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Array of Microfluidic Beam Resonators for Density and Viscosity Analysis of Liquids

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Abstract—This paper reports on the design, fabrication and evaluation of a mass density and viscosity sensor based on an array of polysilicon microbeam resonators integrated with 20 pL fluidic microchannels. When filled with water, resonators exhibit resonant frequencies close to 500 KHz and Q-factor values of 400 operating at atmospheric pressure and ambient temperature. Real-time measurements are highly reproducible and only require 250 μ L of the sample fluid. The built-in interferometric readout enables automatic detection of the beams, increasing the throughput analysis and reducing detection times. The frequency shift response shows a linear behavior in accordance with the density of evaluated solvents, organic solutions and alcoholic drinks, reporting a mass responsivity of 7.4 Hz/pg. Also, the sensor is capable of measuring the viscosity of liquid phase samples with a resolution of 0.15 cP by tracking the Q-factor response of the sensor within a linear regime between 1 cP to 2.6 cP. This approach demonstrates the ability to identify in real-time changes of fluids in the liquid phase that could provide a valuable assessment for bioanalytical applications.

Index Terms—Array, density, liquids, microfluidic channels, Q-factor, resonators, resonant frequency, viscosity.

I. INTRODUCTION

IDENTIFICATION of changes in fluid properties of samples is essential for a wide range of industrial and medical applications. Density and rheological properties of fluids, such as viscosity, are employed to detect food quality [1]–[2], DNA solutions [3] and even single cells [4]. However, current analytical methods generally require large volume samples, need long evaluation times and have a limited sensitivity range [5],[6]. Over the last decade, microelectromechanical sensors (MEMS) have become one of the most promising tools for the achievement of high sensitive sensors due to their miniature size, low mass, and compatibility with CMOS (complementary metal-oxide semiconductor) technologies [7],[8]. The scope of demonstrated applications of MEMS devices for density and rheological parameters of fluids includes: viscoelastic properties

of blood and plasma coagulation [10],[11], viscosity measurements of hydrocarbons, silicone oils, oil/fuel mixtures [11]–[13] and gases [14], characterization of polymer solutions [15], concentration of sugar mixtures [16] and ethanol solutions [1],[17], among others. However, when a MEMS resonator is immersed in a viscous fluid, the overdamped response of the resonator produces a rapid dissipation of energy degrading the sensitivity of the sensor with respect to the viscosity of the medium.

An innovative approach to solve this limitation was proposed by Burg et al. [18]. This approach involved integrating a fluidic channel into a suspended microcantilever, namely suspended microchannel resonator (SMR), thereby avoiding damping and viscous drag produced by the fluid environment. Since then, various solutions have been proposed to improve the sensitivity of SMR devices either by decreasing their effective mass or by proposing different materials and designs on their geometry. For example, Khan et. al [13] used silicon-rich nitride (SRN) as structural material to build transparent microchannels that facilitated the visual inspection of processes taking place inside the channels. This approach further derived density and viscosity measurements in liquid phase reagents. Suspended resonators have also been fabricated based on embedded microchannels in plate Lamé resonators to exhibit higher frequency responses and Q-factor values without the need for vacuum and packaging [19]. Another solution reported the fabrication of a suspended doubly clamped beam sensor at nanoscale dimensions with the aim of reducing the effective mass of the resonator [20]. Despite these promising advantages, reducing the mass and size of resonators in order to achieve sensitive transducers complicates the fabrication process. Furthermore, kinetics plays an important role in these type of devices due to the maximum volume of fluid that can flow through the microchannels and the maximum achievable flow rates to optimize the detection times [21]. Whereas a single SMR device is highly sensitive and suitable for identifying individual reagents, a device approach that includes

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This paper has supplementary downloadable material that consists of a video.

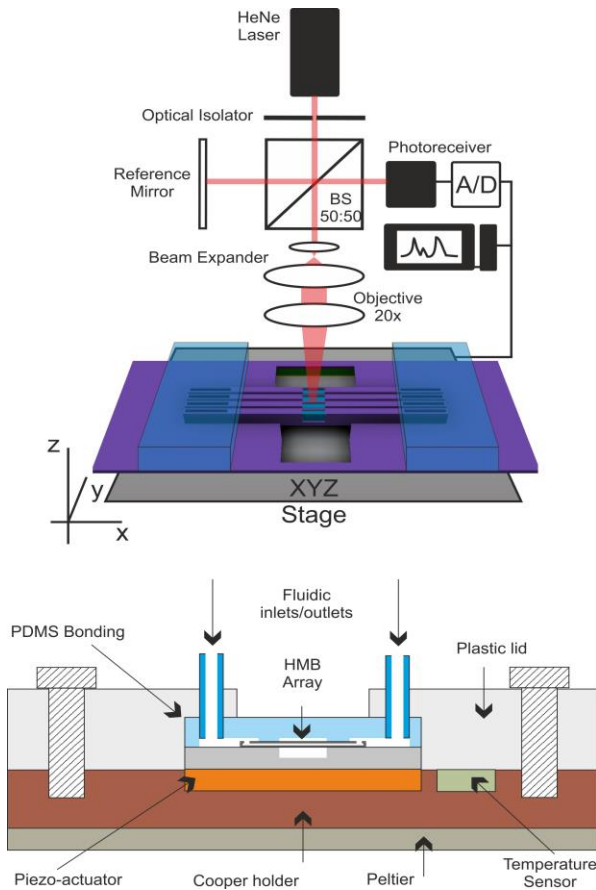


Fig. 1 (Top) Schematic view of the HMB sensor and the experimental setup. A custom free space interferometer acquires the driven excitation response of the resonators while a sample solution is streaming through the embedded microchannels. During experiments, temperature was fixed at $23^{\circ}\text{C} \pm 0.1$. (Bottom) Details of the copper fixture housing the Peltier module and the piezo-ceramic actuator for driving the excitation of the resonators.

1 an array of resonators with single inlet/outlet embedded
 2 microchannels can constitute an efficient sensing platform for
 3 reducing sample evaluation times. Besides, this new approach
 4 can be used for the simultaneous detection of different samples
 5 in real-time.

6 Therefore, in our approach, we have developed a mass density
 7 and viscosity sensor for the real-time identification of Newtonian
 8 fluids in the liquid phase using an array of suspended polysilicon
 9 microbeams, namely Hollow Microbeam (HMB) resonators (see
 10 Fig. 1). The sensor consists of four closely spaced doubly
 11 clamped beams with embedded microfluidic channels where
 12 each microchannel has a volume of around 20 pL. The solution
 13 drawn up in our work includes four main contributions:
 14 fabrication of a sensor array of resonators with embedded
 15 microchannels, implementation of an experimental readout for
 16 automatic detection, increase of the precision of rheological
 17 measurements of liquids, and usage of polysilicon as main
 18 structural material of the sensor. As proof of concept, we have
 19 studied the sensor performance in a flow-through detection mode
 20 to measure density and viscosity of different solvents and
 21 organic solutions. Also, we have analyzed commercial alcoholic
 22 beverages demonstrating that the HMB resonators can reduce to

23 7 min the time employed to measure and validate both the
 24 density and viscosity of liquids (17 min) using conventional
 25 sensors such as pycnometers and U-tube viscometers.

