. Eippert, N. Weiskopf, R. Veit. The human amygdala is sensitive to the valence of pictures and sounds irrespective of arousal: an fMRI study. *Social Cognitive and Affective Neuroscience*, 3 (2008): pp. 233-243, DOI: 10.1093/scan/nsn017 %J Social Cognitive and Affective Neuroscience.
- Styliadis, C., A.A. Ioannides, P.D. Bamidis, C. Papadelis. Amygdala responses to Valence and its interaction by arousal revealed by MEG. *Int J Psychophysiol*, 93 (2014): pp. 121-33, DOI: 10.1016/j.ijpsycho.2013.05.006.
- Hamann, S.B., T.D. Ely, S.T. Grafton, C.D. Kilts. Amygdala activity related to enhanced memory for pleasant and aversive stimuli. *Nature Neuroscience*, 2 (1999): pp. 289, DOI: 10.1038/6404.
- Montag, C., M. Reuter, B. Newport, C. Elger, B. Weber. The BDNF Val66Met polymorphism affects amygdala activity in response to emotional stimuli: evidence from a genetic imaging study. *Neuroimage*, 42 (2008): pp. 1554-9, DOI: 10.1016/j.neuroimage.2008.06.008.
- 73. Molendijk, M.L., M.J. van Tol, B.W.J.H. Penninx, N.J.A. van der Wee, A. Aleman, D.J. Veltman, P. Spinhoven, B.M. Elzinga. BDNF val66met affects hippocampal volume and emotion-related hippocampal memory activity. *Translational psychiatry*, 2 (2012): pp. e74-e74, DOI: 10.1038/tp.2011.72.

- Yonelinas, A.P. The Nature of Recollection and Familiarity: A Review of 30 Years of Research. *Journal of Memory and Language*, 46 (2002): pp. 441-517, DOI: https://doi.org/10.1006/jmla.2002.2864.
- 75. Aggleton, J.P.,M.W. Brown. Interleaving brain systems for episodic and recognition memory. *Trends in Cognitive Sciences*, 10 (2006): pp. 455-463, DOI: https://doi.org/10.1016/j.tics.2006.08.003.
- 76. McKay, N.S., D. Moreau, D.T. Henare, I.J. Kirk. The Brain-Derived Neurotrophic Factor Val66Met Genotype Does Not Influence the Grey or White Matter Structures Underlying Recognition Memory. (2019): pp. 461731, DOI: 10.1101/461731 %J bioRxiv.
- Kaestner, E.J., J.T. Wixted, S.C. Mednick. Pharmacologically increasing sleep spindles enhances recognition for negative and high-arousal memories. *J Cogn Neurosci*, 25 (2013): pp. 1597-610, DOI: 10.1162/jocn_a_00433.
- Hahn, M., A.-K. Joechner, J. Roell, M. Schabus, D.P. Heib, G. Gruber, P. Peigneux, K. Hoedlmoser. Developmental changes of sleep spindles and their impact on sleep-dependent memory consolidation and general cognitive abilities: A longitudinal approach. *Developmental science*, 22 (2019): pp. e12706-e12706, DOI: 10.1111/desc.12706.
- 79. Griessenberger, H., D.P.J. Heib, J. Lechinger, N. Luketina, M. Petzka, T. Moeckel, K. Hoedlmoser, M. Schabus. Susceptibility to Declarative Memory Interference is Pronounced in Primary Insomnia. *PLOS ONE*, 8 (2013): pp. e57394, DOI: 10.1371/journal.pone.0057394.
- Studte, S., E. Bridger, A. Mecklinger. Sleep spindles during a nap correlate with post sleep memory performance for highly rewarded word-pairs. *Brain and Language*, 167 (2017): pp. 28-35, DOI: https://doi.org/10.1016/j.bandl.2016.03.003.

