# Accepted Manuscript

Using polysaccharides for the enhancement of functionality of foods: a review

Xu Lu, Jinghao Chen, Zebin Guo, Yafeng Zheng, Mary C. Rea, Han Su, Xiuhua Zheng, Baodong Zheng, Song Miao

PII: S0924-2244(18)30262-0

DOI: https://doi.org/10.1016/j.tifs.2019.02.024

Reference: TIFS 2430

To appear in: Trends in Food Science & Technology

Received Date: 18 April 2018

Revised Date: 30 October 2018

Accepted Date: 6 February 2019

Please cite this article as: Lu, X., Chen, J., Guo, Z., Zheng, Y., Rea, M.C, Su, H., Zheng, X., Zheng, B., Miao, S., Using polysaccharides for the enhancement of functionality of foods: a review, *Trends in Food Science & Technology*, https://doi.org/10.1016/j.tifs.2019.02.024.

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.



1	Using polysaccharides for the enhancement of functionality of foods: a
2	review
3	Xu Lu <sup>a, b, c, d, e</sup> , Jinghao Chen <sup>a, d</sup> , Zebin Guo <sup>a, c, e</sup> , Yafeng Zheng <sup>a, c, e</sup> , Mary C Rea <sup>b</sup> , Han Su <sup>a</sup> ,
4	<sup>d</sup> , Xiuhua Zheng <sup>a</sup> , Baodong Zheng <sup>a, c, d, e,*</sup> , Song Miao <sup>b, a, d, *</sup>
5	<sup>a</sup> College of Food Science, Fujian Agriculture and Forestry University, Fuzhou 350002, People's
6	Republic of China
7	<sup>b</sup> Teagasc Food Research Centre, Moorepark, Fermoy, Co.Cork, Ireland
8	<sup>c</sup> Institute of Food Science and Technology, Fujian Agriculture and Forestry University, Fuzhou
9	350002, People's Republic of China
0	<sup>d</sup> China-Ireland International Cooperation Centre for Foods Material Science and Structure Design,
1	15 Upper and Lower Store Road, 350002, People's Republic of China
2	<sup>e</sup> Fujian Provincial Key Laboratory of Quality Science and Processing Technology in Special
3	Starch, Fujian Agriculture and Forestry University, Fuzhou 350002, China
4	
5	
6	
7	
8	
9	
21	
2	
3	
4	
5	*Corresponding author.
6	E-mail address: zbdfst@163.com (B. D. Zheng) and song.miao@teagasc.ie (S. Miao).

#### 27 Abstract

Background: Flavor, taste and functional ingredients are important ingredients of food, 28 29 but they are easily lost or react during heating and are stable. not Carbohydrate-carbohydrate interactions (CCIs) and carbohydrate-protein interactions 30 (CPIs) are involved in a variety of regulatory biological processes in nature, including cell 31 32 differentiation, proliferation, adhesion, inflammation and immune responses. 33 Polysaccharides have high molecular weights and many intramolecular hydrogen bonds, can be easily modified chemically and biochemically to enhance bioadhesive and 34 biostability of tissues. Therefore, polysaccharides are the foundation for building complex 35 and stable biosystems that are non-toxic with highydrophilicity and easily biodegradable. 36 Scope and approach: In this review, we summarize the principles and applications of 37 polysaccharide delivery systems in a variety of foods. 38

Key findings and conclusions: This review focuses on the self-assembly of carbohydrates 39 with complex structures and discusses the latest advances in self-assembly systems. The 40 host-guest complexes formed by polyvalent sugar conjugates have the potential to provide, 41 42 control or target delivery or release systems. They can also extend the shelf life of food and prevent oxidation and isomerization during food storage. Moreover, very few studies 43 have outlined a comprehensive overview of the use of various types of food 44 polysaccharide matrixes for the assembly and protection of food ingredients, which is a 45 very important area for further study. 46

47 Keywords: Polysaccharide, self-assembled, food functional ingredients, nanoparticles,
48 directional delivery, bioavailability

1

#### 49 1. Introduction

Nanostructured carbohydrates with self-assembly capabilities are used in both drug 50 51 delivery and functional foods due to their good biocompatibility (Santiago & Castro, 2016). Carbohydrates with different shapes, sizes and structures can achieve different 52 53 self-assembly strategies, resulting in food with abroad range of applications and functions. 54 One of the methods utilizing interactions between bioactive molecules and carbohydrates is the creation of a larger functional unit by multiplexing multiple components under a 55 variety of weaker (non-covalent) interactions. However, the weaker chemical bonds may 56 57 also be destroyed under thermal or mechanical stress, so the chemical bonds are sometimes reinforced by covalent bonding to enhance the nanostructures. Most bioactive 58 molecules in food are often water-soluble with poor bioavailability, reactivity, and 59 60 stability after processing, storage and digestion through the human gastrointestinal tract. It is therefore important to provide an appropriate protective carrier and targeted transport 61 material for the food active ingredient. The carrier matrix should first be food grade, 62 biodegradable and have the ability to withstand harsh processing, storage and directional 63 distribution. Carbohydrate is an important component of food, providing energy and 64 remaining the function integrality of organs. Most carbohydrates are biocompatible within 65 the human body and have good biodegradability, and they can meet the required carrier 66 functional properties using only specific treatments (Fathi, Martin, & McClements, 2014). 67 Polysaccharides usually alter the food matrix to cause changes in rheological 68 properties, which increases water retention and gel formation, leading to thickening of 69 70 the food matrix. Other applications include the stabilization of foam, emulsion and

suspended particulate materials, improving palatability, preventing or reducing the 71 72 formation of ice crystals in frozen foods and interacting with other biomolecules. The viscosity of food has a strong correlation with flavor perception. Highly sticky foods 73 delay the release of food flavor and taste compounds. Thus, the appropriate 74 75 polysaccharide-binding emulsifier formulation can effectively solve the problem of flavor 76 loss. Currently, many food flavor manufacturers are also seeking polysaccharide-based formulations to improve the product's flavor quality and sensory experience. 77 Biocompatibility and biodegradability are key factors for the polymer in the product 78 design when used as a bioactive compound carrier. Natural polysaccharides, such as 79 80 non-toxic and biodegradable polymer materials, have been widely used in the preparation of biological nano-carriers. The wide range of sources, good biocompatibility, 81 82 non-immunogenicity, and the large number of modifiable functional groups render natural polysaccharides a good choice as bioactive nano-carriers. Moreover, natural 83 polysaccharides degrade into oligosaccharides and can easily be absorbed without 84 inducing inflammation. Because the structures of polysaccharides in whole grains, 85 whole-wheat bread/pasta, brown rice, fruits and vegetables are complex and resistant to 86 digestive enzymes, they require long digestion and absorption times. Therefore, it is 87 necessary to alter the carbohydrate carrier's structure to control its solubility, digestion, 88 absorption, specific release, and even targeted delivery to protect nutrients and bioactive 89 compounds (Shibakami, Tsubouchi, Sohma, & Hayashi, 2015). Self-assembly is not a 90 new concept in food sciences; thickened polysaccharides or gelation in sauces and 91 92 puddings are good examples. The assembly can be performed by adjusting the

93 environmental conditions (i.e., temperature, pH, ionic strength, specific substances or 94 ions), processing conditions (i.e., different external forces), or the concentration of molecules/particles in the system. The assembled structure depends on the physical and 95 chemical properties as well as the nutrients and quality of the final products. A 96 high-molecular-weight carbohydrate complex has a chain structure, and its diameter is at 97 98 the nano-level. Such a complex can interact with other food compounds to initiate 99 self-assembly in the presence or absence of external forces to minimize the Gibbs free energy of the mixture. This process requires a balance between repulsive and attractive 100 forces to achieve thermodynamic equilibrium. The uniqueness of carbohydrates, due to 101 102 their various polarities and charged groups, can therefore be used as carriers to bind or capture a large number of bioactive molecules with targeted delivery properties. 103 104 Most bioactive compounds are limited by poor solubility (e.g., curcumin extracted

from turmeric), but some complex polysaccharides can be soluble in water. 105 Polysaccharide carbohydrates can interact with most bioactive compounds through their 106 functional groups, thereby retaining various hydrophilic and hydrophobic bioactive 107 ingredients and increasing their bioavailability and solubility. The bioactivity of a nutrient 108 is easily destroyed during passage through the GIT digestion, resulting in failure to 109 110 achieve the effective concentration in the plasma and tissue. However, polysaccharides 111 have a uniquely slow intestinal digestion rate, which is beneficial for the release of bioactive compounds. More interestingly, the protective effect can be significantly 112 enhanced by a cascaded polysaccharide self-assembled package. Moreover, the stability 113 114 of carbohydrates at high temperatures is beneficial for the protection of bioactive

115 components (Azuma, et al., 2014).

#### 116 **2. Carbohydrate-based self-assembled nanostructures**

117 Chemical complementarity, structural compatibility and weak or covalent interactions create bonds between the self-assembly building blocks to form a hierarchical 118 structure with different specific functions. The self-assembled structures 119 and 120 nanostructures can be subdivided according to their size and phase. Nanoparticles are usually divided by size into zero-dimensional, one-dimensional, two-dimensional and 121 three-dimensional nanoparticles; according to phase, they can be divided into 122 single-phase, multi-phase and complex-phase nanoparticles. Most carbohydrates in food 123 124 are one-dimensional. One-dimensional nanostructure with a higher surface area can be self-assembled by electrostatic spinning, which in turn contributes to higher embedding 125 rates (Rezaei, Nasirpour, & Fathi, 2015). A hydrogel is another means of forming an 126 assembled structure. Hydrogels can form strong gels via physical cross-linking at the 127 appropriate pH or temperature. Adding bioactive molecules (such as polyphenols or 128 flavor compounds) to a three-dimensional network can help solve the problem of delivery 129 of the bioactive to a specific location within the body (Shewan & Stokes, 2013). The 130 combination of xanthan gum-based hydrogels and multifunctional carbohydrate colloidal 131 132 nanofibers is an example of this type of structure (Gökmen, et al., 2015).

Two-dimensional self-assembled nanostructures include thin film coatings, laminates, and nanocomposites, which can be easily formed by interactions based on the local self-assembly of carbohydrate three-dimensional micelles. Micelles are invisible molecularly ordered aggregates in a solution formed by a number of solute molecules or

137 ions with hydrophobic groups as the core and hydrophilic groups as the shell. Amphiphilic polymers are macromolecules with both hydrophilic and hydrophobic chains 138 in the same molecular chain of polymers. Amphiphilic polymers are usually copolymers 139 formed by blocking, grafting or other methods. The hydrophilic and hydrophobic groups 140 141 are mutually incompatible with each other and prone to micro-phase separation; thus, they 142 reduce the surface tension of the solution. The formation of amphiphilic polymer micelles is a spontaneous process of minimizing the contact area between the hydrophobic chains 143 and the aqueous solution. Due to intermolecular forces, such as hydrophobic interactions, 144 electrostatic interactions, hydrogen bonding, and the solvent hydrophobic accumulation 145 146 effect, the hydrophobic region forms a core, and the shell is wrapped by the hydrophilic chains formed outside the core, which stabilizes the micelles in solution. The main 147 148 driving force of micellar formation is the free energy reduction of the system due to the attraction between the hydrophobic chains and the repulsive force between the 149 hydrophilic chains to hydrophobic chains. Polymer micelles are prepared by a molecular 150 self-assembly method. This method does not involve any organic solvents, such as 151 emulsifiers or surfactants, and thus reduces the toxicity of the carrier. Moreover, the 152 self-assembly method is a simple and spontaneous process in which molecules or subunits 153 154 form a supramolecular structure without additional energy input. This type of method includes direct hydrolysis, ultra sonication, dialysis methods and solvent evaporation. The 155 method chosen for a specific case depends on the dissolution and swelling ability of the 156 polymer in water. Direct hydrolysis and ultrasonic methods are usually applied to 157 water-soluble polymers or polymers with good swelling properties; dialysis or solvent 158

159 evaporation methods are usually applied to polymers with poor water solubility, such as nano-bio complexes composed of chitosan and cellulose with covalent bonds. 160 Laver-by-laver assembly is an important technique for the development of nanoscale 161 three-dimensional structures. Negatively charged polyanions and positively charged 162 polycations are assembled layer by layer by electrostatic attraction. This is an emerging 163 164 technology in the food industry and can effectively encapsulation to protect color and flavor substances and prevent the collapse of food structures. For example, biopolymers 165 composed of xanthan gum and galactomannan through non-ionic interactions can extend 166 epidermal growth factor (EGF) release time by 5-fold (Kaminski, Sierakowski, Pontarolo, 167 dos Santos, & de Freitas, 2016). The following sections summarize the advances in 168 delivery systems based on carbohydrates from variable biosources (Fathi, Martín, & 169 170 McClements, 2014).

171 **3. Plant-based carbohydrates** 

172 *3.1. Starch* 

Starch and its derivatives have become the most studied and most popular polymer 173 materials for use as active carriers. The size of natural-origin starch varies between 1 and 174 100  $\mu$ m. Amylose consists of glucose units with  $\alpha$ -1,4 glycosidic bonds while 175 176 amylopectin consists of main chains linked by  $\alpha$ -1,4 glycosidic bonds and side chains linked by  $\alpha$ -1,6 glycosidic bonds. The process of spherulite formation in starch is the 177 result of self-assembly, which produces starch nanoparticles. One assembly method 178 involves inducing the encapsulation of fatty acids by amylose or other surfactants, 179 180 followed by a combination of both to produce a supramolecular amylose spiral complex.

181 Sodium dodecyl sulfate (SDS), polysorbate 80 (Tween 80) and sorbitan monooleate (Span 80) can be used to control the size and surface morphology of starch nanoparticles to 182 183 ensure the production of ultrafine nanoparticles with great thermal stability and dispersion. This process can be generated by mechanically assisted processing, such as ultrasonic, 184 extrusion, autoclaving, or enzymatic treatment and acid hydrolysis (Xiaojing Li, et al., 185 186 2016). The starch nanoparticle complex can help self-assemble the insoluble small molecules together with the soluble protein through electrostatic interactions to form a 187 nanoparticle carrier delivery system for loading the insoluble functional food ingredients 188 (Bhopatkar, et al., 2015). The starch and its derivatives of the polymer micelles are 189 190 generally formed by ultrasound and dialysis methods. Initially, an ultrasonic method is used to dissolve a starch-based polymer in an aqueous solution, followed by an ultrasonic 191 192 method to disperse it evenly in the aqueous solution and to form micelles through intermolecular hydrophobic interactions. The size of the micelles can be controlled by the 193 duration of the ultrasonic treatment. While this method is simple, the stability of the 194 micelles is unsatisfactory. The dialysis method involves first dissolving hydrophobically 195 modified starch in a solvent that is mutually soluble with water, such as dimethyl 196 sulfoxide (DMSO), N,N-dimethylformamide (DMF) or tetrahydrofuran (THF), followed 197 by dialysis in water. Polymer micelles self-assemble to form micelles as the organic 198 solvent is dialyzed away. The size, dispersibility and yield of the polymer micelles are 199 200 related to the organic solvents. Micelles formed by this method are usually of small size and great dispersibility. 201

#### 202 *3.1.1. Formation of starch crystals*

8

Another avenue with good prospects is targeting the release and delivery of bioactive molecules with starch as the matrix under specific controlled conditions. In particular, amylose can also form clathrates with various molecules, such as volatile flavor compounds or fatty acids. Amylose spikes can also be formed by amylose and guest molecules through non-covalent interactions. These amylose spikes have a strong tendency to self-assemble into supramolecular structures via helix-helix synergism.