II. PRINCIPLE OF OPERATION

26 For the operation of the HMB sensor, we evaluated the shift
 27 in the resonant frequency of the resonator due to changes in its
 28 effective mass when a sample fluid of specific density is
 29 streaming through the embedded microchannel. The frequency
 30 behavior of a doubly clamped beam with an embedded
 31 microchannel is modeled as a lump-parameter resonator
 32 according to the Euler-Bernoulli beam theory [28]. The model is
 33 valid under the following assumptions: 1) the material
 34 composition of the resonators is uniformly distributed along the
 35 length of the structures and the cross-section geometry of the
 36 microchannels is constant, 2) the fluid filling of embedded
 37 microchannels does not change the elastic constant of the
 38 resonators and, 3) shearing deformation of the doubly clamped
 39 beams is negligible. For low-damping values, the resonant
 40 frequency of a resonator (ω_r) can be approximated to its natural
 41 frequency (ω_0), as $\omega_r \approx \omega_0$. Then, the resonant frequency of a
 42 linear resonator with elastic constant (k_b) and effective mass (m_b)
 43 is:
 44

$$\omega_r^2 = \frac{k_b}{m_b} \quad (1)$$

45 Here, the elastic constant is defined as $k_b = 192EI/L^3$ where E
 46 is the Young Modulus, I is the moment of inertia of the empty
 47 beam with respect to y -axis and L is the beam length,
 48 respectively. The moment of inertia is defined as $I = (w_b h_b^3 -$
 49 $w_f h_f^3)/12$ being w_f and h_f , the width and height of the embedded
 50 microchannel, and w_b and h_b , the width and height of the beam,
 51 respectively. The total effective mass of the resonator is a
 52 contribution of not only the structural material but also of the
 53 added mass induced by the sample fluid. For this reason, we
 54 modeled the resonators as a multimorph doubly clamped beams
 55 formed by equal length layers in which the effective mass is
 56 defined as [23]:
 57

$$m_b = \frac{26}{70} L \sum_{i=1}^n \rho_i A_i \quad (2)$$

58 where ρ_i is the density of the layer “ i ” with its corresponding
 59 cross-sectional area A_i . By substituting the lump-parameter
 60 definitions of k_b and m_b into (1), the approximated fundamental
 61 resonant frequency of the HMB resonator is:
 62

$$f_{r,b} = \frac{22.736 h_b}{2\pi L^2} \sqrt{\frac{E}{12[\alpha\rho_f + (1-\alpha)\rho_b]}} \quad (3)$$

63 where ρ_b and ρ_f are the structural material and sample fluid
 64 densities, respectively. The parameter α is the ratio of the
 65 microfluidic channel surface to the beam cross-sectional area
 66 defined as $\alpha = w_f h_f / w_b h_b$. By monitoring the HMB resonant

frequency response, we correlated the frequency changes as a function of the sample fluid density.

The quality factor (Q) defines the ratio of stored energy to the dissipated energy on each oscillation cycle of a resonator. For SMR devices, the dissipation of energy is mainly attributed to the shearing of the contained fluid that is able to freely move inside the channel. Besides, as damping is a non-monotonic function of the sample viscosity inside of SMR resonators [24], the Q-factor value can increase or decrease with respect to sample viscosity [25]. Since the embedded microchannel is centered about the beam neutral axis in our approach, the magnitude of the shearing effect can be determined by the dimensionless frequency number $\beta = \rho_{\text{fluid}} \omega h_{\text{fluid}}^2 / \mu$, where h_{fluid} is the channel height, μ is the dynamic viscosity, ρ_{fluid} is the fluid density and ω is the resonator frequency, respectively. Sader et al. [25] derived a non-monotonic function $F(\beta)$ to understand the effect of the energy dissipation of SMR devices as a function of the β number. For the fundamental frequency mode of resonators, the non-monotonic function $F(\beta) \approx 0.152\sqrt{\beta} + 38.7/\beta$ [24] defines two flow transition regimes at its minimum ($\beta=46$). For $\beta < 46$, there is a low-inertia regime whereas for $\beta > 46$, fluid inertia dominates. The dimensionless function $F(\beta)$ yields a maximum error estimation of 13% for all β . According to the fabricated dimensions of our resonators and the used liquid samples, low-inertia regime dominates in the HMB devices. By tracking the quality factor of the resonators while streaming different liquid samples, we analyzed the sensor response to viscosity changes.

Another important design parameter of the HMB resonators is related to the limits of pressure at both ends of the microchannel inlets in order to avoid the collapsing of the inner channel walls. A reasonable approximation with less than 10% error, for aspect ratios of $h/w \leq 0.7$, introduces a pressure difference of [26]:

$$\Delta P \left[1 - \frac{6(2^5) h_f}{\pi^5 w_f} \right] = \frac{12 \mu L}{w h_f^3} Q_{\text{flow}} \quad (4)$$

where ΔP is the pressure difference along the microfluidic channel; μ is the water viscosity (1 cP) and Q_{flow} defines the flow rate ($\mu\text{L}/\text{min}$), respectively. According to this relationship, we can theoretically inject solutions at a pressure difference of up to 7 MPa, yielding flow rates of up to 45 $\mu\text{L}/\text{min}$.

III. FABRICATION AND CHARACTERIZATION

A. Device Design and Fabrication

Four embedded microfluidic channels of 1300 μm in length and cross-sectional area of 20 μm x 4 μm integrate the array of nominally identical resonators, with a distance between each other of 13 μm , as Fig. 2 shows. This design establishes very short separation distances between the hollow doubly clamped beams so that the response of the sensors can be acquired by the optical readout system. The microchannel wall thickness is 1 μm . The effective length of each resonator (about 275 μm) is set at each clamped side of a rectangular trench of 275 μm x 7500 μm . Fluidic inlets of 10 μm x 100 μm in dimension are located

at both end sides of the microchannels. To allow fluid exchange inside of the embedded microchannels, two polydimethylsiloxane (PDMS) delivery channels of cross-sectional area of 200 μm x 35 μm and length of 1.5 mm are integrated on both sides of the inlets of HMB resonators. The serpentine design of the PDMS fluid delivery channels is done in order to facilitate the visual inspection of the meniscus of the liquids during the filling step of the HMB sensors.

