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Research Article

Assessment of *In Vitro* Antidiabetic Potential of Purified Anthocyanin Extract from Floral Petals of Wild Balsam Species

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ABSTRACT

Diabetes is a notorious and growing clinical and public health issue. The International Diabetes Federation assumes that 592 million had diabetes by 2035 and that by 2040 the number will increase to 642 million. Cardiovascular corollary accounts for four million deaths annually attributable to diabetes. Evidence reveals that certain glucose-lowering phytochemicals can improve vascular outcomes with type 2 diabetes, which, together with better understanding of using multiple therapies concurrently, offers opportunities for beneficial personalization of medication regimens. Anthocyanins are coloured pigments and are natural antioxidants. Keeping this in focus, this study was undertaken to evaluate the *in vitro* antidiabetic activity in the petals of wild *Impatiens balsamina* L. The anthocyanin was extracted from floral petals of wild balsam species and purified to homogeneity using chromatographic techniques. Evaluation of *in vitro* antidiabetic properties of anthocyanin extract revealed a dose-dependent increase in the inhibitory effect on the alpha-glucosidase (200 µg/ml) and alpha-amylase enzymes (500 µg/ml) and was comparable with the standard acarbose drug (189 µg/ml and 50 µg/ml). These results indicated that anthocyanin could be used as a source of functional food and nutraceuticals. This information from wild species will be useful in finding more potent antidiabetic principle from the natural resources for the clinical development of antidiabetic therapeutics. Future studies are planned to substantiate the antidiabetic power of anthocyanin using *in vivo* animal models.

Keywords: Alpha amylase, alpha glucosidase, diabetes, herbal remedies, *Impatiens balsamina* L.

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1. INTRODUCTION

Currently, million in the earth suffered from diabetes as per the public health¹. Further, the chance of diabetes is predicted to boom to 592 million by 2035. It is one of the life style disorders that causes increasing mortality and is a global health issue of which type 2 diabetes (T2D) accounting 90%². It featured by hyperglycemia, will cause a severe metabolic disorder in glucose, lipid and protein levels. Furthermore, type 2 diabetes is also connected with many disorders such as nephropathy and cardiovascular and cerebro-vascular diseases³. Most of the diabetics possess hyperlipidemia i.e., blood has too many lipids (or fats), such as cholesterol and triglycerides. Moreover, many documented the induction of oxidative stress related to diabetes. Hyperglycemia induces reactive oxygen species (ROS) and reactive nitrogen species (RNS) will damage the

pancreatic islet β -cell and their by insulin level. Insulin resistance and islet β -cell dysfunction are the critical reasons for diabetes, and therefore, mitigating oxidative stress to protection the cells may be an ideal way for T2D treatment. Nutraceutical along with drug therapy has been an effective strategy against diabetes.

Anthocyanins are flavonoid group of aqueous natural pigments exist in plants widely. They display antioxidant, anti-inflammatory, bactericidal, antiaging, antimetastatic, and protective effects on the liver, cardiovascular and cerebrovascular, and vision. Currently, synthetic drugs with antidiabetic effects like sulfonyleureas, biguanides, and α -glucosidase inhibitors groups are being used orally. The therapeutic support usage has multiple side effects like weight gain, hypoglycemia, gastrointestinal disturbances, liver and kidney damage, and hypersensitivity reactions

which may have a negative impact on the response to treatment⁴. The high cost of antidiabetic therapies together with the myriad of adverse effects prompted the exploration of cheaper and more effective antidiabetic biomolecules from medicinal herbals. Some researchers suggested that the anthocyanins extract inhibits alpha amylase action, indicating that the molecules have unique function to suppress the increase in postprandial glucose content from starch^{5, 6}. The present study is aimed to investigate *in vitro* antidiabetic activities of the purified anthocyanin of wild Balsam species based on local claims regarding its use in the treatment of diabetes.

