

Title

Aqueous outflow imaging techniques and what they tell us about intraocular pressure regulation.

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Abstract

Recent advances in the medical and surgical management of open-angle glaucoma have increased the number of treatment options available. Several new intraocular pressure (IOP)-lowering treatments target the conventional aqueous outflow (AO) system. However, success rates are variable and outcomes in individual patients are often difficult to predict. Variable treatment responses remain unexplained and highlight deficiencies in our current understanding of AO regulation and IOP homeostasis. Imaging is often relied upon to confirm diagnoses and monitor treatment responses in other ocular and systemic pathologies. As yet no suitable AO imaging tool has been developed to fulfill this role in glaucoma. A variety of imaging techniques have been used to study the AO tracts of humans and animals in *ex vivo* and *in vivo* eyes. In this review, results from novel imaging techniques that assess aqueous drainage through the episcleral venous system are considered and we argue these provide new insights into AO regulation. We suggest that the ability to objectively measure AO responses to interventions would be a significant clinical advance, and we have demonstrated that this can be achieved with direct visualisation of aqueous drainage. We predict that the evolution of AO imaging technology will continue to reveal critical components of AO and IOP regulation, and that personalised IOP-lowering treatment in glaucoma care may well become a reality in the near future.

Introduction

Intraocular pressure and aqueous outflow:

The spherical shape of the eye is maintained by the stretching of its elastic, collagenous envelope by its incompressible contents. The force returned by this corneo-scleral extension and the balance between aqueous secretion and drainage determine baseline intraocular pressure (IOP). However, higher-frequency fluctuations are superimposed over this baseline pressure, as a result of variations in the volume of the intraocular circulations during the cardiac cycle, and forces acting on the globe due to contraction of extraocular muscles.

Aqueous outflow (AO) is considered to be the major determinant of baseline intraocular pressure, and increased outflow impedance is an important risk factor for glaucoma. The majority of aqueous leaves the globe by the “conventional” pathway, traversing the trabeculum to enter the circumferential canal of Schlemm, from where it is delivered by drainage channels into episcleral veins on the ocular surface. Those veins in which aqueous maintains a visible, discrete column are called aqueous veins. The episcleral veins disgorge into the superior and inferior ophthalmic veins, which traverse the orbit, delivering their contents into the cavernous sinus.

Imaging of AO has largely concentrated on its emergence from the globe; however, this review will also consider techniques that illustrate its passage through the episcleral circulation.

Imaging of aqueous outflow:

Renewed interest in AO imaging has been generated by the introduction of minimally invasive glaucoma surgery (MIGS). A number of MIGS devices improve conventional AO by reducing outflow resistance, but IOP responses are unpredictable. This raises questions about the functionality of the conventional aqueous outflow system in glaucoma patients and what causes IOP dysregulation.

Elevated IOP is widely accepted as a major risk factor for primary open-angle glaucoma (POAG). IOP reduction has been successfully achieved by means of topical eye drops, laser and surgery. Interventions have emerged that augment conventional or uveoscleral aqueous outflow, or reduce aqueous production. As the size of the glaucoma population increases globally, there is greater demand for effective and consistent treatment responses so that outcomes and efficiency of care can be optimized. Novel treatment solutions are promising, but a greater understanding of functional AO is required to optimise treatment selection and timing. The mechanisms responsible for IOP dysregulation are elusive, although multiple findings from imaging studies support the presence of feedback control of aqueous drainage within the conventional AO system.¹⁻⁶

For the purpose of this review, we concentrated on the imaging of conventional aqueous outflow. Uveoscleral (unconventional) outflow represents a small proportion of total aqueous drainage⁷ and cannot be accurately imaged⁸. AO imaging techniques can be classified in many ways, but are commonly described as structural or functional.^{8,9} Structural imaging techniques identify anatomical

features that may explain how aqueous passes through the conventional outflow system, whereas functional techniques study actual flow through the system. Studies include *ex vivo* and *in vivo* eyes of humans and animals. Available imaging techniques vary in invasiveness and in whether static or dynamic (real-time) assessments of AO are possible. Using a non-invasive imaging system to study AO in physiological conditions is desirable and may create opportunities to move towards more personalised glaucoma care. AO imaging techniques are summarised in Table 1.

Methods

Searches of PubMed, Medline, Google Scholar, Embase, and Scopus were completed by 12th April 2020. Other sources were obtained from reference lists of selected studies, conference proceedings and generally available specialised websites. All relevant materials were included and not limited by date or original language of the publication.

Background

The first aqueous vein was reported in 1942 by Ascher, using slit lamp biomicroscopy¹. Prior to this, multiple descriptions, most likely representing aqueous flow within episcleral vessels, date back to 1921. Gartner published the first aqueous vein photograph in 1944¹, but detailed anatomical representations of AO channels have, until recently, only been possible in *ex-vivo* eyes. In 1951, Ashton elegantly displayed the pathways of aqueous veins, derived from Schlemm's canal (SC), with the use of neoprene casts (Fig. 1).¹⁰ These images are

often a prelude to discussions of AO. Now, with new technologies, AO imaging has been made possible *in-vivo* and this enables functional assessment of flow. The goal of imaging this region of the eye is primarily to explain mechanisms of IOP control that may lead to new therapeutic targets or assist with monitoring responses to treatment. The following section summarises imaging techniques used to study the aqueous outflow system, and discusses how they may help to identify regulatory mechanisms.

Structural aqueous outflow imaging

It has been widely documented that aqueous is secreted by the ciliary body epithelium and drains through the trabecular meshwork (TM) into SC and the episcleral venous system (EVS).^{9,11} Imaging the structural anatomy of the distal AO pathway, from SC into collector channels and episcleral veins, was initially made possible with the use of animal models and human *ex-vivo* eyes.

Microscopy:

Early use of imaging techniques on *ex vivo* primate, bovine and human eyes include light microscopy, scanning electron microscopy, and transmission electron microscopy.¹²⁻¹⁶ These techniques confirmed the drainage of aqueous and the anatomy of the AO drainage pathway (Fig. 2). The major anatomical contributors to AO include: the TM, the juxtacanalicular connective tissue (JCT), SC endothelium, collector channels, and the episcleral venous system.¹⁷ The inner wall of SC, comprised of the JCT and SC endothelium, is widely considered to be the predominant site of AO resistance.¹⁵⁻²⁰ Valve-like structures visualised

after mechanical dilation of SC with viscoelastic^{14,21} are thought to aid pulsatile aqueous drainage through the outflow system.² These anatomical findings form the foundation of major theories of AO regulation that will be discussed later.

Ribbon scanning confocal microscopy:

Ribbon scanning confocal microscopy (RSCM) was initially developed to speed up and improve the quality of brain tissue imaging.²² It is derived from similar principles seen in confocal microscopy; where a laser is used to evoke the emission of fluorescent light from a defined depth within a sample. Single pinhole-focused images are generated and stitched together to form an overall image of a specimen. Instead of imaging multiple points, RSCM sweeps across specimens and creates image “ribbons” that are more easily stitched together and with less artifact.²² As a result, imaging and processing occur ten times faster, and 3D reconstructions are possible (Fig. 3A).²³

Imaging techniques such as spectral-domain optical coherence tomography (SD-OCT) are unable to detect small networks of vessels within the AO tract.²⁴ RSCM shows these networks in porcine eyes labeled with lectin via an intracameral bolus.²⁵ Dense tubular networks and lumens within the TM have been demonstrated, along with uninterrupted vessel branches from the TM to the scleral venous plexus, likely representing aqueous veins.²⁵ Collector channels were larger and more often seen within the nasal hemisphere where the greatest outflow is thought to occur.²⁵ Studies using RSCM in porcine and human eyes show anatomical differences. Human eyes have larger and longer collector channels than porcine eyes, but they are fewer in number.²³

Two-photon microscopy imaging:

Trans-scleral visualisation of the AO system was demonstrated in *ex vivo* rat eyes by two-photon autofluorescence microscopy.^{4,26} Autofluorescence avoids the use of external fluorescent markers that may trigger tissue damage.²⁶ Initially, two-photon microscopy required surgical exposure to visualise SC. The subsequent use of additional filters refined the emitted signal and reduced the impact from scleral light scatter, enabling deep tissue imaging.²⁷ This method prevents the use of tissue fixatives that are often required in tracer studies and may cause tissue distortion.²⁶ A stack of generated images is reconstructed to form images of the TM, SC and aqueous veins.^{4,26} The AO tract was able to be reconstructed in 3-dimensions using specialised software that generated measurements of vessel lumens.⁴ In addition, the cells within aqueous drainage tissues were labeled and profiled to identify their actions. Simulated IOP elevations have been shown by two-photon microscopy to cause local tissue damage to the TM and surrounding tissues.²⁶ Shifting from simulated results in rodent eyes to modelling of the human AO pathway requires further refinement.

Three-dimensional micro-computed tomography:

Many *ex-vivo* imaging techniques produce data in sections or fragments that need to be reconstructed to represent the entire AO system. Conversely, three-dimensional micro-computed tomography (3D micro-CT) is a technique that generates an image of the whole anterior segment including TM, SC, collector channels, and episcleral vasculature (Fig. 3B).²⁸ *Ex vivo* human eyes were fixated with immersion or perfusion techniques and mounted on a rotating device. A

molybdenum X-ray scanner was used to collect raw data that was then converted to allow 3D reconstruction and analysis.^{28,29} Osmium tetroxide 1% was used as a contrast agent to improve X-ray quality. Measurements of some AO components were possible, but perfusion fixation may have artificially opened flow pathways.²⁸ Ex vivo tissue processing can cause collapse of smaller vessels, which may explain the absence of deep scleral plexus vessels, when compared to neoprene casts.²⁸

Spectral-Domain Optical Coherence Tomography:

The anatomy of the AO system is not amenable to standard optical coherence tomography (OCT) imaging. The episcleral vasculature is complex and difficult to resolve due to light scattering from the surrounding sclera.^{30,31} Despite this, the advancement of OCT technology made non-invasive imaging of the AO pathway possible for the first time. Spectral-Domain OCT (SD-OCT), with the assistance of Doppler signals, demonstrated the area of SC was larger in the nasal hemisphere compared to the temporal hemisphere. In a small number of subjects, patients with glaucoma had a smaller SC area than controls.³² No evidence of SC or collector channels could be seen in one subject who had a functioning glaucoma drainage device. However, complete 360 degree imaging of SC was not possible in this case, because images were taken in cross-section and blood vessels caused shadowing artifact.³² Developments in image processing have led to complete reconstructions of the AO system in both ex-vivo and in-vivo human eyes using virtual casting (fig. 3C).³³ However, vasculature appeared less distinct *in vivo*, and vessels less than 10 μ m were poorly-resolved. In addition, arteries and veins could not be reliably differentiated.³³

Novel techniques incorporating OCT have been implemented to minimise image degradation, and to further characterize the relationship between TM and SC. An endoscope was able to image collector channels from within SC in ex vivo human eyes. The SS-OCT images were comparable to subsequent images taken with SEM.³⁴ Similarly, direct imaging of TM and SC through the anterior chamber produced comparable images to SEM. The addition of concurrent fluid pulses forced through SC allowed visualisation of how the TM responded to pressure changes.³⁵

Swept-Source Optical Coherence Tomography:

Successful visualisation of the AO system requires high-resolution imaging due to the complexity of the anatomy. Advancements in OCT technology have achieved resolutions in the tens of micrometers and made full-depth imaging of the anterior segment possible.³⁶⁻³⁸ Swept-source OCT (SS-OCT) has demonstrated the structures of the human drainage angle, as well as the post-trabecular AO pathway within a single B-scan image (Fig. 4).³⁰ Validation of SS-OCT has been made with comparison to histological specimens.³⁹ Vasculature within the episclera and sclera is complex and pathways of vessels are easily lost within a single B-scan. A technique used in computed tomography (CT) scanning, multiplanar reconstruction, provides a solution to this by compiling a continuous cross-section image of the area of interest.³⁰ The path of vessels can be studied using curved multiplanar reconstruction, a technique that involves re-slicing three-dimensional data. This has been applied to SS-OCT to generate a two-dimensional reconstructed image of the pathway between SC and the episcleral

venous plexus.³⁰ However, it may be difficult to differentiate between vessels carrying aqueous and blood.

