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#### Experimental evaluation of the extractability of Fe-bound 1 3 organic carbon in sediments as a function of carboxyl content. 4 5 Ben J. Fisher\*, Oliver W. Moore, Johan C. Faust, Caroline L. Peacock, 6 Christian März 7 School of Earth and Environment, University of Leeds, Leeds, LS2 9JT, UK 8 9 \*Corresponding author (Ben.Fisher@ed.ac.uk) 10 11 Keywords: 12 Iron-bound organic carbon; Carboxyl group bonding; Dithionite reduction; 13 Marine sediments; Ferrihydrite 14 15 **Highlights:** 16 The CBD method can underestimate organic carbon bound to reactive -17 iron in sediments 18 Some organic-ferrihydrite complexes are incompletely reduced by Na 19 dithionite at pH 7 Some organic-ferrihydrite complexes are removable by the NaCl wash in 20 21 the CBD method 22 Carboxyl content of OM bound to iron affects its reactivity towards CBD -23 reduction 24 25 Abstract: The majority of organic carbon (OC) burial in marine sediments occurs on 26 27 continental shelves, of which an estimated 10-20% is associated with 28 reactive iron (Fe<sub>R</sub>). The association of OC with Fe<sub>R</sub> (OC-Fe<sub>R</sub>) is thought to 29 facilitate preservation of organic matter (OM) in sediments and therefore 30 represents an important carbon sink. The citrate-bicarbonate-dithionite 31 (CBD) method is used to quantify OC-Fe<sub>R</sub> in marine sediments by 32 reductively dissolving Fe<sub>R</sub>, thereby releasing bound OC. While the CBD 33 method is widely used, it may be less efficient at measuring OC-Fe<sub>R</sub> than currently thought, due to the incomplete reduction of Fe<sub>R</sub>, resulting from the 34 neutral pH conditions required to prevent OM hydrolysis. Additionally, the 35 36 typical range of values reported for OC-Fe<sub>R</sub> in marine sediments is narrow, 37 despite variation in OM and Fe<sub>R</sub> inputs, OM source types and chemical

- compositions. This suggests a limitation exists on the amount of OC that
- 39 can become associated with Fe<sub>R</sub>, and/or that the CBD method is limited in
- 40 the OC-Fe<sub>R</sub> that it is able to quantify. In assessing the efficiency of the CBD

41 method, we aimed to understand whether methodological errors or 42 (mis)interpretation of these extraction results may contribute to the 43 apparent limitation on OC-Fe<sub>R</sub> values. Here, we synthesised OC-Fe<sub>R</sub> 44 composites with a known Fe<sub>R</sub> phase and known OM moieties, varying in 45 carboxyl content, at neutral pH. These were spiked into OC-free marine 46 sediment, and subject to a CBD extraction to investigate i) the efficiency of 47 CBD for OC extraction; ii) the efficiency of CBD for Fe<sub>R</sub> extraction; ii) how the OC moiety affects the physical parameters of associated Fe<sub>R</sub> minerals; 48 49 and iii) the impact of OM moiety on OC and Fe release. We show that the 50 CBD method results in only partial dissolution of the most susceptible Fe<sub>R</sub> 51 phase (ferrihydrite) and therefore incomplete removal of bound OC. While 52 as little as ~20% of Fe is released from OC-free ferrihydrite, structural 53 disorder of the mineral phase increases with the inclusion of more OC, 54 resulting in greater losses of up to 62% Fe for carboxyl rich OC-Fe<sub>R</sub> 55 complexes. In addition, our results show that the NaCl control step 56 performed in the CBD method is capable of removing weakly bound OC from Fe<sub>R</sub>, such that inclusion of this OC in the total OC-Fe<sub>R</sub> fraction may 57 58 increase marine sediments OC-Fe<sub>R</sub> estimates by ~33%. Finally, we 59 suggest that the structure of OC involved in OC-Fe<sub>R</sub> binding can affect quantification of the OC-Fe<sub>R</sub> pool. Our results have important implications 60 61 for assessing the  $Fe_{R}$  bound OC fraction in marine sediments and the fate 62 of this OC in the global carbon cycle.

63

## 64 **1. Introduction**

65 Marine sediments represent the largest organic carbon (OC) sink on Earth 66 (Hedges and Keil, 1995) and therefore play a crucial role in the regulation of 67 atmospheric oxygen (O<sub>2</sub>) and carbon dioxide (CO<sub>2</sub>) over geological time 68 (Berner, 1989). Despite the importance of OC burial however (Burdige, 2007; 69 Canfield, 1994), the factors that control OC burial and preservation in marine 70 sediments are unclear (Arndt et al., 2013). One important mechanism for OM 71 preservation appears to involve the association of OC with the reactive iron 72 fraction in soils and sediments (Fe<sub>R</sub>, operationally defined as the fraction of 73 total iron that is extractable with sodium dithionate), with up to 40% of total 74 soil OC (Wagai and Mayer, 2007) and 21.5±8.6% of total marine sediment OC (Lalonde et al., 2012) bound to Fe<sub>R</sub>. This Fe<sub>R</sub> "rusty sink" (Lalonde et al.,
2012) is assumed to protect OM from microbial degradation (Jones and
Edwards, 1998), and help preserve OM in the Earth system (Lalonde et al.,
2012).

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80 The mechanisms responsible for OC-Fe<sub>R</sub> associations are poorly 81 understood, but might include chemical sorption, physical occlusion or a 82 combination of both (Kaiser and Guggenberger, 2000; Lalonde et al., 2012; 83 Mu et al., 2016; Wagai and Mayer, 2007). For chemical sorption to protect OC from microbial remineralisation over extended timescales, the 84 mechanism of OC sorption with Fe<sub>R</sub> must involve strong chemical bonds and 85 86 a very high sorption coefficient (Henrichs, 1995). Work has demonstrated this using a ramped pyrolysis or oxidation method to observe the 87 88 temperature decay profile of OC as a proxy for bond strength (Hemingway 89 et al., 2017). Strong chemical bonds between OC and Fe<sub>R</sub> are thought to be 90 mediated via OM carboxyl (COOH) and mineral hydroxyl (OH) groups 91 (Karlsson and Persson, 2010; Karlsson and Persson, 2012; Mikutta, 2011). 92 The importance of carboxyl group bonding has been demonstrated by Fourier Transform Infrared Spectroscopy (FTIR) of ferrihydrite precipitated 93 94 with dissolved organic matter (DOM) (Yang et al., 2012). Carboxyl groups 95 are prevalent in marine DOM, as evidenced by carboxyl rich alicyclic 96 molecules (CRAM) comprising a major component of refractory marine DOM 97 (Hertkorn et al., 2006). Carboxyl groups are thought to facilitate binding of OM to Fe<sub>R</sub> mineral surfaces via ligand exchange, whereby OM carboxyl 98 99 groups replace hydroxyl groups on the mineral surface (Chen et al., 2014; 100 Gu et al., 1994; Gu et al., 1995; Henneberry et al., 2012; Wagai and Mayer, 101 2007).

102

103The citrate-dithionite-bicarbonate (CBD) method for evaluating the amount104of OC-FeR in a sediment sample involves reduction of the present FeR105phases, after which the reduced Fe then becomes dissolved in solution106liberating bound OC. Despite widespread utilisation of the CBD method,107previous studies have indicated that its efficiency may be less than originally108described, whereby it was thought to dissolve all solid reactive iron phases.