2. MATERIALS AND METHODS

2.1 Plant materials

For the whole study, the fresh healthy flowers of *Impatiens balsamina* L. (wild) collected from the natural habitats of hilly regions of Trivandrum, Kerala.

2.2 Estimation of anthocyanin content

1g flower was homogenized with acidified methanol (1% HCl) and the anthocyanin content was quantified by the standard protocol of Sutharut and Sudarat⁷. The absorbance of each dilution was read at 510 and 700 nm against distilled water as blank.

2.3 Purification and characterization of anthocyanin from wild balsam

Anthocyanin from fresh flowers of wild Balsam was extracted by acidified methanol (100% Methanol: 1% HCl, 99:1, v/v) i.e., the petals were immersed in acidified methanol for overnight in dark at low temperature (4°C). Subsequently, the crude solution obtained was filtered through cheese cloth and concentrated in rotary vacuum evaporator under vacuum at temperature not exceeding 40°C. Then the aqueous extract was partitioned with ethyl acetate and the remaining water extract was concentrated in rotary vacuum evaporator, temperature not exceeding 40°C. It was further fractionated by using Sephadex LH-20 followed by C18 as separation matrices. The aqueous extract was loaded on to a Sephadex LH-20 using 100% methanol. Based on the TLC profile the fractions were subjected to purification by C-18 open-column chromatography using acidified water (1% HCl) and the column was eluted with a gradient of acidified methanol (1% HCl) and water [0:100 to 50:50]. From RP C-18 open-column chromatography, based on the TLC profile the pure fractions were concentrated and kept at low temperature (-20°C). Purified anthocyanin from the wild Balsam was subjected to NMR analysis for their structural identification.

2.4 *In vitro* antidiabetic activities

2.4.1 *In vitro* α-glucosidase inhibition assay

The α-amylase and α-glucosidase inhibitory effect of plant extracts was determined according to the protocol of Kim *et al*⁸. For alpha glucosidase inhibition, yeast α-glucosidase was dissolved in 100 mM, pH 7.0 phosphate buffer, containing bovine serum albumin 2 g/l and sodium azide 0.2 g/l which was used as source of enzyme. Substrate used was par-nitrophenyl-α-d-glucopyranoside. Extract was weighed and serial dilutions of 50, 100, 200, 400 and 500 µg/mL were made up with equal volumes of dimethylsulfoxide and distilled water. 10 µl of extract dilutions was incubated for 5 min with 50 µl enzyme source. After incubation, 50 µl of

substrate was added and again incubated for 5 min at room temperature. The pre-substrate and post -substrate addition absorbance was recorded at 405 nm on a microplate reader. The increase in OD on substrate addition was obtained. Each test was carried thrice and the mean OD was used to quantify the percentage of α-glucosidase inhibition. Acarbose was used as positive control with different doses. The dosage of acarbose and anthocyanin extract required to inhibit 50% of α-amylase activity under the conditions was noted as the IC₅₀ value.

Percentage α-glucosidase inhibition was calculated according to the formula:

$$\text{Percentage of inhibition} = \frac{[(\text{Control}_{405} - \text{Extract}_{405})]}{\text{Control}_{405}} \times 100$$

2.4.2 *In vitro* alpha amylase inhibition assay

The assay was performed following the method of Hansawasdi *et al*⁹. 2 mg Starch azure was suspended in 0.2 mL of Tris-HCl buffer (pH 6.9, 0.5 M) containing CaCl₂ (0.01 M, substrate). The tubes containing substrate solution were heated for 5 min and then pre-incubated at 37°C for 5 min. Purified anthocyanin extract from wild balsam species was dissolved in DMSO in order to obtain various doses of 50, 100, 200, 400 and 500 µg/mL. Anthocyanin extract of particular concentration was added to the tube containing the substrate solution. In addition, 0.1 mL of porcine pancreatic amylase in Tris-HCl buffer (2 U/mL) was also added. The reaction was carried out at 37°C for 10 min. The reaction was arrested by adding 0.5 mL of 50% acetic acid in each tube. The reaction mixture was centrifuged at 3000 rpm for 5 min at 4°C. The OD of resulting supernatant was recorded at 595 nm. Acarbose, a known α-amylase inhibitor was used as a standard drug. The experiments were repeated three times. The α-amylase inhibitory activity was calculated by using the following formula:

The α-amylase inhibitory activity =

$$\frac{[(A_{c+} - A_{c-}) - (A_s - A_b)]}{(A_{c+} - A_{c-})} \times 100,$$

where A_{c+}, A_{c-}, A_s, and A_b are the absorbance of 100% enzyme activity (only solvent with enzyme), 0% enzyme activity (only solvent without enzyme), a test sample (with enzyme), and a blank (a test sample without enzyme), respectively. The concentration of acarbose and anthocyanin extract required to inhibit 50% of α-amylase activity under the conditions was defined as the IC₅₀ value.

2.5 Statistical analysis

SPSS 18.0 statistical software was used for data analysis. Data from different assay treatments were analyzed by one-way analysis of variance (ANOVA); The Student-Neuman-Keuls post-hoc test was used to determine significance for individual experimental conditions. Differences with *P* < 0.01 or *P* < 0.05 were regarded as significant.

3. RESULTS AND DISCUSSION

3.1 Anthocyanin content, purification and fractionation

Anthocyanin content in the petals of wild Balsam species was quantified as 5.88 mg/g. Purified anthocyanins after 3 step protocol was subjected to NMR and LC MS analysis (Fig.1, 2, 3) for their structural identification. The major fractions were hesperidin, dimethoxy antirrhinin and trimethoxy antirrhinin (Fig. 4: A, B, C).

M-H



Fig. 1: (a) LC MS and (b) NMR spectrum of Hesperidin from the wild Balsam

M-H

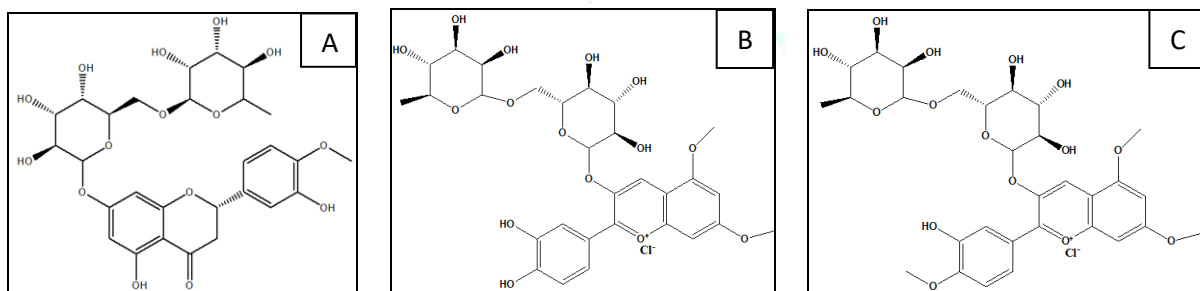


Fig. 2: (a) LC MS and (b) NMR spectrum of Dimethoxy antirrhinin from the wild Balsam

M-H



Fig. 3: (a) LC MS and (b) NMR spectrum of Trimethoxy antirrhinin from the wild Balsam.