Other spherulites are mainly composed of amylose and lightly branched starch 209 polymers formed by "high-temperature regeneration". The long-chain amylose is 210 211 concentrated along the radial direction of the spherulites, and the short-chain amylose is 212 concentrated along the tangential direction of the spherulites. These two types of prepared starch can decrease the digestion rate or increase resistance to enzymatic digestion. The 213 digestion rate depends on the self-assembly morphology and surface characteristics: a 214 dense and smooth surface is not conducive to decomposition by digestive enzymes (Fig. 1) 215 (Kiatponglarp, Rugmai, Rolland-Sabaté, Buléon, & Tongta, 2016). 216

217 *3.1.2. Chemical modification* 

Starch molecules contain large numbers of hydroxyl groups, which makes them hydrophilic. However, the solubility of starch in cold water is quite poor, mainly due to the easy formation of hydrogen bonds among hydroxyl groups. Therefore, the physical and chemical properties of natural starch limit the application of starch in many foods. Chemical modification linking hydrophobic and hydrophilic groups can adjust the self-assembly of starch and thus overcome these limitations. Hydrophobic modification is mainly achieved through an esterification reaction, which not only destroys the

225 intermolecular hydrogen bonds to improve the solubility but also makes the starch amphiphilic. Because starch exhibits bioactivities, including anti-inflammatory, anti-viral, 226 227 nontoxigenic and non immunogenic, with very good biocompatibility, and biodegradability, starch and its derivatives are now considered ideal materials for the 228 preparation of biological nano-carriers and are commonly used to encapsulate insoluble 229 230 bioactive molecules. Modifications mainly improve the physical and chemical properties of the hydroxyl, hydrophilic or hydrophobic groups of starch through grafting, 231 esterification and etherification. Self-assembly in an aqueous solution can easily form 232 micelles and polymer vesicles (Fig. 2) (Marefati, Sjöö, Timgren, Dejmek, & Rayner, 233 2015). 234

For example, hydrophobically modified octenyl succinic anhydride (OSA) starch 235 236 derivatives are most widely used for encapsulating volatile perfumes. The function of OSA starch is primarily based on steric hindrance, which preferentially moves to the 237 gas/water interface when dissolved in water. The hydrophilic carboxyl groups extend into 238 the aqueous phase, while the lipophilic octenyl chains extend into the oil phase; the 239 interface tension of the OSA starch in the oil/water interface through the hydrophobic 240 octenyl action is lower compared to other starch derivatives, which helps form a 241 continuous boundary layer and greatly enhances intermolecular cohesion (Wang, Yuan, & 242 Yue, 2015). The increase in the degree of substitution (DS) and the decrease in the critical 243 concentration at the time of embedding contribute to the decrease in the size of the starch 244 nanospheres and the formation of nanospheres with spherical and/or sharp edges 245 encapsulating the hydrophobic substance resulting in strong stability (Gu, Li, Xia, 246

247 Adhikari, & Gao, 2015).

248 3.1.2.1. Grafted starch polymer micelles

249 There are two types of grafted polymers: one consists of a hydrophobic backbone and hydrophilic side chains, and the other consists of a hydrophilic backbone and 250 hydrophobic side chains. Both grafted polymers can more easily form a core-shell 251 252 structure micelle with one chain directed inwards and the other chains directed outwards in the aqueous solution through self-assembly compared to the block polymer. The size, 253 structure and properties of the grafted polymer micelles can be effectively controlled by 254 the configuration of the polymer, the length and number of side chains and the grafting 255 points. Therefore, it is usually easier to synthesize amphiphilic grafted polymers than 256 block polymers. The grafted starch-based polymer micelles are mostly chemically 257 258 modified by esterification and etherification. Hydrophobically modified starch can be used as a self-assembled biopolymer for the protection of insoluble bioactive substances. 259

260 3.1.2.2. Block starch polymer micelles

Two or more polymers of different compositions and properties are linked by 261 chemical bonds to form a block polymer, and the large number of reactive hydroxyl 262 groups in starch molecules contributes to the introduction of a block polymer. The 263 amphiphilic starch-based polymer has both hydrophilic and hydrophobic chains. When it 264 is placed in a solvent with different dissolution abilities for each type of chain, the 265 polymer can be self-assembled in an aqueous solution due to a large difference in 266 solubility, forming a polymer core with a unique core-shell structure, and the hydrophobic 267 groups in the aqueous environment can aggregate into the core and are surrounded by the 268

269 hydrophilic chains.

Currently, the methods for making amylose-derivative block polymers mainly 270 271 include the enzymatic polymerization method, the coupling method and the active/controlled polymerization method. The latter two methods are research hotspots. 272 Enzymatic polymerization utilizes maltooligosaccharide as a substrate and phosphorylase 273 274 to catalyze the polymerization of the glucose-1-phosphate monomer to make amylose 275 block polymers. This method, while it can control the molecular weight of the starch chain very well is complicated. The coupling method requires the protection of hydroxyl 276 groups in the high-molecular-weight starch, followed by degradation to obtain 277 low-molecular-weight amylose derivatives with reactive functional groups. Finally, 278 coupling reactions among the functional groups result in block polymers, which are 279 280 biodegradable and can be degraded by  $\alpha$ -amylase. However, this coupling method is still cumbersome. 281

The active/controlled polymerization method is the most efficient and simplest 282 method for the synthesis of amphiphilic block polymers. In this method, click chemistry 283 is the main reaction, including the metal ion-catalyzed cycloaddition of azides and 284 alkynes and a condensation reaction between aldehyde and aminooxy groups. In the 285 presence of an active hydrogen compound,  $\varepsilon$ -caprolactone is prone to polymerization. The 286 poly (*ɛ*-caprolactone) (PCL) is then grafted to the backbone of maltose by click chemistry. 287 The size of the nanoparticles can be reduced with a decrease in the molecular weight of 288PCL or an increase in the number of hydrophilic groups (Isono, et al., 2016). 289

290 *3.1.2.3. Polyelectrolyte starch polymer micelles* 

12

291 Micelles that are formed in water-soluble block polymers in an aqueous solution by electrostatic interactions, hydrogen bonding or metal coordination are usually defined as 292 polyelectrolyte micelles. In this process, hydrophilic polymer chains self-assemble to 293 form a tether-like fence and wrap around the outer layer to maintain the spatial stability of 294 the micelles. The core is formed by the aggregation of part of the polymer blocks, which 295 296 is the result of intermolecular forces (hydrophobic interactions, electrostatic interactions, metal complexation and hydrogen bonding between block copolymers). For example, 297 hydrophobic apogossypolone (ApoG2) and hydrophilic adriamycin (doxorubicin, DOX) 298 299 both have good anti-tumor activity; if they are loaded into nanoparticles made of different 300 suitable materials, they can be absorbed by human cells or tissues due to their different sustained-release characteristics to achieve the best combination of treatments. Starch first 301 302 grafts to stearic acid, which is catalyzed by glycidyltrimethylammonium chloride (GTAC) to synthesize cationic stearic acid-grafted starch (CSaSt). This CSaSt amphiphilic 303 conjugate can easily encapsulate ApoG2 to form amphiphilic starch micelles (AAST MCs) 304 during self-assembly. Subsequently, DOX is adsorbed to excess hyaluronic acid (HA) 305 nanoparticles (DHA NPs) through electrostatic interactions. Finally, DHA NPs with 8-9 306 negative charge units, assemble with AAST MCs a positively charged unit by electrostatic 307 interactions to produce mulberry-like bicomponent nano-carriers (MLDC NCs). The 308 therapeutic dose of this two-component delivery system in vitro and in vivo is only 309 one-fifth that of the non-carrier two-component combination and can be used for targeted 310 311 therapy to the tumor (K. Li, et al., 2015).

312 *3.2. Cyclodextrin* 

313 Cyclodextrin (CD) is an enzymatic product of amylase and is included in the US Food and Drug Administration's (USDA) list of products generally recognized as safe 314 (GRAS). The most common natural  $\alpha$ -,  $\beta$ - and  $\gamma$ -CD contain 6, 7 and 8 315 D-(+)-glucopyranose units, respectively, bound by  $\alpha(1-4)$  glycosidic bonds. CD with 316 more than eight glucose units also exists; however, it is extremely difficult to generate and 317 318 purify in the real environment. CD contains a relatively hydrophobic internal cavity 319 structure, which can form complexes with a variety of molecules, including fat, flavor substances and pigments, through molecular self-assembly. The appropriate complex 320 molecular size can result in greater connection strength. Therefore, CD is widely used to 321 322 protect fragrance or unstable flavour compounds of food formulations in harsh environments during food processing, storage and delivery. 323

Because CD contains primary and secondary hydroxyl groups, the surface of CD is 324 hydrophilic, which can effectively promote the conversion of complexed volatile matter 325 from gas or liquid phase into powder. The encapsulation of CD and guest molecules is 326 affected by many factors, including size, geometry and electrical properties. Size refers to 327 the match between the CD cavity and the corresponding guest molecule. The geometry 328 and the stereogenic effect of the guest molecule can also affect the complex, and 329 furthermore, due to the highly hydrophilic effect of ionic guest molecules, they can only 330 form a weak complex with CD, while weakly polar guest molecules can complex with 331 CD more strongly (Jahed, Zarrabi, Bordbar, & Hafezi, 2014). Because of these unique 332 structures and properties, CD can be widely used in the construction and regulation of 333 aggregates in supramolecular self-assembly. 334

335 CD can also be used to protect against oxidative degradation and heat- and light-caused decomposition of edible flavors and bioactive substances. β-cyclodextrin has 336 been found to be an effective aromatic compound retention agent during the heat 337 treatment process due to its relatively suitable cavity volume (Kfoury, Auezova, 338 Greige-Gerges, & Fourmentin, 2015). β-cyclodextrin is also widely used because it is 339 340 reasonablely priced (Celebioglu, Kayaci-Senirmak, İpek, Durgun, & Uyar, 2016). In addition, cyclodextrin can effectively prevent enzymatic oxidation caused by polyphenol 341 oxidase or phenolic compounds in fruit juices by non-covalent interactions with 342 polyphenol oxidases (Aguilera, et al., 2016). 343

CD preserves food aromas and flavors and controls their release during storage and 344 consumption, improving the solubility and retention of these insoluble substances. The 345 encapsulation of cyclodextrins also enhances the stability of fragrance and taste, prolongs 346 the shelf life of products, protects products from isomerization and oxidation during 347 storage, and creates a controlled release system for active substances. Cyclodextrin 348 inclusions are also used to change food flavors; eliminate food bitterness and odors, such 349 as the deodorization of soy milk, soy protein and some fish and rice; and the unpleasant 350 spicy or bitter (Preis, Grother, Axe, & Breitkreutz, 2015) effects of curcumin in food 351 and medicine.  $\beta$ -cyclodextrin is also used as a cholesterol chelator for livestock products 352 (such as egg yolk, milk, butter, lard, cream, and cheese), which can reduce cholesterol by 353 87.54% (Lamas, et al., 2016) in β-cyclodextrin-egg cross-linking derivatives to improve 354 its quality (Y. Li, Chen, & Li, 2017). β-cyclodextrin can also be used to replace egg yolk 355 in bread to maintain its physico-chemical and sensory properties during storage (Marcet, 356

Paredes, & Diaz, 2015). Another application of CD is in the design of nano-bioactive food packaging materials. CD can be incorporated as a fragrant seasoning, a bacteriostatic substance or a bioactive agent, which allows it to be gradually released into food to maintain the sensory nature and to prevent the growth of microorganisms. The slow release of carvacrol in packaging is a good example (Lavoine, et al., 2014).

362 Emulsions are highly valuable because they are widely used in food, pharmaceuticals and cosmetics. Stabilized emulsions are usually obtained by adsorbing 363 surfactants, polymers, or particles at the liquid-liquid interface to inhibit emulsion 364 delamination, flocculation and coalescence. Recently, CD has been used as a substitute 365 for traditional surfactants in emulsion stabilization. In contact with oil components, CD 366 can form amphiphilic supramolecules at the oil/water interface with oil molecules via 367 self-assembly. This supramolecule has a surface activity that can significantly reduce 368 oil/water interfacial tension. Oil-CD inclusion complexes (ICs) are connected to the 369 surface of the emulsion droplets by means of CDs in the aqueous phase. The particles 370 formed by CD and oil are distributed on the surface of the oil droplets. These ICs can 371 further grow into microcrystals and produce dense layers that adhere to the surface of 372 emulsion droplets, similar to Pickering emulsions (solid particle-stabilized emulsion), 373 which can stabilize emulsions. Compared to surfactant-stabilized emulsion, the stability 374 of Pickering emulsions is less affected by pH, salt and temperature and results in none of 375 the environmental pollution or toxicity problems associated with other surfactants. 376 Sometimes, CD and a surfactant are added together through interfacial diffusion, 377 adsorption and rearrangement to form more stable emulsions with various configurations 378

(Fig. 3) (H. Xu, Liu, & Zhang, 2015). However, not all surfactants can stabilize emulsions
as the type, amount, and initial position of a surfactant can all affect their stability (Xue Li,
et al., 2014).