109 For example, Adhikari and Yang (2015) performed the CBD protocol of 110 Lalonde et al. (2012) on synthetic hematite-humic acid complexes and reported incomplete reduction ( $\leq 50\%$ ) of Fe<sub>R</sub>. Further, the CBD method has 111 112 been criticised in the context of Fe speciation for its inability to fully dissolve 113 crystalline Fe<sub>R</sub> minerals in ancient rocks and potentially modern marine 114 sediments (Raiswell et al., 1994). Thompson et al. (2019) showed that a CBD 115 extraction performed at circumneutral pH fails to extract a large proportion of 116 Fe<sub>R</sub> minerals in modern sediments. While these studies have indicated that 117 CBD treatment may be less efficient than currently understood, it is unknown 118 as to why some OC-Fe<sub>R</sub> complexes appear to be unextractable. By 119 evaluating the efficiency of the CBD method for OC-Fe<sub>R</sub> extraction we aim to 120 understand the true extent to which OC-Fe<sub>R</sub> can be extracted, and whether 121 the structural composition of the associated organic compound influences 122 the ability of CBD to successfully extract a particular OC-Fe<sub>R</sub> complex. This 123 may additionally further our understanding as to why only a minority of the 124 sedimentary OC pool is found to be associated with Fe<sub>R</sub> despite the fact that 125 OC-Fe<sub>R</sub> interactions are found to promote OC persistence. For example, the 126 maximal OC-Fe<sub>R</sub> content obtained by Lalonde et al. (2012) across their 42 127 global sediment samples was 41.69% (Wash. Coast 215), with a mean of 128 21.5±8.6%. Other studies employing the same CBD method also concur with 129 this range (e.g., Zhao et al. (2018) (15.6±6.5%), Salvadó et al. (2015)  $(11 \pm 5.5\%)$ . This poses a question as to why there is limited variability in the 130 131 OC-Fe<sub>R</sub> values obtained for sediments under differing environmental 132 conditions, particularly where OC and Fe fluxes to the seafloor vary. To fully 133 understand the fate of OM in marine sediments it is important to determine 134 whether this apparent limitation in OC-Fe<sub>R</sub> presence is a matter of 135 methodological error/interpretation in the quantification of OC-Fe<sub>R</sub> (by CBD), 136 or an inherent feature of the OC-Fe<sub>R</sub> interaction that requires further 137 investigation.

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To address the uncertainty in CBD extraction efficiency we coprecipitated model carboxylic acid compounds, containing either 1, 2 or 3 carboxyl groups, with ferrihydrite, which is ubiquitous in marine sediments and acts as a precursor for more stable Fe oxide phases (e.g., hematite) (Michel et 143 al., 2007). We then spiked OC-free natural sediment with our carboxylic acidferrihydrite coprecipitates and subject them to a CBD extraction. We 144 investigate the efficiency of the CBD method for OC and Fe<sub>R</sub> extraction and 145 146 discuss the implications for evaluating OC-Fe<sub>R</sub> and OM preservation with 147 Fe<sub>R</sub> in the marine system. We also comment on how the carboxyl character 148 of the OC might have a significant influence on OC preservation in marine 149 sediments.

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#### 2. Materials and Methods 151

# 2.1 Ferrihydrite coprecipitate synthesis

153 Following a modified method of Schwertmann and Cornell (2000), 2-line 154 ferrihydrite was precipitated in the presence of either i) pentatonic acid 155  $(C_5H_{10}O_2)$ ; ii) hexanedioic acid  $(C_6H_{10}O_4)$  or iii) butane-1,2,4-tricarboxylic 156 acid acid  $(C_7H_{10}O_6)$ , which increase in their number of carboxyl groups 157 from 1 to 3, respectively. A pure ferrihydrite sample containing no organic 158 carbon was also precipitated as a control.





159 Figure 1: Chemical structures of the organic acids coprecipitated with ferrihydrite. The chemical structures of each carboxylic acid used 160 in synthesising ferrihydrite coprecipitates are shown above their carboxyl 161 162 content abbreviation (1-COOH, 2-COOH etc.) and IUPAC name. 0 COOH 163 shows no structure as the 0 COOH coprecipitate contains no organic and instead represents a 'pure' form of ferrihydrite. 164

- 165
- 166 Saturation concentrations for the three organic acids were determined by
- 167 increasing organic acid concentration in the mineral coprecipitation stage
- 168 and measuring end state wt %C (see Table 1). Saturation concentration
- 169 refers to the point at which additional increases in the amount of organic

170acid added did not result in a further wt% C increase in the resultant171mineral coprecipitate. Coprecipitation of ferrihydrite with the named172organic acids was performed by dissolving organic acids in deionised173water (DI) with Fe (III) nitrate nonahydrate [Fe(NO<sub>3</sub>)<sub>3</sub>.9H<sub>2</sub>O] (20 g/250 ml).1741 M potassium hydroxide (KOH) was then added by titration to achieve a175pH of 7.0 ± 0.3.

177 Following coprecipitation, all of the organomineral composites were rinsed 178 in 5L of DI water and left to gravitationally settle. This rinse step was 179 repeated 5 times. The pentanoic acid (1-COOH) coprecipitate was rinsed 180 three times due to slow settling of the lighter 1-COOH particles and 181 therefore a lower yield. The pH of the resultant slurry was raised to pH 7 182 through dropwise addition of 0.1 M NaOH, then centrifuged (2750 g, 20 minutes) with the precipitate being retained and the supernatant 183 184 discarded. Following this, the precipitate was frozen and freeze-dried. X-185 ray diffraction (XRD) was used to confirm the Fe mineralogy of the 186 coprecipitate as 2-line ferrihydrite.

187 Table 1: Organic acids used to form ferrihydrite coprecipitates.

188

176

| IUPAC Name                                     | COOH Groups             | Stoichiometry  | Amount added <sup>a</sup> |
|--|-------------------------|--|---------------------------|
| Pentanoic acid                                 | 1                       | $C_5H_{10}O_2$                                       | 5 ml                      |
| Hexanedioic acid                               | 2                       | $C_6H_{10}O_4$                                       | 3 g                       |
| butane-1,2,4-tricarboxylic acid                | 3                       | $C_7H_{10}O_6$                                       | 3 g                       |
| <sup>a</sup> values of organic acid addition a | are relative to 20 g of | Fe(NO <sub>3</sub> ) <sub>3</sub> .9H <sub>2</sub> O |                           |

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2.2 Sediment spiking

192 Ferrihydrite-organic coprecipitates, representing iron bound organic 193 carbon (OC-Fe<sub>R</sub>) were spiked into a OC-free natural sediment sample 194 from the Barents Sea (Water Depth 141 m; sediment core depth, 33.5 cm; 195 station B6, E40; Cruise JR16006) which acted as a 'carrier' for the OC-196 Fe<sub>R</sub> compound and replicated the physical properties of natural fine-197 grained (silty clay) sediments. The sediment sample was freeze-dried and 198 homogenised by grinding using a pestle and mortar, followed by ashing 199 (650°C, 12 hours) to oxidatively remove all OC. While ashing affects 200 various natural sediment properties (e.g. dehydration/aggregation of 201 minerals), it is required as a preparatory step in our experiment to remove 202 all natural OC from the sediment. However, the ashed sediment is used 203 simply as a matrix for our OC-Fe<sub>R</sub> additions, to better mimic the physical 204 properties of a natural sediment than an artificial mixture of various marine 205 sediment minerals. Inorganic carbon was removed by fumigation with HCI 206 vapour, resulting in a sediment sample dominated by siliciclastic minerals. 207 Spiked samples were prepared by mixing carbon free sediment with ferrihydrite coprecipitates at different ratios (see matrix in Table 2) in a 208 209 centrifuge tube and agitating for 10 minutes at room temperature in light 210 conditions.