- Mander, B.A., S. Santhanam, J.M. Saletin, M.P. Walker. Wake deterioration and sleep restoration of human learning. *Current Biology*, 21 (2011): pp. R183-R184, DOI: https://doi.org/10.1016/j.cub.2011.01.019.
- Mander, B.A., V. Rao, B. Lu, J.M. Saletin, S. Ancoli-Israel, W.J. Jagust, M.P. Walker. Impaired prefrontal sleep spindle regulation of hippocampal-dependent learning in older adults. *Cerebral cortex (New York, N.Y. : 1991)*, 24 (2014): pp. 3301-3309, DOI: 10.1093/cercor/bht188.
- 83. van der Helm, E., N. Gujar, M. Nishida, M.P. Walker. Sleep-Dependent Facilitation of Episodic Memory Details. *PLOS ONE*, 6 (2011): pp. e27421, DOI: 10.1371/journal.pone.0027421.
- Sopp, M.R., T. Michael, H.-G. Weeß, A.J.C. Mecklinger, Affective, B. Neuroscience.
 Remembering specific features of emotional events across time: The role of REM sleep and prefrontal theta oscillations. 17 (2017): pp. 1186-1209, DOI: 10.3758/s13415-017-0542-8.
- Turriziani, P., M. Oliveri, S. Salerno, F. Costanzo, G. Koch, C. Caltagirone, G.A.
 Carlesimo. Recognition memory and prefrontal cortex: dissociating recollection and familiarity processes using rTMS. *Behav Neurol*, 19 (2008): pp. 23-7.
- Murray, L.J.,C. Ranganath. The Dorsolateral Prefrontal Cortex Contributes to Successful Relational Memory Encoding. 27 (2007): pp. 5515-5522, DOI: 10.1523/JNEUROSCI.0406-07.2007 %J The Journal of Neuroscience.
- 87. Smirni, D., P. Turriziani, G.R. Mangano, L. Cipolotti, M. Oliveri. Modulating Memory Performance in Healthy Subjects with Transcranial Direct Current Stimulation Over the Right Dorsolateral Prefrontal Cortex. *PLoS One*, 10 (2015): pp. e0144838, DOI: 10.1371/journal.pone.0144838.
- 88. Turriziani, P., D. Smirni, M. Oliveri, C. Semenza, L. Cipolotti. The role of the prefrontal cortex in familiarity and recollection processes during verbal and non-verbal recognition

memory: An rTMS study. *NeuroImage*, 52 (2010): pp. 348-357, DOI: https://doi.org/10.1016/j.neuroimage.2010.04.007.

- Brewer, J.B., Z. Zhao, J.E. Desmond, G.H. Glover, J.D.E. Gabrieli. Making Memories:
 Brain Activity that Predicts How Well Visual Experience Will Be Remembered. 281 (1998):
 pp. 1185-1187, DOI: 10.1126/science.281.5380.1185 %J Science.
- 90. Aggleton, J.P.,M.W. Brown. Episodic memory, amnesia, and the hippocampal-anterior thalamic axis. *Behav Brain Sci*, 22 (1999): pp. 425-44; discussion 444-89.
- Andrade, K.C., V.I. Spoormaker, M. Dresler, R. Wehrle, F. Holsboer, P.G. Samann, M. Czisch. Sleep spindles and hippocampal functional connectivity in human NREM sleep. *J Neurosci*, 31 (2011): pp. 10331-9, DOI: 10.1523/jneurosci.5660-10.2011.
- 92. Helfrich, R.F., J.D. Lendner, B.A. Mander, H. Guillen, M. Paff, L. Mnatsakanyan, S. Vadera, M.P. Walker, J.J. Lin, R.T. Knight. Bidirectional prefrontal-hippocampal dynamics organize information transfer during sleep in humans. *Nature Communications*, 10 (2019): pp. 3572, DOI: 10.1038/s41467-019-11444-x.
- Hoedlmoser, K., D.P.J. Heib, J. Roell, P. Peigneux, A. Sadeh, G. Gruber, M. Schabus. Slow Sleep Spindle Activity, Declarative Memory, and General Cognitive Abilities in Children. *Sleep*, 37 (2014): pp. 1501-1512, DOI: 10.5665/sleep.4000.
- 94. Schabus, M., K. Hodlmoser, G. Gruber, C. Sauter, P. Anderer, G. Klosch, S. Parapatics, B. Saletu, W. Klimesch, J. Zeitlhofer. Sleep spindle-related activity in the human EEG and its relation to general cognitive and learning abilities. *Eur J Neurosci*, 23 (2006): pp. 1738-46, DOI: 10.1111/j.1460-9568.2006.04694.x.
- 95. Studte, S., E. Bridger, A. Mecklinger. Nap sleep preserves associative but not item memory performance. *Neurobiology of Learning and Memory*, 120 (2015): pp. 84-93, DOI: https://doi.org/10.1016/j.nlm.2015.02.012.

