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Innovative Fe(III)-dosed Anaerobic Biological Treatment System: From Fe and S Biogeochemical Reactions to Engineering Process

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**Innovative Fe(III)-dosed Anaerobic Biological Treatment System:
From Fe and S Biogeochemical Reactions to Engineering Process**

Musfique Ahmed

**Dissertation submitted
to the Statler College of Engineering and Mineral Resources
at West Virginia University**

**in partial fulfillment of the requirements for the degree of
Doctor of Philosophy in Civil Engineering**

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Keywords: Iron-dosed anaerobic wastewater treatment, Fe/S ratio, Ferric and sulfate reduction rate, Organic oxidation, Iron reducers and sulfate reducers, Resource recovery from sludge, Nutrient recovery

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Abstract

Innovative Fe(III)-dosed Anaerobic Biological Treatment System: From Fe and S Biogeochemical Reactions to Engineering Process

By Musfique Ahmed

In moving toward more sustainable wastewater management, anaerobic treatment is gaining increasing popularity due to its simplicity, low energy requirement, low sludge production and less emission of greenhouse gases compared to typical aerobic wastewater treatment systems. Electron acceptors such as nitrate, sulfate, and CO_2 have been used in various anaerobic processes for removal of organic matters from wastewater under anoxic or anaerobic environments. In energy producing regions, ferric iron, Fe(III), is a predominant element in iron containing wastes such as acid mine drainage (AMD) and coal ash, which can potentially be used as a source of iron in novel anaerobic wastewater treatment. Such an iron-based treatment approach can offer multi-faceted benefits over existing treatment methods including use of iron-containing wastes, no aeration, unique reaction mechanisms for coagulation, sulfide control, organic micropollutant removal, and useful sludge byproducts. The overall goal of this research was to develop an innovative Fe(III)-dosed anaerobic wastewater treatment process through incorporating known and novel biogeochemical reactions of iron in an engineered biological system.

The major research objectives include (1) identifying the critical factors and investigating their effects on the treatment performance of Fe(III)-dosed wastewater treatment; (2) developing a continuous Fe(III)-dosed anaerobic biological treatment system and examining its technical feasibility and potential issues in long-term operations; (3) developing a method for transforming the sludge materials from the Fe(III)-dosed bioreactor into magnetic byproducts; and (4) exploring the applicability of this Fe(III)-dosed treatment method for nutrient removal and recovery.

A detail literature review was first conducted to evaluate the suitability of using iron reduction for wastewater treatment and identify critical factors affecting the treatment. Several factors were identified that affect organics oxidation coupled to iron reduction, including the types of the ferric compound, microorganisms, ferric bioavailability and availability of substrate. Amorphous iron materials (e.g. iron sludge from AMD) with large surface areas and high ferric dissolution rates have great potential to be used in Fe(III)-dosed wastewater treatment process to enhance ferric bioavailability to iron reducers. Given the significant levels of sulfate (SO_4^{2-}) in wastewater, sulfate reduction is expected to be co-occurring with iron reduction in the iron-dosed anaerobic treatment. Shift in microbial composition in relation to ferric and sulfate concentrations (expressed as Fe/S ratio) and their effects on organics removal are important knowledge gaps for developing such treatment technology. In particular, there is a need to understand the nature of the relationships between iron reducing bacteria (IRB) and sulfate reducing bacteria (SRB) (i.e., symbiotic or competitive) to identify optimal operating conditions for this type of wastewater treatment.

Batch experiments on iron-dosed anaerobic biological treatment of wastewater under three different molar Fe/S ratios (1, 2 and 3) showed positive correlation between organics (chemical oxygen demand, COD) oxidation rate and Fe/S ratio. Microbiological analysis suggested that both

iron reducers and sulfate reducers contributed to this organic oxidation. Maximum COD oxidation rate, V_{max} estimated from Michaelis-Menten model ranged from 0.47 mg/L·min to 1.09 mg/L·min as Fe/S ratio increased from 1 to 3. A positive correlation was also observed between COD oxidation rate and the relative abundance of iron reducers, and both increased with the Fe/S ratio.

Long-term continuous wastewater treatment using an anaerobic bioreactor dosed with ferric iron showed satisfactory COD removal of $84 \pm 4\%$, $86 \pm 4\%$ and $89 \pm 2\%$ under Fe/S molar ratio 0.5, 1 and 2 respectively. Fe/S ratio was also observed to regulate the effluent quality by removing excess sulfide from aqueous phase with increasing quantity of ferrous through ferrous sulfide precipitation. The sludge materials contained both biomass (20-40 w/w%) and inorganic precipitates (80-60 w/w%) with the inorganic fraction increasing with Fe/S ratio. Spectroscopic and chemical elemental analyses indicated that the inorganic fraction of the sludge materials mainly contained FeS and FeS₂. Microbiological analyses of the sludge materials identified *Geobacter* sp., *Geothrix* sp. and *Ignavibacteria* sp. as putative iron reducers, and *Desulfovibrio* sp., *Desulfobulbus* sp., *Desulfatirhabdium* sp., *Desulforhabdus* sp. and *Desulfomonile* sp. as putative sulfate reducers.

A simple thermal treatment method was applied to transform the iron sulfide sludge from the bioreactor into magnetic particles. Sludge samples were treated at five different temperatures (300, 350, 400, 450, and 500°C) to evaluate the transformation of iron sulfide sludge into different magnetic phases of iron oxide particles. X-ray Diffraction (XRD) analysis and magnetization measurements showed successful transformation of the sludge to magnetic byproducts and indicated the presence of ferromagnetic magnetite and maghemite phases at different temperatures. The magnetic sludge byproducts have potential applications in biomedicine sector and wastewater treatment (e.g. coagulant, adsorbent). Crystallinity and crystallite size of the thermally derived particles were observed to play a noteworthy role in regulating the magnetization of the byproducts. Adsorption study revealed that both samples baked at 350°C and 500°C had high adsorption capacities to remove phosphate from aqueous solutions.

A study to explore applicability of this Fe(III)-dosed treatment process for nutrient removal and recovery was conducted with synthetic wastewater containing typical concentrations of COD (420 mg/L), phosphate (10 mg/L), SO₄²⁻ (50 mg/L) and ammonium (50 mg/L). Average removal efficiencies of COD, phosphate, SO₄²⁻ and ammonium were $97 \pm 2\%$, $99.7 \pm 0.5\%$, $87.1 \pm 3\%$ and $20.3 \pm 9\%$ respectively. The results showed in addition to organics oxidation, significant phosphate and ammonium removals were achieved in the bioreactor. Potential removal mechanisms include chemical precipitation as ferric phosphate (FePO₄) or ferrous ammonium phosphate (FAP). SEM-EDS and XPS analysis suggested the presence of FAP in the sludge materials.

This innovative treatment process has shown consistent treatment performance and long-term stability under different operating conditions, suggesting its potential for large scale applications. Pilot-scale applications of this treatment approach using iron-containing wastes will give better understanding on the functionality of this process in a field scale environment. Utilizing iron wastes in this novel wastewater treatment process along with recovery of useful sludge byproducts not only can create new avenues to alleviate iron waste disposal, but also improve the sustainability of wastewater treatment.

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Table of Contents

Abstract	ii
Acknowledgements	iv
List of Figures	ix
List of Tables	xi
List of Symbols/ Nomenclature	xii
Chapter 1: Background and Research Objectives	1
1.1 Introduction.....	1
1.2 Applications of Iron in wastewater treatment.....	5
1.3 Ph.D. Research Rationale, Goal, and Objectives	6
1.3.1 Research Objectives	7
References	8
Chapter 2: Ferric Reduction in Organic Matter Oxidation and Its Applicability for Anaerobic Wastewater Treatment: A Review and Future Aspects	12
2.1 Introduction.....	12
2.2 Significance and fundamentals of iron reduction process.....	14
2.3 Organic oxidation by ferric reduction.....	15
2.3.1 Factors effecting the ferric reduction.....	15
2.3.2 Kinetics of ferric reduction in organics oxidation.....	17
2.3.3 Iron reduction process in environmental remediation	19
2.4 Microbial processes of iron reduction and ecology	21
2.4.1 Microbial processes of iron reduction	21
2.4.2 Roles of iron reducing bacteria	23
2.4.3 Diversity of iron reducing microorganisms	25
2.5 Iron reduction in wastewater treatment and future prospect	27
2.5.1 Organics mineralization by ferric reducers.....	28
2.5.2 Ferric iron bioavailability and bioreduction kinetics	29

2.5.3 Microbial ecology	30
2.5.4 Sample treatment process	31
2.6 Conclusion.....	31
References	32
Chapter 3: Effects of Fe/S Ratio on the Kinetics and Microbial Ecology of an Fe(III)-dosed Anaerobic Wastewater Treatment System	38
3.1 Introduction.....	38
3.2 Materials and Methods	40
3.2.1 Batch Bioreactors.....	40
3.2.2 Kinetic Study	41
3.2.3 Analytical Methods.....	41
3.2.4 Kinetics Modeling	42
3.2.5 Sludge Characterization.....	42
3.2.6 Microbiological Analyses.....	43
3.3 Results and Discussion	43
3.3.1 Oxidation Kinetics under different Fe/S ratios.....	43
3.3.2 Factors affecting COD oxidation	45
3.3.3 Sludge Characterization.....	46
3.3.4 Microbial community	49
3.4 Conceptual Framework.....	53
3.5 Conclusion.....	54
3.6 Supplementary Materials.....	55
References	57
Chapter 4: Continuous Ferric Iron-dosed Anaerobic Wastewater Treatment: Treatment Performance, Sludge Characteristics, and Microbial Composition	63
4.1 Introduction.....	63
4.2 Materials and Methods	65
4.2.1 Bench-scale iron-dosed treatment system.....	65

4.2.2 Treatment Experiments.....	66
4.2.3 Analytical Methods.....	67
4.2.4 Sludge Characterization.....	68
4.2.5 Microbiological Analysis.....	69
4.3. Results and Discussion.....	69
4.3.1 Treatment performance.....	69
4.3.2 Sludge Characterization.....	73
4.3.3 Microbiological Composition.....	76
4.4 Conclusion.....	80
4.5 Supplementary Materials.....	81
References.....	84
Chapter 5: Magnetic Sludge Byproducts for Adsorptive Removal of Phosphorus: Resource Recovery from Iron-based Anaerobic Sewage Sludge.....	90
5.1 Introduction.....	90
5.2 Materials and methods.....	93
5.2.1 Sample collection and preparation.....	93
5.2.2 Material Characterization.....	93
5.2.3 Magnetic Measurements.....	94
5.2.4 Adsorption Experiments.....	95
5.2.5 Adsorption Kinetics and Isotherm Modeling.....	95
5.3 Results and Discussion.....	96
5.3.1 Material Characterization.....	96
5.3.2 Magnetic Measurements.....	99
5.3.3 Adsorption Performance.....	101
5.4 Conclusion.....	105
5.5 Supplementary Materials.....	106
References.....	107

Chapter 6: Chemical Fate of Nutrients and Their Recovery Potential Through the Fe(III)-dosed Wastewater Treatment	113
6.1 Introduction.....	113
6.2 Material and Methods.....	116
6.2.1 Experimental Set-up	116
6.2.2 Analytical Methods.....	116
6.2.3 Sludge Characterization.....	117
6.3 Results and Discussion.....	117
6.3.1 Treatment Performance	117
6.3.2 Sludge Characterization.....	118
6.4 Conclusion.....	120
References	120
Chapter 7: Conclusions	124

List of Figures

Fig. 1: Major sources of wastewater	1
Fig. 2: Typical energy consumption in representative aerobic WWTP (adapted from WEF, 2009). Numbers in bracket represent the percentages of energy requirement in each treatment unit.....	3
Fig. 3: Different strategies of microbial and ferric surface interaction (a) Direct Contact, (b) Contact with ligand, (c) Contact with electron shuttle, (d) Contact with nanowires; adapted from Esther et al. (2015).....	22
Fig. 4: Oxidation of organic matters with ferric reduction adapted from Lovley (1993) ..	24
Fig. 5: Schematic diagram of the possible treatment process	31
Fig. 6: Nonlinear kinetic models of COD oxidation under three different Fe/S molar ratios	44
Fig. 7: Dissolved sulfide and ferrous iron concentrations in the bioreactors under different Fe/S ratios	45
Fig. 8: (a) Solid concentrations and (b) VSS/TSS and NVSS/TSS ratios of the sludge materials	47
Fig. 9: Sample SEM micrograph and EDS spectrum of the sludge material under Fe/S molar ratio 3.....	48
Fig. 10: XPS spectra of (a) Fe and (b) S in the sludge material under Fe/S molar ratio 3	49
Fig. 11: (a) Relative abundances of major microbial species and (b) diversity index.....	52
Fig. 12: Conceptual framework of the effects of Fe/S ratio on organic carbon oxidation kinetics.....	53
Fig. 13: (a) COD removal, sulfate reduction and iron retention and (b) COD removal rates under different Fe/S molar ratios	70
Fig. 14: (a) Effluent total iron, ferrous and sulfide concentration and (b) Influent and effluent pHs at different Fe/S molar ratios	71
Fig. 15: Mass flow rates of Fe and S throughout the biological treatment under Fe/S molar ratio 2.....	72
Fig. 16: VSS/TSS and NVSS/TSS as percentage (%) of sludge samples under different Fe/S ratios	73

Fig. 17: (a) SEM micrograph and (b) EDS spectrum for sludge sample under Fe/S molar ratio 2; (c) & (d): SEM images of cells present in anaerobic sludge	74
Fig. 18: XPS spectrum of (a) Fe2p and (b) S2p region of a sludge sample	75
Fig. 19: (a) Microbial distribution and (b) relative abundances of putative IRB and SRB	78
<i>Fig. 20: XRD spectra of the sludge byproducts derived from the thermal treatment. Red: XRD spectrum, Blue: Rietveld fit, Cyan: Background, Pink: Difference between the experimental and Rietveld refinement</i>	<i>97</i>
Fig. 21: Weight percentages of different phases of iron minerals derived from the sludge materials at different temperatures	99
Fig. 22: High-resolution hysteresis measurements of the sludge byproducts derived at different temperatures	100
Fig. 23: Remanent magnetization (Mr) of the sludge byproducts derived at different temperatures	101
Fig. 24: Adsorption kinetic data and pseudo-second-order kinetic models for (a) S350 and (b) S500	102
Fig. 25: Adsorption capacity of S350 for phosphate ions	103
Fig. 26: Adsorption isotherm data, and (a) Langmuir and (b) Freundlich models for S350 and S500 at pH 7.0.....	105
Fig. 27: Adaptation of the N- cycle in the Feammox reaction for NH ₄ ⁺ oxidation	115
Fig. 28: Influent and effluent concentrations of COD, NH ₄ ⁺ -N, PO ₄ ³⁻ -P and SO ₄ ²⁻	118
Fig. 29: SEM micrograph and EDS spectrum of the sludge material	119
Fig. 30: XPS spectra of the sludge sample for (a) Fe2p _{3/2} , (b) N1s, and (c) P2p _{3/2}	120

List of Tables

Table 1: Major iron reducing bacteria and their primary electron donors	27
Table 2: Major bacteria observed with their phyla and averaged relative abundance in all the bioreactors	50
Table 3: Daily loads of Fe and S for three target Fe/S molar ratios	67
Table 4: Major bacteria and their phylum identified in the bioreactor	77
Table 5: Langmuir and Freundlich Isotherm model parameters for S350 and S500	104

List of Symbols/ Nomenclature

AMD – Acid Mine Drainage

WWTP – Wastewater Treatment Plant

COD – Chemical Oxygen Demand

BOD – Biochemical Oxygen Demand

TSS – Total Suspended Solid

VSS – Volatile Suspended Solid

NVSS – Nonvolatile Suspended Solid

IRB – Iron Reducing Bacteria

SRB – Sulfate Reducing Bacteria

EPS – Extra Polymeric Substance

TOC – Total Organic carbon

SEM – Scanning Electron Microscope

EDS - Energy Dispersion Spectroscopy

XPS - X-ray Photoelectron Spectroscopy

VSM – Vibrating Sample Magnetometer

Chapter 1: Background and Research Objectives

1.1 Introduction

Wastewater originates from a variety of different sources including point sources such as domestic, industrial, commercial, agricultural and non-point sources such as groundwater infiltration, stormwater runoff and sewer overflows (Fig. 1). Untreated wastewater contains organic matters which can decompose and lead to nuisance conditions with production of malodorous gases. Additionally, it contains pathogenic microorganisms and nutrients that can have serious effect on human body and environment (Metcalf & Eddy et al., 2014). Therefore, proper treatment of wastewater is required before its discharge to the environment to protect public health and aquatic ecosystems.

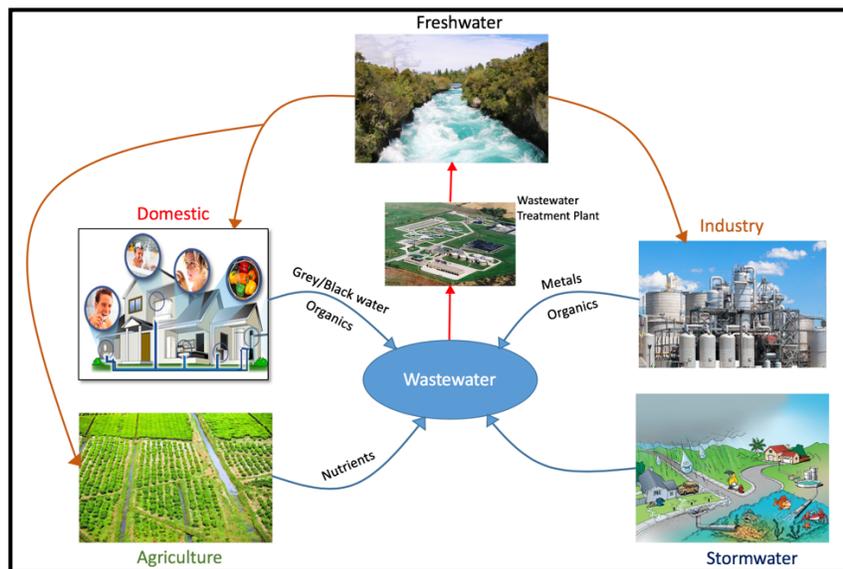


Fig. 1: Major sources of wastewater

Globally, most common wastewater treatment methods are centralized aerobic wastewater treatment plants and lagoons for both domestic and industrial wastewater (Doorn et al., 2006). Domestic wastewater may also be treated in decentralized septic systems which may treat wastewater from one or multiple households. Wastewater treatment plant (WWTP) are designed to satisfy different treatment objectives. Large particles, grits and rags are removed from wastewater by physical operation in primary treatment and remaining particulates are allowed to settle in primary clarifiers. Secondary treatment generally consists of biological and chemical processes that remove organic

matters (Metcalf & Eddy et al., 2014). In general, 85% of chemical oxygen demand (COD) and total suspended solid (TSS) can be removed by secondary treatment. Tertiary treatment is used in WWTPs to remove pathogens, contaminants and remaining nutrients such as nitrogen (N) and phosphorus (P) by using various advanced processes such as disinfection, carbon adsorption, filtration, ion exchange (Doorn et al., 2006; Metcalf & Eddy et al., 2014). Till late 1980s, conventional secondary treatment was the most common method for removal of biological oxygen demand (BOD) and TSS from wastewater. In the U.S., treatment for nutrient removal was used for special circumstances, such as great lakes area, Florida, and the Chesapeake Bay where nutrient related water quality problems were identified. However, with time nutrient removal processes have evolved and now are used more commonly. The changing nature of wastewater, emerging health and environmental concerns, treatment plant performance and reliability, new methods of process analysis and control, and impact of new regulations are some of the important factors that raise new concerns and directions in wastewater treatment (Metcalf & Eddy et al., 2014). New technologies and processes are explored and introduced by the scientists and authorities in wastewater treatment to make it more effective and reliable according to regional environment and setting.

Due to various operational and environmental characteristics such as pollutant loads, plant size, and type of treatment plant, energy consumption in WWTPs varies from 0.26 to 0.84 kWh per m³ of wastewater treated. The average energy consumption in WWTPs in the U.S., United Kingdom, Germany and Italy are 0.45, 0.64, 0.67 and 0.55 kWh/m³ respectively (Guerrini et al., 2017). Approximately 50-65% of the energy consumption is used in the activated sludge process which is the secondary treatment of typical WWTPs (Guerrini et al., 2017). Typical energy consumption of a representative wastewater treatment plant is illustrated in Fig. 2 (WEF, 2009). Typically, to treat wastewater of 500 mg/L COD, energy consumption in conventional activated sludge process is estimated as 3.20 kJ/g COD. On the other hand, average potential energy of typical domestic wastewater has been estimated as 16.2 kJ/g COD which is about five times the electrical energy required to operate wastewater treatment in conventional activated sludge process (Wan et al., 2016). This suggests that WWTPs can be energy self-sufficient if

proper energy recovery technology is implemented, but this can be expensive and less feasible.

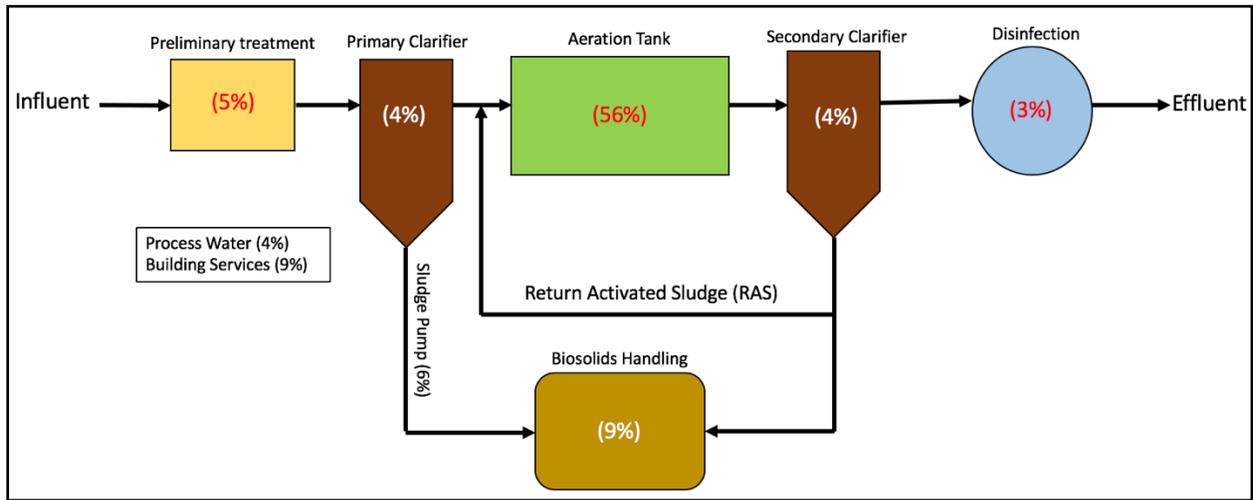


Fig. 2: Typical energy consumption in representative aerobic WWTP (adapted from WEF, 2009). Numbers in bracket represent the percentages of energy requirement in each treatment unit

Because of the high energy consumption in aerobic wastewater treatment, various alternative technologies have been developed to make WWTPs cost effective and energy efficient. Anaerobic treatment process is gaining immense popularity due to its simplicity, low energy requirement, low sludge production and less emission of greenhouse gases. The net operating cost is approximately \$160 per metric ton less for anaerobic process than aerobic process, which can be as high as \$250 for some instances (Speece, 1983). Various high rate anaerobic treatment reactors have been developed in last 40 years, including anaerobic filter (AF), upflow anaerobic sludge blanket (UASB), fluidized bed (FB), and anaerobic baffled reactor (ABR), which can contribute to significant organics removal from both domestic and industrial wastewater (Van Lier et al., 2015). In anaerobic microbiological process, anaerobic bacteria utilize organic matters as electron donor for metabolizing at oxygen-free environment with the help of different electron acceptors like CO₂, sulfate, nitrate (Damianovic and Foresti, 2007; Hubert and Voordouw, 2007; Jeong et al., 2008). Anaerobic treatment with methanogens is the most common treatment process where organics are converted to methane (CH₄) gas through bioconversion. Although, CH₄ is a well-known biogas, production of heat and electricity with this gas

require combined heat and power (CHP) system, which is complex and labor intensive (Mes et al., 2003). For on-site wastewater treatment at community level, treatment system needs to be designed with greater simplicity than centralized treatment systems. Several studies have been reported on the performance of sulfidogenic bioreactors where sulfate is utilized to treat sulfate-rich wastewater and significant organics removal has been observed (Dar et al., 2007; Deng et al., 2016; Oude Elferink et al., 1994; Widdel, 1988). There are opportunities to incorporate other redox active elements in anaerobic treatment and develop new field for research in the context of technology innovation.

An area of such opportunities is utilization of acid mine drainage (AMD) in wastewater treatment. AMD originating from active and abandoned mine, is a significant environmental liability in the mining regions worldwide. It affects the rivers, lakes, estuaries and coastal waters by various direct and indirect pathways. AMD is recognized as a multi-factor pollutant where the main factors are acidity, salinization, metal toxicity and sedimentation processes (Gray, 1997; Hill and Bates, 1979). In the Appalachian region of the U.S., approximately 12,000 km of rivers and streams are impacted by drainage from abandoned mines (Koryak et al., 2004). According to West Virginia Division of Environmental Protection (WVDEP), AMD has affected 484 streams for a total of 2,852 stream miles in West Virginia (USGS, 1997). Surface runoff polluted by iron (Fe), sulfur (S), acid and various heavy metals (copper, zinc, aluminum, manganese) has often leached away from the mines and discharged into the streams. Various active and passive treatment methods have been performed to neutralize the AMD impacts (Johnson and Hallberg, 2005). Although, active treatment can provide effective remediation of AMD, but it requires high operation costs and deals with large amount of sludge production (Chang et al., 2000). Passive biological methods for AMD treatment use wetlands, lagoons and bioreactors, but they require large land areas and higher construction costs (Neculita et al., 2007). In the last decade, co-treatment of AMD and municipal wastewater (MWW) was explored more extensively and recent developments on the co-treatment approach have created new avenues to utilize waste materials as green agent for wastewater treatment in energy producing regions (Deng and Lin, 2013; Hughes and Gray, 2013; Strosnider and Nairn, 2010). Co-treatment studies have shown significant removal of BOD, COD, nutrients and metals through aerobic mixing and

anaerobic biological reactions. As both sulfur (S) and iron (Fe) are prevalent elements of AMD and both are highly redox active elements, they are utilized for degradation of organics from wastewater.

Although numerous studies have been reported in literature which used sulfidogenic bioreactors to treat sulfate-rich wastewater (Dar et al., 2007; Deng and Lin, 2017; Widdel, 1988), not many studies focused on utilization of iron reduction in biological wastewater treatment. In addition to AMD, iron is predominant in many other iron containing waste materials such as coal fly ash, mill industries etc. These iron wastes can potentially be used as a source of iron for wastewater treatment.

1.2 Applications of Iron in wastewater treatment

Iron has been widely used in wastewater treatment for removal of pollutants chemically from wastewater. Various cheap ferric iron salts ($\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$) and ferrous iron salts (FeCl_2 , $\text{FeSO}_4 \cdot 6\text{H}_2\text{O}$) have been used for phosphorous (P) removal from wastewater (Fytianos et al., 1998; Waite, 2002; Zhang et al., 2015). The removal mechanisms include precipitation, co-precipitation, adsorption, formation of ferric-phosphate complexes, depending on the type and concentration of iron particles, and other factors (e.g., pH, ORP) (Takács et al., 2006). P removal from wastewater before discharge into rivers and lakes is very important as it can create eutrophication problem in receiving water bodies. Different types of iron oxide minerals including ferrihydrate ($\text{Fe}(\text{OH})_3$), goethite (FeOOH), hematite (Fe_2O_3), and magnetite (Fe_3O_4) showed varied adsorption capacity for phosphorus removal (Ajmal et al., 2018). In recent years, synthesis and utilization of various iron oxide nanoparticles have also been used in heavy metal and organic contaminant adsorption due to their extremely small size, high surface area to volume ratios and superparamagnetism (Xu et al., 2012).

Fenton is a well-recognized advanced oxidation method that uses iron for removing organic and inorganic compounds from wastewater. This process utilizes iron as catalyst to form active oxygen species like $\bullet\text{OH}$ that can oxidize the larger organics to smaller organics or completely mineralized them into carbon dioxide (CO_2) and water (Babuponnusami and Muthukumar, 2014, 2012; Wang et al., 2016). This process has shown high efficiency of organic removal from different types of wastewaters such as

those containing textile, pesticide, pharmaceutical, pulp mill, olive oil and phenolic compounds (Arslan-Alaton, 2007; Benatti et al., 2006; Bianco et al., 2011). Natural iron-bearing mineral materials, such as goethite, pyrite (FeS_2), hematite, magnetite, and ferrihydrite are used as the catalysts for the Fenton processes (Wang, Zheng, Zhang, & Wang, 2016). Fenton process has been modified by combining different photochemical and electrochemical phenomena to improve the performance of organic degradation.

Ferrous iron is also commonly used to reduce sulfide toxicity and odor problem of the wastewater. Iron can precipitate with sulfide as iron sulfide and precipitate out the soluble sulfide. This can reduce the toxicity to the microorganisms present in the biological treatment systems and improve the treatment efficiency (Davison and Heaney, 1978; Waite, 2002). Ferric iron can be used in anaerobic biological treatment to oxidize organic materials and reduced to ferrous iron by iron reducing bacteria (IRB). As $\text{Fe}^{3+}/\text{Fe}^{2+}$ reduction potential is comparatively high (+0.77 V at pH 2 and +0.2 V at pH 7) than other electron acceptors, IRB can use this energy to respire a wide range of organic compounds. In these redox reactions, ferric iron is utilized as an electron acceptor and organic materials are used as electron donor.

1.3 Ph.D. Research Rationale, Goal, and Objectives

Motivated by abundance of iron and potential of utilizing iron-containing wastes for wastewater treatment, the overall goal of this research is to develop a novel Fe(III)-dosed anaerobic biological wastewater treatment technology to create multi-faceted benefits (no requirement of aeration, simple design and operation, low sludge production, lower emission of CO_2 than aerobic process, unique reaction mechanisms for coagulation, sulfide control and organic pollutant removal, useful by-product from sludge materials).

In this research, a detailed literature review on the prospect and benefits of utilizing ferric iron in wastewater treatment was conducted (Chapter 2). This review covers all the important factors that can affect the reaction of ferric reduction coupled to organics oxidation, microbial ecology of IRB, and their interactions with other microorganisms (e.g., methanogens and SRB). Various knowledge gaps related to the application of ferric reduction in wastewater treatment were identified and research objectives were proposed to address the knowledge gaps. Ferric and Sulfate concentrations (expressed as Fe/S

ratio) have been identified as an important regulating parameter in this kind of Fe(III)-dosed treatment process. In typical wastewater, where organic concentration is abundant, concentration of ferric and sulfate may play the major role in regulating the abundance of IRB and SRB and their subsequent performance in organic oxidation. However, the effect of Fe/S ratio on the relationship of IRB and SRB in wastewater treatment is currently unknown. To design a treatment system like this, it is important to evaluate the performance based on the pollutant removal, kinetics and sludge production, that can help eventually for large scale application. Microbiological study can help to recognize the microbial communities of the system and their roles in pollutant oxidation. The sludge characterization is also an important part of this kind of treatment system, that can help to explore the possibility of resource recovery from the sludge materials. Transformation of the waste sludge into valuable materials and their application in different aspects can make this proposed treatment system more valuable and sustainable.

1.3.1 Research Objectives

The following research objectives are proposed to accomplish the research goal:

1. Identify critical factors and investigate their effects on the treatment performance of the iron-dosed wastewater treatment using batch bioreactors to develop optimal treatment conditions of the treatment method.
2. Develop a continuous Fe(III)-dosed wastewater treatment to investigate technical feasibility and potential issues in long-term operation of the treatment process.
3. Evaluate the potential of resource recovery from the sludge materials produced in this treatment process.
4. Investigate chemical fate of nutrients (ammonium and phosphate) and potential of their recovery from wastewater through the Fe(III)-dosed treatment.

The first research objective is set to identify the major aspects of this Fe(III)-dosed treatment process, specifically the organic oxidation kinetics, microbial distribution, sludge production and their interrelations. This can ultimately help to design the treatment system for long term operation with the goal of large-scale application (Chapter 3). The second objective is expected to investigate the feasibility of the treatment system during

long-term operation with a proto-type treatment system. These results can show the influence of Fe/S ratio on four aspects (organic removal, organic oxidation rate, sludge characterization, microbial composition) of the treatment performance (Chapter 4). The third research objective is expected to explore the opportunity of extracting valuable materials like magnetite, maghemite from the sludge materials and their contribution in removing pollutants from wastewater. This can help to transform the sludge waste into useful resource (Chapter 5). The last objective is set to explore the applicability of this novel Fe(III)-dosed treatment process for nutrient removal and recovery, which will assist to remove both organics and nutrient together with the same treatment system (Chapter 6).

References

- Ajmal, Z., Muhmood, A., Usman, M., Kizito, S., Lu, J., Dong, R., Wu, S., 2018. Phosphate removal from aqueous solution using iron oxides: Adsorption, desorption and regeneration characteristics. *J. Colloid Interface Sci.* 528, 145–155. <https://doi.org/10.1016/j.jcis.2018.05.084>
- Arslan-Alaton, I., 2007. Degradation of a commercial textile biocide with advanced oxidation processes and ozone. *J. Environ. Manage.* 82, 145–154. <https://doi.org/10.1016/j.jenvman.2005.12.021>
- Babuponnusami, A., Muthukumar, K., 2014. A review on Fenton and improvements to the Fenton process for wastewater treatment. *J. Environ. Chem. Eng.* 2, 557–572. <https://doi.org/10.1016/j.jece.2013.10.011>
- Babuponnusami, A., Muthukumar, K., 2012. Advanced oxidation of phenol: A comparison between Fenton, electro-Fenton, sono-electro-Fenton and photo-electro-Fenton processes. *Chem. Eng. J.* 183, 1–9. <https://doi.org/10.1016/j.cej.2011.12.010>
- Benatti, C.T., Tavares, C.R.G., Guedes, T.A., 2006. Optimization of Fenton's oxidation of chemical laboratory wastewaters using the response surface methodology. *J. Environ. Manage.* 80, 66–74. <https://doi.org/10.1016/j.jenvman.2005.08.014>
- Bianco, B., Michelis, I. De, Vegliò, F., 2011. Fenton treatment of complex industrial wastewater: Optimization of process conditions by surface response method. *J. Hazard. Mater.* 186, 1733–1738.
- Chang, I.S., Shin, P.K., Kim, B.H., 2000. Biological treatment of acid mine drainage under sulphate-reducing conditions with solid waste materials as substrate. *Water Res.* 34, 1269–1277. [https://doi.org/10.1016/S0043-1354\(99\)00268-7](https://doi.org/10.1016/S0043-1354(99)00268-7)
- Damianovic, M.H.R.Z., Foresti, E., 2007. Anaerobic degradation of synthetic wastewaters at different levels of sulfate and COD/sulfate ratios in horizontal-flow anaerobic reactors (HAIB). *Environ. Eng. Sci.* 24, 383–393.