Natural cyclodextrin has limited bonding capacity and is even unstable in certain 382 circumstances (i.e., a strongly acidic environment). In particular, the solubility of 383 384  $\beta$ -cyclodextrin in water and other solvents is not high, which limits the application of cyclodextrin. Therefore, it is necessary to modify the structure of cyclodextrin and 385 synthesize cyclodextrin derivatives that can effectively overcome these challenges. 386 Common cyclodextrin derivatization methods include alkylation, acylation, amination, 387 silvlation and azide. Cyclodextrin contains three different types of hydroxyl groups, 388 located at C2, C3 and C6, pointing to two different directions of the cyclodextrin 389 390 molecular ring structure. The reactivity of each hydroxyl group varies under different reaction conditions and results in distinct substitution products. Table 1 lists the assembly 391 objects and methods, basic properties of common starch and cyclodextrin and its 392 derivatives, and their recent applications in food science. 393

394 *3.3. Inulin* 

Inulin is a fructose polymer linked by  $\beta$ -(1-2)-D-fructofuran, and the length of the fructose chain varies from 2 to 60 monomers. The main function of inulin is the storage of carbohydrates in most plants (Apolinario, et al., 2014). Inulin has a variety of nutritional functions, including immune activity, hypolipidemic effects, prebiotic effects (affecting intestinal microbiota), and stimulation of the absorption of minerals (calcium and magnesium). The self assembly of inulin particles has been widely studied in comparison

17

401 to glucan and fructans and it has been shown that the functionality of inulin has great potential for further study. The precipitation of inulin initiates from interactions among 402 hydrogen bonds between adjacent chains and assembly from random coils to a helical 403 one-dimensional conformation. The helical conformation grows into a secondary structure 404 405 due to the interaction between glucose at one end of the inulin chain and fructose on the adjacent chain. Subsequently, the tertiary structure of inulin is formed via cross-linking of 406 every five hydrogen bonds and two-dimensional semi-nano crystal layers (Fig. 4) 407 (Barclay, et al., 2016). Both the degree of polymerization and the process have a large 408 impact on the physical and chemical properties of inulin. The furanosyl content of inulin 409 410 is smaller and more flexible than that of glucan dextran, and the backbone structure has higher molecular elasticity and hydrophobicity and, therefore, relatively low glass 411 412 transition and melting points relative to other oligosaccharides and polysaccharides. However, unlike the other sugars described above, inulin cannot be metabolized by the 413 human body, which favors it for some unique and positive attributes, such as metabolic 414 utilization by the kidney and beneficial colonic bacteria such as lactobacill and 415 bifidobacteria. Inulin is more suitable as a food adhesive than glucose or lactose due to its 416 reduction groups (Mensink, Frijlink, van der Voort Maarschalk, & Hinrichs, 2015). With 417 an increase in inulin polymerization, the glass transition temperature (Tg) of inulin also 418 increases, but the degree of polymerization of the self-assembled natural inulin active 419 compound does not have an impact on Tg or the retention of embedded substances. The 420 particles formed by embedded substances with high DP after drying are of low moisture 421 422 and can help improve the stability of emulsions after re-dissolution (Silva, Zabot, Bargas,

423 & Meireles, 2016). However, the ability of natural inulin as a single packaging material for self-assembling hydrophobic active compounds is limited. Therefore, it is important to 424 425 combine natural inulin with other low-calorie biopolymers (such as maltodextrin, modified starch, chitosan, protein or gum) to improve the rate of interfacial adsorption; 426 427 thus, the wall material can have the dual function of prebiotic functional characteristics 428 and self-assembling embedding ability (Dima, Pătrașcu, Cantaragiu, Alexe, & Dima, 2016). In addition, inulin can be self-assembled after hydrophobic modifications and 429 provide a more useful function. The resulting derivatives can be used as suitable 430 aggregators and stabilizers of emulsions, and they can effectively encapsulate and deliver 431 water-insoluble active compounds in food, pharmaceutical or cosmetic products. Table 1 432 lists common inulin assembly objects, the basic nature of the method and recent 433 434 applications in food science.

435 *3.4. Cellulose* 

Cellulose is the most abundant polysaccharide in nature and includes hundreds to thousands of straight-chain  $\beta$ -(1 $\rightarrow$ 4)-linked D-glucose units. Although the amount of cellulose is vast, humans consume only a small amount of it because it is difficult to digest, making it mainly a nutritional source for animals. Nevertheless, due to the wide range of sources of cellulose, scientists are still working on developing products utilizing cellulose that are acceptable for both ruminant and non-ruminant animals.

442 At present, the most widely used celluloses in the food industry are microcrystalline 443 cellulose and bacterial cellulose. They are non-toxic, tasteless, and safe and have unique 444 physical and chemical properties which do not affect product quality, and they can be

445 used as food additives, such as emulsifiers, thickening agents, foam stabilizers, or nutritional fortifiers or food packaging materials to make cellulose films. Microcrystalline 446 447 cellulose is used in dairy products for thickening and gelling the water phase in an oil-water emulsion, to prevent the aggregation and combination of oil droplets and thus 448 stabilize the suspension. Cellulose is a dietary fiber, and the cellulose self-assembly 449 450 strategy is harnessed mainly to improve its functional applications. However, the rigid 451 molecular chain structure and the formation of hydrogen bonds result in insolubility in common solvents. Therefore, there are relatively few studies on the direct hydrophobic 452 modification of cellulose as a hydrophilic skeleton. However, recent studies have shown 453 that interactions between amphiphilic and hydrophobic parts of unmodified cellulose play 454 an important role in determining the crystal structure and solubility of cellulose, which 455 456 allows hydrophobic modifications at low temperatures in organic, acidic, or alkaline solvents. The hydrophobic modification of cellulose in food is usually a necessary 457 condition for maintaining the stability of the emulsion, which produces amphiphilic 458 cellulose or cellulose derivative-based polymers. These amphiphilic polymers can 459 self-assemble into nanoparticles, which is one type of grafting method. The factors that 460 can impact on self-assembly include the cellulose skeleton length, hydrophobic chain 461 length and density, concentration, polarity, temperature and pH of the solution. 462 Nanocrystalline cellulose is usually used to stabilize emulsions; the amphiphilicity of 463 cellulose crystallization allows it to assemble layer by layer and stabilize the emulsion 464 through the Pickering mechanism. Over the past few decades, dissolvable and 465 regeneratable forms of cellulose have been extensively studied to improve the enzymatic 466

467 hydrolysis rate of cellulose. In the regeneration step, phosphoric acid is often used to remove the solvent (water) from the cellulose molecule allowing cellulose to 468 self-assemble into nanostructured cellulose. Moreover, some species of bacteria can also 469 biosynthesize cellulose nanocrystals using glucose as the substrate (Dayal & Catchmark, 470 471 2016), but the method has proven difficult to commercialize thus far. Nanostructured 472 cellulose plays an important role in flavor delivery (Hao, et al., 2015). Electrostatic technology can also effectively assist cellulose in assembling functional substances 473 (Rezaei, et al., 2015). Table 1 lists the basic properties of cellulose, methods for its 474 475 assembly and recent developments in its application in food science and industry.

476 *3.5. Pectin* 

Pectin is an important food gelation agent that is most commonly found in the cell 477 478 wall of terrestrial plants. Pectin is composed of a number of  $\alpha$ -D-(1 $\rightarrow$ 4) galacturonic acid residues through linear linkage. DE (degree of esterification) is an important factor in 479 characterizing the esterification of the carboxyl and methanol groups of pectin. DE is the 480 ratio of the esterified GalA groups to the total GalA groups, and the DE value of pectin in 481 general ranges from 60% to 90%. Pectins with a DE value greater than 50% are known as 482 high-methoxy pectins (HM pectins), and pectins with DE value less than 50% are known 483 as low-methoxy pectins (LM pectins). LM pectin can be further subdivided into two 484 categories: amidated LM and conventional LM pectin. LM pectin has the characteristics 485 of anti-hydrolysis on oral, gastric and intestinal microbial-specific enzymolysis. Therefore, 486 it can be used as an effective carrier for bioactive substances that are sensitive to acids. 487 488 However, as a protective carrier, its solubility is often high.

489 The hydrogen bonding and hydrophobic interactions are the main forces driving the aggregation of pectin molecules. For pectin molecules in the neutral or slightly acidic, 490 most of the unesterified carboxyl groups are present in the form of ionized salts and this 491 causes repulsion between negative charges preventing pectin from forming a network. 492 This scenario also reduces the attractive forces between pectin and water molecules and 493 494 the repulsive forces between pectin molecules. However, LM pectin requires calcium (or 495 other polyvalent cations) for proper gel formation. The affinity and the capability of divalent cations to form a gel can be ranked as follows: barium> strontium> calcium. The 496 mechanism of LM pectin gelation is well known as the "egg box" model. GalA monomers 497 in a specific sequence of galacturonic acid in adjacent chains are interlinked by 498 electrostatic and ionic bonding through carboxyl groups. A large amount of sugar (i.e., 499 500 60% or higher) forms hydrogen bonds by dehydration to reduce gel formation. Many studies have added calcium salts and LM pectin (to increase the binding capacity of 501 pectin to calcium) to matrix tablets to delay the release of embedded substances (Fig. 5a 502 and 5b) (Sriamornsak, 2011). Gel beads are formed by LM pectin with calcium, which 503 can cross-link to form a poly-Gal chain. Pectin beads are produced by ionic gel 504 preparation and often perform as a delivery system for the sustained release of active 505 506 materials. However, their release in vitro is usually accelerated. By changing the LM pectin DE, the calcium pectin gel beads can be changed and the release mode can be 507 modified. 508

509 Therefore, scientists often follow several strategies to develop pectin-derivative 510 carriers. First, pectin needs to be biodegradable by bacteria and highly soluble. An

511 alternative is to allow interactions between functional components and pectin molecules 512 to modify the local structure, for example, electrostatic interactions with proteins (such as lactoglobulin) to form 150-500 nm nanoparticles and then become a delivery system for 513 active substances, including  $\omega$ -3 unsaturated fatty acids and others (Jones & McClements, 514 515 2010; Zimet & Livney, 2009). A polyelectrolyte complex (PEC) is formed by electrostatic 516 interactions between the polymer electrolytes and their counterparts with opposite charges in aqueous solution. The structure of pectin contains many carboxyl groups, thus allowing 517 pectin to interact with the oppositely charged membrane or liposome. Self-assembled 518 pectin-liposomal nanocomposites (PLNS) can be prepared by simple mixing of a pectin 519 520 solution with cationic liposomes. The carboxyl group in galacturonic acid in pectin can be methyl esterified or reacted with an ammonia group to form an amide group. The 521 522 esterification degree (DE) and amidation degree (DA) are important methods of pectin classification. High-methoxylated pectin (CU201, 70% DE, 200 kDa), low-methoxylated 523 pectin (CU201, 38% DE, 80 kDa) and low-methoxylated amide pectin (CU020, 29% DE, 524 20% DA, 150 kDa) can interact with amine groups of liposomes (SA). AFM images show 525 a small amount of branched pectin and spherical liposomes (Fig. 5c) (Sriamornsak, et al., 526 527 2008).

528 Changes in zeta potential and particle size are important factors for verifying the 529 success of liposome coating. The high charge density on the surface of liposomes 530 promotes the adsorption of pectin. Moreover, a high concentration of pectin with an 531 increase in the coated liposome size results in better coverage of liposomes. The increase 532 in the high-fat pectin ratio may be due to bridging flocculation. The appropriate ratio of

533 pectin can help produce stable monolayered liposomes and prevent bridging flocculation

534 (Fig. 5d) (Alund, Smistad, & Hiorth, 2013).

Moreover, the interactions between pectin and a cationic solution environment can be used to change the gel properties to improve the embedding rate of functional substances. With an increase in the salt ion content, unlike the alginate gel, the number of bar-like interfaces decreases, while the number of dot-like crosslinks increases, and semi-flexible chains are also present (Fig. 5e and 5f) (Ventura, Jammal, & Bianco-Peled, 2013). Table 1 lists pectin assembly objects, the basic properties of the method and recent

541 developments in the food industry.

542 3.6. Animal polysaccharides

543 *3.6.1. Chitosan* 

Chitosan is mainly found in shells of shrimp and other crustaceans. Chitosan essentially consists of a large number of (1-4)-glycosidic bonds linking N-acetyl-2amino-2-deoxy-d-glucopyranose (glucosamine) and 2-amino-2-deoxyd-glucopyranose. Chitosan is widely used in the food industry for its antibacterial, clarification, and deacidification properties and its mouth-feel. Moreover, chitosan can be used as a controlled release medium for antioxidants, nutrients, spices and drugs.

550 Chitosan's cations and hydrophobic sites make it a polyelectrolyte and amphiphilic 551 compound. In acidic media, chitosan is highly protonated due to the positively charged 552 amino chitosan groups. Moreover, both intramolecular and intermolecular hydrogen 553 bonds (due to the presence of -OH single bonds and  $NH_2$  single-bond groups in the 554 chitosan backbone) contribute to self-assembly. The cationicity of chitosan is

555 advantageous for its incorporation into the surface of the anionic species by electrostatic forces and hydrogen bonding. Biopolymers with the opposite charge, such as alginates, 556 557 pectins, xanthan gum, carrageenan, acacia and anionic lipids, can provide interesting nanostructures in delivery systems for various components. For example, the NH<sub>3</sub> groups 558 559 of chitosan and the phosphate groups of modified lecithin can assemble via electrostatic 560 interactions to deliver hydrophilic compounds. Nanoparticles show excellent stability at certain pH (3-6) and ionic strength ranges, and nanoparticles can be easily made into 561 freeze-dried powders (Chuah, Kuroiwa, Ichikawa, Kobayashi, & Nakajima, 2009). A 562 self-assembled complex of chitosan with  $\alpha$ -lactalbumin or with  $\beta$ -lactoglobulin can also 563 be used as potential delivery vehicles (Lee & Hong, 2009). Moreover, since the chitosan 564 backbone has many functional groups, it can assemble better into nanomaterials after 565 566 chemical modification to assemble active ingredients in food or drugs (Y. Yang, et al., 2014). The specific methods include the following: (1) the ionic cross-linking method. In 567 an acidic medium, the amino group in the chitosan chain is easily protonated, so that it 568 has a certain positive charge. Under certain conditions, chitosan with positive charges is 569 mixed with anionic cross-linking agents (such as carboxylic acid or tripolyphosphate 570 (TPP)) in aqueous solution, and the protonated amino group can interact with the anionic 571 cross-linking agents to wrap the core materials to form chitosan-based nano-capsules. 572 Cross-linking between TPP and low-molecular-weight chitosan can produce nano-573 capsules with an average particle size of 138 nm. Nano-capsules made using this method 574 do not require organic solvents, and the cross-linking agent is of low toxicity. Therefore, 575 this process is widely used to make sustained-release capsules, but the disadvantage is 576

577 that the particle size distribution is wide and the stability is poor (Fan, Yan, Xu, & Ni, 2012). (2) Layer-by-layer self-assembly method. This method is suitable for the 578 preparation of double- or multi- layer nano-capsules. The main principle is to use 579 easy-to-remove or need-to-be-covered nano-particles as a template and to alternately 580 deposit polyelectrolytes with different charges on the surface of nanoparticle. Finally, the 581 582 particles are removed to obtain nano-capsules. Liposomes are an artificial membrane with targeted drug delivery properties. They have good biocompatibility, but their structure is 583 unstable and can be easily damaged by changes in external temperature and pH. To 584 improve the stability of liposomes, nano-lipid particles are wrapped in layers of chitosan 585 and alginate capsules by the layer-by-layer method, which not only improves the stability 586 of liposomes but also enhances their sustained release to a certain extent. This method not 587 588 only effectively controls the thickness of capsule shells but can also impart the directional release properties of capsules through the introduction of various capsule wall materials 589 (W. Liu, Liu, Li, & Liu, 2013). (3) Re-coagulation method. In this method, core 590 materials are distributed into two or more packaging materials with different charges, 591 followed by interactions among packaging materials by the adjusting pH, temperature, 592 and concentration of the system, which results in a low-solubility complex that 593 precipitates to form nano-capsules. The nano-capsules are prepared on the surface of 594 nanotubes re-coagulation 595 carbon by the method between chitosan and ethylenediaminetetracarboxylic acid. This nano-capsule not only achieves the effective 596 coating of enzymes and some proteins but can also be used as a biosensor or bioresponder 597 (Fig. 6) (H. Liu, et al., 2013). Table 1 lists the assembly objects and methods of chitosan 598