Hollow microbridges were fabricated using 4-in, type p silicon wafers of 500 μm thickness. Polysilicon was used as structural layer and boron phosphorus doped silicon oxide (BPSG) as sacrificial layer because of its high selectivity to polysilicon and high etching rate with HF 49%. Thereafter, a 1 μm layer of polysilicon was deposited by low-pressure chemical vapor deposition (LPCVD) at 580°C and 380 mtorr. The 4 μm topology of microfluidic channels was patterned by reactive ion etching and hard contact photolithography. As follows, another layer of 1 μm polysilicon was coated in order to enclose the microchannels. Access holes, located at both ends of the structures were etched on top of the polysilicon layer in order to

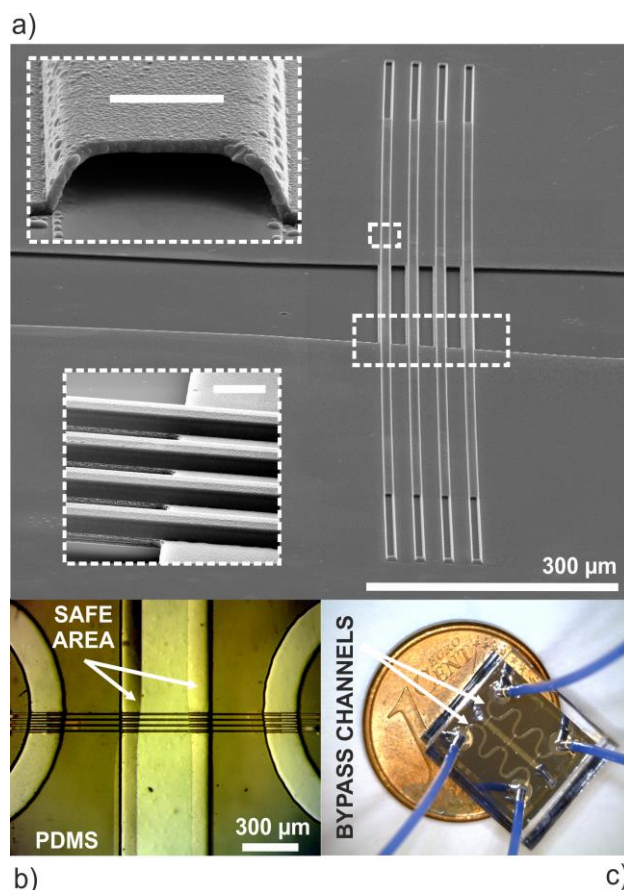


Fig. 2 a) Scanning electron microscope image of an array of four HMB resonators; inset 1 shows the cross section of an embedded microchannel of 20 μm x 4 μm with 1 μm thick polysilicon structural layer; inset 2 illustrates the 4 μm topology and lateral distance of 13 μm between the suspended array of resonators. Scale bar represents 10 μm . b) Optical microscope image illustrating the PDMS bypass channels connecting with the inlets of the HMB resonators. A safe area was set along the edges of the sensors (50 μm) to avoid any leakage of uncured PDMS onto the structures. c) Chip with integrated microfluidics and external tubing connections for injection of sample solutions (dimensions 1 cm x 1 cm).

1 dissolve the sacrificial layer. Also, in this step, the length of the
2 resonators was defined. Using HF 49%, up to 1300 μm long
3 channels were emptied after approximately 25 min of isotropic
4 wet etching. The resonators were released in this procedure by
5 dissolving the SiO_2 PECVD TEOS 2:1 beneath the polysilicon
6 microbeams at a lower etching rate than the BPSG sacrificial
7 layer. Finally, the wafer was manually diced to have chips of 63
8 cm x 1 cm. A scanning electron image of the final array of double
9 clamped beams is illustrated in Fig. 2a. Further details of device
10 fabrication are described in [27]. 66

11 For injection of liquids inside the array of resonators, PDMS
12 bypass channels were replicated from a master mold of SU-8
13 negative photoresist (Microchem SU-8 2025). The master mold
14 was fabricated by standard photolithography [28]. First, the
15 photoresist was spun onto a silicon substrate to obtain a 35 μm
16 thick layer and by soft-photolithography, the bypass channels
17 were patterned. The 4 mm thick PDMS replica was prepared by
18 a 10:1 ratio of elastomer and curing agent from "Sylgard 1844
19 Silicon elastomer Kit". PDMS was cured in a hot plate at 80° C
20 for 2 hrs. Thereafter, four through holes were perforated on each
21 PDMS reservoir of the bypass channels with a biopsy punch
22 (Harris Uni-Core™ 1 mm I.D.) for external access of tubing
23 connections and using a razor blade the PDMS was cut to have
24 dimensions of 1 cm x 1 cm. Permanent bonding of PDMS to the
25 devices was done by a stamp-and-stick technique in which
26 uncured PDMS of 10:3 ratio was used as an adhesive [29].
27 Applying soft pressure to make the bonding process faster
28 resulted in reflow of the uncured polymer. Also, any increase in
29 temperature caused clogging of the microchannels. Once the
30 uncured PDMS covered all over the surface, except those areas
31 corresponding to the microfluidic channels and the array of
32 resonators, the 10:3 PDMS was let to cure for 48 h at room
33 temperature. This methodology yielded a good sealing of the
34 topology of the structures and prevented any structural damage
35 to the resonators as shown in Fig. 2b. The maximum pressure
36 that this permanent union can withstand is up to 38 psi, which is
37 adequate for flow rates below 100 $\mu\text{l}/\text{min}$. Finally, four PEEK
38 tubes (Valco Instruments Co. Inc. JR-T-6009) of 250 μm of i.d.
39 were affixed to each delivery channel reservoir. Fig. 2c
40 illustrates the chip device with the integrated bypass channels. 96

42 B. Experimental set-up

43 Measurement of the HMB resonators was performed by
44 custom designed optical free-space interferometer. S1 from
45 supporting information shows a detailed scheme of our
46 homebuilt experimental setup. A 632.8 nm HeNe laser beam
47 1 mm diameter (JDSU 1101, 1.5mW) passes through an optical
48 isolator (Thorlabs IO-3D-633-VLP) to cancel undesired back
49 reflections and noise fluctuations. A 50:50 beamsplitter
50 (Thorlabs CM1-BS1) splits the laser beam by half to form
51 reference path, which reflects back from a reference mirror onto
52 a high-bandwidth photoreceiver (New Focus 1801). On the
53 active arm of the interferometer, the beam is expanded five times
54 with a beam expander (Thorlabs BE05M-A) for imaging
55 purposes and for reducing the final spot size. The laser beam
56 focused tightly through a 20x microscope objective (Olympus

20x, NA = 0.4) onto the middle point of the structures with the
optical axis parallel to the bending motion of the resonators. The
spot size of 1.3 μm is calculated according to the Rayleigh
criteria: $\omega_0 = 1.64\lambda/2\text{NA}$. Finally, the photoreceiver collects the
interference pattern of the reflected light from a beam and the
light from the reference mirror. All optical components of the
experimental setup are rigidly assembled to an optical table with
active mechanical isolation to compensate for undesired
vibration drifts.