- Dar, S.A., Yao, L., Van Dongen, U., Kuenen, J.G., Muyzer, G., 2007. Analysis of diversity and activity of sulfate-reducing bacterial communities in sulfidogenic bioreactors using 16S rRNA and *dsrB* genes as molecular markers. *Appl. Environ. Microbiol.* 73, 594–604. <https://doi.org/10.1128/AEM.01875-06>
- Davison, W., Heaney, S.I., 1978. Ferrous iron-sulfide anoxic hypolimnetic interactions waters. *Limnol. Oceanogr.* 23, 1194–1200.
- Deng, D., Lin, L.S., 2017. Continuous sulfidogenic wastewater treatment with iron sulfide sludge oxidation and recycle. *Water Res.* 114, 210–217. <https://doi.org/10.1016/j.watres.2017.02.048>
- Deng, D., Lin, L.S., 2013. Two-stage combined treatment of acid mine drainage and municipal wastewater. *Water Sci. Technol.* 67, 1000–1007. <https://doi.org/10.2166/wst.2013.653>
- Deng, D., Weidhaas, J.L., Lin, L.-S., 2016. Kinetics and microbial ecology of batch sulfidogenic bioreactors for co-treatment of municipal wastewater and acid mine drainage. *J. Hazard. Mater.* 305, 200–8. <https://doi.org/10.1016/j.jhazmat.2015.11.041>
- Doorn, M.R.J., Towprayoon, S., Vieira, S.M.M., Irving, S.W., Palmer, C., Pipatti, R., Wang, C., 2006. Wastewater Treatment and Discharge, 2006 IPCC Guidelines for National Greenhouse Gas Inventories.
- Fytianos, K., Voudrias, E., Raikos, N., 1998. Modelling of phosphorus removal from aqueous and wastewater samples using ferric iron. *Environ. Pollut.* 101, 123–130. [https://doi.org/10.1016/S0269-7491\(98\)00007-4](https://doi.org/10.1016/S0269-7491(98)00007-4)
- Gray, N.F., 1997. Environmental impact and remediation of acid mine drainage: A management problem. *Environ. Geol.* 30, 62–71. <https://doi.org/10.1007/s002540050133>
- Guerrini, A., Romano, G., Indipendenza, A., 2017. Energy efficiency drivers in wastewater treatment plants: A double bootstrap DEA analysis. *Sustainability* 9, 1–13. <https://doi.org/10.3390/su9071126>
- Hill, R.D., Bates, E.R., 1979. Acid mine drainage and subsidence: effects of increased coal utilization. *Environ. Health Perspect.* Vol. 33, 177–190.
- Hubert, C., Voordouw, G., 2007. Oil field souring control by nitrate-reducing *Sulfurospirillum* spp. that outcompete sulfate-reducing bacteria for organic electron donors. *Appl. Environ. Microbiol.* 73, 2644–2652. <https://doi.org/10.1128/AEM.02332-06>
- Hughes, T.A., Gray, N.F., 2013. Co-treatment of acid mine drainage with municipal wastewater: Performance evaluation. *Environ. Sci. Pollut. Res.* 20, 7863–7877. <https://doi.org/10.1007/s11356-012-1303-4>
- Jeong, T.Y., Cha, G.C., Seo, Y.C., Jeon, C., Choi, S.S., 2008. Effect of COD/sulfate ratios on batch anaerobic digestion using waste activated sludge. *J. Ind. Eng. Chem.* 14, 693–697. <https://doi.org/10.1016/j.jiec.2008.05.006>

- Johnson, D.B., Hallberg, K.B., 2005. Acid mine drainage remediation options: A review. *Sci. Total Environ.* 338, 3–14. <https://doi.org/10.1016/j.scitotenv.2004.09.002>
- Koryak, M., Stafford, L.J., Reilly, R.J., 2004. Declining intensity of acid mine drainage in the northern Appalachian bituminous coal fields: Major Allegheny River tributaries. *J. Am. Water Resour. Assoc.* 40, 677–689. <https://doi.org/10.1111/j.1752-1688.2004.tb04452.x>
- Mes, T.Z.D. de, Stams, A.J.M., Reith, J.H., Zeeman, G., 2003. Methane production by anaerobic digestion of wastewater and solid wastes, in: Reith, J.H., Wijffels, R.H., Barten, H. (Eds.), *Biomethane and Biohydrogen. Status and Perspectives of Biological Methane and Hydrogen Production.* pp. 58–94. <https://doi.org/10.1016/j.biortech.2010.08.032>
- Metcalf & Eddy, I., Tchobanoglous, G., Stensel, H.D., Tsuchihashi, R., Burton, F., 2014. *Wastewater Engineering: Treatment and Resource Recovery.* McGraw-Hill, New York.
- Neculita, C.-M., Zagury, G.J., Bussi re, B., 2007. Passive Treatment of Acid Mine Drainage in Bioreactors using Sulfate-Reducing Bacteria. *J. Environ. Qual.* 36, 1. <https://doi.org/10.2134/jeq2006.0066>
- Oude Elferink, S.J.W.H., Visser, A., Pol, L.W.H., Stams, A.J., 1994. Sulfate reduction in methanogenic bioreactors. *FEMS Microbiol. Rev.* 15, 119–136.
- Speece, R.E., 1983. Anaerobic biotechnology for industrial wastewater treatment. *Environ. Sci. Technol.* 17, 416A-427A. <https://doi.org/10.1021/es00115a001>
- Strosnider, W.H., Nairn, R.W., 2010. Effective passive treatment of high-strength acid mine drainage and raw municipal wastewater in Potos , Bolivia using simple mutual incubations and limestone. *J. Geochemical Explor.* 105, 34–42. <https://doi.org/10.1016/j.gexplo.2010.02.007>
- Tak cs, I., Murthy, S., Smith, S., McGrath, M., 2006. Chemical phosphorus removal to extremely low levels: Experience of two plants in the Washington, DC area. *Water Sci. Technol.* 53, 21–28. <https://doi.org/10.2166/wst.2006.402>
- USGS, 1997. *U.S. Geological Survey Programs in West Virginia.* Charleston.
- Van Lier, J.B., Van der Zee, F.P., Frijters, C.T.M.J., Ersahin, M.E., 2015. Celebrating 40 years anaerobic sludge bed reactors for industrial wastewater treatment. *Rev. Environ. Sci. Biotechnol.* 14, 681–702. <https://doi.org/10.1007/s11157-015-9375-5>
- Waite, T.D., 2002. Challenges and opportunities in the use of iron in water and wastewater treatment. *Rev. Environ. Sci. Biotechnol.* 1, 9–15. <https://doi.org/10.1023/A:1015131528247>
- Wan, J., Gu, J., Zhao, Q., Liu, Y., 2016. COD capture: A feasible option towards energy self-sufficient domestic wastewater treatment. *Sci. Rep.* 6, 1–9. <https://doi.org/10.1038/srep25054>
- Wang, N., Zheng, T., Zhang, G., Wang, P., 2016. A review on Fenton-like processes for organic wastewater treatment. *J. Environ. Chem. Eng.* 4, 762–787.

<https://doi.org/10.1016/j.jece.2015.12.016>

WEF, 2009. Energy Conservation in Water and Wastewater Treatment Facilities. McGraw-Hill, Alexandria.

Widdel, F., 1988. Microbiology and ecology of sulfate and sulfur-reducing bacteria, in: Zehnder, A. (Ed.), *Biology of Anaerobic Microorganisms*. Wiley, New York, pp. 459–586.

Xu, P., Zeng, G.M., Huang, D.L., Feng, C.L., Hu, S., Zhao, M.H., Lai, C., Wei, Z., Huang, C., Xie, G.X., Liu, Z.F., 2012. Use of iron oxide nanomaterials in wastewater treatment: A review. *Sci. Total Environ.* 424, 1–10. <https://doi.org/10.1016/j.scitotenv.2012.02.023>

Zhang, Z., Wang, Y., Leslie, G.L., Waite, T.D., 2015. Effect of ferric and ferrous iron addition on phosphorus removal and fouling in submerged membrane bioreactors. *Water Res.* 69, 210–222. <https://doi.org/10.1016/j.watres.2014.11.011>

Chapter 2: Ferric Reduction in Organic Matter Oxidation and Its Applicability for Anaerobic Wastewater Treatment: A Review and Future Aspects

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2.1 Introduction

A wide range of treatment technologies have been developed and explored to dwindle the impacts of wastewater generated from household and industries on aquatic ecosystems. Although aerobic treatment process has been used worldwide to achieve high degree of treatment efficiency, anaerobic wastewater treatment has been gaining increasing attention because of its simplicity, energy efficiency, low sludge production (McCarty and Smith, 1986; Speece, 1983; Van Lier, 2008; Van Lier et al., 2015). Anaerobic treatment has distinct advantages over aerobic treatment when treating high concentrated organic wastewater including emission of significantly less amounts of greenhouse gas (Chan et al., 2009). Additional benefits such as lower capital, operational and maintenance costs and technological advancement of resource recovery have helped to justify the use of anaerobic treatment over aerobic treatment (Chan et al., 2009; Manariotis and Grigoropoulos, 2002). Currently, anaerobic treatment systems have not been efficaciously in practice around the world because of inaptness of their process and inadequate capacity of the reactors to treat great amount of water (Saravanane and Murthy, 2016). But, with recent scientific advancements in microbiological processes and environmental biotechnologies, these shortcomings are readily to be overcome if have not already been. In particular, continuous development in anaerobic treatment has led to the applications of different high-rate reactor systems. Among those, anaerobic fluidized bed reactor (AFBR), fixed film reactor, anaerobic baffled reactor (ABR) and upflow anaerobic sludge blanket (UASB) have demonstrated impressive organic removal rates (Chan et al., 2009; Saravanane and Murthy, 2016).

Given the reducing conditions, opportunities exist for innovative approaches of incorporating natural redox active elements in anaerobic treatment systems. In this

context, co-treatment of acid mine drainage (AMD) and municipal wastewater (MWW) is an example of an innovative treatment approach. It was first explored in the early twentieth century to reduce pathogens by low pHs and elevated metal concentrations in AMD (Roetman, 1932). Recent developments of this co-treatment approach have produced impressive results of mitigating AMD and reduction of organic materials from wastewater (Deng and Lin, 2013; Hughes and Gray, 2013; Strosnider et al., 2011). Strosnider et al. (2011) studied a co-treatment of synthetic AMD and municipal wastewater to reduce the biological oxygen demand (BOD), nutrients and metals in a four stage passive process. A two stage co-treatment consisting of aerobic mixing and anaerobic biological treatment was recently tested and has shown significant chemical oxygen demand (COD) removal and sulfate reduction (>80%) under COD/sulfate ratios ranging from 0.6 to 5.4 in anaerobic biological reactors (Deng and Lin, 2013). The two stage treatment also resulted in an average 75% reduction of a range of metals and excellent phosphorus removal. The kinetics and microbial ecology of the biological system were later quantified and characterized, with both affected by COD/sulfate ratio and iron concentrations (Deng et al., 2016). Hughes and Gray (2013) examined a co-treatment process by injecting AMD in different forms of organic wastes (i.e., raw AMD, pre-treated AMD and pre-treated AMD with screened MWW) and reported substantial removal of COD, BOD, total organic carbon (around 90%) with continuous loading of AMD into an activated sludge reactor.

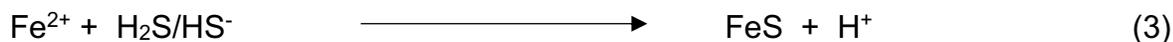
Given high levels of sulfate in AMD, treatment of AMD has mainly focused on using sulfate reducing bacteria (SRB) to reduce sulfate to (bi)sulfide and metal sulfide precipitation by providing organics as electron donors. Iron is also a prevalent element in AMD as well as in a wide range of iron containing wastes. While numerous studies have been conducted to evaluate the performance of sulfidogenic bioreactors for treating sulfate-rich wastewater (Dar et al., 2007; Oude Elferink et al., 1994; Widdel, 1988), only limited studies on iron amendment and its effects on sulfidogenic treatment can be found in literature. The studies examining iron reduction processes and iron reducers in the literature mostly focused on sediment and soil systems (Jensen et al., 2003; Lovley, 1987; Roden and Wetzel, 2002; Urrutia et al., 1998) and studies related to wastewater is extremely scarce.

This review focuses on fundamentals of iron reduction process in organics oxidation and potential utilities of iron reducing bacteria (IRB) for removing organic matters from wastewater. Specifically, significance of ferric reduction process and its applicability in oxidizing organic matters are reviewed. This review also covers applications of iron reduction process in environmental remediation and microbial ecology of IRB and their interactions with other microorganisms such as SRB and methanogens. Knowledge gaps in applying iron reduction in wastewater treatment are identified and its future research scope is discussed.

2.2 Significance and fundamentals of iron reduction process

A main feature of anaerobic/anoxic wastewater treatment is the use of alternative electron acceptors other than oxygen for organics oxidation. In such environments, metabolic energy is supplied by oxidation of organic carbons and reduction of ions like sulfate, nitrate, and ferric ions. Microbial reducers utilizing these alternative electron acceptors such as SRB, IRB, denitrifying bacteria can provide diverse metabolic pathways for oxidation of organic and inorganic wastes. Given its abundance in the environment, reduction of ferric iron has long been recognized as an important biologically-mediated process with significant influence on the fate of organic and inorganic pollutants.

The fundamental iron reduction process involves $\text{Fe}^{3+}/\text{Fe}^{2+}$ redox pair in which ferric ion is reduced to ferrous ion by receiving an electron from an electron donor. Dissimilatory IRB gain their energy for their metabolic functions and growth. This reduction half reaction (i.e., $\text{Fe}^{3+} + e^- \rightarrow \text{Fe}^{2+}$) has a standard redox potential of +0.77 V at pH 2 and +0.2 V at pH 7 (Madigan et al., 2015). However, this potential can vary from -1 V to +1 V due to the instability of complex compounds and dissimilar stability constants (Pierre et al., 2002). As $\text{Fe}^{3+}/\text{Fe}^{2+}$ reduction potential is comparatively high than other electron acceptors, ferric reducers can use this energy to respire a wide range of organic compounds. As a result, large number of organic materials can be used as a substrate by IRB as a mechanism of decomposing the organic matters. In the case of sulfate rich wastewater, ferric reduction and sulfate reduction can occur concurrently (Eq. 1 & Eq. 2) and form ferrous sulfide (FeS) precipitation (Eq. 3).



Previous studies have shown various Fe^{3+} compounds as electron acceptors are effective for organic matter mineralization in groundwater, soils and sediments (Lovley, 1995, 1987). This ferric to ferrous transformation can be an effective mechanism for organic oxidation in addition to its contribution to iron cycle in aquatic environments. For example, sediments found with comparatively higher concentration of Fe^{3+} than the other electron acceptors have the potential to use iron as oxidant to mineralize organic matters with releasing nutrients such as phosphate and trace metals adsorbed on ferric oxyhydroxides. In such sediment systems, most of the organic carbon has been found to be retained in the fermentation products (Lovley and Phillips, 1986). With amorphous ferric as terminal electron acceptor, major fermentation products of acetate could be oxidized (Kamura et al., 1963).

Another oxidation pathway involving ferric reduction is known as ferric ammonium oxidation (Feammox). In strict anaerobic condition, ferric reduction has been found to be an effective process to oxidize ammonium (Clément et al., 2005; Sawayama, 2006; Yang et al., 2012). Sawayama (2006) reported the evidence of ammonium oxidation to nitrate by IRB with Fe^{3+} EDTA as an oxidizing agent. This process was also observed to be thermodynamically feasible by Clément et al. (2005) by using goethite as the iron source for ferric reduction in wetland soils. All these examples substantiated the significance of ferric reduction process in remedying environmental pollutants.

2.3 Organic oxidation by ferric reduction

2.3.1 Factors effecting the ferric reduction

A number of factors have been reported to affect organics oxidation in iron reduction process, including the types of the ferric compound, microorganisms, and availability of substrate (Lovley, 1987). In general, increasing degree of crystallinity of the iron source materials results in lower iron reduction. Less crystalline or amorphous compounds have larger surface areas and higher solubility rate compared to highly crystalline ferric

materials (Lovley, 1987). A sequence of preference had been established on different ferric compounds as $\text{FePO}_4 \cdot 4\text{H}_2\text{O} > \text{Fe}(\text{OH})_3 > \gamma\text{-FeOH} > \alpha\text{-FeOH} > \text{Fe}_2\text{O}_3$ with respect to their microbial reduction rate (Munch and Ottow, 1983). This sequence corroborated on the hypothesis of decrease in microbial reduction rate with high crystallinity. Solubility of ferric compounds is another major factor to be considered in ferric reduction process. Most of the ferric compounds are highly insoluble and found as solid forms in nature. Due to this reason, ferric reduction may be hindered in sediments by other reducers such as denitrifiers, sulfate reducers, and methanogens that use highly soluble substrates (i.e., nitrate, sulfate, and carbon dioxide). Because of the low solubility of Fe^{3+} compounds, IRB may require direct contact with the surface of compounds to achieve the reduction process (Lovley, 1987). In this regard, addition of Fe^{3+} chelating agents (e.g. Fe^{3+} NTA, Fe^{3+} citrate) can increase bacterial iron reduction (Lovley and Phillips, 1988) and enhance ferric reduction rate of the cultures (Arnold et al., 1986; Jones et al., 1983).

Ferric reduction process has also been found to be pH dependent. Percentage of iron reduction was decreased when the seawater-nutrient medium was acidified and pH was decreased (De Castro and Ehrlich, 1970). With soil humic acid as a substrate, Chen et al. (2003) reported slower and lower ferric reduction capacity under a low pH condition (< 4), and an improved reduction capacity with a higher pH (> 4).

Different pure ferric reducing cultures have been utilized in carbon oxidation studies (Arnold et al., 1988; Hyun et al., 1999; Lonergan et al., 1996; Lovley, 1995). These microorganisms gain their energy by the oxidation of organic materials or other available substrates via extracellular iron reduction, a process which is known as 'extracellular respiration' (Esther et al., 2015). *Geobacter metallireducens*, also known as GS-15 was first isolated and used as the model ferric reducing microorganism in several studies (Lovley, 1995; Lovley and Lonergan, 1990; Lovley and Phillips, 1988). This iron reducer was found to generate different biogenic iron products with different ferric compounds. For example, with acetate as the electron donor, amorphous ferric oxide was reduced to magnetite (Fe_2O_3), and ferric citrate was reduced to ferrous compound. *Shewanella Sp.* has also been identified as an important and effective ferric reducer (Hyun et al., 1999; Lovley, 1993; Nealson and Myers, 1990). The direct contact between the cells of

microorganism and solid surface might be required for the ferric oxyhydroxide reduction by *Shewanella putrefaciens*, which reveals the occurrence of ferric reduction at membrane-bound sites (Arnold et al., 1988; Beliaev and Saffarini, 1998). A detail section has been presented on the microbial iron reduction process and ecological diversity of these iron reducers in later part of this paper.

The effectiveness of Fe^{3+} reduction process in organic matter decomposition also depends on the competition of IRB with SRB and methanogens for electron donors. With amorphous ferric oxide added to sediments, Fe^{3+} reduction process was observed to inhibit sulfate reduction by 90% and methane production by 50-90% where sulfate reduction and methane production was the major terminal electron acceptors respectively (Lovley and Phillips, 1987). In sediments, IRB can divert the electron flow away from SRB and methanogens and maintain low concentration of major substrates. As a result, SRB and methanogens do not get the enough substrate for metabolizing. Thermodynamically, ferric iron is a more favorable electron acceptor than sulfate and CO_2 (Madigan et al., 2015), causing this electron flow diversion. In some cases, however, ferric reduction was not found to hinder the sulfate reduction and methane production with sufficient substrate available in the environment (Lovley and Phillips, 1987) and co-existence of ferric reduction and sulfate reduction were observed (Acht nich et al., 1995; Thomsen et al., 2004). Distinct to sediments, wastewater contains high amount of organic materials and availability of substrate may not be a limiting factor for sulfate reduction and methane production. However, evidences have been found where ferric reduction process inhibited the sulfate reduction and methane production in wastewater (Zhang et al., 2009). Utgikar et al. (2002) suggested that precipitation of metal sulfide on the surface of sulfate reducers and methanogens might be a possible reason behind such inhibition. But, the authors recommended further research at cellular and enzymatic levels to validate this hypothesis.

2.3.2 Kinetics of ferric reduction in organics oxidation

Most of the studies examining the kinetics of ferric reduction in organic matter oxidation focused on aquatic and freshwater sediment systems (Bonneville et al., 2004; Jensen et al., 2003; Roden and Wetzel, 2002; Thamdrup, 2000). Bioavailability of amorphous or

poorly crystalline Fe^{3+} is an important factor and their concentration has been used as a variable for quantifying the ferric reduction rate as well as a predictor of the contribution of Fe^{3+} reduction to carbon mineralization (Lovley and Phillips, 1987, 1988). Jensen et al. (2003) quantified the ferric reduction rate in a marine sediment system and found that the organics mineralization was highly correlated with the concentration of poorly crystalline Fe^{3+} . The authors then justified the use of Fe^{3+} concentration for estimating the ferric reduction rate. The study assumed only iron and sulfate reductions contributed to anaerobic carbon oxidation and the individual reduction rates were combined to derive a ferric reduction rate expression.

The Monod model has been used to characterize microbial ferric reduction by several iron reducers. Liu et al. (2001) studied goethite ($\alpha\text{-FeOOH}$) reduction rate with lactate as the electron donor using a *S. putrefaciens* culture. The ferric bioreduction rate and extent were quantified with respect to the electron donor and electron acceptor. The initial ferric reduction rate was found to increase with increasing goethite and lactate concentrations. The authors used FeOOH sorption capacity for Fe(II) as a surrogate measure of the electron acceptor concentration to normalize the Fe(III) bioreduction rate. The normalized bioreduction rate was fairly constant ($\sim 0.027 \pm 0.0023 \text{ h}^{-1}$) over varied FeOOH concentrations, indicating a first-order relationship of the ferric bioreduction with the goethite surface area for the concentration range examined. Whereas, the normalized bioreduction rate was well characterized by the Monod rate expression as a function of the lactate concentration with the maximum rate $0.029 \pm 0.002 \text{ h}^{-1}$ and half-saturation constant $0.52 \pm 0.1 \text{ mM}$. Another Fe(III)EDTA - reduction study has been done by Van der Maas (2005) in BioDeNOx reactors with different electron donors. The highest reduction rate has been observed by glucose ($13.9 \text{ mM}\cdot\text{h}^{-1}$) followed by ethanol ($8.2 \text{ mM}\cdot\text{h}^{-1}$), acetate ($5.1 \text{ mM}\cdot\text{h}^{-1}$), hydroge ($5.1 \text{ mM}\cdot\text{h}^{-1}$) and methanol ($4.1 \text{ mM}\cdot\text{h}^{-1}$). So, the ferric reduction rate can fluctuate respect to the type of electron donors.

Bonneville et al. (2004) investigated the changes in ferric reduction rate with different forms of ferric oxyhydroxide with lactate as an electron donor. Using the Michaelis-Menten kinetics with Fe^{3+} concentration as the substrate for the rate expression, the maximum reduction rate (V_{max}) varied from 0.2×10^{-11} to $399 \times 10^{-11} \mu\text{mol h}^{-1} \text{ cell}^{-1}$. The

highest rates were observed with ferric citrate as the reductant and the lowest rate was obtained with hematite. A positive correlation was observed between the solubility of the ferric hydroxides with the maximum ferric reduction rate. Postma (1993) showed that the kinetic reactivity of iron oxides can be distinctive from each other by comparing the reduction rate of ferrihydrate with reductive dissolution of hematite and goethite. At same pH, the ferric reduction rate of ferrihydrate had been observed much higher than hematite and goethite. At pH 3, the initial reduction rate of ferrihydrate was $1.2 \times 10^{-8} \text{ mol s}^{-1} \text{ m}^{-2}$ whereas the initial reduction rate was $6.1 \times 10^{-11} \text{ mol s}^{-1} \text{ m}^{-2}$ for hematite and $1.8 \times 10^{-11} \text{ mol s}^{-1} \text{ m}^{-2}$ for goethite (Banwart et al., 1989; Zinder et al., 1986). It was concluded that ferrihydrate dissolved much faster than hematite and goethite, resulting in 180 times faster reaction rate than hematite and 100 times faster than goethite.

Roden and Wetzel (2002) used a first-order rate expression to model Fe^{3+} oxide concentration as a function of time in sediments from a freshwater wetland system. The rate expression contained a fraction of ferric oxide as non-reacting. The study found that the exponential decrease in ferric reduction with time was most likely due to limited availability of Fe^{3+} oxide rather than organic matters. This study also provided an evidence to support the hypothesis that in wetland sediments, ferric reduction rate followed a first order relationship with amorphous Fe^{3+} oxide concentration.

2.3.3 Iron reduction process in environmental remediation

The potential of IRB to be employed in environmental remediation can be measured by the degree to which the IRB can utilize ferric compounds in successful organic oxidization and removal of organic pollutants from environment. Several aliphatic and aromatic organic compounds have been reported to be successfully oxidized by ferric reduction (Azam and Finneran, 2013; Lovley and Lonergan, 1990; Lovley and Phillips, 1988). In this section, evidences of aliphatic and aromatic organic matter oxidation reported in the literature are summarized.

Fe³⁺ reduction in aliphatic compound oxidation

Azam and Finneran, (2013) successfully demonstrated that various ferric amendments increased the rates and extent of mineralization of carbon compounds to different degrees in a septic tank system. Lepidocrocite and Fe^{3+} EDTA were found as the most effectual

ferric compounds with high mineralization rate compared to Fe^{3+} citrate, ferrihydrate, Fe^{3+} NTA and Fe^{3+} pyrophosphate. Lepidocrocite achieved maximum mineralization rate for acetate (92%), lipid (98%) and lactate (82%) and Fe^{3+} EDTA attained the highest mineralization of glucose (74%), starch (93%) and butyrate (88%). The ferric amendment increased the generation of carbon dioxide and reduced the rate of greenhouse gas methane (CH_4) production. While most of the ferric compounds showed better results in carbon mineralization, there were exceptions such as ferric citrate which actually had a decreased mineralization rate of acetate. Fermentation of citrate to acetate was probably the reason of this anomaly (Azam and Finneran, 2013). Finneran and Lovley (2001) studied anaerobic degradation of methyl tert-butyl ether (MTBE) and tert-butyl alcohol (TBA) in aquatic sediments amended with poorly crystalline ferric oxide. The study found that approximately 30 moles of Fe^{3+} compound was required to completely oxidize MTBE, and addition of humic substances stimulated the degradation. In their study, rapid degradation of TBA was observed in a strictly anaerobic condition which differed the observation of slow degradation of TBA in previous studies.

Fe^{3+} reduction in aromatic compound oxidation

Groundwater contaminated by organic materials develops an anaerobic environment in the aquifer and aromatic hydrocarbons present in this anaerobic condition can be oxidized by reducers using nitrate, sulfate and iron as electron acceptor (Lovley et al., 1989; Major et al., 1988; Reinhard et al., 1984; Wilson et al., 1986). Geochemical evidences of accretion of Fe^{2+} precipitation in groundwater systems and depletion of Fe^{3+} from the aquifer sediments have suggested the applicability of ferric reduction as a remedial mechanism for groundwater systems contaminated by aromatic hydrocarbons (Major et al., 1988; Reinhard et al., 1984; Wilson et al., 1986). Previous studies have shown the effective decomposition of aromatic carbons by injecting ferric compounds (Lovley, 1995; Lovley and Lonergan, 1990; Lovley and Phillips, 1988).

Various monoaromatic compounds (benzene, xylene, toluene) have been found to be oxidized by microorganisms such as GS-15 using Fe^{3+} as the major electron acceptor. In one of the preliminary experiments by Lovley et al. (1989), toluene was successfully oxidized with poorly crystalline ferric oxide to carbon dioxide. More different aromatic

compounds (toluene, phenol and p-cresol) were later shown oxidized by an enriched culture of GS-15 in the ferric reduction process (Lovley and Lonergan, 1990). Thirty six (36) moles of ferric oxide was required to completely oxidize 1 mole of toluene to carbon dioxide where ferric oxide was the sole oxidant. Likewise, 28 and 34 moles of ferric compound were required for oxidation of phenol and p-cresol respectively. The oxidation rates were highly varied with the type of ferric compounds that were used in the experiment. In contrast to the toluene oxidation with ferric oxide, 108 moles of ferric hydroxide, $\text{Fe}(\text{OH})_3$ were required to oxidize 1 mole of toluene. However, these stoichiometric ratios do not represent the actual amounts as these metabolic reactions are much more complicated. Various forms of Fe^{2+} and Fe^{3+} and incorporation of some organic compounds on the cells of reducing microorganisms may be the reason for this complexity. Furthermore, these reactions are endothermic as good amount of energy is produced for the cell growth (Lovley and Lonergan, 1990).

2.4 Microbial processes of iron reduction and ecology

Iron reducing bacteria gain their energy by the oxidation of organic materials or other available substrates via extracellular iron reduction, a process known as 'extracellular respiration' (Esther et al., 2015). Different mechanisms of microbial iron reduction have been proposed which include direct and indirect contact of IRB with the iron minerals (Esther et al., 2015; Weber et al., 2006). Also, a wide range of microorganisms belong to different taxa have been identified as iron reducers which can adapt to different chemical environments.

2.4.1 Microbial processes of iron reduction

The microbial iron reduction is mainly transpired by extracellular electron transfer from iron reducers to the solid surface of the Fe^{3+} compounds, typically insoluble Fe^{3+} oxide minerals. Several strategies of electron transference had been investigated and reported in the literature (Esther et al., 2015; Weber et al., 2006), and the strategies are dependent on the type of microorganism and the surrounding environment. In microbial ferric reduction, interaction of iron reducers with ferric mineral surface is often interceded by the formation of biofilm that is composed of extra polymeric substance (EPS) matrix (Franks et al., 2010; Thormann et al., 2004). Electron transfer is then mediated by the

EPS as it creates support for electron transport proteins to attach with the microbes and improves the conductivity of the biofilm. Four approaches have been hypothesized by researchers on the interactions of microbial cells and Fe^{3+} surface which are illustrated in Fig. 3:

1. Direct contact between the bacteria surface and Fe^{3+} surface
2. Contact through pili, which is also known as 'protein nanowires'
3. Contact by using complex ligands which solubilize the Fe^{3+} compound
4. Electron shuttle which facilitates the electron transfer from and to solid Fe^{3+} surface

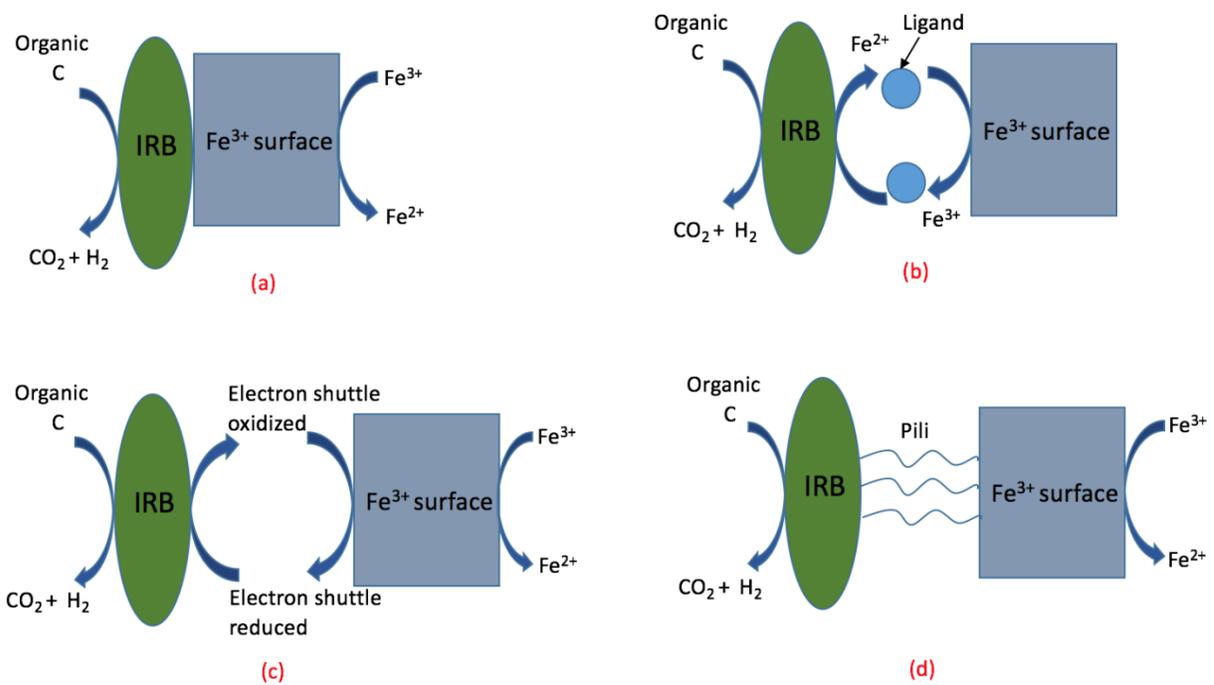


Fig. 3: Different strategies of microbial and ferric surface interaction (a) Direct Contact, (b) Contact with ligand, (c) Contact with electron shuttle, (d) Contact with nanowires; adapted from Esther et al. (2015).