- Mohn, C., K. Sundet, B.R. Rund. The relationship between IQ and performance on the MATRICS consensus cognitive battery. *Schizophrenia research. Cognition*, 1 (2014): pp. 96-100, DOI: 10.1016/j.scog.2014.06.003.
- 97. Jensen, A.R. The relationship between learning and intelligence. *Learning and Individual Differences*, 1 (1989): pp. 37-62, DOI: https://doi.org/10.1016/1041-6080(89)90009-5.
- 98. Ujma, P. Sleep spindles and general cognitive ability A meta-analysis. *Sleep Spindles and Cortical Up States*, 0 (2018): pp. 1-17, DOI: 10.1556/2053.2.2018.01.
- Ackermann, S., F. Hartmann, A. Papassotiropoulos, D.J. de Quervain, B. Rasch. No Associations between Interindividual Differences in Sleep Parameters and Episodic Memory Consolidation. *Sleep*, 38 (2015): pp. 951-9, DOI: 10.5665/sleep.4748.
- Piantoni, G., S.S. Poil, K. Linkenkaer-Hansen, I.M. Verweij, J.R. Ramautar, E.J. Van Someren, Y.D. Van Der Werf. Individual differences in white matter diffusion affect sleep oscillations. *J Neurosci*, 33 (2013): pp. 227-33, DOI: 10.1523/jneurosci.2030-12.2013.
- Pezawas, L., B.A. Verchinski, V.S. Mattay, J.H. Callicott, B.S. Kolachana, R.E. Straub,
 M.F. Egan, A. Meyer-Lindenberg, D.R. Weinberger. The Brain-Derived Neurotrophic
 Factor val66met Polymorphism and Variation in Human Cortical Morphology. 24 (2004):
 pp. 10099-10102, DOI: 10.1523/JNEUROSCI.2680-04.2004 %J The Journal of
 Neuroscience.
- 102. Legge, R.M., S. Sendi, J.H. Cole, S. Cohen-Woods, S.G. Costafreda, A. Simmons, A.E. Farmer, K.J. Aitchison, P. McGuffin, C.H.Y. Fu. Modulatory effects of brain-derived neurotrophic factor Val66Met polymorphism on prefrontal regions in major depressive disorder. *The British journal of psychiatry : the journal of mental science*, 206 (2015): pp. 379-384, DOI: 10.1192/bjp.bp.113.143529.
- 103. Kim, S.N., D.-H. Kang, J.-Y. Yun, T.Y. Lee, W.H. Jung, J.H. Jang, J.S. Kwon. Impact of the BDNF Val66Met Polymorphism on Regional Brain Gray Matter Volumes: Relevance to

the Stress Response. *Psychiatry investigation*, 10 (2013): pp. 173-179, DOI: 10.4306/pi.2013.10.2.173.

