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DOI: 10.20933/100001167

Publication date: 2020

Document Version Other version

Link to publication in Discovery Research Portal

Citation for published version (APA): Schor, A., Kankova , K., Woolston, A-M., Vojtesek, B., Felts, P., Norman, D., & Harada, K. (2020, Sep). Characterisation of antibodies to Migration Stimulating Factor (MSF). Detection of MSF isoforms. University of Dundee. https://doi.org/10.20933/100001167

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DRAFT. REPORT TO BE COMPLETED

Title: Characterisation of antibodies to Migration Stimulating Factor (MSF). Detection of MSF isoforms.

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https://doi.org/10.20933/100001167

Time period during which data was collected: 1994-2019

<u>Abstract</u>

Migration Stimulating Factor (MSF) is a 70kDa truncated isoform of fibronectin (FN). Unlike FN, MSF is not a matrix molecule but a soluble factor which exhibits a range of potent cytokine-like bioactivities not displayed by full-length FN. Two isoforms of human MSF (MSF+aa and MSF-aa), as well as murine Migration Stimulating Factor (mMSF) have been cloned. In this communication we report the characterisation of various polyclonal and monoclonal antibodies to human and murine MSF. In particular:

- (i) Specific MSF-identification antibodies that recognise both MSF+aa and MSF-aa;
- (ii) Specific mMSF-identification antibodies;
- (iii) Identification antibodies that recognise MSF-aa but not MSF+aa;
- (iv) MSF-function-neutralising antibodies that recognise MSF+aa, MSF-aa, mMSF and the gelatin-binding domain of FN/MSF (Gel-BD) but not full-length FN.

Description of the antigens:

Fibronectin (FN) is a modular glycoprotein consisting of the following functional domains: Hep 1/Fib-1 (N-terminal low affinity binding to heparin and fibrin), Gel-BD (binding to gelatin/collagen), Cell-BD (RGD-mediated binding to integrins), Hep-2 (high affinity heparin binding) and Fib-2 (C-terminal fibrin binding site). Each functional domain is composed of a different number of three possible homology modules, called type I, II and III (Fig 1). Human Migration Stimulating Factor (referred to as MSF) is a 70kDa truncated isoform of FN. MSF RNA is generated from the fibronectin gene by a two-stage processing mechanism. In the first stage, an MSF-specific primary transcript is generated from the fibronectin gene by read-through of intron 12, separating exons III-1a and III-1b. This is followed by intraintronic cleavage to produce a 5.9 kb MSF pre-message that remains sequestered within the nucleus, where it is rapidly degraded. In cells that express MSF protein a second stage takes place, whereby the intron-derived 3' UTR of the pre-message is cleaved a second time to produce a 2.1 kb mature MSF message. This has a shorter (195bp) intron-derived 3' sequence containing a 30bp in-frame coding sequence (immediately contiguous with exon III-1a), followed by a 165bp 3'-UTR containing several in-frame stop codons and a cleavage/polyadenylation signal. The mature message is rapidly exported to the cytoplasm for translation [Schor et al, 2003; Kay et al, 2005]. Therefore, MSF is identical to the N terminus of full-length fibronectin, up to and including the amino acid sequence coded by exon III-1a, with the addition of an MSF-unique (intron-coded) 10 amino acid C-terminus: VSIPPRNLGY [Schor et al, 2003; Kay et al, 2005] (Fig 1).

FN is a major component of the extracellular matrix. Unlike FN, MSF is a monomer and is not a matrix molecule but a soluble factor which exhibits a range of cytokine-like bioactivities, including the stimulation of cell migration and angiogenesis. The motogenic activity of MSF is mediated by the IGD motifs, present in modules I3, I5, I7 and I9 of the Gel-BD domain and, in some cases, by the HEEGH motif present in module I8 [Schor et al, 1999, 2003; Houard et al, 2005; Millard et al, 2007] (**Fig 1, Fig 2**). The bioactivities of MSF are not expressed by full-length fibronectin, due to steric hindrance [Millard et al, 2007; Vakonakis et al 2009].