Understanding the complex interactions between iron surface and microorganism is a convoluted task. Most of the research on microbial iron reduction has been progressed around two model iron reducers: *Geobacter* and *Shewanella* (Esther et al., 2015; Fredrickson and Gorby, 1996; Lovley, 1993; Nealson and Myers, 1990; Weber et al.,

2006). The interactions of these two reducers with the ferric surface are quite dissimilar from each other. *Shewanella* has been found to have direct and indirect electron transfer to insoluble Fe³⁺ surface, including protein nanowires. In contrast, *Geobacter* species are strictly anaerobe and do not contain enough electron shuttling or chelating compounds to solubilize Fe³⁺ and hence have been found to mostly rely on pilin filaments (Esther et al., 2015). Direct electron transfer by *Shewanella oneidensis* occurs by following the 'porin-cytochrome model' via a Mtr pathway. This Mtr pathway was formed by four multi-heme cytochromes (MtraA, MtrB, CymA and OmcA) and one non-heme protein MtrB (Beliaev et al., 2001; Myers and Myers, 1997, 2001; Pitts et al., 2003) that help in transferring electrons to the cell surface (Esther et al., 2015). The development of nanowire like pili was assumed as a requirement for the attachment of bacteria with ferric surface. But, later evidences showed, it works more likely as an electron conduit rather than an attachment medium (Weber et al., 2006). These conductive nanowires help to increase the spatial area outside the cell membrane and improve the cell to cell communications. Eventually, it improves the electron transfer to insoluble ferric oxide surface and other potential electron acceptors. Soluble redox-active compounds can serve as exogenous and endogenous electron shuttles mediating indirect electron transfer between iron reducers and the ferric mineral surfaces (Weber et al., 2006). Exogenous electron shuttles (e.g., humic substances and sulfur compounds) are compounds present in the surrounding environment and endogenous electron shuttles are mainly secreted from the microorganism itself (Esther et al., 2015). These compounds are reduced by the ferric reducers upon oxidization of an electron donor and that reduced electron shuttle subsequently diffuses and donates electrons to ferric oxide abiotically. These strategies are mainly investigated in sediments, submerged soils and aquifers, and are conceptually applicable in new applications of ferric reduction such as wastewater treatment.

2.4.2 Roles of iron reducing bacteria

Various bacterial strains have shown effectiveness to catalyze organics oxidation reactions coupled with ferric reduction. Dissimilatory ferric reducers have been shown to facilitate oxidation of glucose, amino acid, aromatics, long and short chain fatty acids (Lovley, 1993; Lovley and Phillips, 1989). Fermented sugar and amino acid could be

metabolized by Fe^{3+} reducers to produce two major fermentative products: acetate and hydrogen (H_2). Other fermentative products include propionate and formate which could subsequently be transformed into CO_2 with the help of *Geobacter* *Desulfuromonas* and *Shewanella desulfovibrio*, respectively (Lovley, 1993). Glucose can be oxidized directly to CO_2 with Fe^{3+} as the electron acceptor but the microorganisms that conduct this transformation are still unknown. However, these bacterial strains would be at detriments compared to the fermentative bacterial strain as glucose is always found to be fermented rather than directly oxidized to CO_2 in Fe^{3+} reducing sediments (Lovley and Phillips, 1989). Fig. 4 illustrates different pathways of organic matter oxidation coupled with ferric reduction and various microorganisms that have been found to catalyze the reactions. Although, most of the attempts to isolate these unknown species were unsuccessful, thermodynamic consideration has supported the assumption. Strains that help oxidize long chain fatty acids are still unknown whereas the aromatics are generally oxidized by *Geobacter* species with Fe^{3+} as the solitary electron acceptor (Lovley, 1993).

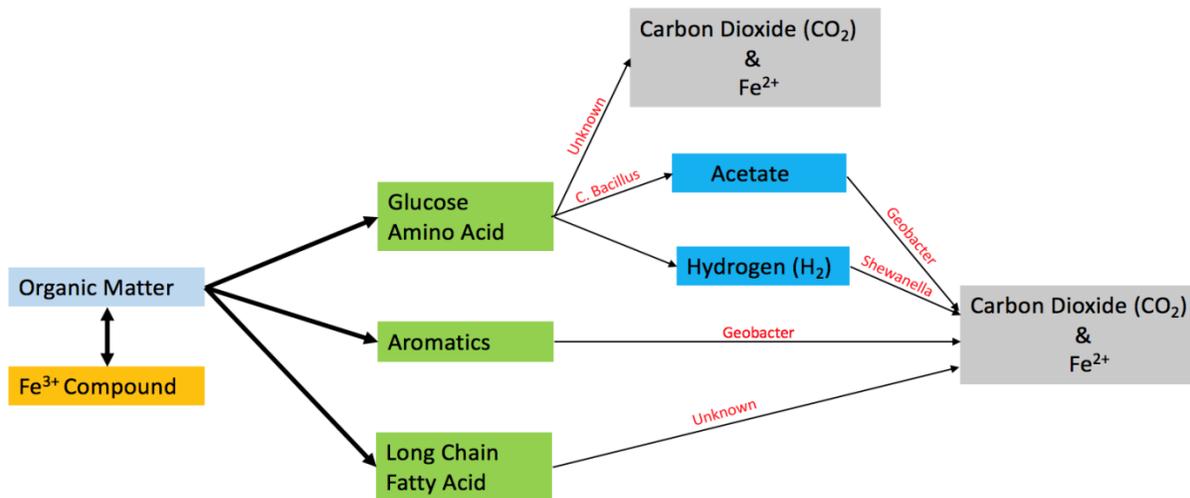


Fig. 4: Oxidation of organic matters with ferric reduction adapted from Lovley (1993)

Apart from IRB, the iron reduction competency of anaerobic ammonium oxidation (Anammox) bacteria in wastewater has been investigated in several studies (Park et al., 2009; Zhao et al., 2014). The iron reduction ability of anammox bacteria showed impressive results when organic matters were used as electron donors and can be an impending way of removing ammonium from wastewater treatment. The heterotrophic

IRB and autotrophic anammox bacteria compete against each other for utilizing ferric, but with the rising of nitrate production, the anammox bacteria outcompete the heterotroph (Park et al., 2009). However, the iron reduction activity of anammox bacteria was inhibited to around 93% when nitrite was coinciding with ferric compound. The performance of anammox bacteria in iron reduction varied with respect to the electron donors and acceptors (Zhao et al., 2014). Among formate, acetate and propionate, formate had been found as the most effective electron donor with the highest production of Fe^{2+} concentration ($\sim 179.64 \text{ mol L}^{-1}$). Between Fe^{3+} NTA and Fe^{3+} EDTA, first complex had better results as an electron donor at pH 7.

2.4.3 Diversity of iron reducing microorganisms

A wide variety of bacteria and archaea belonging to diverse taxa have been found to perform the ferric reduction in different physical and chemical conditions (Esther et al., 2015; Fredrickson and Gorby, 1996; Weber et al., 2006), suggesting prevalence of these ferric reducers in the environment. Most of the bacterial iron reducers belong to the Firmicutes, Proteobacteria, Deferibacteres and Actinobacteria taxa where the proteobacteria are categorized in different classes as α - Proteobacteria, β Proteobacteria, Δ Proteobacteria and γ Proteobacteria. They can be categorized as thermophilic, mesophilic, alkaliphilic, alkali-thermophilic, acido-thermophilic according to different adaptable conditions (Esther et al., 2015; Weber et al., 2006).

Model iron reducers *Geobacter Sp.* and *Shewanella Sp.* both belong to the group of Proteobacteria where *Geobacter Sp.* resides in delta sub division and *Shewanella sp.* in gamma sub division. *Shewanella Sp.* generally use lactate as carbon source and oxidize it to acetate, and *Geobacter Sp.* use acetate and completely oxidize it to CO_2 (Esther et al., 2015; Fredrickson and Gorby, 1996). *Geobacter metallireducens*, also known as GS-15 is one of the oldest and common ferric reducers which showed the capacity to reduce not only iron oxides but also Mn^{4+} and NO_3^- . GS-15 has been found to oxidize several carbon compounds to CO_2 by using Fe^{3+} (Nealson and Myers, 1990). Some of the iron reducers have been found to be closely linked with sulfur reducers belonging to the same taxa and using same electron acceptor for growth. For example, *Geobacter acetoxidans*, another member of delta Proteobacteria have shown similarity with *Desulfuromonas*

acetoxidans, a sulfur reducing bacteria (Fredrickson and Gorby, 1996) as both can use Fe^{3+} as electron acceptor for growth. *Pelobacter carbinolicus* has also shown ability to couple Fe^{3+} with oxidation of fermentation products and has an analogous phylogenetic relationship with *Geobacter* and *Desulfuromonas*. *Pelobacter Sp.* generally uses formate, ethanol and H_2 as electron donors with Fe^{3+} as the electron acceptor. These bacterial strains were mainly isolated from iron-rich sediments. Apart from these, *G. sulfurreducens* and *D. palmitatis* which were isolated from submerged soils of a ditch and marine sediments, respectively, have been discovered to use Fe^{3+} as sole electron acceptor. *Desulfuromusa kysingii* and *Desulfuromusa bakaii* have also found to utilize Fe^{3+} as electron acceptor but were isolated as sulfur reducers. These findings supported the hypothesis that there are some sulfate reducers which can reduce Fe^{3+} as electron acceptor. Nevertheless, sulfate reducers with only iron as sole electron acceptor has not been found in any literature (Lonergan et al., 1996).

Shewanella putrefaciens, also known as MR-1, is a facultative anaerobe, in contrast to the *Geobacter Sp.* which is a strict anaerobe. This bacterial strain was isolated from a culture that used non-fermentable carbon as electron donor (Nealson and Myers, 1990). But, it has the capacity to use a wide range of electron donors including iron, manganese etc. *Shewanella Sp.* is most effective in ferric reduction with lactate as the electron donor and the least effective with succinate. Although, these bacteria are proficient to preserve energy by ferric reduction, but possess extremely limited capacity to utilize organic matters as electron donors. Incomplete oxidation of lactate and pyruvate to acetate is commonly found redox reaction facilitated by these bacterial species (Lonergan et al., 1996). Similar to *S. putrefaciens*, BrY and *Desulfovibrio species* have also been noticed for their ability to oxidize lactate to acetate and CO_2 coupling with Fe^{3+} reduction (Lovley, 1993). *Shewanella Sp.* is also capable of reducing more crystalline form of ferric oxides compared to other iron reducers which reduce only amorphous or poorly crystalline ferric oxides. This revelation has helped to support the supposition that structure of ferric compound can also control the diversity of ferric reducing bacteria (Fredrickson and Gorby, 1996). A thermodynamically unfavorable magnetite reduction by *S. putrefaciens* has also been discovered, which has made them a bacterial strain of significant importance. In this case, formate or lactate could have been used by the *S. putrefaciens*

to reduce magnetite. These finding can ultimately help to understand the ecological diversity of iron reducers in the wastewater with different compositions. A list of major iron reducers that can reduce Fe^{3+} compounds is provided below with their major characteristics (Table 1):

Table 1: Major iron reducing bacteria and their primary electron donors

Sl No.	Bacterial strain	Adaptable condition	Electron donors	Reference
1	<i>Geobacter fertireducens</i>	Mesophilic	Acetate, volatile fatty acid, alcohol	Lovley (1993)
2	<i>Shewanella putrefaciens</i>	Mesophilic	Formate, lactate, pyruvate	Hyun et al. (1999)
3	<i>Aquaspirillum magnetotacticum</i>	Microaerophilic	Succinate	Guerin and Blakemore (1992)
4	<i>G.metallireducens</i>	Mesophilic	Mono aromatic compounds like toluene, p-cresol, and phenol	Lovley et al. (1989)
5	<i>G. chappelleii</i>	Mesophilic	Acetate, Hydrogen	Lonergan et al. (1996)
6	<i>G. hydrogenophilus</i>	Mesophilic	Acetate, Hydrogen	Lonergan et al. (1996)
7	<i>S. frigidimarina</i>	Psychrotropic	Sodium lactate, sodium acetate	Bowman et al. (2009)
8	<i>Geogemma pacifica sp.</i>	Hyper thermophilic	Formate, acetate,	Esther et al. (2015)
9	<i>Desulfovibrio sp.</i>	Mesophilic	Hydrogen	Lovley (1993)
10	<i>Shewanellasp. HN-41</i>	Mesophilic	Formate, lactate, pyruvate	Esther et al. (2015)
11	<i>Pelobacter carbinolicus</i>	Mesophilic	Fermentable substrate, formate, ethanol and H_2	Lonergan et al. (1996)

2.5 Iron reduction in wastewater treatment and future prospect

Given that typical untreated municipal wastewater has COD 339–1016 mg/L and sulfate 24-72 mg/L, free ammonia 14-41 mg/L, insignificant nitrate and nitrite levels, and

phosphate 3.7-11 mg/L (Table 3-18, (Metcalf & Eddy et al., 2014), using ferric Fe(III) in wastewater treatment has the following advantages: Low solubility of iron phosphate can be an effective mechanism for retaining phosphorus from wastewater and reducing P loads to receiving waters. This could be achieved by mixing wastewater and iron to allow formation and settling of the chemical precipitation prior to the anaerobic treatment (Berner, 1973; Ivanov et al., 2005). This is an important treatment aspect that nitrate and sulfate can not provide. Given the significant levels of sulfate in wastewater, sulfate reduction is expected to co-exist with iron reduction in the proposed iron-dosed anaerobic treatment. Precipitation of iron sulfide due to its low solubility (amorphous ferrous sulfide $K_{sp} \approx 10^{-3.05}$) can limit potential sulfide toxicity on iron reducers and sulfate reducers. With abundance and widespread presence of iron (in particular, various forms of Fe-containing wastes such as chemical sludge from acid mine drainage treatment and wastes from steel industry), tremendous opportunities exist for incorporating iron as a green agent in innovative wastewater treatment technologies.

In light of the literature findings that showed effectiveness of IRB for organic matter oxidation in sediments, groundwater, soil systems, applicability of ferric reduction for organic matter removal from wastewater is explored in this section. A few studies that evaluated the utilization of ferric reduction in wastewater treatment have been reported and future scope of this research is discussed here with the context of identified knowledge gaps. Although the composition of wastewater is distinctively different from the other environmental sources, the mechanisms of the reduction process in organic oxidation in the above-mentioned studies are beneficial for the applications of iron reduction process in wastewater treatment.

2.5.1 Organics mineralization by ferric reducers

Previous studies on co-treatment of AMD and municipal wastewater discussed in the introduction part of this paper have shown that substantial amount of COD, BOD, total organic carbon (TOC), sulfate and other nutrients can be reduced from the wastewater through the co-treatment process. These results have suggested the potential benefits of using iron in anaerobic wastewater treatment. The study of Azam and Finneran (2013) has explored the pertinence of ferric amendments in mineralization of various carbon

compounds in on-site anaerobic wastewater treatment. The mineralization of wide range of carbon molecules by different ferric compounds was one of the major illustrations of the study. As the composition of wastewater can vary significantly depending on the source of origin, there is a possibility that different ferric compounds will differ in performance of removing organic matter from wastewater of different composition. Other reduction processes such as fermentation and methanogens have limited substrate range and can not oxidize carbon molecules with high molecular mass, whereas ferric reduction process can be effective for mineralizing a broad array of carbon molecules (Chang et al., 2010; Lowe and Siegrist, 2008; McKinley and Siegrist, 2010). Further studies are required to examine biodegradability of different categories of organic matters typically found in wastewater.

2.5.2 Ferric iron bioavailability and bioreduction kinetics

Previous studies have indicated ferric bioavailability for IRB is a critical factor affecting ferric bioreduction kinetics and consequently carbon oxidation rate (Bonneville et al., 2004; Liu et al., 2001; Roden and Wetzel, 2002). For iron-dosed wastewater treatment, identification of the chemical morphology and structure of ferric compounds which support and disfavor the ferric bioreduction can be used as a baseline for selecting iron source materials. To promote bioavailability and reduce environmental footprint of iron-dosed wastewater treatment processes, iron-containing materials with large surface areas and fast ferric dissolution rates are required. In this regard, chemical sludge from alkaline treatment of AMD could be suitable materials for such applications due to their amorphous nature and large surface areas. Use of AMD sludge would also reduce the environmental burdens and cost of disposing the sludge materials. However, presence of toxic heavy metals in the AMD sludge need to be examined and addressed. Formation of metal sulfides (e.g., PbS and CuS) with biogenic sulfide in the bioreactor can potentially be a mechanism used to remove heavy metals and address the concern. Alternatively, selective precipitation in AMD treatment to obtain high-purity iron hydroxide and exclude heavy metals in the chemical sludge (Wei et al., 2005) can be used to prevent introducing AMD heavy metals to the wastewater.

Another critical factor for ferric bioavailability is pH as ferric compounds are highly

insoluble at circumneutral pH and most of the bacteria need an adaptable pH level of 5-8 (Straub et al., 2001). Soluble and insoluble ferric compounds can differ greatly in oxidizing the organic matters in wastewater. Hence, there is scope of future study to investigate optimal pH range for the biological treatment and variability in organics removal efficiency and kinetics in relation to pH.

Kinetic models that characterize ferric bioreduction and/or carbon oxidation are needed for design the iron-dosed treatment process. The kinetic models can then be used to estimate hydraulic and biomass retention times in the design of the treatment process. Both retention times are critical design parameters to achieve sufficient wastewater treatment.

2.5.3 Microbial ecology

In wastewater treatment applications, organic substrate availability is typically not a limiting factor as wastewaters have relatively higher levels of organic matters available compared to electron acceptors (e.g., nitrate and sulfate). In the iron-dosed anaerobic treatment, IRB and SRB are expected to be the dominant bacterial species in the bioreactors. Iron/sulfate ratio can be a suitable parameter for gauging microbial activities of iron and sulfate reducers in the bioreactors, and for developing kinetic models. Shift in microbial species distributions in relation to iron/sulfate ratio, and how the variations in microbial ecology affect organics removal represent important knowledge gaps that require studies for developing such treatment technology. In particular, there is a need to understand the nature of the relationships between IRB and SRB (i.e., symbiotic or competitive) to identify optimal operating conditions for the treatment.

In addition, studies aim to identify predominant microbial strains that truly responsible for ferric bioreduction and carbon oxidation are needed. The understanding of the microbial species and their ecology would help identify their mechanisms for utilizing ferric iron and carbon compounds. Although common strategies of IRB for utilizing ferric iron from iron material surface can be found in the literature, it is likely that new strategies and mechanisms can also be learned in wastewater treatment applications. Therefore, advancement in microbial studies is required to identify effective ferric reducers and their

ecology, and their relationship with wastewater composition to better employ iron reduction in wastewater treatment.

2.5.4 Sample treatment process

A possible treatment train of the proposed iron-dosed wastewater treatment may consist of primary, secondary, and a polishing treatment unit (Fig. 5). The primary unit is a clarifier where the iron is mixed with raw wastewater to form iron phosphate precipitation and settle chemical sludge. The secondary treatment is an anaerobic bioreactor that receives the effluent from the clarifier. And the polishing unit is to remove the remaining biological instability (e.g., ferrous Fe, sulfide, etc.) of the effluent from the biological system. Chlorine can be used as an oxidant that achieve both removal of biological instability and control of pathogens. The treated effluent can possibly be reused for a range of different purposes, which may or may not require additional treatment.

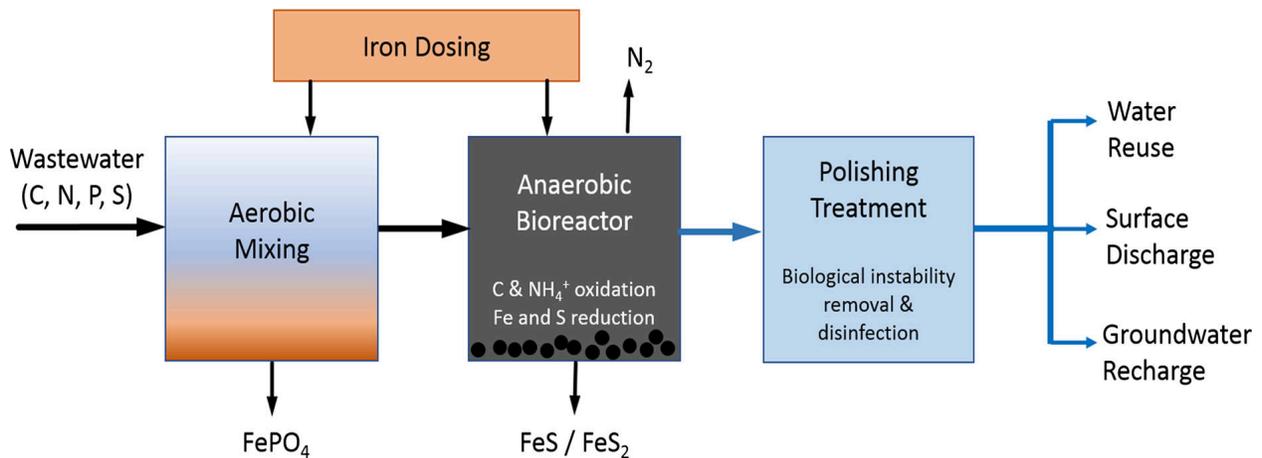


Fig. 5: Schematic diagram of the possible treatment process

2.6 Conclusion

Microbial reduction processes are drawing increasing attention because of their benefits over conventional aerobic processes. Ferric reduction process has shown astonishing results in organic degradation in natural systems and has a great potential to be used in novel wastewater treatment applications that provide multiple energy and environmental benefits. Majority of the studies on ferric reducers focused on their fundamental biochemical mechanisms in soil and groundwater systems. Better understanding on ferric

reduction process in engineering systems can broaden its applications such as wastewater treatment. Future research in the areas identified in this paper is required to further develop this treatment concept and capitalize the benefits that are made available through adopting the biochemical reactions of IRB in wastewater treatment.

References

- Achtnich, C., Bak, F., Conrad, R., 1995. Competition for electron donors among nitrate reducers, ferric iron reducers, sulfate reducers, and methanogens in anoxic paddy soil. *Biol. Fertil. Soils* 19, 65–72. <https://doi.org/10.1007/BF00336349>
- Arnold, R.G., DiChristina, T.J., Hoffmann, M.R., 1986. Inhibitor studies of dissimilative Fe(III) reduction by *Pseudomonas* sp. strain 200 ('*Pseudomonas ferrireductans*'). *Appl. Environ. Microbiol.* 52, 281–289.
- Arnold, R.G., DiChristina, T.J., Michael R. Hoffmann, 1988. Reductive dissolution of Fe(III) oxides by *Pseudomonas* sp. 200. *Biotechnol. Bioeng.* 32, 1081–1096.
- Azam, H.M., Finneran, K.T., 2013. Ferric iron amendment increases Fe(III)-reducing microbial diversity and carbon oxidation in on-site wastewater systems. *Chemosphere* 90, 1435–1443. <https://doi.org/10.1016/j.chemosphere.2012.09.002>
- Banwart, S., Davies, S., Stumm, W., 1989. The role of oxalate in accelerating the reductive dissolution of hematite (α -Fe₂O₃) by ascorbate. *Colloids and Surfaces* 39, 303–309. [https://doi.org/10.1016/0166-6622\(89\)80281-1](https://doi.org/10.1016/0166-6622(89)80281-1)
- Beliaev, A.S., Saffarini, D.A., 1998. *Shewanella putrefaciens* mtrB encodes an outer membrane protein required for Fe(III) and Mn(IV) reduction. *J. Bacteriol.* 180, 6292–6297.
- Beliaev, A.S., Saffarini, D.A., McLaughlin, J.L., Hunnicutt, D., 2001. MtrC, an outer membrane decahaem c cytochrome required for metal reduction in *Shewanella putrefaciens* MR-1. *Mol. Microbiol.* 39, 722–730. <https://doi.org/10.1046/j.1365-2958.2001.02257.x>
- Berner, R.A., 1973. Phosphate removal from sea water by adsorption on volcanogenic ferric oxides. *Earth Planet. Sci. Lett.* 18, 77–86. [https://doi.org/10.1016/0012-821X\(73\)90037-X](https://doi.org/10.1016/0012-821X(73)90037-X)
- Bonneville, S., Van Cappellen, P., Behrends, T., 2004. Microbial reduction of iron(III) oxyhydroxides: Effects of mineral solubility and availability. *Chem. Geol.* 212, 255–268. <https://doi.org/10.1016/j.chemgeo.2004.08.015>
- Bowman, J.P., McCammon, S.A., Nichols, D.S., Skerratt, J.H., Rea, S.M., Nichols, P.D., McMeekin, T.A., 2009. *Shewanella gelidimarina* sp. nov. and *Shewanella frigidimarina* sp. nov., Novel Antarctic Species with the Ability To Produce Eicosapentaenoic Acid (20:5 3) and Grow Anaerobically by Dissimilatory Fe(III) Reduction. *Int. J. Syst. Bacteriol.* 47, 1040–1047. <https://doi.org/10.1099/00207713-47-4-1040>
- Chan, Y.J., Chong, M.F., Law, C.L., Hassell, D.G., 2009. A review on anaerobic-aerobic treatment of industrial and municipal wastewater. *Chem. Eng. J.* 155, 1–18. <https://doi.org/10.1016/j.cej.2009.06.041>
- Chang, N.-B., Xuan, Z., Daranpob, A., Wanielista, M., 2010. A Subsurface Upflow Wetland System for Removal of Nutrients and Pathogens in On-Site Sewage

- Treatment and Disposal Systems. *Environ. Eng. Sci.* 28, 11–24. <https://doi.org/10.1089/ees.2010.0087>
- Chen, J., Gu, B., Royer, R.A., Burgos, W.D., 2003. The roles of natural organic matter in chemical and microbial reduction of ferric iron. *Sci. Total Environ.* 307, 167–178. [https://doi.org/10.1016/S0048-9697\(02\)00538-7](https://doi.org/10.1016/S0048-9697(02)00538-7)
- Clément, J.C., Shrestha, J., Ehrenfeld, J.G., Jaffé, P.R., 2005. Ammonium oxidation coupled to dissimilatory reduction of iron under anaerobic conditions in wetland soils. *Soil Biol. Biochem.* 37, 2323–2328. <https://doi.org/10.1016/j.soilbio.2005.03.027>
- Dar, S.A., Yao, L., Van Dongen, U., Kuenen, J.G., Muyzer, G., 2007. Analysis of diversity and activity of sulfate-reducing bacterial communities in sulfidogenic bioreactors using 16S rRNA and *dsrB* genes as molecular markers. *Appl. Environ. Microbiol.* 73, 594–604. <https://doi.org/10.1128/AEM.01875-06>
- De Castro, A.F., Ehrlich, H.L., 1970. Reduction of iron oxide minerals by a marine *Bacillus*. *Antonie Van Leeuwenhoek* 36, 317–327. <https://doi.org/10.1007/BF02069033>
- Deng, D., Lin, L.S., 2013. Two-stage combined treatment of acid mine drainage and municipal wastewater. *Water Sci. Technol.* 67, 1000–1007. <https://doi.org/10.2166/wst.2013.653>
- Deng, D., Weidhaas, J.L., Lin, L.-S., 2016. Kinetics and microbial ecology of batch sulfidogenic bioreactors for co-treatment of municipal wastewater and acid mine drainage. *J. Hazard. Mater.* 305, 200–8. <https://doi.org/10.1016/j.jhazmat.2015.11.041>
- Esther, J., Sukla, L.B., Pradhan, N., Panda, S., 2015. Fe (III) reduction strategies of dissimilatory iron reducing bacteria. *Korean J. Chem. Eng.* 32, 1–14. <https://doi.org/10.1007/s11814-014-0286-x>
- Franks, A.E., Malvankar, N., Nevin, K.P., 2010. Bacterial biofilms: the powerhouse of a microbial fuel cell. *Biofuels* 1, 589–604. <https://doi.org/10.4155/bfs.10.25>
- Fredrickson, J.K., Gorby, Y.A., 1996. Environmental processes mediated by iron-reducing bacteria. *Curr. Opin. Biotechnol.* 7, 287–294.
- Guerin, W.F., Blakemore, R.P., 1992. Redox cycling of iron supports growth and magnetite synthesis by *Aquaspirillum magnetotacticum*. *Appl. Environ. Microbiol.* 58, 1102–1109.
- Hughes, T.A., Gray, N.F., 2013. Co-treatment of acid mine drainage with municipal wastewater: Performance evaluation. *Environ. Sci. Pollut. Res.* 20, 7863–7877. <https://doi.org/10.1007/s11356-012-1303-4>
- Hyun, M., Kim, B., Chang, I., Park, H., Kim, H., Kim, G., Kim, M., DH, P., 1999. Isolation and identification of metalreducing bacterium, *Shewanella putrefaciens* IR-1. *J. Microbiol.* 37, 206–212.
- Ivanov, V., Stabnikov, V., Zhuang, W.Q., Tay, J.H., Tay, S.T.L., 2005. Phosphate removal from the returned liquor of municipal wastewater treatment plant using iron-reducing bacteria. *J. Appl. Microbiol.* 98, 1152–1161. <https://doi.org/10.1111/j.1365-2672.2005.02567.x>
- Jensen, M.M., Thamdrup, B., Rysgaard, S., Holmer, M., Fossing, H., 2003. Rates and Regulation of Microbial Iron Reduction in Sediments of the Baltic-North Sea Transition 65, 295–317.
- Jones, J.G., Gardener, S., Simon, B.M., 1983. Bacterial Reduction of Ferric Iron in a

- Stratified Eutrophic Lake. *J. Gen. Microbiol.* 129, 131–139. <https://doi.org/10.1099/00221287-129-1-131>
- Kamura, T., Takai, Y., Ishikawa, K., 1963. Microbial reduction mechanism of ferric iron in paddy soils (part I). *Soil Sci. Plant Nutr.* 9, 5–9. <https://doi.org/10.1080/00380768.1963.10431048>
- Liu, C., Kota, S., Zachara, J.M., Fredrickson, J.K., Brinkman, C.K., 2001. Kinetic analysis of the bacterial reduction of goethite. *Environ. Sci. Technol.* 35, 2482–2490. <https://doi.org/10.1021/es001956c>
- Lonergan, D., Jenter, H., Coates, J., Phillips, E., Schmidt, T., Lovley, D., 1996. Phylogenetic analysis of dissimilatory Fe (III)-reducing bacteria. *J. Bacteriol.* 178, 2402–2408. <https://doi.org/10.1128/JB.178.8.2402-2408.1996>
- Lovley, D.R., 1995. Bioremediation of organic and metal contaminants with dissimilatory metal reduction. *J. Ind. Microbiol.* 14, 85–93. <https://doi.org/10.1007/BF01569889>
- Lovley, D.R., 1993. Dissimilatory metal reduction. *Annu. Rev. Microbiol.* 47, 20–29.
- Lovley, D.R., 1987. Organic matter mineralization with the reduction of ferric iron: A review. *Geomicrobiol. J.* 5, 375–399. <https://doi.org/10.1080/01490458709385975>
- Lovley, D.R., Baedeker, M.J., Lonergan, D.J., Cozzarelli, I.M., Phillips, E.J.P., Siegel, D.I., 1989. Oxidation of aromatic contaminants coupled to microbial iron reduction. *Nature* 339, 297–300. <https://doi.org/10.1038/339297a0>
- Lovley, D.R., Lonergan, D.J., 1990. Anaerobic oxidation of toluene, phenol, and p-cresol by the dissimilatory iron-reducing organism, GS-15. *Appl. Environ. Microbiol.* 56, 1858–1864. <https://doi.org/10.1128/aem.72.5.3236-3244.2006>
- Lovley, D.R., Phillips, E.J., 1987. Competitive mechanisms for inhibition of sulfate reduction and methane production in the zone of ferric iron reduction in sediments. *Appl. Environ. Microbiol.* 53, 2636–41.
- Lovley, D.R., Phillips, E.J.P., 1989. Requirement for a microbial consortium to completely oxidize glucose in Fe(III)-reducing sediments. *Appl. Environ. Microbiol.* 55, 3234–3236.
- Lovley, D.R., Phillips, E.J.P., 1988. Novel Mode of Microbial Energy Metabolism: Organic Carbon Oxidation Coupled to Dissimilatory Reduction of Iron or Manganese. *Appl. Environ. Microbiol.* 54, 1472–1480. <https://doi.org/10.1103/PhysRevLett.50.1998>
- Lovley, D.R., Phillips, E.J.P., 1986. Organic Matter Mineralization with Reduction of Ferric Iron in Anaerobic Sediments. *Appl. Environ. Microbiol.* 51, 683–689. <https://doi.org/10.1080/01490458709385975>
- Lowe, K.S., Siegrist, R.L., 2008. Controlled Field Experiment for Performance Evaluation of Septic Tank Effluent Treatment during Soil Infiltration. *J. Environ. Eng.* 134, 93–101.
- Madigan, M., Martinko, J., Bender, K., Buckley, D., Stahl, D., 2015. Brock biology of microorganism, 14th ed. Pearson Education, Illionis.
- Major, D.W., Mayfield, C.I., Barker, J.F., 1988. Biotransformation of Benzene by Denitrification in Aquifer Sand. *Groundwater* 26, 8–14.
- Manariotis, I.D., Grigoropoulos, S.G., 2002. Low-Strength Wastewater Treatment Using an Anaerobic Baffled Reactor. *Water Environ. Res.* 74, 170–176.
- McCarty, P.L., Smith, D.P., 1986. Anaerobic wastewater treatment processes. *Environ. Sci. Technol.* 20, 1200–1206. <https://doi.org/10.1021/es00154a002>
- McKinley, J.W., Siegrist, R.L., 2010. Accumulation of Organic Matter Components in Soil