The actuation scheme of the array of resonators was done
through a frequency sweep methodology that allowed a fast and
high-throughput excitation over a range of frequencies around
the central peak response of the resonators [30]. The excitation
was achieved by means of a piezoceramic actuator (PIMICO
PIC181) placed beneath the chip, as Fig. 1 shows. The driven
response of the resonators was acquired by a synchronization
protocol between a function generator (Agilent 33220A) and a
fast acquisition card (National Instruments PXI-5922) that
avoided cross-talk interference between actuated and acquired
signals. The function generator controlled the amplitude (1.5
Vpp) and sweep time (100 ms) of a sine-wave excitation signal
over a 200 KHz bandwidth around the central peak response of
the resonators. Sequential evaluation of every resonator response
was performed by the optical readout by transversally scanning
each of the middle points of the beam resonators under the laser
spot using an automated computer controlled 3-axis stage.
Finally, we computed the Fast Fourier transform (FFT) of the
acquired signals in LabVIEW to obtain the frequency spectrum
response. The FFT was averaged three times to reduce noise
fluctuations and a Lorentzian curve fitting algorithm computed
in real-time the peak frequency response and Q-factor values.

The filling of embedded microchannels was done through an
H-shaped microfluidic configuration as Fig. 2b shows. In each
measurement, 250 μL of sample volume was loaded and
delivered into one of the bypass channels by a low-pressure valve
(Valco Instruments Co. Inc. C22-3186) at a constant flow rate
of 10 $\mu\text{L}/\text{min}$ using a syringe pump (New Era Pump Systems Inc.
NE-300). On the other bypass channel, a constant flow rate of 1
 $\mu\text{L}/\text{min}$ rinsed the output of the microchannels continuously.

The calibration protocol of our sensor consisted of measuring
the response of the resonators filled with air and water before
determining the density and viscosity of samples. The response
of the HMB devices was automatically captured by the optical
readout interferometer while the resonators were actuated
through the sweep frequency methodology. The temperature of
the sensor was stabilized at 23° C by a closed loop temperature
controller (Thorlabs T-Cube TEC Controller) with a resolution
of 0.1° C. Measurements were taken after 5 min of sample
injection to ensure the liquid exchange inside the resonators and
temperature stabilization. Then, the resonance frequency and the
Q-factor were extracted from the resonance peak for each
resonator. A calibration fit was done for the measurements of
water and air. Before measuring a new sample fluid, the HMB
resonators were cleaned with sodium dodecyl sulfate (SDS),
hydrochloric acid 0.1 M (HCl) and rinsed with plenty of water to
reduce systematic instabilities on frequency. All measurements
were done at least in triplicates.

IV. RESULTS AND DISCUSSION

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By considering polysilicon as structural material with density of 2331 kg/m³ and Young Modulus of 160 GPa [31], calculations from (3) showed that the expected fundamental resonant frequency of resonators ($L \approx 275 \mu\text{m}$) was approximately 918 KHz when filled with air. Instead, resonators showed a resonance frequency close to 650 KHz ± 10 Hz operating at 23°C and at atmospheric pressure conditions. This frequency variation was attributed to a modification of the effective length of the structures while releasing them from the substrate with the treatment of HF 49% acid during the fabrication process. By an optical characterization with microscope images, we measured a lateral over-etching distance of approximately 25 μm , which modified the final effective length of each beam. Based on calculations from (3), the resonant frequency of resonators with an over-length of 25 μm at each clamped side was approximately of 651 KHz when filled with air. An additional photolithographic mask to empty and release the resonators in two consecutive steps can further reduce the lateral over-etching effect.

After characterizing the resonators in the unfilled state, we studied the performance of the sensors when filled with water. The array of beams exhibited a frequency response close to 502 KHz ± 11 Hz, which represents a frequency shift of 148 kHz (22.77%) with respect to their response in air. In the Electronic Supplementary Information (ESI) we show a video of the fluid exchange inside the HMB device. The filling capabilities of the embedded microchannels were successfully proven owing to the physical properties of the internal sidewalls, which presented low porosity and a high planarization level. The frequency shift of the resonators was in accordance with the change in density of the fluid contained inside the microfluidic channels. Sensor noise was estimated based on the evaluation of the standard deviation of the frequency response of the resonators when filled with water, specifically when no frequency variation was expected ($\Delta f_R = 0$). Over a period of 45 min, the resonant frequency peak showed a standard deviation of $\sigma_R = \pm 10$ Hz. Although random sources of noise were present in the acquisition system, such as laser amplitude fluctuations and variations of the optical focusing of resonators, other sources of error could potentially be controlled. For example, by increasing the number of acquired samples and by reducing the excitation frequency bandwidth, the systematic errors introduced by the Lorentzian curve fitting of the frequency response were minimized.

Thereafter, to calculate the mass responsivity of the device we used four samples with different and well-known densities (from Sigma Aldrich): diethyl ether (713.4 kg/m³), isopropanol (786 kg/m³), ethanol (789 kg/m³), and acetic acid (1049 kg/m³). The latter one was chosen to measure the linear response of the sensor beyond the density of the reference liquid (water). Fig. 3 shows the frequency response depicted by these sample solutions with respect to their reference density values for a single resonator. The results demonstrated a clear relationship between the measured resonant frequency peak shift and the sample solution density, according to the model proposed by (3). To determine the sensitivity of the sensor, a linear curve fitting of

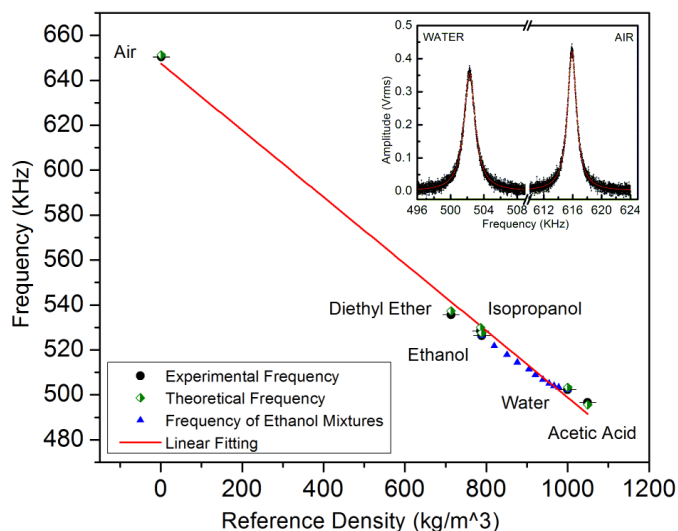


Fig. 3 Resonant frequency response of a resonator when filled with different solutions with well-known densities, including water/ethanol mixtures (blue triangle markers). The inset shows the peak frequency response of the resonator before and after filling it with water indicating a frequency shift of 148 KHz with a 41.1% quality factor decay. Error bars are smaller than black dots and represent a standard deviation of ± 10 Hz.