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212

#### Table 2: Concentration matrix for spiked sediments.

| %OC-Fe: Sedimer | nt O | 10  | 20  | 30  | 40  | 50  |  |
|-----------------|------|-----|-----|-----|-----|-----|--|
| OC-Fe (mg)      | 0    | 25  | 50  | 75  | 100 | 125 |  |
| Sediment (mg)   | 250  | 225 | 200 | 175 | 150 | 125 |  |

213 Values shown are contextualised for a total experimental sample mass of 250 mg (Lalonde et al., 2012).

215

### 216 **2.3 Extraction of iron bound carbon.**

Extraction of dithionite reducible Fe<sub>R</sub> and associated OC was conducted 217 218 according to the citrate-dithionite-bicarbonate (CBD) method (Lalonde et 219 al., 2012; Salvadó et al., 2015). Briefly, 0.25 g of spiked sediment was 220 weighed into centrifuge tubes. Following this, 13 ml of 0.11 M sodium 221 bicarbonate (NaHCO<sub>3</sub>) and 0.27 M trisodium citrate (Na<sub>3</sub>C<sub>6</sub>H<sub>5</sub>O<sub>7</sub>) solution 222 were added and the mixture pre-heated to 80°C in a water bath. Sodium dithionite (0.25 g) was then dissolved in 2 ml of 0.11 M NaHCO<sub>3</sub> and 0.27 223 224 M Na<sub>3</sub>C<sub>6</sub>H<sub>5</sub>O<sub>7</sub> solution, added, and the mixture was agitated and heated 225 again (80°C, 15 minutes). In parallel, a control extraction was conducted 226 replacing sodium dithionite and trisodium citrate with sodium chloride at 227 an equivalent ionic strength; 13 ml of 1.6 M NaCl and 0.11 M NaHCO<sub>3</sub>, 228 followed by 0.22 g NaCl dissolved in 2 ml of the NaCl and NaHCO<sub>3</sub> 229 solution. All samples were centrifuged (10 minutes) and the supernatant 230 was retained. This was followed by three rinses of the precipitate in 231 artificial seawater with 15 ml of each of these three supernatants retained

and combined with one another. All supernatants were acidified to pH <2</li>
with 12 N HCl to prevent Fe precipitation. Precipitates were oven dried at
60°C for 12 hours and decarbonated by acid fumigation to remove
inorganic carbon. Sample mass was recorded prior to elemental analysis
to correct for any mass loss.

# 237 **2.5 Elemental analysis - Iron**

238 Iron concentrations for samples following CBD extraction were 239 determined by atomic absorption spectroscopy (Thermo Fisher iCE3300 AAS). Calibration was performed using matrix matched standards and 240 241 quality control was confirmed following every 10 samples by repeat 242 sampling of calibration standards to check for drift. Total sedimentary Fe 243 content was determined by digesting  $\sim 2$  mg of the spiked sediment mixes 244 in 1 ml 12N HCl at room temperature followed by an initial 10-fold dilution 245 with 1% HCl solution. Subsequent dilutions were made, dependent on Fe 246 content, using MilliQ water to produce a subsample within the detectable 247 window (1-10 ppm Fe). Control supernatants were diluted 20-fold to 248 prevent salt blockages and washed supernatants were undiluted except 249 for where the Fe concentration was >10 ppm, whereby these were diluted 250 10-fold. Extraction of Fe ( $\Delta$ Fe) was calculated as total Fe – CBD 251 extractable Fe, Fe liberated in the NaCl wash stage was subtracted from 252 the CBD extractable total however the NaCl values were negligibly small. 253 Error was determined by propagating the standard error of the mean (SEM) calculated from duplicate results for the total Fe and CBD 254 255 extractable Fe pools.

# 256 **2.6 Elemental analysis – Carbon**

257 Initial carbon content of synthetic samples was measured using a LECO-258 SC144DR C&S analyser. Post extraction carbon was determined by 259 LECO analysis for the majority of sediment samples (OC-Fe 260 concentrations  $\leq$  50%) or by a Vario PYRO cube (Elementar Analysis) 261 where post extraction sample mass was below the limits required for 262 LECO analysis (OC-Fe concentration >50%). Both instruments were corrected for drift by running a known standard (LECO 502-694 and 263 264 Elementar sulfanilamide) following every 10-15 samples throughout the

265analytical run. All carbon samples were analysed in a dried (oven or266freeze-dried) state following the removal of IC as described above.267Carbon loss was calculated according to supplementary equation 1,268adapted from Peter and Sobek (2018) and Salvadó et al. (2015).269Instrument error for C measurement is minimal ( $\leq$  1% RSD) and in all270cases was less than the SEM calculated for independent repeat271measurements.

272 **2.7 X-Ray diffraction analysis** 

Ferrihydrite-organic composites were structurally characterised for crystallinity by powder x-ray diffraction (XRD). The pattern was collected over a 2-90° 2θ range in 0.01969° intervals with a step time of 930 ms using a Bruker D8 diffractometer with Cu-Kα radiation source ( $\lambda \approx 0.154$ nm). Comparative analysis between the different coprecipitates was achieved by normalising for relative intensity.

# 279 **2.8 Particle sizing**

Sizing of particle aggregates within the range of 0.1-10000 µm was conducted via laser diffraction using a Malvern Mastersizer 2000. Wet ferrihydrite coprecipitates were added dropwise to water in the suspension mixer until within the detection limit of the machine. Detection of the particle scattering pattern was conducted in triplicate by the detector array and interpreted using the Malvern Mastersizer 2000 software (v6.01).

286

# **3. Results**

# 288 **3.1 OC content calibration of spiked sediments**

289 Coprecipitation of the three organic acids with ferrihydrite produced 290 coprecipitates with an increasing wt% C content, 1-COOH = 1.15%, 2-291 COOH = 4.20%, 3-COOH = 8.51%. The molar C/Fe ratio similarly 292 increased (1-COOH = 0.2, 2-COOH = 0.5, 3-COOH = 1). These 293 coprecipitates were then spiked into the treated sediment across a 294 concentration gradient (as detailed in Table 2), producing a range of 295 composite samples variable in their C content (Fig. 2). The 15 unique 296 sample compositions represent a range of C concentrations from 0.33% 297 (10%, 1-COOH) to 6.94% (50%, 3-COOH). The C content of these 298 composites varied by both the number of C molecules in the three organic 299 acids and the amount of OC-Fe<sub>R</sub> spiked into the sample relative to 300 sediment (see **Table 2**).



301Figure 2 - Calibration of sediment coprecipitate content and sediment wt% C.302Coprecipitates show a tight linear regression trend across the series with R-squared303values of: 1-COOH = 0.97; 2-COOH = 0.98; 3-COOH = 0.99. 1/2/3 COOH refers to304the number of carboxyl groups per molecule in the three different organic acids305coprecipitated with ferrihydrite. %C refers to the concentration of carbon in the306overall sediment composite by wt%. Instrument error is too small (≤1% RSD) to be307visualised.