- under Conditions Imposed by Wastewater Infiltration. *Sci. Soc. Am. J.* 74, 1690–1700.
- Metcalf & Eddy, I., Tchobanoglous, G., Stensel, H.D., Tsuchihashi, R., Burton, F., 2014. *Wastewater Engineering: Treatment and Resource Recovery*. McGraw-Hill, New York.
- Munch, J.C., Ottow, J.C.G., 1983. Reductive Transformation Mechanism of Ferric Oxides in Hydromorphic Soils. *Environ. Biogeochem.* 35, 383–394.
- Myers, C.R., Myers, J.M., 1997. Cloning and sequence of *cymA*, a gene encoding a tetraheme cytochrome *c* required for reduction of iron(III), fumarate, and nitrate by *Shewanella putrefaciens* MR-1. *J. Bacteriol.* 179, 1143–1152. <https://doi.org/10.1128/jb.179.4.1143-1152.1997>
- Myers, J.M., Myers, C.R., 2001. Role for outer membrane cytochromes *OmC*A and *OmC*B of *Shewanella putrefaciens* MR-1 in reduction of manganese dioxide. *Appl. Environ. Microbiol.* 67, 260–269. <https://doi.org/10.1128/AEM.67.1.260-269.2001>
- Nealson, K.H., Myers, C.R., 1990. Iron reduction by bacteria: A potential role in the genesis of banded iron formations. *Am. J. Sci.* 290, 35–45.
- Oude Elferink, S.J.W.H., Visser, A., Pol, L.W.H., Stams, A.J., 1994. Sulfate reduction in methanogenic bioreactors. *FEMS Microbiol. Rev.* 15, 119–136.
- Park, W., Nam, Y.K., Lee, M.J., Kim, T.H., 2009. Anaerobic ammonia-oxidation coupled with Fe³⁺ reduction by an anaerobic culture from a piggery wastewater acclimated to NH₄⁺/Fe³⁺ medium. *Biotechnol. Bioprocess Eng.* 14, 680–685. <https://doi.org/10.1007/s12257-009-0026-y>
- Pierre, J.L., Fontecave, M., Crichton, R.R., 2002. Chemistry for an essential biological process: The reduction of ferric iron. *BioMetals* 15, 341–346. <https://doi.org/10.1023/A:1020259021641>
- Pitts, K.E., Dobbin, P.S., Reyes-Ramirez, F., Thomson, A.J., Richardson, D.J., Seward, H.E., 2003. Characterization of the *Shewanella oneidensis* MR-1 Decaheme Cytochrome *MtrA*. *J. Biol. Chem.* 278, 27758–27765. <https://doi.org/10.1074/jbc.m302582200>
- Postma, D., 1993. The reactivity of iron oxides in sediments: A kinetic approach. *Geochim. Cosmochim. Acta* 57, 5027–5034. [https://doi.org/10.1016/S0016-7037\(05\)80015-8](https://doi.org/10.1016/S0016-7037(05)80015-8)
- Reinhard, M., Barker, J.F., Goodman, N.L., 1984. Occurrence and distribution of organic chemicals in two landfill leachate plumes. *Environ. Sci. Technol.* 18, 953–961. <https://doi.org/10.1021/es00130a011>
- Roden, E.E., Wetzell, R.G., 2002. Kinetics of microbial Fe(III) oxide reduction in freshwater wetland sediments. *Limnol. Oceanogr.* 47, 198–211. <https://doi.org/10.4319/lo.2002.47.1.0198>
- Roetman, E., 1932. The sterilization of sewage by acid mine water. West Virginia University.
- Saravanane, R., Murthy, D.V.S., 2016. Application of anaerobic fluidized bed reactors in wastewater treatment: a review. *Environ. Manag. Heal.* 11, 97–117.
- Sawayama, S., 2006. Possibility of anoxic ferric ammonium oxidation. *J. Biosci. Bioeng.* 101, 70–72. <https://doi.org/10.1263/jbb.101.70>
- Speece, R.E., 1983. Anaerobic biotechnology for industrial wastewater treatment. *Environ. Sci. Technol.* 17, 416A–427A. <https://doi.org/10.1021/es00115a001>

- Straub, K.L., Benz, M., Schink, B., 2001. Iron metabolism in anoxic environments at near neutral pH. *FEMS Microbiol. Ecol.* 34, 181–186. [https://doi.org/10.1016/S0168-6496\(00\)00088-X](https://doi.org/10.1016/S0168-6496(00)00088-X)
- Strosnider, W.H., Winfrey, B.K., Nairn, R.W., 2011. Biochemical oxygen demand and nutrient processing in a novel multi-stage raw municipal wastewater and acid mine drainage passive co-treatment system. *Water Res.* 45, 1079–1086. <https://doi.org/10.1016/j.watres.2010.10.026>
- Thamdrup, B., 2000. Bacterial Manganese and Iron Reduction in Aquatic Sediments. *Adv. Microb. Ecol.* 16, 41–84. https://doi.org/10.1007/978-1-4615-4187-5_2
- Thomsen, U., Thamdrup, B., Stahl, D.A., Canfield, D.E., 2004. Pathways of organic carbon oxidation in a deep lacustrine sediment, Lake Michigan. *Limnol. Oceanogr.* 49, 2046–2057. <https://doi.org/10.4319/lo.2004.49.6.2046>
- Thormann, K.M., Shukla, S., Pelletier, D.A., Spormann, A.M., 2004. Initial Phases of Biofilm Formation in *Shewanella oneidensis* MR-1. *J. Bacteriol.* 186, 8096–8104. <https://doi.org/10.1128/JB.186.23.8096>
- Urrutia, M.M., Roden, E.E., Fredrickson, J.K., Zachara, J.M., 1998. Microbial and surface chemistry controls on reduction of synthetic Fe(III) oxide minerals by the dissimilatory iron-reducing bacterium *Shewanella* alga. *Geomicrobiol. J.* 15, 269–291.
- Utgikar, V.P., Harmon, S.M., Chaudhary, N., Tabak, H.H., Govind, R., Haines, J.R., 2002. Inhibition of sulfate-reducing bacteria by metal sulfide formation in bioremediation of acid mine drainage. *Environ. Toxicol.* 17, 40–48. <https://doi.org/10.1002/tox.10031>
- Van der Maas, P., 2005. Fe(III) EDTA reduction in BioDeNOx reactors: kinetics and mechanism, in: *Chemically Enhanced Biological NOx Removal from Flue Gases*. Wageningen University, pp. 109–124.
- Van Lier, J.B., 2008. High-rate anaerobic wastewater treatment: Diversifying from end-of-the-pipe treatment to resource-oriented conversion techniques. *Water Sci. Technol.* 57, 1137–1148. <https://doi.org/10.2166/wst.2008.040>
- Van Lier, J.B., Van der Zee, F.P., Frijters, C.T.M.J., Ersahin, M.E., 2015. Celebrating 40 years anaerobic sludge bed reactors for industrial wastewater treatment. *Rev. Environ. Sci. Biotechnol.* 14, 681–702. <https://doi.org/10.1007/s11157-015-9375-5>
- Weber, K.A., Achenbach, L.A., Coates, J.D., 2006. Microorganisms pumping iron: Anaerobic microbial iron oxidation and reduction. *Nat. Rev. Microbiol.* 4, 752–764. <https://doi.org/10.1038/nrmicro1490>
- Wei, X., Viadero, R.C., Buzby, K.M., 2005. Recovery of Iron and Aluminum from Acid Mine Drainage by Selective Precipitation. *Environ. Eng. Sci.* 22, 745–755. <https://doi.org/10.1089/ees.2005.22.745>
- Widdel, F., 1988. Microbiology and ecology of sulfate and sulfur-reducing bacteria, in: Zehnder, A. (Ed.), *Biology of Anaerobic Microorganisms*. Wiley, New York, pp. 459–586.
- Wilson, B.H., Smith, G.B., Rees, J.F., 1986. Biotransformations of Selected Alkylbenzenes and Halogenated Aliphatic Hydrocarbons in Methanogenic Aquifer Material: A Microcosm Study. *Environ. Sci. Technol.* 20, 997–1002. <https://doi.org/10.1021/es00152a005>
- Yang, W.H., Weber, K.A., Silver, W.L., 2012. Nitrogen loss from soil through anaerobic ammonium oxidation coupled to iron reduction. *Nat. Geosci.* 5, 538–541. <https://doi.org/10.1038/ngeo1530>

- Zhang, L., Keller, J., Yuan, Z., 2009. Inhibition of sulfate-reducing and methanogenic activities of anaerobic sewer biofilms by ferric iron dosing. *Water Res.* 43, 4123–4132. <https://doi.org/10.1016/j.watres.2009.06.013>
- Zhao, R., Zhang, H., Li, Y., Jiang, T., Yang, F., 2014. Research of Iron Reduction and the Iron Reductase Localization of Anammox Bacteria. *Curr. Microbiol.* 69, 880–887. <https://doi.org/10.1007/s00284-014-0668-7>
- Zinder, B., Furrer, G., Stumm, W., 1986. The coordination chemistry of weathering: II. Dissolution of Fe(III) oxides. *Geochim. Cosmochim. Acta* 50, 1861–1869.

Chapter 3: Effects of Fe/S Ratio on the Kinetics and Microbial Ecology of an Fe(III)-dosed Anaerobic Wastewater Treatment System

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3.1 Introduction

Anaerobic biological treatment of wastewater using alternative electron acceptors to oxygen can offer benefits of cost effectiveness and energy efficiency (Van Lier, 2008; Van Lier et al., 2015). It also provides opportunities of utilizing redox active elements such as iron and sulfate from waste materials in wastewater treatment. Previous studies have shown that iron reducing microorganisms are effective in organic matter mineralization in natural systems such as groundwater, soils, and sediments (Bonneville et al., 2004; Jensen et al., 2003; Lovley, 1995, 1987; Roden and Wetzel, 2002; Thamdrup, 2000). Therefore, engineering processes that couple reduction of ferric iron to organic carbon oxidation can potentially be an effective wastewater treatment method that provides multiple energy and environmental benefits (Ahmed and Lin, 2017).

Previous studies showed that ferric reduction kinetics was influenced by the type of electron donors. Van der Maas (2005) showed variations in ferric reduction rate by changing the electron donors in mixed liquor bioreactors, which ranged from 4.1 mM.h⁻¹ with methanol to 13.9 mM.h⁻¹ with glucose. Microbial iron reduction rates are also affected by factors such as the crystallinity, mineralogy and solubility of ferric compounds, contact between ferric surface and iron reducing bacteria (IRB), and electron transfer mechanisms (Ahmed and Lin, 2017; Liu et al., 2008; Nevin and Lovley, 2002). In general, more poorly crystalline ferric compounds result in higher reduction rates than crystalline compounds (Bonneville et al., 2009), and less stable minerals such as ferrihydrite and lepidocrocite showed higher reactivity than stable minerals such as goethite and hematite (Postma, 1993). Bonneville et al. (2006) studied the variations in maximum specific reduction rates with different forms of ferric hydroxide, and reported a range of values from 0.27 X 10⁻¹¹ μmolh⁻¹ cell⁻¹ with LSA hematite to 399 X 10⁻¹¹ μmolh⁻¹ cell⁻¹ with soluble ferric citrate. The solubility of these different ferric compounds was considered as a factor

causing the differences in the reduction rate as it affects the bioavailability of these compounds to IRB. While phylogenetically diverse microorganisms may be capable of dissimilatory iron reduction, majority of previous studies have focused on characterizing the metabolism and physiology of *Shewanella* and *Geobacter* species that can be easily cultivated in the laboratory (Aklujkar et al., 2010; Liu et al., 2002; Lovley et al., 1993; Mahadevan et al., 2011; Myers and Myers, 2001; Urrutia et al., 1998).

In anaerobic processes using ferric iron reduction for treating sulfate-containing wastewater, IRB and sulfate reducing bacteria (SRB) may co-exist and both contribute to organic carbon oxidation. Zhang et al. (2013) have observed a symbiotic relationship between SRB and IRB in an anaerobic wastewater treatment process, in which addition of ferric oxide into a sulfidogenic bioreactor elicited microbial iron reduction and improved both sulfate reduction and organic carbon oxidation. However, IRB can outcompete SRB under limited carbon substrate conditions by diverting electron flow away from SRB (Lovley and Phillips, 1989). Such symbiotic and/or competitive dynamics between IRB and SRB is regulated by the availability of carbon substrate and electron acceptors (ferric iron and sulfate), and largely unknown for engineering applications.

Production of ferrous iron and (bi)sulfide from the reductive microbial reactions along with the resulting iron sulfide sludge in the iron-dosed anaerobic bioreactors is another aspect concerning the treatment method. Excessive production of aqueous and gaseous sulfides at levels toxic to microorganisms may limit the extent of carbon oxidation (Cirne et al., 2008; Mosey et al., 1971). Aqueous sulfides can form sparingly soluble metal sulfide precipitates and reduce contact area between reactants and microbial enzymes (Utgikar et al., 2002). In general, biogenic iron sulfide precipitating as FeS (mackinawite) is poorly crystalline (Herbert et al., 1998). This iron mono sulfide is also known as a precursor to well crystallized pyrite (FeS_2) as it provides an initial nucleation site for pyrite growth (Gramp et al., 2010; Herbert et al., 1998). Greigite (Fe_3S_4), smythite (Fe_9S_{11}), and pyrrhotite (Fe_{1-x}S) are some of the intermediate iron sulfide forms that may exist between the transformation of amorphous FeS to crystalline FeS_2 . Gramp et al. (2010) reported that mackinawite (FeS) was the dominant phase in the biologically produced precipitates with lower Fe/S molar ratio (1:3.25, 1:20, 1:50). The crystallinity of mackinawite was

enhanced when the iron concentration increased (Fe/S ratio 0.5), which eventually increased the proportion of pyrite formation in the precipitates. Understanding the composition of iron sulfides produced in these systems may be critical given their potential beneficial applications due to magnetic and electrical properties.

This study addressed the knowledge gaps in the effects of iron-dosing on organic carbon oxidation kinetics, competitive/synergistic relationship of IRB and SRB, correlations of microbial ecology with the carbon oxidation, and sludge characteristics. Specifically, anaerobic batch bioreactors were used to treat sulfate-containing wastewater under three different Fe/S molar ratios while maintaining the same total equivalent of electron acceptors (i.e., ferric iron and sulfate). Microbiological analyses were performed to characterize the microbial communities, and to investigate their relationships with the organic carbon oxidation kinetics. Sludge samples from the biological treatment were collected and characterized for their morphology, chemical composition, and elemental oxidation states. A conceptual framework was proposed to illustrate the major process factors that influence the organic carbon oxidation and to assist further development of this treatment method.

3.2 Materials and Methods

3.2.1 Batch Bioreactors

Six batch bioreactors (1.15 L glass bottles) were used to investigate the effects of Fe/S ratio on the kinetics of organic carbon oxidation ([Supplementary material, Fig. S1](#)). The bioreactors were packed with plastic media (Kaldnes K1, specific surface area = 500 cm²/m³, Evolution Aqua Ltd, UK) for attached growth of microorganisms. Each reactor was packed with 550 plastic media with a working volume of 0.7 L. A mixture of anaerobic sludge from a wastewater treatment plant (Star City, West Virginia) and acid mine drainage (AMD) from St. Thomas, Morgantown, West Virginia at 1:1 volume ratio was used to inoculate the bioreactors. Enrichment of biomass lasted for approximately 3 months during which fresh synthetic wastewaters containing organic carbon, ferric iron and sulfate were prepared to replace the solutions in the bioreactors every week. The synthetic wastewater contained sodium acetate anhydrous (C₂H₃NaO₂), ethanol (C₂H₆O),

lactose monohydrate ($C_{12}H_{22}O_{11} \cdot H_2O$), sodium bicarbonate ($NaHCO_3$) and trace elements (Deng and Lin, 2017). During this enrichment period, the concentration of chemical oxygen demand (COD) in the synthetic wastewaters was kept constant (400 mg/L) and iron and sulfate concentrations were varied to obtain three Fe/S molar ratios (1, 2 and 3) using ferric chloride ($FeCl_3 \cdot 6H_2O$) and sodium sulfate (Na_2SO_4) solutions. For all three Fe/S ratios, the ferric iron and sulfate concentrations were predetermined to obtain the same total equivalent concentration of the electron acceptors (4.68 mN). Specifically, ferric iron concentrations were 29, 53 and 71 mg/L and sulfate concentrations were 50, 45 and 41 mg/L for Fe/S molar ratio 1, 2 and 3 respectively. Each bioreactor was sealed with a rubber stopper and bubbled with N_2 gas regularly to ensure an anaerobic condition in the bioreactors. Each bioreactor was equipped with two sampling ports through the rubber stopper to allow periodic sample collection and pH/redox potential monitoring. The sampling ports were sealed except during sample collection.

3.2.2 Kinetic Study

After the biomass enrichment, the bioreactors were used to examine the effects of Fe/S ratio on organic carbon oxidation using the synthetic wastewater described above. The concentrations of organic carbons were varied to obtain different COD levels (50, 100, 200, 400, 500, 800, and 1000 mg/L). The pH of the mixed solutions was maintained at a moderate level (6-6.2) using a sodium hydroxide solution (5N) to ensure a favorable environment for biological growth. The samples were collected after every three hours over a nine-hour period to monitor the COD, sulfate, sulfide, ferric and ferrous iron concentrations. Each sample was filtered with a 0.2 μm membrane filter and stored at 4°C until analysis. Changes in COD concentrations during first three hours was used to estimate the COD oxidation rate (V , mg/L*min) using the initial rate method (Hegyi et al., 2013). The concentrations of dissolved sulfide and ferrous iron were monitored to help examine the degree of iron and sulfate reduction in the bioreactors and their stoichiometric relationships.

3.2.3 Analytical Methods

All the chemical analyses were conducted by following the Standard Methods (APHA et al., 2005). COD concentrations were measured by following a closed reflux, colorimetric

method (Standard Method 5220 D) and using a spectrophotometer (HACH, DR 2800). The ferrous and sulfide concentrations were determined by the 1, 10 phenanthroline method (Standard Method 3500 B) and methylene blue method (Standard Method 4500 D) respectively using the spectrophotometer. Soluble sulfate was measured with a GENESYS 10UV (Thermo Scientific) spectrophotometer following the turbidimetric method (USEPA method 375.4). Total iron concentration was measured using an atomic absorption spectrophotometer (Perkin Elmer 3100) after the samples were acidified with a 70% nitric acid solution. Total sulfur in the sludge samples was analyzed using an optical emission spectrometer (Optima 2100 DV). Sample pH and redox potential were measured using a pH meter (AB15 Plus, Fisher Scientific) and a VWR traceable ORP meter.

3.2.4 Kinetics Modeling

The calculated COD oxidation rates (V , mg/L*min) at different substrate (S , mg/L) levels were used to characterize the COD oxidation kinetics of the biological treatment using the Michaelis-Menten model:

$$V = \frac{V_{\max} \times S}{K_m + S} \dots\dots\dots (1)$$

where V_{\max} (mg/L*min) is the maximum reaction rate and K_m (mg/L) is the Michaelis-Menten constant. The data were used to fit the model using both linear and nonlinear methods and estimate the model parameters (K_m and V_{\max}). Specifically, Lineweaver-Burk transformation of the model, a double reciprocal plot, was used as the linear model (Deng et al., 2016; Kaksonen et al., 2006) and Microsoft Excel’s “Solver” program was used for the nonlinear fitting.

3.2.5 Sludge Characterization

Once the kinetics experiment at different COD concentrations were concluded, the accumulated sludge samples were collected from the bioreactors for physical, chemical, and biological characterization. Total suspended solid (TSS), volatile suspended solid (VSS) and non-volatile suspended solid (NVSS) in the sludge materials were measured

(Standard Method 2540). Sludge samples were also acidified using a 70% HNO₃ solution to quantify total Fe and S. A scanning electron microscope (SEM) equipped with an energy dispersion spectroscopy (SEM-EDS, Hitachi S 4700) was used to study the morphology and elemental composition of sludge materials. Specifically, the sludge samples were dried in a desiccator filled with calcium sulfate and flushed with nitrogen gas to prevent sludge oxidation. The sludge samples were prepared on aluminum mounts and coated with Au-Pd. For SEM analysis, both filtered and powdered sludge samples were used and analyzed under an accelerating voltage of 10 kV. Selected areas of the sludge samples were analyzed by the EDS for elemental information under the same accelerating voltage with online ZAF correction. Powdered samples of the sludge were also used for X-ray photoelectron spectroscopy (XPS) analysis (PHI 5000 Versaprobe) to identify the chemical states of Fe and S. The sludge samples were prepared following the same method as that for the SEM analysis. Elemental scans of Fe and S were conducted using a pass energy of 23.5 eV.

3.2.6 Microbiological Analyses

DNA was extracted from biofilm samples using a DNeasy Powersoil DNA extraction kit (Qiagen, Maryland, USA) and quantified using a Qubit fluorometer (Invitrogen, Carlsbad, CA, USA). 16S rRNA genes in the extracted DNA were sequenced at Argonne National Laboratory using the bacterial/archaeal primer set 515F/806R that targets the V4 region of the gene. Resulting reads were checked for chimeras (DADA2) and subsequently clustered into exact sequence variant (ESV) classifications at 100% similarities using the DADA2 tool in the QIIME2 pipeline (Qiime2-2018.4) and SILVA 16S rRNA gene database. Sequences used in this study have been deposited in the NCBI Sequence Read Archive under accession number PRJNA504045.

3.3 Results and Discussion

3.3.1 Oxidation Kinetics under different Fe/S ratios

The nonlinear least square method (i.e., Microsoft Solver) consistently produced better fits of the data (Fig. 6) than the linear method ([Supplementary Materials, Fig. S2](#)) in this study. The Lineweaver-Burk plot can suffer from a highly-biased weighting of points and

is discouraged to be used in many cases of enzyme kinetics (Martin, 1997; Perrin, 2017). The V_{max} values estimated by the nonlinear model were 0.47, 0.73 and 1.09 mg/L*min for Fe/S ratio 1, 2 and 3, respectively.

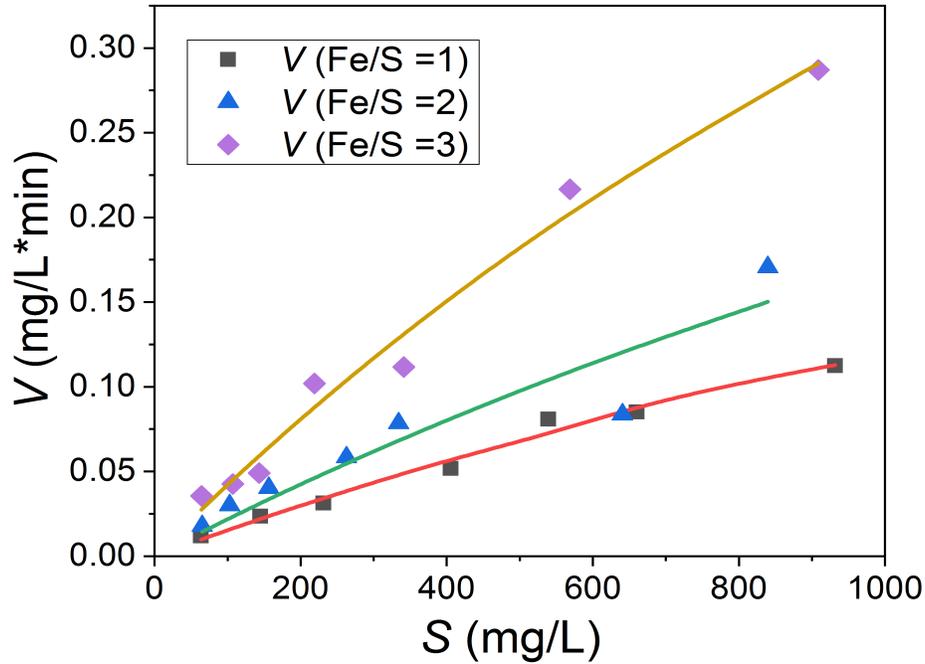


Fig. 6: Nonlinear kinetic models of COD oxidation under three different Fe/S molar ratios

At any substrate level in the range examined, increased Fe/S ratios resulted in higher carbon oxidation rates. For example, at substrate level 400 mg/L, the oxidation rate obtained from nonlinear model was 0.05, 0.08 and 0.15 mg/L*min for ratio 1, 2 and 3 respectively. This indicates that with same total equivalent concentration of the electron acceptors, a higher Fe^{3+} concentration along with its corresponding lower sulfate concentration improved the overall COD oxidation rate. From the wastewater treatment perspective, the improved COD oxidation kinetics driven by the increasing percentage of ferric iron would allow shorter hydraulic retention time. The COD oxidation rates were relatively less variable at low substrate levels and showed significantly larger differences at high substrate levels among the three Fe/S ratios. K_m values ranged from 2,503 mg/L to 3,267 mg/L and they reflected the combined enzymatic activities of both iron reducers and sulfate reducers in the bioreactors. These K_m values were substantially higher than the COD levels typically found in municipal wastewaters, indicating that this treatment

method can be operated at higher COD oxidation rates under higher COD levels. These results indicate that Fe/S ratio can play a significant role in regulating the kinetics of the COD oxidation processes with higher COD oxidation rates under increased Fe/S ratios.

3.3.2 Factors affecting COD oxidation

3.3.2.1 pH & redox potential

The pH value in the bioreactors ranged in 6.5-7, which was conducive to microbial activities of both iron and sulfate reducers. Redox potential of these bioreactors was low and varied from -123.4 mV to -241.5 mV. These conditions were favorable for iron and sulfide to precipitate as ferrous sulfide (Gerardi, 2006).

3.3.2.2 Dissolved ferrous and sulfide

For each Fe/S ratio, all samples collected in the first nine hours under all tested COD concentrations ($n = 50$) were filtered ($0.2 \mu\text{m}$) and analyzed for both dissolved ferrous iron and sulfide. Dissolved sulfide was consistently low under all Fe/S ratios with median concentrations ranging in $0.02 - 0.034 \text{ mg/L}$ (Fig. 7). The median concentration of ferrous iron increased from 0.02 mg/L to 5.6 mg/L , indicating increasingly excessive quantities of ferrous iron over sulfide as Fe/S ratio increased (Fig. 7).

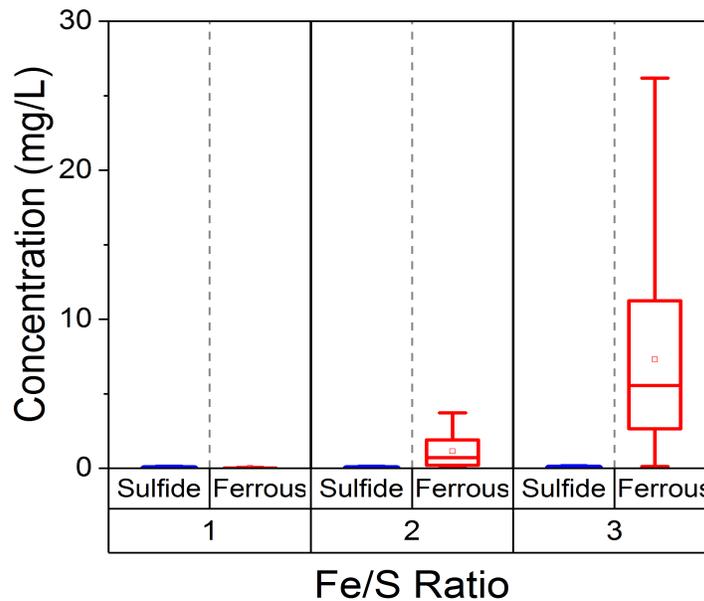


Fig. 7: Dissolved sulfide and ferrous iron concentrations in the bioreactors under different Fe/S ratios

The mean concentration products of ferrous iron and sulfide under different Fe/S ratios ranged from $1.3 \times 10^{-10} \text{ M}^2$ to $1.6 \times 10^{-13} \text{ M}^2$, which were very low reflecting high degrees of iron sulfide precipitation in the bioreactors. Increasingly excessive ferrous production over sulfide was observed with increased Fe/S ratio (Fig. 7). For practical applications, the presence of ferrous iron and sulfide in the treated water can cause biological instability in its receiving water because of their oxygen demands. However, they can be removed through oxidation which can be conveniently achieved by chlorination in the disinfection treatment of the effluent (Cadena and Peters, 1988; Khadse et al., 2015). Other physical and chemical methods such as precipitation and absorption can also be used to remove these chemicals (Horikawa et al., 2004; Kapdi et al., 2005; Petersson and Wellinger, 2009).

3.3.3 Sludge Characterization

3.3.3.1 Sludge Solid Contents

The solid concentrations of the cumulated sludge collected after the kinetic tests varied significantly among the three different Fe/S ratios. TSS concentrations of the samples (n = 30) increased with Fe/S ratio (Fig. 8a), and were $339 \pm 28 \text{ mg/L}$, $505 \pm 97 \text{ mg/L}$ and $652 \pm 143 \text{ mg/L}$ for Fe/S ratio 1, 2 and 3, respectively. The increases were mostly caused by NVSS and relatively smaller variations caused by VSS. Mean VSS/TSS ratio varied from 0.32 to 0.45 and correspondingly NVSS/TSS ratio varied from 0.68 to 0.55 (Fig. 8b). These results showed the range of the inorganic and organic fractions of the sludge materials, and that their differences increased with Fe/S ratio (Fig. 8a). They also showed that higher Fe/S ratios would increase overall sludge production rate from the treatment process.

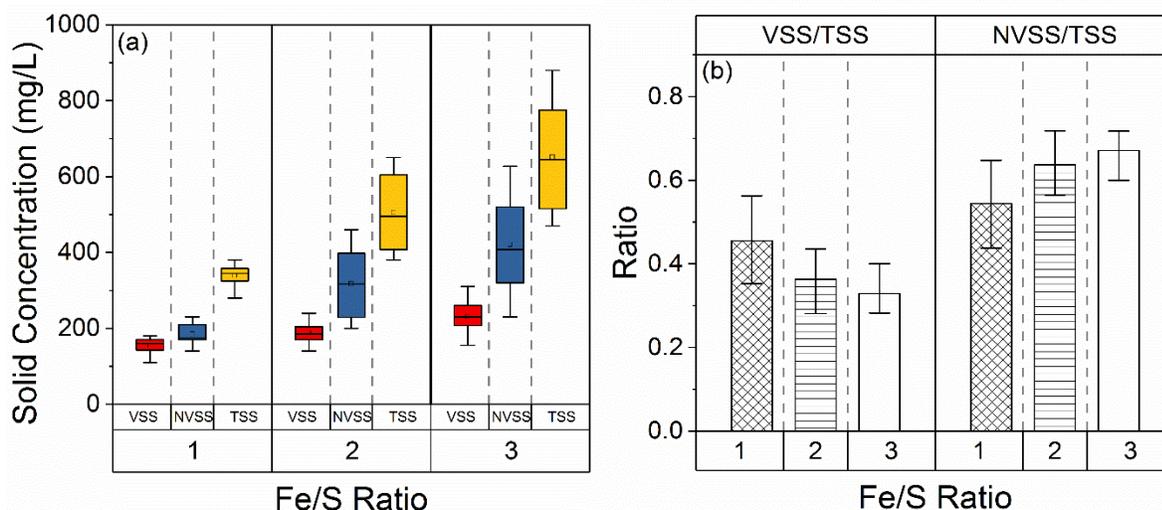


Fig. 8: (a) Solid concentrations and (b) VSS/TSS and NVSS/TSS ratios of the sludge materials

3.3.3.2 Iron and Sulfur

In the feed solutions, sulfur (S) was in a soluble form, but ferric iron was in suspension as $\text{Fe}(\text{OH})_3$ flocs given the solution pH (6.5-7). Upon ferrous sulfide precipitation, Fe and S were mostly in particulate forms and only a small portion was in soluble forms given the low solubility of ferrous sulfide. In the sludge materials, approximately 95.5% of the total iron was in a solid form leaving 4.5% in a soluble form mostly as Fe^{2+} (Supplementary Material, Fig. S3). Similarly, particulate S accounted for a much greater fraction of the sludge (90 – 97%) and dissolved S made up only a small fraction (3-10%). This chemically reduced inorganic material is a by-product of the treatment system and can possibly be extracted from the sludge for beneficial uses such as production of magnetite particles (Thorpe et al., 1984; Waters et al., 2008).

3.3.3.3 Sludge morphology and chemical composition

SEM-EDS analysis gave a synopsis on the morphology and chemical composition of the sludge. SEM images showed small grains of precipitate particles ($< 1 \mu\text{m}$) that were poorly ordered in structure (Fig. 9). The EDS analysis showed that major sludge elements included carbon (C), oxygen (O), Fe and S (Fig. 9). Chemical elements with a high atomic number generally reflect brighter contrast in SEM images due to their larger backscattered electron yield (Li et al., 2015). Therefore, the bright elements were

attributed to precipitates of Fe and S as they have higher atomic number than C and O. The structural arrangement of inferred iron sulfide was comparable to those reported previously (Bratkova et al., 2018; Csákberényi-Malasics et al., 2012; Vaclavkova et al., 2014).

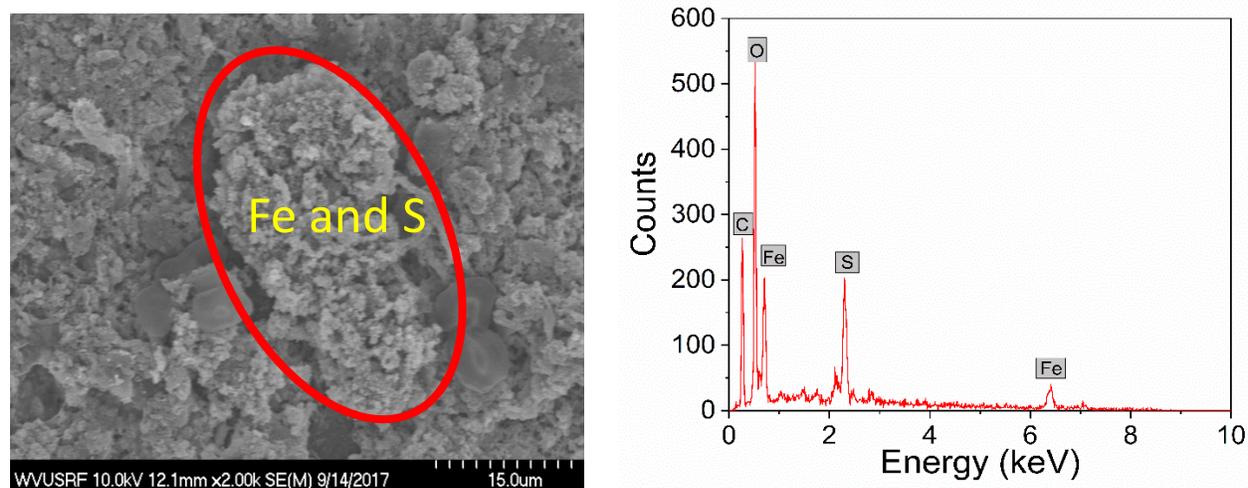


Fig. 9: Sample SEM micrograph and EDS spectrum of the sludge material under Fe/S molar ratio 3

The ZAF correction quantitative analysis indicated the weight and atomic percentage ratios of C, O, Fe, and S were comparable among the three Fe/S ratios ([Supplementary material, Fig. S4](#)). The variations in weight and atomic percentages of each element were observed as measurements taken at different locations of the sludge samples. The atomic percentage ratios of C: O: Fe: S were estimated as 47:34:9:10, 47:36:8:9 and 41:40:10:9 for Fe/S ratio 1, 2 and 3, respectively (Fig. S4b). The atomic percentage ratios of Fe and S were close to 1 suggesting the presence of iron sulfide (FeS) as the major inorganic constituent of the biogenic sludge materials. The sludge was primarily precipitate as amorphous FeS, but can be transformed to more crystalline form of FeS and FeS₂ during long-term operation (Deng et al., 2019).

For all the Fe/S ratios, similar XPS results were observed and XPS fitting for Fe/S ratio 3 is illustrated as a representative example. The peak fitting of Fe and S binding energies showed significant presence of ferrous iron and moderate presence of sulfide in the sludge (Fig. 10). The strong peaks observed for ferrous iron at 708.5 kV and sulfide at

162.4 kV and 161.2 kV suggested the presence of FeS and FeS₂ in the sludge sample (Descostes et al., 2000; Lennie and Vaughan, 1996). The excessive iron that was not utilized by microbial process was mostly in the form of ferric hydroxide and showed a peak at 711 eV in the Fe2p spectrum.