- and its recent applications in the food industry.
- 600 *3.7. Algae polysaccharides*

601 Acid polysaccharides are another specially constructed type of polysaccharide found in seaweed, including fucoidans and laminarans in brown algae (Phaeophyceae), 602 carrageenans in red algae (Rhodophyceae), and ulvans in green algae (Chlorrigyceae) 603 604 (Wrigstedt, et al., 2010). Research into the delivery of active substances using acid polysaccharides is ongoing (Venkatesan, Anil, Kim, & Shim, 2016). These 605 polysaccharides have a stronger ability to bind to proteins in self-assembly and therefore 606 are used in dairy and meat products. For example, the interaction between fucoidan and 607 bovine serum albumin at pH 4.0 through -SO<sup>3-</sup> and NH<sup>3+</sup> favours the formation of 608 complexes with very dense and intimate internal structures; the complex can dissociate 609 into the soluble state in the presence of 0.01 M NaCl at pH 4.5 (D.-Y. Kim & Shin, 2015). 610 Moreover, the thermostability, solubility and emulsification around PI of complexes 611 prepared at high temperature can be improved (D.-Y. Kim & Shin, 2016). Moreover, 612 chitosan and fucoidan can form multilayered hydrated nanocomposites via electrostatic 613 attraction, which can further embed bioactive compounds. Carrageenan is another 614 polysaccharide gel widely used in the food industry, which can thicken and stabilize 615 liquids. Carrageenan can also bind strongly to food proteins. In an in vitro model, 616 carrageenan could bind to protamine to form carrageenan/protamine polyelectrolyte 617 nanocomposites (Dul, et al., 2015). Carrageenan can also interact with surfactants, the 618 cationic glycine betaine amide, of different concentrations to form multiscale nanoparticle 619 complexes, which can significantly reduce the electrostatic interactions between the 620

621 surfactant and the polymer and gradually dissociate the polymer nanostructures (Gaillard, et al., 2017). Alginate is another brown anionic acidic polysaccharide and is widely 622 distributed in the cell wall of brown algae. Alginate is mainly composed of  $\alpha$ -l-guluronic 623 acid and  $\beta$ -D-mannuronic acid residues linked by 1,4-glycosidic linkages. Alginate can be 624 combined with 200-300 times its own weight of water to form a sticky gel. The formed 625 626 gel does not provide any nutrients, but it can stabilize the emulsion system. In the self-assembly process, alginate is often used as a base carrier and complexes with 627 chitosan to carry flavors, such as capsaicin and other flavor substances. Chitosan/alginate 628 self-assembles layer by layer to form a biofilm with an anti-fungal or anti-bacterial 629 coating (F. Jiang, Yeh, Wen, & Sun, 2015). Table 1 lists the algae polysaccharide 630 assembly objects, the basic methods and its applications in the food industry. 631

#### 632 3.8 Applications in Biomedicine and Environment

Polysaccharides and their derivatives are superior to synthetic polymers because of their non-toxicity, biodegradability, compatibility and low cost. They are also widely used in biomedical and environmental fields, such as tissue engineering, biological imaging or environmental utilization.

Polysaccharides and their derivatives, applied in tissue engineering, are often used in biological signal transduction, cell adhesion, cell proliferation, cell differentiation, cell responsive degradation, re-modeling, regeneration or planning of the shape and structure of cell growth, etc. Sugar as a biological scaffold in tissue engineering can meet the requirements of bio-compatibility, non-toxicity, biodegradable rate, appropriate porosity and structural integrity (Khan & Ahmad, 2013).

643 Biological imaging is an important tool for understanding key physiological information and pathological processes, such as cancer detection and treatment, stem cell 644 transplantation, immunogenicity and tissue engineering. Biological imaging has 645 advantages in using fluorescence as signal output. Compared with small organic dyes, 646 fluorescent nanomaterials exhibit excellent light stability, adjustable size emission, 647 648 multi-functional potential and ideal pharmacokinetic behavior. An alternative luminescent nanometer material of (FONs) fluorescent organic nanoparticles type appeared recently. 649 Chitosan is used to prepare ultra-small cross-linked chitosan nanoparticles by "one-pot" 650 multi-component reaction or atom transfer radical polymerization (ATRP), the particles 651 can show strong yellow or red color and have better water dispersibility (Wan, et al., 652 2016). β-cyclodextrin can be used to prepare red fluorescent organic nanoparticles with 653 654 aggregation-induced emission (AIE) characteristics (FON) (D. Xu, et al., 2017), or preparing an amphiphilic AIE active copolymer with strong green fluorescence through 655 host-guest interaction (H. Huang, et al., 2017). Starch produces AIE active polymer 656 nanoprobes with strong blue-green or green-yellow fluorescence through a "one-pot" 657 strategy with pH and glucose responsiveness and good biocompatibility (internalization 658 within 3 hours) (M. Liu, et al., 2015; Shi, et al., 2018). Oxidized sodium alginate (OSA) 659 can be used to prepare probes with red fluorescent AIE activity (Wan, et al., 2017). 660

The pollution of heavy metal ions in the environment has become a serious environmental problem and can exist in the natural environment for a long time, thus creating long-term risks to ecosystems and human. Copper ion is the most typical of heavy metal ions because it is involved in electroplating, paint, electricity, fertilizers,

wood manufacturing and pigments. Many adsorbents with small size and high specific surface area, including carbon nanotubes (CNT), magnetic nanoparticles, graphene oxide, silica nanoparticles, etc. have been used. Chitosan is rich in amino, carboxyl and hydroxyl functional groups and can be used as a coordination site. Carboxymethyl chitosan was immobilized on CNT to form CNT-based chitosan nanocomposites by the combination of mussel adsorption chemistry and Michael addition reaction, which could overcome the shortcomings of traditional CNT (Zeng, et al., 2016).

**6**72 **4** 

#### 4. Summary and future trends

Recent developments in healthcare products require the integration of new 673 technologies into customized functional food, which poses new challenges to food 674 scientists. The self-assembly strategy is an effective technique for the design and 675 676 manufacture of delivery agents for bioactive substances to effectively control interactions among food ingredients. Self-assembly technology plays a vital role in nutritional and 677 functional foods because of the great variation in structural, functional and physical and 678 chemical properties. Self-assembly technology can be used to assemble existing 679 micro-components to form larger structural units, which requires the manipulation or 680 control of structures. The appropriate substrates, environmental conditions and chemical 681 or physiological functions must be identified in advance. Carbohydrates, especially 682 polysaccharides, are a very suitable base matrix due to their structure and special 683 functions. Self-assembly has proven to be an attractive technology for the delivery of 684 685 food nutrients and functional components. Additionally, the application of nano-technology combined with self-assembly is the key to the manipulation of food 686

687 polymers to improve their functional structure, which can effectively improve the quality, feel, functional ingredients and shelf life of food. However, different monosaccharide 688 689 structures are used as building units, including isomeric stereoisomers, sequence changes, side chain connection, branching and distribution, and modified functional groups leading 690 to wide structural diversity of polysaccharides. More studies are needed to address the 691 692 problem of assembly and stability to improve the functions of assembled complexes. Parallel to developments in the laboratory, computer simulation and theoretical modeling 693 technology can also help solve key issues in the assembly process. More work on the 694 695 biomedical applications of carbohydrates/polysaccharides and their biological behavior in vivo, including adsorption, distribution, metabolism and excretion is 696 essential. This review explained recent developments in self-assembly strategies, 697 698 behaviors and methods, which can provide a useful reference for further studies of carbohydrate carrier delivery systems. When designing a delivery system, more 699 consideration needs be given to the requirements of the human body's internal 700 environment, in order to adjust the system more intelligently. This may be an area for the 701 702 future direction of development in this field.

703 5. Acknowledgments

This project was supported by the Research Fund for the National Natural Science Foundation of China (No. 31601473), the China-Ireland International Cooperation Centre for Food Material Science and Structure Design (No. KXGH17001), the Doctoral Program of Higher Education (No. 108/k41mke02a), the Natural Science Foundation of Fujian Province (No. 2018J01697), Spark Plan (No. 2017S0008), High Level

31

709	Construction Projects of Fujian Agriculture and Forestry University (No. 612014042),
710	Projects for Scientific and Technological Development of Fujian Agriculture and Forestry
711	University (No. KF2015099), the Program for Innovative Research Team in Science and
712	Technology in Fujian Province University (No. 2012-03), and the Scientific and
713	Technological Innovation Team Support Plan of Fujian Agriculture and Forestry
714	University (No. cxtd12009).
715	References
716	Abbas, S., Karangwa, E., Bashari, M., Hayat, K., Hong, X., Sharif, H. R., & Zhang, X.
717	(2015). Fabrication of polymeric nanocapsules from curcumin-loaded
718	nanoemulsion templates by self-assembly. Ultrasonics sonochemistry, 23, 81-92.
719	Aguilera, Y., Mojica, L., Rebollo-Hernanz, M., Berhow, M., de Mejía, E. G., &
720	Martín-Cabrejas, M. A. (2016). Black bean coats: new source of anthocyanins
721	stabilized by $\beta$ -cyclodextrin copigmentation in a sport beverage. Food chemistry,
722	212, 561-570.
723	Alund, S. J., Smistad, G., & Hiorth, M. (2013). A multivariate analysis investigating
724	different factors important for the interaction between liposomes and pectin.
725	Colloids and Surfaces A: Physicochemical and Engineering Aspects, 420, 1-9.
726	Apolinario, A. C., de Lima Damasceno, B. P. G., de Macêdo Beltrão, N. E., Pessoa, A.,
727	Converti, A., & da Silva, J. A. (2014). Inulin-type fructans: A review on different
728	aspects of biochemical and pharmaceutical technology. Carbohydrate polymers,
729	101, 368-378.

730 Auriemma, G., Mencherini, T., Russo, P., Stigliani, M., Aquino, R. P., & Del Gaudio, P.

731	(2013). Prilling for the development of multi-particulate colon drug delivery
732	systems: Pectin vs. pectin-alginate beads. Carbohydrate polymers, 92, 367-373.
733	Azuma, K., Osaki, T., Ifuku, S., Saimoto, H., Morimoto, M., Takashima, O., Tsuka, T.,
734	Imagawa, T., Okamoto, Y., & Minami, S. (2014). Anti-inflammatory effects of
735	cellulose nanofiber made from pear in inflammatory bowel disease model.
736	Bioactive Carbohydrates and Dietary Fibre, 3, 1-10.
737	Barclay, T. G., Rajapaksha, H., Thilagam, A., Qian, G., Ginic-Markovic, M., Cooper, P. D.,
738	Gerson, A., & Petrovsky, N. (2016). Physical characterization and in silico
739	modeling of inulin polymer conformation during vaccine adjuvant particle
740	formation. Carbohydrate polymers, 143, 108-115.
741	Beyki, M., Zhaveh, S., Khalili, S. T., Rahmani-Cherati, T., Abollahi, A., Bayat, M.,
742	Tabatabaei, M., & Mohsenifar, A. (2014). Encapsulation of Mentha piperita
743	essential oils in chitosan-cinnamic acid nanogel with enhanced antimicrobial
744	activity against Aspergillus flavus. Industrial Crops and Products, 54, 310-319.
745	Bhopatkar, D., Feng, T., Chen, F., Zhang, G., Carignano, M., Park, S. H., Zhuang, H.,
746	Campanella, O. H., & Hamaker, B. R. (2015). Self-Assembled Nanoparticle of
747	Common Food Constituents That Carries a Sparingly Soluble Small Molecule.
748	Journal of agricultural and food chemistry, 63, 4312-4319.
749	Celebioglu, A., Kayaci-Senirmak, F., İpek, S., Durgun, E., & Uyar, T. (2016).
750	Polymer-free nanofibers from vanillin/cyclodextrin inclusion complexes: high
751	thermal stability, enhanced solubility and antioxidant property. Food & Function,
752	7, 3141-3153.

33

753	Chen, M., Gao, C., Lü, S., Chen, Y., & Liu, M. (2016). Dual redox-triggered
754	shell-sheddable micelles self-assembled from mPEGylated starch conjugates for
755	rapid drug release. RSC Advances, 6, 9164-9174.
756	Cheong, A. M., & Nyam, K. (2016). Improvement of physical stability of kenaf seed
757	oil-in-water nanoemulsions by addition of $\beta$ -cyclodextrin to primary emulsion
758	containing sodium caseinate and Tween 20. Journal of Food Engineering, 183,
759	24-31.
760	Chopra, M., Kaur, P., Bernela, M., & Thakur, R. (2014). Surfactant assisted nisin loaded
761	chitosan-carageenan nanocapsule synthesis for controlling food pathogens. Food
762	Control, 37, 158-164.
763	Chuah, A. M., Kuroiwa, T., Ichikawa, S., Kobayashi, I., & Nakajima, M. (2009).
764	Formation of Biocompatible Nanoparticles via the Self-Assembly of Chitosan and
765	Modified Lecithin. Journal of food science, 74, N1-N8.
766	Chun, J., Jo, Y., Bjrapha, P., Choi, M., & Min, S. (2015). Antimicrobial Effect of α-or
767	$\beta$ -Cyclodextrin Complexes with Trans-Cinnamaldehyde Against Staphylococcus
768	aureus and Escherichia coli. Drying Technology, 33, 377-383.
769	Dayal, M. S., & Catchmark, J. M. (2016). Mechanical and structural property analysis of
770	bacterial cellulose composites. Carbohydrate polymers, 144, 447-453.
771	Dey, S., & Sreenivasan, K. (2014). Conjugation of curcumin onto alginate enhances
772	aqueous solubility and stability of curcumin. Carbohydrate polymers, 99,
773	499-507.