Two isoforms of MSF (referred to as MSF) have been cloned. Both contain the same unique 10 amino acid C-terminus as well as same bioactive IGD and HEEGH amino acid motifs. The two isoforms differ solely in terms of a 45bp deletion in exon II-1 and are consequently referred to as MSF+aa and MSF-aa to indicate the retention or deletion of a 15 amino acid

sequence in module II-1 (**Fig 1** and **Fig 2**). The term MSF or total MSF will be employed to denote both isoforms.

We have isolated and cloned a murine MSF (mMSF) transcript by PCR. This is homologous to its human counterpart, consisting of the 5-terminus of mouse FN, up to and including exon III-1a, and terminating in a unique 3'coding sequence derived from the intron separating exons III-1a and -1b. The 3'UTR ends in a polyA tail. mMSF protein consequently has a molecular mass of 70kDa and terminates in a unique 12 amino acid C-terminus: VSNSSAALDSDP (**Fig 3**). The murine FN coding sequence is over 90% homologous to human FN; the four IGD motifs and the HEEGH motif are similarly located in modules I3, I5, I7, I9 and I8, respectively. However, there is no significant homology between the human and mouse intron-derived C-terminal MSF-unique peptides.

Eukaryotic and prokaryotic recombinant MSF and mMSF were produced as described [Schor et al 2003].

We have raised polyclonal (Pab) and monoclonal (Mab) antibodies to human and murine MSF. The peptides used as antigens to raise these antibodies and an overview of the results obtained are shown in **Table 1**. The antibodies were characterised by ELISA, immunoblotting, IHC and their ability to abrogate or remove MSF/mMSF bioactivity [Schor et al 2003, 2012]. As expected, different antibodies were useful for certain techniques and not for others.

Production of antibodies

To be completed

Characterisation of VSI antibodies

To be completed

<u>Conclusions</u>: VSI are MSF-specific identification antibodies that recognise the unique 10-mer C-terminal sequence of MSF+aa and MSF-aa; that is, total MSF.

Characterisation of TYN antibodies

To be completed

Conclusions: TYN are identification antibodies that recognise MSF-aa but not MSF+aa.

Characterisation of VSN antibodies

To be completed

<u>Conclusions</u>: VSN are mMSF-specific identification antibodies that recognise the unique 12mer C-terminal sequence of mMSF.

Characterisation of pepQ antibodies

To be completed

<u>Conclusions:</u> pepQ are MSF-function-neutralising antibodies that recognise MSF+aa, MSF-aa, mMSF and Gel-BD but not FN.

Methods

To be completed

- 1. ELISA
- 2. Dot Blots
- 3. Western Blots
- 3. Immunohistochemistry (IHC)

4. Cell migration

5. Cell proliferation

ACKNOWLEDGEMENTS

This work has been funded by Cancer Research UK, Breast Cancer Campaign, Engineering and Physical Sciences Research Council, Biotechnology and Biological Sciences Research Council, Scottish Chief Scientist Office and Medical Research Council. We thank the many collaborators that have contributed to this work by providing specimens and experimental data.

Table 1. Overview of the antibodies raised and results obtained.

The peptides indicated were used as antigens to raise monoclonal (Mab) and polyclonal (Pab) antibodies.

Peptide used as antigen	Ab code	Reactivity of Abs
VSIPPRNLGY	VSI	Mab and Pab recognise MSF+aa
10 mer, MSF-unique C-terminus		and MSF-aa.
		Do not recognise FN, Gel-BD or
		Hep 1/Fib-1 domains.
TYNDRTDSTTSNY	TYN	Mab and Pab recognise MSF-aa.
13 mer, present in MSF-aa, II-1. In MSF+aa these		Do not recognise MSF+aa, FN or
amino acids are adjacent to the sequence deleted in		Gel-BD
MSF-aa (6 before and 7 after)		
VSNSSAALDSDP	VSN	Pab recognise mMSF. Do not
12 mer, mMSF-unique C-terminus		recognise MSF, FN or Gel-BD
TNEGVMYRIGDQWDKQHDMGH	pepQ	Mab recognise MSF+aa, MSF-aa
21-mer, IGD-containing peptide in module I-7		and Gel-BD.
		Do not recognise FN.