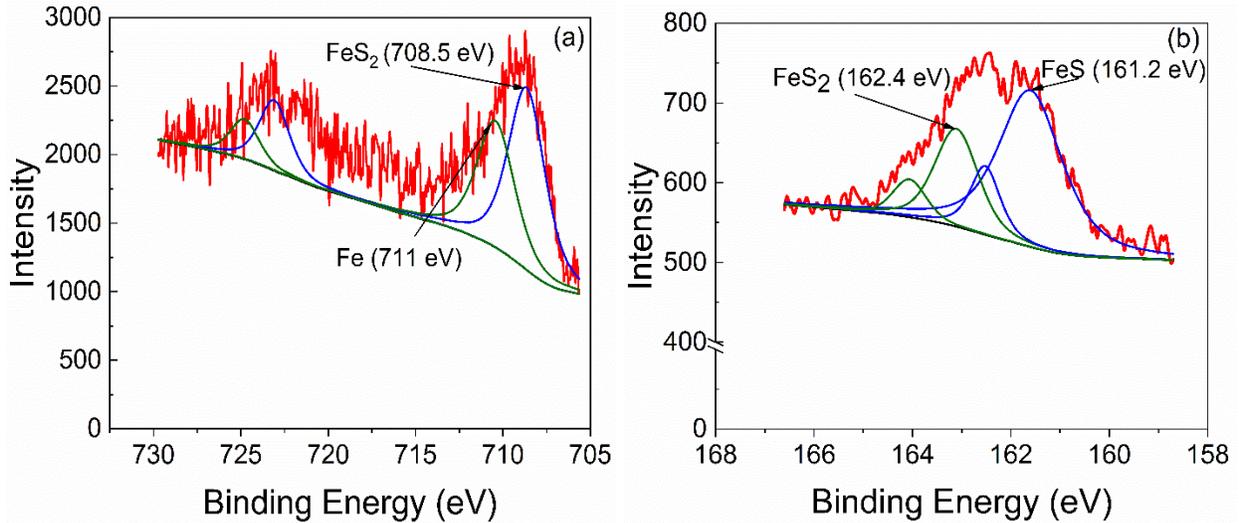


Fig. 10: XPS spectra of (a) Fe and (b) S in the sludge material under Fe/S molar ratio 3

3.3.4 Microbial community

16S rRNA genes affiliated with taxa spanning eight phyla were detected in the biofilm and sludge samples, including members of the Alphaproteobacteria, Gammaproteobacteria, Deltaproteobacteria, Acidobacteria, Actinobacteria, Bacteroidetes, Firmicutes and Euryarchaeota (Table 2).

Table 2: Major bacteria observed with their phyla and averaged relative abundance in all the bioreactors

Iron Reducing Bacteria	Sulfate Reducing Bacteria	Fermentative Bacteria
<ul style="list-style-type: none"> ▪ <i>Geothrix</i> sp. (.5%) (Acidobacteria) ▪ <i>Geobacter</i> sp. (5%) (Deltaproteobacteria) ▪ <i>Ignavibacteria</i> sp. (2%) (Bacteroidetes) 	<ul style="list-style-type: none"> ▪ <i>Desulfovibrio</i> sp. (40%) (Deltaproteobacteria) ▪ <i>Desulfobulbus</i> sp. (2%) (Deltaproteobacteria) 	<ul style="list-style-type: none"> ▪ <i>Sphingomonas</i> sp. (1%) (Alphaproteobacteria) ▪ <i>Micropepsaceae</i> sp. (7%) (Alphaproteobacteria) ▪ <i>Paludibacter</i> sp. (4%) (Bacteroidetes) ▪ <i>Clostridium sensu stricto</i> 9 (8%) (Firmicutes) ▪ <i>Ruminiclostridium</i> sp. (4%) (Firmicutes) ▪ <i>Acetivibrio</i> sp. (1%) (Firmicutes) ▪ <i>Cellulomonas</i> sp. (2.2%) (Actinobacteria)
Nitrogen Fixing Bacteria	Methanogens	Nitrate Reducing Bacteria
<ul style="list-style-type: none"> ▪ <i>Pleomorphomonas</i> sp. (19%) (Alphaproteobacteria) ▪ <i>Azovibrio</i> sp. (.3%) (Gammaproteobacteria) ▪ <i>Azospira</i> sp. (.1%) (Gammaproteobacteria) 	<ul style="list-style-type: none"> ▪ <i>Methanobacterium</i> sp. (1%) (Euryarchaeota) 	<ul style="list-style-type: none"> ▪ <i>Prolixibacteraceae</i> sp. (.4%) (Bacteroidetes)

Assigning functional roles to the detected microorganisms on the basis of 16S rRNA genes can be challenging, due to both the metabolic flexibility of some taxa and the limited knowledge of microbial metabolism in other taxonomic groups. However, conservative functional roles may be assigned in some instances where laboratory studies have identified conserved traits for specific taxonomic groups. Building on this concept, here we have classified *Geobacter* sp., *Geothrix* sp. and *Ignavibacteria* sp. as IRB. Members of the *Geobacteraceae* and *Geothrix* sp. are able to reduce ferric iron in laboratory studies

(Childers et al., 2002; Coates et al., 2001, 1999; Nevin et al., 2005), while a number of recent studies have implicated *Ignavibacteria* in iron reduction (Fortney et al., 2018; Podosokorskaya et al., 2013). Similarly, we have classified both *Desulfovibrio* sp. and *Desulfobulbus* sp. as SRB in this study, due to their well-characterized ability to reduce sulfate (El Houari et al., 2017; Rabus et al., 2015). Both *Geobacter* sp. and *Desulfovibrio* sp. were predominant across all the bioreactors, while *Desulfobulbus* sp. relative abundances ranged between 0-24 %.

With the exception of *Ignavibacteria*, these putative IRB and SRB are heterotrophic, and belong to the Deltaproteobacteria. Dominant *Geobacter* species generally utilize small organic substrates (e.g., acetate) that are oxidized to CO₂ (Esther et al., 2015; Lovley, 1993; Lovley et al., 1993), and may reduce insoluble iron oxides via direct contact with outer-membrane cytochromes and conductive pili structures. Acetate was present in the synthetic wastewater used in this experiment (30-600 mg/L), and may at least partly explain the dominance of *Geobacter* sp. in the communities. Similarly, both *Geobacter* sp. and *Desulfovibrio* sp. grow well at moderate temperatures (20 to 45°C) and near neutral pH (6-8.5) conditions found within the bioreactors (Straub et al., 2001).

SRB were identified as the major contributor for COD oxidation in the bioreactors although the higher Fe/S ratios promoted the contribution of IRB to COD oxidation. Thermodynamically, IRB have a competitive advantage over SRB for organic carbon substrates due to higher energy gain from the coupled redox reactions, which should have resulted in higher IRB biomass production than SRB. However, at a near neutral pH, ferric iron is present as Fe(OH)₃, which reduces its bioavailability to IRB. Quantification of microbial populations indicated that approximately 50 % of the community were putative SRB while the inferred IRB constituted 2 % of the total population under Fe/S ratio 1 (Fig. 11a). The SRB populations decreased to 34 % and 36 %, and correspondingly IRB increased to 8 % and 10 % relative abundances as the Fe/S ratio increased to 2 and 3, respectively. This result indicates a positive correlation between the inferred IRB and the COD oxidation rate. The dominance of *Desulfovibrio* sp. in all the bioreactors was likely linked to the presence of lactate in the synthetic wastewater used in this study (Jones et al., 1984), and had additional implications for in situ redox reactions. In addition to sulfate

reduction, *Desulfovibrio* sp. have been implicated in performing enzymatic ferric reduction in aquatic sediments and groundwater (Coleman et al., 1993; Lovley, 2000; Tebo and Obraztsova, 1998) and so may have contributed to the observed enzymatic iron reduction in the bioreactors.

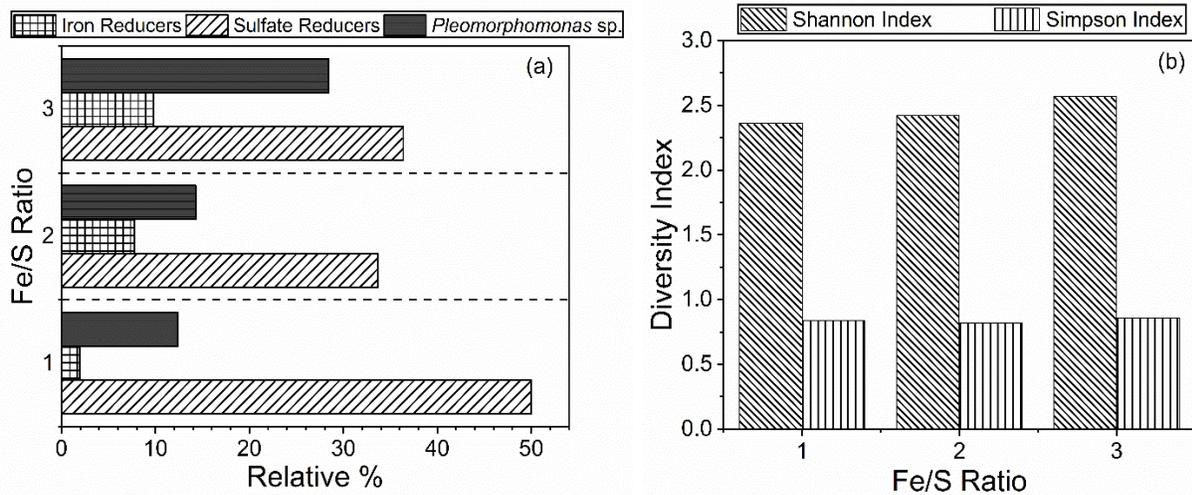


Fig. 11: (a) Relative abundances of major microbial species and (b) diversity index

Microbial diversity indices generally increased with the Fe/S ratio (Fig. 11b). Shannon's H, which measures both the diversity and evenness of a community, increased from low to high Fe/S ratio, and was calculated as 2.36, 2.42 and 2.57 for ratio 1, 2 and 3, respectively. Simpson's diversity index was estimated as 0.84, 0.82 and 0.86 for ratio 1, 2 and 3 respectively. Increases in microbial diversity within the community may support both increased functionality that results in more efficient substrate utilization, and resilience to disturbance of the environmental and chemical conditions (e.g., temperature, electron donors). Here, the reactor with Fe/S ratio 3 supported the most microbial diverse community – indicated by the highest Shannon's H index (2.57) and Simpson index (0.86) – and reported the highest COD oxidation rate. The low relative abundance of putative methanogens (~1 %) in these bioreactors suggests that IRB and SRB were able to successfully outcompete these microorganisms for methanogenic substrates (e.g., acetate, H₂).

Members of the genus *Pleomorphomonas* were also abundant in the bioreactors and their population increased with Fe/S ratio. Characterized isolates from this genus are able to

fix atmospheric nitrogen, suggesting that strains present in the bioreactor might be able to perform this same role where available N becomes limiting (Esquivel-Elizondo et al., 2018). N₂ gas sparged to maintain the anaerobic condition in this study was likely a nitrogen source for these N-fixing bacteria.

Fermentative bacteria affiliated with the genus *Clostridium* sp. (8 %) and genus *Ruminiclostridium* sp. (4 %) were also identified in the bioreactors, and likely played a key role in degrading more complex carbon substrates into labile compounds for oxidation by IRB and SRB. Fermentative clostridial species have previously been shown to play roles in iron reduction through the disposal of reducing equivalents, and may have contributed to iron reduction through such a mechanism in the bioreactors (Dobbin et al., 1999; Shah et al., 2014).

3.4 Conceptual Framework

Based on the findings of this study, a conceptual framework is proposed to illustrate the effects of Fe/S ratio on organic carbon oxidation rate of this iron-dosed treatment system (Fig. 12).

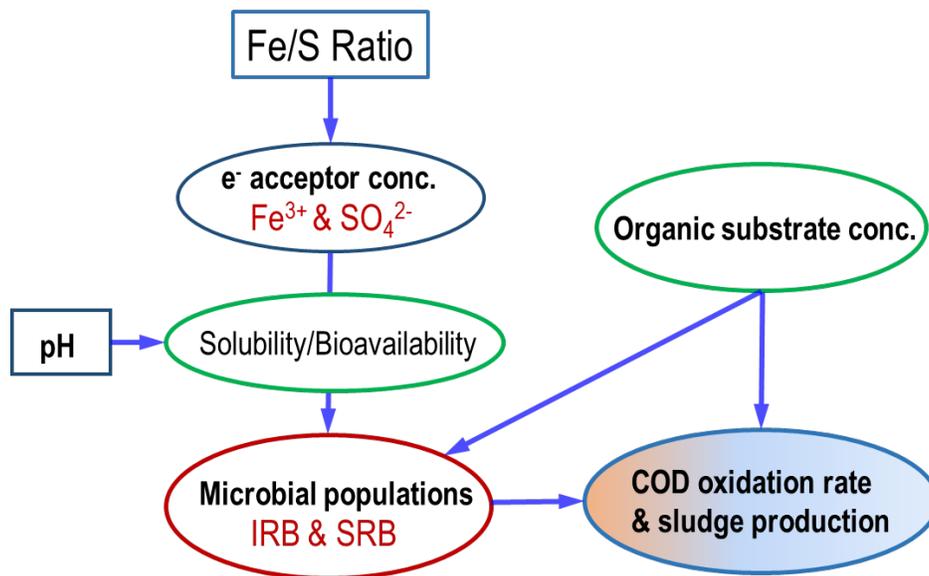


Fig. 12: Conceptual framework of the effects of Fe/S ratio on organic carbon oxidation kinetics

This study finds that Fe/S ratio plays a significant role in shaping multiple aspects of this iron-dosed wastewater treatment system. It regulates the quantities of ferric iron and sulfate as electron acceptors whose bioavailability to microorganisms is governed by their solubility and relevant environmental factors such as pH. Populations of the major reducers (IRB and SRB) and other microbial species are then regulated by the concentrations and bioavailability of the electron acceptors and the organic substrates as electron donors. The overall COD oxidation rate is dependent on the individual oxidation rates of IRB and SRB and their respective populations. The results showed that overall COD oxidation and sludge production rates were enhanced by increased Fe/S ratios. The COD oxidation rate has a positive correlation with abundance of IRB, suggesting that IRB facilitated faster COD oxidation kinetics than SRB.

3.5 Conclusion

Fe/S ratio was found to be an important factor that affects critical aspects of this Fe(III)-dosed treatment system, which include bioavailability of the two electron acceptors, microbial populations and distribution of IRB and SRB, organics oxidation rate, sludge production, and treated water quality. For practical applications of wastewater treatment and design, this ratio can be used for regulating the treatment performance and optimizing cost-effectiveness. Enhancing the organics oxidation rate can be achieved by increasing iron dosing and help reduce environmental footprint of the treatment process. However, excessive amounts of ferrous iron that may present in the treated water and cause biological instability in receiving waters need to be considered. Removing ferrous iron from the effluent can be achieved concurrently with disinfection using oxidants such as chlorine and provide an opportunity for collecting precipitated ferric hydroxide for recycled use of ferric iron. The inorganic fraction of the produced sludge consisted of mostly ferrous sulfides, and can potentially render beneficial applications given their magnetic and electrical properties. The proposed conceptual framework helps illustrate how iron dosing affects treatment kinetics, microbial ecology, and their interplays. It also provides a basis for further developing the treatment process. Future studies are required on continuous, long-time operations of this novel process, sludge composition, microbial ecology evolution in the bioreactors, and methods for enhancing ferric iron bioavailability.

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3.6 Supplementary Materials

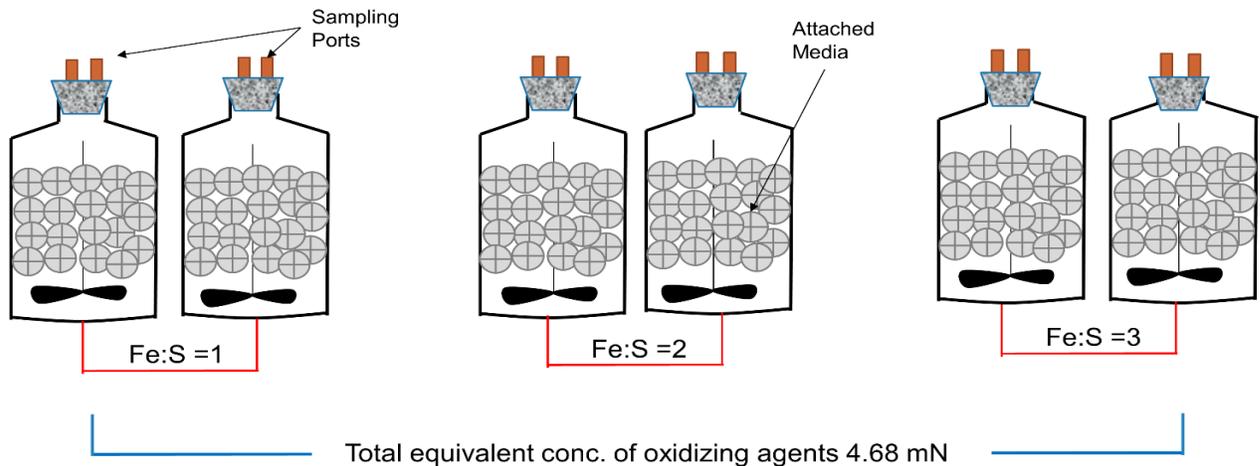


Fig. S1: Experimental Set-up of Batch Reactors

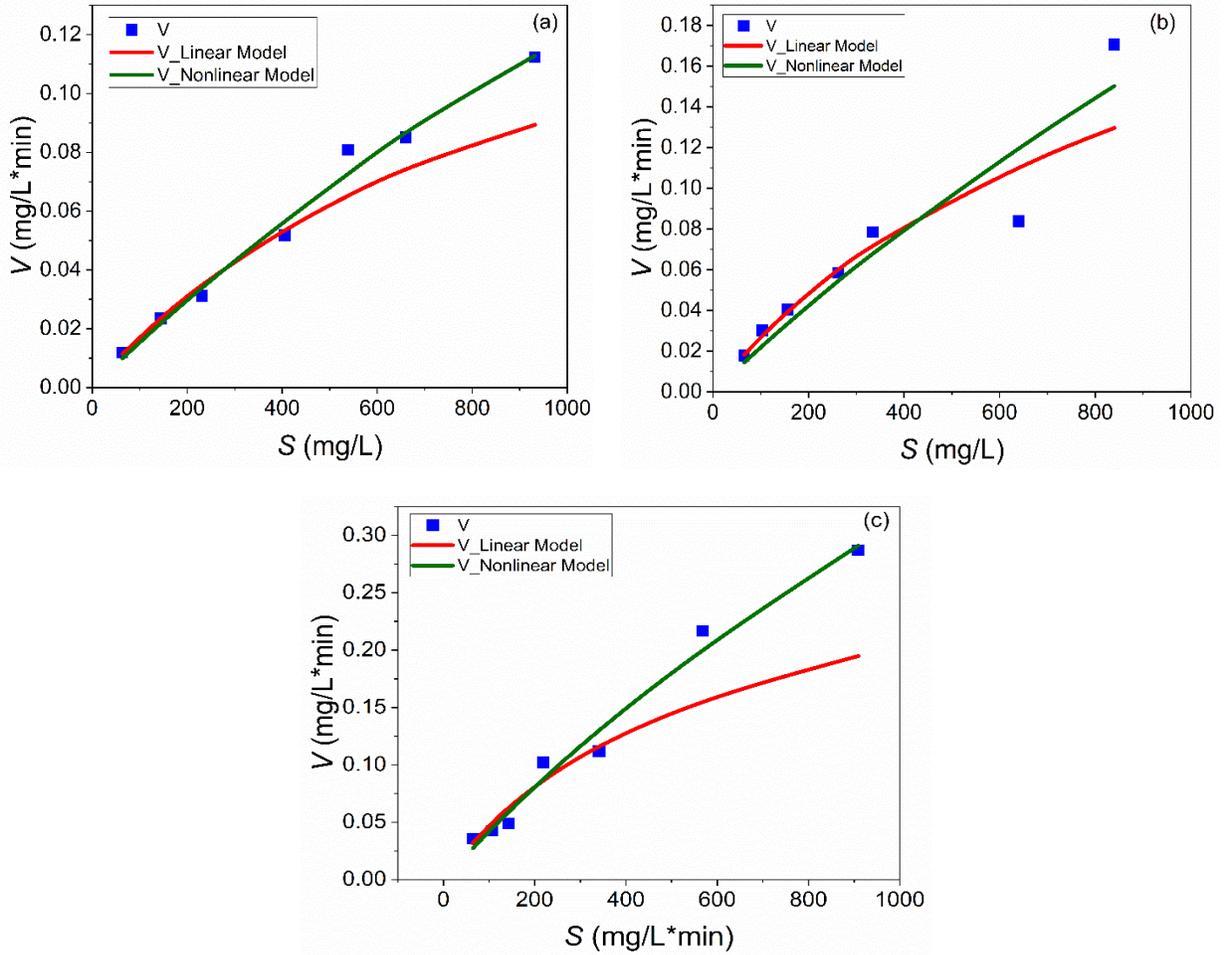


Fig. S2: Linear and Nonlinear Models for three different ratios (a) Fe/S =1 (b) Fe/S =2 (c) Fe/S=3

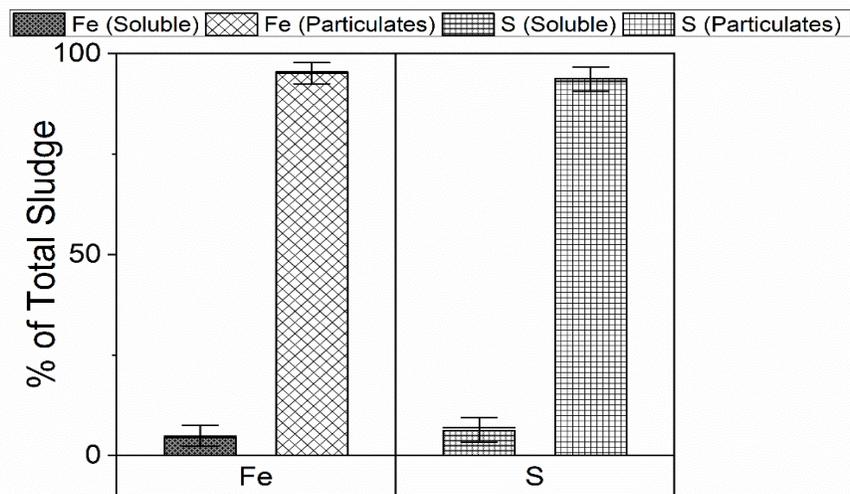


Fig. S3: Fe and S percentages in the sludge materials from the bioreactors

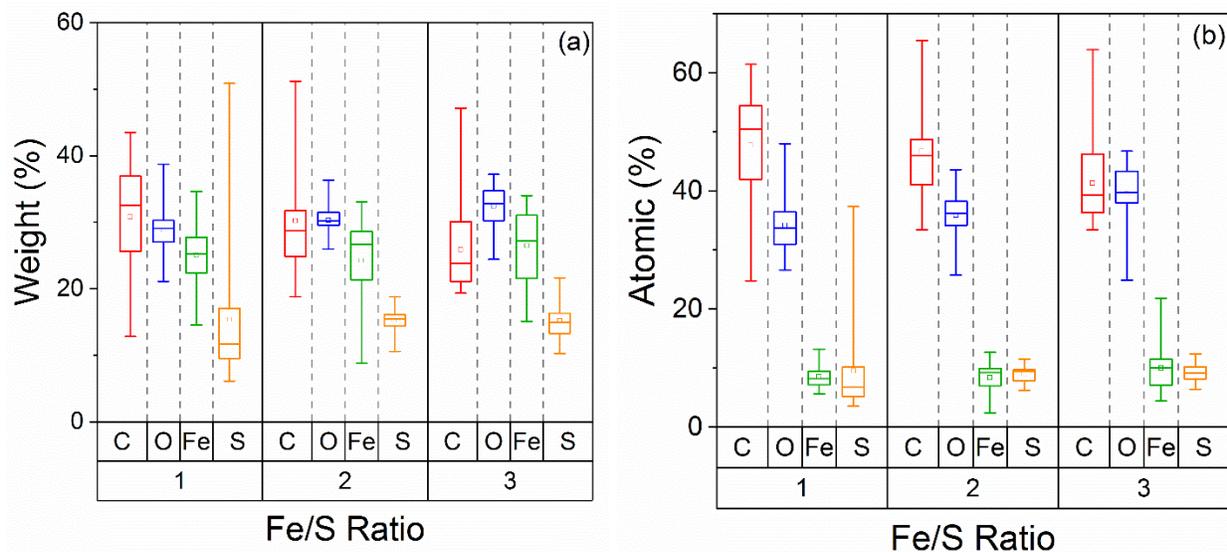


Fig. S4: (a) Weight and (b) atomic percentages of selected chemical elements in the sludge materials from EDS analysis

References

- Ahmed, M., Lin, L.-S., 2017. Ferric reduction in organic matter oxidation and its applicability for anaerobic wastewater treatment: a review and future aspects. *Rev. Environ. Sci. Biotechnol.* 16, 273–287. <https://doi.org/10.1007/s11157-017-9424-3>
- Aklujkar, M., Young, N.D., Holmes, D., Chavan, M., Risso, C., Kiss, H.E., Han, C.S., Land, M.L., Lovley, D.R., 2010. The genome of *Geobacter bemidjensis*, exemplar for the subsurface clade of *Geobacter* species that predominate in Fe(III)-reducing subsurface environments. *BMC Genomics* 11, 1–18. <https://doi.org/10.1186/1471-2164-11-490>
- APHA, AWWA, WEF, 2005. Standard methods for the examination of water and wastewater, 21st ed. American Public Health Association, American Water Works Association, and Water Environment Federation.
- Bonneville, S., Behrends, T., Cappellen, P. Van, Hyacinthe, C., Röling, W.F.M., 2006. Reduction of Fe(III) colloids by *Shewanella putrefaciens*: A kinetic model. *Geochim. Cosmochim. Acta* 70, 5842–5854. <https://doi.org/10.1016/j.gca.2006.04.029>
- Bonneville, S., Behrends, T., Van Cappellen, P., 2009. Solubility and dissimilatory reduction kinetics of iron(III) oxyhydroxides: A linear free energy relationship. *Geochim. Cosmochim. Acta* 73, 5273–5282. <https://doi.org/10.1016/j.gca.2009.06.006>
- Bonneville, S., Van Cappellen, P., Behrends, T., 2004. Microbial reduction of iron(III) oxyhydroxides: Effects of mineral solubility and availability. *Chem. Geol.* 212, 255–268. <https://doi.org/10.1016/j.chemgeo.2004.08.015>
- Bratkova, S., Lavrova, S., Angelov, A., Nikolova, K., Ivanov, R., Koumanova, B., 2018.

- Treatment of wastewaters containing Fe, Cu, Zn and as by microbial hydrogen sulfide and subsequent removal of COD, N and P. *J. Chem. Technol. Metall.* 53, 245–257.
- Cadena, F., Peters, R.W., 1988. Evaluation of Chemical Oxidizers for Hydrogen Sulfide Control. *Water Pollut. Control Fed.* 60, 1259–1263. <https://doi.org/10.2307/25043633>
- Childers, S.E., Ciufo, S., Lovley, D.R., 2002. *Geobacter metallireducens* accesses insoluble Fe(III) oxide by chemotaxis. *Nature* 416, 767–769. <https://doi.org/10.1038/416767a>
- Cirne, D.G., Van Der Zee, F.P., Fernandez-Polanco, M., Fernandez-Polanco, F., 2008. Control of sulphide during anaerobic treatment of S-containing wastewaters by adding limited amounts of oxygen or nitrate. *Rev. Environ. Sci. Biotechnol.* 7, 93–105. <https://doi.org/10.1007/s11157-008-9128-9>
- Coates, J.D., Bhupathiraju, V.K., Achenbach, L.A., McInerney, M.J., Lovley, D.R., 2001. *Geobacter hydrogenophilus*, *Geobacter chappellei* and *Geobacter grbciaie*, three new, strictly anaerobic, dissimilatory Fe(III)-reducers. *Int. J. Syst. Evol. Microbiol.* 51, 581–588.
- Coates, J.D., Ellis, D.J., Gaw, C. V, Lovley, D.R., 1999. *Geothrix fermentans* gen. nov., sp. nov., a novel Fe(III)-reducing bacterium from a hydrocarbon-contaminated aquifer. *Int. J. Syst. Evol. Microbiol.* 49, 1615–1622.
- Coleman, M.L., Hedrick, D.B., Lovley, D.R., White, D.C., Pye, K., 1993. Reduction of Fe(III) in sediments by sulphate-reducing bacteria. *Nature* 361, 436–438. <https://doi.org/10.1038/361436a0>
- Csákberényi-Malasics, D., Rodriguez-Blanco, J.D., Kis, V.K., Rečnik, A., Benning, L.G., Pósfai, M., 2012. Structural properties and transformations of precipitated FeS. *Chem. Geol.* 294–295, 249–258. <https://doi.org/10.1016/j.chemgeo.2011.12.009>
- Deng, D., Lin, L.S., 2017. Continuous sulfidogenic wastewater treatment with iron sulfide sludge oxidation and recycle. *Water Res.* 114, 210–217. <https://doi.org/10.1016/j.watres.2017.02.048>
- Deng, D., Lin, O., Rubenstein, A., Weidhaas, J.L., Lin, L.-S., 2019. Elucidating biochemical transformations of Fe and S in an innovative Fe(II)-dosed anaerobic wastewater treatment process using spectroscopic and phylogenetic analyses. *Chem. Eng. J.* 358, 1208–1217. <https://doi.org/10.1016/J.CEJ.2018.10.030>
- Deng, D., Weidhaas, J.L., Lin, L.-S., 2016. Kinetics and microbial ecology of batch sulfidogenic bioreactors for co-treatment of municipal wastewater and acid mine drainage. *J. Hazard. Mater.* 305, 200–8. <https://doi.org/10.1016/j.jhazmat.2015.11.041>
- Descostes, M., Mercier, F., Thromat, N., Beaucaire, C., Gautier-Soyer, M., 2000. Use of XPS in the determination of chemical environment and oxidation state of iron and sulfur samples: Constitution of a data basis in binding energies for Fe and S reference compounds and applications to the evidence of surface species of an

- oxidized py. *Appl. Surf. Sci.* 165, 288–302. [https://doi.org/10.1016/S0169-4332\(00\)00443-8](https://doi.org/10.1016/S0169-4332(00)00443-8)
- Dobbin, P.S., Carter, J.P., Juan, C.G.S.S., Von Höbe, M., Powell, A.K., Richardson, D.J., 1999. Dissimilatory Fe(III) reduction by *Clostridium beijerinckii* isolated from freshwater sediment using Fe(III) maltol enrichment. *FEMS Microbiol. Lett.* 176, 131–138. [https://doi.org/10.1016/S0378-1097\(99\)00229-3](https://doi.org/10.1016/S0378-1097(99)00229-3)
- El Houari, A., Ranchou-Peyruse, M., Ranchou-Peyruse, A., Dakdaki, A., Guignard, M., Idouhammou, L., Bennisse, R., Bouterfass, R., Guyoneaud, R., Qatibi, A.I., 2017. *Desulfobulbus oligotrophicus* sp. Nov., a sulfate-reducing and propionate-oxidizing bacterium isolated from a municipal anaerobic sewage sludge digester. *Int. J. Syst. Evol. Microbiol.* 67, 275–281. <https://doi.org/10.1099/ijsem.0.001615>
- Esquivel-Elizondo, S., Maldonado, J., Krajmalnik-Brown, R., 2018. Anaerobic carbon monoxide metabolism by *Pleomorphomonas carboxyditropha* sp. nov., a new mesophilic hydrogenogenic carboxydrotroph. *FEMS Microbiol. Ecol.* 94, 1–12. <https://doi.org/10.1093/femsec/fiy056>
- Esther, J., Sukla, L.B., Pradhan, N., Panda, S., 2015. Fe (III) reduction strategies of dissimilatory iron reducing bacteria. *Korean J. Chem. Eng.* 32, 1–14. <https://doi.org/10.1007/s11814-014-0286-x>
- Fortney, N.W., He, S., Kulkarni, A., Friedrich, M.W., Holz, C., Boyd, E.S., Roden, E.E., 2018. Stable isotope probing for microbial iron reduction in Chocolate Pots hot spring, Yellowstone National Park. *Appl. Environ. Microbiol.* 84. <https://doi.org/10.1128/AEM.02894-17>
- Gerardi, M.H., 2006. *Wastewater bacteria*. John Wiley & Sons, New Jersey.
- Gramp, J.P., Bigham, J.M., Jones, F.S., Tuovinen, O.H., 2010. Formation of Fe-sulfides in cultures of sulfate-reducing bacteria. *J. Hazard. Mater.* 175, 1062–1067. <https://doi.org/10.1016/j.jhazmat.2009.10.119>
- Hegyi, G., Kardos, J., Kovács, M., Málnási-Csizmadia, A., Nyitray, L., Pál, G., Radnai, L., Reményi, A., Venekei, I., 2013. *Introduction to Practical Biochemistry*. Eötvös Loránd University, Hungary.
- Herbert, R.B., Benner, S.G., Pratt, A.R., Blowes, D.W., 1998. Surface chemistry and morphology of poorly crystalline iron sulfides precipitated in media containing sulfate-reducing bacteria. *Chem. Geol.* 144, 87–97. [https://doi.org/10.1016/S0009-2541\(97\)00122-8](https://doi.org/10.1016/S0009-2541(97)00122-8)
- Horikawa, M.S., Rossi, F., Gimenes, M.L., Costa, C.M.M., Da Silva, M.G.C., 2004. Chemical absorption of H₂S for biogas purification. *Brazilian J. Chem. Eng.* 21, 415–422. <https://doi.org/10.1590/S0104-66322004000300006>
- Jensen, M.M., Thamdrup, B., Rysgaard, S., Holmer, M., Fossing, H., 2003. Rates and Regulation of Microbial Iron Reduction in Sediments of the Baltic-North Sea Transition 65, 295–317.
- Jones, J., Davison, W., Gardener, S., 1984. Iron reduction by bacteria: range of