- Bueller, J.A., M. Aftab, S. Sen, D. Gomez-Hassan, M. Burmeister, J.K. Zubieta. BDNF
 Val66Met allele is associated with reduced hippocampal volume in healthy subjects. *Biol Psychiatry*, 59 (2006): pp. 812-5, DOI: 10.1016/j.biopsych.2005.09.022.
- 105. Szeszko, P.R., R. Lipsky, C. Mentschel, D. Robinson, H. Gunduz-Bruce, S. Sevy, M. Ashtari, B. Napolitano, R.M. Bilder, J.M. Kane, D. Goldman, A.K. Malhotra. Brain-derived neurotrophic factor val66met polymorphism and volume of the hippocampal formation. *Mol Psychiatry*, 10 (2005): pp. 631-6, DOI: 10.1038/sj.mp.4001656.
- Ziegler, E., A. Foret, L. Mascetti, V. Muto, A. Le Bourdiec-Shaffii, J. Stender, E. Balteau,
 V. Dideberg, V. Bours, P. Maquet, C. Phillips. Altered white matter architecture in BDNF
 met carriers. *PloS one*, 8 (2013): pp. e69290-e69290, DOI: 10.1371/journal.pone.0069290.
- 107. Cao, L., A. Dhilla, J. Mukai, R. Blazeski, C. Lodovichi, C.A. Mason, J.A. Gogos. Genetic modulation of BDNF signaling affects the outcome of axonal competition in vivo. *Current biology : CB*, 17 (2007): pp. 911-921, DOI: 10.1016/j.cub.2007.04.040.
- 108. Singh, K.K., K.J. Park, E.J. Hong, B.M. Kramer, M.E. Greenberg, D.R. Kaplan, F.D. Miller. Developmental axon pruning mediated by BDNF-p75NTR–dependent axon degeneration. *Nature Neuroscience*, 11 (2008): pp. 649, DOI: 10.1038/nn.2114
- 109. Gaudreault, P.-O., N. Gosselin, M. Lafortune, S. Deslauriers-Gauthier, N. Martin, M. Bouchard, J. Dubé, J.-M. Lina, J. Doyon, J. Carrier. The association between white matter and sleep spindles differs in young and older individuals. *Sleep*, 41 (2018): pp. zsy113-zsy113, DOI: 10.1093/sleep/zsy113.
- 110. Mander, B.A., A.H. Zhu, J.R. Lindquist, S. Villeneuve, V. Rao, B. Lu, J.M. Saletin, S. Ancoli-Israel, W.J. Jagust, M.P. Walker. White Matter Structure in Older Adults Moderates the Benefit of Sleep Spindles on Motor Memory Consolidation. *The Journal of neuroscience*

: the official journal of the Society for Neuroscience, 37 (2017): pp. 11675-11687, DOI: 10.1523/JNEUROSCI.3033-16.2017.