308

### 309 **3.2 Extraction of iron bound organic carbon.**

310 The Fe<sub>R</sub> bound OC fraction (%OC-Fe<sub>R</sub>) lost from the composite material during the extraction was determined for each of the carboxyl 311 coprecipitate containing samples at concentrations of 20-50% OC-Fe, 312 since C loss for coprecipitates at 10% OC-Fe was too small to be 313 314 accurately determined (Fig. 3). The wt% C values displayed in black 315 indicate those which have been processed through Supplementary 316 equation 1, a minor adaptation to the common equation used to remove 317 OC that is extractable with NaCl and believed to be unbound to FeR 318 (Barber et al., 2017; Lalonde et al., 2012; Peter and Sobek, 2018; Salvadó 319 et al., 2015). In blue, the maximal 'true' wt% C extracted are shown, these refer to data which has only been transformed for mass loss, i.e. the 320 321 control and reduction extractions were not subtracted from one another. 322 For the 1- and 2-COOH samples the value shown in blue represents OC

323 loss from just the NaCl extraction, as this value was greater than OC loss by dithionite reduction, while for the 3-COOH sample this shows OC loss 324 from reduction as this was higher; all values can be found in 325 326 **Supplementary table 3**. Consideration of why some samples 327 experienced greater OC loss from treatment with NaCl than by dithionite 328 reduction is explored in 4.1. Overall, these two sets of values can be 329 grouped separately; the "true" extraction values are similar in nature (shown in the blue box), while the interpreted data (shown in black) 330 331 become spread after inclusion of both the NaCl control and dithionite reduction values. For the interpreted values ~46% of the total OC-Fe was 332 333 extracted from the 3-COOH coprecipitate, but 'negative' extractions of ~ -334 6% and ~ -90% were achieved for the 1- and 2-COOH coprecipitates, 335 respectively. Negative extraction values can be explained as being an 336 artificial feature of subtracting a greater OC loss in the control experiment 337 from a lesser OC loss in the reduction. The mean difference between all 338 three carboxyl coprecipitates was found to be significantly different for all 339 pairwise combinations via a one-way ANOVA and Tukey's test post hoc 340 (99% significance level) (Supplementary Table S1). Precise values for all extractions including duplicate errors are detailed in **Supplementary** 341 342 Table S3.



343 Figure 3- Dithionite extractable fraction of OC from carboxyl coprecipitate 344 spiked sediments. %OC-Fe extracted from artificial sediments spiked with 1,2 or 3 345 carboxyl organic acids. Error bars show standard error of the mean (SEM) of 346 duplicate values, where error bars are not shown this is because the error is too 347 small to be visualised on this scale. %OC-Fe extracted refers to wt% C removed by 348 reductive dissolution of the reactive iron phase for iron bound carbon. Key 349 descriptors, 1/2/3 COOH, represent the variation in the number of carboxyl groups 350 of the 3 different organic acids coprecipitated with ferrihydrite.

351352

# 3.3 Iron extraction

353 Extractable Fe shows an increasing stepwise trend with the number of 354 carboxyl groups per coprecipitated acid molecule for all concentrations of 355 OC-Fe<sub>R</sub> relative to sediment (Fig. 4). All extractions for easily reducible 356 Fe were incomplete (< 100% of added Fe) with a maximum yield of 61.6% 357 achieved for the 3-COOH coprecipitate at 50% OC-Fe, with a mean range of 50.5-61.6% across all concentrations. In decreasing order, the 2-COOH 358 359 sediment yield was 40.0-45.7% Fe and 1-COOH sediment yield was 17.9-360 29.9%. The OC free (0-COOH) sediment composite achieved the lowest 361 rate of Fe recoverability, between 6 and 14%. Fe removed by the NaCl 362 control treatment was subtracted from the Fe removed by CBD, however 363 the contribution of Fe removed by NaCl was minimal ( $\leq 0.3\%$ ). Fe liberated in wash stages for both CBD and NaCl treatment are included in the 364 365 overall %Fe extracted values. Relative to Fe liberated from the initial chemical treatment, the wash removed a further 6.7-34.0% Fe for CBD 366 367 and 0.4 to 27.8% for NaCl. All sample means were calculated from the average of independent duplicates. A significant difference was found 368 369 between the means of all coprecipitate pairwise combinations (>99% 370 significance) by one-way ANOVA analysis and Tukey's test post hoc. A 371 statistical summary is included in Supplementary Table S2



Figure 4- Dithionite Figure 4- CBD extraction efficiency for reactive Fe in sediment samples differing in carboxyl richness. %Fe extracted from each carboxyl spiked sediment (0-3 COOH) is shown, values are control corrected. Error bars represent propagated error calculated from the duplicate values as detailed in 3.1.

378

# **379 3.4 Coprecipitate grain size**

380 Grain size of ferrihydrite composites was determined by laser diffraction 381 analysis in the 0.01 to 10000 µm range. The 3-COOH coprecipitate 382 showed a slight peak shift from 12.04 to 28.66 µm compared to the 383 carboxyl free, pure ferrihydrite phase (0-COOH) (Fig. 5). One and two 384 COOH coprecipitates showed no determinable difference in grain size 385 distribution and sat between the 0- and 3-COOH sediments. The 0- and 386 3-COOH precipitates also had marginally taller, slimmer distributions 387 indicating a narrower range of grain sizes and a greater proportion of the class existed at the mid-point compared to 1- and 2-COOH. These two 388 389 samples have almost identical normal distribution curves and show no 390 determinable difference in grain size or distribution. Due to the 391 overlapping of all distributions, there is no statistically significant 392 difference in the grain size of coprecipitates.



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Figure 5- Grain size distribution of carboxyl coprecipitates. Grain size distribution is shown as percent by volume in class. Values shown along the solid lines are the mean of triplicate measurements, dashed lines represent the SEM. A small number of data points in the range 0.01-0.1  $\mu$ m and 1000-10000  $\mu$ m are excluded from this graph to improve resolution, all these values were ~0.

398 399

### 400 **3.5 X-ray diffraction analysis (XRD)**

401 All samples show a characteristic two peak signature (Supplementary 402 Fig. S1), indicating successful synthesis, and carboxyl free ferrihydrite 403 resembles a characteristic two-line sample. As the number of carboxyl 404 groups per coprecipitated acid molecule increases, a decrease in relative 405 intensity and softening of the two peaks can be seen.

406

### **4**07 **4. Discussion**

408 Organic carbon bound to Fe<sub>R</sub> represents an important mechanism by 409 which OC is preserved in marine sediments with ~22% of total sediment 410 OC bound to Fe<sub>R</sub> (Lalonde et al., 2012). This "rusty sink" is important for regulating atmospheric O<sub>2</sub> and CO<sub>2</sub> (Torn et al., 1997) and understanding 411 412 the fate of OM is crucial in accurately predicting the future of carbon 413 cycling under global climate change (Adhikari and Yang, 2015). The CBD 414 method, previously used to quantify OC-Fe<sub>R</sub> in terrestrial environments, 415 has been applied to marine sediments providing the first quantification of 416 OC preserved in association with Fe<sub>R</sub> on a global scale (Lalonde et al.,

417 2012). Here we applied the same CBD method to synthetic samples, in an experimental investigation of OC-Fe<sub>R</sub>, to investigate the performance of 418 the CBD method for OC and Fe extractability as a function of the 419 420 molecular characteristics of the associated OM. Previous studies have 421 typically produced organomineral coprecipitates using natural sources of 422 OM, e.g., forest floor extract (Chen et al., 2014; Eusterhues et al., 2014a; 423 Eusterhues et al., 2014b; Eusterhues et al., 2011). Whilst using naturally 424 sourced OM more closely replicates OC-Fe<sub>R</sub> coprecipitates formed in 425 environmental settings, the speciation and structure of natural OM are 426 largely unknown and therefore the effect of OM molecular composition on the efficiency of the CBD method cannot be easily evaluated. By using 427 428 short chain organic molecules, differing only in the number of carboxyl 429 groups per acid molecule, we explore how the CBD method is influenced 430 by the carboxylic character of the OM. We demonstrate important 431 inefficiencies in the CBD method through calibration against synthetic 432 standards which has the potential to transform our interpretation of CBD extractions performed on natural sediments and thus our understanding 433 434 of marine sediment carbon preservation on a global scale (4.1, 4.2). We 435 also comment on how the carboxyl character of the OM might have a 436 significant influence on OM preservation (4.3).

