

Article ID: 8923
DOI: 10.5586/asbp.8923

Publication History
Received: 2019-03-30
Accepted: 2020-04-17
Published: 2020-06-24

Handling Editor
Grażyna Kłobus; University of Wrocław, Poland;
<https://orcid.org/0000-0001-9337-0745>

Authors' Contributions
JCB and JD participated in the design of the research; MF and IZ carried out the measurement of Cd content; RH and JCB carried out the experiments concerning seeds physiological state; JCB and SI conducted measurements of germination rate and seedlings growth; JCB, MF, IZ, and JD analyzed the data; JCB wrote the manuscript





Funding
The study was financed by a grant of the Plenipotentiary Representative of Poland in the Joint Institute of Nuclear Research (JINR) in Dubna, Russia (topics 03-4-1104-2011/2016 and 03-4-1128-2017/2019, ordinance No. 389 of 07/06/2017, p. 28).

Competing Interests
No competing interests have been declared.

Copyright Notice
© The Author(s) 2020. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits redistribution, commercial and noncommercial, provided that the article is properly cited.

ORIGINAL RESEARCH PAPER in PHYSIOLOGY AND BIOCHEMISTRY

Tough Sprouting – Impact of Cadmium on Physiological State and Germination Rate of Soybean Seeds

Jagna Chmielowska-Bąk ^{1*}, Renata Holubek²,
Marina Frontasyeva ³, Inga Zinicovscaia ^{3,4}, Selin İşıdoğru⁵,
Joanna Deckert ¹

¹ Department of Plant Ecophysiology, Institute of Experimental Biology, Faculty of Biology, Adam Mickiewicz University in Poznań, Poland

² Department of Zoology and Anthropology, Faculty of Natural Sciences, Constantine the Philosopher University, Slovakia

³ Frank Laboratory of Neutron Physics, Division of Nuclear Physics, Department of Neutron Activation Analysis and Applied Research, Joint Institute for Nuclear Research, Russia

⁴ Horia Hulubei National Institute for R&D in Physics and Nuclear Engineering, MG-6, Bucharest – Magurele, Romania

⁵ Bülent Ecevit University, Turkey

*To whom correspondence should be addressed. Email: jagna.chmielowska@amu.edu.pl

Abstract

Seed germination is the earliest process in plant development and is crucial for further plant growth and fitness. The process is regulated by various internal and external factors, including soil pollutants such as nonessential metals. In the present study, we examined in detail the impact of short-term imbibition in Cd solutions at several concentrations (5, 10, and 25 mg/L) on germination rate and physiological state of soybean seeds. The results showed that although Cd was readily absorbed by the seeds, the metal had no effect on seeds cell viability, oxidative stress intensity, or germination percentage. In contrast, imbibition in Cd solution led to slight reduction in antioxidant capacity of seeds. Seedlings grown from seeds pretreated with metal showed no differences in growth in relation to the control. Taken together, the results indicate that soybean seeds are relatively tolerant even to high Cd concentration (up to 25 mg/L).

Keywords

metal stress; antioxidants; cell viability; lipid peroxidation

1. Introduction

Seed germination is the earliest and one of the most important processes in plant development. Its proper timing is crucial for future plant fitness and survival. Premature or delayed germination might lead to disturbances in development processes as well as to the exposure to stress conditions and decreased chances in intra- and interspecies competition for environmental resources. The germination process is divided into three phases of which the first phase is characterized by significant water uptake during the process of imbibition accompanied by early activation of metabolism and the second phase comprises mobilization of storage material, enhanced transcription, translation, and respiration, and increased growth of embryo cells. The final third phase includes further water uptake, seedlings growth, and final rupture of the seed coat and radical emergence (Wietbrecht et al., 2011). The course of the germination process is regulated by several internal and external factors. Internal regulation is mediated by plant hormones, namely abscisic acid, ethylene, auxins, gibberellins, cytokinins, and brassinosteroids and other signaling molecules including reactive oxygen species (ROS) and NO (Arc et al., 2013; Miransari & Smith, 2014; Shu et al., 2016; Wojtyła et al., 2016).

In the case of external factors, the main germination modulators include water status and light and temperature conditions (Donohue, 2005; Finch-Savage & Leubner-Metzger, 2006). However, the process might be similarly affected by environmental contaminants. Several studies have shown that exposure to metals such as Ni, Pb, Cd, and Cr decrease germination rate and/or hamper growth of young seedlings. The inhibitory effect is associated with disturbances in water uptake, oxidative stress, decreased level and activity of proteins, changes in membrane permeability, and DNA damage (Kranner & Colville, 2011; Sethy & Ghosh, 2013). In contrast, there are reports showing the stimulatory effect of metals at low concentrations on germination of several plant species (Ilić et al., 2015; Kavuličova et al., 2012; Lefèvre et al., 2009). This phenomenon might be explained by hormesis – stimulation of growth by toxic ions at low concentration through, among other processes, perturbation in signaling network and activation of defense mechanisms (Poschenrieder et al., 2013). One of the important metal contaminants is Cd, which has been detected at elevated levels in soil samples from various regions of the world (Khan et al., 2011, 2017; Wang et al., 2015). This highly mobile element is readily absorbed by plants leading to the development of severe toxicity symptoms including growth inhibition, disturbances in photosynthesis, and damage of proteins, membranes, and nucleic acids (Gallego et al., 2012; Khan et al., 2017; Tran & Popova, 2013). Absorption of Cd by crops poses a threat to human health, as this metal is considered a class I carcinogen and causes alterations in structure and functioning of bones, kidneys, and lungs (Khan et al., 2017; Rizwan et al., 2017). The aim of the present study was to examine in detail the impact of Cd on the physiological state of soybean seeds during the earliest germination phase. The effect on seed cell viability, membrane integrity, intensity of oxidative stress, antioxidant properties, and germination rate are examined and discussed.