Dima, C., Pătrașcu, L., Cantaragiu, A., Alexe, P., & Dima, Ş. (2016). The kinetics of the

34

775	swelling process and the release mechanisms of Coriandrum sativum L. essential
776	oil from chitosan/alginate/inulin microcapsules. Food chemistry, 195, 39-48.
777	Dul, M., Paluch, K. J., Kelly, H., Healy, A. M., Sasse, A., & Tajber, L. (2015).
778	Self-assembled carrageenan/protamine polyelectrolyte nanoplexes-Investigation
779	of critical parameters governing their formation and characteristics. Carbohydrate
780	polymers, 123, 339-349.
781	Fan, W., Yan, W., Xu, Z., & Ni, H. (2012). Formation mechanism of monodisperse, low
782	molecular weight chitosan nanoparticles by ionic gelation technique. Colloids and
783	Surfaces B: Biointerfaces, 90, 21-27.
784	Fathi, M., Martín, Á., & McClements, D. J. (2014). Nanoencapsulation of food
785	ingredients using carbohydrate based delivery systems. Trends in Food Science &
786	Technology, 39, 18-39.
787	Fathi, M., Martin, A., & McClements, D. J. (2014). Nanoencapsulation of food
788	ingredients using carbohydrate based delivery systems. Trends in Food Science &
789	Technology, 39, 18-39.
790	Fioramonti, S. A., Perez, A. A., Aringoli, E. E., Rubiolo, A. C., & Santiago, L. G. (2014).
791	Design and characterization of soluble biopolymer complexes produced by
792	electrostatic self-assembly of a whey protein isolate and sodium alginate. Food
793	Hydrocolloids, 35, 129-136.
794	Fuenzalida, J. P., Nareddy, P. K., Moreno-Villoslada, I., Moerschbacher, B. M., Swamy,
795	M. J., Pan, S., Ostermeier, M., & Goycoolea, F. M. (2016). On the role of alginate
796	structure in complexing with lysozyme and application for enzyme delivery. Food

- 797 *Hydrocolloids*, *53*, 239-248.
- 798 Gökmen, F. Ö., Rzayev, Z. M., Salimi, K., Bunyatova, U., Acar, S., Salamov, B., & Türk,
- M. (2015). Novel multifunctional colloidal carbohydrate nanofiber electrolytes
  with excellent conductivity and responses to bone cancer cells. *Carbohydrate polymers*, *133*, 624-636.
- Gaillard, C., Wang, Y., Covis, R., Vives, T., Benoît, M., & Benvegnu, T. (2017).
  Monitoring the architecture of anionic κ-carrageenan/cationic glycine betaine
  amide surfactant assemblies by dilution: A multiscale approach. *Carbohydrate polymers*, 155, 49-60.
- Ge, W., Li, D., Chen, M., Wang, X., Liu, S., & Sun, R. (2015). Characterization and
  antioxidant activity of β-carotene loaded chitosan-graft-poly (lactide)
  nanomicelles. *Carbohydrate polymers*, 117, 169-176.
- Gu, F., Li, B., Xia, H., Adhikari, B., & Gao, Q. (2015). Preparation of starch nanospheres
  through hydrophobic modification followed by initial water dialysis. *Carbohydrate polymers*, *115*, 605-612.
- Han, L., Ratcliffe, I., & Williams, P. A. (2015). Self-Assembly and Emulsification
  Properties of Hydrophobically Modified Inulin. *Journal of agricultural and food chemistry*, 63, 3709-3715.
- 815 Hao, X., Shen, W., Chen, Z., Zhu, J., Feng, L., Wu, Z., Wang, P., Zeng, X., & Wu, T.
- 816 (2015). Self-assembled nanostructured cellulose prepared by a dissolution and
- 817 regeneration process using phosphoric acid as a solvent. *Carbohydrate polymers*,
- 818 *123*, 297-304.

819	Higueras, L., López-Carballo, G., Gavara, R., & Hernández-Muñoz, P. (2016). Effect of
820	hydroxypropyl- $\beta$ -cyclodextrin and coadjuvants on the sorption capacity of
821	hydrophilic polymer films for monoterpene alcohols. Carbohydrate polymers, 151,
822	1193-1202.
823	Hu, H., Xing, L., Hu, Y., Qiao, C., Wu, T., Zhou, G., & Zhang, W. (2016). Effects of
824	regenerated cellulose on oil-in-water emulsions stabilized by sodium caseinate.
825	Food Hydrocolloids, 52, 38-46.
826	Hu, Y., Zhang, J., Yu, C., Li, Q., Dong, F., Wang, G., & Guo, Z. (2014). Synthesis,
827	characterization, and antioxidant properties of novel inulin derivatives with
828	amino-pyridine group. International journal of biological macromolecules, 70,
829	44-49.
830	Huang, H., Xu, D., Liu, M., Jiang, R., Mao, L., Huang, Q., Wan, Q., Wen, Y., Zhang, X.,
831	& Wei, Y. (2017). Direct encapsulation of AIE-active dye with $\beta$ cyclodextrin
832	terminated polymers: Self-assembly and biological imaging. Materials Science
833	and Engineering: C, 78, 862-867.
834	Huang, Y. C., & Kuo, T. H. (2016). O-carboxymethyl chitosan/fucoidan nanoparticles
835	increase cellular curcumin uptake. Food Hydrocolloids, 53, 261-269.
836	Hundre, S. Y., Karthik, P., & Anandharamakrishnan, C. (2015). Effect of whey protein
837	isolate and $\beta$ -cyclodextrin wall systems on stability of microencapsulated vanillin
838	by spray-freeze drying method. Food chemistry, 174, 16-24.
839	Isono, T., Miyachi, K., Satoh, Y., Nakamura, R., Zhang, Y., Otsuka, I., Tajima, K.,
840	Kakuchi, T., Borsali, R., & Satoh, T. (2016). Self-Assembly of

- Maltoheptaose-block-polycaprolactone Copolymers: Carbohydrate-Decorated
  Nanoparticles with Tunable Morphology and Size in Aqueous Media. *Macromolecules*.
- Jahed, V., Zarrabi, A., Bordbar, A. K., & Hafezi, M. S. (2014). NMR (1 H, ROESY)
  spectroscopic and molecular modelling investigations of supramolecular complex

846 of  $\beta$ -cyclodextrin and curcumin. *Food chemistry*, *165*, 241-246.

- Jiang, F., Yeh, C. K., Wen, J., & Sun, Y. (2015). N-trimethylchitosan/Alginate
  Layer-by-Layer Self Assembly Coatings Act as "Fungal Repellents" to Prevent
  Biofilm Formation on Healthcare Materials. *Advanced healthcare materials, 4*,
- 850 469-475.
- Jiang, J., Chen, Y., Wang, W., Cui, B., & Wan, N. (2016). Synthesis of superparamagnetic
  carboxymethyl chitosan/sodium alginate nanosphere and its application for
  immobilizing α-amylase. *Carbohydrate polymers*, *151*, 600-605.
- Jones, O. G., & McClements, D. J. (2010). Biopolymer Nanoparticles from Heat-Treated
- 855 Electrostatic Protein-Polysaccharide Complexes: Factors Affecting Particle
  856 Characteristics. *Journal of food science*, *75*, N36-N43.
- Kaminski, G. A., Sierakowski, M. R., Pontarolo, R., dos Santos, L. A., & de Freitas, R. A.
  (2016). Layer-by-layer polysaccharide-coated liposomes for sustained delivery of
  epidermal growth factor. *Carbohydrate polymers*, *140*, 129-135.
- Kfoury, M., Auezova, L., Greige-Gerges, H., & Fourmentin, S. (2015). Promising
  applications of cyclodextrins in food: Improvement of essential oils retention,
  controlled release and antiradical activity. *Carbohydrate polymers*, *131*, 264-272.

863	Kfoury, M., Auezova, L., Ruellan, S., Greige-Gerges, H., & Fourmentin, S. (2015).
864	Complexation of estragole as pure compound and as main component of basil and
865	tarragon essential oils with cyclodextrins. Carbohydrate polymers, 118, 156-164.
866	Khalili, S. T., Mohsenifar, A., Beyki, M., Zhaveh, S., Rahmani-Cherati, T., Abdollahi, A.,
867	Bayat, M., & Tabatabaei, M. (2015). Encapsulation of Thyme essential oils in
868	chitosan-benzoic acid nanogel with enhanced antimicrobial activity against
869	Aspergillus flavus. LWT-Food Science and Technology, 60, 502-508.
870	Khan, F., & Ahmad, S. R. (2013). Polysaccharides and their derivatives for versatile tissue
871	engineering application. Macromolecular bioscience, 13, 395-421.
872	Kiatponglarp, W., Rugmai, S., Rolland-Sabaté, A., Buléon, A., & Tongta, S. (2016).
873	Spherulitic self-assembly of debranched starch from aqueous solution and its
874	effect on enzyme digestibility. Food Hydrocolloids, 55, 235-243.
875	Kim, D. Y., & Shin, W. S. (2015). Unique characteristics of self-assembly of bovine
876	serum albumin and fucoidan, an anionic sulfated polysaccharide, under various
877	aqueous environments. Food Hydrocolloids, 44, 471-477.
878	Kim, D. Y., & Shin, W. S. (2016). Functional improvements in bovine serum
879	albumin-fucoidan conjugate through the Maillard reaction. Food chemistry, 190,
880	974-981.
881	Kim, E. S., Lee, J. S., & Lee, H. G. (2016). Nanoencapsulation of red ginseng extracts
882	using chitosan with polyglutamic acid or fucoidan for improving antithrombotic
883	activities. Journal of agricultural and food chemistry, 64, 4765-4771.

885	octenyl-and dodecenyl-succinylated inulins. Food Hydrocolloids, 50, 145-149.
886	Lamas, A., Anton, X., Miranda, J. M., Roca-Saavedra, P., Cardelle-Cobas, A., Ibarra, I.,
887	Franco, C., & Cepeda, A. (2016). Technological strategies for the development of
888	egg-derived products with reduced content of cholesterol. Food and Bioprocess
889	<i>Technology</i> , 9, 81-90.
890	Lavoine, N., Givord, C., Tabary, N., Desloges, I., Martel, B., & Bras, J. (2014).
891	Elaboration of a new antibacterial bio-nano-material for food-packaging by
892	synergistic action of cyclodextrin and microfibrillated cellulose. Innovative Food
893	Science & Emerging Technologies, 26, 330-340.
894	Lavoine, N., Guillard, V., Desloges, I., Gontard, N., & Bras, J. (2016). Active bio-based
895	food-packaging: Diffusion and release of active substances through and from
896	cellulose nanofiber coating toward food-packaging design. Carbohydrate
897	polymers, 149, 40-50.
898	Lee, A. C., & Hong, Y. H. (2009). Coacervate formation of $\alpha$ -lactalbumin–chitosan and
899	β-lactoglobulin–chitosan complexes. <i>Food Research International</i> , 42, 733-738.
900	Li, H., & Peng, L. (2015). Antimicrobial and antioxidant surface modification of cellulose
901	fibers using layer-by-layer deposition of chitosan and lignosulfonates.
902	Carbohydrate polymers, 124, 35-42.
903	Li, K., Liu, H., Gao, W., Chen, M., Zeng, Y., Liu, J., Xu, L., & Wu, D. (2015).
904	Mulberry-like dual-drug complicated nanocarriers assembled with
905	apogossypolone amphiphilic starch micelles and doxorubicin hyaluronic acid
906	nanoparticles for tumor combination and targeted therapy. Biomaterials, 39,

907 131-144.

- 908 Li, X., Li, H., Xiao, Q., Wang, L., Wang, M., Lu, X., York, P., Shi, S., & Zhang, J. (2014).
- 909 Two-way effects of surfactants on Pickering emulsions stabilized by the 910 self-assembled microcrystals of  $\alpha$ -cyclodextrin and oil. *Physical Chemistry* 911 *Chemical Physics*, 16, 14059-14069.
- Li, X., Qin, Y., Liu, C., Jiang, S., Xiong, L., & Sun, Q. (2016). Size-controlled starch
   nanoparticles prepared by self-assembly with different green surfactant: The
- 914 effect of electrostatic repulsion or steric hindrance. *Food chemistry*, *199*, 356-363.
- Li, Y., Chen, Y., & Li, H. (2017). Recovery and purification of cholesterol from
  cholesterol-β-cyclodextrin inclusion complex using ultrasound-assisted extraction. *Ultrasonics sonochemistry*, *34*, 281-288.
- Liu, H., Cui, Y., Li, P., Zhou, Y., Chen, Y., Tang, Y., & Lu, T. (2013). Polyphosphonate
- 919 induced coacervation of chitosan: encapsulation of proteins/enzymes and their
  920 biosensing. *Analytica chimica acta*, 776, 24-30.
- 921 Liu, M., Zhang, X., Yang, B., Li, Z., Deng, F., Yang, Y., Zhang, X., & Wei, Y. (2015).
- 922 Fluorescent nanoparticles from starch: Facile preparation, tunable luminescence
  923 and bioimaging. *Carbohydrate polymers*, 121, 49-55.
- Liu, W., Liu, J., Liu, W., Li, T., & Liu, C. (2013). Improved physical and in vitro digestion
  stability of a polyelectrolyte delivery system based on layer-by-layer
  self-assembly alginate-chitosan-coated nanoliposomes. *Journal of agricultural and food chemistry*, *61*, 4133-4144.
- Liu, W., Liu, W., Ye, A., Peng, S., Wei, F., Liu, C., & Han, J. (2016). Environmental stress

929	stability of microencapsules based on liposomes decorated with chitosan and
930	sodium alginate. Food chemistry, 196, 396-404.
931	Liu, W., Tian, M., Kong, Y., Lu, J., Li, N., & Han, J. (2017). Multilayered vitamin C
932	nanoliposomes by self-assembly of alginate and chitosan: Long-term stability and
933	feasibility application in mandarin juice. LWT-Food Science and Technology, 75,
934	608-615.
935	Liu, Y., Liang, J., Wei, S., Liu, L., & Liao, M. (2015). Nanoparticles based on
936	$\beta$ -conglycinin and chitosan: Self-assembly, characterization, and drug delivery.
937	Journal of Applied Polymer Science, 132.
938	Lu, K. Y., Li, R., Hsu, C. H., Lin, C. W., Chou, S. C., Tsai, M. L., & Mi, F. L. (2017).
939	Development of a new type of multifunctional fucoidan-based nanoparticles for
940	anticancer drug delivery. Carbohydrate polymers, 165, 410-420.
941	Marcet, I., Paredes, B., & Diaz, M. (2015). Egg yolk granules as low-cholesterol replacer
942	of whole egg yolk in the preparation of gluten-free muffins. LWT-Food Science
943	and Technology, 62, 613-619.
944	Marefati, A., Sjöö, M., Timgren, A., Dejmek, P., & Rayner, M. (2015). Fabrication of
945	encapsulated oil powders from starch granule stabilized W/O/W Pickering
946	emulsions by freeze-drying. Food Hydrocolloids, 51, 261-271.
947	Marras-Marquez, T., Peña, J., & Veiga-Ochoa, M. (2015). Robust and versatile
948	pectin-based drug delivery systems. International journal of pharmaceutics, 479,
949	265-276.
950	Mauro, N., Campora, S., Scialabba, C., Adamo, G., Licciardi, M., Ghersi, G., &

- Giammona, G. (2015). Self-organized environment-sensitive inulin-doxorubicin
  conjugate with a selective cytotoxic effect towards cancer cells. *RSC Advances*, 5,
  32421-32430.
- Mensink, M. A., Frijlink, H. W., van der Voort Maarschalk, K., & Hinrichs, W. L. (2015).
  Inulin, a flexible oligosaccharide I: review of its physicochemical characteristics. *Carbohydrate polymers, 130*, 405-419.
- Muley, P., Kumar, S., El Kourati, F., Kesharwani, S. S., & Tummala, H. (2016).
  Hydrophobically modified inulin as an amphiphilic carbohydrate polymer for
  micellar delivery of paclitaxel for intravenous route. *International journal of pharmaceutics*, 500, 32-41.
- 961 Nguyen, A. T. B., Winckler, P., Loison, P., Wache, Y., & Chambin, O. (2014).
- 962 Physico-chemical state influences in vitro release profile of curcumin from pectin
  963 beads. *Colloids and Surfaces B: Biointerfaces, 121*, 290-298.
- 964 Pan, Y., Huang, X., Shi, X., Zhan, Y., Fan, G., Pan, S., Tian, J., Deng, H., & Du, Y. (2015).
- 965 Antimicrobial application of nanofibrous mats self-assembled with quaternized 966 chitosan and soy protein isolate. *Carbohydrate polymers*, *133*, 229-235.
- Peng, H., Chen, S., Luo, M., Ning, F., Zhu, X., & Xiong, H. (2016). Preparation and
  Self-Assembly Mechanism of Bovine Serum Albumin-Citrus Peel Pectin
  Conjugated Hydrogel: A Potential Delivery System for Vitamin C. *Journal of agricultural and food chemistry*, 64, 7377-7384.
- 971 Phunpee, S., Saesoo, S., Sramala, I., Jarussophon, S., Sajomsang, W., Puttipipatkhajorn,
- 972

S., Soottitantawat, A., & Ruktanonchai, U. R. (2016). A comparison of eugenol

973	and menthol or	n encapsu	lation cha	racteristics	with	water-solu	ıble	quaternized
974	$\beta$ -cyclodextrin	grafted	chitosan.	Internati	onal	journal	of	biological
975	macromolecules,	84, 472-4	180.					

