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Research Article

Evaluation of acute oral toxicity study of essential oils (Eos) from *Pogostemon benghalensis* and *P. cablin* in Wistar rats

Pradeep D P^{1*}, Murugan K² and Manoj G S³^{1,3} Department of Botany, Mahatma Gandhi College, Trivandrum, Kerala, India² Center for Innovation in Science and Social Action, Trivandrum, Kerala, India

ABSTRACT

The use of crude herbal decoctions in the traditional treatment of diseases is a common practice. *Pogostemon benghalensis* and *P. cablin* are commonly used for treatment of diverse categories of diseases such as infectious and non-infectious disease. Native people use the crude decoctions as bactericidal, antimalarial, anti-leishmania, anti-diarrheal and insecticidal activities. Its safety profile is not yet elucidated and therefore, this study was to analyze the acute toxicity of essential oils (Eos) from *P. benghalensis* and *P. cablin* as medicinal. Methods include acute toxicity study using male and female Wistar albino rats with single oral dose and followed up to 14 days as per the guidelines of OECD. Visual observations were carried regularly during the experimental period while body weight was measured weekly. Organ weight, clinical chemistry and hematology data were collected on the 7th and 14th days. Results were presented as mean \pm standard deviation. One-way analysis of variance (ANOVA) was carried. Oral administration of Eos from *P. benghalensis* and *P. cablin* revealed no treatment-related mortality in female rats up to the dose of 5000 mg/kg. In acute toxicity studies, no remarkable treatment related anomalies were observed compared to negative controls. Food consumption, body weight, organ weight, hematology did not showed sound variation between controls and treatment groups. However, creatinine, triglycerides, and monocytes were lower in the treated groups in 7th day as compared to control groups. No significant variations between male and female groups in relative organ weight, hematology were noticed. In conclusion, the Eos from *P. benghalensis* and *P. cablin* showed LD₅₀ > 3000 mg/kg in acute toxicity studies.

Keywords: *Pogostemon benghalensis*, *P. cablin*, traditional medicine, safety, plant medicine, adverse effect, acute oral toxicity

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*Address for Correspondence:

Pradeep D P, Department of Botany, Mahatma Gandhi College, Trivandrum, Kerala, India

INTRODUCTION

Globally, secondary metabolites from plants are employed as therapeutics in primary health care system. Most of the crude extracts used by the native communities are non-validated scientifically. The plant extracts can be harmful at certain concentrations, but may be safe and curative in other doses. Therefore, there is a range of possible impacts from acute to sub-acute level of toxicity. The plants synthesize the pool of phytochemicals like protein, sugar, lipids, polyphenols, alkaloids, saponins, nitrogen conjugates etc. as a part of defense mechanisms against grazing animals or invading pathogens. Many synthetic compounds that have been proven as toxic even in minute quantities, while the plant-based drugs have been employed to prevent, diagnose, and treat diverse disorders. This therapy has been practiced out for the past several decades. Exposure of a phytochemical in human beings can be studied by analyzing

the cumulative effects and concentrations that may be carcinogenic, mutagenic, teratogenic, skin or eye irritation and others¹. Toxicity testing is essential to quantify the damage level caused by the molecules to biological and non-biological compounds of the cell system. This acute oral toxicity test is usually attempted on prospective products to develop novel drugs and to determine their therapeutic potentials².

Lack of evidence-based approaches like the legal and regulatory framework, pharmacovigilance, non-standardization and lack of toxicological profiling of herbal products form the greatest concern of medicinal plants usage^{3, 4}. Plants synthesize diverse metabolites that form complex molecules that may be beneficial or harmful to humans^{5, 6}. The general view is that natural plant based products are safe and without side effects and is proven false in some cases⁷. Investigations carried out on many plant

species revealed toxic substances. Despite the widespread crude usage of *Pogostemon benghalensis* and *P. cablin* in traditional medicine, there is a lack of experimental data on its possible toxicity. In addition, allelopathic enthusiasts criticize herbal specialties in terms of organ damage. To eliminate these issues and to ensure the safety of the herbals like *P. benghalensis* and *P. cablin*, the present study was carried in terms of acute toxicity analysis of essential oils extract *via* administered orally in male and female Wistar rats.