2. Material and Methods

2.1. Seeds Treatment and Growth Procedures

Soybean (*Glycine max* L. 'Naviko') seeds, kindly supplied by the Department of Genetics and Plant Breeding, University of Life Sciences in Poznań, Poland, were surface sterilized with 75% ethanol (5 min) and 1% sodium hypochlorite (10 min) and washed for 30 min under running water. Next, for every experimental treatment, 50 seeds were imbibed for 2 hr in 30 mL of distilled water (control) or CdCl₂ (P.O.Ch.; Gliwice, Poland) with Cd at the concentration 5, 10, or 25 mg/L. After 2 hr, the seeds were washed thoroughly under running water and used for further analysis.

2.2. Measurement of Cd Uptake

Cd uptake by seeds was traced using atomic absorption spectrometry (AAS) (iCe 3000 series; Thermo Scientific, Waltham, Massachusetts, U.S.) with electrothermal atomization. AAS-Cd standard solution with a concentration of 1,000 mg/L was used for preparation of the standard (AAS standard solution; Merck, Kenilworth, New Jersey, U.S.). The quality control of AAS results was ensured using the standard reference materials 1570a (Trace Elements in Spinach Leaves) and 1575a (Pine Needles).

For analysis, approximately 0.2 g of dried sample was added to a Teflon vessel with 5 mL of concentrated HNO₃ and 1 mL of H₂O₂. Subsequently, the samples were digested using a two-step temperature program. During the first step, the temperature was linearly increased to 160 °C for 15 min; the maximum power of the rotating magnetron was 400 W and pressure 2,000 kPa. During the second step, the temperature was maintained at 190 °C for 10 min. After digestion and cooling, digests were quantitatively transferred to 100-mL calibrated flasks and made up to the volume with double distilled water. The results are reported as the average of three repeated measurements, and all digestions were conducted in triplicate. All the reagents used for this study were of analytical grade.

For detection of Cd in the seeds, the seeds were cut in half, washed for 10 min in deionized water on a rocking platform and incubated for 30 min in 0.5% dithizone

(Sigma-Aldrich; Saint-Louise, Missouri, U.S.) solution with 20 μL of ice acetic acid. In reaction to dithizone staining, tissues containing Cd acquire a pinkish-red color. Thereafter, the seeds were washed again with deionized water, dried with blotting paper, and photographed.

2.3. Evaluation of Cell Viability

Cell viability was evaluated based on two methods: Staining with Evans blue (EB; Sigma-Aldrich) and tetrazolium chloride (TTC) (Serva; Heidelberg, Germany). In the first method, based on the method described by Lehotai et al. (2011), with some modification, accumulation of EB indicates cell death. Seeds of similar fresh weight (200 mg) were incubated for 20 min in 0.25% EB solution, washed twice for 15 min in distilled water and homogenized using a mortar and pestle with destaining solution containing 50 mL of ethanol, 49 mL of distilled water, and 1 mL of 10% dodecyl sulfate (SDS; Serva). Samples were incubated in a heating block for 15 min at 50 °C and centrifuged for 15 min at 13,400 g. EB uptake was measured at $\lambda = 600$ nm using Biomate 3S spectrophotometer (Thermo Scientific). Destaining solution was used as the blank. Measurements were performed on samples from five independent experiments.

The second method of cell viability evaluation is based on the fact that in living cells dehydrogenase reduces TTC to insoluble red formazan (Towill & Mazur, 1974). The seeds were weighed, transferred to Eppendorf tubes, and incubated with 1 mL of 0.4% TTC dissolved in phosphate buffer pH 7.5 for 24 hr at 20–22 °C in the dark. Thereafter, they were washed with distilled water, transferred to new Eppendorf tubes, supplemented with 0.5 mL of 95% ethanol and homogenized in Tissue Lyser II (Qiagen; Hilden, Germany). The samples were incubated for 1 hr at 55 °C, supplemented again with 0.5 mL of 95% ethanol, mixed on vortex, and centrifuged for 3 min at 10,000 g. The absorbance of the mixture containing 800 μL of supernatant and 1,400 μL of 95% ethanol was measured at $\lambda = 485$. The blank contained 95% methanol. Measurements were performed on samples from four independent experiments.