TABLE I
PERFORMANCES OF THE ARRAY OF HMB RESONATORS

Bridge	Sensor mass (ng)	Resonance frequency f_0 (kHz) in air	Extracted slope (S_A) Hz/kgm ⁻³	Experimental mass responsivity Hz/pg
B1	17.69	650.119	147.77	7.38
B2	17.30	650.422	147.86	7.39
B3	16.90	651.094	148.19	7.40
B4	16.50	651.262	148.75	7.43

data within this range of densities was calculated for each HMB resonator, as Table 1 shows. The mass responsivity was obtained by dividing the sensitivity with respect to the volume of a single embedded microchannel (~20 pL). The results show a better mass responsivity than that achieved by two previous SMR approaches [32],[13]. However, the mass responsivity of HMB resonators is surpassed by that of SMR devices with smaller effective mass [19],[20]. Also, our findings demonstrated that a minimum resolvable density change of 0.068 kg/m³ was achieved for a frequency resolution of ± 10 Hz.

Owing that the mass responsivity of the array of HMB resonators resulted quite similar (~7.4 Hz/pg), the average response of the four sensors was calculated in order to increase the throughput in the analysis of the following samples. To study the sensor performance as a rheology analyzer we prepared binary solutions of ethanol and water. We injected solutions of ethanol with concentrations in volume from 0% to 100%, with increments of 10%. Besides, the density of ethanol/water mixtures was calculated with a commercial pycnometer (10 ml pycnometer, Brand) at a fixed temperature of 23°C to evaluate the frequency response of the sensor as a function of the density of the samples. Fig. 4a shows the average frequency peak measurement over a time period of 10 min for each ethanol concentration. As expected, the interplay between the frequency

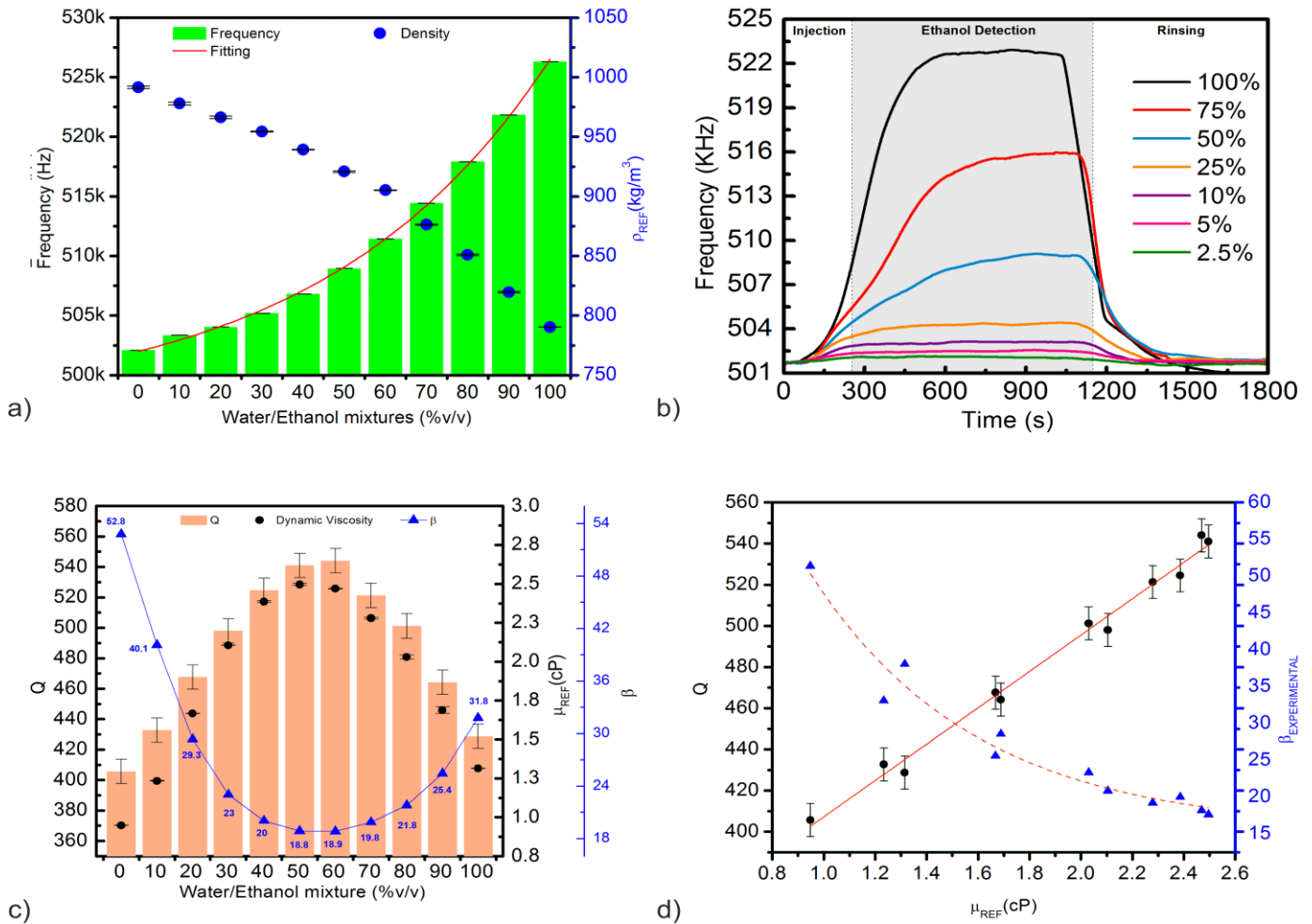


Fig. 4 a) Average resonant frequency response of the HMB resonators with respect to binary mixtures of ethanol and water at different volumetric concentrations ranging from 0% to 100%. Data also shows the inversely proportional interplay of density values measured with a commercial pycnometer. b) Real-time measurements of the shift in the resonant frequency to detect the minimum resolvable %v/v concentration. Dilution effects were noticeable during the first minutes of injection for higher ethanol concentrations. c) A linear tendency is depicted by the Q-factor values as the viscosity of the binary mixtures increases within the viscosity regime from 1 cP to 2.6 cP. d) The experimental values for β demonstrate the interplay between fluid inertia and Q-factor values.

1 of the sensor and the density was inversely proportional with
 2 respect to the content of water/ethanol mixtures. This tendency
 3 was also observed in other calibration protocols [33]. The linear
 4 curve fitting of the frequency with respect to the density within
 5 this range showed a slope of 146 Hz/kgm^{-3} . As Fig. 4b shows, a
 6 complete sample exchange inside of resonators took about 75
 7 minutes for a $10 \mu\text{L}/\text{min}$ flow rate in the inlet bypass channel.
 8 We note that this time could be reduced by controlling the inlet
 9 bypass channel inflow to diminish dilution effects of mixtures
 10 with the initial reference liquid (water) that was contained inside
 11 of resonators. Furthermore, the results demonstrated an
 12 increasing exponential trend in frequency as the ethanol
 13 concentration increased while modifying the sample density
 14 according to the following curve fitting equation:

$$f(\%v/v) = 497178.74 + 4824.114e^{-0.01806(\%v/v)} \quad (5)$$

15
 16
 17 From here, the minimum resolvable ethanol concentration of
 18 0.115% was computed for a frequency resolution of $\sigma_R = \pm 10$

19 Hz. Sensor Signal-to-Noise ratio (SNR) was calculated for the
 20 minimum ethanol concentration as $\Delta f_{2.5\%}/\sigma_R$; where $\Delta f_{2.5\%}=418$
 21 Hz is the frequency shift derived from a 2.5% ethanol
 22 concentration, as shown in Fig. 4b, yielding a SNR of 41.8.