Measurements of SC area and its perimeter obtained with SS-OCT are comparable to estimates generated by human perfusion and histological studies.^{36,40} Chen et al⁴¹ used a commercially available SS-OCT (CASIA SS-1000; Tomey Corporation, Nagoya, Japan) to demonstrate a reduction in the diameter and area of SC with ageing.⁴¹ Variations within SC were seen, the widest measurements being in the nasal and temporal quadrants. Increasing age correlated with TM thickening in the nasal and temporal quadrants.⁴¹ SC was visible in 80.6% of eyes between 60-83 years of age. The authors suggest that reduced SC size and detection rates may result from tissue degeneration or shrinkage, a decrease in aqueous production with age, or distortion of the corneoscleral contour due to loss of ciliary muscle movement.⁴¹ Optimisation of scan accuracy is required to completely validate this technique.³⁹ Images are taken with the patient gazing to the opposite direction of the target quadrant, and not in primary position. There may also be diurnal variations in the morphology of SC that have not been identified in the current studies.⁴¹

Swept-source OCT has been used to reconstruct images of the AO pathway before and after Trabectome and 360-degree suture trabeculotomy, techniques used to reduce AO resistance.⁴² No significant structural changes were detected as a result of these surgical techniques, despite good visualisation of the AO pathway.⁴² The ability to compare pre- and post-operative SS-OCT data for an individual is challenging. Variability in diurnal AO and focus quality, as well as

difficulty identifying the corresponding pre-operative AO pathway are limitations identified by the authors of this technique.⁴² In another study, a 90% increase in SC area within the nasal and temporal quadrants was demonstrated 8 hours after instillation of travoprost 0.004% eye drops.⁴³ However, similar difficulties arose with identifying the corresponding pre-intervention site for comparison.

Optical Coherence Tomography Angiography:

The introduction of OCT angiography (OCT-A) has enabled the detection of blood flow within vessels of *in vivo* human eyes.^{38,44} Aqueous is more challenging to visualise with OCT-A than blood, so concurrent use of contrast is required. Akagi et al³⁸ compared OCT-A with fluorescein scleral angiography and aqueous angiography using indocyanine green (ICG).³⁸ En face OCT-A images at varying depths were reconstructed to represent the circumferential flow of superficial conjunctival vessels (> 200 µm) and deeper intrascleral vessels (200 to 1000 µm) (Fig. 3D).³⁸ The deeper vasculature represents the episcleral and intrascleral vessels responsible for draining aqueous from SC. Deep OCT-A vessel patterns reasonably correlated reasonably well with those generated by aqueous angiography, but further work is required to refine the technique into a reliable clinical tool.

Many aqueous-carrying vessels also contain red blood cells.¹ OCT-A will detect these vessels as long as there is concurrent blood flow^{31,38}, but will not detect vessels containing only aqueous. Contrasts are used to improve visualisation with many other imaging modalities, but there is no dedicated contrast for OCT-

A.³¹ Gold nanorods have been shown to generate measurable Doppler OCT signals⁴⁵ and lipid emulsions (milk and propofol) induced strong OCT-A signals³¹. The findings from both of these studies are limited to *ex vivo* porcine eyes^{31,45} OCT contrast studies are yet to be undertaken in human eyes.

Functional aqueous outflow imaging

Imaging techniques are evolving to encompass the dynamic nature of aqueous outflow. However, little is known about the physiology of AO distal to the trabecular meshwork and how drainage of aqueous is affected in glaucoma.^{25,46} Further study is warranted because the episcleral venous system is likely to be responsible for 25-50% of AO resistance.^{18,47} Visualisation of aqueous drainage is challenging and traditionally has been made possible with dyes such as fluorescein.⁴⁶ Recent imaging advances have facilitated the analysis of functional AO.

Fluorophotometry and tracers:

The imaging techniques discussed thus far provide structural representations of the AO system, but they are unable to demonstrate functional flow. Early attempts to study aqueous flow originate from fluorophotometry, a technique to measure aqueous concentrations of fluorescein in animal and human eyes.⁴⁸⁻⁵⁰ Fluorescein may enter the anterior chamber of the eye via intravenous, oral, or topical eye drop administration.⁵⁰⁻⁵² Disappearance of fluorescein from the anterior chamber is measured over time to reflect the rate of aqueous exchange.

Multiple studies have recorded the rate of bulk aqueous flow in normal human subjects to be 1.6 to 3.4 $\mu\text{l}/\text{min}$.⁵² However, fluorescein clearance can be affected by physiological states, such as pupil dilation. Some fluorescein may be lost posteriorly in the eye and that may impact aqueous flow calculations.⁵³ In addition, direct visualisation of fluorescein drainage from the eye is not possible with this technique.

A variety of tracers have been used to study aqueous drainage in more detail; particularly, to identify flow variations around the circumference of the eye and anatomical responses to varying perfusion pressures.⁵⁴ Cationic ferritin, gold particles, fluorescent microspheres and nanospheres, latex beads and quantum dots have all been used as tracers.⁵⁴⁻⁶¹ Most of these studies perfuse *ex vivo* animal or human eyes with tracer. Confocal, light, or electron microscopy techniques are then used to generate images of the labeled and processed tissue sections.⁵⁴⁻⁶¹ These techniques rely on tissue adherence or active cellular uptake of tracers to help identify structures within the AO pathway. Tracers have a larger molecular weight than water, and so they accumulate at the TM and collector channels.⁸

Segmental labeling of the TM in multiple studies suggests regions of preferential aqueous outflow in all eyes.^{54,58,59,61} However, one study demonstrated homogenous cationic ferritin labeling in normal eyes compared to segmental patterns of uptake in glaucomatous eyes.⁶² Further work in healthy *ex vivo* human eyes suggests preferential outflow where pigmented TM is in close proximity to collector channels.⁶¹ Tracer imaging has also been used to illustrate

the IOP-lowering mechanisms of Rho kinase inhibitors, which include TM and JCT expansion, and vasodilation of episcleral veins.⁵⁶ Despite the significance of the findings from tracer studies, there are a number of limitations due to the use of *ex vivo* eyes. Conditions are not physiological and do not account for other variables such as; aqueous production, ciliary body tone, EVP, and uveoscleral drainage.⁵⁶ Processing of histological sections is required to assess tracer response, which makes this imaging technique incompatible with studying *in vivo* human eyes.⁸ Real-time analysis of AO is required to best reflect the pathophysiology of IOP dysregulation as seen in glaucoma.

A similar technique, canalography, is another form of tracer study, but histological section is not required. Fluorescent microspheres have been injected into the anterior chambers of *ex vivo* porcine eyes and visualised using a dissecting microscope capable of fluorescence imaging.^{63,64} In studies of non-glaucomatous porcine eyes, canalograms have demonstrated regional and circumferential outflow patterns with an increase in fluorescence nasally (Fig. 5).⁶³ This finding was accentuated following *ab interno* trabeculectomy, but improvement of flow and peak fluorescence was seen in all quadrants.⁶⁴ Another *ex vivo* porcine study used canalography to demonstrate the effects of trabectome-mediated *ab interno* trabeculectomy and trabecular micro-bypass implantation.⁶⁵ Fluorescein perfusion was initially administered to identify AO, and then Texas red was perfused post-procedure to demonstrate the effects of each surgery. Both surgical techniques achieved a regional improvement in AO.⁶⁵ These studies provide validation for surgically reducing trabecular resistance,

but limitations of the use of *ex vivo* primate eyes prevent comparison with what occurs in the episcleral venous system.

Intra-operative Techniques:

Channelography:

Surgical access to SC using techniques such as viscocanulostomy and canaloplasty have created opportunities to image the AO system more clearly. Viscocanulostomy is achieved by dilating SC with a viscoelastic,⁶⁶ whereas canaloplasty includes the additional step of leaving behind an intracanalicular suture thread to maintain SC.⁶⁷ The goal of both techniques is to reduce AO resistance by expanding SC, however, the success of these surgical techniques depends on the functionality of the distal AO system.⁴⁶ The desire to improve surgical success rates has led to further steps being added at the beginning of the surgery to ensure AO patency. Provocative gonioscopy or ocular decompression cause blood to reflux into SC, and this can be easily visualised with an intraoperative gonioscope. Fluorescein may be injected from a microcatheter into SC to enable downstream visualisation within the EVS and reflux into the anterior chamber (Fig. 6A).^{46,67} The latter technique became known as channelography, in which fluorescein is directly visualised with the operating microscope.⁶⁷ The health and permeability of the TM may be represented by the amount of fluorescein that leaks into the anterior chamber.⁴⁶ Similarly, a greater number of episcleral vessels that accept fluorescein within an eye may reflect superior functional capacity within the distal drainage system.⁴⁶ However, channelography bypasses outflow resistance generated by the TM and requires

manual perfusion of contrast directly into SC.⁶⁸ Both of these factors mean that patterns of outflow are non-physiological and do not replicate aqueous outflow, despite providing clear images of the outflow tracts.

Indocyanine green (ICG) channelography has been achieved with a near-infrared camera in human *in vivo* eyes.⁶⁸ The choice to use ICG combined with viscoelastic was to prevent excessive leakage from SC and collateral fluorescence. In some eyes, areas of SC appeared non-patent as evidenced by incomplete fluorescence. Collector channels only filled in corresponding areas of SC patency, suggesting possible intracanalicular obstruction.⁶⁸ Increasing the perfusion pressure and volume enabled visualisation of collector channels not previously evident. Retrograde filling by ICG of glomerular patterns within the ring of SC were not fully explained. The authors state that the structures did not leak and suggest they may form part of the iris or even the lymphatic system.⁶⁸ Although there is no clear explanation for these structures, it is conceivable that they, or potentially other undetected structures, contribute to IOP regulation.

Operating microscope:

Surgical intervention provides a unique opportunity to study the AO system and the effects of procedures that target the trabecular meshwork. Variable MIGS results have driven studies that particularly focus on advancements in real-time image acquisition and AO visualisation. A number of intra-operative techniques have been reported that help evaluate aqueous drainage. In one technique, known as episcleral venous fluid wave (EVFW), a pulse of balanced salt solution was suddenly released by the irrigation and aspiration hand-piece to examine

the patency of the distal episcleral vessels following cataract surgery combined with trabectome (ab interno trabeculotomy).⁶⁹ The authors postulate that the presence of an EVFW indicates that the episcleral venous system is able to accept aqueous. Conversely, the absence of an EVFW may predict a higher chance of surgical failure, possibly due to a blockage, stenosis, or atrophy within the system.⁶⁹ In a retrospective study, the extent of EVFW around the circumference of the eye inversely correlated with IOP in the short-term post-operative period.⁷⁰ Eyes with poor or no visible EVFW were associated with a poor trabectome result and a one third chance of requiring further glaucoma surgery.⁷⁰ Furthermore, the extent of EVFW was compared between trabectome and iStent, and both were found to generate segmental flow improvement.⁷¹ Trypan blue has been used to highlight flow within the EVS following ab interno TM excision with the Kahook Dual Blade (New World Medical, Rancho Cucamonga, CA).⁷² This technique essentially improves visualisation of the EVFW. Whilst the detection of EVFW and its extent may help to assess surgical success or prognosis on the operating table, it does not reflect true physiological drainage or functionality of the AO system. The potential effects of phacoemulsification on AO are also not addressed.

Aqueous Angiography:

Another intraoperative imaging technique, aqueous angiography (Fig. 6B), was developed to enhance visualisation of AO pathways in real-time. Initial work was on *ex vivo* porcine⁷³, bovine⁷⁴ and human eyes^{73,75}, but intraoperative studies on *in vivo* non-human primate eyes⁷⁶ and then human eyes⁷⁷ soon followed.

Aqueous angiography incorporates steps from a number of imaging techniques

that have already been discussed. It takes advantage of intraoperative access to the proximal aqueous drainage structures via the anterior chamber, and incorporates a tracer that can be externally detected with a bespoke angiographic imaging system. A specialised arm supports the angiography equipment (Spectralis HRA+OCT Flex module; Heidelberg Engineering, Heidelberg, Germany) above the patient's eye, which is centred with the assistance of confocal scanning laser ophthalmoscopic infrared images.⁷⁶⁻⁷⁹ The anterior chamber is supported by a continuous infusion whilst a fluorophore is introduced intracamerally. Although the anterior chamber dynamics are non-physiological, this method examines the functionality of the TM and subsequent flow into distal outflow channels. Angiographic images are recorded as the patient is asked to rotate the eyes in various directions to avoid obstruction by the eyelids. This technique enables real-time assessment of circumferential aqueous drainage in live human eyes.⁷⁷⁻⁷⁹

Both ICG and fluorescein have been safely administered as intracameral tracers. Sequential use of tracers (ICG first) is possible and yields similar results.^{74,75,78,79} This permits angiography to demonstrate the flow response before, and immediately after, an intervention. Aqueous angiography has been applied to support established AO anatomy findings, identify spontaneous dynamic changes within the AO system, and to demonstrate that areas with poor angiographic signal can be recovered with trabecular bypass surgery.^{77,78} Although aqueous angiography comes closer to visualising physiological AO than other imaging techniques there are still a number of limitations that prevent this goal from being achieved. It is an invasive process and requires patient sedation, pupil

dilation and topical anaesthesia prior to surgery, all of which are likely to affect AO and IOP. The use of an intraoperative speculum and anterior chamber maintainer alter IOP and aqueous dynamics.⁷⁷ Fluctuations in IOP during surgery also affect aqueous dynamics and imaging.⁷⁸ Despite these limitations, aqueous angiography has contributed significantly to our understanding of the AO system and how MIGS can interact with AO. Nevertheless, non-invasive imaging is required to assess physiological AO.