2.4. Examination of Membrane Integrity

Membrane integrity was estimated based on ion leakage. Fifty seeds were transferred to glass beaker for every experimental treatment and imbibed for 2 hr in distilled water or CdCl_2 solutions on a shaker at 20–22 °C. Thereafter, the solutions were transferred to new beakers and their conductivity was measured using a conductometer (Tel-Eko; Wrocław, Poland). Measurements were performed on samples from four independent experiments.

2.5. Lipid Peroxidation

Oxidative stress intensity was estimated based on changes in the level of lipid peroxidation products, thiobarbituric acid reactive substances, according to Cuypers et al. (2011) with minor modifications. The seeds (200 mg) were homogenized using a mortar and pestle in 3 mL of 10% trichloroacetic acid (Sigma-Aldrich) buffer and centrifuged for 10 min at 12,000 g. Thereafter, 1 mL of supernatant was transferred to glass tubes, filled with 4 mL of 0.05% thiobarbituric acid (Sigma-Aldrich) dissolved in 10% trichloroacetic acid and incubated for 30 min in 95 °C. Afterwards, the samples were cooled, mixed by inversion, and centrifuged for 1 min at 5,000 g. The absorbance of the supernatant was measured at $\lambda = 532$ nm and corrected for unspecific absorbance at $\lambda = 600$ nm. Measurements were performed on samples from three independent experiments.

2.6. Measurements of Antioxidant Capacity

The antioxidant capacity was estimated based on the reduction rate of 2,2-diphenyl-1-picrylhydrazyl (DPPH; Sigma-Aldrich), which changes color from dark violet to pinkish during the reduction process (Brand-Williams et al., 1995). The seeds (200 mg) were homogenized with 2 mL of 80% methanol, incubated for 2 hr at 37 °C on a shaker, and centrifuged for 10 min at 12,000 g. The absorbance of the

solution containing 100 μL of sample, 250 μL of DPPH, and 2 mL of 80% methanol was measured spectrophotometrically at $\lambda = 517$ nm immediately after preparation and after 10 min of incubation in the dark. The blank contained 250 μL of methanol instead of DPPH. The antioxidant capacity is expressed as the difference between the absorbance before and after incubation in the dark. Measurements were performed on samples from four independent experiments.

2.7. Measurements of the Level of Nonenzymatic Antioxidants

The level of total soluble phenolics was measured using Folin–Ciocalteu (Merck) reagent according to Díaz et al. (2005). Seeds of similar weight (200–300 mg) were homogenized with 4 mL of 80% methanol, incubated for 15 min at 70 °C and after cooling, centrifuged at 2,500 g. The supernatant was diluted to 5 mL with 80% methanol. Mixtures containing 3,750 μL of distilled water, 250 μL of Folin–Ciocalteu reagent, and 250 μL of supernatant were prepared and supplemented after 3 min with 750 μL of 20% Na_2CO_3 . Immediately afterwards, the absorbance of the prepared mixture was measured at $\lambda = 760$ nm.

The level of flavonoids was determined according to Jia et al. (1998). Seeds of similar weight (200–300 mg) were homogenized with 2 mL of 80% methanol, incubated for 2 hr at 37 °C, and centrifuged for 10 min at 13,400 g at 10–12 °C. Thereafter, mixtures containing 250 μL of supernatant, 1,250 μL of distilled water, and 75 μL of 5% NaNO_3 (Merck) were prepared. After 5 min incubation at 20–22 °C, 150 μL of 10% AlCl_3 (Merck) was added. Finally, after an additional 6 min of incubation at 20–22 °C, 500 μL of 1 M NaOH (Merck) was added and the absorbance of the mixture was measured at $\lambda = 510$ nm.

The level of anthocyanins was assessed using the method described by Sims and Gamon (2002) with some modifications. Seeds of similar weight (200–300 mg) were homogenized with a 1 mL mixture containing methanol/distilled water/HCl in the proportion 60:1:1. The samples were incubated for 24 hr at 4 °C. Thereafter, the samples were centrifuged at 13,400 g for 10 min and the absorbance of the supernatant was measured at two wavelengths: $\lambda = 529$ nm and $\lambda = 650$ nm. The level of anthocyanins was calculated according to the formula $AA = A_{529} - (0.288) \times A_{650}$, where AA is the corrected absorbance of anthocyanins.

Ascorbic acid level was measured based on the methods described by Mukherjee and Choudhuri (1985). Seeds of similar weight (200–300 mg) were homogenized with 2 mL of 5% trichloroacetic acid, transferred to glass tubes, and incubated for 15 min at 100 °C with 2 mL of 2% 2,4-dinitrophenylhydrazine (Sigma-Aldrich) dissolved in methanol and 40 μL of 10% thiourea (P.O.Ch) dissolved in 70% ethanol. The mixtures were cooled on ice and supplemented with 2.5 mL of 80% H_2SO_4 (P.O.Ch). Thereafter, the absorbance of the mixtures was measured at $\lambda = 530$ nm.

2.8. Measurements of Germination Rate and Growth Parameters

The seeds were transferred to glass Petri dishes, 30 cm in diameter, and covered with two layers of lignin and one layer of blotting paper. Then, they were watered with 30 mL of distilled water and kept in the cultivation room in the dark at 22 °C. The number of germinated seedlings was counted after 24, 48, and 72 hr. The roots of seedlings germinating for 72 hr were used for measurements of length and fresh and dry weight. Measurements were performed in five–six independent repetitions.