Figure 1. The structure of fibronectin and MSF. Fibronectin domains include: Hep 1/Fib-1 (N-terminal low affinity binding to heparin and fibrin), Gel-BD (binding to gelatin/collagen), Cell-BD (RGD-mediated binding to integrins), Hep-2 (high affinity heparin binding) and Fib-2 (C-terminal fibrin binding site). Each domain is composed of three possible homology modules, called type I, II and III. MSF is identical to the N terminus of fibronectin, up to and including the amino acid sequence coded by exon III-1a, with the addition of an MSF-unique (intron-coded) 10 amino acid C-terminus. Two isoforms of MSF have been cloned. These differ solely in terms of a 45bp deletion in exon II-1 and are consequently referred to as MSF+aa and MSF-aa to indicate the retention or deletion of a 15 amino acid sequence in module II-1. The location of IGD motifs (\downarrow) and HEEGH motif (*) is indicated.

MSF+aa. Accession number AJ535086

	<u>http://wv</u>	<u>vw.ncbi.nlm.n</u>	<u>ih.gov/entrez/</u>	<u>/viewer.fcgi?d</u>	<u>b=Protein&id</u>	<u>=27227743</u>
1	MLRGPGPGLL	LLAVQCLGTA	VPSTGASKSK	RQAQQMVQPQ	SPVAVSQSKP	GCYDNGKHYQ
61	INQQWERTYL	GNALVCTCYG	GSRGFNCESK	PEAEETCFDK	YTGNTYRVGD	TYERPKDSMI
121	WDCTCIGAGR	GRISCTIANR	CHEGGQSYKI	GD TWRRPHET	GGYMLECVCL	GNGKGEWTCK
181	PIAEKCFDHA	AGTSYVVGET	WEKPYQGWMM	VDCTCLGEGS	GRITCTSRNR	CNDQDTRTSY
241	R <mark>IGD</mark> TWRKKD	NRGNLLQCIC	TGNGRGEWKC	ERHTSVQTTS	SGSGPFTDVR	AAVYQPQPHP
301	QPPPYGHCVT	DSGVVYSVGM	QWLKTQGNKQ	MLCTCLGNGV	SCQETAVTQT	YGGNSNGEPC
361	VLPFTYNGRT	FYSCTTEGRQ	DGHLW CSTTS	NYEQDQKYSF	CTDHTVLVQT	RGGNSNGALC
421	HFPFLYNNHN	YTDCTSEGRR	DNMKWCGTTQ	NYDADQKFGF	CPMAAHEEIC	TTNEGVMYRI
481	GD QWDKQHDM	GHMMRCTCVG	NGRGEWTCIA	YSQLRDQCIV	DDITYNVNDT	FHKRHEEGHM
541	LNCTCFGQGR	GRWKCDPVDQ	CQDSETGTFY	Q IGD SWEKYV	HGVRYQCYCY	GRGIGEWHCQ
601	PLQTYPSSSG	PVEVFITETP	SQPNSHPIQW	NAPQPSHISK	YILRWRP <mark>VSI</mark>	PPRNLGY

MSF-aa: Accession number AJ276395 (15 amino acid deletion in module II-1) <u>http://www.ncbi.nlm.nih.gov/entrez/viewer.fcgi?db=Protein&id=12053817</u>

1	MLRGPGPGLL	LLAVQCLGTA	VPST GASKSK	RQAQQMVQPQ	SPVAVSQSKP	GCYDNGKHYQ
61	INQQWERTYL	GNALVCTCYG	GSRGFNCESK	PEAEETCFDK	YTGNTYRVGD	TYERPKDSMI
121	WDCTCIGAGR	GRISCTIANR	CHEGGQSYKI	GD TWRRPHET	GGYMLECVCL	GNGKGEWTCK
181	PIAEKCFDHA	AGTSYVVGET	WEKPYQGWMM	VDCTCLGEGS	GRITCTSRNR	CNDQDTRTSY
241	R <mark>IGD</mark> TWSKKD	NRGNLLQCIC	TGNGRGEWKC	ERHTSVQTTS	SGSGPFTDVR	AAVYQPQPHP
301	QPPPYGHCVT	DSGVVYSVGM	QWLKTQGNKQ	MLCTCLGNGV	SCQETAVTQT	YGGNSNGEPC
361	VLPFTYNDRT	DSTTSNYEQD	QKYSFCTDHT	VLVQTRGGNS	NGALCHFPFL	YNNHNYTDCT
421	SEGRRDNMKW	CGTTQNYDAD	QKFGFCPMAA	HEEICTTNEG	VMYR <mark>IGD</mark> QWD	KQHDMGHMMR
481	CTCVGNGRGE	WTCIAYSQLR	DQCIVDDITY	NVNDTFHKRH	EEGH MLNCTC	FGQGRGRWKC
541	DPVDQCQDSE	TGTFYQ <mark>IGD</mark> S	WEKYVHGVRY	QCYCYGRGIG	EWHCQPLQTY	PSSSGPVEVF
601	ITETPSQPNS	HPIQWNAPQP	SHISKYILRW	RP <mark>VSIPPRNL</mark>	GY	