437

# 438 4.1 Current extraction methods may underestimate the extent of OC 439 Fe<sub>R</sub> interactions in soils and sediments

Preservation of OM by Fe<sub>R</sub> is thought to result from protection of OC from 440 microbial degradation, mediated through chemical bonding between OC 441 442 and Fe<sub>R</sub> (Kaiser and Guggenberger, 2000). Protection is believed to 443 require high energy bonding between OC and Fe<sub>R</sub> to maintain a 444 preservative effect (Henrichs, 1995). Weaker interactions (e.g., Van der 445 Waals forces) may exist but are assumed to be of insufficient strength to 446 facilitate OM preservation (Burdige, 2007). Treatment with CBD has been 447 described to dissolve "all solid reactive iron phases and the organic 448 carbon associated with these phases" in natural sediments (Lalonde et 449 al., 2012). Therefore, we would expect full liberation of Fe and C given 450 that the C content of our synthetic sample was equal to the amount of OC

451 associated with ferrihydrite, a solid reactive Fe phase. Across all 452 concentrations and C structures we observe incomplete recovery of Fe<sub>R</sub>, not exceeding 50.51 ± 1.32% of the added Fe content (3-COOH, 40% OC-453 454 Fe<sub>R</sub> addition) (Fig. 2). As the amount of OC extracted is less than total 455 amount of OC added to the sediment samples this indicates either the 456 CBD method or the interpretation of CBD results contain a source of error. 457 However, the OC-Fe<sub>R</sub> values obtained, in particular for the 3-COOH containing sediment, are comparable to observations from many 458 459 environmental samples.

460

461 Where CBD has been applied to marine sediments the %OC-Fe<sub>R</sub> (= 462 fraction of total OC bound to Fe<sub>R</sub>) ranges between 0-42% on continental 463 shelves (e.g. Lalonde et al., 2012; Ma et al., 2018; Salvadó et al., 2015) 464 with typical averages for sediments under oxic bottom waters of 21.7  $\pm$ 465 7.8% (Lalonde et al., 2012). While the calculation for our 3-COOH 466 coprecipitate exceeds these ranges for %OC-Fe<sub>R</sub> in marine sediments, it is agreeable with terrestrial environments. For example, Zhao et al. (2016) 467 468 determined an %OC-Fe<sub>R</sub> pool of 37.8 ± 20% for forest soils. The 469 appearance of a less than complete OC extraction might be explained by 470 the large loss of OC-Fe<sub>R</sub> from NaCl treatment, designed to remove OC 471 not bound to Fe<sub>R</sub>. Lalonde et al. (2012) note 7.2 ± 5.4% of OC is released 472 from treatment of natural sediments with NaCl, while we recorded 43.81 473 ± 3.06% for our 3-COOH coprecipitate. The subtraction of this OC pool, 474 thought to be unbound to Fe<sub>R</sub>, reduces the value for %OC-Fe<sub>R</sub> from 90.00 ± 2.50 % removed by CBD treatment to 46.19 ± 7.57 %. Therefore, we 475 476 suggest that OC removed by CBD gives a more accurate assessment of 477 OC-Fe<sub>R</sub>, and that the subtraction of the NaCl 'control' pool in previous 478 studies may have led to a severe underestimation of the actual size of the 479 %OC-Fe<sub>R</sub> pool.

480

481 The effect of NaCl removal of OC is amplified for the 1- and 2-COOH 482 sediment composites compared to the 3-COOH composite. Here, the 483 substantial extraction of OC by NaCl results in final %OC-Fe<sub>R</sub> calculations 484 of ~-90% and ~-6% respectively (**Fig. 2**), which is physically implausible. 485 This demonstrates that the NaCl control treatment is capable of removing OC from a Fe<sub>R</sub> mineral interface. While the effect of NaCl on the OC-Fe<sub>R</sub> 486 487 association has not been previously investigated and is largely based on 488 assumptions, research in soils has indicated that NaCl has the ability to 489 interfere with  $Fe_R$  minerals, with Pereira et al. (2019) showing that 8-10% 490 of Na<sup>+</sup> and Cl<sup>-</sup> ions in a NaCl solution can adsorb to ferrihydrite. 491 Subsequently the presence of NaCl can affect the adsorption kinetics of 492 organic compounds to Fe<sub>R</sub> with a positive relationship observed between 493 increasing NaCl concentration and adsorption of glyphosate (an organic 494 herbicide) to ferrihydrite (Pereira et al., 2019), while (Orcelli et al., 2018) 495 show an inverse trend for glyphosate adsorption to goethite.

496

497 In our study we are able to show NaCl is capable of removing OC-Fe<sub>R</sub> 498 because by synthesising coprecipitates, we have ensured that all OC in 499 the sample is associated with Fe<sub>R</sub>: i) ferrihydrite is the only reactive 500 interface OC is exposed to; ii) residual OC not associated with ferrihydrite 501 is already removed by multiple rinses following precipitation, at which 502 point C/Fe ratio determination is recommended (Eusterhues et al., 2011; 503 Han et al., 2019). Presumably C/Fe determination would not be 504 recommended following rinses if it was plausible that non-Fe associated OC remained in the coprecipitate, as this would skew these ratios by 505 506 including excess OC.

507

508 In addition to a high OC loss in the control, we may also be observing an 509 artificially low OC loss in the CBD treatment stage. This can explain 510 negative mean values for C% loss of -12.53% and -19.61% following CBD extraction at 20 and 40% (1-COOH), respectively. Under reducing 511 512 conditions, the Fe<sub>R</sub> phase is dissolved by sodium dithionite and the 513 coprecipitate experiences Fe loss, as shown in Fig. 4. However, this Fe 514 loss is incomplete and much of the phase (~77%) remains within the 515 sediment. Liberation of OC from the surface of ferrihydrite during reductive 516 dissolution will increase the number of available surface binding sites and 517 therefore could allow for either re-adsorption of liberated OC or sorption 518 of C containing reagents involved in the reduction stage (sodium 519 bicarbonate, trisodium citrate). While sorption of bicarbonate to Fe 520 (hydr)oxides at neutral pH is a thermodynamically favourable process via monodentate inner-sphere complexation (Acelas et al., 521 2017). 522 bicarbonate contamination is not expected to be significant due to acid 523 fumigation of samples to remove carbonates. Lalonde et al. (2012) also 524 determined residual bicarbonate to be insignificant, contributing  $\leq 0.08\%$ 525 of dry sediment weight. Sorption of OC or citrate onto reduced Fe<sub>R</sub> phases could elevate the sediment carbon concentration determined and 526 527 therefore appear to artificially decrease %C loss in the reduction stage, 528 below what is observed in the non-dissolved control.