Functional Imaging in the Clinic:

Phase-sensitive Optical Coherence Tomography:

Pulsatile AO was initially reported by Ascher⁸⁰ and was thought to originate from SC in synchrony with ocular pulsation.^{80,81} The static nature of OCT and OCT-A is a significant limitation when imaging dynamic aspects of the AO system. Li et al^{35,82} described a novel OCT technique to enable assessment of TM motion in living human eyes.^{35,82} Phase-sensitive OCT (PhS-OCT) is able to quantify tissue motion to the nanometer, a measurement not possible with traditional OCT.³⁵ The technology is derived from the use of Doppler OCT to measure blood flow.⁸³ Each standard OCT B scan contains 500 A scans, and it is the measurement of the phase changes between successive A scans that reflect tissue movement.^{83,84} IOP variations were generated in an *ex vivo* primate model to simulate ocular pulsations. The TM was displaced towards SC in synchrony with each generated pulsation. Artificial elevation of IOP reduced TM displacement and SC area was reduced.³⁵ PhS-OCT has advanced to permit imaging of human subjects on a system similar to a regular outpatient slit-lamp. This technology has been used

to support the findings of Johnstone et al^{3,85} who identified a model of IOP regulation controlled by pulse-dependent TM motion.^{3,85}

Haemoglobin Video Imaging:

Until recently, imaging of conventional aqueous outflow pathways has required the introduction of a marker into the anterior chamber. However, Haemoglobin Video Imaging (HVI), a slit-lamp technique originally developed to investigate limbal microvascular blood flow⁸⁶, can demonstrate these pathways without the need for more invasive methods^{86, 87}.

HVI exploits the absorption spectrum of haemoglobin to demonstrate red blood cells in the conjunctival and episcleral circulations^{86,88,89}, and can illustrate wherever venous blood is parted by clear cylindrical columns of aqueous: “aqueous veins”.

The circulations are illuminated by a broad bandwidth (540-580nm), which corresponds with the long-wavelength absorption peak of haemoglobin, causing erythrocytes to appear dark against the background of bright, reflective sclera. In contrast, aqueous is seen as a paraxial, translucent lamina within the blood column.

The images are captured with a sensitive, high-resolution, monochrome video camera and bespoke stabilisation software eliminates motion artefact due to micro-saccades.⁸⁸

Patients are examined with HVI in a darkened clinic room and are positioned as they would be for standard slit lamp examination. The entire limbus can be recorded by asking the patient to adjust fixation.

Familiarity with the ocular surface microcirculations⁸⁹ is helpful to differentiate their complex hierarchy (Fig. 7). In general, vessels delivering blood to microcirculations tend to be more tortuous, sometimes with intermittency of flow, and they may demonstrate advancing constrictions. Blood in drainage vessels appears granular, and converging venules add successive laminae to the blood column.

The smallest vessels in the episcleral circulation often form an interconnecting lattice and appear larger than conjunctival capillaries. Constrictions are not visible and flow may freely reverse. Perfusion of any segment of the ocular surface microcirculation may be periodic, erythrocytes remaining static for long periods.⁸⁹

Aqueous veins are a consequence of laminar flow within venous blood. As successive venous channels converge, they contribute to the periphery of the venous column, remaining discreet for considerable distances before they ultimately combine.

Since aqueous usually enters the venous system early in its course, aqueous laminae tend to lie centrally. However, when aqueous is added to an established blood column, it usually remains peripheral until displaced axially by subsequent contributions of venous blood (Fig. 8).⁹⁰

Visualisation of aqueous using HVI was first reported in 1989⁸⁶ and creates a unique opportunity to study physiological AO in living human eyes⁸⁹. Although aqueous is detected indirectly, by the displacement of erythrocytes within episcleral collecting veins (Fig. 7B), a number of aqueous vein characteristics previously reported by Ascher¹ have been demonstrated.^{87,90} These include: pulsatile flow corresponding with the heart rate, parallel stratification of flowing blood and aqueous, disruption of the aqueous stream during eye movements and gentle globe compression, and redirection of aqueous flow into adjacent episcleral vessels following localised aqueous vein compression.

HVI has recently graduated from a qualitative to a quantitative modality⁸⁷ (Fig. 9A). In HVI, absorption of incident light increases with thickness of the erythrocyte barrier between the light source and sclera. Computer modelling of the intensity of light absorbed by transepts of aqueous veins demonstrates that the transition between blood and aqueous corresponds with the transmission minima. Therefore, from light intensity transepts (ImageJ - open source software), it is possible to calculate aqueous column cross-section area (AqCA), a statistic presumed to be proportional to bulk flow in the cylindrical aqueous column^{87,90} (Fig. 9B,C). At least three transepts are measured and the calculations, averaged. Measurements of AqCA have been used to demonstrate improvements in AO following selective laser trabeculoplasty and trabecular bypass surgery (iStent Inject).^{87,90}

Although HVI has many potential clinical applications, its use in studying AO is in its infancy. At the time of writing, the technology has not yet developed into a turnkey ancillary test. Visualisation of AO within the EVS with HVI is straightforward, but quantification techniques are currently laborious and need to be streamlined. Measurements vary between vessels within the same eye, but the same vessel can easily be identified in successive scans for comparison and to assess the effects of interventions. A normative database is not yet available for AqCA and further work is required to establish diurnal variations in aqueous flow. Despite this, HVI is the only aqueous outflow imaging system that can generate anatomical and flow data non-invasively in physiological conditions in human eyes *in vivo*. The proximal drainage structures, TM and SS, cannot be seen with HVI, meaning complimentary imaging methods such as OCT will be needed to provide functional assessments of the entire outflow system.

Future directions for HVI include three broad approaches. Firstly, the definition of normal AO is required to contextualise changes seen in varying levels of IOP and glaucoma severity. The purpose of this is to assist with glaucoma staging and it may lead to more detailed classification of open-angle glaucoma. Secondly, it is necessary to study the effects of medical, laser, and surgical interventions on AO in order to identify factors associated with success, enabling a personalised approach to IOP-lowering treatment. In particular, information on the patency of the AO system may help to refine our choice of surgical tool. Or in the case of TBS, the location of device insertion may be optimized to improve IOP-lowering outcomes. Lastly, HVI creates an opportunity to observe, manipulate, and study

physiological aqueous drainage. HVI findings may advance the current understanding of mechanisms that control AO and IOP regulation.

The role of imaging in theories of aqueous outflow regulation:

The eye is amenable to imaging and this has been exploited, particularly for retinal conditions. However, the AO system is a far greater challenge due to its complex and concealed pathways through varying densities of ocular tissue. Emerging technologies have exposed the anatomy and some functional aspects of the AO system. The presence of homeostatic mechanisms responsible for AO drainage and IOP control has been widely speculated in the literature for some time.^{2-6,90} It is the regulatory mechanism that continues to elude researchers and its discovery will likely revolutionise glaucoma diagnosis, prognosis, and treatment. Imaging techniques that define structure have become non-invasive in the form of various types of OCT scanning, but functional AO assessment has, until recently, been invasive. Despite many challenges, imaging studies have provided researchers with critical information, leading to detailed theories of AO regulation. Our understanding of IOP homeostasis is likely to invoke evidence from all these imaging modalities.

What have imaging studies taught us about IOP regulation?

Static *ex vivo* and *in vivo* imaging studies have described the structure of the AO system in great detail. It is generally understood that aqueous flows from the anterior chamber through TM and into SC. From there, aqueous enters collector channels and flows into the deep scleral plexus. Intrasceral collector vessels

drain aqueous into episcleral veins that flow into the ophthalmic veins, which traverse the orbit to join the cavernous sinus. More detail has been uncovered by recent imaging studies to begin explaining the intricacies of aqueous flow through this pathway and to test theories of AO. It is important to remember that histological data contribute considerably to our understanding, but are beyond the scope of this review.

Limitations of the Passive Flow Theory:

There is now a large volume of corroborative evidence to refute the theory that aqueous drainage through the conventional outflow pathway is purely passive,^{11,20} down a pressure gradient.⁵ Variations of flow patterns have been visualised throughout the system. Regions of preferential flow exist within the TM⁵⁴ and evidence of asymmetrical SC and EVS drainage patterns, as well as the discovery of valves within SC have been uncovered by AO imaging.^{54,61,67,68,71,75-77,90} Intraoperative channelography and episcleral venography studies support findings from tracer studies that SC is discontinuous. Recurring anatomical findings suggest the eye is structured to drain aqueous predominantly in the nasal and inferior quadrants.^{71,75-77,90,91} There is no specific pattern to the distribution of collector channels identified with 3D micro-CT²⁷, but variations in anatomy throughout the circumference of the eye have been demonstrated with multiple imaging techniques to indicate preferential drainage.^{25,32,39-41,91} RSCM identified that scleral venous plexus vessels receiving aqueous were larger in the inferior limbus and the largest collector channels were in the nasal quadrant.²⁵ SC diameter in living human eyes was shown to be larger in the nasal quadrant when imaged with S-D OCT³² and SS-OCT³⁹⁻⁴¹. 3D micro-CT showed that SC was

attenuated superiorly, but appeared dilated at collector channel openings.²⁸ Deep vasculature was more dense and complex within the nasal and temporal quadrants than elsewhere, perhaps indicating sites of preferred AO.³⁸ However, anterior segment OCT-A identifies luminal structures regardless of their function, so it is difficult to draw definitive conclusions.³⁸ Intra-nasal AO has been shown to predominate in human flow studies.^{68,61,75-77,90} This information not only reassures surgeons to continue targeting the nasal quadrant when performing TBS, but also cultivates the notion that aqueous drainage may be regulated, rather than passively leaving the eye.

The Funneling Theory:

The trabecular meshwork is thought to account for approximately 50-75% of AO resistance in human eyes.^{11,20,22,32} The ultrastructural relationship of the JCT and SC endothelium has been visualised with electron microscopy.¹⁷ Elastic fibres of the JCT lie tangentially to SC endothelial cells. This anatomical relationship may be responsible for baseline TM resistance to AO, but the specific site of resistance continues to be debated in the scientific literature.^{17,92} Out-pouchings of SC endothelium into the canal lumen are often referred to as 'giant vacuoles'. Electron microscopy has also detected pores at the distal end of vacuoles, which are thought to aid transcellular fluid movement.⁹² The presence of these pores forms the basis of the 'funneling theory' of AO.^{93,94} Essentially, aqueous is forced to drain preferentially through the JCT in regions where giant vacuoles and pores are present. Based on this theory; the JCT provides most of the resistance to AO, but giant vacuoles and pores within SC endothelium can moderate resistance and flow.⁹²⁻⁹⁵

The funneling theory is supported by the finding of asymmetrical and segmental trabecular outflow. Tracer studies confirm segmental flow through the TM with labelling of only 10-20% of the SC inner wall.⁹² Variations in TM thickness; specifically thickening of TM in regions of active outflow compared with inactive areas, and higher numbers of collector channels in regions of active flow, substantiate segmental AO.⁹¹ Thickening of the TM may be due to JCT expansion. This has been highlighted by fluorescein bead deposition adjacent to collector channels in response to raised IOP in perfused *ex vivo* human eyes.⁶¹ Although pores have been implicated in the control of outflow facility in a number of studies,⁹⁶ the funneling theory is limited by the potential for tissue fixation processes to create artefactual pores.⁹² There is also poor correlation between measured AO resistance and SC inner wall pore density in a number of studies.⁹³

The Mechanical Pump Theory:

Anatomical variations throughout the circumference of the anterior segment provide clues about how aqueous may flow through the conventional outflow system. However, real-time observation is required to appreciate the dynamic nature of the AO system and understand its physiology. Evidence of pulsatile aqueous flow in synchrony with cardiac pulse dates back to Asher's observations¹, and the phenomenon continues to be reported as imaging technologies develop.^{2,3,76,77,81,88,90} The TM does not contain smooth muscle, but contractions may originate from nearby connections of ciliary muscle fibres.⁴ Pulsatile aqueous flow has been observed in normal and glaucomatous eyes, but with reduced pulsatility in the latter group.³ Some IOP-lowering medications

have been shown to increase pulsatile AO.³ More recently, TM movement, in synchrony with ocular pulse, has been demonstrated with PhS-OCT.^{35,82,97,98} This reflects the elastic properties of the TM and indicates its possible role in maintaining IOP homeostasis.⁵

TM motion, as detected by PhS-OCT, provides evidence to support the 'mechanical pump' theory. The basis of the theory is that ocular pulse, blinking, and sudden changes in eye position create transient increases in IOP that propagate through the AO system. Outward deformation of the TM causes SC narrowing and expulsion of aqueous into collector channels. As the pulse subsides, the TM rebounds to its initial position and SC widens. Aqueous from the anterior chamber then moves down a pressure gradient through valves to fill SC.^{2,84,98} The TM may contribute to maintenance of IOP homeostasis by varying its compliance with each pulsation. Ongoing insults to TM integrity could possibly cause permanent anatomical and functional changes. For example, in glaucomatous eyes, a reduction in pulsatile aqueous flow may be caused by an increase of TM stiffness.^{6,98} Also, IOP changes have been shown to alter AO pulsatility. Ocular hypertension (>28 mmHg) causes a reduction in visible pulsation.³ It has been hypothesized that progressive apposition of SC walls due to TM movement during raised IOP increases AO resistance.⁵ Temporary lowering of IOP below a physiological level also reduces pulsatile flow.³ All of this evidence favours mechanical pumping of aqueous into collector channels, but other active drainage mechanisms are equally likely to be present within the distal outflow system. Despite the contribution of the TM, 25-50% of residual AO

resistance^{11,20,22,32} remains unexplained and must occur within SC or the distal outflow tract.