- 976 Pinheiro, A. C., Bourbon, A. I., Cerqueira, M. A., Maricato, É., Nunes, C., Coimbra, M.
- A., & Vicente, A. A. (2015). Chitosan/fucoidan multilayer nanocapsules as a
  vehicle for controlled release of bioactive compounds. *Carbohydrate polymers*, *115*, 1-9.
- Preis, M., Grother, L., Axe, P., & Breitkreutz, J. (2015). In-vitro and in-vivo evaluation of
  taste-masked cetirizine hydrochloride formulated in oral lyophilisates. *International journal of pharmaceutics*, 491, 8-16.
- 983 Rezaei, A., Nasirpour, A., & Fathi, M. (2015). Application of cellulosic nanofibers in food
- 984 science using electrospinning and its potential risk. *Comprehensive Reviews in*985 *Food Science and Food Safety, 14*, 269-284.
- 986 Santiago, L. G., & Castro, G. R. (2016). Novel technologies for the encapsulation of

987 bioactive food compounds. *Current Opinion in Food Science*, 7, 78-85.

- Sathuvan, M., Thangam, R., Gajendiran, M., Vivek, R., Balasubramanian, S., Nagaraj, S.,
  Gunasekaran, P., Madhan, B., & Rengasamy, R. (2016). κ-Carrageenan: An
  effective drug carrier to deliver Curcumin in Cancer cells and to Induce Apoptosis.
- 991 *Carbohydrate polymers.*
- 992 Shah, B. R., Li, Y., Jin, W., An, Y., He, L., Li, Z., Xu, W., & Li, B. (2016). Preparation and
- 993 optimization of Pickering emulsion stabilized by chitosan-tripolyphosphate
  994 nanoparticles for curcumin encapsulation. *Food Hydrocolloids*, *52*, 369-377.

995	Shewan, H. M., & Stokes, J. R. (2013). Review of techniques to manufacture
996	micro-hydrogel particles for the food industry and their applications. Journal of
997	Food Engineering, 119, 781-792.

- 998 Shi, Y., Xu, D., Liu, M., Fu, L., Wan, Q., Mao, L., Dai, Y., Wen, Y., Zhang, X., & Wei, Y.
- 999 (2018). Room temperature preparation of fluorescent starch nanoparticles from
- starch-dopamine conjugates and their biological applications. *Materials Science and Engineering: C*, 82, 204-209.
- 1002 Shibakami, M., Tsubouchi, G., Sohma, M., & Hayashi, M. (2015). Preparation of 1003 transparent self-standing thin films made from acetylated euglenoid  $\beta$ -1, 1004 3-glucans. *Carbohydrate polymers*, *133*, 421-428.
- Silva, E. K., Zabot, G. L., Bargas, M. A., & Meireles, M. A. A. (2016).
  Microencapsulation of lipophilic bioactive compounds using prebiotic
  carbohydrates: Effect of the degree of inulin polymerization. *Carbohydrate polymers*, 152, 775-783.
- Smistad, G., Bøyum, S., Alund, S. J., Samuelsen, A. B. C., & Hiorth, M. (2012). The
  potential of pectin as a stabilizer for liposomal drug delivery systems. *Carbohydrate polymers, 90*, 1337-1344.
- Sriamornsak, P. (2011). Application of pectin in oral drug delivery. *Expert opinion on drug delivery*, 8, 1009-1023.
- 1014 Sriamornsak, P., Thirawong, N., Nunthanid, J., Puttipipatkhachorn, S., Thongborisute, J.,
- 1015 & Takeuchi, H. (2008). Atomic force microscopy imaging of novel
  1016 self-assembling pectin–liposome nanocomplexes. *Carbohydrate polymers*, 71,

1017 324-329.

- Tan, C., Feng, B., Zhang, X., Xia, W., & Xia, S. (2016). Biopolymer-coated liposomes by
  electrostatic adsorption of chitosan (chitosomes) as novel delivery systems for
  carotenoids. *Food Hydrocolloids*, 52, 774-784.
- 1021 Tang, D. W., Yu, S. H., Ho, Y. C., Huang, B. Q., Tsai, G. J., Hsieh, H. Y., Sung, H. W., &
- 1022 Mi, F. L. (2013). Characterization of tea catechins-loaded nanoparticles prepared 1023 from chitosan and an edible polypeptide. *Food Hydrocolloids*, *30*, 33-41.
- Tedeschi, C., Leuenberger, B., & Ubbink, J. (2016). Amorphous–amorphous phase
  separation in hydrophobically-modified starch–sucrose blends I. Phase behavior
- and thermodynamic characterization. *Food Hydrocolloids*, 58, 75-88.
- 1027 Thirawong, N., Thongborisute, J., Takeuchi, H., & Sriamornsak, P. (2008). Improved 1028 intestinal absorption of calcitonin by mucoadhesive delivery of novel 1029 pectin–liposome nanocomplexes. *Journal of controlled release*, *125*, 236-245.
- 1030 Tian, J., Tu, H., Shi, X., Wang, X., Deng, H., Li, B., & Du, Y. (2016). Antimicrobial
- application of nanofibrous mats self-assembled with chitosan and epigallocatechin
  gallate. *Colloids and Surfaces B: Biointerfaces, 145*, 643-652.
- 1033 Venkatesan, J., Anil, S., Kim, S.-K., & Shim, M. S. (2016). Seaweed
   1034 polysaccharide-based nanoparticles: preparation and applications for drug delivery.
   1035 *Polymers*, 8, 30.
- 1036 Ventura, I., Jammal, J., & Bianco-Peled, H. (2013). Insights into the nanostructure of
  1037 low-methoxyl pectin-calcium gels. *Carbohydrate polymers*, 97, 650-658.
- 1038 Wan, Q., Jiang, R., Guo, L., Yu, S., Liu, M., Tian, J., Liu, G., Deng, F., Zhang, X., & Wei,

- Y. (2017). Novel Strategy toward AIE-Active Fluorescent Polymeric
  Nanoparticles from Polysaccharides: Preparation and Cell Imaging. ACS *Sustainable Chemistry & Engineering, 5*, 9955-9964.
- 1042 Wan, Q., Liu, M., Xu, D., Mao, L., Tian, J., Huang, H., Gao, P., Deng, F., Zhang, X., &
- 1043 Wei, Y. (2016). Fabrication of aggregation induced emission active luminescent
- 1044 chitosan nanoparticles via a "one-pot" multicomponent reaction. *Carbohydrate* 1045 *polymers*, *152*, 189-195.
- Wang, X., Yuan, Y., & Yue, T. (2015). The application of starch-based ingredients in
  flavor encapsulation. *Starch*, 67, 225-236.
- 1048 Wen, C., Yuan, Q., Liang, H., & Vriesekoop, F. (2014). Preparation and stabilization of
- 1049 d-limonene Pickering emulsions by cellulose nanocrystals. *Carbohydrate*1050 *polymers*, 112, 695-700.
- 1051 Wen, P., Zhu, D., Feng, K., Liu, F., Lou, W., Li, N., Zong, M., & Wu, H. (2016).
- 1052 Fabrication of electrospun polylactic acid nanofilm incorporating cinnamon
- essential oil/β-cyclodextrin inclusion complex for antimicrobial packaging. *Food chemistry*, *196*, 996-1004.
- Wrigstedt, P., Kylli, P., Pitkanen, L., Nousiainen, P., Tenkanen, M., & Sipilä, J. (2010).
  Synthesis and antioxidant activity of hydroxycinnamic acid xylan esters. *Journal of agricultural and food chemistry*, 58, 6937-6943.
- 1058 Wu, L., Liao, Z., Liu, M., Yin, X., Li, X., Wang, M., Lu, X., Lv, N., Singh, V., & He, Z.
- 1059 (2016). Fabrication of non-spherical Pickering emulsion droplets by cyclodextrins
   1060 mediated molecular self-assembly. *Colloids and Surfaces A: Physicochemical and*

- 1061 *Engineering Aspects, 490, 163-172.*
- 1062 Xu, D., Liu, M., Zou, H., Huang, Q., Huang, H., Tian, J., Jiang, R., Wen, Y., Zhang, X., &
- 1063 Wei, Y. (2017). Fabrication of AIE-active fluorescent organic nanoparticles 1064 through one-pot supramolecular polymerization and their biological imaging.
- 1065 *Journal of the Taiwan Institute of Chemical Engineers*, 78, 455-461.
- 1066 Xu, H., Liu, H., & Zhang, L. (2015). Blocking and Blending: Different Assembly Models
- 1067 of Cyclodextrin and Sodium Caseinate at the Oil/Water Interface. *Langmuir*, *31*,1068 9061-9069.
- 1069 Xu, W., Jin, W., Zhang, C., Li, Z., Lin, L., Huang, Q., Ye, S., & Li, B. (2014). Curcumin
  1070 loaded and protective system based on complex of κ-carrageenan and lysozyme.
  1071 *Food Research International*, *59*, 61-66.
- Yang, J., Gao, C., Lü, S., Zhang, X., Yu, C., & Liu, M. (2014). Physicochemical
  characterization of amphiphilic nanoparticles based on the novel
  starch–deoxycholic acid conjugates and self-aggregates. *Carbohydrate polymers*, *1075 102*, 838-845.
- Yang, Y., Wang, S., Wang, Y., Wang, X., Wang, Q., & Chen, M. (2014). Advances in
  self-assembled chitosan nanomaterials for drug delivery. *Biotechnology advances*, *32*, 1301-1316.
- Zeng, G., Liu, X., Liu, M., Huang, Q., Xu, D., Wan, Q., Huang, H., Deng, F., Zhang, X.,
  & Wei, Y. (2016). Facile preparation of carbon nanotubes based carboxymethyl
  chitosan nanocomposites through combination of mussel inspired chemistry and
  Michael addition reaction: Characterization and improved Cu<sup>2+</sup> removal capability.

- 1083 *Journal of the Taiwan Institute of Chemical Engineers*, 68, 446-454.
- 1084 Zhang, T., Zhou, P., Zhan, Y., Shi, X., Lin, J., Du, Y., Li, X., & Deng, H. (2015).
- 1085 Pectin/lysozyme bilayers layer-by-layer deposited cellulose nanofibrous mats for 1086 antibacterial application. *Carbohydrate polymers*, *117*, 687-693.
- 1087 Zhang, W., Li, X., Yu, T., Yuan, L., Rao, G., Li, D., & Mu, C. (2015). Preparation,
- 1088 physicochemical characterization and release behavior of the inclusion complex
- 1089 of trans-anethole and  $\beta$ -cyclodextrin. *Food Research International*, 74, 55-62.
- 1090 Zhang, X., Zhang, Y., Zhang, H., Yang, Q., Wang, H., & Zhang, G. (2015). Preparation,
- 1091 characterization and antibacterial activity of octenyl succinic anhydride modified
  1092 inulin. *International journal of biological macromolecules*, 78, 79-86.
- 1093 Zhao, D., Wei, W., Zhu, Y., Sun, J., Hu, Q., & Liu, X. (2015). Stable Emulsions Prepared
- by Self-assembly of Hyaluronic Acid and Chitosan for Papain Loading. *Macromolecular bioscience*, 15, 558-567.
- 1096 Zimet, P., & Livney, Y. D. (2009). Beta-lactoglobulin and its nanocomplexes with pectin
- 1097 as vehicles for  $\omega$ -3 polyunsaturated fatty acids. Food Hydrocolloids, 23,
- 1098

1120-1126.

1099

- 1102 1103 1104
- 1105
- 1106
- 1107
- 1108
- 1109

1111		delivery sys	tems	
Biopolymers	Modified solvent and assembly method	Potential objects	Main results	References
	Octenyl succinic anhydride (OSA)	Highly moisture-sensitive substances	The encapsulation of bioactive compounds was achieved using a glass transition temperature of different amorphous phases in a phase separation array (hydrophobically modified starch and sucrose binary blend	(Tedeschi, Leuenberger, ۂ Ubbink, 2016)
	OSA	Shea oil with liquid and solid	system). Powder filled with shea oil was prepared via the freeze-drying method using water-in-oil-in-water (W/O/W) double emulsions (Fig. 2). The stability after freeze-drying was enhanced by OSA-modified starch Pickering emulsions. Layer-by-layer (LBL) self-assembly:	(Marefati, et al 2015)
Starch	OSA	Curcumin	ultrasonic-aided chitosan (polycation) and Na-CMC (polyanionic) materials were continuously adsorbed onto the starch surface to form a stable curcumin nanoemulsion	(Abbas, et al. 2015)
	Deoxycholic acid	Antitumor active drugs	polymer multi-shell. Novel spherical nanoparticles with amphipathic properties; the average size of the aggregates ranged from 182 to 247 nm, with a good response to pH.	(J. Yang, et al 2014)
	Poly (ethylene glycol) (mPEG) capped with a primary amino group	Adriamycin	Grafted; diselenium bonds instead of disulfur bonds were used to cross-link and synthesize new compounds, which had a better embedding load rate, stability and glutathione (GSH) response capability. This could help accelerate the release of embedded substances at the targeted site.	(Chen, Gao, L Chen, & Liu, 2016)
β-CD	Co-precipitation and freeze-drying	Trans-anethole (AT)	The obtained β-cyclodextrin solid complex had good thermal stability while controlling the slow evaporation of AT.	(W. Zhang, et a 2015)
β-CD	Co-precipitation method	Antibacterial polylactic acid (PLA) and cinnamon essential oil (CEO)	First, the CEO/ $\beta$ -CD-IC complex was prepared using co-precipitation, followed by the preparation of the PLA/CEO/ $\beta$ -CD-IC nanofilm using electrostatic spinning technology. This film was used for pork packaging, which had very good antibacterial effects against <i>E. coli</i> and <i>Staphylococcus</i> <i>aureus</i> .	(P. Wen, et al 2016)
β-CD	Non-thermal spraying - freeze-drying (SFD)	Vanillin	Preparation of microcapsule complexes using SFD exhibited better thermal stability than spray drying and freeze-drying techniques.	(Hundre, Karth & Anandharamak hnan, 2015)
β-CD	High-pressure homogenization method	Kenaf ( <i>Hibiscus</i> <i>cannabinus</i> L.) seed oil	Sodium caseinate (SC), Tween 20 (T20) and β-cyclodextrin (β-CD) were used as emulsifiers for preparing stable nano-emulsions.	(Cheong & Nyam, 2016
α-CD, β-CD	Freeze-drying Method	Trans-cinnamalde hyde against	Substances embedded in β-CD exhibited better antibacterial activity against <i>Staphylococcus aureus</i> and <i>Escherichia coli</i> than substances embedded in α-CD.	(Chun, Jo, Bjrapha, Choi, Min, 2015)
α-CD, β-CD	High-speed cutting method	Wheat germ oil (WO), olive oil, coconut oil, rice	Changing the ratio of oil to CD produced non-spherical Pickering emulsion droplets, such as disc-shaped, oval and rod-shaped	(Wu, et al., 20)