MATERIALS AND METHODS

Plant Material

Fresh leaves and twigs of *Pogostemon benghalensis* and *P. cablin* used in the present study were collected from the natural habitats of Munnar Hills of Idukki district, Kerala, India on July 2019. The collected species were identified using flora and were authenticated at the Herbarium of Jawaharlal Nehru Tropical Botanic Garden and Research Institute (JNTBGRI), Palode, Trivandrum, Kerala, India. The voucher specimens were numbered (MGH 1456 and 1457 for *Pogostemon benghalensis* and *P. cablin* respectively) and kept in the herbarium of the Department of Botany, Mahatma Gandhi College, Trivandrum, Kerala, India. The collected plants were washed thoroughly with running tap water and the hydro-distillation of fresh leaves were carried out using a Clevenger-type apparatus to extract the essential oil.

Animals

The healthy Wistar rats of 8 to 10 weeks aged with the body weight of 120–140 g were used for the present acute oral toxicity studies. The rats were caged in standard lab conditions for 5 days at the temperature of 23°C (\pm 1°C), with 12 h light/ dark cycle. They were given standard diet and water *ad libitum*^{8,9}.

Rats were randomly categorized in to control and experimental groups. A total of 30 rats containing 6 rats per group (three male and three female, I to V group) were used for acute toxicity study a control (water alone), tween-20 (solvent) and six treatment classes (of Eos from *P. benghalensis* and *P. cablin* solubilized i.e., 500, 1500 and 3000 mg/kg b.w). All classes of the rats were fasted overnight prior to Eos administrations. After the fasting period, all rats were weighed, and the concentrations were carried based on their body weight. The extracts were prepared in tween-20. Eos of the two species were separately mixed in tween-20 at the ratio of 1: 1 v/v in glass tubes and labelled carefully. Rats in groups III to V were given single oral dose of the respective oil by using feeding needle and rats were kept without food for 3–4 h prior to administration. Initially, the rats were closely observed for first 60 min, intermittently for 4 h and thereafter once every 24 h for 14 days. Food was given after 1–2 h of dosing. The same procedure was carried for the vehicle group of 6 rats to whom tween-20 was administered in same volume as that of treated groups. Both the groups were observed closely for any toxic effect at regular intervals for a total period of 14 days. Surviving rats were observed to determine the toxic reactions onset. Weights of animals were monitored and documented as well. At the end of study, rats were weighed and blood samples were collected by cardiac puncture under anesthesia with isoflurane and serum was separated for biochemical and hematological evaluations. Vital organs were excised after killing mice by cervical dislocation and the weight of organs was noted.

Biochemical assay

The biochemical parameters include serum urea, serum creatinine, alanine aminotransferase (ALT), aspartate aminotransferase (AST), alkaline phosphate (AP), total bilirubin, total protein, albumin, globulins, cholesterol, triglycerides (TGA), high density lipoprotein (HDL), low density lipoprotein (LDL) and very low density lipoprotein (VLDL) were measured by using Randox kits.

Hematological evaluations

The blood samples from animals (both treated and vehicle control groups) were collected in EDTA containing tubes for hematological study. Hemoglobin (Hb), total red blood cells RBC, hematocrit (HCT), mean corpuscular volume (MCV), mean corpuscular hemoglobin (MCH), mean corpuscular hemoglobin concentration (MCHC), platelet count, white blood cells count (WBC), neutrophils, lymphocytes, monocytes and eosinophils were determined with automatic hematology analyzer.

Statistical analysis

Experimental results were presented as mean \pm SD and the statistical significance between the groups was analyzed by means of one way ANOVA followed by Tukey's multiple comparison test. $P \leq 0.05$ was considered as statistically significant.

RESULTS AND DISCUSSION

The doses of Eos from *P. benghalensis* and *P. cablin* employed were 500, 1500 and 3000 mg/kg body weight of rats in the test groups. Control class (I and II) received water and tween-20 respectively. After feeding the Eos, the rats were kept under close vision continuously for 1 h and intermittently for 4 h and thereafter once every 24 h for 14 days.

During this course of experimental periods, medical observations were carried for mortality, behavioral changes, neurological features and other anomalies. The body weight was measured weekly. Finally, the gross physical examinations were attempted. The animals were then anesthetized under isoflurane. After sacrificing, gross pathological features of the rats were attempted on the vital organs.