2.9. Statistical Analysis

Statistically significant differences between experimental treatments were calculated using the ANOVA test, at $p = 0.05$ using Free Statistics Calculators version 4.0 (<https://www.danielsoper.com/statcalc/default.aspx>). The statistically significant differences are denoted with an asterisk (*) on the graphs.

3. Results

Even relatively short (2 hr) imbibition in Cd solutions resulted in significant ($p = 0.05$) uptake of this metal by the seeds. The level of Cd in the seeds was dependent

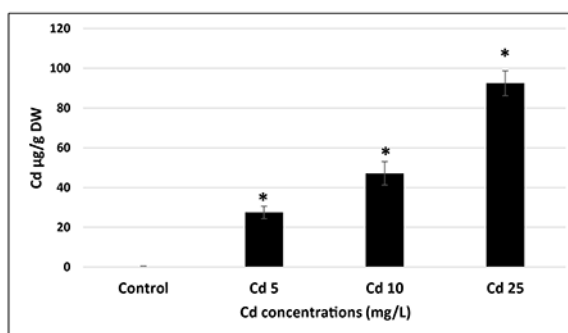


Figure 1 Cd content in the seeds imbedded for 2 hr in CdCl₂ solutions with Cd at concentrations of 5 (Cd 5), 10 (Cd 10), and 25 (Cd 25) mg/L expressed in µg per g of dry weight (DW). The results are means of three independent repetitions ± SE. Significant differences ($p = 0.05$) in relation to the control are marked with an asterisk (*).

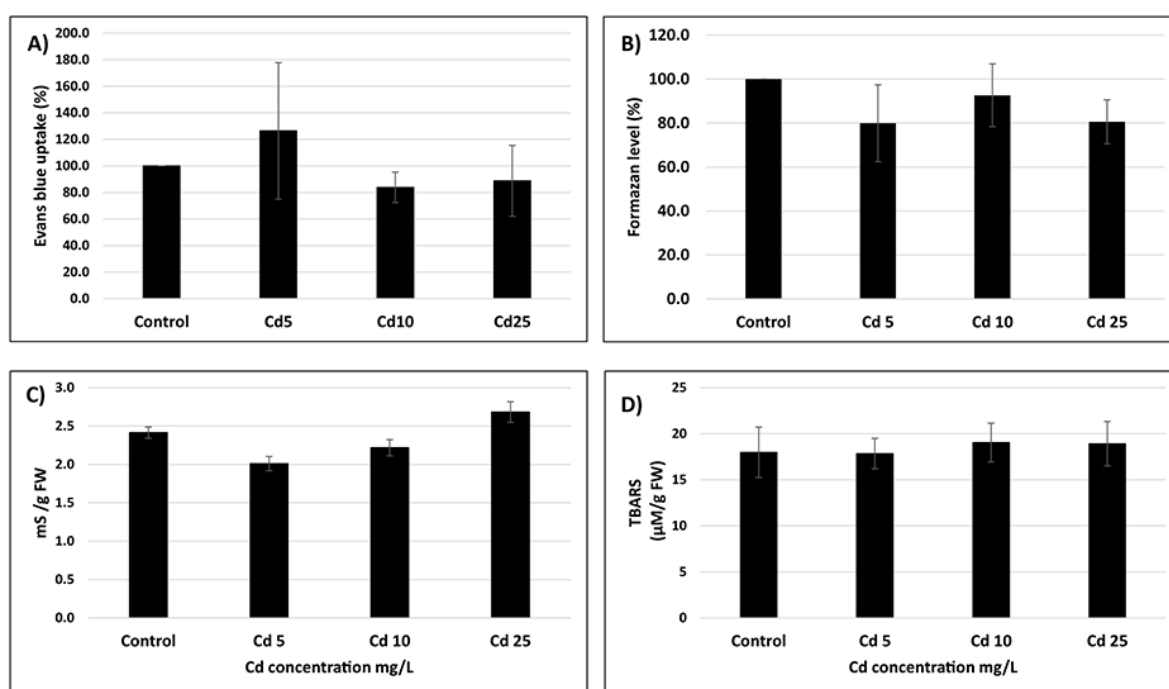


Figure 2 Amount of dead cells expressed as % of the level of Evans blue (A) or tetrazolium chloride (B) changes in electrolyte leakage (C) and thiobarbituric acid reactive substances level (D) in seeds exposed to Cd at concentration 5 (Cd 5), 10 (Cd 10), and 25 (Cd 25) mg/L. The results are means of three–five independent repetitions ± SE. No significant differences were noted.

on the applied metal concentration and reached nearly 100 µg/g dry weight in the case of the most severe treatment (25 mg/L) (Figure 1). Cd uptake by the seeds was also confirmed by histochemical staining with dithizone reagent (Figure S1). The images show that Cd is mainly accumulated in the cotyledons.