529

530 The CBD method assumes that treatment of sediment with NaCl removes 531 OC not associated with Fe<sub>R</sub>. However, our results suggest that this NaCl 532 leached pool of OC is bound to Fe<sub>R</sub> phases but less strongly than the CBD 533 extractable pool, with the susceptibility of an individual OC compound 534 towards removal with NaCl being linked to its carboxyl content. The extent to which comparatively weak OC-Fe<sub>R</sub> associations, removed from Fe<sub>R</sub> by 535 536 NaCl, confer protection from microbial degradation upon OC is unknown, 537 however, a large co-localisation of OC with mineral phases (including Fe) 538 has been observed at depths far beyond quantification of OC-Fe<sub>R</sub> (23.9 metres below seafloor) (Estes et al., 2019). This potentially suggests the 539 540 extent of OC association with Fe<sub>R</sub> is larger than determined by CBD 541 extraction. We suggest that the OC pool extractable by NaCl is of importance when calculating the overall OC-Fe<sub>R</sub> budget. The scale of this 542 543 importance is demonstrated by the additional 7.2 ± 5.4% OC removed 544 with NaCl in treatment of surface marine sediment, previously exposed to 545 seawater (Lalonde et al., 2012). When accounting for the mass of C lost 546 in the extraction (as in this study and in Salvadó et al. (2015)) the removal 547 of OC by NaCl is even greater with 16.4%±7.1% of OC-Fe<sub>R</sub> being 548 discounted from an initial CBD extraction of 27.4%±5.4% (Salvadó et al., 549 2015). Therefore, the subtraction of OC removed by NaCl from the overall 550 OC-Fe pool has a large influence on the resulting value for OC-FeR 551 calculation. It is a question as to why exposure to NaCl should remove 552 such a large fraction of OC from marine sediments which have existed in

553 environments surrounded by seawater for many years. However, the NaCl wash employed in the CBD method is of a much greater molar 554 strength than seawater (1.85 M vs 0.49 M). Our results show that NaCl 555 556 treatment does have the ability to remove Fe<sub>R</sub> bound OC and therefore 557 subtraction of this value from CBD extractions has the potential to 558 underestimate the global OC-Fe<sub>R</sub> budget. Given the ability of NaCl to 559 remove loosely bound OC, caution should be taken in interpreting OC 560 extracted in this way before removal from the dithionite extractable pool.

561

562 A recalculation on the presumption that all NaCl removable OC extracted 563 by Lalonde et al. (2012) represents weakly bound OC-Fe<sub>R</sub> suggests that 564 the role of Fe<sub>R</sub> in preserving OC could be underestimated by up to  $\sim$ 33%. 565 However, while in a synthetic experiment it appears that NaCl removable 566 OC is bound to Fe<sub>R</sub> phases, this may not be fully translated to natural 567 sediments due to the loss of OC from other sources, such as those bound 568 to manganese oxides and other mineral surfaces (Allard et al., 2017; 569 Bernard, 1997; Tipping and Heaton, 1983) or DOC dissolved in the pore 570 waters (Fox et al., 2018; Rossel et al., 2016). A change in the extent to 571 which OC-Fe<sub>R</sub> contributes to OM preservation has ramifications for both 572 the global OC-Fe<sub>R</sub> budget and for biogeochemical models aiming to 573 include OM preservation mechanisms to reduce uncertainties related to 574 the fate of OM (Schmidt et al., 2011; Zhao et al., 2016). The bias towards 575 removal of organic compounds with a lower number of carboxyl groups by 576 NaCl suggests that these moieties are the most likely organic compounds to be excluded from OC-Fe<sub>R</sub> quantifications by the CBD method. 577 578 However, the underlying mechanisms controlling OC-Fe<sub>R</sub> bonding and the 579 strength of these OC-Fe<sub>R</sub> bonds requires further investigation.

580

# 581**4.2 Current extraction methods may incompletely extract reactive**582iron phases in sediments.

583 Quantifying the efficiency of Fe<sub>R</sub> extraction techniques has largely been 584 neglected within the literature due to the nature of chemical methods 585 being operationally defined, making it difficult to calibrate these against 586 alternative methods of measuring Fe<sub>R</sub> phases such as Mössbauer

587 spectroscopy (Hepburn et al., 2020). In marine sediments, total Fe is 588 comprised of phases of differing stability, requiring extraction techniques 589 of increasing strength to remove them (Poulton and Canfield, 2005). Here 590 we isolated ferrihydrite, which is considered easily reducible by sodium 591 dithionite as part of the Fe<sub>R</sub> pool (Lalonde et al., 2012). However, our 592 measure of Fe extractability shows a maximum extraction of only ~60 wt% 593 Fe when coprecipitated with a 3-COOH acid; comparable with the 594 incomplete (~31-50%) sodium dithionite reduction of hematite-humic acid 595 complexes observed by Adhikari and Yang (2015). We find that Fe 596 extractability appears to be even less efficient, for 1- and 2-COOH 597 coprecipitates (Fig. 4).

598

599 While reduction by Na dithionite is a well-established method, the 600 adoption of CBD treatment at circumneutral pH, to avoid hydrolysis of OC, 601 remains unique to the OC extraction application. Thompson et al. (2019) 602 found that a CBD extraction at pH 7.6 fails to extract a large proportion of 603 crystalline Fe oxide minerals in both modern and ancient sedimentary 604 rocks. We make a similar observation for OC-free ferrihydrite phases, 605 where only  $9.91 \pm 0.95\%$  Fe is recovered post extraction. While it is 606 apparent that CBD treatment cannot fully extract easily reducible Fe 607 (hydr)oxide phases, and therefore the entire associated OC pool, there is no obvious improvement that can be made to the current protocol due to 608 609 the constraints of maintaining a neutral pH.

610

611 The apparent trend between the number of carboxyl groups per 612 coprecipitated acid molecule and Fe extracted (Fig. 2) may be explained 613 by considering how the association of OC with iron (hydr)oxides effects 614 the structural order of the resulting mineral phases. Previous work shows 615 that with increasing OC content, the mineral is more poorly ordered (Cismasu et al., 2011; Mikutta, 2011; Mikutta et al., 2008). Thus, because 616 617 we see increased OC uptake with the number of carboxyl groups per acid 618 molecule, it is expected that our ferrihydrite becomes more poorly ordered 619 and thus less chemically stable compared to OC-free ferrihydrite. Our 620 XRD analysis supports this theory as a weakening of peaks identifying 2-

621 line ferrihydrite is observed when OC content in the coprecipitates is increased (Supplementary Fig. S1). Decreasing crystallinity was 622 similarly observed for samples where total OC (polygalacturonic acid) 623 624 content was increased (C/Fe 0-2.5) which inhibits transformation of 625 ferrihydrite to more stable phases (ThomasArrigo et al., 2018). In addition, 626 upon extraction the presence of organic matter in OM-Fe<sub>R</sub> complexes has 627 previously been attributed to the reductive release of Fe through electron 628 shuttling effects, due to the ability of organic compounds to act as electron 629 carriers between redox reactions (Adhikari and Yang, 2015).