The Role of Schlemm's Canal:

Schlemm's canal is a complex structure that was previously thought to act merely as a reservoir to collect aqueous from the anterior chamber and distribute it into aqueous veins.¹ Through *ex vivo* imaging studies we now understand that there are microstructures within SC that may influence AO control. Adhesions and septal columns have been visualised within areas of SC using scanning electron microscopy. They likely maintain patency of the canal and prevent inner wall tissue from causing herniating blockages into nearby collector channels.¹⁶ In addition; a lumen within septal columns may facilitate one-way drainage of aqueous across SC to collector channel openings.^{16,6} Valve-like structures have been identified at the collector channel openings, which may prevent backflow into SC.^{5,16} The intraluminal structure of SC further supports an underlying mechanism behind AO regulation. Experimental manipulations of IOP and SC also demonstrate a relationship between the two. IOP was artificially elevated using an ophthalmodynamometer and S-D OCT imaged SC.^{99,100} Raised IOP reduced SC cross-sectional area by 32% in healthy eyes,⁹⁹ compared to a 50% reduction in cadaver eyes.¹⁰⁰ Furthermore, increasing SC-CSA with an intraluminal scaffold caused a reduction in IOP.¹⁰¹ These findings indicate that there is likely a baseline structural stiffness of SC inner wall that helps to maintain SC patency and IOP control. This creates a “chicken and egg” scenario where loss of TM and SC integrity over time (TM thickening and shrinking of

SC)⁴¹ may cause raised IOP, or pathologically raised IOP may cause loss of structural integrity.

Potential for distal outflow regulation:

Further proof of the dynamic nature of aqueous drainage is seen with real-time aqueous angiography. Spontaneous variations in regions of outflow appear as increasing or decreasing angiographic signal.^{76,77} Similar AO fluctuations are seen under physiological conditions with HVI and are thought to be due to diurnal variation.⁹⁰ It is possible that these changes reflect homeostatic responses in the form of localised vessel wall contractions, control of flow through TM or SC, or both. An initial absence of flow does not necessarily reflect whether a vessel is able to accept aqueous.⁷⁸ This can be confirmed by applying localised pressure to the globe or directly to an aqueous vein. Aqueous flow is then directed into adjoining blood-filled vessels.^{1,87} It is also possible that as structures, such as TM, SC, or collector channels, become pathologically altered over time, flow is diverted towards healthier pathways within the system to reduce the burden and maintain IOP.⁶¹

Recovery of AO in low flow regions has been achieved with TBS and verified with aqueous angiography.^{75,78} Furthermore, HVI has illustrated that TBS can re-establish AO in regions of the microcirculation previously devoid of aqueous.⁹⁰ The non-invasive nature of HVI enabled serial follow-up of post-operative TBS (iStent Inject®) patients. Recovery or improvement of AqCA was not apparent immediately after surgery. In some cases it took 4 weeks for an aqueous stream to become established following periods of fluctuation.⁹⁰ This may indicate a

gradual process of SC or collector channel reperfusion, or realignment of homeostatic mechanisms. Unpredictable variations of IOP within the initial weeks after TBS likely occur due to these adjustments.⁹⁰ Variable longer-term IOP results seen with all forms of TBS and trabectome⁹³ prove the significance of post-trabecular outflow resistance and support the initial work of Grant^{18,20}.

The episcleral venous system is a complex and heavily innervated venous web with multiple arterio-venous anastomoses (AVAs), characteristics that favour its potential involvement in IOP regulation.¹⁰² There is some evidence that the EVS may modulate outflow resistance in mice. Smooth muscle was identified within vein walls of the distal AO tract using tissue labeling and two-photon microscopy.⁴ This may indicate that veins within the episcleral system are built to respond to dynamic pressure changes in a similar way to blood pressure control.

None of the aforementioned theories of AO regulation consider the influence of episcleral venous pressure (EVP). The resistance generated by EVP opposes aqueous outflow and acts as a natural mechanism to prevent hypotony.

Pathologically elevated EVP is well known to raise IOP.¹⁰³ EVP is challenging to measure accurately, especially non-invasively and in physiological conditions.^{103,104} This limits the study of EVP in the context of IOP regulation.

However, the interaction between blood and aqueous within the EVS is evidence of a potential role for EVP in IOP control. Now, with HVI, visualisation of this dynamic relationship is possible.^{87,90}

Future directions:

The goal of AO imaging is to create an understanding of IOP control, which may enable targeted and personalised IOP-lowering treatments. We need to understand AO regulation to interpret inconsistent responses to current IOP-lowering interventions; knowledge that may guide future treatment decisions. Vital information may be uncovered by observing physiological AO in an outpatient setting. Anterior segment OCT-A is an evolving technology, but aqueous cannot be resolved without the use of a contrast agent. Early work on *ex vivo* porcine eyes demonstrated the potential use of contrast in conjunction with anterior segment OCT-A.^{31,43} However, it may be our ability to visualise the response of AO to surgical and pharmacological interventions that allows us to unlock critical elements of IOP control. Real-time imaging techniques such as PhS-OCT, aqueous angiography, and HVI provide unique opportunities to capture such moments. Correlating imaging findings taken from the same patients may provide further clarity. None of these systems are currently available commercially and aqueous angiography is an invasive procedure. Technologies will evolve over time to improve efficiency and accessibility of AO imaging.

HVI is the most recent addition to the AO imaging techniques, and the first platform to deliver non-invasive imaging in the clinic setting under physiological conditions. This generates unique opportunities to quantify and compare AO within the EVS in many clinical scenarios. The immediate effects following selective laser trabeculoplasty⁸⁷ and short-term follow-up after TBS with iStent

Inject⁹⁰ have been documented with HVI. Other interventions can be examined in a similar way. However, the identification of patients with glaucoma or staging the disease, is far more challenging. A normative database is required to achieve this goal and move towards personalised diagnosis and treatment. Quantification of AO needs to be refined, or even automated, before the technique can be used more widely. Nevertheless, HVI has the potential to uncover important discoveries that contribute to our understanding of IOP regulation.

The AO system shares similarities with blood and lymphatic systems, both of which are regulated.⁶ IOP regulation is most probably controlled by structures within the AO system;⁶ therefore, imaging studies have concentrated on this region. Significant evidence has been amassed to support this theory including: segmental aqueous flow through TM, movement of TM in synchrony with ocular pulsation, the variable presence of pores along SC inner wall, valve-like structures permitting aqueous to enter and leave SC, AO pulsatility within episcleral veins, regional fluctuations of aqueous entering the EVS, the presence of AVAs within the EVS, and the variable re-establishment of distal AO following TBS. Despite this evidence there is no clarity regarding the influence of EVP and how the system functions in its entirety.

The contribution of aqueous production to IOP homeostasis also warrants consideration and may actively participate in the control of IOP. Theoretical feedback mechanisms between structures within the outflow system and the ciliary epithelium may exist to coordinate IOP regulation. Direct visualisation of a connection or communication between inflow and outflow systems is unlikely,

but corroborating evidence may be uncovered, just as it has been within the AO system. Mechanosensing receptors may occur within sites of AO resistance; TM^{6,11}, SC or within episcleral vessel walls. Only recently has it become possible to image aqueous outflow within the EVS, making it possible at last to demonstrate feedback control of aqueous homeostasis.

Conclusion:

Imaging is routinely relied upon to support medical diagnoses, determine treatment decisions and prognoses, and monitor progress. This stage has not yet been reached in glaucoma care largely due to an incomplete understanding of IOP dysregulation. Once that knowledge is attained then we predict that AO imaging will be used to improve patient outcomes in clinical glaucoma care. A body of knowledge has been built over a number of generations, driven by AO imaging and using a variety of imaging techniques. Each study acts as a building block for the next. Concurrent use of structural and functional imaging is likely to provide useful additional information. Non-invasive imaging techniques that assess structure and/or function of the AO pathways now exist (anterior segment OCT-A, PhS-OCT, and HVI) and they can be performed under physiological conditions. Their commercial accessibility will promote further study of the distal AO pathway. We are in the unique position of using AO imaging to discover possible mechanisms for IOP homeostasis. Many challenges remain, but the ongoing development of novel or adapted imaging technologies will continue to assist the search for these mechanisms; information that may

lead to the explanation of IOP homeostasis and elucidate the pathophysiology of primary open-angle glaucoma.

We have provided a detailed account of aqueous outflow imaging techniques and how they have contributed to our current understanding of aqueous outflow regulation. Non-invasive, physiological aqueous outflow analysis is now possible, and unlocks a new frontier in the study of IOP dysregulation in open-angle glaucoma.

Compliance with ethical standards

Conflict of interest:

There are no conflicts of interest for any author.

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References:

1. Ascher KW. The Aqueous Veins: I. Physiologic importance of the visible elimination of intraocular fluid. *Am J Ophthalmol.* 1942;25(10):1174-1209
2. Johnstone, MA. The aqueous outflow system as a mechanical pump: evidence from examination of tissue and aqueous movement in human and non-human primates. *J Glaucoma.* 2004 Oct;13(5):421-38.
3. Johnstone MA, Martin E, Jamil A. Pulsatile flow into the aqueous veins: Manifestations in normal and glaucomatous eyes. *Exper. Eye Res.* 2011;92(5), 318–327
4. Gonzalez Jr JM, Ko MK, Hong Y-K, Weigert R, and Tan JCH. Deep tissue analysis of distal aqueous drainage structures and contractile features. *Sci Rep.* 2017;7(1):1701
5. Johnstone MA. Intraocular pressure regulation: findings of pulse-dependent trabecular meshwork motion lead to unifying concepts of intraocular pressure homeostasis. *Journal of Ocular Pharmacology and Therapeutics.* 2014;30(2):88-93
6. Carreon T, van der Merwe E, Fellman R, Johnstone M, and Bhattacharya SK. Aqueous outflow – a continuum from trabecular meshwork to episcleral veins. *Prog Retin Eye Res.* 2017;57:108-133
7. Bill A and Phillips CI. Uveoscleral drainage of aqueous humor in human eyes. *Experimental Eye Research.* 1971;12:275–281
8. Huang AS, Francis BA, and Weinreb RN. Structural and functional imaging of aqueous humour outflow: a review. *Clin Experimen Ophthalmol.* 2018;46:158-168
9. Fautsch MP, Johnson DH, and Second ARVO/Pfizer Research Institute Working Group. Aqueous humor outflow: what do we know? Where will it lead us? *Invest Ophthalmol Vis Sc.* 2006;47(10):4181-4187
10. Ashton N. Anatomical study of Schlemm's canal and aqueous veins by means of neoprene casts, Part I. *Brit J Ophth.* 1951;35:291
11. Goel M, Picciani RG, Lee RK, and Bhattacharya SK. Aqueous humor dynamics: a review. *The Open Ophthalmology Journal.* 2010;4:52-59
12. Ujiie K and Bill A. The drainage routes for aqueous humor in monkeys as revealed by scanning electron microscopy of corrosion casts. *Scan Electron Microsc* 1984; (Pt 2): 849–56
13. Smit BA and Johnstone MA. Effects of viscoelastic injection into Schlemm's canal in primate and human eyes: potential relevance to viscocanalostomy. *Ophthalmology.* 2002;109:786–792.
14. Van Buskirk EM. Anatomic correlates of changing aqueous outflow facility in excised human eyes. *Invest Ophthalmol Vis Sci.* 1982;22:625– 632.
15. Overby D, Gong H, Qiu G, Freddo TF, and Johnson M. The mechanism of increasing outflow facility during washout in the bovine eye. *Invest Ophthalmol Vis Sci.* 2002;43(11): 3455-3464
16. Bentley MD, Hann CR, and Fautsch MP. Anatomical variation of human collector channel orifices. *Invest Ophthalmol Vis Sci.* 2016;57:1153-1159
17. Tamm ER. The trabecular meshwork outflow pathways: structural and functional aspects. *Experimental Eye Research.* 2009;88:648-655
18. Grant WM. Experimental aqueous perfusion in enucleated human eyes.