The acute toxicity analyzes did not revealed any toxic symptoms or signs among the experimental groups of male and female rats from 500 to 3000 mg/kg b.w. No morbidity or mortality was noticed in the administrated rats at the above specified doses during the acute toxicity studies. As a result, the LD₅₀ of the Eos from *P. benghalensis* and *P. cablin* should be higher than 3000 mg/kg body weight.

The gross pathological analyzes of the kidneys and liver of Eos from *P. benghalensis* and *P. cablin* administrated rats revealed no anomalous physical changes in terms of size, shape, color, and texture as compared with the control groups. The absolute mean liver weights were 5.52 g (at 1500 mg/kg *P. benghalensis*), 5.67g (at 3000 mg/kg *P. benghalensis*) compared with the control (5.43 g). Similarly, the absolute mean kidney weights were 1.18 g (at 1500 mg/kg *P. benghalensis*) and 1.17 g (at 3000 mg/kg *P. benghalensis*) as compared with the control group (1.17 g) (Fig.1). Similarly, the liver weights were 5.59 g (at 1500 mg/kg *P. cablin*) and 5.71g (at 3000 mg/kg *P. cablin*). Similarly, the absolute mean kidney weights were 1.21 g (at 1500 mg/kg *P. cablin*) and 1.22 g (at 3000 mg/kg *P. cablin*) (Fig.1).

There was a gradual increase in the body weight of both the treated and control rats (Table 1). The initial mean body of control rats was 123.67g; at the end of the experiment their final mean body weight was 136.21 g. The mean body weight gain for the control rats was 12.54 g. The initial mean body weights of rats treated with doses of 1500 mg/kg, and 3000 mg/kg of Eos from *P. benghalensis* were 126.2 g, and 127.8g

respectively. At the end of the experiment (after 14 days) the final mean body weight of rats treated with 1500 mg/kg and 3000 mg/kg was 139.1 g and 140.4 g, respectively. The mean body weight gain for rats treated with 1500 mg/kg, and 3000 mg/kg of *P. cablin* was 140.2 g and 141.1 g respectively (after 14 days) (Table 1).

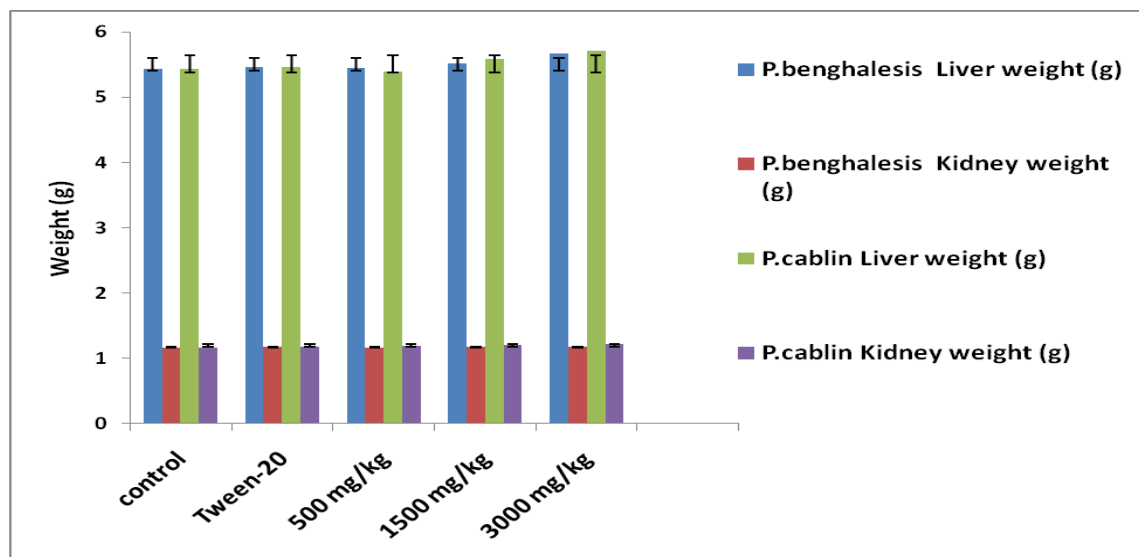


Figure 1: Liver and kidney weights of rats treated with Eos from *P. benghalensis* and *P. cablin*

Table 1: Body weights of rats treated with Eos from *P. benghalensis* and *P. cablin*