23 As regards the energy dissipation of the HMB device, the
 24 reported Q-factor value of resonators was 692 ± 10 in air.
 25 However, after filling the microfluidic channels with water, the
 26 Q-factor decreased to 405.70 ± 10 for all the resonators, which
 27 represents a decay of 41.1%. This effect is similar to another
 28 approach of SMR devices [24] when the air was replaced with
 29 water showing a 40% decay of the quality factor. To better
 30 understand this behavior, we compared the average response of
 31 the quality factors of the array of resonators when filled with
 32 binary mixtures of water/ethanol, with the corresponding
 33 dynamic viscosities. The viscosities were calculated with a
 34 commercial Ubbelohde viscometer (UBBEL02UKC, Sigma-
 35 Aldrich) at a fixed temperature of 23°C , as Fig. 4c shows.
 36 Interestingly, the experimental results showed a decrease in the
 37 dissipation of energy of the resonators (enhancement of quality
 38

1 factor) as a function of increasing viscosity. This effect is
 2 contrary to the Q-factor response depicted by low-stress silicon
 3 nitride SMR devices [13] in which the quality factor decreased
 4 as a function of increasing viscosity of water/ethanol mixtures.
 5 This can be explained in terms of the dimensionless frequency
 6 number β by considering the height of our microfluidic channels
 7 ($h_{\text{fluid}} = 4 \mu\text{m}$) and taking into account the values of density and
 8 viscosity calculated from the reference sensors. For instance, the
 9 value of β after filling the microchannel with ethanol was
 10 approximately 31.81. Fig. 4c shows that for our device, the
 11 computed values of β for water/ethanol mixtures are within the
 12 low inertia regime ($\beta < 46$). In this regime, since there is little
 13 inertia, the fluid follows the solid displacement of the resonator
 14 resembling a rigid-body oscillation. This explains the
 15 improvement in Q-factor values with increasing viscosity from
 16 1 cP to 2.6 cP. From here, we correlated the Q-factor values of
 17 water/ethanol samples with respect to their reference viscosity
 18 values as shown in Fig. 4d, which resulted in an empirical linear
 19 behavior for the water/ethanol mixtures according to the
 20 following curve fitting approximation:

$$Q(\mu) = 325.78 + 82.5(\mu) \quad (6)$$

21
 22
 23 The non-monotonic interplay between the water/ethanol samples
 24 and the Q-factor, along with the polar nature of the ethanol and
 25 water molecules indicate that (6) is applicable within a short
 26 range of viscosities. From here, the minimum resolvable
 27 viscosity change of 0.15 cP was computed for a Q-factor
 28 resolution of ± 10 . Experimentally, β decreased exponentially as
 29 a function of sample viscosity, which confirms the low-inertia
 30 regime of the sensor, as Fig. 4d shows.

31
 32 Finally, we tested the device performance to measure the density
 33 and viscosity of a variety of alcoholic beverages. We compared
 34 four distilled beverages that contained no added sugar (spirits),
 35 and one distilled beverage with added sugar and flavorings. For
 36 the group of distilled alcoholic beverages, there was a close
 37 correlation as density increased with a percentage error below
 38 0.56%. In particular, beverages such as vodka, whisky and rum,
 39 which contain 40% of Alcohol by Volume (ABV), depicted
 40 close frequency values among them with an average resonant
 41 frequency of $506.6 \text{ KHz} \pm 11 \text{ Hz}$, as Fig. 5a shows. However,
 42 measurements with coffee liquor exhibited a frequency response
 43 ($496.105 \text{ KHz} \pm 11 \text{ Hz}$) below the frequency response of water,
 44 showing that sugar concentration and flavorings influence
 45 density values more than the ethanol content. In comparison with
 46 the time used by U-tube viscometers ($\sim 15 \text{ min}$) and pycnometers
 47 ($\sim 2 \text{ min}$) to determine the viscosity and density of alcoholic
 48 drinks, the HMB device significantly reduced the time for
 49 calculating both parameters. In a single measurement both the
 50 viscosity and density of alcoholic drinks were determined with
 51 our methodology in 7 min requiring sample volumes of only 250
 52 μl . Afterwards, we calculated by (6) the dynamic viscosity of the
 53 alcoholic drinks by tracking the response of the Q-factor of the
 54 sensor (see Fig. 5b). We compared the calculated dynamic
 55 viscosity of each alcoholic drink with their corresponding
 56 dynamic viscosity obtained using the U-tube sensor. The results
 57 showed a percentage error below 2%. Deviations from the