- Arch Ophthalmol. 1963;69:783–801.
19. Maepea O and Bill A. Pressures in the juxtacanalicular tissue and Schlemm's canal in monkeys. *Exp Eye Res.* 1992;54:879–883.
 20. Grant, W.M. Further studies on facility of flow through the trabecular meshwork. *Arch. Ophthalmol.* 60:523–533, 1958.
 21. Smit BA and Johnstone MA. Effects of viscocanalostomy on the histology of Schlemm's canal in primate eyes. *Invest Ophthalmol Vis Sci.* 2000; 41:S578.
 22. Watson AM, Rose AH, Gibson GA, Gardner CL, Sun C, Reed DS, et al. Ribbon scanning confocal for high-speed high-resolution volume imaging of brain. *PLoS One.* 2017;12:e0180486
 23. Loewen RT, Waxman S, Wang C, Atta S, Chen S, Watkins SC, and Watson AM. 3D-Reconstruction of the human conventional outflow system by ribbon scanning confocal microscopy. *PLOS ONE.* 2020;15(5): e0232833. <https://doi.org/10.1371/journal.pone.0232833>
 24. Francis AW, Kagemann L, Wollstein G, Ishikawa H, Folz S, and Overby DR. Morphometric analysis of aqueous humor outflow structures with spectral-domain optical coherence tomography. *Invest Ophthalmol Vis Sci.* 2012;53:5198–5207.
 25. Waxman S, Loewen RT, Dang Y, Watkins SC, Watson AM, and Loewen NA. High-resolution, three-dimensional reconstruction of the outflow tract demonstrates segmental differences in cleared eyes. *Invest Ophthalmol Vis Sci.* 2018;59:2371-2380
 26. Zhang J, Ren L, Mei Xi, Xu Q, Zheng W, and Liu Z. Microstructure visualization of conventional outflow pathway and finite element modeling analysis of trabecular meshwork. *BioMed Eng OnLine.* 2016;15(Suppl 2):162
 27. Li D, Zheng W, and Qu JY. Two-photon autofluorescence microscopy of multicolor excitation. *Opt Lett.* 2009;34(2):202–4.
 28. Hann CR, Bentley MD, Vercnocke A, Ritman EL, and Fautsch MP. Imaging the aqueous humor outflow pathway in human eyes by three-dimensional micro-computed tomography (3D micro-CT). *Experimental Eye Research.* 2011;92:104-111
 29. Jorgensen SM, Demirkaya O, and Ritman EL. Three-dimensional imaging of vasculature and parenchyma in intact rodent organs with X-ray micro-CT. *Am J Physiol.* 1998;275(3):H1103-H1114.
 30. Uji A, Muraoka Y, and Yoshimura N. In Vivo Identification of the Posttrabecular Aqueous Outflow Pathway Using Swept-Source Optical Coherence Tomography. *Invest Ophthalmol Vis Sci.* 2016;57:4162-9
 31. Gottschalk HM, Wecker T, Khattab MH, Fischer CV, Callizo J, Rehfeldt F, et al. Lipid emulsion-based OCT angiography for ex vivo imaging of the aqueous outflow tract. *Invest Ophthalmol Vis Sci.* 2019;60:397-406
 32. Kagemann L, Wollstein G, Ishikawa H, Bilonick RA, Brennen PM, Folio LS, et al. Identification and assessment of Schlemm's canal by spectral-domain optical coherence tomography. *Invest Ophthalmol Vis Sci.* 2010;51(8):4054-9
 33. Kagemann L, Wollstein G, Ishikawa H, Sigal IA, Folio LS, Xu J, et al. 3D visualization of aqueous humor outflow structures in-situ in humans. *Exp Eye Res.* 2011;93(3):308-315

34. Ren J, Gille HK, Wu J, and Yang C. Ex vivo optical coherence tomography imaging of collector channels with a scanning endoscopic probe. *Invest Ophthalmol Vis Sci.* 2011;52(7):3921-5
35. Li P, Reif R, Zhi Z, Martin E, Shen TT, Johnstone MA, et al. Phase-sensitive optical coherence tomography characterization of pulse-induced trabecular meshwork displacement in ex vivo nonhuman primate eyes. *J Biomed Opt.* 2012;17(7):076026
36. Asrani S, Sarunic M, Santiago C, and Izatt J. Detailed Visualization of the Anterior Segment Using Fourier- Domain Optical Coherence Tomography. *Arch Ophthalmol.* 2008 June ; 126(6): 765–771
37. Sarunic MV, Asrani S, and Izatt JA. Imaging the ocular anterior segment with real-time, full-range Fourier-domain optical coherence tomography. *Arch Ophthalmol* 2008;126(4):537–542.
38. Akagi T, Uji A, Huang AS, Weinreb RN, Yamada T, Miyata M, et al. Conjunctival and intrascleral vasculatures assessed using anterior segment optic coherence tomography angiography in normal eyes. *Am J Ophthalmol.* 2018;196:1-9
39. Shi G, Wang F, Li X, Lu J, Ding Z, Sun X, et al. Morphometric measurement of Schlemm’s canal in normal human eye using anterior segment swept source optical coherence tomography. *Journal of Biomedical Optics.* 2012;17(1):016016
40. Allingham RR, deKater AW, and Ethier CR. Schlemm’s canal and primary open angle glaucoma: correlation between Schlemm’s canal dimensions and outflow facility. *Exp. Eye Res.* 1996;62(1): 101–109
41. Chen Z, Sun J, Li M, Liu S, Chen L, Jing S, et al. Effect of age on the morphologies of the human Schlemm’s canal and trabecular meshwork measured with swept-source optical coherence tomography. *Eye.* 2018;32:1621-1628
42. Yoshikawa M, Akagi T, Uji A, Nakanishi H, Kameda T, Suda K, et al. Pilot study assessing the structural changes in posttrabecular aqueous humor outflow pathway after trabecular meshwork surgery using swept-source optical coherence tomography. 2018. *PLoS ONE*; 13(6):e0199739
43. Wang B, Kagemann L, Schuman JS, Ishikawa H, Bilonick RA, Ling Y, et al. Gold nanorods as a contrast agent for Doppler optical coherence tomography. *PLoS One.* 2014;9:e90690.
44. Gao SS, Jia Y, Zhang M, Su JP, Liu G, Hwang TS, et al. Optical coherence tomography angiography. *Invest Ophthalmol Vis Sci.* 2016;57:OCT27–OCT36.
45. Chen J, Huang H, Zhang S, Chen X, and Sun X. Expansion of Schlemm’s Canal by Travoprost in Healthy Subjects Determined by Fourier-Domain Optical Coherence Tomography. *Invest Ophthalmol Vis Sci.* 2013;54(2):1127-1134
46. Grieshaber MC, Pienaar A, Olivier J, and Stegmann R. Clinical evaluation of the aqueous outflow system in primary open-angle glaucoma for canaloplasty. *Invest Ophthalmol Vis Sci.* 2010; 51(3): 1498-1504
47. Rosenquist R, Epstein D, Melamed S, Johnson M, and Grant WM. Outflow resistance of enucleated human eyes at two different perfusion pressures and different extents of trabeculotomy. *Curr Eye Res.* 1989;8:1233–1240.
48. Maurice DM. A new objective fluorophotometer. *Exp Eye Res.* 1963;2:33-

49. Langham M and Wybar KC. Fluorophotometric apparatus for the objective determination of fluorescence in the anterior chamber of the living eye. *Br J Ophthalmol*. 1954;38:52-57
50. Jones R and Maurice D. New methods of measuring the rate of aqueous flow in man with fluorescein. *Exp Eye Res* 1966;5:208–220
51. Nagataki S. Aqueous humor dynamics of human eyes as studied using fluorescein. *Jpn J Ophthalmol*. 1975;19:235
52. Brubaker RF. Flow of aqueous humor in humans [The Friedenwald Lecture]. *Invest Ophthalmol Vis Sci*. 1991;32(13):3145–3166.
53. Maus TL and Brubaker RF. Measurement of aqueous humor flow by fluorophotometry in the presence of a dilated pupil. *Invest Ophthalmol Vis Sci*. 1999;40(2):542-6
54. Hann CR, Bahler CK, and Johnson DH. Cationic ferritin and segmental flow through the trabecular meshwork. *Invest Ophthalmol Vis Sci*. 2005;46:1-7
55. Sabanay I, Gabelt BT, Tian B, Kaufman PL, and Geiger B. H-7 effects on the structure and fluid conductance of monkey trabecular meshwork. *Arch Ophthalmol*. 2000;118:955-962
56. Ren R, Li G, Le TD, Kopczynski C, Stamer WD, and Gong H. Netarsudil increases outflow facility in human eyes through multiple mechanisms. *Invest Ophthalmol Vis Sci*. 2016;57:6197-6209
57. Battista SA, Lu Z, Hofmann S, Freddo T, Overby DR, and Gong H. Reduction of the Available Area for Aqueous Humor Outflow and Increase in Meshwork Herniations into Collector Channels Following Acute IOP Elevation in Bovine Eyes. *Invest Ophthalmol Vis Sci*. 2008;49(12):5346-5352
58. Braakman ST, Read AT, Chan D W-H, Ethier CR, and Overby DR. Colocalization of outflow segmentation and pores along the inner wall of Schlemm's canal. *Exp Eye Res*. 2015;130:87-96
59. Keller KE, Bradley JM, Vranka JA, and Acott TS. Segmental versican expression in the trabecular meshwork and involvement in outflow facility. *Invest Ophthalmol Vis Sci*. 2011;52(8):5049-5057
60. Karg SJ, Garron LK, Feeney ML, and McEwen WK. Perfusion of human eyes with latex microspheres. *Arch Ophthalmol*. 1959;61:68–71.
61. Hann CR and Fautsch MP. Preferential fluid flow in the human trabecular meshwork near collector channels. *Invest Ophthalmol Vis Sci*. 2009;50:1692–1697.
62. De Kater AW, Melamed S, and Epstein DL. Patterns of aqueous humor outflow in glaucomatous and non-glaucomatous human eyes. A tracer study using cationized ferritin. *Arch Ophthalmol*. 1989;107(4):572-6
63. Loewen RT, Brown EN, Roy P, Schuman JS, Sigal IA, and Loewen NA. Regionally discrete aqueous humor outflow quantification using fluorescein canalograms. *PLoS ONE*. 2016;11(3):e0151754
doi:10.1371/journal.pone.0151754
64. Loewen RT, Brown EN, Scott G, Parikh H, Schuman JS, and Loewen NA. Quantification of focal outflow enhancement using differential canalograms. *Invest Ophthalmol Vis Sci*. 2016;57:2831-2838

65. Parikh HA, Loewen RT, Roy P, Schuman JS, Lathrop KL, and Loewen NA. Differential Canalograms Detect Outflow Changes from Trabecular Micro-Bypass Stents and Ab Interno Trabeculectomy. *Sci Rep*. 2016;6:34705.
66. Mendrinós E, Mermoud A, and Shaarawy T. Nonpenetrating glaucoma surgery. *Surv Ophthalmol*. 2008;53(6):592-630
67. Grieshaber MC. Ab externo Schlemm's canal surgery: viscocanalostomy and canaloplasty. *Dev Ophthalmol*. 2012;50:109-124
68. Zeppa L, Ambrose L, Guerra G, Fortunato M, and Costagliola G. *In vivo* near-infrared fluorescence imaging of aqueous humor outflow structures. *Journal of ophthalmology*. 2016;2016:8706564. doi:10.1155/2016/8706564
69. Fellman RL and Grover DS. Episcleral venous fluid wave: intraoperative evidence of patency of the conventional outflow system. *J Glaucoma*. 2014;23:347-350
70. Fellman RL, Feuer WJ, and Grover DS. Episcleral venous fluid wave correlates with trabectome outcomes: intraoperative evaluation of the trabecular outflow pathway. *Ophthalmology*. 2015;122(12):2385-2392
71. Fellman RL and Grover DS. Episcleral Venous Fluid Wave in the Living Human Eye Adjacent to Microinvasive Glaucoma Surgery (MIGS) Supports Laboratory Research: Outflow is Limited Circumferentially, Conserved Distally, and Favored Inferonasally. *J Glaucoma*. 2019;28(2):139-145
72. Laroche D, Nortey A, and Ng C. A novel use of trypan blue during canalicular glaucoma surgery to identify aqueous outflow to episcleral and intrascleral veins. *J Glaucoma*. 2018;27:e158-e161
73. Saraswathy S, Tan JCH, Yu F, Francis BA, Hinton DR, Weinreb RN, et al. Aqueous angiography: real-time and physiologic aqueous humor outflow imaging. *PLoS One* 2016; 11: e0147176
74. Huang AS, Saraswathy S, Dastiridou A, Begian A, Legaspi H, Mohindroo C, et al. Aqueous angiography with fluorescein and indocyanine green in bovine eyes. *Transl Vis Sci Technol* 2016; 5: 5. [DOI](#)
75. Huang AS, Saraswathy S, Dastiridou A, Begian A, Mohindroo C, Tan JCH, et al. Aqueous angiography-mediated guidance of trabecular bypass improves angiographic outflow in human enucleated eyes. *Invest Ophthalmol Vis Sci* 2016; 57: 4558–65. [DOI](#)
76. Huang AS, Li M, Yang D, Wang H, Wang N, and Weinreb RN. Aqueous angiography in living nonhuman primates shows segmental, pulsatile, and dynamic angiographic aqueous humor outflow. *Ophthalmology* 2017; 124: 793–803. [DOI](#)
77. Huang AS, Camp A, BY X, Penteadó RC, and Weinreb RN. Aqueous angiography: aqueous humor outflow imaging in live human subjects. *Ophthalmology* 2017; 124: 1249–51. [DOI](#)
78. Huang A, Penteadó RC, Papoyan V, Voskanyan L, and Weinreb RN. Aqueous angiographic outflow improvement after trabecular microbypass in glaucoma patients. *Ophthalmol Glaucoma*. 2019;2(1):11-21
79. Huang AS, Penteadó RC, Saha SK, Do JL, Ngai P, Hu Z, et al. Fluorescein aqueous angiography in live normal human eyes. *J Glaucoma*. 2018;27(11):957-964