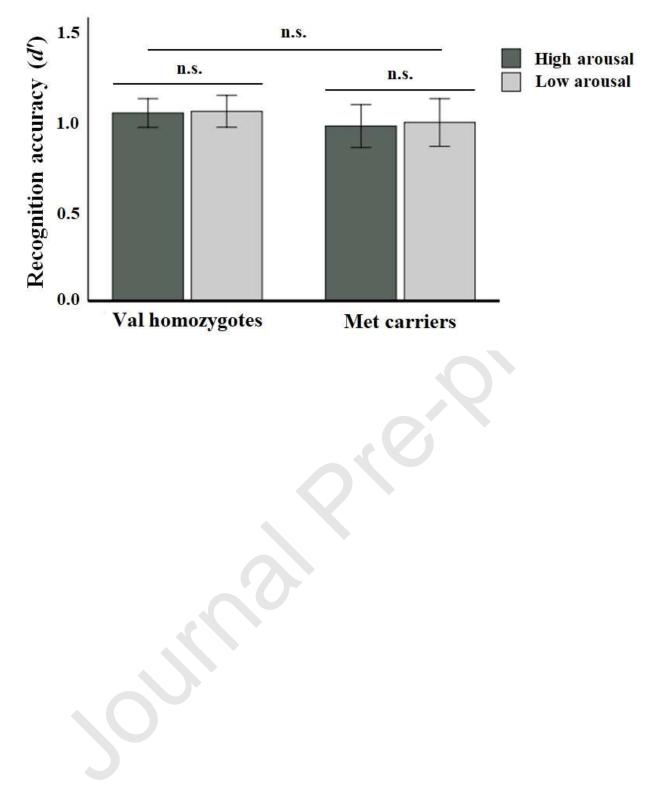
- 111. Chiang, M.-C., M. Barysheva, A.W. Toga, S.E. Medland, N.K. Hansell, M.R. James, K.L. McMahon, G.I. de Zubicaray, N.G. Martin, M.J. Wright, P.M. Thompson. BDNF gene effects on brain circuitry replicated in 455 twins. *NeuroImage*, 55 (2011): pp. 448-454, DOI: 10.1016/j.neuroimage.2010.12.053.
- 112. Huang, C.-C., M.-E. Liu, K.-H. Chou, A.C. Yang, C.-C. Hung, C.-J. Hong, S.-J. Tsai, C.-P. Lin. Effect of BDNF Val66Met polymorphism on regional white matter hyperintensities and cognitive function in elderly males without dementia. *Psychoneuroendocrinology*, 39 (2014): pp. 94-103, DOI: https://doi.org/10.1016/j.psyneuen.2013.09.027.
- Helfrich, R.F., B.A. Mander, W.J. Jagust, R.T. Knight, M.P. Walker. Old Brains Come Uncoupled in Sleep: Slow Wave-Spindle Synchrony, Brain Atrophy, and Forgetting. *Neuron*, 97 (2018): pp. 221-230.e4, DOI: 10.1016/j.neuron.2017.11.020.
- 114. Mikutta, C., B. Feige, J.G. Maier, E. Hertenstein, J. Holz, D. Riemann, C. Nissen. Phaseamplitude coupling of sleep slow oscillatory and spindle activity correlates with overnight memory consolidation. 0, DOI: 10.1111/jsr.12835.
- 115. Weaver, K.E., J.D. Wander, A.L. Ko, K. Casimo, T.J. Grabowski, J.G. Ojemann, F. Darvas. Directional patterns of cross frequency phase and amplitude coupling within the resting state mimic patterns of fMRI functional connectivity. *NeuroImage*, 128 (2016): pp. 238-251, DOI: 10.1016/j.neuroimage.2015.12.043.
- Lago-Fernandez, L.F., F.J. Corbacho, R. Huerta. Connection topology dependence of synchronization of neural assemblies on class 1 and 2 excitability. *Neural Netw*, 14 (2001): pp. 687-96.

- 117. Kitano, K.,T. Fukai. Variability v.s. synchronicity of neuronal activity in local cortical network models with different wiring topologies. *J Comput Neurosci*, 23 (2007): pp. 237-50, DOI: 10.1007/s10827-007-0030-1.
- Fujisawa, S., M.K. Yamada, N. Nishiyama, N. Matsuki, Y. Ikegaya. BDNF boosts spike fidelity in chaotic neural oscillations. *Biophysical journal*, 86 (2004): pp. 1820-1828, DOI: 10.1016/S0006-3495(04)74249-6.
- 119. Gosselin, N., L. De Beaumont, K. Gagnon, A.A. Baril, V. Mongrain, H. Blais, J. Montplaisir, J.F. Gagnon, S. Pelleieux, J. Poirier, J. Carrier. BDNF Val66Met
 Polymorphism Interacts with Sleep Consolidation to Predict Ability to Create New
 Declarative Memories. *J Neurosci*, 36 (2016): pp. 8390-8, DOI: 10.1523/jneurosci.4432-15.2016.
- Grant, L.K., S.W. Cain, A.-M. Chang, R. Saxena, C.A. Czeisler, C. Anderson. Impaired cognitive flexibility during sleep deprivation among carriers of the Brain Derived Neurotrophic Factor (BDNF) Val66Met allele. *Behavioural brain research*, 338 (2018): pp. 51-55, DOI: 10.1016/j.bbr.2017.09.025.
- 121. Konrad, K., C. Firk, P.J. Uhlhaas. Brain development during adolescence: neuroscientific insights into this developmental period. *Deutsches Arzteblatt international*, 110 (2013): pp. 425-431, DOI: 10.3238/arztebl.2013.0425.
- 122. Fair, D.A., D. Bathula, K.L. Mills, T.G.C. Dias, M.S. Blythe, D. Zhang, A.Z. Snyder, M.E. Raichle, A.A. Stevens, J.T. Nigg, B.J. Nagel. Maturing thalamocortical functional connectivity across development. *Frontiers in systems neuroscience*, 4 (2010): pp. 10-10, DOI: 10.3389/fnsys.2010.00010.
- 123. Goldstone, A., A.R. Willoughby, M. de Zambotti, D.B. Clark, E.V. Sullivan, B.P. Hasler,P.L. Franzen, D.E. Prouty, I.M. Colrain, F.C. Baker. Sleep spindle characteristics in