Despite significant uptake of Cd, the seeds showed no symptoms of metal toxicity. No differences were observed in the amount of dead cells assessed by two distinct methods – staining with EB and TTC assay (Figure 2A,B). The conductometric measurements of electrolyte leakage were similar in the control and Cd-treated seeds, implicating that the membrane integrity was intact (Figure 2C). Additionally, no symptoms of oxidative stress were observed as indicated by similar thiobarbituric acid reactive substances levels in all experimental treatments (Figure 2D).

The only statistically significant difference ($p = 0.05$) between control and Cd-treated seeds was a decrease in the antioxidant activity noted in response to the highest applied Cd concentration (Figure 3).

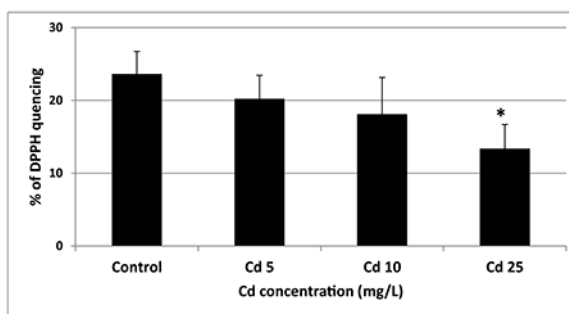


Figure 3 Antioxidant activity of seeds exposed to Cd at concentration 5 (Cd 5), 10 (Cd 10), and 25 (Cd 25) mg/L expressed as % of DPPH quenching. The results are means of five independent repetitions \pm SE. Statistically significant differences ($p = 0.05$) in relation to the control are marked with an asterisk (*).

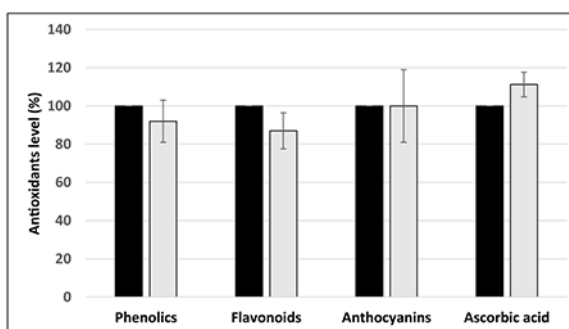


Figure 4 Level of nonenzymatic antioxidants in control seeds (black bars) and seeds treated with Cd at the concentration 25 mg/L (light grey bars) expressed as % in relation to the control. The results are means of three–four independent repetitions \pm SE. No significant differences were noted.

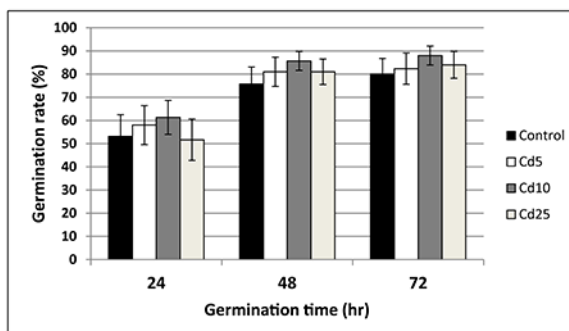


Figure 5 Germination rate of control seeds (black bars) and seeds preincubated in Cd solutions with Cd at concentration 5 mg/L (white bars), 10 mg/L (grey bars), and 25 mg/L (creamy bars) measured after 24, 48, and 72 hr. The results are means of six independent repetitions \pm SE. No significant differences were noted.

However, the level of several nonenzymatic antioxidants, namely total soluble phenolics, flavonoids, anthocyanins, and ascorbic acid, did not differ between the control and the seeds treated with Cd at the concentration 25 mg/L (Figure 4).

Seeds from all experimental treatments showed a similar germination rate measured at three time points – after 24, 48, and 72 hr (Figure 5). The germinated seedlings showed no differences in the terms of root length and fresh and dry weight (Figure 6A–C).