# 1110 Table 1 Recent studies regarding the use of polysaccharides matrix active compound

		bran oil, rapeseed oil and castor oil (CO)	droplets. Excessive microcrystals led to more stable non-spherical, rather than spherical, droplets.	
$\alpha$ -cyclodextrin ( $\alpha$ -CD), $\beta$ -cyclodextrin ( $\beta$ -CD), Hydroxypropyl $\beta$ -cyclodextrin (HP- $\beta$ -CD), Random methylated $\beta$ -cyclodextrin (RAMEB), Low-methylation- $\beta$ -cy clodextrin (CRYSMEB) and $\gamma$ -cyclodextrin	Freeze-drying method	Estragole (ES) and tarragon essential oils (EOs)	The obtained β-cyclodextrin solid complex was light insensitive and antioxidant active, and its release was controllable.	(Kfoury, Auezova, Ruellan, Greige-Gerges, & Fourmentin, 2015)
(γ-CD) Hydroxypropyl □ cyclodextrin (HP-β-CD)	Hydroxyalkylated cyclodextrins	Carvacrol or monoterpene alcohols	Chitosan infiltrated HP-β-CD and glycerol-encapsulated (carvacrol or monoterpene alcohols) to produce antibacterial films that acted on <i>Staphylococcus aureus</i> and <i>Escherichia coli</i> . The relative humidity of the environment controlled the rate of release of antimicrobial substances.	(Higueras, López-Carballo, Gavara, & Hernández-Muño z, 2016)
Water-soluble quaternized β-CD grafted with chitosan (QCD-g-CS)	Ionized cyclodextrin	Eugenol and (–)-menthol	Eugenol and (-)-menthol could self-assemble with QCD-g-CS; eugenol had a slower sustained release rate than (-)-menthol.	(Phunpee, et al., 2016)
Inulin	Alkenyl succinic anhydride with different olefinic lengths (C8-C18) 2-octen-1-yl-succinic anhydride and 2-dodecen-1-yl-succinic anhydride (DDSA)	Water-insoluble active compounds Medium-chain triglycerides (MCT)	Alkenyl chrysanthemums with different degrees of polymerization were synthesized by alkenyl succinic anhydride with different olefinic lengths in aqueous solution via hydrophobic modifications. The critical aggregation concentration (CAC) and hydrodynamic diameter of the derivatives were proportional and inversely proportional to the chain length, respectively. All derivatives were capable of producing micellar aggregates and oil-in-water emulsions that provided good storage stability in the presence of Tween 20. DDSA-inulin in the resulting emulsion gave a smaller droplet size than OSA-inulin and had higher storage stability. The self-assembly force was mainly achieved by the electrostatic repulsion of the carboxylate ion of the derivatives. The antibacterial activity of octenyl succinic anhydride-modified inulin (In-OSA) against <i>Staphylococcus aureus</i> and <i>Escherichia coli</i> was mainly caused by In-OSA damaging proteins and nucleic acids on the surface of	(Han, Ratcliffe, & Williams, 2015) (Kokubun, Ratcliffe, & Williams, 2015) (X. Zhang, et al., 2015)
			proteins and nucleic acids on the surface of bacteria, which resulted in the leakage of these two bacteria through the cell membrane and cell wall.	2015)
	Lauryl carbamate	Paclitaxel (PTX)	Modified inulin and PTX (amphiphilic compounds) could self-assemble to form micelles to improve their blood compatibility and stability, reducing cytotoxicity and	(Muley, Kumar, El Kourati, Kesharwani, &

	Ethylenediamine (EDA)	Doxorubicin	maintaining an effective dose of PTX for a longer period. Adriamycin and pentanoic acid inulin could assemble to form a polymerizable amphiphilic complex with pH sensitivity. The net charge of the complex changed from negative to	Tummala, 2016) (Mauro, et al.,
	and pentynoic acid (P)	(Doxo)	positive at 5.5 <ph <6.4,="" allowed<br="" which="">changes in its conformation and biological behaviors and released free doxorubicin. Inulin derivatives had better oxidation</ph>	2015)
	Amino-pyridines	No	resistance and water solubility than inulin. Their capability for cleaning was related to the position of the pyridine amino groups grafted to the inulin derivatives.	(Y. Hu, et al., 2014)
		D-Limonene	Ultrasonic homogenization was used to form a	(C. Wen, Yuan,
	Ammonium persulfate	(4-isopropenyl-1- methylcyclohexen	highly stable Pickering emulsion consisting of nano-microcrystalline cellulose and	Liang, &
		e)	D-limonene.	Vriesekoop, 2014)
	Cold phosphoric acid	Soybean oil	Sodium caseinate together with regenerated cellulose nanoparticles functioned as emulsifiers to stabilize emulsions. Proteins combined with polysaccharides could reduce interfacial tension and enhance the thickness of the adsorbed layer of the surrounding fat globules to improve the stability of the emulsion system. In O/W emulsions, regenerated cellulose could improve the adsorption of the droplet onto the surface and reduce the interactions among droplets. The effect was better than microcrystalline	(H. Hu, et al., 2016)
Cellulose	No	Lysozyme (LZ) and pectin	cellulose. Electrospinning allowed positively charged lysozyme and negatively charged pectin to be alternately deposited onto the surface of the cellulose nanofiber pad via layer-by-layer self-assembly. This fiber (the outermost layer is lysozyme) had a strong inhibitory effect on <i>Escherichia coli</i> and <i>Staphylococcus aureus</i> . Chitosan and lignosulfonate were superimposed on the surface of the fiber via	(T. Zhang, et al., 2015)
	No	Chitosan (CS) and lignin sulfonate (LS)	layer-by-layer self-assembly. The anticoagulant activity of <i>Escherichia coli</i> was best when the outermost layer was chitosan, and the antioxidant activity was also improved.	(H. Li & Peng, 2015)
	No	Caffeine and chlorhexidine digluconate (CHX)	Caffeine and chlorhexidine digluconate underwent controlled release in the carrier of cellulose nanofibers (CNFs). Interactions among CNF nanoporous networks, embedded molecules and nanofibers slowed the release of molecules.	(Lavoine, Guillard, Desloges, Gontard, & Bras, 2016)
Pectin	Natural agitation or freeze-drying	Tolbutamide	Pluronic, Tween and sodium lauryl sulfate were used as surfactants to control the dispersibility and pH release properties of pectin hydrogels in water, and the release properties were good. Two different strategies were developed: blending with agarose (natural) and freeze-drying. Agarose was added to increase the embedding system of the	(Marras-Marquez, Peña, & Veiga-Ochoa, 2015)

Agitation and mixing method	Liposome	<ul> <li>gel network. Freeze-drying produced a porous structure that accelerated the immersion of water, and the freshly prepared blend had a slower release rate.</li> <li>Among LM, HM and AM pectin, LM and HM pectin were the best choices for stabilizing liposomal systems.</li> <li>HM pectin, due to having the greatest hydrophobicity, lowest charge density and more branched structure, could encapsulate liposomes in the form of long tails and/or rings to form a larger adsorbent layer. The interaction between liposomes coated with HM pectin and mucin was the most pronounced, which could improve the adhesion to intestinal mucosa.</li> <li>Pectin-liposomal nanocomposites (PLNS)</li> </ul>	(Smistad, Bøyum, Alund, Samuelsen, & Hiorth, 2012)
Co-precipitation method	Calcitonin (eCT)	<ul> <li>could increase calcitonin absorption in the intestine (ECT). Highly esterified PLNS with a highly negative charge ratio exhibited lower DE. Low-DE pectin PLNS showed strong mucoadhesive properties. Low-DE pectin PLNS remained at the site of mucosal adhesion after 6 hours of administration.</li> <li>Pectin adhered to the mucus layer and to the intestinal mucosa to extend calcitonin retention at the mucosa, especially in the duodenum and jejunum.</li> <li>Curcumin encapsulated by low-ester pectin</li> </ul>	(Thirawong, Thongborisute, Takeuchi, & Sriamornsak, 2008)
Ionic gelation method	Curcumin	beads prepared by SOLUTOL was rapidly released in an amorphous form. Curcumin encapsulated by low-ester pectin beads prepared by sodium caseinate could not be dissolved during the encapsulation process, which was related to its casein micelles. Transcrackers prepared by Transcutol exhibited pH-dependent release in simulated gastric fluid, simulating slower intestinal fluid.	(Nguyen, Winckler, Loison, Wache, & Chambin, 2014)
Granulation method and an enteric coating	Piroxicam	Zinc-pectin/alginate beads prepared by the Piroxicam loading delivery system could	(Auriemma, et al., 2013)
Polyelectrolyte self-assembly method	The functional agent vitamin C (Vc)	extend the drug release time. Bovine serum albumin (BSA) and citrus pectin could self-assemble to form a pH-responsive natural hydrogel (BSA-Pectin Gel, BPH). This hydrogel could carry Vc via electrostatic and covalent interactions between hydrophobic groups of BAS and amide groups of pectin.	(Peng, et al., 2016)
Ionic gelation method	Nisin	Chitosan nano-capsules loaded with nisin and its antibacterial activity in tomato juice was studied. Through the observation of bacterial activity in tomato juice for 6 months, it was found that chitosan nano-capsules loaded with nisin could effectively inhibit the biological activity of <i>Pseudomonas aeruginosa</i> and, to a	(Chopra, Kaur, Bernela, & Thakur, 2014)
Thin-layer dispersion method combined with micro-fluidization	Vitamin C	certain extent, extend food's shelf life. Positive chitosan (CH) and negative sodium alginate (AL) were added to the surface of anionic nanoliposomes (NLs). After 90 days of storage, this self-assembled compound	(W. Liu, et al., 2017)

Chitosan

		could delay lipid peroxidation and vitamin C release at 4°C.	
High-pressure	Papain	A stable complex was made through electrostatic interactions among hyaluronic acid (HA)/chitosan (CS) and papain, which	(Zhao, et al.,
homogenization method	rupum	have been shown to be a suitable enzyme delivery system in emulsions.	2015)
		5-fluorouracil (5-FU) nanoparticles were provided based on β-glycinin (7S) and	
	5-fluorouracil	chitosan (CS). Nanoparticles were mainly formed by electrostatic interactions between amine groups (interacting NH <sup>3+</sup> ) of CS and 7S	(Y. Liu, Liang,
Freeze-drying method	(5-FU)	carboxyl groups (-COO- and intermolecular hydrogen bonds). 5-FU release was	Wei, Liu, & Liao, 2015)
		pH-dependent, which is in accordance with the Fick diffusivity of the Ritger-Peppas	,
		model. CS and catechin (EGCG) double-layer composites or CS-rectorite (REC) composites	
Layer-by-layer		(CS-REC) and EGCG double-layer composites can be self-assembled through	(Tian, et al.,
self-assembly using freeze-drying method	Epigallocatechin gallate (EGCG)	layer-by-layer (LBL) self-assembly manufacturing technology. CS and EGCG can	(11ail, et al., 2016)
neeze-urying memou		increase the contact time on the surface of	,
		adsorbent materials to inhibit <i>Staphylococcus aureus</i> and extend the EGCG release time.	
		Nanoparticles were assembled by pH-responsive chitosan and poly (γ-glutamic	
Polyelectrolyte	Tea catechins	acid) (γ-PGA) for delivery of tea catechins. These nanoparticles improved the delivery	(Tang, et al.,
self-assembly method		capacity of EGCG in the cell monolayer due to their ability to tightly connect to Caco-2	2013)
		cells. Positively charged N-[(2-hydroxy-3-	
		trimethylammonium) propyl] chitosan chloride (HTCC) and negatively charged soy	
		protein isolate (SPI) were self-assembled via a layer-by-layer mechanism on cellulose acetate	
Electrofiber technology	НТСС	(CA). The antibacterial effect of nanofiber	(Pan, et al., 2015)
		mats was positively correlated with the number of double-layers on CA, which had a	
		good antibacterial effect against <i>Escherichia</i> coli and <i>Staphylococcus aureus</i> (mainly by	
		HTCC). Amphiphilic chitosan-graft-poly (lactide)	
Micro-phase separation		(CS-g-PLA) copolymer was synthesized via homogeneous ring-opening polymerization	
method	β-Carotene (β-C)	(ROP) in ionic liquid. This significantly improved the stability of $\beta$ -carotene and	(Ge, et al., 2015)
		antioxidant activity. Chitosan was adsorbed onto the surface of	
$\mathbf{Y}$		liposomes by electrostatic and hydrophobic forces, maintaining the spherical shape,	
	Carotenoids,	inducing a charge reversal, enhancing the orderly distribution of the polysaccharides in	(Tan, Feng,
Thin-film evaporation	lycopene,	the polar region and the hydrophobic core of	Zhang, Xia, &
method	$\beta$ -carotene, lutein and canthaxanthin	lipid, and limiting the free movement of lipid molecules. This enhanced the stability of	Xia, 2016)
		carotenoids under heating, gastrointestinal	
		stress and centrifugal sedimentation	