- organisms involved and metals reduced. *FEMS Microbiol. Lett.* 21, 133–136.
- Kaksonen, A.H., Plumb, J.J., Robertson, W.J., Riekkola-Vanhanen, M., Franzmann, P.D., Puhakka, J.A., 2006. The performance, kinetics and microbiology of sulfidogenic fluidized-bed treatment of acidic metal- and sulfate-containing wastewater. *Hydrometallurgy* 83, 204–213. <https://doi.org/10.1016/j.hydromet.2006.03.025>
- Kapdi, S.S., Vijay, V.K., Rajesh, S.K., Prasad, R., 2005. Biogas scrubbing, compression and storage: Perspective and prospectus in Indian context. *Renew. Energy* 30, 1195–1202. <https://doi.org/10.1016/j.renene.2004.09.012>
- Khadse, G.K., Patni, P.M., Labhasetwar, P.K., 2015. Removal of Iron and Manganese from drinking water supply. *Sustain. Water Resour. Manag.* 1, 157–165. <https://doi.org/10.1021/ie50297a006>
- Lennie, A.R., Vaughan, D.J., 1996. Spectroscopic studies of iron sulfide formation and phase relations at low temperatures, in: Dyar, M.D., McCammon, C., Schaefer, M.W. (Eds.), *Mineral Spectroscopy: A Tribute to Roger G. Burns*. pp. 117–131.
- Li, C., Jun, X., Jing, C., 2015. Applications of scanning electron microscopy in earth sciences. *Sci. China Earth Sci.* 58, 1768–1778. <https://doi.org/10.1007/s11430-015-5172-9>
- Liu, C., Gorby, Y.A., Zachara, J.M., Fredrickson, J.K., Brown, C.F., 2002. Reduction kinetics of Fe(III), Co(III), U(VI), Cr(VI), and Tc(VII) in cultures of dissimilatory metal-reducing bacteria. *Biotechnol. Bioeng.* 80, 637–649. <https://doi.org/10.1002/bit.10430>
- Liu, J., Zhao, Z., Jiang, G., 2008. Coating Fe₃O₄ Magnetic Nanoparticles with Humic Acid for High Efficient Removal of Heavy Metals in Water. *Environ. Sci. Technol.* 42, 6949–6954. <https://doi.org/10.1021/es800924c>
- Lovley, D.R., 2000. Fe(III) and Mn(IV) Reduction, in: Lovley, D.R. (Ed.), *Environmental Microbe-Metal Interactions*. ASM Press, Washington D.C., pp. 3–30.
- Lovley, D.R., 1995. Bioremediation of organic and metal contaminants with dissimilatory metal reduction. *J. Ind. Microbiol.* 14, 85–93. <https://doi.org/10.1007/BF01569889>
- Lovley, D.R., 1993. Dissimilatory metal reduction. *Annu. Rev. Microbiol.* 47, 20–29.
- Lovley, D.R., 1987. Organic matter mineralization with the reduction of ferric iron: A review. *Geomicrobiol. J.* 5, 375–399. <https://doi.org/10.1080/01490458709385975>
- Lovley, D.R., Giovannoni, S.J., White, D.C., Champine, J.E., Phillips, E.J.P., Gorby, Y.A., Goodwin, S., 1993. *Geobacter metallireducens* gen. nov. sp. nov., a microorganism. *Arch. Microbiol.* 159, 336–344.
- Lovley, D.R., Phillips, E.J.P., 1989. Requirement for a microbial consortium to completely oxidize glucose in Fe(III)-reducing sediments. *Appl. Environ. Microbiol.* 55, 3234–3236.
- Mahadevan, R., Palsson, B., Lovley, D.R., 2011. In situ to in silico and back: Elucidating the physiology and ecology of *Geobacter* spp. using genome-scale modelling. *Nat. Rev. Microbiol.* 9, 39–50. <https://doi.org/10.1038/nrmicro2456>

- Martin, R.B., 1997. Disadvantages of Double Reciprocal Plots. *J. Chem. Educ.* 74, 1238. <https://doi.org/10.1021/ed074p1238>
- Mosey, F., Swanwick, J., Hughes, D.A., 1971. Factors Affecting the Availability of Heavy Metals to Inhibit Anaerobic Digestion. *J. Water Pollut. Control Fed.* 71, 668–678.
- Myers, J.M., Myers, C.R., 2001. Role for outer membrane cytochromes OmcA and OmcB of *Shewanella putrefaciens* MR-1 in reduction of manganese dioxide. *Appl. Environ. Microbiol.* 67, 260–269. <https://doi.org/10.1128/AEM.67.1.260-269.2001>
- Nevin, K.P., Holmes, D.E., Woodard, T.L., Hinlein, E.S., Ostendorf, D.W., Lovley, D.R., 2005. *Geobacter bemidjiensis* sp. nov. and *Geobacter psychrophilus* sp. nov., two novel Fe(III)-reducing subsurface isolates. *Int. J. Syst. Evol. Microbiol.* 55, 1667–1674. <https://doi.org/10.1099/ijs.0.63417-0>
- Nevin, K.P., Lovley, D.R., 2002. Mechanisms for Fe(III) Oxide Reduction in Sedimentary Environments. *Geomicrobiol. J.* 19, 141–159.
- Perrin, C.L., 2017. Linear or Nonlinear Least-Squares Analysis of Kinetic Data? *J. Chem. Educ.* 94, 669–672. <https://doi.org/10.1021/acs.jchemed.6b00629>
- Petersson, A., Wellinger, A., 2009. Biogas upgrading technologies – developments and innovations Task 37 -Energy from biogas and landfill gas IeA Bioenergy aims to accelerate the use of environmental sound and cost-competitive Bioenergy on a sustainable basis, and thereby achieve a substant. IEA Bioenergy 13.
- Podosokorskaya, O.A., Kadnikov, V. V., Gavrilov, S.N., Mardanov, A. V., Merkel, A.Y., Karnachuk, O. V., Ravin, N. V., Bonch-Osmolovskaya, E.A., Kublanov, I. V., 2013. Characterization of *Melioribacter roseus* gen. nov., sp. nov., a novel facultatively anaerobic thermophilic cellulolytic bacterium from the class Ignavibacteria, and a proposal of a novel bacterial phylum Ignavibacteriae. *Environ. Microbiol.* 15, 1759–1771. <https://doi.org/10.1111/1462-2920.12067>
- Postma, D., 1993. The reactivity of iron oxides in sediments: A kinetic approach. *Geochim. Cosmochim. Acta* 57, 5027–5034. [https://doi.org/10.1016/S0016-7037\(05\)80015-8](https://doi.org/10.1016/S0016-7037(05)80015-8)
- Rabus, R., Venceslau, S.S., Wöhlbrand, L., Voordouw, G., Wall, J.D., Pereira, I.A.C., 2015. A Post-Genomic View of the Ecophysiology, Catabolism and Biotechnological Relevance of Sulphate-Reducing Prokaryotes. *Adv. Microb. Physiol.* 66, 55–321. <https://doi.org/10.1016/bs.ampbs.2015.05.002>
- Roden, E.E., Wetzal, R.G., 2002. Kinetics of microbial Fe(III) oxide reduction in freshwater wetland sediments. *Limnol. Oceanogr.* 47, 198–211. <https://doi.org/10.4319/lo.2002.47.1.0198>
- Shah, M., Lin, C.-C., Kukkadapu, R., Engelhard, M.H., Zhao, X., Wang, Y., Barkay, T., Yee, N., 2014. Syntrophic Effects in a Subsurface Clostridial Consortium on Fe(III)- (Oxyhydr)oxide Reduction and Secondary Mineralization. *Geomicrobiol. J.* 31, 101–115.
- Straub, K.L., Benz, M., Schink, B., 2001. Iron metabolism in anoxic environments at near

- neutral pH. *FEMS Microbiol. Ecol.* 34, 181–186. [https://doi.org/10.1016/S0168-6496\(00\)00088-X](https://doi.org/10.1016/S0168-6496(00)00088-X)
- Tebo, B.M., Obraztsova, A.Y., 1998. Sulfate-reducing bacterium grows with Cr (VI), U (VI), Mn (IV), and Fe (III) as electron acceptors. *Source* 162, 193–198. <https://doi.org/10.1111/j.1574-6968.1998.tb12998.x>
- Thamdrup, B., 2000. Bacterial Manganese and Iron Reduction in Aquatic Sediments. *Adv. Microb. Ecol.* 16, 41–84. https://doi.org/10.1007/978-1-4615-4187-5_2
- Thorpe, A.N., Senftle, F.E., Alexander, C.C., Dulong, F.T., 1984. Oxidation of pyrite in coal to magnetite. *Fuel* 63, 662–668. [https://doi.org/10.1016/0016-2361\(84\)90163-7](https://doi.org/10.1016/0016-2361(84)90163-7)
- Urrutia, M.M., Roden, E.E., Fredrickson, J.K., Zachara, J.M., 1998. Microbial and surface chemistry controls on reduction of synthetic Fe(III) oxide minerals by the dissimilatory iron-reducing bacterium *Shewanella* alga. *Geomicrobiol. J.* 15, 269–291.
- Utgikar, V.P., Harmon, S.M., Chaudhary, N., Tabak, H.H., Govind, R., Haines, J.R., 2002. Inhibition of sulfate-reducing bacteria by metal sulfide formation in bioremediation of acid mine drainage. *Environ. Toxicol.* 17, 40–48. <https://doi.org/10.1002/tox.10031>
- Vaclavkova, S., Jørgensen, C.J., Jacobsen, O.S., Aamand, J., Elberling, B., 2014. The Importance of Microbial Iron Sulfide Oxidation for Nitrate Depletion in Anoxic Danish Sediments. *Aquat. Geochemistry* 20, 419–435. <https://doi.org/10.1007/s10498-014-9227-x>
- Van der Maas, P., 2005. Fe(III) EDTA reduction in BioDeNOx reactors: kinetics and mechanism, in: *Chemically Enhanced Biological NOx Removal from Flue Gases*. Wageningen University, pp. 109–124.
- Van Lier, J.B., 2008. High-rate anaerobic wastewater treatment: Diversifying from end-of-the-pipe treatment to resource-oriented conversion techniques. *Water Sci. Technol.* 57, 1137–1148. <https://doi.org/10.2166/wst.2008.040>
- Van Lier, J.B., Van der Zee, F.P., Frijters, C.T.M.J., Ersahin, M.E., 2015. Celebrating 40 years anaerobic sludge bed reactors for industrial wastewater treatment. *Rev. Environ. Sci. Biotechnol.* 14, 681–702. <https://doi.org/10.1007/s11157-015-9375-5>
- Waters, K.E., Rowson, N.A., Greenwood, R.W., Williams, A.J., 2008. The effect of heat treatment on the magnetic properties of pyrite 21, 679–682. <https://doi.org/10.1016/j.mineng.2008.01.008>
- Zhang, J., Zhang, Y., Chang, J., Quan, X., Li, Q., 2013. Biological sulfate reduction in the acidogenic phase of anaerobic digestion under dissimilatory Fe (III) - Reducing conditions. *Water Res.* 47, 2033–2040. <https://doi.org/10.1016/j.watres.2013.01.034>

Chapter 4: Continuous Ferric Iron-dosed Anaerobic Wastewater Treatment: Treatment Performance, Sludge Characteristics, and Microbial Composition

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4.1 Introduction

In the efforts of moving towards more sustainable wastewater management, anaerobic treatment is increasing in popularity due to its energy efficiency, cost effectiveness, low sludge production and greenhouse emission, as well as the potential for resource recovery (Chan et al., 2009; Van Lier et al., 2015). Nitrate, sulfate and carbon dioxide are electron acceptors commonly used in anaerobic biological processes to treat wastewater (Damianovic and Foresti, 2007; Hubert and Voordouw, 2007). Due to its highly redox-active nature, the prospect of using ferric iron as an electron acceptor in wastewater treatment was recently reviewed and demonstrated in a batch study (Ahmed et al., 2019; Ahmed and Lin, 2017). Iron, the second most abundant mineral on earth, is often found in wastes (e.g., acid mine drainage and coal ash) and these wastes can be readily used as a cheap source of iron for wastewater treatment. Iron has been commonly used for coagulation, Fenton's reagent, and sulfide toxicity and odor control in wastewater treatment (Waite, 2002; Wang et al., 2016; Zhang et al., 2015). A recent study found that Fe(III) dosing in wastewater treatment can also promote removal of organic micropollutants via adsorption of the compounds on iron sulfide (FeS) surface and subsequent biodegradation (Kulandaivelu et al., 2019). In addition, biogenic iron sulfide sludge produced as a byproduct of iron-dosed wastewater treatment has great applications in remediation of soil and groundwater contamination as well as in wastewater treatment. Iron sulfides and nanoparticles synthesized from iron sulfides were found effective in removal of organic contaminants (benzene, chlorinated organic pollutants, aromatic hydrocarbons), toxic metals (As, Pb, Hg, Cd, Cr), nutrients (N and P), and radionuclides (U and Se) (Gong et al., 2016; Yang et al., 2017). Moreover, this iron sulfide sludge can potentially be converted into useful products with magnetic properties (e.g., magnetite) for phosphorus recovery from wastewater (Thorpe et al.,

1984a; Wang et al., 2008; Waters et al., 2008; Xiang et al., 2014). This iron-based wastewater treatment does not produce biogas as in methanogenic process. However, its multiple potential benefits (use of iron-containing wastes, no aeration, unique reaction mechanisms for coagulation, sulfide control, organic micropollutant removal, and useful sludge byproduct) render this treatment method versatile. It can be tailored to meet treatment needs such as decentralized treatment with low operation and maintenance requirements.

In Fe(III)-dosed anaerobic wastewater treatment, both Iron reducing bacteria (IRB) and sulfate reducing bacteria (SRB) can contribute to microbial oxidation of organics given the prevalent presence of sulfate in wastewater. Under substrate limiting conditions, IRB can outcompete SRB by diverting the electron flow away from SRB (Lovley and Phillips, 1989). However, for treatment applications of wastewaters, in which substrate limiting is unlikely (Chemical Oxygen Demand, COD, 339-1,016 mg/L, (Metcalf & Eddy et al., 2014)), the symbiotic relationship between IRB and SRB is currently unknown. In such a treatment system, availability of ferric iron and sulfate is expected to regulate the microbial activities of IRB and SRB, and the overall treatment performance. In addition, the treatment effectiveness also depends on the types of ferric compound, pH, organic compounds and their concentrations as well as reactor configuration (Azam and Finneran, 2013; Lovley, 1987).

Geobacter sp. and *Shewanella sp.* are two of the most well-studied iron-reducing taxa in the natural environment (Weber et al., 2006). The interactions of these iron reducers with insoluble ferric surface are quite dissimilar from each other. *Shewanella sp.* was observed to have direct and indirect (ligand, electron shuttle) electron transfer to insoluble ferric surface, whereas *Geobacter sp.* was often found to rely on pilin filaments for electron transfer (Esther et al., 2015; Weber et al., 2006). Some of the IRB were observed to be closely linked with SRB belonging to the same taxa and utilizing same electron acceptor for growth. For example, *Desulfovibrio sp.*, a common SRB, was observed to perform ferric reduction and organic compound oxidation in an iron reducing environment (Lovley, 1993). In an engineering treatment system, the abundances of these reducers and other microorganisms are dependent on the concentrations and bioavailability of the electron

acceptors (i.e., ferric iron vs. sulfate) and organic compounds. The respective populations of IRB and SRB in turn affect the treatment performance of organics oxidation kinetics (Ahmed et al., 2019).

Building upon a previous fed-batch study of reaction kinetics (Ahmed et al., 2019), the objective of this study was to investigate the long-term performance of continuous Fe(III)-dosed anaerobic biological treatment of sulfate-containing wastewater under varied ferric and sulfate concentrations (expressed as Fe/S ratios). Specifically, the study focused on how Fe/S ratio affects treatment performance, sludge production, and microbial composition. The treatment performance was characterized in terms of organics removal efficiency, removal rate, and effluent quality. In addition, mass flow rates of Fe and S were estimated for understanding the biogeochemical transformations of the elements throughout the treatment system. Microbiological analyses were performed to elucidate the relationship between microbial communities and both treatment performance and sludge production.

4.2 Materials and Methods

4.2.1 Bench-scale iron-dosed treatment system

The treatment system consists of a wastewater tank, a ferric iron solution reservoir, a bioreactor and an effluent collection tank ([Supplementary Material, Fig. S5](#)).

4.2.1.1 Wastewater Tank

A 4-L tank was used to feed wastewater in the bioreactor of the treatment system. A solution containing sodium acetate anhydrous ($C_2H_3NaO_2$, 3 mM), 1.54 mM ethanol (C_2H_6O , 1.54 mM), lactose monohydrate ($C_{12}H_{22}O_{11} \cdot H_2O$, 0.32 mM), sodium bicarbonate ($NaHCO_3$, 1.57 mM), and trace elements (4.75 mL/L) (Deng and Lin, 2017) was prepared as a synthetic wastewater in this study. This synthetic wastewater was used as a base solution to maintain a consistent level of organics (COD=420 mg/L). A sodium sulfate solution (Na_2SO_4) was used to adjust the sulfate concentration in the wastewater for different Fe/S ratios.

4.2.1.2 Ferric Chloride Reservoir

A 2-L tank containing a ferric chloride solution ($\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$, 1.32 mM, 2.50 mM and 4.50 mM) was used to feed ferric iron to the bioreactor to obtain Fe/S molar ratio 0.5, 1 and 2 respectively. The ferric solution pH was adjusted to 4-4.2 using a sodium hydroxide solution (NaOH, 5N).

4.2.1.3 Bioreactor

The bioreactor (1.4 L) was made of acrylic cylinder with ports on the top for wastewater and ferric iron inflows. Two ports on the side of the reactor were used for internal recirculation to enhance hydraulic mixing in the reactor. The reactor was packed with five hundred plastic media (Evolution Aqua Ltd., UK, Kaldness K1 Biomedia, specific surface area = $500 \text{ m}^2/\text{m}^3$) for attached growth of microorganisms. This resulted in a working volume of 0.9 L in the reactor. A perforated acrylic plate was used to support the packing media and a cone-shaped bottom was used for sludge settling and collection. The reactor was first inoculated with anaerobic sludge from a wastewater treatment plant (Star City, West Virginia) and acid mine drainage (St. Thomas, Morgantown, West Virginia) at 1:1 volume ratio. The bioreactor was purged with N_2 gas regularly prior to the operation and sealed airtight to maintain an anaerobic condition. The bioreactor was operated at room temperature and given three-month enrichment period with continuous feeding of the synthetic wastewater containing a Fe/S molar ratio of 0.5.

4.2.2 Treatment Experiments

The treatment performance was evaluated under three different Fe/S molar ratios (0.5, 1, and 2) while maintaining the same total equivalent concentration of Fe(III) and sulfate (Table 3). The bioreactor was operated continuously for 500 days, during which the flow rates of influent (0.67 L/d), ferric solution (0.53 L/d), and internal recirculation (3.4 L/d) were kept constant. During the treatment, the ferric chloride and sulfate loadings to the bioreactor were varied to obtain the three target Fe/S molar ratios. The total equivalent concentration of the electron acceptors was maintained the same for all three Fe/S molar ratios (11.9 mN/d) to have the same total reducing capacity for organic matter oxidation.

Table 3: Daily loads of Fe and S for three target Fe/S molar ratios

Fe/S molar ratio	Fe loading (mmol/d)	S loading (mmol/d)
0.5	0.70	1.40
1	1.32	1.32
2	2.38	1.19

Before changing the ratio of Fe/S, all solutions and sludges were removed from the bioreactor and replaced with fresh synthetic wastewater. Under each Fe/S ratio, the first month was allowed for microorganisms to adapt to the chemical condition and to reach a steady state and treatment in the following three months was considered under the steady state condition. The influent and effluent samples were collected daily and later intermittently for COD, sulfate, total iron, sulfide and ferrous iron analyses. All samples were stored at 4°C until analysis except sulfide and ferrous iron samples which were analyzed immediately to avoid oxidation. Influent and effluent COD, sulfate and total iron concentrations were used to estimate the COD removal efficiency, sulfate reduction and total iron retention. In this study, iron retention is referred to iron retained in the bioreactor and the sludge. COD removal rate was calculated using the influent and effluent COD concentrations under the steady-state condition (Deng and Lin, 2017). The sludge samples were collected monthly for solid measurements, microscopic and microbiological analyses.

4.2.3 Analytical Methods

Standard methods were used for all the chemical analyses (APHA et al., 2005). A pH meter (AB15 Plus, Fisher Scientific) was used to measure the pH of wastewater and ferric solution. COD concentration was measured by following a closed reflux, colorimetric method (Standard Method 5220 D) using a spectrophotometer (HACH, DR 2800). Soluble sulfate concentration was measured by a turbidimetric method (USEPA method 375.4) using a spectrophotometer (Thermo Scientific, GENESYS 10UV). Total iron concentration was measured with an atomic absorption spectrophotometer (Perkin Elmer 3100) after the samples were acidified with a 70% nitric acid (HNO₃) solution. Ferrous iron concentration was determined using 1, 10 phenanthroline method (Standard Method

3500 B) and ferric iron concentration was calculated by the differences between the total and ferrous iron concentrations. Sulfide concentrations were measured by a methylene blue method (Standard Method 4500 D) using a HACH spectrophotometer (DR 2800). An optical emission spectrometer (Optima 2100 DV) was used to estimate the total sulfur in the sludge sample after acidified with a 70% HNO₃ solution.

4.2.4 Sludge Characterization

After running the experiment under each Fe/S ratio, the accumulated sludge sample was collected from the bioreactor for physical, chemical and biological characterization. Total suspended solid (TSS), volatile suspended solid (VSS), and non-volatile suspended solid (NVSS) in the sludge samples were measured (Standard Method 2540). A scanning electron microscope (SEM) equipped with an energy dispersion spectroscopy (SEM-EDS, Hitachi S 4700) was used to study the morphological and elemental composition of the sludge. The sludge samples were dried in a closed desiccator filled with calcium sulfate and flushed with N₂ gas to prevent sludge oxidation. After grinding the dried sludge, the powdered samples were mounted on aluminum stubs and coated with Au-Pd by using a sputter (Denton Desk V) to avoid surface charging. The SEM scanning was performed under an accelerating voltage of 10-20 kV and qualitative elemental analysis of the sludge samples was conducted by EDS spectrometry under an accelerating voltage of 10 kV. Biological fixation of the sludge sample was done to take SEM images of the microorganisms ([Supplementary material](#)).

The powdered sludge samples were also used to determine the chemical states of Fe and S by X-ray photoelectron spectroscopy (XPS) analysis (PHI 5000 Versaprobe). The sample powder was mounted on a sample holder with a zero reflective quartz plate (MTI corporation, CA) located underneath. XPS spectra were obtained with a monochromatized Al K α X-ray source (1487 eV) while base pressure of the analytical chamber was on the order of 10⁻⁷ Pa. A pass energy of 23.5 eV was used to conduct elemental scans of Fe and S. X-ray diffraction (XRD) analysis was conducted to determine the crystallinity of the sludge samples using an X-ray diffractometer (PANalytical X'Pert Pro). The samples were prepared by separating the sludge particles

from the solution using high-speed centrifugation (5000 x *g*) for 10 minutes. The XRD analysis was performed with a Cu K α X-ray source operated under 45 kV and 40 mA.

4.2.5 Microbiological Analysis

A DNeasy Powersoil DNA extraction kit (Qiagen, Maryland, USA) was used to extract the DNA from sludge samples and a Qubit fluorometer (Invitrogen, Carlsbad, CA, USA) was used for quantification. The bacterial/archaeal primer set 515F/806R that targets the V4 region of the gene was used to sequence the 16S rRNA genes in the extracted DNA at Argonne National Laboratory. Resulting reads were checked for chimeras (DADA2) and subsequently clustered into exact sequence variant (ESV) classifications at 100% similarities using the DADA2 tool in the QIIME2 pipeline (Qiime2-2018.4) and SILVA 16S rRNA gene database. Sequences used in this study have been deposited in the NCBI Sequence Read Archive under accession number PRJNA528092.

4.3. Results and Discussion

4.3.1 Treatment performance

4.3.1.1 Acclimation to new Fe/S ratios

Acclimation of the bioreactor to a new Fe/S ratio was evaluated by monitoring COD concentration periodically to determine when a steady state could be reached. COD concentration was observed to stabilize after approximately 23 days ([Supplementary material, Fig. S6](#)). A one-month acclimation period was therefore given before regular sampling of the influent and effluent to evaluate the treatment performance under each Fe/S ratio.

4.3.1.2 COD removal, sulfate reduction and iron retention

The bioreactor was found to provide fairly consistent treatment for COD removal during the study ([Supplementary material, Fig. S7](#)). The average COD removal efficiencies were $84 \pm 4 \%$, $86 \pm 4 \%$ and $89 \pm 2 \%$ for Fe/S ratios of 0.5, 1 and 2, respectively (Fig. 1). The COD removal increased slightly with Fe/S ratio showing the effect of increasing Fe(III) dosing on the organics oxidation. Sulfate reduction and iron retention also followed an increasing trend with the Fe/S ratio (Fig. 13a). Higher than 90% sulfate reduction and

close to 100% iron retention under all the Fe/S ratios indicated that both ferric iron and sulfate reduction played a significant role in COD oxidation.

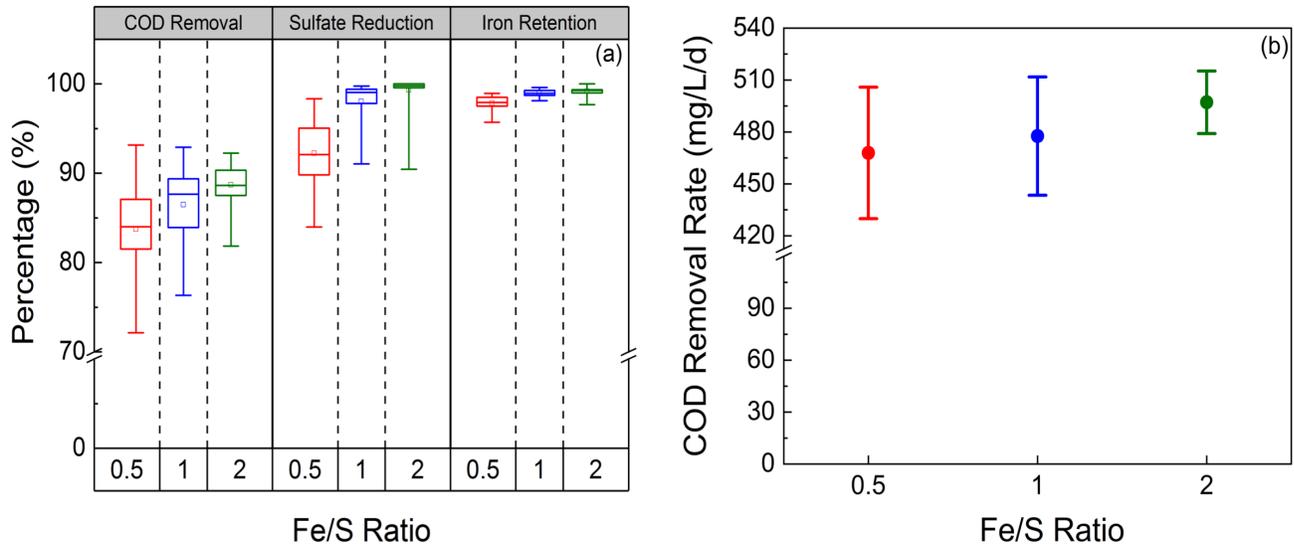


Fig. 13: (a) COD removal, sulfate reduction and iron retention and (b) COD removal rates under different Fe/S molar ratios

4.3.1.3 COD Removal Rate

The COD removal rates were calculated as 468 ± 25 mg/L/d, 478 ± 23 mg/L/d and 497 ± 12 mg/L/d for Fe/S ratio 0.5, 1 and 2 respectively (Fig. 13b). The increasing removal rate was attributed to evolving microbial community in the bioreactor that facilitated faster organics oxidation resulting from the increased ferric loadings. The result shows how Fe/S ratio can be used to regulate organic oxidation rates in the Fe(III)-dosed treatment system.

4.3.1.4 Effluent Quality

Mean total iron concentrations of the 200 effluent samples were observed as 1.6 ± 0.6 , 1.6 ± 0.5 and 2.2 ± 0.8 mg/L under Fe/S ratio 0.5, 1 and 2, respectively. Soluble ferrous iron was consistently low under all Fe/S ratios with mean concentrations ranging from 0.02-0.05 mg/L. Sulfide concentrations were higher under Fe/S ratio 0.5 (0.42 ± 0.3 mg/L) and decreased to 0.04 ± 0.1 and 0.03 ± 0.02 mg/L as the ratio increased to 1 and 2, respectively (Fig. 14a). These results indicated that under Fe/S ratio 0.5, ferrous iron

produced from ferric reduction was insufficient to precipitate out all the biogenic sulfide. Under the two higher Fe/S ratios (1 and 2), sufficient amounts of ferrous iron were produced to precipitate out soluble sulfide. The residual iron represented a small fraction (2%, 1%, and 0.8% for Fe/S ratios 0.5, 1 and 2 respectively) of the iron dose, and mostly was in the form of ferric iron. The residual ferrous iron and sulfide in the effluent represent an oxygen demand and can be oxidized by chlorine in a disinfection unit (Cadena and Peters, 1988; Khadse et al., 2015). The resultant ferric iron is expected to form a hydroxide solid which can be removed by sedimentation or filtration before discharge of the effluent to the environment.

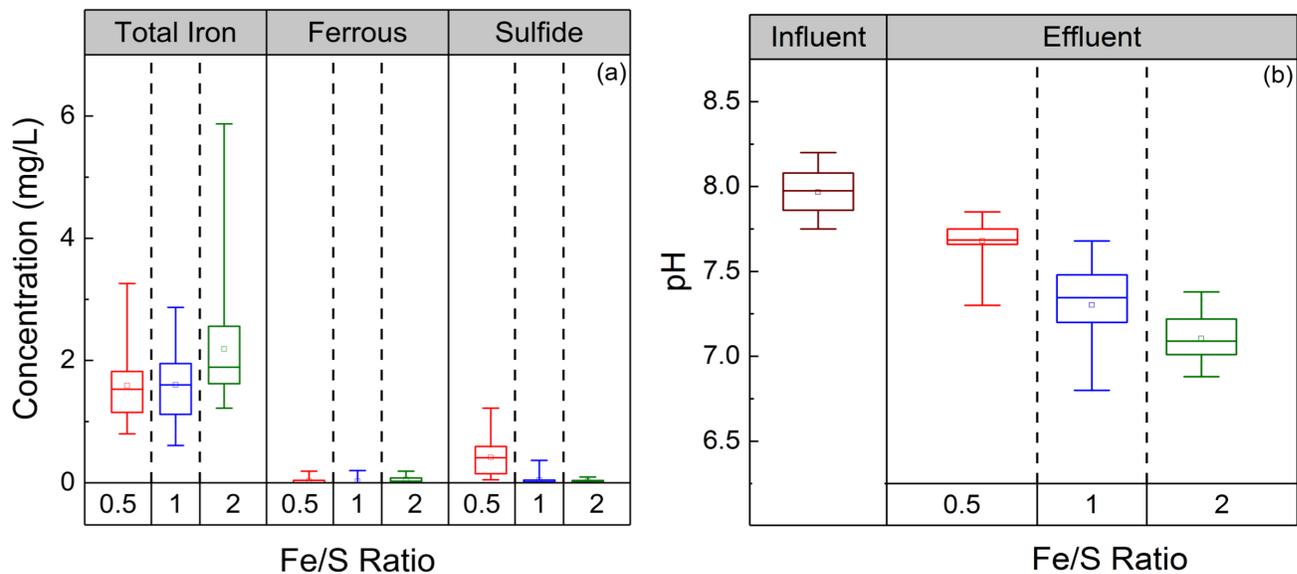


Fig. 14: (a) Effluent total iron, ferrous and sulfide concentration and (b) Influent and effluent pHs at different Fe/S molar ratios

The influent pH (8.0 ± 0.1) was consistently lowered to those in the effluent as a result of the biological treatment (Fig. 14b), which was attributed partly to addition of the ferric iron solution (pH 4-4.2) to the bioreactor. The pH trend also suggests a positive correlation between net acidity production and the Fe/S ratio. This is supported by previous studies that showed production of acidity from organic carbon oxidation was higher than alkalinity generation by ferric iron reduction (Deng et al., 2016; Lovley and Lonergan, 1990; Lovley and Phillips, 1988).

4.3.1.5 Mass balance of Fe and S in the bioreactor

A mass balance was performed on Fe and S under Fe/S ratio 2 to understand the chemical flows throughout the biological treatment. In the influent, sulfur (S) was in the dissolved form (38.2 mg/d) and iron (Fe, 134.3 mg/d) was in suspended flocs (pH 4 – 4.2). During this operation, total 50 samples used for mass balance calculations showed that only 1.3% of the total S load and 2% of the total Fe load were discharged through the effluent. On the other hand, 69% of the total S load and 48% of the total Fe load was accounted as particulates in the sludge sample (Fig. 15). This high Fe and S content of sludge samples were due to ferrous sulfide precipitation. The remaining chemical masses were calculated as unaccounted fractions that can be recognized by several possible mechanisms, including iron sulfide precipitation retained in the bioreactor, evaporative loss of sulfide or some loss during sampling and sample preparation for chemical analyses (Deng and Lin, 2017). This mass balance of Fe and S revealed that, most of the fed Fe and S formed chemical precipitates as a result of the biogeochemical transformations occurring in the bioreactor and only a small fraction of the elements was discharged in the effluent.

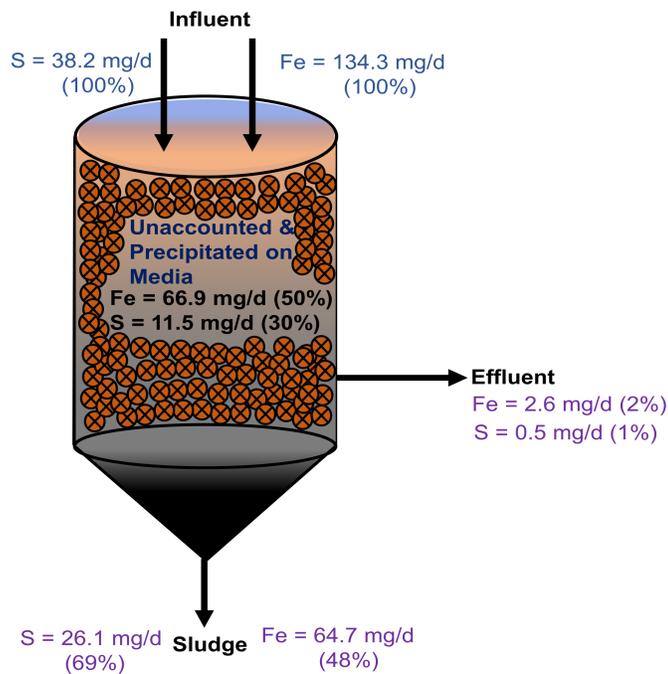


Fig. 15: Mass flow rates of Fe and S throughout the biological treatment under Fe/S molar ratio 2

4.3.2 Sludge Characterization

4.3.2.1 Solid Concentrations

Mean TSS concentration of the sludge samples ranged from 1,341 – 21,946 mg/L. The mean TSS concentration increased with the Fe/S ratio, indicating an increase in the sludge production under the higher Fe/S ratios compared to Fe/S ratio 0.5. The solid analyses showed that the mean VSS/TSS decreased from 40% to 20% and correspondingly the mean NVSS/TSS increased from 60% to 80% as Fe/S ratio increased from 0.5 to 2 (Fig. 16). This revealed that most of the solids were inorganic materials and the inorganic fraction of the solids increased with the Fe/S ratio. These results showed the range of volatile fractions of the sludge samples (0.2 – 0.4) and their corresponding nonvolatile fractions (0.8 – 0.6) resulting from this treatment and their fluctuations under different dosing scenarios. The increasing inorganic fraction of the sludge solid with the Fe/S ratio indicated more iron sulfide production with increasing iron dosing.