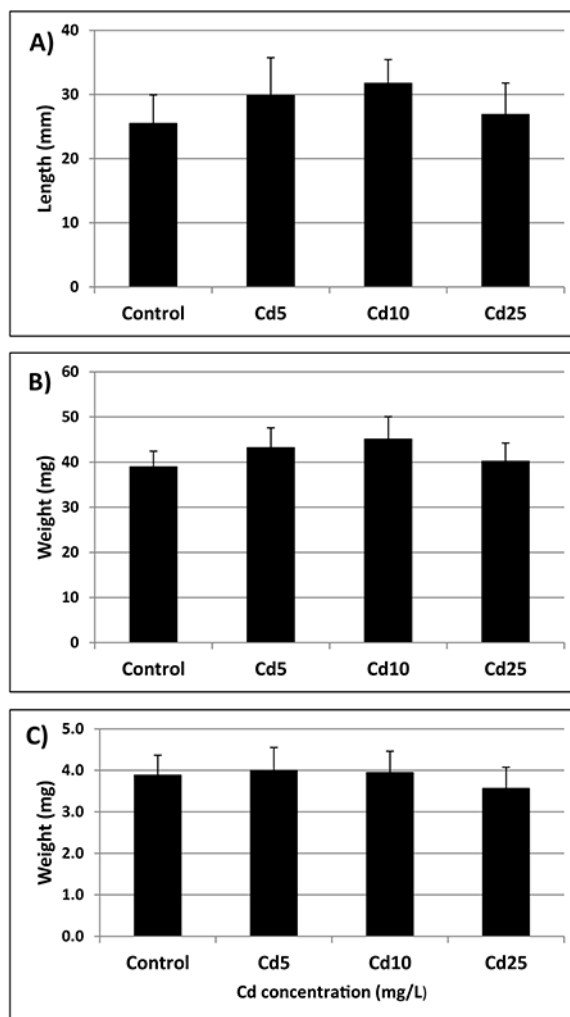


Figure 6 Root length (A), fresh weight (B), and dry weight (C) of seedlings grown from seeds preincubated in Cd solution with Cd at concentrations of 5 (Cd 5), 10 (Cd 10), and 25 (Cd 25) mg/L measured after 72 hr of germination. The results are means of five independent repetitions \pm SE. No significant differences were noted.

4. Discussion

High Cd levels have been detected in various regions of the world (Khan et al., 2011, 2017; Wang et al., 2015). This metal is readily taken up by plants thereby altering their functioning and limiting growth. Importantly, Cd absorbed by edible plants can enter the human food chain (Rizwan et al., 2017). According to the Agency for Toxic Substances and Disease Registry (2020), Cd is among the top 10 substances which pose the most significant threat to human health.

The present study assessed the impact of Cd on the germination process in an important crop plant species – soybean. The results show that even short-term imbibition in Cd solution results in significant uptake of this element by soybean seeds (Figure 1). The amount of Cd reached nearly 100 μ g/g of dry weight in the case of the treatment with the highest applied metal concentration (25 mg/L). However, metal treatment did not affect cell viability assessed by two distinct methods – staining with EB and TTC assay (Figure 2A,B). Additionally, seeds imbibed in metal solutions showed no differences in the level of electrolytes leakage or lipid peroxidation products (thiobarbituric acid reactive substances) (Figure 2C,D). These results indicate that the cellular membranes were not damaged under Cd treatment. In contrast, imbibition in the highest Cd concentration (25 mg/L) led to significant decrease in seeds antioxidant activity (Figure 3). The fact that Cd affects antioxidant

machinery is well documented. However, so far, its impact on the antioxidant system has been examined mainly in seedlings or fully developed plants (reviewed by Chmielowska-Bąk et al., 2014). In the present study, we show that Cd already decreases antioxidant capacity in seeds during the earliest germination phase. Further investigation showed that the observed attenuation of antioxidant activity is not associated with the decrease in the level of major nonenzymatic antioxidants including total soluble phenolics, flavonoids, anthocyanins, and ascorbic acid (Figure 4). Therefore, it can be suspected that the main reason for the observed depletion in the antioxidant defense are changes in the activity of antioxidant enzymes. This assumption would need further evaluation.

According to the “oxidative window for germination” hypothesis, precise regulation of ROS levels, among others by the antioxidant system, is indispensable for the proper progression of the germination process (Bailly et al., 2008). The beneficial roles played by ROS during germination include involvement in signaling network, selective oxidation of transcripts and proteins associated with mobilization of storage material, endosperm weakening, and enhancing protection against pathogens. In turn, ROS overproduction is associated with loss of membrane integrity, DNA damage, aging, and decreased seeds vigor (Bailly et al., 2008; Waterworth et al., 2015; Wojtyła et al., 2016). Therefore, it could be expected that the observed decrease in antioxidant activity in Cd-treated soybean seeds will affect their future germination. However, no perturbations in the germination process were observed. Preincubation in Cd did not affect germination rate at any of the measured timepoints – after 24, 48, or 72 hr (Figure 5). Additionally, seedlings grown from Cd pretreated seeds showed no differences in any of the measured growth parameters (Figure 6A–C). The effect of Cd treatment on germination rate and growth of young seedlings varies depending on plant species, applied concentration, duration of treatment, and method of metal application. For instance, a study on six species of pulses showed that in general, metal treatment inhibited germination rate and led to decrease in root and shoot length (Tao et al., 2015). Similarly, treatment of pigeon pea, fenugreek, and maize with Cd led to reduced germination rate and/or seedlings growth (Sneideris et al., 2015; Wahis et al., 2015; Zayneb et al., 2015). However, in the case of flax and China aster the inhibitory effect was observed only at the highest metal concentrations (Kavuličova et al., 2012). Moreover, in *Dorycnium pentaphyllum* application of Cd at low concentration resulted in slightly higher final germination percentage, although severe Cd stress led to drastic disturbances in germination (Lefèvre et al., 2009).

In summary, it can be concluded that although cadmium is considered a highly toxic compound, soybean seeds are tolerant to its action, at least during the first phase of germination. The only observed difference between control seeds and seed exposed to Cd was a decrease in the total antioxidant activity observed in response to the highest metal concentration. It would be interesting to assess if the early exposure to the metal affects future development or stress response of soybean plants.

5. Supporting Material

The following supporting material is available for this article:

- Figure S1. Detection of Cd in soybean seeds imbibed for 2 hr in Cd solutions with Cd at the concentration 5, 10, and 25 mg/L.