	Freeze-drying method	<i>Mentha piperita</i> essential oils	Chitosan-cinnamic acid could embed <i>Mentha</i> <i>piperita</i> essential oil to enhance its stability and antimicrobial activity against <i>Aspergillus</i> <i>flavus</i> . Moreover, 1000 ppm nano-gel on the surface of fruit could inhibit the growth of bacteria on the tomato surface.	(Beyki, et al., 2014)
	Ultrasonic method	Thyme essential oils	Chitosan and benzoic acid nanogel (CS-BA) could embed thyme essential oil to enhance its stability and antimicrobial activity against <i>Aspergillus flavus</i> . Moreover, 700 mg/L nano-gel on the surface of fruit could inhibit	(Khalili, et al., 2015)
	High-speed cutting method	Curcumin and Medium-chain triglyceride (MCT)	bacterial growth on the tomato surface. Curcumin was successfully encapsulated in CS-TPP nanoparticles and had good stability and sustained release in the Pickering emulsion system over a long period of time.	(Shah, et al., 2016)
	Ionic gelation method	Red ginseng extract (RG)	Fucoidan itself had anticoagulant activity, and a nanoparticle in the form of an ionic gel composed of fucoidan and chitosan helped enhance the anti-thrombotic activity of red ginseng extract in mice while improving the poor bioavailability of oral ginsenosides.	(E. S. Kim, Lee, & Lee, 2016)
Fucoidan	Ion cross-linking method and freeze-drying method	Curcumin	O-carboxymethyl chitosan (O-CMC) and fucoidan formed a pH-sensitive nanoparticle, which then encapsulated curcumin. This nanoparticle had a 92.8% encapsulation degree, and the complex was stable under acidic conditions. Under alkaline conditions, the complex could reduce cytotoxicity after release and enhanced the uptake capability of	(YC. Huang & Kuo, 2016)
	Agitation and mixing method	Doxorubicin	cells. Nanoparticles formed by fucoidan and a cationic polypeptide (protamine) could be released by enzymatic digestion in an acidic intracellular microenvironment (pH 4.5-5.5). P-selectin-induced endocytosis could improve a metastatic breast cancer cell line system.	(Lu, et al., 2017)
	Ultrasonic method	Poly-1-lysine	Hollow multilayered nano-capsules were formed by chitosan and fucoidan in a layer-by-layer manner. The encapsulated poly-l-lysine was released in accordance with Fick's diffusion law at pH 7.	(Pinheiro, et al., 2015)
	Solvent evaporation method	Curcumin	The cumulative release of curcumin in a <i>K</i> -carrageenan vector was 78% at pH 5.0, which thereby induced a decline in the mitochondrial membrane potential of cancer cells and eventually resulted in apoptosis.	(Sathuvan, et al., 2016)
к-Carrageenan	Agitation and mixing method	Curcumin	K-carrageenan and lysozyme self-assembled to form a micro-complex and embed curcumin. The solubility of the complex increased significantly, and the stability increased by approximately 2.7-fold, 2-fold and 1.7-fold compared to un-self-assembled curcumin, respectively, which could maintain its biological activity after Pasteurization and ultraviolet radiation.	(W. Xu, et al., 2014)
Alginate	Agitation and mixing method	Curcumin	The prepared alginate-curcumin (Alg-Ccm) conjugate had great solubility and stabilization in water at pH 7.4, and the cytotoxicity remained.	(Dey & Sreenivasan, 2014)
	Agitation and mixing	β-lactamase	Sodium alginate (Alg) and lysozyme (Lyz)	(Fuenzalida, et
	-			

		AC	CCEPTED MA	NUSCRIPT	
		method	(BLA)	formed a nanopolymer electrolyte complex, and the addition of Alg molecular weight or Ca <sup>2+</sup> resulted in higher cross-linking with Lyz, the associated second enzyme BLA. The activity was related to the ionic strength of the solution. A polyelectrolyte delivery system (PDS) based	al., 2016)
		Dynamic high-pressure microfluidization (DHPM)	Nanoliposomes	on sodium alginate (AL) and chitosan (CH) coated on the surface of nanoliposomes (NLs) was prepared, which could resist lipolytic degradation under simulated gastrointestinal conditions.	(W. Liu, et al., 2013)
		Ultrasonic method	α-amylase	Superparamagnetic carboxymethyl chitosan/sodium alginate nanosphere (SM/CMC/SA) could fix α-amylase and increase enzymatic activity by 4.67-fold as well as its stability against acid, alkali, heat and storage conditions.	(J. Jiang, Chen, Wang, Cui, & Wan, 2016)
Sodiu	um alginate	Agitation and mixing method	Vitamin C	A polymer of liposomes (LPs), chitosan (CH) and sodium alginate (AL) had the ability to reduce the release of vitamin C from simulated intestinal fluid. The release rate was highest after mixing with pancreatic and bile salts.	(W. Liu, et al., 2016)
		Agitation and mixing method	Potential lipophilic agents	Sodium alginate and whey protein isolate (WPI) self-assembled to form a soluble biopolymer complex. At high temperature, more hydrophobic groups were exposed on the surface of the whey protein isolate, which benefited the interactions between the complex and potential lipophilic bioactive agents.	(Fioramonti, Perez, Aringoli, Rubiolo, & Santiago, 2014)
1	112			ugents.	
1	113				
1	114				
1	115				
1	116				
1	117				
1	118				
1	119				
1	120				
1	121				
1	122				
1	123				
1	124				
1	125	X '			
1	126	7			
1	127				
1	128				
1	129				
1	130				
1	131				
1	132				
1	133				

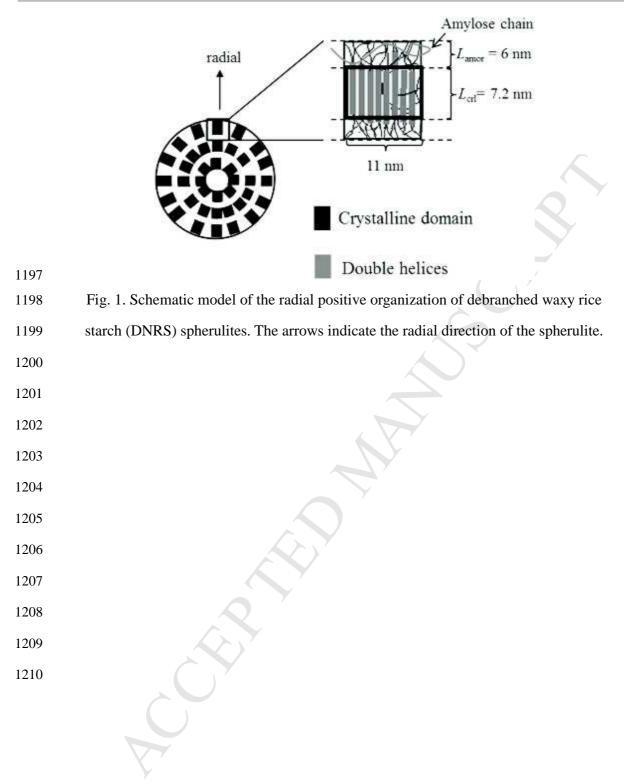
### 1134 List of Figures:

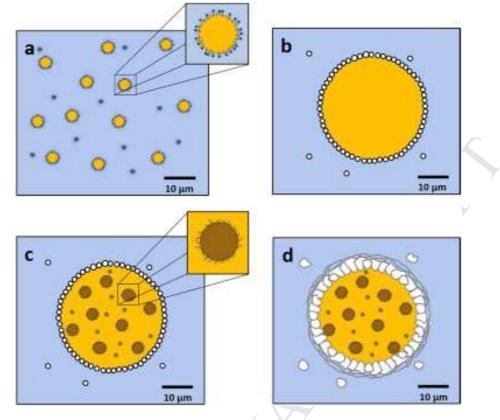
- 1135 Fig. 1. Schematic model of the radial positive organization of debranched waxy rice
- starch (DNRS) spherulites. The arrows indicate the radial direction of the spherulite.
- 1137 Fig. 2. Schematic representation of different types of emulsions: (a) surfactant-stabilized
- 1138 emulsion, (b). particle-stabilized emulsion, (c) starch granule-stabilized double emulsion

and (d) heat-treated starch granule-stabilized double emulsion.

- 1140 Fig. 3. Schematic representation of the assembly model for the  $\beta$ -CD (used as solid
- 1141 colloidal particles)/SC (sodium caseinate, used as an emulsifier)/TG (triglyceride) system
- 1142 at the oil/water interface (H. Xu, et al., 2015)
- Fig. 4. Schematic representation of inulin particle formation: (a) inulin chains withrandom coils; (b) formation of glucose–fructose links; (c) antiparallel arrangement of
- 1145 inulin helices in ribbons (arrow indicates the long axis of the ribbon); (d) inulin ribbons
- 1146 combined, likely through spherulite
- 1147 Fig. 5. Schematic representation of (a) calcium binding to polygalacturonate sequences of
- 1148 LM pectin ('egg box' dimer and 'egg-box' cavity) and (b) a model for the gelation of
- amidated LM pectin; (c) topographical (left) and equivalent processed (right) images from
- atomic force microscopy (AFM) of pectin–liposome nanocomplexes (PLNs) using pectins;
- (d) different proportions of liposomes and pectin interactions; (e) calcium–pectin gel with
- 1152 low calcium concentration; (f) calcium–pectin gel with high calcium concentration.
- 1153 Fig. 6. Schematic illustration of the synthetic process and mechanism of EDTMP-Hb-
- 1154 chitosan-MWCNT composites. Key points: (i) The interaction between chitosan and
- 1155 MWCNTs results in the dispersion of MWCNTs. (ii) The strong electrostatic and/or

1156	hydrogen bonding interactions between EDTMP and chitosan result in the coacervation of
1157	chitosan. (iii) The coacervation of chitosan results in the in situ encapsulation of Hb. (iv)
1158	The interaction between EDTMP and Hb facilitates the encapsulation of Hb.
1159	
1160	
1161	
1162 1163	
1164	
1165	
1166	
1167	
1168	
1169	
1170	
1171 1172	
1172	
1174	
1175	
1176	
1177	
1178	
1179 1180	
1181	
1182	
1183	
1184	
1185	
1186 1187	
1187 1188	
1189	
1190	
1191	
1192	
1193	
1194	
1195 1196	
1190	

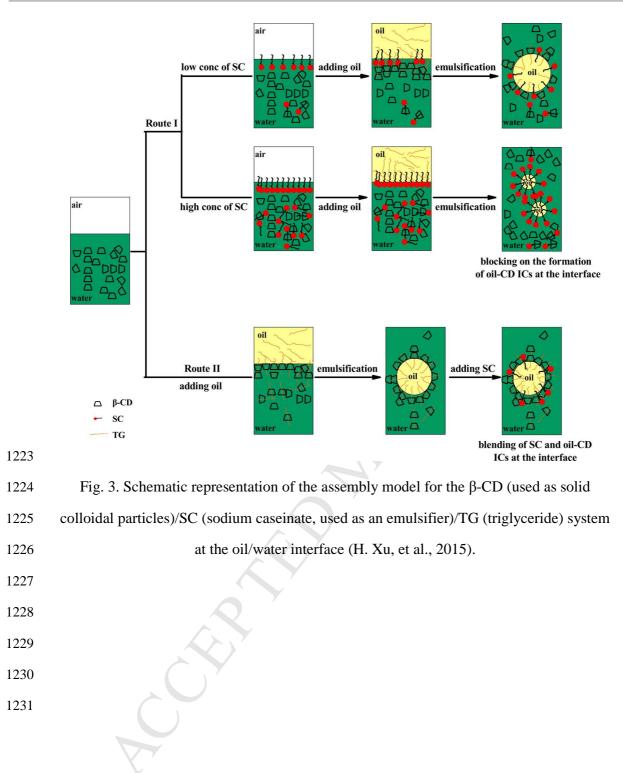




1212 Fig. 2. Schematic representation of different types of emulsions: (a) surfactant-stabilized

1213 emulsion, (b) particle-stabilized emulsion, (c) starch granule-stabilized double emulsion

and (d) heat-treated starch granule-stabilized double emulsion.



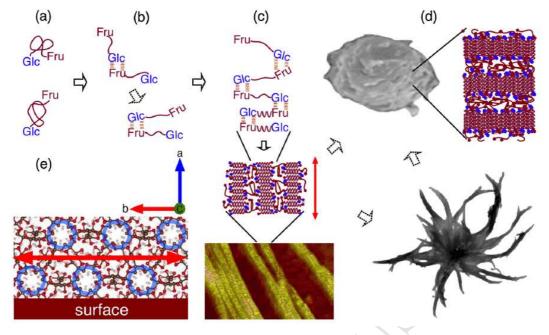


Fig. 4. Schematic representation of inulin particle formation: (a) inulin chains with
random coils; (b) formation of glucose–fructose links; (c) antiparallel arrangement of
inulin helices in ribbons (arrow indicates the long axis of the ribbon); (d) inulin ribbons
combined, likely through spherulite intermediates, forming semicrystalline particles; (e)
inulin ribbons lying flat on a surface (rings accentuate inulin helices).

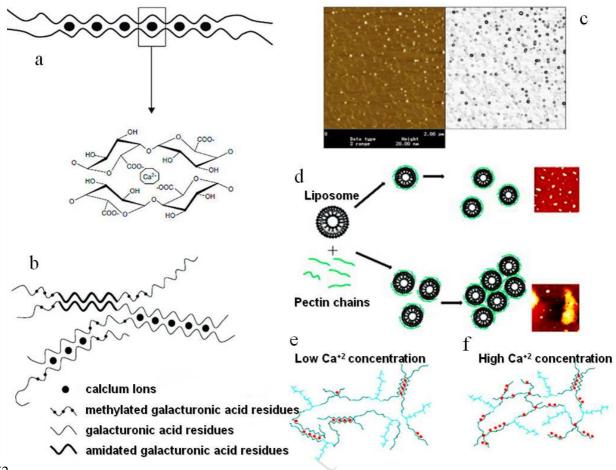


Fig. 5. Schematic representation of (a) calcium binding to polygalacturonate sequences of
LM pectin ('egg box' dimer and 'egg-box' cavity) and (b) a model for the gelation of
amidated LM pectin; (c) topographical (left) and equivalent processed (right) images from
atomic force microscopy (AFM) of pectin–liposome nanocomplexes (PLNs) using pectins;
(d) different proportions of liposomes and pectin interactions; (e) calcium–pectin gel with
low calcium concentration; (f) calcium–pectin gel with high calcium concentration.

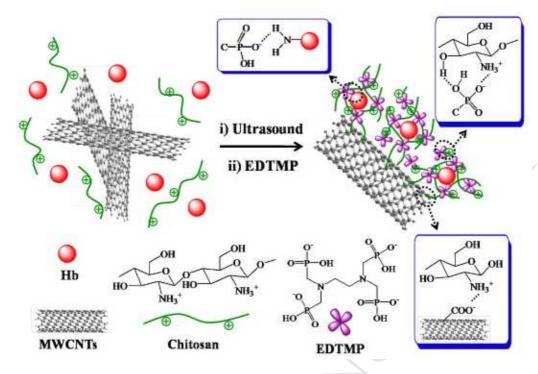


Fig. 6. Schematic illustration of the synthetic process and mechanism of EDTMP-Hb-chitosan-MWCNT composites. Key points: (i) The interaction between chitosan and MWCNTs results in the dispersion of MWCNTs. (ii) The strong electrostatic and/or hydrogen bonding interactions between EDTMP and chitosan result in the coacervation of chitosan. (iii) The coacervation of chitosan results in the in situ encapsulation of Hb. (iv) The interaction between EDTMP and Hb facilitates the encapsulation of Hb. 

		ACCEPTED MANUSCRIPT
1294		Highlights
1295	•	The activity of bioactive compounds is limited due to their instability.
1296	•	An overview of delivery systems based on various carbohydrates is provided.
1297	•	The assembling food ingredients using carbohydrate biopolymers is reviewed.
1298	•	The protective effects of the corresponding delivery systems are highlighted.

<text><text><text>

### Highlights

- 2 The activity of bioactive compounds is limited due to their instability.
- 3 An overview of delivery systems based on various carbohydrates is provided.
- 4 The assembling food ingredients using carbohydrate biopolymers is reviewed.
- 5 The protective effects of the corresponding delivery systems are highlighted.
- 6