630

# 631 **4.3 Carboxyl content of Fe associated OC effects OC extraction**

632 As the carboxyl content of our coprecipitates increases, the ability of dithionite to remove OC increases, while the ability of NaCl to remove OC 633 634 diminishes. For the sample spiked with 50% OC-Fe, the 1-COOH sample was extractable for  $92.74 \pm 4.39\%$  of its OC-Fe content by treatment with 635 636 NaCl while dithionite removed only 10.85 ± 9.32% (Supplementary Table S3.) The balance between OC liberated by dithionite vs NaCl 637 638 subsequently shifts in favour of dithionite with increasing carboxyl content, 639 for the 2-COOH sample the values are roughly equal (dithionite =  $70.42 \pm$ 640 1.23%, NaCl =  $77.89 \pm 2.12\%$ ). Finally, for the 3-COOH sample dithionite 641 is twice as effective than NaCl, removing  $89.60 \pm 0.49\%$  of OC compared 642 to  $43.04 \pm 0.49\%$ , the trend described for the 50% OC-Fe sample was 643 similarly observed across all coprecipitate concentrations (Fig. 3).

644

It is assumed that NaCl is only likely to remove weakly bound OC, since 645 646 OC-Fe bonds are known to be very strong (Barber et al., 2017; Henrichs, 647 1995) and NaCl was previously attributed to only removing OC not 648 associated with Fe<sub>R</sub> (Lalonde et al., 2012). Further, the extraction of less 649 OC from sediments upon treatment with NaCl compared to dithionite (e.g. 650 Lalonde et al., 2012; Salvadó et al., 2015) demonstrates that the NaCl 651 treatment must be less aggressive than dithionite, which is to be expected 652 since NaCI has no reducing power. Therefore, the greater loss of OC by 653 NaCl treatment compared to dithionite for the 1- and 2-COOH containing samples suggests a greater proportion of this OC is weakly bound 654

655compared to the 3-COOH sample. In turn this suggests that polycarboxylic656acids and more carboxyl rich compounds are more strongly bound to FeR,657possibly via the formation of multiple strong carboxyl ligand exchange658bonds, and hence that carboxyl rich OM might be preferentially preserved659in soils and sediments (Kaiser and Guggenberger, 2007).

660

661 For our coprecipitation experiments, the OC source was isolated to 662 carboxyl groups as far as possible, with unavoidable inclusion of one 663 additional short chain C molecule in the 1- and 2-COOH acids (4 non-COOH C's) compared to the 3-COOH acid (3 non-COOH C's). As COOH 664 and OH groups are thought to be important for OM-Fe binding (Karlsson 665 666 and Persson, 2010; Karlsson and Persson, 2012; Mikutta, 2011; Yang et al., 2012) individual moieties of carboxylic acid were used to reproduce 667 668 natural OM binding to Fe<sub>R</sub>, but in a controlled system where the potential 669 for different sorption mechanisms due to different OM functional groups 670 was reduced. If polycarboxylic acids and more carboxyl rich OM compounds are more strongly bound to Fe<sub>R</sub> then environmental conditions 671 672 favouring high concentrations of Fe, such as continental margins and hydrothermal vents (Tagliabue et al., 2017), as well as carboxyl rich OM 673 674 inputs (e.g., regions of high primary productivity and shallow waters) are 675 likely to be more efficient at preserving OC.

676

677 The translation of this implication into an environmental context requires an understanding of the moieties present in marine DOM. Much of the 678 679 work aimed at probing the OC-Fe<sub>R</sub> interaction has involved 680 coprecipitations of Fh with terrestrial DOM (e.g., Eusterhues et al., 2014a; Eusterhues et al., 2014b; Eusterhues et al., 2011). Typical coprecipitation 681 682 experiments for Fe<sub>R</sub> bound OC use DOM from forest floor extracts or 683 standards (e.g., Suwannee River). Molecular level work using scanning 684 transmission X-ray microscopy coupled with near-edge X-ray absorption 685 fine structure (STXM NEXAFS) spectroscopy suggests the importance of 686 OM carboxyl content for OC binding to Fe<sub>R</sub> by using forest floor extract 687 enriched for carboxyl content (Chen et al. (2014). Coprecipitation with 688 terrestrial organic matter may not be representative of marine DOM

689 however, as marine DOM might be enriched in carboxyl content, for 690 example, a significant fraction of refractory marine DOM is composed of 691 carboxyl-rich alicyclic molecules 'CRAM' (Hertkorn et al., 2006). 692 Additionally, marine (algae) OM further differs from terrestrial sources by 693 exhibiting a lower C/N ratio (4-10) than terrestrial OM (>20) (Meyers, 694 1994). Higher plants, the largest contributor to terrestrial OM, contain less 695 than 20% protein, hence have lower nitrogen contents and higher C/N 696 ratios than protein rich (~80%) marine bacteria and benthic animals 697 (Müller, 1977). OM preserved in ancient sediments (since 11.6 Ma) shows 698 increased C/N ratios (>10) relative to marine surface (algal) OM (Twichell 699 et al., 2002). This change is attributed to partial OM degradation during 700 sinking and potentially post deposition. Verardo and McIntyre (1994), 701 however, suggest that alteration of C/N ratios from shallow to deep burial 702 in sediments occurs due to favourable decomposition of N-rich (C/N low) 703 OM by microbial communities. This would imply that N-rich (C/N low) 704 marine OM undergoes more extensive structural alteration during 705 decomposition than C/N high terrestrial OM. Future work should focus on 706 coprecipitation of carboxyl rich OM and marine OM with Fe<sub>R</sub> to further 707 elucidate the controls on OC-Fe<sub>R</sub> associations.

708 709

# 5.0 Conclusion

710 Determining of the amount of OM associated with Fe<sub>R</sub> in sediments is an 711 important parameter in understanding the fate of OM in the marine 712 environment and the biogeochemical controls on the global C cycle more 713 widely. Currently, only the CBD method exists for quantifying the extent of 714 this interaction, however, by estimating OC-Fe<sub>R</sub> using a chemical 715 extraction, this means any resulting value is operationally defined by its 716 susceptibility to chemical treatment and not by its persistence or 717 association with Fe<sub>R</sub>. Despite this uncertainty, little work had been 718 performed to understand the limitations of the CBD method and therefore 719 results are currently considered to be representing all of the Fe<sub>R</sub> and 720 associated OC in a sediment sample, with some studies using this value 721 to estimate regional or global OC-Fe<sub>R</sub> budgets. Here, we performed CBD 722 extractions on synthetic ferrihydrite-organic acid coprecipitates varying in

723 their carboxyl context to understand how samples which should be recoverable for their total Fe and C contents performed under the CBD 724 method. In contrast with assertions that CBD treatment was able to fully 725 726 reduce all solid Fe<sub>R</sub> phases and associated OC, we found that CBD is 727 unable to fully reduce ferrihydrite. Further, we show Fe reduction to be 728 variable based on the chemical composition of OM, specifically carboxyl 729 group content. As an increasing number of carboxyl groups become 730 associated with a ferrihydrite-organic coprecipitate this exerts a strong 731 control on the crystallinity and therefore reactivity of Fe, with a significant 732 increase in the amount of Fe reduced for carboxyl rich coprecipitates. 733 Additionally, we found that the NaCl wash stage has the ability to remove 734 a large amount of Fe bound OM and that further consideration should be 735 given in interpreting OC loss from the NaCl stage before subtracting this, 736 often large value, from the dithionite extractable pool. These conclusions 737 have significant implications for the quantification of OC-Fe<sub>R</sub> as NaCl 738 extractable OC was originally thought to only extract non-Fe bound OC 739 and therefore this sum is subtracted from the dithionite extractable pool of 740 OC-Fe<sub>R.</sub> Our results therefore indicate that a combination of incomplete Fe<sub>R</sub> reduction by CBD and removal of Fe<sub>R</sub> bound OC by NaCl have likely 741 742 contributed towards an underestimation of the extent to which OC-FeR 743 interactions persist in marine sediments.