References

- Agency for Toxic Substances and Disease Registry. (2020). *ATSDR's substance priority list*. <https://www.atsdr.cdc.gov/spl/>
- Arc, E., Galland, M., Godin, B., Cueff, C., & Rajjou, L. (2013). Nitric oxide implication in the control of seed dormancy and germination. *Frontiers in Plant Science*, 4, Article 346. <https://doi.org/10.3389/fpls.2013.00346>
- Bailly, C., El-Maarouf-Bouteau, H., & Corninea, U. (2008). From intracellular signaling network to cell death: The dual role of reactive oxygen species in seed physiology. *Comptes Rendus Biologies*, 331, 806–814. <https://doi.org/10.1016/j.crv.2008.07.022>

- Brand-Williams, W., Cuvelier, M. E., & Berset, C. (1995). Use of free radical method to evaluate antioxidant activity. *LWT – Food Science and Technology*, 28, 25–30. [https://doi.org/10.1016/S0023-6438\(95\)80008-5](https://doi.org/10.1016/S0023-6438(95)80008-5)
- Chmielowska-Bąk, J., Gzyl, J., Rucińska-Sobkowiak, R., Arasimowicz-Jelonek, M., & Deckert, J. (2014). The new insights into cadmium sensing. *Frontiers in Plant Science*, 5, Article 245. <https://doi.org/10.3389/fpls.2014.00245>
- Cuypers, A., Smeets, K., Ruytinx, J., Opdenakker, K., Keunen, E., Remens, T., Horemans, N., Vanhoudt, N., Sanden, S., F., B., Guisez, Y., Colpaert, J., & Vangronsveld, J. (2011). The cellular redox state as a modulator in cadmium and copper responses in *Arabidopsis thaliana* seedlings. *Journal of Plant Physiology*, 168, 309–316. <https://doi.org/10.1016/j.jplph.2010.07.010>
- Díaz, J., Silvar, C., Varela, M. M., Bernal, A., & Merino, F. (2005). *Fusarium* confers protection against several mycelial pathogens of pepper plants. *Plant Pathology*, 54, 773–780. <https://doi.org/10.1111/j.1365-3059.2005.01285.x>
- Donohue, K. (2005). Seeds and seasons: Interpreting germination timing in the field. *Seed Science Research*, 15, 175–187. <https://doi.org/10.1079/SSR2005208>
- Finch-Savage, W. E., & Leubner-Metzger, G. (2006). Seed dormancy and the control of germination. *New Phytologist*, 171, 501–523. <https://doi.org/10.1111/j.1469-8137.2006.01787.x>
- Gallego, S. M., Pena, L. B., Barcia, R. A., Azpillicueta, C. E., Iannone, M. F., Rosales, E. P., Zawoznik, M. S., Groppa, M. D., & Benavides, M. P. (2012). Unravelling cadmium toxicity and tolerance in plants: Insight into regulatory mechanisms. *Environmental and Experimental Botany*, 83, 33–46. <https://doi.org/10.1016/j.envexpbot.2012.04.006>
- Ilić, S. Z., Mirecki, N., Trajković, R., Kapoulas, N., Milenković, L., & Šunić, L. (2015). Effect of Pb on germination of different seeds and his translocation in bean seed tissues during sprouting. *Fresenius Environmental Bulletin*, 24, 670–675.
- Jia, Z., Tang, M., & Wu, J. (1998). The determination of flavonoid contents in mulberry and their scavenging effects on superoxides radicals. *Food Chemistry*, 64, 555–559. [https://doi.org/10.1016/S0308-8146\(98\)00102-2](https://doi.org/10.1016/S0308-8146(98)00102-2)
- Kavuličová, J., Kaduková, J., & Ivánová, J. (2012). The evaluation of heavy metal toxicity in plants using the biochemical tests. *Nova Biotechnologica et Chimica*, 11, 101–109. <https://doi.org/10.2478/v10296-012-0011-2>
- Khan, M. A., Khan, S., Khan, A., & Alam, M. (2017). Soil contamination with cadmium, consequences and remediation using organic amendments. *Science of The Total Environment*, 601-602, 1591–1605. <https://doi.org/10.1016/j.scitotenv.2017.06.030>
- Khan, S., Khan, M. A., & Rehman, S. (2011). Lead and cadmium contamination of different roadside soils and plants in Peshawar City, Pakistan. *Pedosphere*, 21, 351–357. [https://doi.org/10.1016/S1002-0160\(11\)60135-5](https://doi.org/10.1016/S1002-0160(11)60135-5)
- Kranner, I., & Colville, L. (2011). Metals and seeds: Biochemical and molecular implications and their significance for seed germination. *Environmental and Experimental Botany*, 72, 93–105. <https://doi.org/10.1016/j.envexpbot.2010.05.005>
- Lefèvre, I., Marchal, G., Gorréal, E., Zanuzzi, A., & S, L. (2009). Variation in response to heavy metals during vegetative growth in *Dorycnium pentaphyllum* Scop. *Plant Growth Regulation*, 59, 1–11. <https://doi.org/10.1007/s10725-009-9382-z>
- Lehotai, N., Petö, A., Bajkán, S., Tari, I., & Kobert, Z. (2011). In vivo and in situ visualization of early physiological events induced by heavy metals in pea root meristem. *Acta Physiologiae Plantarum*, 33, 2199–2207. <https://doi.org/10.1007/s11738-011-0759-z>
- Miransari, M., & Smith, D. L. (2014). Plant hormones and seed germination. *Environmental and Experimental Botany*, 99, 110–121. <https://doi.org/10.1016/j.envexpbot.2013.11.005>
- Mukherjee, S. P., & Choudhuri, M. A. (1985). Implication of hydrogen peroxide–ascorbate system on membrane permeability of water stressed *Vigna* seedlings. *New Phytologist*, 99, 355–360. <https://doi.org/10.1111/j.1469-8137.1985.tb03663.x>
- Poschenrieder, C., Cabot, C., Martos, S., Gallego, B., & Barceló, J. (2013). Do toxic ions induce hormesis in plants? *Plant Science*, 212, 15–25. <https://doi.org/10.1016/j.plantsci.2013.07.012>
- Rizwan, M., Alis, S., Adrees, M., Ibrahim, M., Tsang, D. C. W., Zia-Ur-Rehman, M., Zahir, Z. A., Rinklebe, J., Tack, F. M. G., & Ok, Y. S. (2017). A critical review on effects, tolerance mechanisms and management of cadmium vegetables. *Chemosphere*, 182, 90–105. <https://doi.org/10.1016/j.chemosphere.2017.05.013>
- Sethy, S. K., & Ghosh, S. (2013). Effect of heavy metals on germination of seeds. *Journal of Natural Science, Biology and Medicine*, 2, 272–275. <https://doi.org/10.4103/0976-9668.116964>