Treatment groups	<i>P. benghalensis</i>		<i>P. cablin</i>	
	1 week	2 week	1 week	2 week
Control	123.67 ±0.56	136.21 ±0.18	123.67 ±0.23	136.21±0.44
Tween-20	127.45 ±0.71	134.5 ±0.87	127.45±0.25	134.5±0.03
500 mg/kg	125.1 ±0.27	134.4 ±0.34	126.3±0.23	133.9±0.99
1500 mg/kg	126.2 ±0.67	139.1 ±0.24	127.0 ±0.18	140.2±0.21
3000 mg/kg	127.8 ±0.38	140.4 ±0.31	127.93±0.26	141.1±0.36

Acute toxicity analysis is a pre-requisite to evaluate the adverse impacts that happen within a short time of administration of a herbal drug. Principally, such tests are carried in rats/mice and are usually attempted in designing a new drug or crude product to gain amble knowledge on its toxicity potentials⁴.

For acute toxicity study, 500, 1500 and 3000 mg/kg of Eos from *P. benghalensis* and *P. cablin* were given to rats. The rats did not show any signs/symptoms of toxicity at all the tested concentrations. No mortality was seen in the administrated rats, i.e., even at 3000 mg/kg during the course of the study period. Thus, the LD₅₀ of the Eos could be higher than 3000 mg/kg. The Eos from *P. benghalensis* and *P. cablin* was considered safe on acute dose exposure.

Biochemical parameters

No sound serum creatinine level change was noticed; meanwhile serum urea level in acute toxicity group was slightly higher as compared to vehicle control group (Table 2). Biochemical markers of liver function test revealed significant ($p < 0.05$) changes (Table 2). There was no remarkable variation in the creatinine and urea level in the control and treated groups. Alanine aminotransferase (ALT), aspartate aminotransferase (AST) levels were increased. Meanwhile, alkaline phosphatase (AP) was reduced in acute toxicity group as compared to vehicle control group.

Table 2: Biochemical parameters of rats treated with Eos from *P. benghalensis* and *P. cablin*

Parameters	Vehicle control group	Acute toxicity group (Eos from <i>P. benghalensis</i> - 3000 mg/kg b.w)	Acute toxicity group (Eos from <i>P. cablin</i> - 3000 mg/kg b.w)
Creatinine serum (mg/dl)	0.396 ±0.065	0.392 ±0.01	0.402 ±0.05
serum urea (mg/dl)	17.86 ±0.14	18.05 ±0.28	18.84 ±0.43
ALT (U/L)	210.0 ±2.55	248.7 ±1.54	251.0 ±2.65
AST (U/L)	336.6 ±1.89	339.0 ±2.38	340.6 ±2.55
AP (U/L)	165.2 ±0.56	124.3 ±0.09	128.4 ±0.54

(ALT- Alanine aminotransferase, AST- Aspartate aminotransferase, AP- Alkaline phosphatase)

Total protein and globulin levels were increased. However, no remarkable changes were seen in total bilirubin and albumin among the tested groups. It was also noticed that when the Eos treated groups was compared with vehicle

control group in terms of HDL and VLDL showed no sound variations, however there was significant ($p < 0.05$) increase in cholesterol, triglycerides (TGA) and LDL (Table 3).

Table 3: Biochemical parameters of rats treated with Eos from *P. benghalensis* and *P. cablin*

Parameters	Vehicle control group	Acute toxicity group (Eos from <i>P. benghalensis</i> - 3000 mg/kg b.w)	Acute toxicity group (Eos from <i>P. cablin</i> - 3000 mg/kg b.w)
Total bilirubin (mg/dl)	0.91 ±0.07	0.915±0.006	0.920±0.013
Total protein (U/dl)	6.7 ±0.21	7.59±0.09	7.96±0.25
Albumin (U/dl)	4.62±0.054	4.59 ±0.21	4.69±0.05
Globulin (U/dl)	2.4±0.38	2.64 ±0.12	2.72 ±0.021
Cholesterol (mg/dl)	164.36±0.89	173 ±0.02	174.89 ±0.12
TGA (mg/dl)	126.2 ±0.18	148 ±0.31	150 ±0.27
HDL (mg/dl)	31 ±0.21	35 ±0.29	36.2 ±0.33
LDL (mg/dl)	111 ±0.056	129.56 ±0.18	131±0.65
VLDL (mg/dl)	24.5±0.34	28.9 ±0.21	29.2 ±0.28