80. Ascher KW. Glaucoma and the aqueous veins. *Am J Ophthalmol* 1942;25:11:1309-15.
81. Johnstone M, Stegmann R, Martin E, and Jamil A. Pulsatile Circumferential Aqueous Flow Into Schlemm's Canal (SC) Is Synchronous With the Cardiac Pulse. *Invest. Ophthalmol. Vis. Sci.* 2009;50(13):4855
82. Li P, Shen TT, Johnstone M, and Wang RK. Pulsatile motion of the trabecular meshwork in healthy human subjects quantified by phase-sensitive optical coherence tomography. *Biomed Opt Express.* 2013;4(10):2051-65
83. Wang RK, Ma Z, and Kirkpatrick SJ. Tissue Doppler optical coherence elastography for real time strain rate and strain mapping of soft tissue. *Appl Phys. Lett.* 2006;89:144103
84. Wang RK, Kirkpatrick SJ, and Hinds M. Phase-sensitive optical coherence elastography for mapping tissue microstrains in real time, *Appl Phys. Lett.* 2007;90(16): 164105
85. Johnstone MA. The aqueous outflow system as a mechanical pump: Evidence from examination of tissue and aqueous movement in human and non-human primates. *J Glaucoma* 2004;13:5:421-38.
86. Meyer PA. The circulation of the human limbus. *Eye* 1989;3:121-127.
87. Khatib TZ, Meyer PA, Lusthaus JA, Manyakin I, Mushtaq Y, and Martin KR. Haemoglobin Video Imaging provides novel in vivo high-resolution imaging and quantification of human aqueous outflow in glaucoma patients. *Ophthalmology Glaucoma.* 2019;2(5):327-335
88. Meyer PA. Re-orchestration of blood flow by micro-circulations. *Eye.* 2018;32:222-229
89. Meyer PA. Anterior segment vascular imaging. In: Easty DL, Sparrow JM(eds) *Oxford Textbook of Ophthalmology.* Oxford University Press: Oxford, UK, 1999 pp 160-174.
90. Lusthaus JA, Meyer PAR, Khatib TZ, and Martin KR. The effects of trabecular bypass surgery on conventional aqueous outflow, visualized by Hemoglobin Video Imaging. *J Glaucoma.* May 2020 accepted for publication.
91. Cha EDK, Xu Jia, Gong L, and Gong H. Variations in active outflow along the trabecular outflow pathway. *Exp Eye Res.* 2016;146:354-360
92. Johnson M. What controls aqueous humour outflow resistance? *Exp Eye Res.* 2006;82(4):545-557
93. Overby DR, Stamer DW, and Johnson M. The changing paradigm of outflow resistance generation: towards synergistic models of the JCT and inner wall endothelium. *Exp Eye Res.* 2009;88(4):656-670
94. Johnson M, Shapiro A, Ethier CR, and Kamm RD. Modulation of outflow resistance by the pores of the inner wall endothelium. *Invest Ophthalmol Vis Sci.* 1992;33:1670 -1675.
95. Ethier C and Chan D. Cationic ferritin changes outflow facility whereas anionic ferritin does not. *Invest Ophthalmol Vis Sci.* 2001; 42:1795-1802.
96. Stamer WD and Acott TS. Current understanding of conventional outflow dysfunction in glaucoma. *Curr Opin Ophthalmol.* 2012;23(2):135-143
97. Wang K, Johnstone MA, Xin C, Song S, Padilla S, Vranka JA, et al. Estimating human trabecular meshwork stiffness by numerical modeling and advanced OCT imaging. *Invest Ophthalmol Vis Sci.* 2017 Oct; 58(11):

4809–4817

98. Xin C, Song S, Johnstone M, Wang N, and Wang RK. Quantification of pulse-dependent trabecular meshwork motion in normal humans using phase-sensitive OCT. *Invest Ophthalmol Vis Sci.* 2018;59(8):3675-81
99. Kagemann L, Wang B, Wollstein G, Ishikawa H, Nevins JE, Nadler Z, et al. IOP elevation reduces Schlemm's canal cross-sectional area. *Invest Ophthalmol Vis Sci.* 2014;55:1805-9
100. Johnstone MA and Grant WG. Pressure-dependent changes in structures of the aqueous outflow system of human and monkey eyes. *Am J Ophthalmol.* 1973;75:365–383.
101. Gulati V, Fan S, Hays CL, Samuelson TW, Ahmed IIK, and Toris CB. A novel 8-mm Schlemm's canal scaffold reduces outflow resistance in a human anterior segment perfusion model. *Invest Ophthalmol Vis Sci.* 2013;54:1698–1704.
102. Selbach JM, Rohen JW, Steuhl KP, and Lutjen-Drecoll E. Angioarchitecture and innervation of the primate anterior episclera. *Curr Eye Res.* 2005;30:337–344. [↗](#)
103. Sit AJ and McLaren JW. Measurement of episcleral venous pressure. *Experimental Eye Research.* 2011;93:291-298
104. Lee SS, Robinson MR, and Weinreb RN. Episcleral venous pressure and the ocular hypotensive effects of topical and intracameral prostaglandin analogs. *J Glaucoma.* 2019;28(9):846-857

Figure and Table Legends:

Figure 1. A: Microscopic photograph of neoprene cast representing Schlemm's canal and associated distal aqueous outflow system. **B:** Drawing of neoprene cast seen in **A** with green vessels representing the deep scleral plexus and suture loops identifying aqueous veins. Both figures reproduced from Anatomical study of Schlemm's canal and aqueous veins by means of neoprene casts, Ashton N, 35, 291-303, ©1951 with permission from BMJ Publishing Group Ltd.

Figure 2. Scanning electron microscopy image demonstrating the outflow pathway of aqueous (blue arrows) in a primate eye. Anterior chamber (AC), trabecular meshwork (TM), intrascleral collector channels (ISCC), episcleral aqueous vein (AV-E), and conjunctival aqueous vein (AV-C). Image reproduced with permission from Johnstone, MA. Intraocular pressure control through linked trabecular meshwork and collector channel motion. In: Samples, JR., Knepper, PA, editors. Glaucoma Research and Clinical Advances: 2016 to 2018. Kugler Publications; Amsterdam ©2016.

Figure 3. Three-dimensional reconstructions of the human aqueous outflow system using multiple structural imaging techniques. **A:** Ribbon-scanning confocal microscopy (collector channels in blue). **B:** 3D micro-CT reconstruction demonstrating Schlemm's canal (purple ring) with narrowing superiorly in a perfused eye. Black arrows represent sites of collector channels evenly distributed throughout the circumference of the eye. **C:** Virtual casting reconstruction of collector channels from an actively perfused in situ human eye imaged with SD-OCT. **D:** OCT-A reconstructed image of merged superficial (green) and deep (red) vascular networks. Aqueous carrying vessels are not identifiable. (A) Adapted image reprinted from Loewen RT, et al²² with permission under the terms of the CC BY license (<https://creativecommons.org/licenses/by/4.0/>). (B) Adapted image reprinted from Experimental Eye Research, 92, Hann CR, et al, Imaging the aqueous humor outflow pathway in human eyes by three-dimensional micro-computed tomography (3D micro-CT), 104-111, ©2011, with permission from Elsevier. (C) Adapted image reprinted from Experimental Eye Research, 93(3), Kagemann L, et al, 3D visualization of aqueous humor outflow structures in-situ in human eyes, 308-315, ©2011, with permission from Elsevier. (D) Adapted image reprinted from Am J Ophthalmol, 196, Akagi et al, Conjunctival and intrascleral vasculatures assessed using anterior segment optical coherence tomography in normal eyes, 1-9, ©2018, with permission from Elsevier.

Figure 4. A: Swept-source optical coherence tomography (SS-OCT) of the human drainage angle. Trabecular meshwork and Schlemm's canal (black arrow) are pictured within the magnified box. **B:** A single B-scan demonstrating the pathway of a post-trabecular aqueous outflow vessel (red arrowheads) emanating from Schlemm's canal (yellow arrowhead). The image was reconstructed using SS-OCT with flattening, *en face* imaging and reslicing techniques.³⁰ (A) Adapted image reprinted from Chen Z et al⁴¹ with permission under the terms of the CC BY license (<https://creativecommons.org/licenses/by/4.0/>). (B) Adapted image republished with permission of Association for Research in Vision &

Ophthalmology (ARVO) from In Vivo Identification of the Posttrabecular Aqueous Outflow Pathway Using Swept-Source Optical Coherence Tomography. Uji A et al³⁰, 57, ©2016

Figure 5. Fluorescein canalogram taken from a perfused whole porcine eye. Adapted image reprinted from Loewen RT, et al⁶³ with permission under the terms of the CC BY license (<https://creativecommons.org/licenses/by/4.0/>).

Figure 6. Intraoperative techniques to visualise distal aqueous drainage and immediately confirm surgical success. **A:** Fluorescein-filled episcleral vessels after injection directly into Schlemm's canal. **B:** Aqueous angiography demonstrating increasing signal in the supero-nasal quadrant during cataract surgery. Images are taken 176 seconds (B1) and 182 (B2) seconds after introduction of 0.04% indocyanine green. (A) Reprinted by permission from Springer Nature. *Ophthalmologe*. Grieshaber MC. Kanalographie und Wirkmechanismus der Kanaloplastik [Channelography and mechanism of action in canaloplasty]. 112(4):319-324 ©2015. doi:10.1007/s00347-014-3163-4 (B) Adapted image reprinted from Huang AS, et al, Structural and functional imaging of aqueous humour outflow: a review. *Clin Experimen Ophthalmol*, 46:158-168, ©2018 with permission from John Wiley and Sons.

Figure 7. A, B: Anterior segment microcirculations. eac, episcleral arterial circle; lvc, limbal venous circle; ecv, episcleral collecting vein; ca, conjunctival arteriole; cv, conjunctival venule; cap, capillary. **B:** The black arrows demonstrate blood striation and the white arrows identify an aqueous column.

Figure 8. Two examples in which blood and aqueous drain into a common vessel to create an asymmetrical flow pattern. The points where blood and aqueous combine are marked with white stars. The direction of distal aqueous flow is indicated by the white arrows.

Figure 9. A: Diagrammatic representation of aqueous column measurement from light intensity transepts. **B and C:** An aqueous vein is pictured with a linear transept cutting across the vessel. The transept and corresponding graph are generated by Image J software. The aqueous column diameter equals the distance between the troughs (vertical blue arrows). The diameter in pixels is converted to aqueous column cross-sectional area (AqCA) in micrometres squared.