adolescents. *Clinical Neurophysiology*, 130 (2019): pp. 893-902, DOI: https://doi.org/10.1016/j.clinph.2019.02.019.

 Yonelinas, A.P., L.J. Otten, K.N. Shaw, M.D. Rugg. Separating the Brain Regions Involved in Recollection and Familiarity in Recognition Memory. 25 (2005): pp. 3002-3008, DOI: 10.1523/JNEUROSCI.5295-04.2005 %J The Journal of Neuroscience.





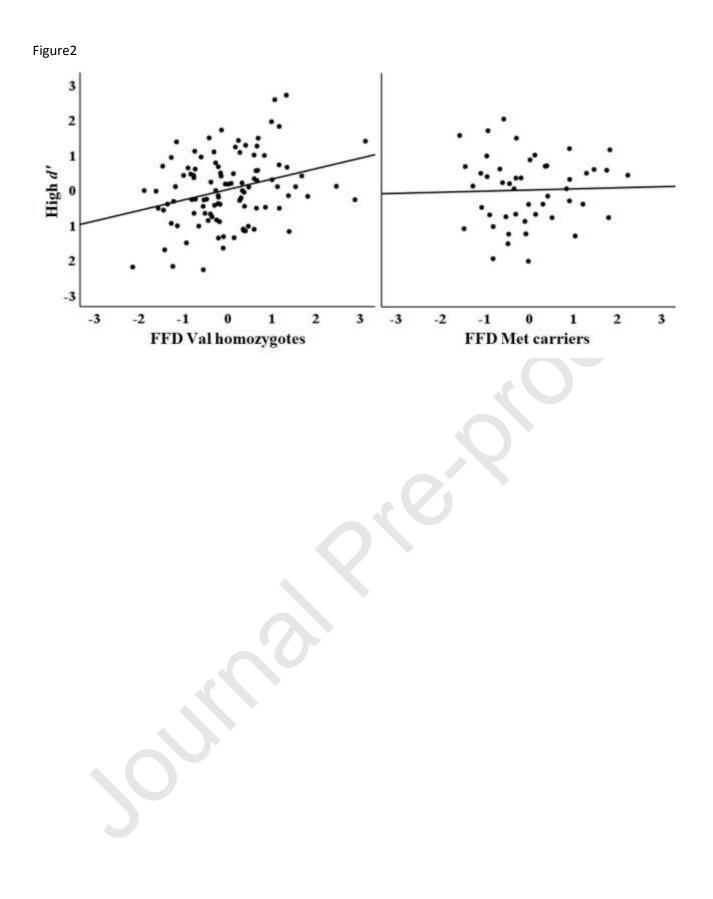


Table 1 presents the age, pubertal development, FSIQ, sleep measures and spindle densities. No differences in the variables emerged between the genotype groups (Val homozygotes and Met carriers). Furthermore, spindle duration, amplitude and frequency showed no significant differences between the genotypes in N2 and N3 spindles. (all p values \geq .11; see Supplementary Material 1).