(TGA- Triglycerides, HDL- High density lipoprotein, LDL- Low density lipoprotein, VLDL-Very low density lipoprotein)

In acute toxicity evaluation of Eos from *P. benghalensis* and *P. cablin*, the health status of the animal was evaluated by biochemical parameters including serum biomarkers activity levels. Ramaiah¹⁰ and Ozer¹¹ reported that liver damages due to hepatotoxic chemicals can elevate the ALT, AST and total proteins levels. Remarkable elevation in ALT, AST, total proteins and globulin levels were seen in the present study (Table. 2). Adedapo¹² reported the toxic effects of *Euphorbia* species on hematological and biochemical parameters of rats. Similarly, Adeoye¹³ correlated the genotoxic and systemic toxicity of pharmaceutical effluent in Wistar rats with oxidative stress. Ogunlana¹⁴ evaluated the toxicological profile of the *Caesalpinia bonduc* extract. Ali¹⁵ documented cytogenetic damage among the female agricultural workers of Pakistan exposed to pesticides. Hepato cellular damage may leads to increased cell membrane permeability and release of amino transferases into the blood stream. AP is treated as the indicator of biliary tract obstruction¹⁶. In the present study, there was remarkable reduction in AP levels was noticed (Table. 2) which is a sign of herbal hepato protective effect. Multiple hyperlipidemias are often secondary to many factors like over diet, alcohol intake, therapies or to diseases like nephrosis, diabetes, hypothyroidism or tumors¹⁷. Enhanced cholesterol, triglycerides and LDL levels noticed in the experimental group suggests the multiple

hyperlipidemic impacts of Eos of *Pogostemon* species. Goldstein¹⁸ reported the hyperlipidemia in coronary heart disease in 176 families and delineation of a new inherited disorder. Travlos¹⁹ reported that renal function impairment is reflected by increased levels of serum creatinine and urea. In the present data, serum urea levels were found elevated (Table 2) revealing that there is a marginal renal injury.

Hematological data

Blood parameters are sensitive indicators of the physiological changes in response to any pollutant or stress in animals²⁰. Blood platelets have key role in blood clotting processes. This study revealed significant increase in platelet count levels as compared to control vehicle group (Table 4) reflecting hemostatic potential of tested Eos. Li²¹ reported similar stand on the hemostatic activity of the Tibetan herb *Lamiophlomis rotata*. In the present study increased level of mean corpuscular volume (MCV) and reduced mean corpuscular hemoglobin concentration (MCHC) was noticed (Table 4). Elevated WBC count and lymphocytes refers the potentiality of Eos defending nature against the pathogens and also its contribution to enhance cellular inflammatory events. The hematological data of vehicle group and Eos treated group are illustrated in Table 4.

Table 4: Hematological parameters of rats treated with Eos from *P. benghalensis* and *P. cablin*

Parameters	Vehicle control group	Acute toxicity group (Eos from <i>P. benghalensis</i> - 3000 mg/kg b.w)	Acute toxicity group (Eos from <i>P. cablin</i> - 3000 mg/kg b.w)
Hb (g/dl)	12.1±0.21	12.78 ±0.23	12.9 ±0.31
Total RBC (10 ¹² /l)	7.42 ± 0.04	7.89 ±0.08	7.92 ±0.06
HCT (%)	33.5 ±0.11	41.69 ±1.32	42.3 ±1.28
MCV (fl)	45.4 ±0.25	54.3 ±0.78	55 ±0.99
MCH (pg)	16.02 ±0.45	16.09 ±0.21	16.08 ±0.33
MCHC (g/dl)	35.3 ± 0.254	27.8 ±0.28	29 ±0.93
Platelet count (x10 ⁹ /l)	246 ±1.43	490 ±2.34	493 ±3.01
WBC Count (10 ⁹ /l)	3.3 ±0.09	5.23 ±0.76	5.4 ±0.54
Neutrophil (%)	9.9 ±0.03	10.3 ±0.73	10.5 ±0.27
Lymphocytes %	85 ±0.76	87 ±0.58	89 ±0.87
Monocytes %	2.9 ±0.1	3.9 ±0.03	4.09 ±0.04
Eosinophils %	1.02 ±0.04	2.03 ±0.21	2.06 ±0.06