Figure 2. MSF+aa and MSF-aa protein sequences. The sequences highlighted are: MSFunique decamer in red (sequence MSF does not share with FN). MSF 15 amino acid region present in MSF+aa and absent in MSF-aa in blue. IGD and HEEGH motifs in purple.

L G Ρ G Ρ G R L L T. 77 C Τ. Т 1 Μ Т R Α T. G 1 ATGCTCAGGGGTCCGGGACCCGGGCGGCTGCTGCTGCTGGCAGTCCTGTGCCTGGGGACC 21 61 К S V R C т E Α G K S R 0 Α 0 0 Т ٦7 P \cap TCGGTGCGCTGCACCGAAGCCGGGAAGAGCAAGAGGCÅGGCTCÅGCÅAATCGTGCÅGCCT F 41 V V S D S Ρ Α 0 S Κ Ρ G С Ν G Κ Η Y 0 121 CÂATCCCCGGTGGCTGTCAGTCÂGAGCAAGCCTGGCTGTTTTGACAATGGGAAGCACTAT 61 Ν W Т Y G N v 0 0 Ε R T. Α C Т Y Τ. 181 CÂGATAAATCÂGCÂGTGGGAACGGACCTACCTAGGCAACGCCCTGGTTTGTACCTGCTAT 81 G S R G F Ν С Ε S Κ Ρ Ε Ρ Ε Е F D Т C G 241 GGAGGAAGCCGGGGTTTTAACTGCGAGAGCAAGCCTGAGCCTGAAGAGACTTGCTTTGAC 101 G Ν Т Υ Κ v G D Т Y Ε Y Т R P Κ D AAATACACTGGGAACACTTACAAAGTGGGTGACACTTATGAGCGCCCTAAAGATTCCATG 301 121 W D C т C Т G Α G R G R Т S C T ATCTGGGÃCTGTACCTGCATCGGGGCTGGGAĜAGGCAĜGATCAGCTGTACCATTGCAAAT R C H E G G Q S Y K I G D K W R R P H E 361 141 CGCTGCCATGAAGGGGGTCAGTCCTACAAGATTGGCGACAAGTGGAGGAGGCCACATGAG 421 161 Ν W Т G G Y Μ Τ. E C Τ. С L G G Κ G E C 481 ACTGGTGGCTACATGTTAGAGTGTCTGTGTCTGGGAAATGGAAAAGGGGAATGGACCTGC 181 Ρ E D Н G Т S V Κ Т Α K C F Α Α 541 AAACCTATAGCTGAGAAGTGTTTTGATCATGCTGCTGGGACGTCCTACGTCGTGGGGGGAG 201 W Ε G W М Μ V D C Т 0 G 601 ACCTGGGAAAAGCCCTACCAAGGCTGGATGATGGTGGACTGTACTTGTCTAGGCGAAGGC 221 Т N С N G Т C S R R D Q D T AATGGACGCATCACCTGTACCTCCAGAAACAGATGCAACGATCÃGGACACCCGGACATCC 661 241 721 R G D Т W S K К D N R G N Τ. Τ. Q C V Т TATAGGATTGGAGACACGTGGAGCAAGAAGGACAACCGAGGAAACCTGCTTCAGTGTGTC 261 Ν W C Т G G R G E K E R H Α Τ. 0 S Α 781 TGCACAGGCAATGGCAGAGGGGAGTGGAAGTGTGAGCGACATGCTCTACÃAAGTGCTTCA 281 G S G S F Т D V R Т Α Ι Y Q Ρ Q Т Η Α GCCGGATCTGGCTCCTTCACTGATGTCCGAACAGCTATTTACCÄACCGCÄGACTCACCCC 841 301 Ρ G Η V Т D S G V V Α G 0 901 CÂGCCCGCTCCCTACGGCCACTGTGTCACCGACAGTGGTGTGGTCTACTCTGTGGGAATG Q W L K S Q G N K Q M L C T C L G N G V