	Val homozygotes			Met carriers			
	Mean	Range	SD	Mean	Range	SD	р
Age	16.9	16.6–17.2	0.1	16.9	16.7–17.2	0.1	.53
Pubertal development	3.2	2.4-3.8	0.4	3.3	2.2–3.8	0.4	.79
FSIQ	0.0	-1.4-1.5	0.6	-0.1	-2.5-0.9	0.7	.24
Sleep variables							
Total Sleep Time (hh:mm)	7:41	3:10-9:52	1:02	7:29	3:49-10:46	1:22	.33
N1 %	10.4	3.0-27.0	4.4	9.8	2.7-18.0	4.3	.47
N2 %	38.3	23.2-51.2	6.0	37.9	20.5-50.6	6.3	.76
N3 %	25.0	12.4–39.6	5.7	26.0	15.7–39.6	6.3	.36
REM %	19.7	4.4-30.9	5.1	18.7	4.3–28.4	4.9	.25
Sleep efficiency (%)	93.4	70.7–98.4	4.8	93.1	60.8–98.9	6.9	.34
N2 Spindle density (n/min)							
Central slow	3.4	2.3–4.6	0.6	3.6	2.2-4.6	0.7	.23
Frontal slow	4.4	1.6–6.3	0.8	4.5	2.9-6.2	0.7	.39
Central fast	3.6	1.8–6.2	0.9	3.5	2.1-5.7	0.9	.65
Frontal fast	2.9	0.9–5.7	0.9	3.0	1.6-4.8	0.8	.79
N3 spindle density (n/min)							
Central slow	2.4	0.6–3.8	0.6	2.5	1.1–3.2	0.8	.20
Frontal slow	3.3	1.0-5.5	0.9	3.3	1.5-6.3	1.0	.85
Central fast	3.6	1.5–5.9	0.9	3.5	1.7–5.1	0.8	.87
Frontal fast	2.5	1.0-5.6	0.8	2.5	1.3–5.1	0.9	.86

Table 1. Sample Characteristics.

Pubertal development estimated using the Pubertal Development Scale (PDS) [66]. FSIQ = Full-Scale Intelligence

Quotient. REM = Rapid Eye Movement sleep; N1-N3 = stages of non-REM sleep.

Spindle	Genotype							Genotype
density	All	VH	MZ	x spindle	All	VH	MZ	x spindle
High d'	B (SE)	B (SE)	B (SE)	F	B (SE)	B (SE)	B (SE)	F
Central slow	0.02 (.06)	0.07 (.07)	-0.04 (.11)	0.55	0.01 (.06)	0.05 (.07)	-0.04 (.11)	0.33
Frontal slow	0.09* (.04)	0.11* (.04)	0.07 (.10)	3.3	0.08 (.04)	0.10* (.05)	0.03 (.10)	1.03
Central fast	0.08 (.04)	0.07 (.05)	0.11 (.10)	1.83	0.07 (.04)	0.06 (.05)	0.11 (.10)	0.31
Frontal fast	0.09* (.04)	0.12** (.04)	0.01 (.09)	3.89*	0.09* (.04)	0.12** (.04)	0.02 (.09)	4.66*
Low d'	В	В	В	F	В	В	В	F
Central slow	0.06 (.07)	0.11 (.09)	0.00 (.11)	0.54	0.05 (.07)	0.06 (.08)	0.00 (.11)	0.23
Frontal slow	0.10** (.05)	0.15** (.06)	0.07 (.10)	0.72	0.10* (.05)	0.11* (.05)	0.03 (.10)	1.02
Central fast	0.07 (.05)	0.06 (.06)	0.07 (.10)	0.01	0.05 (.05)	0.05 (.05)	0.07 (.10)	0.00
Frontal fast	0.06 (.05)	0.10 (.05)	-0.07 (.09)	2.31	0.06 (.04)	0.10* (.05)	-0.05 (.09)	3.23

Table 2. Regression and interaction analyses between picture learning scores and N2 spindle densities.

B = Regression analysis coefficient B for spindle density variables in the entire sample (All) and separately for Val homozygotes (VH) and Met Carriers (MC). SE = Standard Error. F = F-value of the interaction term 'genotype x spindle'. High d' = recognition accuracy as d' for high arousal pictures. Low d' = recognition accuracy as d' for low arousal pictures. Model 1 covariates: sex. Model 2 covariates: sex. FSIQ, total time awake and sleep duration. * = p < .05, ** = p < .01.