Table 1. Imaging techniques and their contributions to aqueous outflow knowledge

AO = aqueous outflow

SC = Schlemm's canal

JCT = Juxtacanalicular tissue

TM = Trabecular meshwork

OCT = Optical coherence tomography

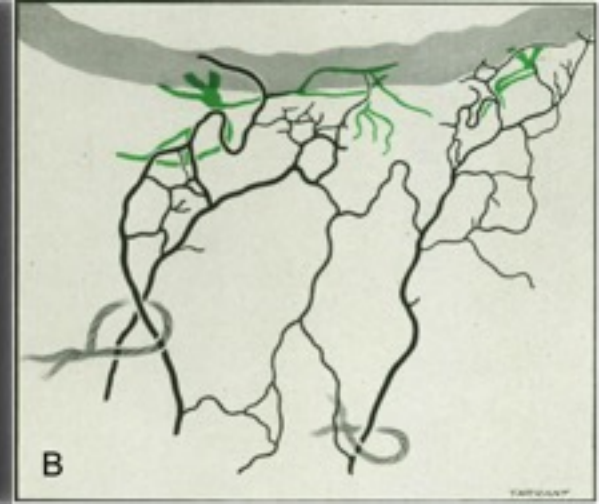
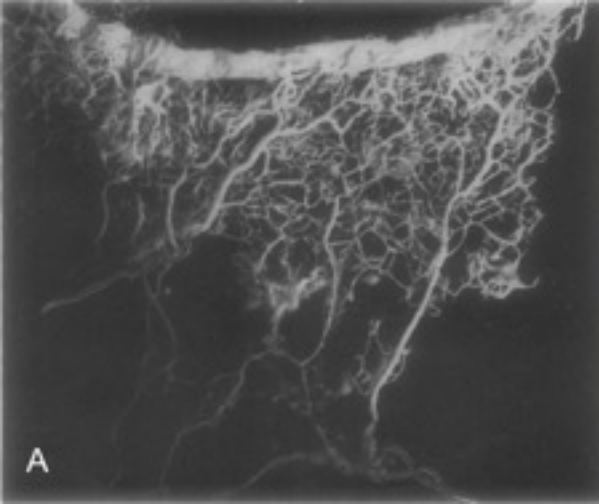
IOP = Intraocular pressure

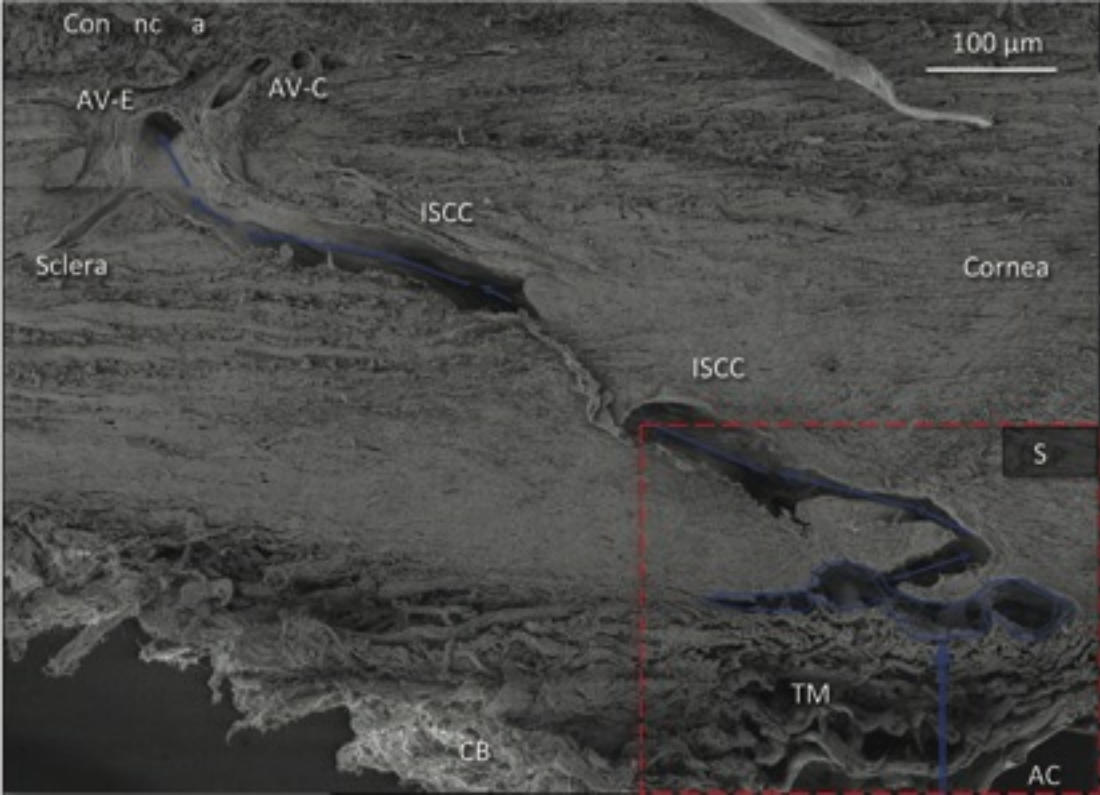
EVFW = Episcleral venous fluid wave

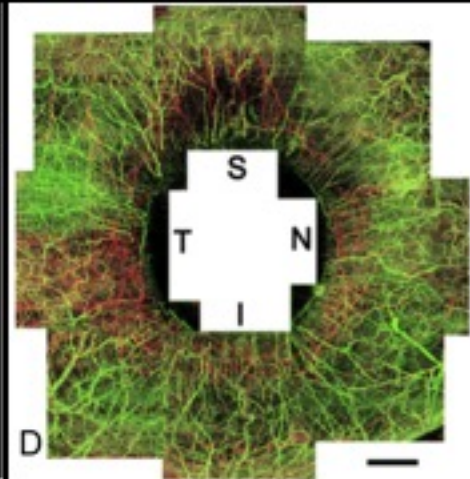
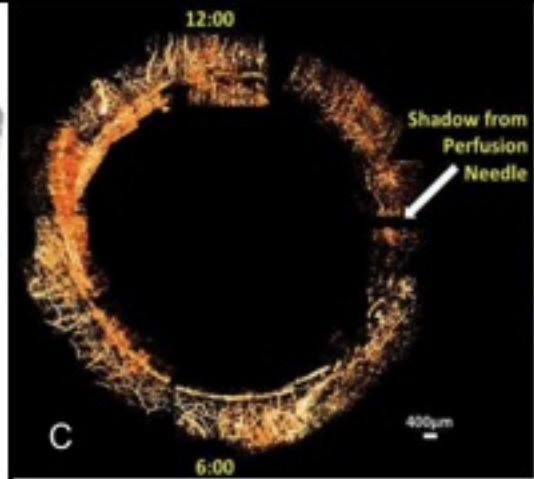
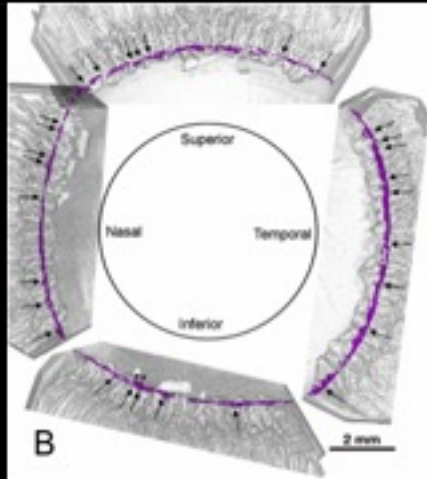
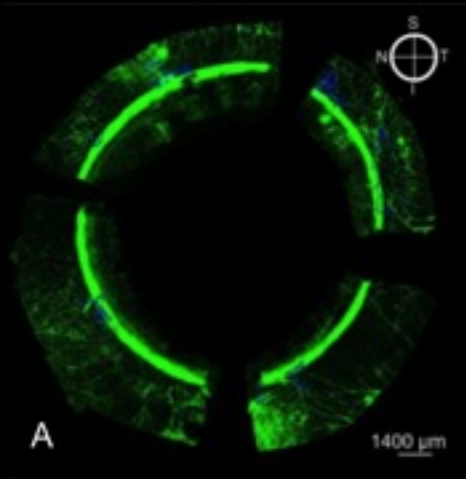
ICG = Indocyanine green

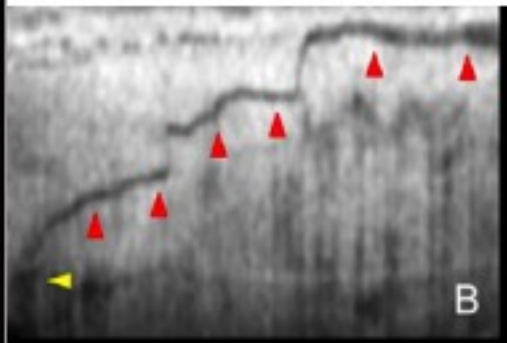
TBS = Trabecular bypass surgery

3D micro-CT = Three-dimensional micro-computed tomography



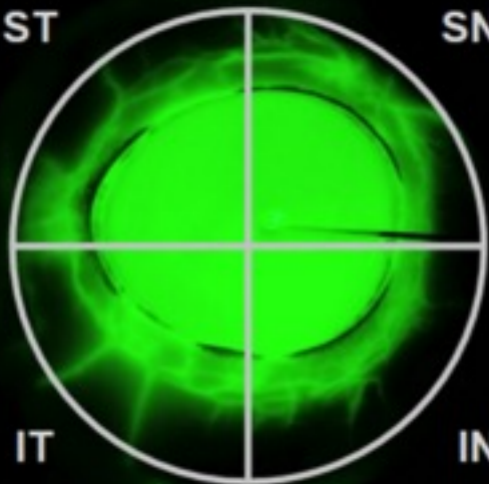






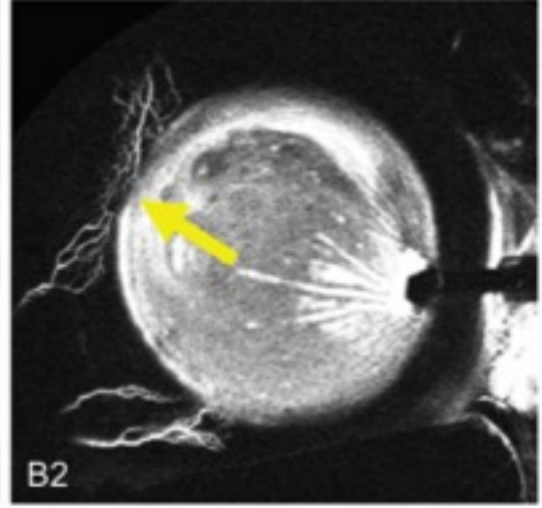
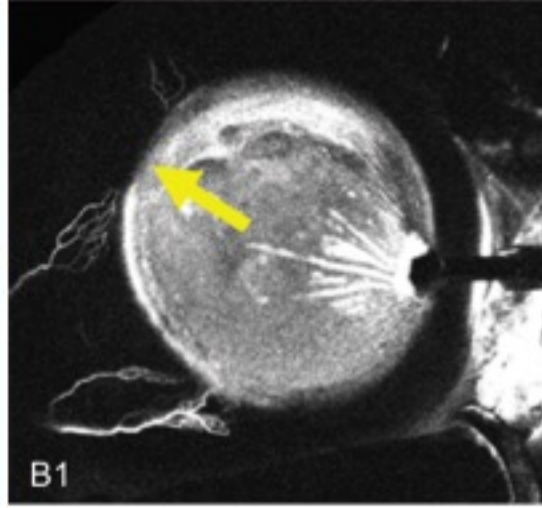
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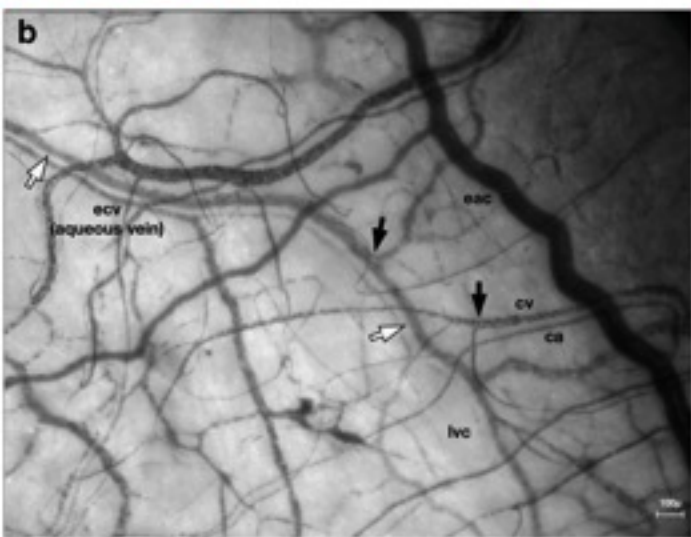
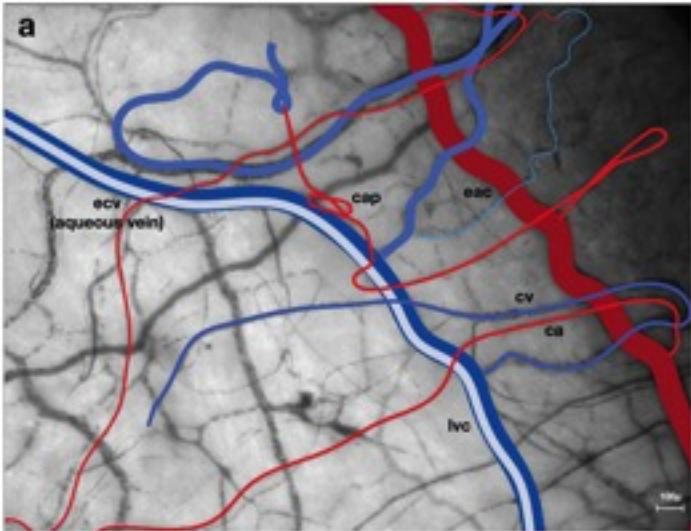
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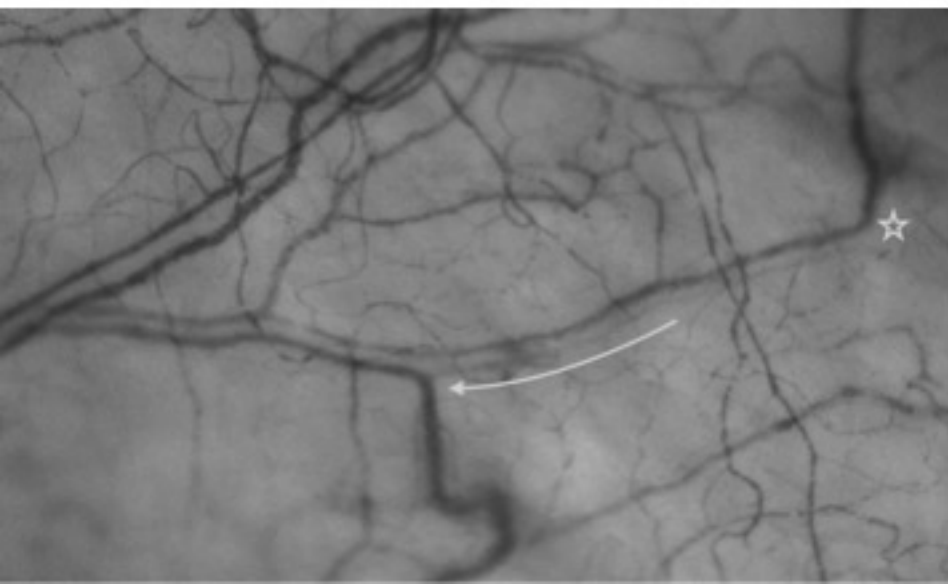
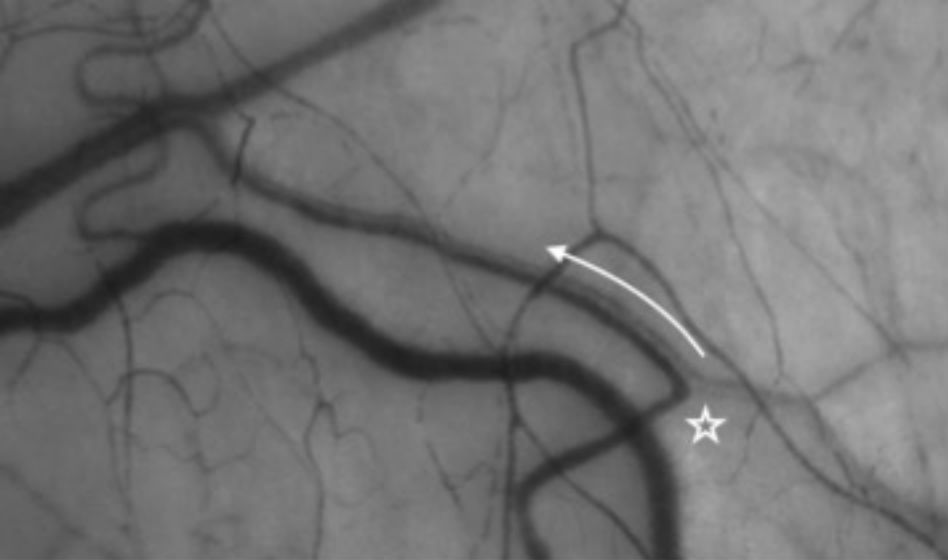