Fig. 4: Chemical structure of (A) Hesperidin ($C_{28}H_{34}O_{15}$) (B) Dimethoxy antirrhinin ($C_{29}H_{35}ClO_{15}$) (C) Trimethoxy antirrhinin ($C_{30}H_{37}ClO_{15}$)

3.2 In vitro Antidiabetic Activity

3.2.1 In vitro α -amylase inhibitory activity

The *in vitro* α -amylase inhibitory potentiality of the purified anthocyanin extracts of wild balsam species was evaluated. The data displays a dose-dependent increase in the percentage of inhibitory activity against the α -amylase enzyme. The purified anthocyanin extract (50, 100, 200, 400 and 500 μ g/ml) of wild balsam exhibited potent α -amylase inhibitory activity from 24.2 to 80.7% with an IC_{50} value of 200 μ g/ml extract (Table 1). Acarbose, the standard drug for α -amylase inhibitor at a concentration of 50-500 μ g/ml, showed α -amylase inhibitory activity from 27.1 to 83.2% with an IC_{50} value 189 μ g/ml extract. So, it is possible to

suggest that the extract may be used as starch blockers since it inhibits or delays the starch absorption into the body mainly via blocking the lysis of 1,4-glycosidic linkages of starch and other oligosaccharides into maltose, maltotriose and other sugars. In the present study, the extract of wild balsam showed maximum α -amylase inhibitory activity (IC_{50} = 200 μ g/ml extract) which could be attributed by the purified anthocyanin (5.88 mg/g). It is possible to suggest that polyphenolics are not only capable of mitigating oxidative stress but also inhibits carbohydrate hydrolyzing enzymes due to their ability to bind with proteins. The obtained results were in accordance with the studies in the species such as Vietnamese edible plants, *Gymnema montanum*, and *Psidium guajava* leaves respectively^{10, 11, 12}.

Table 1. *In vitro* α -amylase inhibitory potentiality of the purified anthocyanin extracts of wild balsam species

Concentration ($\mu\text{g/mL}$)	Percentage inhibition	
	Anthocyanin	Standard: Acarbose
50	24.2	27.1
100	39.60	33.31
200	50.0	56.48
400	63.92	73.12
500	80.7	83.2

3.2.2 *In vitro* α -glucosidase inhibitory activity

Antidiabetic activity using α -glucosidase inhibitory assay of the purified anthocyanin extracts was narrated in the Table 2. The anthocyanin extract showed remarkable inhibitory

action of α -glucosidase enzyme. The percentage of inhibition at 50-500 $\mu\text{g/ml}$ concentrations of wild balsam anthocyanin extract showed a concentration dependent increase in inhibition.

Table 2. *In vitro* α -glucosidase inhibitory potentiality of the purified anthocyanin extracts of wild balsam species

Sample concentration ($\mu\text{g/mL}$)	Percentage inhibition	
	Anthocyanin	Standard: Acarbose
50	18.5	50.01
100	30.20	65.51
200	34.40	68.66
400	38.02	74.72
500	50.03	85.27

The percentage of inhibition varied from 18.5 – 50.03% for highest concentration to the lowest dose with an IC_{50} value 500 $\mu\text{g/ml}$. Thus the inhibition of the activity of α -glucosidase by anthocyanin would retard the carbohydrate degradation which in turn reduce the absorption of glucose, as a result the reduction of post-prandial blood glucose level elevation.

In this study, the standard drug acarbose for the α -glucosidase inhibitor at a concentration of 50-500 $\mu\text{g/ml}$ showed α -glucosidase inhibitory activity from 59.28 to 85.27% with an IC_{50} value 50 $\mu\text{g/ml}$. This indicates that the anthocyanin is potent α -amylase and α -glucosidase inhibitor and comparable with acarbose. Thus, there is a positive relationship between the polyphenol content and their ability to inhibit intestinal α -glucosidase and pancreatic α -amylase.