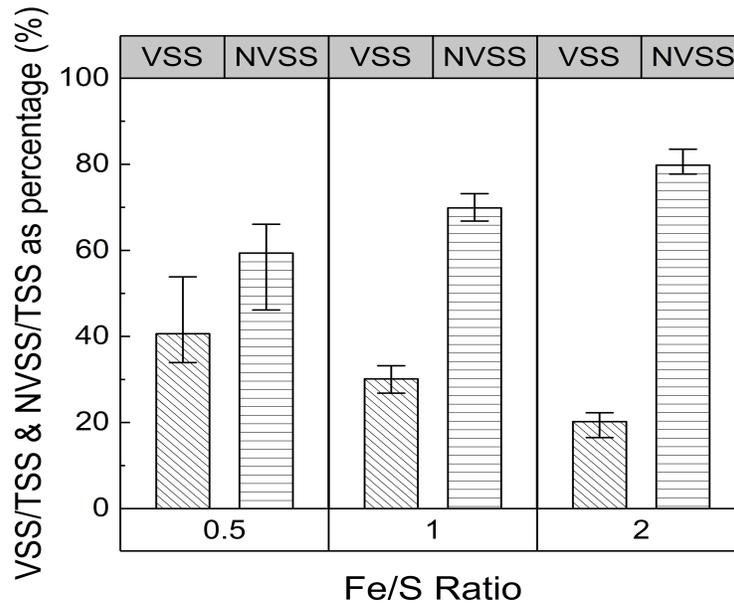


Fig. 16: VSS/TSS and NVSS/TSS as percentage (%) of sludge samples under different Fe/S ratios

4.3.2.2 Sludge Morphology

SEM-EDS analysis in this study revealed major elements of the sludge sample as carbon (C), oxygen (O), Fe, and S (Supplementary material, Fig. 17a and b). The small grains observed in the SEM image (Fig. 17a) were recognized as the precipitate particles which had a very disordered morphology with no specific pattern. The small sizes ($\approx 1\text{-}2\ \mu\text{m}$) of the sludge particles as spherical aggregates were in line with the structural descriptions of iron sulfide presented in previous studies (Bratkova et al., 2018; Csákberényi-Malasics et al., 2012; Vaclavkova et al., 2014).

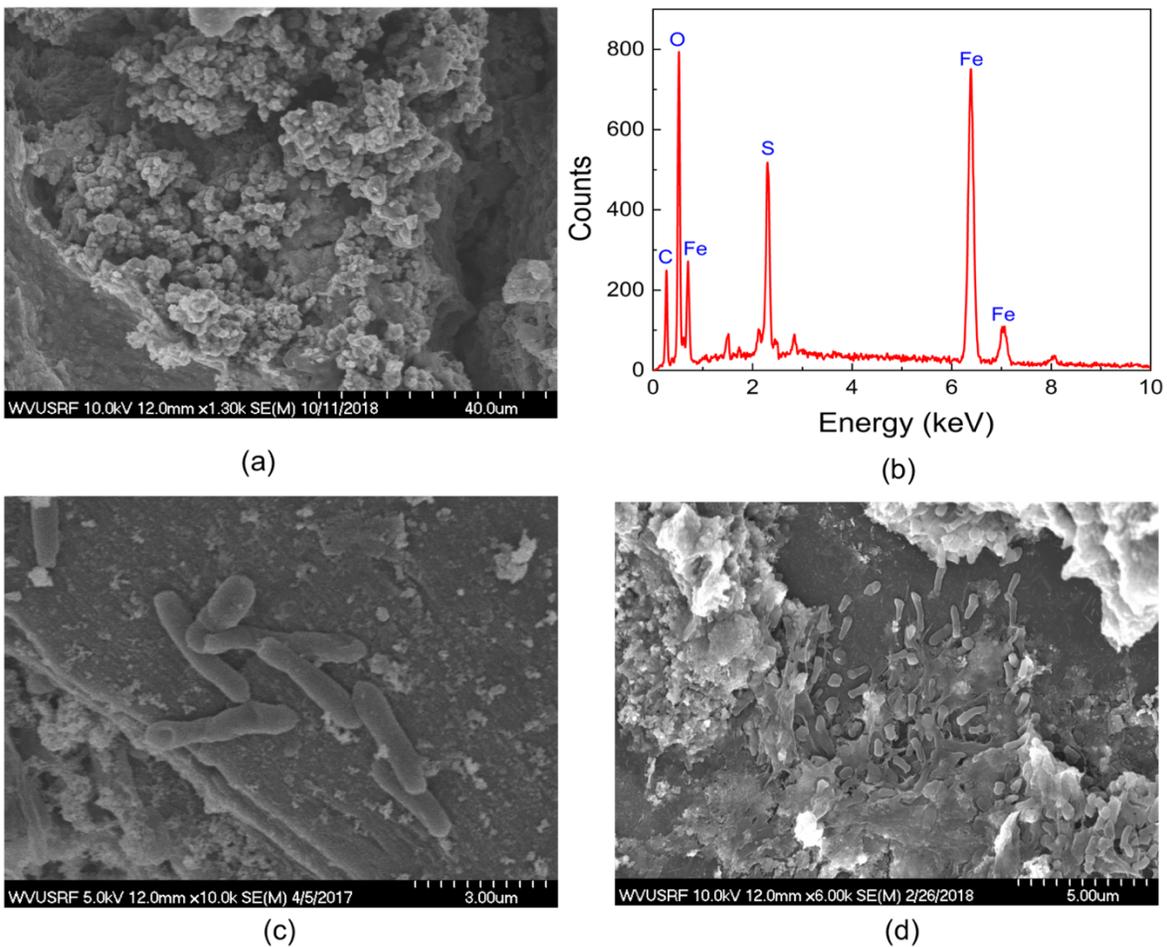


Fig. 17: (a) SEM micrograph and (b) EDS spectrum for sludge sample under Fe/S molar ratio 2; (c) & (d): SEM images of cells present in anaerobic sludge

SEM images taken after the biological fixation of sludge samples revealed the presence of various microbial cells in the sludge. Majority of the observed microorganisms were rod

shaped and curved, and had an approximate length of 2-3 μm (Supplementary Material, Fig. 17c). The size and shape observed in these images were similar to those in the previous studies describing the physical characteristics of *Geobacter* sp. (Caccavo Jr et al., 1994; Nevin et al., 2005) and *Desulfovibrio* sp. (Zellner et al., 1989). Cells were observed to be in high density (Fig. 17d).

4.3.2.3 Sludge Composition

XPS analysis showed that two different iron sulfide forms, FeS and FeS₂, were the major inorganic constituents of the sludge samples. The narrow region in the spectrum of Fe2p revealed a major peak at 707.4 eV (Fig. 18a), which represents both FeS and FeS₂. As the binding energies of FeS and FeS₂ are very similar, it is often difficult to separate the peaks of these two different forms in an Fe2p spectrum (Descostes et al., 2000; Han et al., 2013). However, the S2p spectrum was fit with two distinct peaks at 160.9 eV and 162.3 eV, in agreement with the typical binding energies of FeS and FeS₂ respectively (Fig. 18b) (Lennie and Vaughan, 1996; Naumkin et al., 2012).

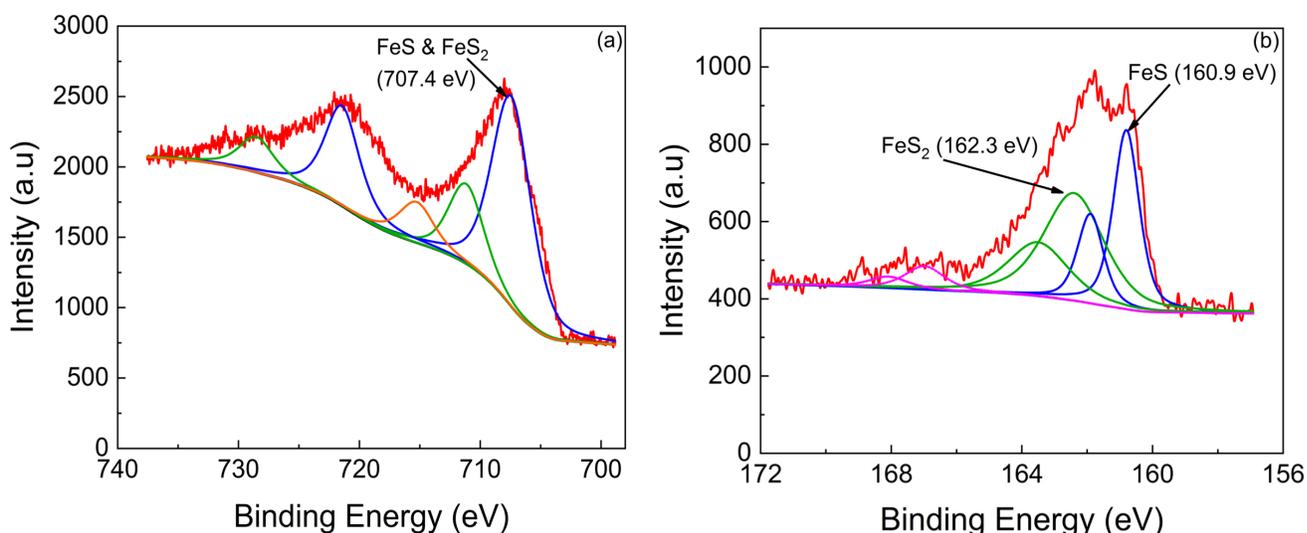


Fig. 18: XPS spectrum of (a) Fe2p and (b) S2p region of a sludge sample

4.3.2.4 Sludge Crystallinity

An XRD spectrum of the sludge samples showed sharp peaks representing crystalline forms of FeS (mackinawite), FeS₂ (pyrite), Fe₃S₄ (greigite), FeOOH (goethite), and Fe₃O₄ (magnetite) (Supplementary Material, Fig. S8). However, the low intensity of the sharp

peaks and dominant presence of broad hump peaks suggest that most of the sludge samples were in an amorphous form. Biogenic sludge generally precipitates as amorphous FeS, which may be transformed to stable crystalline form of FeS and FeS₂ under a long-term operation. Greigite (Fe₃S₄) was considered an intermediate iron sulfide form between the transformation of amorphous FeS to well crystallized FeS₂ in anaerobic environments (Csákberényi-Malasics et al., 2012; Gramp et al., 2010). Therefore, the sludge precipitates can be described as a mixture of both amorphous and crystalline iron sulfides with amorphous phases being prevailing. Crystallographic information of the obtained peaks revealed tetragonal crystal structure of FeS and cubic structure of FeS₂ and Fe₃S₄.

Sharp peaks of FeOOH represent crystalline ferric oxyhydroxides that occurred due to the surface oxidation of ferrous sulfide. The presence of Fe₃O₄ (magnetite) can be attributed to biologically-induced mineralization (BIM) in which microbes sorb solutes onto their cell surface or extrude organic polymers resulting in mineral formation (Bazylinski et al., 2007). *Geobacter* sp. has been reported to produce magnetite extracellularly through BIM during FeOOH reduction (Moskowitz et al., 1989). Magnetite can be a very useful element because of its physical properties and various heat treatments have been investigated for generating magnetite from iron sulfide (Thorpe et al., 1984b; Waters et al., 2008). These results suggest the sludge produced in this Fe(III)-dosed biological treatment can potentially be further processed for beneficial applications.

4.3.3 Microbiological Composition

Dominant ESVs identified in the sludge samples belonged to ten different phyla, including members of the Alphaproteobacteria, Deltaproteobacteria, Gammaproteobacteria, Acidobacteria, Bacteroidetes, Chloroflexi, Firmicutes, Spirochaetes, Actinobacteria and Euryarchaeota. The average relative abundances of these phyla are listed in Table 4.

Table 4: Major bacteria and their phylum identified in the bioreactor

Deltaproteobacteria	Acidobacteria	Bacteroidetes	Gammaproteobacteria	Actinobacteria
<i>Geobacter sp.</i> (22%)	<i>Geothrix sp.</i> (0.5%)	<i>Ignavibacteria sp.</i> (4%)	<i>Methylomonas sp.</i> (0.2%)	<i>Cellulomonas sp.</i> (0.1%)
<i>Desulfobulbus sp.</i> (4%)		<i>Paludibacter sp.</i> (2%)	<i>Azospira sp.</i> (0.04%)	
<i>Desulfovibrio sp.</i> (5%)			<i>Azobacter sp.</i> (0.1%)	
<i>Desulfatirhabdium sp.</i> (3%)				
<i>Desulforhabdus sp.</i> (1%)				
<i>Desulfomonile sp.</i> (0.2%)				
<i>Syntrophobacter sp.</i> (2 %)				
<i>Smithella sp.</i> (0.2%)				
Firmicutes	Chloroflexi	Alphaproteobacteria	Spirochaetes	Euryarchaeota
<i>Clostridium sp.</i> (4%)	<i>Anaerolineae sp.</i> (4%)	<i>Pleomorphomonas sp.</i> (7%)	<i>Treponema sp.</i> (6%)	<i>Methanosaeta sp.</i> (2%)
<i>Ruminiclostridium 1</i> (0.04%)			<i>Brevinema sp.</i> (2%)	
<i>Veillonellaceae sp.</i> (1%)				

Although caution must be taken when assigning functional roles to identified taxa from 16S rRNA gene data, some inferred conserved functions may be assigned based on laboratory studies of model microorganisms within specific taxonomic groups. *Geobacter sp.*, *Geothrix sp.* and *Ignavibacteria sp.* were classified as IRB, and *Desulfovibrio sp.*, *Desulfobulbus sp.*, *Desulfatirhabdium sp.*, *Desulforhabdus sp.* and *Desulfomonile sp.* were classified as SRB based on this concept. The ability of *Geobacter sp.* and *Geothrix*

sp. to perform ferric reduction has been observed in laboratory studies (Childers et al., 2002; Coates et al., 2001; Nevin et al., 2005; Nevin and Lovley, 2002). *Ignavibacteria* sp. was recently identified to facilitate iron reduction as well (Fortney et al., 2018; Podosokorskaya et al., 2013). With 83% relative abundance of the total identified IRB, *Geobacter* sp. was inferred to perform the majority of iron reduction in the bioreactor (Fig. 19a). The dominance of *Geobacter* sp. can be linked with presence of acetate as one of the major organic compounds in synthetic wastewater. *Geobacter* sp. typically oxidize small organic substrates such as acetate to CO₂, coupled to iron reduction (Esther et al., 2015; Lovley, 1993; Lovley et al., 1993). *Ignavibacteria* sp. and *Geothrix* sp. comprised 15% and 2% relative abundance of the total IRB respectively, and likely contributed to the ferric reduction in the bioreactor (Fig. 19a). Similar to *Geobacter*, *Ignavibacteria* has been observed to grow well in acetate-amended incubations (Fortney et al., 2018). Due to the insolubility of ferric compounds at the pH range 7-8 in the bioreactor, these IRBs perform the ferric reduction either by direct contact with outer-membrane cytochromes or via conductive pili structures. In addition, *Geothrix* is able to produce chelators that solubilize Fe(III) and release electron-shuttling compounds for the microbial interaction of Fe(III) surface with cell structure (Nevin and Lovley, 2002).

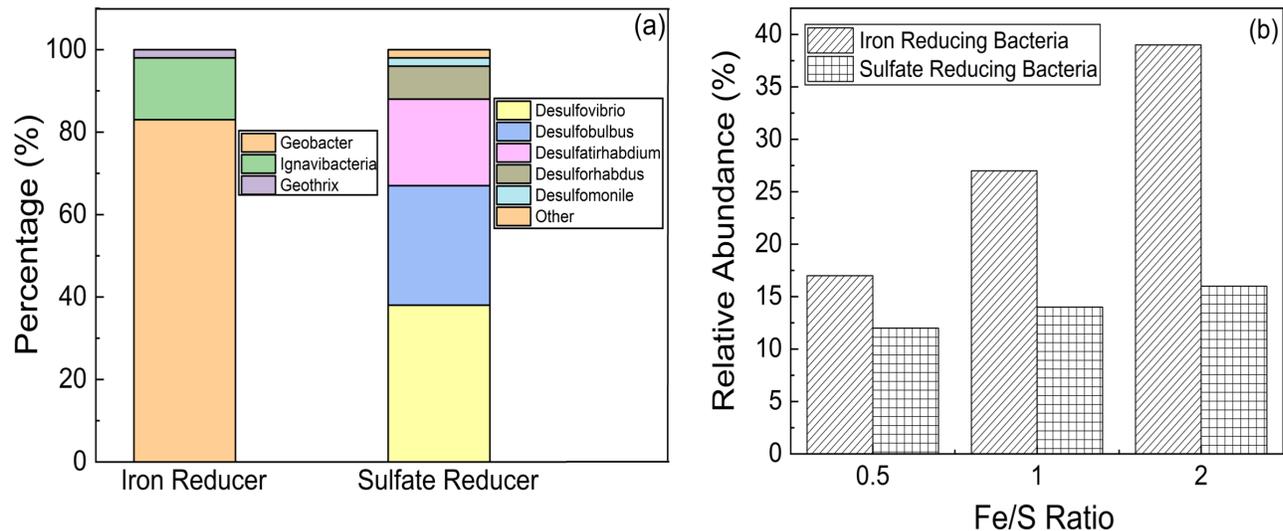


Fig. 19: (a) Microbial distribution and (b) relative abundances of putative IRB and SRB

Desulfovibrio sp., *Desulfobulbus* sp., *Desulfatirhabdium* sp., *Desulforhabdus* sp., *Desulfomonile* sp. were the major putative SRB observed in the bioreactor and comprised 38%, 30%, 21%, 8% and 2% relative abundance of the total SRB respectively (Fig. 19a). All these gram-negative SRB belong to the Deltaproteobacteria phyla, and previously have been shown to use sulfate as electron acceptor, which is subsequently reduced to sulfide (Balk et al., 2008; DeWeerd et al., 1990; El Houari et al., 2017; Elferink et al., 1995; Rabus et al., 2015). *Desulfovibrio* sp. and *Desulfobulbus* sp. can catalyze incomplete oxidation of larger organic substrates such as lactate, yielding acetate which can subsequently be used by other IRB and SRB (i.e. *Desulfatirhabdium* sp. and *Desulforhabdus* sp.) as an electron donor (El Houari et al., 2017; Rabus et al., 2015). Characterized representatives of the putative SRB identified here are mesophilic, growing well under moderate temperatures (20 - 45°C) and circumneutral pH.

In terms of relative abundances, IRB were observed to be the dominant functional group over SRB under all the Fe/S ratios. In general, IRB have a competitive edge over SRB for organic substrate utilization due to the thermodynamically favorable redox reactions with ferric iron. The relative abundance of putative IRB was 17%, 27% and 39% for Fe/S 0.5, 1 and 2 respectively (Fig. 19b), indicating a positive correlation of IRB with COD removal as organic oxidation was also improved with increasing Fe/S ratio. Although sulfate concentrations decreased slightly with the increasing Fe/S ratio, the relative abundances of putative SRB increased from 12% to 16%. A major reason may be the presence of both *Desulfovibrio* sp. and *Desulfobulbus* sp. as the dominant SRB in the bioreactor. Both of the taxonomic groups have been observed to grow well under iron reducing conditions, and perform enzymatic Fe(III) reduction (Coleman et al., 1993; Holmes et al., 2004; Tebo and Obraztsova, 1998). Therefore, the possibility exists that these SRB were also performing ferric reduction under higher Fe/S ratios, and concurrently increasing in relative abundance.

In addition to IRB and SRB, many other bacteria were also detected in the bioreactor. Comparatively low abundance of methanogens, *Methanosaeta* sp. (2%) in the bioreactor suggests that IRB and SRB outcompeted methanogens for substrate utilization. Bacteria previously characterized as fermenters, such as *Clostridium* sp., *Treponema* sp., and

Paludibacter sp. were present with average relative abundance of 4%, 6% and 2% respectively. These fermentative bacteria can break the complex organic compounds into small organic substrates that can be easily utilized by IRB and SRB (Chen et al., 2005; Dröge et al., 2008; Ueki et al., 2006) . Some strains of *Clostridium* have previously been found to indirectly participate in dissimilatory Fe(III) reduction by disposing of reducing equivalents (Dobbin et al., 1999; Shah et al., 2014). *Anaerolineae* ESVs were also identified within the bioreactor; members of this group are generally found in the anaerobic sludge granules of bioreactors (Yamada et al., 2005).

Shannon's diversity index (H) was calculated as 3.26, 3.36 and 3.34 for Fe/S ratio 0.5, 1 and 2 respectively and Simpson's diversity index was measured as 0.88, 0.92 and 0.92 for the same ratio values. These high values of diversity indices (higher than 2 for Shannon's H and close to 1 for Simpson's index) reflect the diverse microbial community in the bioreactor (Grabchak et al., 2017; Ifo et al., 2016). Higher microbial diversity of the bioreactor at higher ratios may also support increase in functionality and tolerance level against environmental and chemical disturbances (e.g. temperature, electron donors) (Hiibel et al., 2011; Koschorreck et al., 2010).

4.4 Conclusion

The long-term operation of a continuous Fe(III)-dosed anaerobic biological treatment was investigated in this study to understand the Fe and S biogeochemical transformations in the context of wastewater treatment. The Fe/S ratio was found to play a significant role in regulating major treatment aspects of the biological system including organics removal rate and efficiency, effluent quality, sludge production and microbial composition. For practical implementation of this treatment method, the iron dosing requirement is primarily to be determined by the organics and sulfate loads of the wastewater to provide sufficient electron acceptors for organics oxidation. The results showed increasing Fe(III) dosing resulted in enhanced organics removal rates and efficiency . Another beneficial outcome of higher iron dosing is that excessive quantity of ferrous iron can lower the sulfide level in the effluent through forming iron sulfide precipitates. This precipitation mechanism is significant because it eliminates toxicity and oxygen demand possibly caused by sulfide in the receiving water. The biogenic ferrous iron and sulfide primarily precipitated as

amorphous FeS, which may undergo long-term transformations to crystalline FeS and FeS₂. These sludge byproducts can directly be used for environmental remediation and wastewater treatment or for recovering valuable materials (e.g magnetite). The microbiological analyses indicated the presence of putative IRB and SRB along with fermentative bacteria in the bioreactor. Their known functional activities suggest synergistic relationships among these bacterial species in organics degradation.

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4.5 Supplementary Materials

Biological fixation of sludge samples for SEM photographs

The 6 ml sludge sample was taken on a Microscope cover glass (Fisherbrand) and processes several steps of biological fixation to preserve the integrity of bacterial cell walls. The sample was washed with 2.5% Glutaraldehyde for one hour and rinsed three times with phosphate buffer saline (PBS) at 15-minute intervals. Then the sample was dehydrated with a series of ethanol solution at different concentrations (30%, 50%, 70%, 90%, 100%). This dehydration procedure was executed with gentle agitation for 15 minutes in each step. Then the ethanol was removed from the sample and dried with hexamethyldisilazane (HMDS) for another 15 minutes. This technique was applied to eliminate surface tension effects by raising the temperature of the sample above the critical temperature for CO₂ and reducing the distortion of morphology and surface structure.

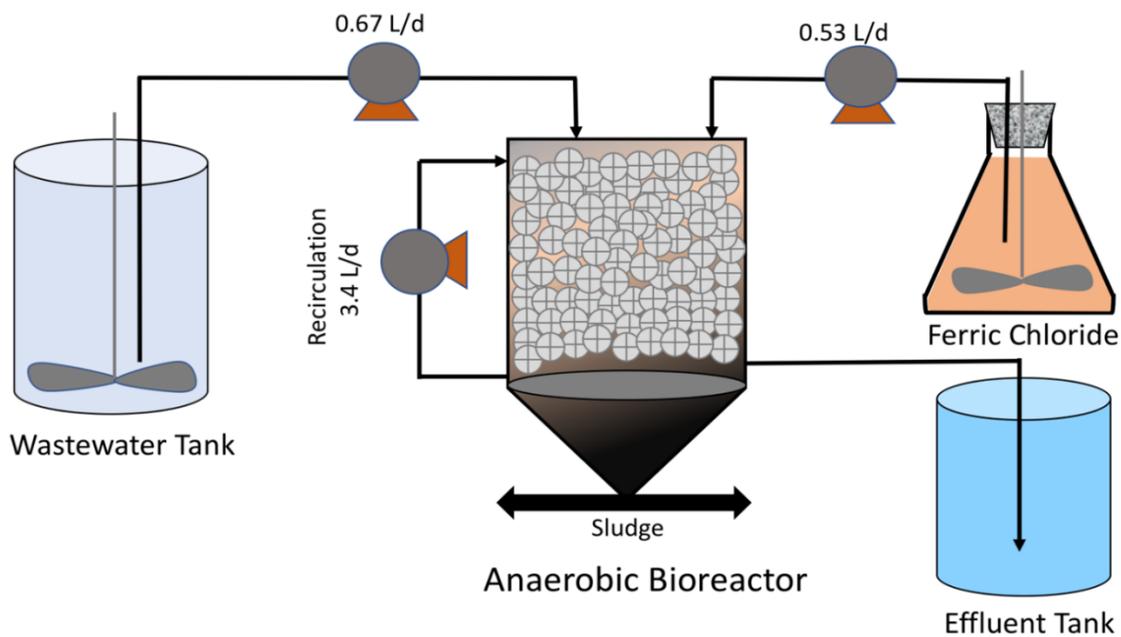


Fig. S5: Experimental setup of the Fe(III)-dosed treatment system

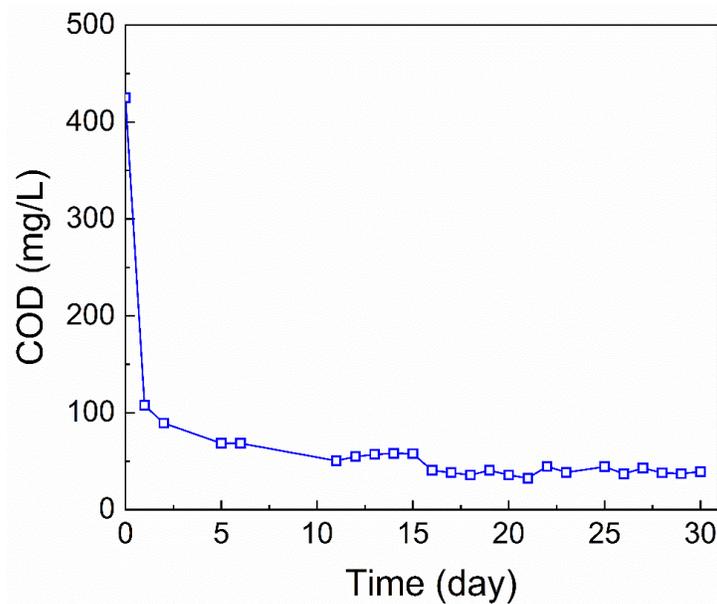


Fig. S6: COD concentration of the effluent under Fe/S ratio 2 for microbial acclimation evaluation

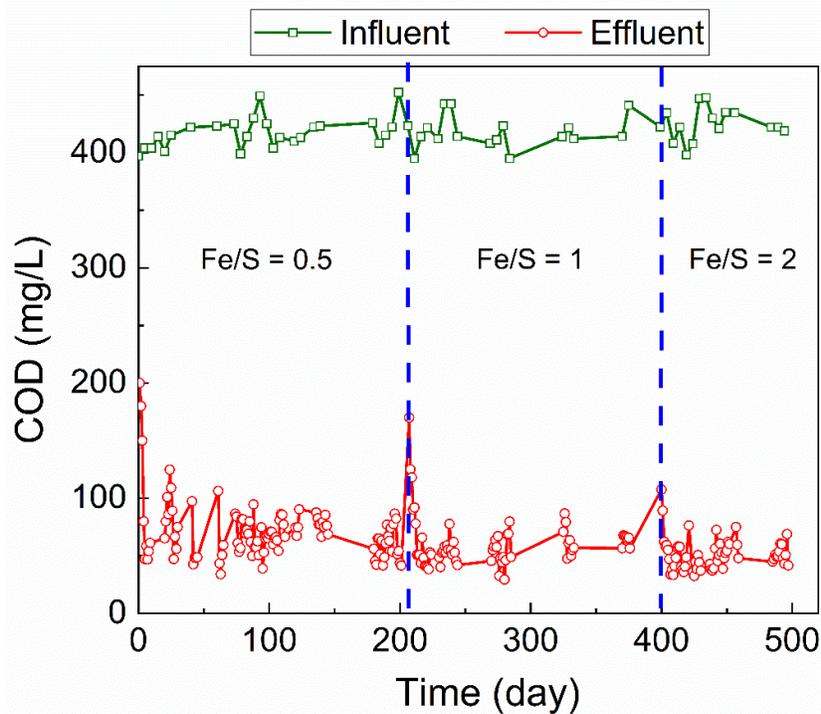


Fig. S7: COD concentration of effluent from the Fe(III)-dosed biological treatment

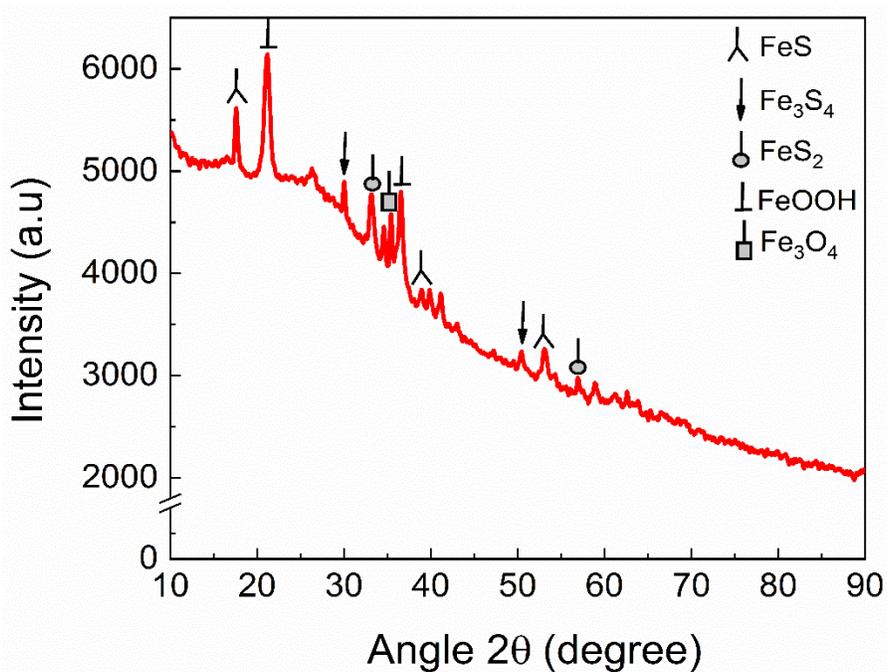


Fig. S8: XRD spectrum of the anaerobic sludge sample (Fe/S = 2)

References

- Ahmed, M., Lin, L.-S., 2017. Ferric reduction in organic matter oxidation and its applicability for anaerobic wastewater treatment: a review and future aspects. *Rev. Environ. Sci. Biotechnol.* 16, 273–287. <https://doi.org/10.1007/s11157-017-9424-3>
- Ahmed, M., Lin, O., Saup, C.M., Wilkins, M.J., Lin, L.S., 2019. Effects of Fe/S ratio on the kinetics and microbial ecology of an Fe(III)-dosed anaerobic wastewater treatment system. *J. Hazard. Mater.* 369, 593–600. <https://doi.org/10.1016/j.jhazmat.2019.02.062>
- APHA, AWWA, WEF, 2005. Standard methods for the examination of water and wastewater, 21st ed. American Public Health Association, American Water Works Association, and Water Environment Federation.
- Azam, H.M., Finneran, K.T., 2013. Ferric iron amendment increases Fe(III)-reducing microbial diversity and carbon oxidation in on-site wastewater systems. *Chemosphere* 90, 1435–1443. <https://doi.org/10.1016/j.chemosphere.2012.09.002>
- Balk, M., Altınbaş, M., Rijpstra, W.I.C., Damsté, J.S.S., Stams, A.J.M., 2008. *Desulfatirhabdium butyratorans* gen. nov., sp. nov., a butyrate-oxidizing, sulfate-reducing bacterium isolated from an anaerobic bioreactor. *Int. J. Syst. Evol. Microbiol.* 58, 110–115. <https://doi.org/10.1099/ijs.0.65396-0>
- Bazylinski, D.A., Frankel, R.B., Konhauser, K.O., 2007. Modes of biomineralization of magnetite by microbes. *Geomicrobiol. J.* 24, 465–475. <https://doi.org/10.1080/01490450701572259>
- Bratkova, S., Lavrova, S., Angelov, A., Nikolova, K., Ivanov, R., Koumanova, B., 2018. Treatment of wastewaters containing Fe, Cu, Zn and As by microbial hydrogen sulfide and subsequent removal of COD, N and P. *J. Chem. Technol. Metall.* 53, 245–257.
- Caccavo Jr, F., Lonergan, D.J., Lovley, D.R., Davis, M., Stolz, J.F., McInerney, M.J., 1994. *Geobacter sulfurreducens* sp. nov., a Hydrogen- and Acetate- Oxidizing Dissimilatory Metal-Reducing Microorganism. *Appl. Environ. Microbiol.* 60, 3752–3759. <https://doi.org/10.1111/j.1758-2229.2009.00055.x>
- Cadena, F., Peters, R.W., 1988. Evaluation of Chemical Oxidizers for Hydrogen Sulfide Control. *Water Pollut. Control Fed.* 60, 1259–1263. <https://doi.org/10.2307/25043633>
- Chan, Y.J., Chong, M.F., Law, C.L., Hassell, D.G., 2009. A review on anaerobic-aerobic treatment of industrial and municipal wastewater. *Chem. Eng. J.* 155, 1–18. <https://doi.org/10.1016/j.cej.2009.06.041>
- Chen, W.M., Tseng, Z.J., Lee, K.S., Chang, J.S., 2005. Fermentative hydrogen production with *Clostridium butyricum* CGS5 isolated from anaerobic sewage sludge. *Int. J. Hydrogen Energy* 30, 1063–1070. <https://doi.org/10.1016/j.ijhydene.2004.09.008>
- Childers, S.E., Ciuffo, S., Lovley, D.R., 2002. *Geobacter metallireducens* accesses insoluble Fe(III) oxide by chemotaxis. *Nature* 416, 767–769.