(Hb-Hemoglobin, RBC-Red blood cells, HCT-Hematocrit, MCV-Mean corpuscular volume, MCH-Mean corpuscular hemoglobin, MCHC-Mean corpuscular hemoglobin concentration, WBC-White blood cells)

The present results from the acute toxicity study of Eos from *P. benghalensis* and *P. cablin* supports the nontoxic nature of herbal extracts and was comparable with the reports by other researchers^{10, 11, 12}. Further, the results of Eos extracts showed no symptom of toxicity or mortality even at 3000 mg/kg concentration of the species. Ior²² showed that the LD₅₀ of ethanolic extract from the leaves of *Syzygium guineense* was 4000 mg/kg.

Variations in body weight are a marker character for evaluating the toxicity of plant drugs. In the present work, there was no remarkable increase in the absolute mean body weight of the treated rats as compared to control groups. The absolute mean body weight for the Eos treated rats was 140.4 and 141.1 g as compared to control 136.2 g (after 14 days). The mean body weight gain for rats treated with 1500 mg/kg and 3000 mg/kg were statistically insignificant.

Liver and kidneys of animals are employed by many researchers to analyze the safety or toxicity of drugs or phytochemicals. In the present acute toxicity study, gross pathological results of the kidneys and liver of treated rats did not show much visual variation in terms of size, shape, color, and texture when compared with control groups. Further, there were no remarkable variations in the absolute weight of liver and kidneys of Eos administered animals as compared to control groups. Vahalia²³ analyzed the chronic toxicity versus generic ayurvedic mineral formulation tamra bhasma in laboratory animals and substantiated its safety as drugs. Abba²⁴ analyzed the hemorrhagic centrilobular necrosis and cytoplasmic vacuolation of the hepatocytes in *Syzygium guineense* treated mice. Loha²⁵ authenticated methanolic leaf extract of *Syzygium guineense* in terms of histology of the liver, kidney and biochemical compositions of blood in rats used in acute and sub acute toxicity studies. *Chlorophytum alismifolium* tuber extract's non toxic nature was established by acute and sub-acute toxicity studies in Wistar rats by Abubakar²⁶. Chebaibi²⁷ reviewed the acute toxicity of plant mixture employed in traditional treatment of kidney diseases in Morocco. Jatsa²⁸ evaluated the acute and sub-chronic oral toxicity studies of the aqueous leaf extract of *Clerodendrum umbellatum* on mice. Hemalatha²⁹

confirmed the safety of *Trema orientalis* methanol extract in rats by acute and sub-acute toxicity studies. Gebrehiwot³⁰ proved the non-toxic potential of root extract of *Carissa spinarum* by *in vivo* acute and sub-acute toxicity studies in Swiss albino mice. Abotsi³¹ compared the acute and sub-acute toxicity of the ethanolic extract of the aerial parts of *Hillieria latifolia* in rodents. Jothy³² documented the acute oral toxicity of methanolic seed extract of *Cassia fistula* in mice. Kpemissi³³ proved the acute and sub chronic oral toxicity assessments of *Combretum micranthum* in Wistar rats. Pal and Mishra³⁴ evaluated acute toxicity of the methanolic extract of *Dhatryadi ghrita* in Wistar rats. Acute and sub-acute toxicity of *Echinops kebericho* decoction in rats was validated by Deyno³⁵. Gaelle³⁶ documented acute and sub acute toxicity profiles of the methanol extract of *Lycopersicon esculentum*. All the above plant based works justify the present data on rats treated with Eos from *P. benghalensis* and *P. cablin*.

CONCLUSION

In the background of findings of acute toxicity studies it was possible to suggest that Eos from *P. benghalensis* and *P. cablin* is safe and non toxic based on the mortality, behavioural evaluation, mass of organs, biochemical and hematological parameters. However, the results should be further evaluated for long term usage and repeated dose effects to ensure safety of these aromatic species.

Conflict of interest

The authors declare that they have no conflict of interest.