CAGTGGCTGAAGTCGCAAGGAAACAAGCAAATGCTGTGCACGTGCCTGGGCAATGGCGTC 321 961 341 S С 0 E Т Α V Т 0 Т Y G G Ν S Ν G E P AGCTGCCÄGGAGACAGCCGTGACCCÄGACTTATGGTGGCAATTCAAACGGGGAGCCCTGT V L P F T Y N G R T F Y S C T T E G R Q 1021 361 Т S C GTCCTCCCGTTCACCTACAACGGTAGGACCTTCTATTCCTGCACCACCGAAGGGCGGCAA D G H L W C S T T S N Y E Q D Q K Y S F 1081 381 1141 GACGGACATCTGTGGTGTAGCACAACTTCCAATTACGAACÃAGACCÃGAAGTATTCCTTC 401 Т D Н Α V L V 0 Т R G G Ν S Ν G Α 1201 TGCACAGACCATGCGGTTTTGGTTCAGACTCGAGGCGGAAATTCCAATGGTGCTCTGTGC 421 Ν Ν R Ν Y Т D F L C Т Ε G R ${\tt CACTTCCCCTTCCTGTACAACAACCGGAATTACACCGACTGTACTTCTGAGGGTCGCAGG$ 1261 441 D N Μ K W C G Т Т Q Ν Υ D Α D Q Κ F G GĂCAĂCATGAĂATGGTĞCGĞCAĊCAĊCCĂGAĂCTĀCGĀTGCCGATCÃGAAGTTTGGATTC 1321 461 V Ρ Μ Α Α Η Ε Ε Ι C Т Т Ν Ε G Μ Y R С 1381 TGCCCAATGGCTGCCCACGAGGAGATCTGCACAACCAATGAAGGGGTCATGTATCGCATT 481 W D Κ 0 Η D G Η М Μ R G D 0 L C Т C G GGGGATCAGTGGGATAAGCAGCATGACCTGGGCCACATGATGAGGTGCACGTGTGTGGGG 1441 501 Ρ Y Ν G R G Ε W Α С Ι S Q R D 0 L AACGGTCGTGGAGAATGGGCCTGCATCCCCTACTCCCÄGCTCCGAGACCÄGTGCATCGTT 1501 Ν 521 D Т Т Y N 77 D Т F Η K R H E GATGACATTACTTACAATGTGAACGACACGTTCCACAAGCGTCACGAGGAGGGACATATG 1561 541 C Т С F G G R G R W С D Ρ Ν 0 Κ Т D CTGAACTGTACCTGCTTTGGTCÂGGGCCGGGGCAGATGGAAGTGTGACCCCATTGACCÂG 1621 561 Т F Y 0 D S Е R Т 0 т G D S W E Κ 1681 TGCCAAGATTCAGAGACCCGGACATTTTACCAGATTGGTGACTCCTGGGAGAAGTTTGTG 581 V Y С Y С Y G R Т W н G R 0 G G E Η C 0 1741 CATGGTGTCCGATACCÃGTGTTACTGCTACGGCCGTGGCATCGGGGGGGGGCACTGTCÃA 601 V V Т Υ Ρ Т Т G P Τ L 0 G 0 Τ 1801 CCTCTGCÃGACCTACCCAGGCACAACTGGACCTGTCCÂAGTAATTATCACGGAGACCCCC 621 Ν Η 0 W Ν Α Ρ Ε Ρ S Η 0 1861 AGCCÂGCCCAATTCCCACCCCATCCÂGTGGAATGCCCCGGAGCCTTCACACATCACCAAG 641 Ρ TAT R V S N S S Δ D S D TACATTCTCAGATGGAGACCTGTGAGTAATAGCTCCGCAGCCTTGGACTCTGACCCCTGA 1921

Figure 3. Murine MSF protein and nucleotide sequences. The protein sequences highlighted are: MSF-unique decamer in red (sequence mMSF does not share with mouse FN). mMSF 15 amino acid region absent in MSF-aa in blue. IGD and HEEGH motifs in purple.