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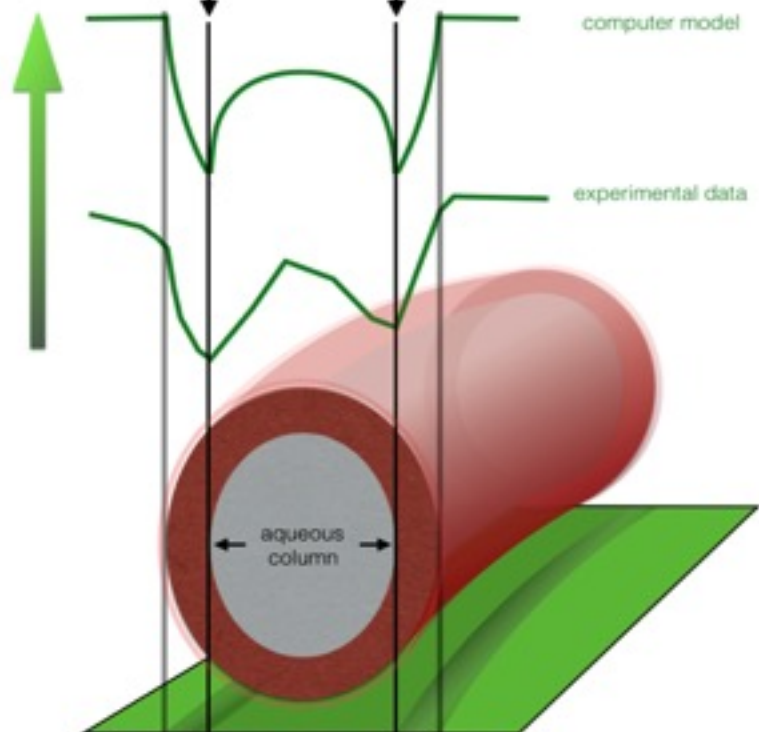




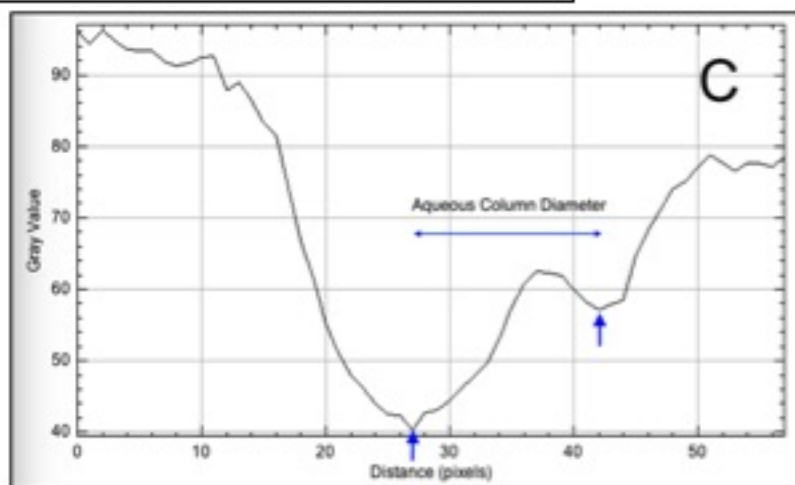
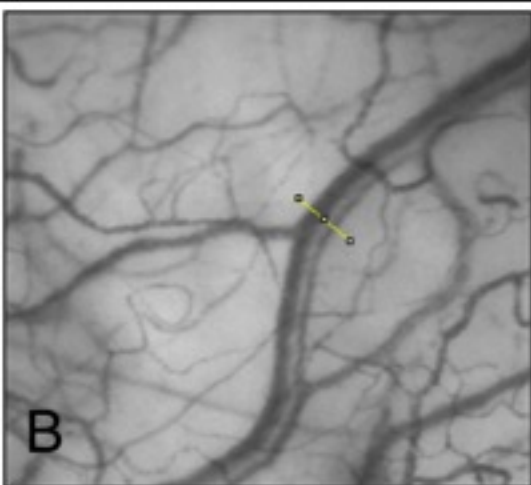


Intensity transects

(light transmitted by blood and aqueous;
reflected by sclera)



A



<i>Imaging Technique</i>	<i>Specimen Type</i>	<i>Specimen Condition</i>	<i>Tissue Preparation</i>	<i>Imaging Condition</i>	<i>Classification of Findings</i>	<i>Significant Findings</i>	<i>References</i>
Photography and sketch	Human	<i>Ex vivo</i>	Neoprene casts	Static	Structural	Defined AO anatomy	Ashton ¹⁰
Light and electron microscopy	Primate and human	<i>Ex vivo</i>	Corrosion casts, perfusion fixed, viscoelastic injection	Static	Structural	Confirmed anatomy of proximal AO system	Ujiie et al ¹² Smit et al ¹³ Van Buskirk ¹⁴
	Bovine	<i>Ex vivo</i>	Controlled AC tracer perfusion for 3 hours	Static	Structural and functional	Adhesive tissue between JCT and SC inner wall helps regulate AO resistance	Overby et al ¹⁵
	Human	<i>Ex vivo</i>	Perfusion fixed	Static	Structural	Variations in collector channel orifices may assist AO regulation	Bentley et al ¹⁶
	Human	<i>Ex vivo</i>	Tracer perfusion and histological tissue preparation	Static	Structural and functional	Segmental labeling of TM suggests preferential AO ^{54,58,59,61,93} Thickening of TM and a higher number of collector channels in regions of active outflow ⁹¹	Hann et al ⁵⁴ Braakman et al ⁵⁸ Keller et al ⁵⁹ Hann et al ⁶¹ Cha et al ⁹¹ Overby et al ⁹³
Dissecting microscope with fluorescent imaging capabilities	Porcine	<i>Ex vivo</i>	Fluorescein tracer introduction with canalography	Static	Functional	Perilimbal fluorescence is greatest in nasal quadrant ^{63,64} Fluorescence increased after trabectome ⁶⁵	Loewen et al ⁶³ Loewen et al ⁶⁴ Parikh et al ⁶⁵

Two-photon microscopy	Rodent	<i>Ex vivo</i>	Specimen stabilisation	Static	Structural	TM lacked smooth muscle, but structure of veins supports compliance	Gonzalez et al ⁴ Zhang et al ²⁶
3D micro-CT	Human	<i>Ex vivo</i>	Immersion or perfusion fixation	Static	Structural	Collector channels are unevenly dispersed around the circumference of the eye	Hann et al ²⁸ Jorgensen et al ²⁹
OCT	Spectral-Domain OCT	Human	<i>In vivo</i>	Nil	Static	SC is larger in the nasal quadrant. ³² Manipulations of IOP and SC affect one another. ⁹⁹⁻¹⁰¹	Kagemann et al ³² Kagemann et al ⁹⁹ Johnstone et al ¹⁰⁰ Gulati et al ¹⁰¹
	Swept-Source OCT		<i>In vivo</i>	Nil	Static	Enabled full-depth imaging of anterior segment. ^{36,37} SC reduces in size with age. ⁴¹ SC is widest in nasal ³⁹⁻⁴¹ and temporal quadrants. ⁴¹	Asrani et al ³⁶ Sarunic et al ³⁷ Shi et al ³⁹ Allingham et al ⁴⁰ Chen et al ⁴¹
	OCT Angiography	Human	<i>In vivo</i>	Nil	Static	The presence of blood is required to identify vessels. ^{31,38} Deep vasculature was more detailed in the nasal	Gottschalk et al ³¹ Akagi et al ³⁸ Gao et al ⁴⁴

							and temporal quadrants. ³⁸	
	Phase-sensitive OCT	Primate and human	<i>Ex vivo</i> and <i>in vivo</i>	Nil	Dynamic	Functional	TM motion occurs in synchrony with the ocular pulse. ^{3,35,82,97,98} Artificial IOP elevation reduced TM movement. ³⁵	Li et al ³⁵ Li et al ⁸² Johnstone et al ³ Wang et al ⁹⁷ Xin et al ⁹⁸
Operating microscope		Human	<i>In vivo</i>	Fluorescein injected into SC (channelography)	Dynamic	Functional	Poor diffusion of fluorescein may indicate SC collapse	Grieshaber et al ⁴⁶
		Human	<i>In vivo</i>	EVFW – Sudden intraoperative release of balanced salt solution	Dynamic	Functional	Presence of EVFW may indicate the AO system can accept aqueous	Fellman et al ⁶⁹⁻⁷¹
		Human	<i>In vivo</i>	Trypan blue injection following excisional goniotomy	Dynamic	Functional	Highlights EVFW	Laroche et al ⁷²
Near-infrared camera		Human	<i>In vivo</i>	ICG injected into SC	Dynamic	Functional	Flow in SC is segmental Unexplained retrograde filling patterns in SC	Zeppa et al ⁶⁸
Aqueous angiography		Porcine, bovine, non-human primate,	<i>Ex vivo</i> and <i>in vivo</i>	Intracameral injection of fluorescein or ICG	Dynamic	Functional	AO is dynamic ⁷⁸ , pulsatile ^{76,77} and predominantly nasal ⁷⁵⁻⁷⁷ Areas of poor	Sarawathy et al ⁷³ Huang et al ⁷⁵ Huang et al ⁷⁶ Huang et al ⁷⁷

	human					angiographic signal can be recovered with TBS ^{77,78}	Huang et al ⁷⁸
Haemoglobin Video Imaging	Human	<i>In vivo</i>	Nil	Dynamic and physiological	Functional	AO can be detected non-invasively ⁸⁸ AO is pulsatile ^{87,90} and predominantly nasal ⁹⁰ AO fluctuates following TBS ⁹⁰	Khatib et al ⁸⁷ Meyer ⁸⁸ Lusthaus et al ⁹⁰