REFERENCES

- Saleem U, Amin S, Ahmad B, Azeem H, Anwar F, Mary S, "Acute oral toxicity evaluation of aqueous ethanolic extract of *Saccharum munja* Roxb. roots in albino mice as per OECD 425 TG", *Toxicol Rep*, 2017; 4:580-585.
- Falya Y, Sumiwi SA, Levita L, "Toxicity study of plant extracts", *J Pharm Biol Sci*, 2020; 15(2):25-32.
- Oguejiofor CJ, Okoli CO, Ugwah MO, Umaru ML, Ogbulie CS, Mshelia HE, *et al.*, "Acute and sub-acute toxicity of aqueous

- extract of aerial parts of *Caralluma dalzielii* N. E. Brown in mice and rats", *Heliyon*, 2019; 5(1).
4. Kale OE, Awodele O, Akindele AJ, "Sub acute and sub chronic oral toxicity assessments of *Acridocarpus smeathmannii* (DC.) Guill. & Perr. root in Wistar rats", *Toxicol Rep*, 2019; 6:161-175.
 5. Takke A, Shende P, "Nanotherapeutic silibinin: an insight of phytomedicine in healthcare reformation", *Nanomedicine*, 2019; 21.
 6. Friedman LS, Martin P, Munoz SJ. Liver function tests and the objective evaluation of the patient with liver disease. 3rd ed. Philadelphia: WB Saunders; 1996. P. 791-833.
 7. Singh RP, Gangadharappa HV, Mruthunjaya K, "Phospholipids: Unique carriers for drug delivery systems", *J. Drug Deliv Sci Technol*, 2017; 39:166-179.
 8. Organisation for economic co-operation and development (OECD), Guidelines for the testing of new chemicals revised draft guideline; acute and subacute oral toxicity, 2008.
 9. National Research Council (NRC), Guide for the Care and Use of Laboratory Animals. 8th ed. Washington DC, USA: National Academy Press; 2011.
 10. Ramaiah SK, "Preclinical safety assessment: current gaps, challenges, and approaches in identifying translatable biomarkers of drug-induced liver injury", *Clin Lab Med*, 2011; 31(1):161-172.
 11. Ozer J, Ratner M, Shaw M, Bailey W, Schomaker S, "The current state of serum biomarkers of hepatotoxicity", *Toxicol*, 2008; 245(3):194-205.
 12. Adedapo AA, Abatan MO, Olorunsogo OO, "Toxic effects of some plants in the genus euphorbia on haematological and biochemical parameters of rats", *Veterinarski arhiv*, 2004; 74(1):53-62.
 13. Adeoye GO, Alimba CG, Oyeleke OB, "The genotoxicity and systemic toxicity of a pharmaceutical effluent in Wistar rats may involve oxidative stress induction", *Toxicol Rep*, 2015; 2:1265-1272.
 14. Ogunlana OO, Ogunlana OE, Adeneye AA, Chijioke OAC, Olipede TI, Olagunju JA, *et al.*, "Evaluation of the toxicological profile of the leaves and young twigs of *Caesalpinia bonduc* (linn) roxb", *Afr J Tradit Complement Altern Med*, 2013; 10(6):504-512.
 15. Ali T, Bhalli JA, Rana SM, Khan QM, "Cytogenetic damage in female pakistani agricultural workers exposed to pesticides", *Environ Mol Mutagen*, 2008; 49(5):374-380.
 16. Manjunatha BK, Vidya SM, Dhiman P, Pallavi R, Mankani KL, "Hepatoprotective activity of *Leucas hirta* against Ccl-4 induced hepatic damage in rats", *Indian J Exp Biol*, 2005; 43(8):722-7.
 17. Havel RJ, "Pathogenesis, differentiation and management of hyper triglyceridemia", *Adv Intern Med*, 1969; 15:117.
 18. Goldstein JL, Schrott HG, Hazzard WR, Bierman EL, Motulsky AG, "Hyperlipidemia in coronary heart disease ii. Genetic analysis of lipid levels in 176 families and delineation of a new inherited disorder, combined hyperlipidemia", *J Clin Invest*, 1973; 52(7):2544.
 19. Travlos GS, Morris RW, Elwell MR, Duke A, Rosenblum S, Thompson MB, "Frequency and relationships of clinical chemistry and liver and kidney histopathology findings in 13-week toxicity studies in rats", *Toxicol*, 1996; 107(1):17-29.