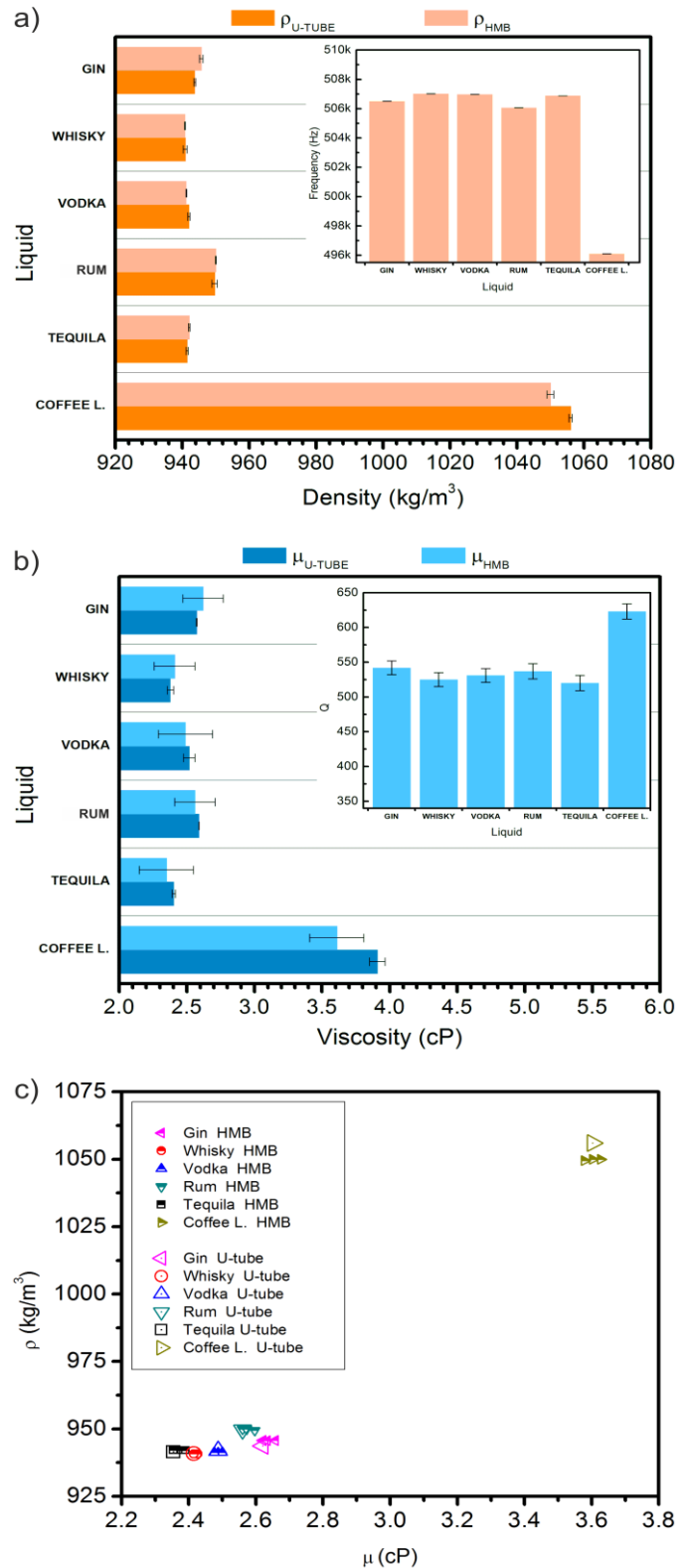


Fig. 5 a) Resonant frequency response of various alcoholic drinks. b) Calculated viscosity of distilled alcoholic drinks (the error increases for viscosities beyond 2.6 cP as is the case for the coffee liquor). c) Comparison of experimental values from the HMB sensor with respect to the values from reference sensors for identification of alcoholic drinks.

1 expected value (7.67%) were observed for the coffee liquo57
 2 beverage given that its dynamic viscosity is out of the linea58
 3 viscosity regime of the HMB sensor. Error bars of the HMB9
 4 viscosity measurements in Fig. 5b make challenging the0
 5 identification of individual alcoholic beverages with close ABV61
 6 percentages. The accuracy of the dynamic viscosity2
 7 measurements can be improved by operating the sensors in a3
 8 moderate vacuum environment. Identification of alcoholic84
 9 drinks with the HMB devices is only feasible as a function of
 10 both the obtained density and dynamic viscosity as Fig. 5c85
 11 shows.

12 Regarding the material properties of the sensor, a structural66
 13 material such as polysilicon can be easily oxidized by differen67
 14 methods such as thermal oxidation or by direct streaming of68
 15 oxidation agents. This advantage is of importance in order to69
 16 modify the hydrophilic properties of the inner microfluidic70
 17 walls to facilitate the filling of resonators with fluids of different
 18 viscosities. Another advantage of polysilicon is the reduced time71
 19 for etching the sacrificial layer in only a few minutes.72
 20

21 V. SUMMARY AND CONCLUSIONS 76

22 We have validated the fabrication and performance of an array78
 23 of polysilicon doubly clamped resonators with embedded79
 24 microfluidic channels to work as a mass density and viscosity80
 25 sensor. The fabrication of the devices was accomplished using81
 26 standard surface micromachining techniques. The double83
 27 clamped configuration of the resonators provided more84
 28 flexibility for fabricating straight microchannels with different85
 29 dimensions to facilitate the input and output of liquid phase86
 30 solutions and also to reduce the clogging of the microfluidic88
 31 channels.89
 32

33 We have achieved a proper on-chip integration of polymer91
 34 based channels using a permanent bonding strategy with PDMS92
 35 This included the sealing with good step coverage of93
 36 microstructures with features that protruded out from the base94
 37 substrate. On the other hand, the implemented optical readout96
 38 granted the efficient acquisition of the nanometric out-of-plane97
 39 displacements of the HMB devices automatically. Enhancing the98
 40 frequency resolution of the detection system by implementing99
 41 parametric feedback oscillator methodologies and operating the101
 42 devices in a moderate vacuum can continuously improve the102
 43 sensitivity and resolution of HMB devices at least one order103
 44 magnitude as it has been demonstrated by a previous approach104
 45 [34].105
 46

47 The experimental results demonstrated that the system could108
 48 be used for high-throughput measurement of liquid phase109
 49 analytes. We have experimentally proven the linearity between110
 50 the resonance frequency shifts of the resonators and the liquid111
 51 density, when testing different samples ranging from solvents113
 52 organic solutions to alcoholic beverages, streamed through the114
 53 embedded microchannels in real-time. Furthermore, we found115
 54 correlation of the viscosity of the samples as a function of116
 55 increasing the Q-factor value of the resonators. Due to the non-118
 56 monotonic energy dissipation of the HMB device, sample119
 57 viscosity measurements were feasible in a short linear regime120
 58 between 1 cP to 2.6 cP, with a resolution of 0.15 cP. Importantly121
 59

the array of HMB resonators could characterize low volumes of
 liquids at atmospheric pressure conditions with a better mass
 responsivity (7.4 Hz/pg) than current SMR devices [13] and with
 a comparable response to another competitive approach [32].
 Thus, this work is a step towards the development of a
 multiplexed platform capable of rapid monitoring of rheological
 properties of distinct fluid samples.