- Shu, K., Liu, X. D., Xie, Q., & He, Z. H. (2016). Two faces of one seed: Hormonal regulation of dormancy and germination. *Molecular Plant*, 9, 34–45. <https://doi.org/10.1016/j.molp.2015.08.010>
- Sims, D. A., & Gamon, J. A. (2002). Relationship between leaf pigment content and spectral reflectance across a wide range of species, leaf structures and developmental stages. *Remote Sensing of Environment*, 87, 337–354. [https://doi.org/10.1016/S0034-4257\(02\)00010-X](https://doi.org/10.1016/S0034-4257(02)00010-X)
- Sneideris, L. C., Gavassi, M. A., Campos, M. L., D'Amico-Damião, V., & Carvalho, R. F. (2015). Effects of hormonal priming on seed germination of pigeon pea under cadmium stress. *Anais da Academia Brasileira de Ciências*, 87, 1847–1852. <https://doi.org/10.1590/0001-3765201520140332>
- Tao, L., Guo, M., & Ren, J. (2015). Effects of cadmium on seed germination, coleoptile growth, and root elongation in six pulses. *Polish Journal of Environmental Studies*, 24, 295–299. <https://doi.org/10.15244/pjoes/29942>
- Towill, E. L., & Mazur, P. (1974). Studies on the reduction of 2,3,5-triphenyltetrazolium chloride as a viability assay for plant tissue cultures. *Canadian Journal of Botany*, 53, 1097–1102. <https://doi.org/10.1139/b75-129>
- Tran, T. A., & Popova, L. P. (2013). Functions and toxicity of cadmium in plants: Recent advances and future prospects. *Turkish Journal of Botany*, 37, 1–13. <https://doi.org/10.3906/bot-1112-16>
- Wahis, A., & Khaliq, S. (2015). Architectural and biochemical changes in embryonic tissues of maize under cadmium toxicity. *Plant Biology*, 17, 1005–1012. <https://doi.org/10.1111/plb.12326>
- Wang, L., Cui, X., Cheng, H., Chen, F., Wang, J., Zhao, X., Lin, C., & Pu, X. (2015). A review of soil contamination in China including health risk assessment. *Environmental Science and Pollution Research*, 22, 16441–16452. <https://doi.org/10.1007/s11356-015-5273-1>
- Waterworth, W. M., Bray, C. M., & West, C. E. (2015). The importance of safeguarding genome integrity in germination and seed longevity. *Journal of Experimental Botany*, 66, 3549–3558. <https://doi.org/10.1093/jxb/erv080>
- Wietbrecht, K., Müller, K., & Leubner-Metzger, G. (2011). First off the mark: Early seed germination. *Journal of Experimental Botany*, 62, 3289–3309. <https://doi.org/10.1093/jxb/err030>
- Wojtyła, Ł., Lechowska, K., Kubala, S., & Garnczarska, M. (2016). Different modes of hydrogen peroxide action during seed germination. *Frontiers in Plant Science*, 7, Article 66. <https://doi.org/10.3389/fpls.2016.00066>
- Zayneb, C., Bassem, K., Zeineb, K., Grubb, C. D., Nureddine, D., Hafedh, M., & Amine, E. (2015). Physiological responses of fenugreek seedlings and plants treated with cadmium. *Environmental Science and Pollution Research*, 22, 10679–10689. <https://doi.org/10.1007/s11356-015-4270-8>