Lee *et al* screened antidementia acetylcholinesterase inhibitors from the fruits and designed an optimal extraction conditions¹³. *In vitro* antidiabetic and antioxidant activities of methanol extract of *Tinospora sinensis* was described¹⁴. Antihyperglycemic, antidiabetic, and antioxidant effects of *Garcinia pedunculata* in rats was also proved¹⁵. Different studies have reported that dietary polyphenols including flavonoids and phenolic acids may inhibit α -amylase and α -glucosidase which in turn inhibit glucose absorption in the intestine by sodium-dependent glucose transporter 1 (SGLT1), stimulate insulin secretion, and reduce hepatic glucose output. Polyphenols may also increase insulin-independent glucose uptake, activate 5' adenosine monophosphate-activated protein kinase (AMPK), modify the microbiome, and have anti-inflammatory effects. Detailed experimentation should be conducted to identify the exact mechanism. Some researchers showed that catechin treatment ameliorates diabetes and its complications in streptozotocin-induced diabetic rats¹⁶. Madhuri and Naik analyzed modulatory effect of garcinol in streptozotocin-induced diabetic Wistar rats¹⁷. Some studies documented *in vitro* antioxidant, antidiabetic, cholinesterase and tyrosinase inhibitory potential of fresh juice from *Citrus hystrix* and *C.*

maxima fruits¹⁸. *Ocimum* genera have been reported to have antidiabetic potentials. *O. basilicum*, *O. tenuiflorum*, *O. sanctum*, *O. canum* leaves have all been reported to exhibit strong inhibition against α -amylase and α -glucosidase¹⁹. Studies revealed the preventive effects of *Morus alba* anthocyanins on diabetes in Zucker diabetic fatty rats²⁰. It was proposed that inhibition of the activity of such alpha-amylase and alpha-glucosidase would delay the degradation of carbohydrate, which would in turn cause a decrease in the absorption of glucose, as a result the reduction of postprandial blood glucose level elevation²¹. The antidiabetic and antioxidative potential of the blue congo variety of purple potato extract in streptozotocin-induced diabetic rats was explained²². Sundarrajan *et al* studied *in-vitro* hypoglycemic and glucose uptake activity of isolated compound from ethanolic leaf extract of *Amaranthus tristis*²³. Researchers showed *in vitro* antidiabetic activity affecting glucose uptake in Hep G2 cells following their exposure to extracts of *Lauridia tetragon*²⁴. Antidiabetic effect of ethanolic leaf extract of *Phyllanthus amarus* in alloxan induced diabetic mice was studied²⁵. Recently, evaluated of hypoglycemic and antihyperglycemic effect of aqueous-methanolic leaves extract of two medicinal plants of Meghalaya in mice²⁶. Similarly, confirmed the antidiabetic and antiobesity effects of plant extract from *Cleome droserifolia*²⁷. The antidiabetic activity of *Terminalia catappa* leaf extracts in alloxan-induced diabetic rats was also proved²⁸.

Antidiabetic activity of selected herbals such as bark of *Eugenia jambolana*, the whole plants of *Phyllanthus niruri*, seeds of *Momordica charantia* and leaves of *Nyctanthes arbor-tristis* was investigated. The results revealed that the aqueous extract showed better efficacy than the ethanolic extracts in all the treated extract in lowering the blood glucose levels in alloxan induced diabetic rats²⁹. *In vitro* antioxidant and antidiabetic activity of methanolic fruit extract of medicinal plant *Piper cubeba* was evaluated. The treatment of the yeast cells with the methanolic extract revealed that the glucose uptake was found to increase in a dose dependent manner which amplifies its antidiabetic

potentiality. This probable attribute is due to the bioactive polyphenols and flavonoids in the extract³⁰. Thus, the obtained data in this study substantiates the antidiabetic possibilities of anthocyanins from the wild balsam species. This is the first report of *in vitro* antidiabetic activity of anthocyanin from the species.

4. CONCLUSION

The results of the present study showed that the wild balsam possess optimal level of anthocyanin content. *In vitro* alpha amylase and alpha glucosidase activity of anthocyanin from the *I. balsamina* floral extract was evaluated. Anthocyanin of Balsam species has been traditionally used as herbal medicine in folk remedies for curing multiple ailments. So, further *in vivo* studies are warranted to declare the anthocyanin of Balsam species as novel antidiabetic target for new drug designing.

Conflict of interest

The authors declare that they have no conflict of interest.

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