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REFERENCES

- [1] R. Paxman, J. Stinson, A. Dejaridin, R. A. McKendry, and B. W. Hoogenboom, "Using Micromechanical Resonators to Measure Rheological Properties and Alcohol Content of Model Solutions and Commercial Beverages," *Sensors*, vol. 12, no. 5, pp. 6497–6507, 2012.
- [2] J. F. Vélaz-Ruiz and G. V. Barbosa-Cánovas, "Rheological properties of concentrated milk as a function of concentration, temperature and storage time," *J. Food Eng.*, vol. 35, no. 2, pp. 177–190, 1998.
- [3] P. Rust, D. Cereghetti, and J. Dual, "A micro-liter viscosity and density sensor for the rheological characterization of DNA solutions in the kilohertz range," *Lab. Chip*, vol. 13, no. 24, pp. 4794–4799, 2013.
- [4] M. Godin, A. K. Bryan, T. P. Burg, K. Babcock, and S. R. Manalis, "Measuring the mass, density, and size of particles and cells using a suspended microchannel resonator," *Appl. Phys. Lett.*, vol. 91, no. 12, pp. 123121-123121-3, 2007.
- [5] J. Zhang, C. Dai, X. Su, and S. J. O'Shea, "Determination of liquid density with a low frequency mechanical sensor based on quartz tuning fork," *Sens. Actuators B Chem.*, vol. 84, no. 2–3, pp. 123–128, 2002.
- [6] P. Enoksson, G. Stemme, and E. Stemme, "Fluid density sensor based on resonance vibration," *Sens. Actuators Phys.*, vol. 47, no. 1–3, pp. 327–331, 1995.
- [7] I. Etchart et al., "MEMS sensors for density–viscosity sensing in a low-flow microfluidic environment," *Sens. Actuators Phys.*, vol. 141, no. 2, pp. 266–275, 2008.
- [8] B. A. Bircher et al., "Real-Time Viscosity and Mass Density Sensors Requiring Microliter Sample Volume Based on Nanomechanical Resonators," *Anal. Chem.*, vol. 85, no. 18, pp. 8676–8683, 2013.
- [9] L. Müller et al., "Investigation of Prothrombin Time in Human Whole-Blood Samples with a Quartz Crystal Biosensor," *Anal. Chem.*, vol. 82, no. 2, pp. 658–663, 2010.
- [10] O. Cakmak et al., "Microcantilever based disposable viscosity sensor for serum and blood plasma measurements," *Methods*, vol. 63, no. 3, pp. 225–232, 2013.
- [11] M. Yousry, N. Belmiloud, B. Caillard, C. Ayela, C. Pellet, and I. Dufour, "A straightforward determination of fluid viscosity and density using microcantilevers: From experimental data to analytical expressions," *Sens. Actuators Phys.*, vol. 172, no. 1, pp. 40–46, 2011.
- [12] J. Toledo et al., "Comparison of quartz tuning forks and AlN-based extensional microresonators for viscosity measurements in oil/fuel mixtures," in *Proc. SPIE 8763, Smart Sensors, Actuators, and MEMS VI*, 2013, p. 87630L.
- [13] M. F. Khan et al., "Online measurement of mass density and viscosity of pL fluid samples with suspended microchannel resonator," *Sens. Actuators B Chem.*, vol. 185, pp. 456–461, 2013.
- [14] S. Boskovic, J. W. M. Chon, P. Mulvaney, and J. E. Sader, "Rheological measurements using microcantilevers," *J. Rheol.* 1978-Present, vol. 46, no. 4, pp. 891–899, 2002.
- [15] N. McLoughlin, S. L. Lee, and G. Hähner, "Temperature dependence of viscosity and density of viscous liquids determined from thermal noise spectra of uncalibrated atomic force microscope cantilevers," *Lab. Chip*, vol. 7, no. 8, p. 1057, 2007.
- [16] M. Hennemeyer, S. Burghardt, and R. W. Stark, "Cantilever Micro-rheometer for the Characterization of Sugar Solutions," *Sensors*, vol. 8,

1 no. 1, pp. 10–22, 2008. 68
 2 [17] N. McLoughlin, S. L. Lee, and G. Hähner, “Simultaneous determination 69
 3 of density and viscosity of liquids based on resonance curves of 70
 4 uncalibrated microcantilevers,” *Appl. Phys. Lett.*, vol. 89, no. 18, p. 71
 5 184106, 2006.
 6 [18] T. P. Burg et al., “Vacuum-Packaged Suspended Microchannel Resonant 72
 7 Mass Sensor for Biomolecular Detection,” *J. Microelectromechanical 73
 8 Syst.*, vol. 15, no. 6, pp. 1466–1476, 2006.
 9 [19] V. Agache, G. Blanco-Gomez, F. Baleras, and P. Caillat, “An embedded 74
 10 microchannel in a MEMS plate resonator for ultrasensitive mass sensing 75
 11 in liquid,” *Lab. Chip*, vol. 11, no. 15, pp. 2598–2603, 2011. 76
 12 [20] R. A. Barton, B. Ilic, S. S. Verbridge, B. R. Cipriany, J. M. Parpia, and H. 77
 13 G. Craighead, “Fabrication of a Nanomechanical Mass Sensor Containing 78
 14 a Nanofluidic Channel,” *Nano Lett.*, vol. 10, no. 6, pp. 2058–2063, 2010.
 15 [21] J. L. Arlett and M. L. Roukes, “Ultimate and practical limits of fluid-based 79
 16 mass detection with suspended microchannel resonators,” *J. Appl. Phys.* 80
 17 vol. 108, no. 8, pp. 84701–84701–11, 2010.
 18 [22] S. M. Han, H. Benaroya, and T. Wei, “Dynamics of Transversely 81
 19 Vibrating Beams Using Four Engineering Theories,” *J. Sound Vib.*, vol. 225, no. 5, pp. 935–988, 1999.
 20 [23] N. Lobontiu, *Mechanical Design of Microresonators: Modeling and 82
 21 Applications*, 1 edition. New York: McGraw-Hill Professional, 2005.
 22 [24] T. P. Burg, J. E. Sader, and S. R. Manalis, “Nonmonotonic Energy 83
 23 Dissipation in Microfluidic Resonators,” *Phys. Rev. Lett.*, vol. 102, no. 84
 24 22, p. 228103, 2009.
 25 [25] J. E. Sader, T. P. Burg, and S. R. Manalis, “Energy Dissipation in 85
 26 Microfluidic Beam Resonators,” *J. Fluid Mech.*, vol. 650, pp. 215–250, 86
 27 2010.
 28 [26] A. Folch, *Introduction to BioMEMS*. CRC Press, 2012.
 29 [27] S. Marquez, M. Alvarez, D. Farina, C. Dominguez, and L. M. Lechuga, 87
 30 “Towards a biosensing multiple platform based on an array of hollow 88
 31 microbridge resonators,” in *2014 IEEE SENSORS*, 2014, pp. 329–331. 89
 32 [28] D. Qin, Y. Xia, and G. M. Whitesides, “Rapid prototyping of complex 90
 33 structures with feature sizes larger than 20 μm ,” *Adv. Mater.*, vol. 8, no. 91
 34 11, pp. 917–919, 1996.
 35 [29] R. W. R. L. Gajasinghe et al., “Experimental study of PDMS bonding to 92
 36 various substrates for monolithic microfluidic applications,” *J. 93
 37 Micromechanics Microengineering*, vol. 24, no. 7, p. 75010, 2014.
 38 [30] C. I. Enriquez-Flores, J. J. Gervacio-Arciniega, E. Cruz-Valeriano, P. de 94
 39 Urquijo-Ventura, B. J. Gutierrez-Salazar, and F. J. Espinoza-Beltran, 95
 40 “Fast frequency sweeping in resonance-tracking SPM for high-resolution 96
 41 AFAM and PFM imaging,” *Nanotechnology*, vol. 23, no. 49, p. 495705, 97
 42 2012.
 43 [31] S. D. Senturia, *Microsystem Design*. Springer US, 2005.
 44 [32] J. Lee et al., “Suspended microchannel resonators with piezoresistive 98
 45 sensors,” *Lab Chip*, vol. 11, no. 4, pp. 645–651, 2010.
 46 [33] I. S. Khattab, F. Bandarkar, M. A. A. Fakhree, and A. Jouyban, “Density, 99
 47 viscosity, and surface tension of water+ethanol mixtures from 293 to 100
 48 323K,” *Korean J. Chem. Eng.*, vol. 29, no. 6, pp. 812–817, 2012.
 49 [34] S. Olcum et al., “Weighing nanoparticles in solution at the attogram 100
 50 scale,” *Proc. Natl. Acad. Sci.*, p. 201318602, 2014.
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