 20. Jain N, Sharma P, Sharma N, Joshi SC, "Haemato-biochemical profile following sub acute toxicity of in male albino rats", *Avicenna J Phytomed*, 2009; 2:500-506.
 21. Li M, Jia Z, Zhang R, Hu Z, Tian X, "The structure of an iridoid glycoside, 8-deoxyshanzhiside, from *Lamiophlomis rotata*", *Carbohydr Res*, 2008; 343:561-565.
 22. Ior LD, "Anti-inflammatory and analgesic activities of the ethanolic extract of the leaf of *Syzygium guineense* in rats and mice", *J Pharm*, 2012; 2(4):33-36.
 23. Vahalia MK, Thakur KS, Nadkarni S, Sangle VD, "Chronic toxicity study for tamra bhasma (a generic ayurvedic mineral formulation) in laboratory animals", *Recent Res Sci Technol*, 2011; 3(11):76-79.
 24. Abba S, Omotoso OD, Joseph MI, "Hemorrhagic centrolobar necrosis and cytoplasmic vacuolation of the hepatocytes in *Syzygium guineense* chronic treated mice", *Int J Anat Appl Physiol*, 2018; 4(4):99-102.
 25. Loha M, Mulu A, Abay SM, Ergete W, Geleta B, "Acute and subacute toxicity of methanol extract of *Syzygium guineense* leaves on the histology of the liver and kidney and biochemical compositions of blood in rats", *Evid Based Complement Altern Med*, 2019; 1-15.
 26. Abubakar A, Nazifi AB, Hassan FI, Duke KA, Edoh TD, "Safety assessment of *Chlorophytum alismifolium* tuber extract (Liliaceae): Acute and sub-acute toxicity studies in Wistar rats", *J of Acute Toxicity studies*, 2019; 8(1):21-27.
 27. Chebaibi M, Bousta D, Chbani L, Iken I, Achour S, "Evaluation of acute toxicity of plant's mixture used in traditional treatment of kidney diseases in Morocco", *Phcog Res*, 2019; 11:155-161.
 28. Jatsa HB, Fassi JB, Kenfack MC, Feussom NG, Kameni MP, Simo ND, *et al.*, "Acute and sub-chronic oral toxicity studies of the leaves aqueous extract of *Clerodendrum umbellatum* Poir. on mice", *A J Physiol Biochem Pharmacol*, 2018; 7(2):75-85.
 29. Hemalatha T, Mary DA, Ganthi AS, "Acute and sub-acute toxicity study of *Trema orientalis* (L.) Bl. methanol extract in rats", *Journal of Drug Delivery and Therapeutics*, 2019; 9(1-s):307-311.
 30. Gebrehiwot S, *In vivo* acute and sub-acute toxicity study of root extract of *Carissa spinarum* Linn. in Swiss albino mice", *Int J Pharm*, 2019; 11(6):62-65.
 31. Abotsi WKM, Ainooson GK, Gyasi EB, "Acute and sub-acute toxicity studies of the ethanolic extract of the aerial parts of *Hillieria latifolia* (Lam.) H. Walt. (Phytolaccaceae) in rodents", *West Afr J Pharm*, 2011; 22:27-35.
 32. Jothy SL, Zakaria Z, Chen Y, Lau YL, Latha LY, Sasidharan S, Acute oral toxicity of methanolic seed extract of *Cassia fistula* in mice", *Molecules*, 2011; 16:5268-82.
 33. Kpemissi M, Metowogo K, Melila M, Veerapur VP, Negru M, Taulescu M, *et al.*, "Acute and sub chronic oral toxicity assessments of *Combretum micranthum* (Combretaceae) in Wistar rats", *Toxicol Rep*, 2020; 7:162-168.
 34. Pal RS, Mishra A, Evaluation of acute toxicity of the methanolic extract of *Dhatryadi ghrta* in Wistar rats", *The Open Pharm J*, 2019; 9:1-4.
 35. Deyno S, Abebe A, Tola MA, Hymete A, Bazira J, Makonnen E, Alele PE, "Acute and sub-acute toxicity of *Echinops kebericho* decoction in rats", *Complement Ther Med*, 2020; 20(2).
 36. Gaele S, Nguenang, Arsene SM, Ntyam, Kuete V, "Acute and subacute toxicity profiles of the methanol extract of *Lycopersicon esculentum* L. Leaves (Tomato), a botanical with promising *in vitro* anticancer potential", *Evid Based Complement Altern Med*, 2020; (2):1-10.