744

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942

| 943 | Supplementary Information  |
|-----|--|
| 944 |  |
| 945 | Mass balance calculation   |
| 946 | To account for the mass loss during the extraction experiment we   |
| 947 | applied the mass balance calculation of Salvadó et al. (2015) combined   |
| 948 | with Peter and Sobek (2018). The %OC loss is calculated by applying  |
| 949 | bulk %C to the pre extraction mass and post extraction %C to the final   |
| 950 | mass. Comparison by change in raw %C is likely to overestimate the   |
| 951 | final %C if start and end carbon concentrations are calibrated to the  |
| 952 | initial mass, as when %C reduces so does the sample mass.  |
| 953 | Supplementary Equation 1 was devised to determine %OC-Fe loss.   |
| 954 |  |
| 955 | Supplementary Equation 1   |
| 956 | $\Delta \mathbf{C(\%)} = \frac{\left(M^{Pre(R)} x \ \% C^{Bulk}\right) - \left(M^{Post(R)} x \ \% C^{Post(R)}\right)}{M^{Pre(R)} x \ \% C^{Bulk}} - $                      |
| 957 | $\frac{\left(M^{Pre(C)}x\ {}^{\!\!\!\!%}C^{Bulk}\right) - \left(M^{Post(C)}x\ {}^{\!\!\!\%}C^{Post(C)}\right)}{M^{Pre\ (C)}x\ {}^{\!\!\!\%}C^{Bulk}} \ \textbf{\times100}$ |
| 958 | M <sup>Pre/Post(R/C)</sup> = Mass pre/post (R) reduction / (C) control.  |
| 959 | %C <sup>Post(R/C)</sup> = %C Post reduction/control experiment.  |
| 960 | %C <sup>Bulk</sup> = %C in the sediment pre extraction (same for control and   |
| 961 | reduction).  |
| 962 |  |

X-Ray Diffraction Analysis



964

965 966

967

Figure S.1- Stacked XRD of coprecipitates with increasing carboxyl rich organic content.

968 Statistical Analysis

# 969 Ordinary one-way ANOVA

970 A one-way ANOVA test was conducted for %OC-Fe extraction values 971 (Figure 2) and %Fe extraction values (shown in Figure 4) across the 972 carboxyl range. The one-way ANOVA was applied to these two data sets 973 as a comparative measure of sample means across the groups. The 974 one-way ANOVA is limited by only having the ability to report whether there is a difference in means between two groups. Hence, a Tukey 975 976 honest significant difference (HSD) test was applied post-hoc to 977 determine the exact groups for which a statistically significant difference 978 in means occurred for at the 99% significance level (Table S1 and S2). 979 All analyses were conducted in GraphPad Prism version 8.30, GraphPad 980 Software, La Jolla California USA, www.graphpad.com.

981

963

| 982        | Table S1: Tukey's multiple comparisons test of extracted %OC-Fe for carboxyl |                              |               |  |  |  |
|------------|--|------------------------------|---------------|--|--|--|
| 983        | coprecipitates.  | coprecipitates.              |               |  |  |  |
| 985        | Comparison   | Adjusted P Value             | Significant?* |  |  |  |
| 986        | 1 COOH vs 2 COOH   | <0.0001                      | Yes           |  |  |  |
| 987<br>988 | 1 COOH vs 3 COOH   | <0.0001                      | Yes           |  |  |  |
| 989        | 2 COOH vs 3 COOH   | 0.0001                       | Yes           |  |  |  |
| 990        | *Significance was determined   | d at the 99% significance le | vel.          |  |  |  |
| 991        |  |                              |               |  |  |  |
| 992        |  |                              |               |  |  |  |
| 993        | Table S2- Tukey's multiple comparisons test of extracted %Fe for carboxyl    |                              |               |  |  |  |
| 994        | coprecipitates.  |                              |               |  |  |  |
| 996        |  |                              |               |  |  |  |
|            | Comparison   | Adjusted P Value             | Significant?* |  |  |  |

| Companson        | Aujusteu i Value | Significan |   |
|------------------|------------------|------------|---|
| 0 COOH vs 1 COOH | 0.0050           | Yes        | 999   |
| 0 COOH vs 2 COOH | <0.0001          | Yes        | $\begin{array}{c} 1000 \\ 1001 \end{array}$ |
| 0 COOH vs 3 COOH | <0.0001          | Yes        | 1002  |
| 1 COOH vs 2 COOH | <0.0001          | Yes        | 1005  |
| 1 COOH vs 3 COOH | <0.0001          | Yes        | 1005<br>1006                                |
| 2 COOH vs 3 COOH | 0.0019           | Yes        | 1007<br>1008                                |
|                  |                  |            |   |

1009

\*Significance was determined at the 99% significance level.

# 1010 Determination of %OC-Fe

1011Table S3 shows the raw data for the reduction and control stages of the1012CBD treatment which were used as inputs to Equation 1 in order to1013calculate %OC-Fe as shown in Figures 2 and 3

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- 1015
- 1016

Table S3- Raw carbon data for determining %OC-Fe.

| wt %OC-Fe<br>in sample | Bulk %OC* | Reduction (Δ %OC)  | Control (∆<br>%OC)   | %OC-Fe extracted    |
|------------------------|-----------|--------------------|----------------------|---------------------|
|                        |           | 1 COC              | ЭН                   |                     |
| 20                     | 0.636     | -12.527 (± 21.056) | 78.606 (±<br>11.716) | -91.132 (± 9.340)   |
| 30                     | 0.968     | 17.398 (± 7.741)   | 95.968 (± 1.162)     | -65.189 (± 9.346)   |
| 40                     | 0.978     | -19.607 (± 2.243)  | 87.622 (± 9.317)     | -107.229 (± 11.560) |
| 50                     | 1.357     | 10.854 (± 9.315)   | 92.742 (± 4.386)     | -81.888 (± 13.702)  |
|                        |           | 2 000              | ער                   |                     |
| 20                     | 1.681     | 74.066 (± 5.818)   | 75.944 (± 1.457)     | -1.879 (± 7.275)    |
| 30                     | 2.192     | 71.080 (± 2.150)   | 70.613 (± 9.976)     | 0.467 (± 12.126)    |
| 40                     | 3.237     | 63.203 (± 8.030)   | 78.079 (± 2.245)     | -14.876 (± 10.275)  |
| 50                     | 3.639     | 70.423 (± 1.234)   | 77.892 (± 2.123)     | -7.468 (± 3.357)    |
|                        |           | 2 000              | NU                   |                     |
| 20                     | 2.761     | 87.043 (± 3.224)   | 46.620 (± 4.390)     | 40.423 (± 7.615)    |
| 30                     | 4.339     | 91.764 (± 1.881)   | 44.503 (± 3.625)     | 47.260 (± 5.506)    |
| 40                     | 5.420     | 91.628 (± 0.686)   | 41.114 (± 0.629)     | 50.514 (± 1.315)    |
| 50                     | 6.935     | 89.602 (± 0.487)   | 43.036 (± 0.485)     | 46.566 (± 0.973)    |
|                        |           |                    |                      |                     |

1017

1018\*Bulk %OC refers to the OC content prior to any treatment. Values shown are a mean<br/>of duplicates, ± indicates the standard error of the mean (SEM).

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