<https://doi.org/10.1038/416767a>

- Coates, J.D., Bhupathiraju, V.K., Achenbach, L.A., Mcinerney, M.J., Lovley, D.R., 2001. *Geobacter hydrogenophilus*, *Geobacter chapellei* and *Geobacter grbciaae*, three new, strictly anaerobic, dissimilatory Fe(III)-reducers. *Int. J. Syst. Evol. Microbiol.* 51, 581–588.
- Coleman, M.L., Hedrick, D.B., Lovley, D.R., White, D.C., Pye, K., 1993. Reduction of Fe(III) in sediments by sulphate-reducing bacteria. *Nature* 361, 436–438. <https://doi.org/10.1038/361436a0>
- Csákberényi-Malasics, D., Rodriguez-Blanco, J.D., Kis, V.K., Rečnik, A., Benning, L.G., Pósfai, M., 2012. Structural properties and transformations of precipitated FeS. *Chem. Geol.* 294–295, 249–258. <https://doi.org/10.1016/j.chemgeo.2011.12.009>
- Damianovic, M.H.R.Z., Foresti, E., 2007. Anaerobic degradation of synthetic wastewaters at different levels of sulfate and COD/sulfate ratios in horizontal-flow anaerobic reactors (HAIB). *Environ. Eng. Sci.* 24, 383–393.
- Deng, D., Lin, L.S., 2017. Continuous sulfidogenic wastewater treatment with iron sulfide sludge oxidation and recycle. *Water Res.* 114, 210–217. <https://doi.org/10.1016/j.watres.2017.02.048>
- Deng, D., Weidhaas, J.L., Lin, L.-S., 2016. Kinetics and microbial ecology of batch sulfidogenic bioreactors for co-treatment of municipal wastewater and acid mine drainage. *J. Hazard. Mater.* 305, 200–8. <https://doi.org/10.1016/j.jhazmat.2015.11.041>
- Descostes, M., Mercier, F., Thromat, N., Beaucaire, C., Gautier-Soyer, M., 2000. Use of XPS in the determination of chemical environment and oxidation state of iron and sulfur samples: Constitution of a data basis in binding energies for Fe and S reference compounds and applications to the evidence of surface species of an oxidized py. *Appl. Surf. Sci.* 165, 288–302. [https://doi.org/10.1016/S0169-4332\(00\)00443-8](https://doi.org/10.1016/S0169-4332(00)00443-8)
- DeWeerd, K.A., Mandelco, L., Tanner, R.S., Woese, C.R., Suflita, J.M., 1990. *Desulfomonile tiedjei* gen. nov. and sp. nov., a novel anaerobic, dehalogenating, sulfate-reducing bacterium. *Arch. Microbiol.* 154, 23–30. <https://doi.org/10.1103/PhysRev.85.816>
- Dobbin, P.S., Carter, J.P., Juan, C.G.S.S., Von Höbe, M., Powell, A.K., Richardson, D.J., 1999. Dissimilatory Fe(III) reduction by *Clostridium beijerinckii* isolated from freshwater sediment using Fe(III) maltol enrichment. *FEMS Microbiol. Lett.* 176, 131–138. [https://doi.org/10.1016/S0378-1097\(99\)00229-3](https://doi.org/10.1016/S0378-1097(99)00229-3)
- Dröge, S., Rachel, R., Radek, R., König, H., 2008. *Treponema isoptericolens* sp. nov., a novel spirochaete from the hindgut of the termite *Incisitermes tabogae*. *Int. J. Syst. Evol. Microbiol.* 58, 1079–1083. <https://doi.org/10.1099/ijs.0.64699-0>
- El Houari, A., Ranchou-Peyruse, M., Ranchou-Peyruse, A., Dakdaki, A., Guignard, M., Idouhamou, L., Bennisse, R., Bouterfass, R., Guyoneaud, R., Qatibi, A.I., 2017.

- Desulfobulbus oligotrophicus* sp. Nov., a sulfate-reducing and propionate-oxidizing bacterium isolated from a municipal anaerobic sewage sludge digester. *Int. J. Syst. Evol. Microbiol.* 67, 275–281. <https://doi.org/10.1099/ijsem.0.001615>
- Elferink, S.J.W.H.O., Maas, R.N., Harmsen, H.J.M., Stams, A.J.M., 1995. *Desulforhabdus amnigenus* gen. nov. sp. nov., a sulfate reducer isolated from anaerobic granular sludge. *Arch. Microbiol.* 164, 119–124.
- Esther, J., Sukla, L.B., Pradhan, N., Panda, S., 2015. Fe (III) reduction strategies of dissimilatory iron reducing bacteria. *Korean J. Chem. Eng.* 32, 1–14. <https://doi.org/10.1007/s11814-014-0286-x>
- Fortney, N.W., He, S., Kulkarni, A., Friedrich, M.W., Holz, C., Boyd, E.S., Roden, E.E., 2018. Stable isotope probing for microbial iron reduction in Chocolate Pots hot spring, Yellowstone National Park. *Appl. Environ. Microbiol.* 84. <https://doi.org/10.1128/AEM.02894-17>
- Gong, Y., Tang, J., Zhao, D., 2016. Application of iron sulfide particles for groundwater and soil remediation: A review. *Water Res.* 89, 309–320. <https://doi.org/10.1016/j.watres.2015.11.063>
- Grabchak, M., Marcon, E., Lang, G., Zhang, Z., 2017. The generalized Simpson's entropy is a measure of biodiversity. *PLoS One* 12, 1–11. <https://doi.org/10.1371/journal.pone.0173305>
- Gramp, J.P., Bigham, J.M., Jones, F.S., Tuovinen, O.H., 2010. Formation of Fe-sulfides in cultures of sulfate-reducing bacteria. *J. Hazard. Mater.* 175, 1062–1067. <https://doi.org/10.1016/j.jhazmat.2009.10.119>
- Han, D.S., Batchelor, B., Abdel-Wahaba, A., 2013. XPS Analysis of Sorption of Selenium(IV) and Selenium(VI) to Mackinawite (FeS). *Environ. Prog. Sustain. Energy* 32, 84–93. <https://doi.org/10.1002/ep>
- Hiibel, S.R., Pereyra, L.P., Breazeal, M.V.R., Reisman, D.J., Reardon, K.F., Pruden, A., 2011. Effect of Organic Substrate on the Microbial Community Structure in Pilot-Scale Sulfate-Reducing Biochemical Reactors Treating Mine Drainage. *Environ. Eng. Sci.* 28, 563–572. <https://doi.org/10.1089/ees.2010.0237>
- Holmes, D.E., Bond, D.R., Lovley, D.R., 2004. Electron Transfer by *Desulfobulbus propionicus* to Fe (III) and Graphite Electrodes. *Environ. Microbiol.* 70, 1234–1237. <https://doi.org/10.1128/AEM.70.2.1234>
- Hubert, C., Voordouw, G., 2007. Oil field souring control by nitrate-reducing *Sulfurospirillum* spp. that outcompete sulfate-reducing bacteria for organic electron donors. *Appl. Environ. Microbiol.* 73, 2644–2652. <https://doi.org/10.1128/AEM.02332-06>
- Ifo, S.A., Moutsambote, J.M., Koubouana, F., Yoka, J., Ndzai, S.F., Bouetou-Kadilamio, L.N.O., Mampouya, H., Jourdain, C., Bocko, Y., Mantota, A.B., Mbemba, M., Mouanga-Sokath, D., Odende, R., Mondzali, L.R., Wenina, Y.E.M., Ouissika, B.C.,

- Joel, L.J., 2016. Tree Species Diversity, Richness, and Similarity in Intact and Degraded Forest in the Tropical Rainforest of the Congo Basin: Case of the Forest of Likouala in the Republic of Congo. *Int. J. For. Res.* 2016. <https://doi.org/10.1155/2016/7593681>
- Khadse, G.K., Patni, P.M., Labhasetwar, P.K., 2015. Removal of Iron and Manganese from drinking water supply. *Sustain. Water Resour. Manag.* 1, 157–165. <https://doi.org/10.1021/ie50297a006>
- Koschorreck, M., Geller, W., Neu, T., Kleinstaub, S., Kunze, T., Trosiener, A., Wendt-Potthoff, K., 2010. Structure and function of the microbial community in an in situ reactor to treat an acidic mine pit lake. *FEMS Microbiol. Ecol.* 73, 385–395. <https://doi.org/10.1111/j.1574-6941.2010.00886.x>
- Kulandaivelu, J., Gao, J., Song, Y., Shrestha, S., Li, X., Li, J., Doederer, K., Keller, J., Yuan, Z., Mueller, J.F., Jiang, G., 2019. Removal of Pharmaceuticals and Illicit Drugs from Wastewater Due to Ferric Dosing in Sewers. *Environ. Sci. Technol.* 53, 6245–6254. <https://doi.org/10.1021/acs.est.8b07155>
- Lennie, A.R., Vaughan, D.J., 1996. Spectroscopic studies of iron sulfide formation and phase relations at low temperatures, in: Dyar, M.D., McCammon, C., Schaefer, M.W. (Eds.), *Mineral Spectroscopy: A Tribute to Roger G. Burns*. pp. 117–131.
- Lovley, D.R., 1993. Dissimilatory metal reduction. *Annu. Rev. Microbiol.* 47, 20–29.
- Lovley, D.R., 1987. Organic matter mineralization with the reduction of ferric iron: A review. *Geomicrobiol. J.* 5, 375–399. <https://doi.org/10.1080/01490458709385975>
- Lovley, D.R., Giovannoni, S.J., White, D.C., Champine, J.E., Phillips, E.J.P., Gorby, Y.A., Goodwin, S., 1993. *Geobacter metallireducens* gen. nov. sp. nov., a microorganism. *Arch. Microbiol.* 159, 336–344.
- Lovley, D.R., Lonergan, D.J., 1990. Anaerobic oxidation of toluene, phenol, and p-cresol by the dissimilatory iron-reducing organism, GS-15. *Appl. Environ. Microbiol.* 56, 1858–1864. <https://doi.org/10.1128/aem.72.5.3236-3244.2006>
- Lovley, D.R., Phillips, E.J.P., 1989. Requirement for a microbial consortium to completely oxidize glucose in Fe(III)-reducing sediments. *Appl. Environ. Microbiol.* 55, 3234–3236.
- Lovley, D.R., Phillips, E.J.P., 1988. Novel Mode of Microbial Energy Metabolism: Organic Carbon Oxidation Coupled to Dissimilatory Reduction of Iron or Manganese. *Appl. Environ. Microbiol.* 54, 1472–1480. <https://doi.org/10.1103/PhysRevLett.50.1998>
- Metcalf & Eddy, I., Tchobanoglous, G., Stensel, H.D., Tsuchihashi, R., Burton, F., 2014. *Wastewater Engineering: Treatment and Resource Recovery*. McGraw-Hill, New York.
- Moskowitz, B.M., Frankel, R.B., Bazylinski, D.A., Jannasch, H.W., Lovley, D.R., 1989. A comparison of magnetite particles produced anaerobically by magnetotactic and dissimilatory iron-reducing bacteria. *Geophys. Research Lett.* 16, 665–668.
- Naumkin, A. V., Kraut-Vass, A., Gaarenstroom, S.W., Powell, C.J., 2012. NIST X-ray

Photoelectron Spectroscopy Database [WWW Document].
<https://doi.org/http://dx.doi.org/10.18434/T4T88K>

- Nevin, K.P., Holmes, D.E., Woodard, T.L., Hinlein, E.S., Ostendorf, D.W., Lovley, D.R., 2005. *Geobacter bemidjiensis* sp. nov. and *Geobacter psychrophilus* sp. nov., two novel Fe(III)-reducing subsurface isolates. *Int. J. Syst. Evol. Microbiol.* 55, 1667–1674. <https://doi.org/10.1099/ijs.0.63417-0>
- Nevin, K.P., Lovley, D.R., 2002. Mechanisms for accessing insoluble Fe(III) oxide during dissimilatory Fe(III) reduction by *Geothrix fermentans*. *Appl. Environ. Microbiol.* 68, 2294–2299. <https://doi.org/10.1128/AEM.68.5.2294-2299.2002>
- Podosokorskaya, O.A., Kadnikov, V. V., Gavrilov, S.N., Mardanov, A. V., Merkel, A.Y., Karnachuk, O. V., Ravin, N. V., Bonch-Osmolovskaya, E.A., Kublanov, I. V., 2013. Characterization of *Melioribacter roseus* gen. nov., sp. nov., a novel facultatively anaerobic thermophilic cellulolytic bacterium from the class Ignavibacteria, and a proposal of a novel bacterial phylum Ignavibacteriae. *Environ. Microbiol.* 15, 1759–1771. <https://doi.org/10.1111/1462-2920.12067>
- Rabus, R., Venceslau, S.S., Wöhlbrand, L., Voordouw, G., Wall, J.D., Pereira, I.A.C., 2015. A Post-Genomic View of the Ecophysiology, Catabolism and Biotechnological Relevance of Sulphate-Reducing Prokaryotes. *Adv. Microb. Physiol.* 66, 55–321. <https://doi.org/10.1016/bs.ampbs.2015.05.002>
- Shah, M., Lin, C.-C., Kukkadapu, R., Engelhard, M.H., Zhao, X., Wang, Y., Barkay, T., Yee, N., 2014. Syntrophic Effects in a Subsurface Clostridial Consortium on Fe(III)-(Oxyhydr)oxide Reduction and Secondary Mineralization. *Geomicrobiol. J.* 31, 101–115.
- Tebo, B.M., Obratzsova, A.Y., 1998. Sulfate-reducing bacterium grows with Cr (VI), U (VI), Mn (IV), and Fe (III) as electron acceptors. *Source* 162, 193–198. <https://doi.org/10.1111/j.1574-6968.1998.tb12998.x>
- Thorpe, A.N., Senftle, F.E., Alexander, C.C., Dulong, F.T., 1984a. Oxidation of pyrite in coal to magnetite. *Fuel* 63, 662–668. [https://doi.org/10.1016/0016-2361\(84\)90163-7](https://doi.org/10.1016/0016-2361(84)90163-7)
- Thorpe, A.N., Senftle, F.E., Alexander, C.C., Dulong, F.T., 1984b. Oxidation of pyrite in coal to magnetite. *Fuel* 63, 662–668. [https://doi.org/10.1016/0016-2361\(84\)90163-7](https://doi.org/10.1016/0016-2361(84)90163-7)
- Ueki, A., Akasaka, H., Suzuki, D., Ueki, K., 2006. *Paludibacter propionicigenes* gen. nov., sp. nov., a novel strictly anaerobic, Gram-negative, propionate-producing bacterium isolated from plant residue in irrigated rice-field soil in Japan. *Int. J. Syst. Evol. Microbiol.* 56, 39–44. <https://doi.org/10.1099/ijs.0.63896-0>
- Vaclavkova, S., Jørgensen, C.J., Jacobsen, O.S., Aamand, J., Elberling, B., 2014. The Importance of Microbial Iron Sulfide Oxidation for Nitrate Depletion in Anoxic Danish Sediments. *Aquat. Geochemistry* 20, 419–435. <https://doi.org/10.1007/s10498-014-9227-x>
- Van Lier, J.B., Van der Zee, F.P., Frijters, C.T.M.J., Ersahin, M.E., 2015. Celebrating 40 years anaerobic sludge bed reactors for industrial wastewater treatment. *Rev.*

- Environ. Sci. Biotechnol. 14, 681–702. <https://doi.org/10.1007/s11157-015-9375-5>
- Waite, T.D., 2002. Challenges and opportunities in the use of iron in water and wastewater treatment. *Rev. Environ. Sci. Biotechnol.* 1, 9–15. <https://doi.org/10.1023/A:1015131528247>
- Wang, L., Pan, Y.X., Li, J.H., Qin, H.F., 2008. Magnetic properties related to thermal treatment of pyrite. *Sci. China, Ser. D Earth Sci.* 51, 1144–1153. <https://doi.org/10.1007/s11430-008-0083-7>
- Wang, N., Zheng, T., Zhang, G., Wang, P., 2016. A review on Fenton-like processes for organic wastewater treatment. *J. Environ. Chem. Eng.* 4, 762–787. <https://doi.org/10.1016/j.jece.2015.12.016>
- Waters, K.E., Rowson, N.A., Greenwood, R.W., Williams, A.J., 2008. The effect of heat treatment on the magnetic properties of pyrite. *Miner. Eng.* 21, 679–682. <https://doi.org/10.1016/j.mineng.2008.01.008>
- Weber, K.A., Achenbach, L.A., Coates, J.D., 2006. Microorganisms pumping iron: Anaerobic microbial iron oxidation and reduction. *Nat. Rev. Microbiol.* 4, 752–764. <https://doi.org/10.1038/nrmicro1490>
- Xiang, H., Liu, C., Pan, R., Han, Y., Cao, J., 2014. Magnetite for phosphorus removal in low concentration phosphorus-contained water body. *Adv. Environ. Res.* 3, 163–172.
- Yamada, T., Sekiguchi, Y., Imachi, H., Kamagata, Y., Ohashi, A., Harada, H., 2005. Diversity, Localization, and Physiological Properties of Filamentous Microbes Belonging to. *Microbiology* 71, 7493–7503. <https://doi.org/10.1128/AEM.71.11.7493>
- Yang, Y., Chen, T., Sumona, M., Gupta, B. Sen, Sun, Y., Hu, Z., Zhan, X., 2017. Utilization of iron sulfides for wastewater treatment: a critical review. *Rev. Environ. Sci. Biotechnol.* 16, 289–308. <https://doi.org/10.1007/s11157-017-9432-3>
- Zellner, G., Messner, P., Kneifel, H., Winter, J., 1989. *Desulfovibrio simplex* spec. nov., a new sulfate-reducing bacterium from a sour whey digester. *Arch. Microbiol.* 152, 329–334. <https://doi.org/10.1007/BF00425169>
- Zhang, Z., Wang, Y., Leslie, G.L., Waite, T.D., 2015. Effect of ferric and ferrous iron addition on phosphorus removal and fouling in submerged membrane bioreactors. *Water Res.* 69, 210–222. <https://doi.org/10.1016/j.watres.2014.11.011>

Chapter 5: Magnetic Sludge Byproducts for Adsorptive Removal of Phosphorus: Resource Recovery from Iron-based Anaerobic Sewage Sludge

(M. Ahmed, M. Azizah, R. Anwar, M. Johnson, L.-S. Lin, Magnetic Sludge Byproducts for Adsorptive Removal of Phosphorus: Resource Recovery from Iron-based Anaerobic Sewage Sludge, Manuscript under review in Waste Management Journal)

5.1 Introduction

Sludge management is an important aspect of biological wastewater treatment as it represents a difficult and expensive issue in wastewater engineering (Ciešlik et al., 2015; Uggetti et al., 2010; Yang et al., 2015). Worldwide, sludge management strategies mostly fall in three categories: land application, landfill disposal and incineration or thermal technologies (Campbell, 2000). In many instances, sludge is applied for land application at a high loading rate which exceeds the nutrient requirements of the vegetation. Excess nutrients from the sludge can contaminate groundwater and surface water through leaching from soil and via runoff generating from rainfall respectively (EPA, 2000). Severe environmental impact was observed by landfill gas and leachate emitted from landfill disposal, and local soil contamination with metals and pathogens due to land application. Incineration has a major drawback of air pollution due to the emission of contaminants, that can cause severe health and environmental risk (Hu et al., 2015; Roberts and Chen, 2006). Anaerobic digestion and pyrolysis are among the most promising processes practiced for energy generation from sewage sludge (Cao and Pawłowski, 2012). However, energy generation through these processes is highly dependent on the amount of organic matters present in the sludge. Biogas production using anaerobic digestion is a complex process as several parameters (e.g. pH, temperature, reactor type, retention time, organic loading rate) are needed to be controlled for an efficient process (Van et al., 2019). Moreover, the presence of incombustible gases like carbon dioxide (CO₂), hydrogen sulfide (H₂S) and water vapor reduce the calorific value of biogas and make the compression, transportation and energy generation expensive (Olugasa et al., 2014). Therefore, it is important to develop simple and cost-effective methods to extract valuable resources from the sludge. Various chemical and biological treatment approaches (e.g. stripping/absorption of ammonium, partial nitrification/anammox, struvite precipitation)

have been employed for nutrient recovery from sewage sludge (Ahmed et al., 2015; Antakyali et al., 2013; Booker et al., 1999; Vrieze et al., 2016). These approaches create new avenues of valuable resource recovery from sludge.

A novel, energy-efficient Fe(III)-dosed anaerobic biological wastewater treatment system was recently developed and showed effective treatment performance (Ahmed et al., 2020, 2019). In this biological treatment, organics oxidation is coupled to both iron- and sulfate-reduction facilitated by iron- and sulfate-reducing bacteria. The sludge materials produced from this treatment mostly contained inorganic precipitates and smaller amounts of biomass. Electronic microscopic and spectroscopic analyses of the sludge materials revealed that the inorganic precipitates are iron sulfide (FeS and FeS₂, Ahmed et al., 2020, 2019). Although sludge production rate in this anaerobic treatment system is lower than those of typical aerobic treatment processes, beneficial applications of the sludge materials can significantly improve the sustainability of the treatment method. Valuable resource recovery from the iron sulfide sludge with simple treatment approach can reduce the sludge management problem and minimize environmental footprint.

Different forms of iron sulfides (FeS, FeS₂, Fe_{1-x}S) and synthesized nanoparticles from iron sulfides were found effective for removing organic compounds (e.g., trichloroethylene, tetrachloroethylene, nitroaromatic compounds), heavy metals (e.g., Hg, Cd, Cr), oxyanions (arsenite/arsenate, selenite/selenate), nutrients (N and P), radionuclides (e.g., U, Tc, Np) through chemical precipitation, chemical reduction, surface sorption, and complexation from aqueous solutions (Gong et al., 2016; Yang et al., 2017). These iron sulfides known as weakly paramagnetic with very small and positive susceptibility (Waters et al., 2008), can be transformed into iron oxides with high magnetic properties. Iron oxides such as magnetite- and maghemite-based nanoparticles have been recognized as useful materials due to their technological and industrial applications (Khan et al., 2015; Teja and Koh, 2009). Their unique properties such as superparamagnetism, higher surface area to volume ratio, morphology and chemical composition, lead to broad applications in the field of environment, agriculture and biomedicine (Ali et al., 2016; Farhanian et al., 2018). Magnetite is identified to have major potential in biomedicine sector, especially in phototherapy, bacterial detection, drug

delivery, and bioimaging agents (Ansari et al., 2015; Ansari et al., 2019; Saeed et al., 2018; Xu et al., 2019). In the field of wastewater treatment and pollution control, iron oxide-based particles have been used as a coagulant for rapid sludge clarification and process intensification (Anderson et al., 1987; Booker et al., 1991), and as an effective adsorbent for contaminant removal from aqueous wastes due to their smaller size, and higher surface area (Nassar, 2012; Oliveira et al., 2002). Their magnetic properties allow separation of the adsorbents from the aqueous phase for further applications (Dave and Chopda, 2014). In particular, iron oxide particles (e.g., magnetite, maghemite, and hematite) have been extensively used for phosphate removal from aqueous system (Ajmal et al., 2018; Berner, 1973; Shahid et al., 2019). Magnetite particles were found effective for removing heavy metals such as lead, copper, zinc, manganese, and cadmium (El-Dib et al., 2019; Giraldo et al., 2013). More than 90% adsorptive removal of lead, zinc, cadmium, chromium, copper, and nickel from aqueous system by maghemite were also reported (Ahmadi et al., 2014; Hu et al., 2006). A synthesized magnetite-hematite mixture was also observed as a proficient adsorbent with maximum adsorption efficiency of 98, 99.50, and 99.84% for removal of lead (II), chromium (III) and cadmium (II) respectively from water (Ahmed et al., 2013).

Thermal treatment has been used to convert iron sulfide (FeS_2) into strong ferrimagnetic substances such as magnetite and maghemite (Thorpe et al., 1984; Wang et al., 2008; Waters et al., 2008). Weakly antiferromagnetic hematite ($\alpha\text{-Fe}_2\text{O}_3$) and strongly ferrimagnetic maghemite ($\gamma\text{-Fe}_2\text{O}_3$) were observed as the oxidation products of pyrite in coal treated at dry temperatures, as high as 600°C (Marusak et al., 1976). Another study by Nishihara and Kondo, (1959) revealed that at low partial pressure of oxygen, FeS_2 was converted stepwise to pyrrhotite (Fe_{1-x}S), FeO , Fe_3O_4 and finally to $\alpha\text{-Fe}_2\text{O}_3$ when treated at high temperature ($700\text{-}900^\circ\text{C}$). In another study, pyrite of bituminous coal was partially converted to magnetite when baked at $393\text{-}455^\circ\text{C}$ at low oxygen atmosphere (Thorpe et al., 1984). At low oxygen concentration, FeS_2 was observed to oxidize to Fe_3O_4 and $\alpha\text{-Fe}_2\text{O}_3$, but no magnetite phase was observed when temperature and oxygen concentration were too high, and only $\alpha\text{-Fe}_2\text{O}_3$ phase prevailed. Thus, temperature and partial pressure of oxygen both act as regulating factors for thermal conversion of pyrite

into magnetic particles. However, controlling the partial pressure of oxygen at high temperature is a complex task compared to baking samples with a simple muffle furnace.

The objective of this study was to improve sustainability of the newly developed Fe(III)-dosed anaerobic biological wastewater treatment system (Ahmed et al., 2020) by generating useful products from its sludge materials. A thermal method was applied to transform the sludge materials to products that have magnetic properties for beneficial uses (Supplementary Material, Fig. S9). Spectroscopic and magnetization analyses were conducted to characterize the chemical structures and magnetic properties of the sludge byproducts. Adsorptive removal of phosphate using the magnetic materials was examined as one of the beneficial applications of the sludge byproducts.

5.2 Materials and methods

5.2.1 Sample collection and preparation

Sludge samples were collected from a Fe(III)-dosed anaerobic bioreactor that was used for treatment of synthetic wastewaters for more than three years. Details of the bioreactor and experimental conditions were previously described (Ahmed et al., 2020). Sludge samples were collected every two months from the bioreactor under mass loading to COD 281 mg/d, Fe(III) 134.3 mg/d, and sulfur 38.2 mg/d (Fe/S molar ratio 2). The sludge solids were first separated from the solution using high-speed centrifugation (5000 x *g*) for 10 min. The solid samples were then heated for an hour in a muffle furnace (Lindberg Blue M, Thermo Scientific) at five different temperatures (300°C, 350°C, 400°C, 450°C, 500°C). After the thermal treatment, the products were crushed and milled to obtain a uniform size and denoted by S300, S350, S400, S450 and S500, respectively.

5.2.2 Material Characterization

The thermally transformed sludge byproducts were characterized for their composition using PANalytical X'Pert Pro X-ray Diffractometer (XRD) with a Cu-K α source ($\lambda = 1.542 \text{ \AA}$). Rietveld refinement for phase quantification was done by HighScore Pro software (Bish and Howard, 1988; De La Torre et al., 2001) for four different phases: magnetite (Fe₃O₄), hematite (α -Fe₂O₃), maghemite (γ -Fe₂O₃) and pyrite (FeS₂). Rietveld refinement for powder-diffraction data is a multi-parameter curve-fitting procedure used to quantify

the weight percentage (%) of the different crystalline phases present in a sample. Many diffraction peaks or lines of the diffraction pattern for each phase were considered to reduce the discrepancies of overlapping lines from different patterns. A fitted spectrum was determined by minimizing the χ -squared difference between the fit and experimental spectra, based on the known crystal structure parameters and fitting parameters (weight percent, crystallite size and strain) for each phase. This calculated spectrum includes the contributions from the sharp Bragg peaks from the crystalline components of the sample and the contribution of the smooth background that underlies the peaks. The observed, refined, background, and difference between observed and refined data are labeled I_O , I_C , I_{BG} and $I_O - I_C$ respectively. The χ -squared, goodness of fit (GOF) values are also presented for each spectrum. The particle size was calculated by Scherrer equation $L=K\lambda/\beta\cos\theta$ (Aziziha et al., 2019a; Patterson, 1939; Scherrer, 1912) where L is the particle size, K is a constant equal to 0.9 for spherical particles, λ is the X-ray wavelength, β is the full-width at half-maximum of the peak, and θ is the peak position.

5.2.3 Magnetic Measurements

A Physical Property Measurement System (PPMS[®]) Evercool II (Quantum Design) was used for the magnetic measurements of the sludge byproducts. This system allows the sample temperature to be varied from 1.8 to 400 K (with 1% accuracy) and the magnetic field (H) from -90 to 90 kOe (with an accuracy of 0.2 Oe). The estimated error in M/m (emu/g) was less than 0.1% for the samples used in this study. For the vibrating sample magnetometer (VSM) magnetic measurements, 10 mg of each powder sample was used. For each sample, the powder was compressed and sealed in a plastic tube sample holder (P/N 4096-388) and the sample holder was snapped into a brass half tube, which was appropriately positioned in the magnetic field. The small diamagnetic background related to the sample holder and the brass piece was measured (without the powder sample) and subtracted from the measured data associated with the powder samples.

5.2.4 Adsorption Experiments

Batch adsorption experiments were conducted on sludge byproducts baked at 350°C and 500°C using synthetic phosphate (PO_4^{3-}) solutions that were prepared at $\text{pH } 7.0 \pm 0.1$ using sodium phosphate dibasic (Na_2HPO_4) and sodium phosphate monobasic ($\text{NaH}_2\text{PO}_4 \cdot 2\text{H}_2\text{O}$). Phosphorous (P) concentrations were monitored during a 24-hr period to examine the adsorption kinetics. A range of PO_4^{3-} -P concentrations (20, 50, 100, 200, 300, 500, 800 and 1000 mg/L) were used to develop adsorption isotherms by letting the reaction reach equilibrium. For each batch, a predetermined amount of adsorbent (0.01 g) was added into a 50 ml Erlenmeyer flask, followed by the addition of 15 mL of the PO_4^{3-} solution resulting in an adsorbent loading rate of 0.67 g/L. The flasks were covered with parafilm and placed in an incubator shaker (MaxQ 4000, Thermo Scientific) at 150 rpm for mixing. After adsorption, the suspension samples were filtered through a 0.45 μm membrane to remove the solids and the P concentrations in the filtrate samples were measured by the Ascorbic Acid method (Standard Method 4500 E, APHA et al., 2005). An experiment was also conducted to study the effects of pH on the adsorption by using 0.1N HCl and 0.02N NaOH solutions to control the pH.

5.2.5 Adsorption Kinetics and Isotherm Modeling

A pseudo-second-order kinetic model was applied on the kinetic data to gain a better insight into the adsorption kinetics. This model takes the form

$$\frac{t}{q} = \frac{1}{kq_e^2} + \frac{t}{q_e} \quad (1)$$

where q_e and q (mg PO_4^{3-} -P/g-adsorbent) are the amount of PO_4^{3-} -P adsorbed by the adsorbent at equilibrium and at given time t (min), respectively. Adsorption rate constant k [g/ (mg \times min)] and adsorption capacity q_e (mg- PO_4^{3-} -P/g-adsorbent) were calculated from the slope and intercept of the linear regression line.

Non-linear Langmuir (Eq. 2) and Freundlich (Eq. 3) models were used to characterize the adsorption isotherms (Ajmal et al., 2018; Desta, 2013; Krishna and Swamy, 2012). Langmuir model is based on the assumption that adsorption can occur only at a finite number of surface sites and maximum adsorption corresponds to a saturated monolayer

of adsorbate molecules on the adsorbent surface (Dada et al., 2012). The Freundlich model can be applied to the multilayer adsorption, based on an assumption of heterogenous surface energies (Krishna and Swamy, 2012). The Langmuir and Freundlich equations used are

$$q_e = q_m K_L \frac{C_e}{1 + K_L C_e} \quad \text{and} \quad (2)$$

$$q_e = K_F C_e^{1/n} \quad , \quad (3)$$

where, C_e (mg/L), q_m (mg PO_4^{3-} -P/g-adsorbent) and K_L (L/mg) represent the equilibrium aqueous P concentration, the maximum adsorption capacity, and Langmuir adsorption constant, respectively. The Freundlich parameter K_F ((mg/g) (L/mg) $^{1/n}$) quantifies the adsorption capacity and $1/n$ (dimensionless) is indicative of adsorption intensity. Non-linear fitting of both the models were done using Microsoft Excel Solver.

5.3 Results and Discussion

5.3.1 Material Characterization

5.3.1.1 Material size and structure

The X-ray diffraction spectra of the sludge byproducts baked at 300, 350, 400, 450, and 500 °C indicate that the samples were a mixture of amorphous and crystalline structures (Fig. 20). Sharp peaks and broad hump peaks of the spectra represent the crystalline and amorphous from of the samples respectively. The intensity of the sharp peaks increased with the temperature, which indicates the increase of the sample's crystallinity with temperature. The crystallite size of the materials ranged from 7 to 15 nm, where size was observed to increase with the baking temperature (data not shown). These sizes are comparable to the size of the iron oxide nanocrystals reported in previous studies (Demortière et al., 2011). Magnetic iron oxide nanocrystals have many technological and industrial applications due to their unique physical and chemical properties (Demortière et al., 2011; Liu et al., 2016; Tian et al., 2015; Willis et al., 2005). Particularly, they have significant contribution in the field of biomedicine such as drug delivery, DNA detection, magnetic resonance imaging (MRI), cell separation technique etc. (Hogemann et al., 2000; Josephson et al., 2001; Willis et al., 2005). There is an enormous scope for using

magnetic nanocrystals for development of magnetic or optical devices, including permanent magnets, recording media (Chaubey et al., 2008; Weller and McDaniel, 2006). Moreover, these particles can also be used as adsorptive material for pollutant removal for air emission and wastewater treatment due to their high surface area to size ratio (Chiavola et al., 2016; Karatza et al., 2013; Molino et al., 2013).

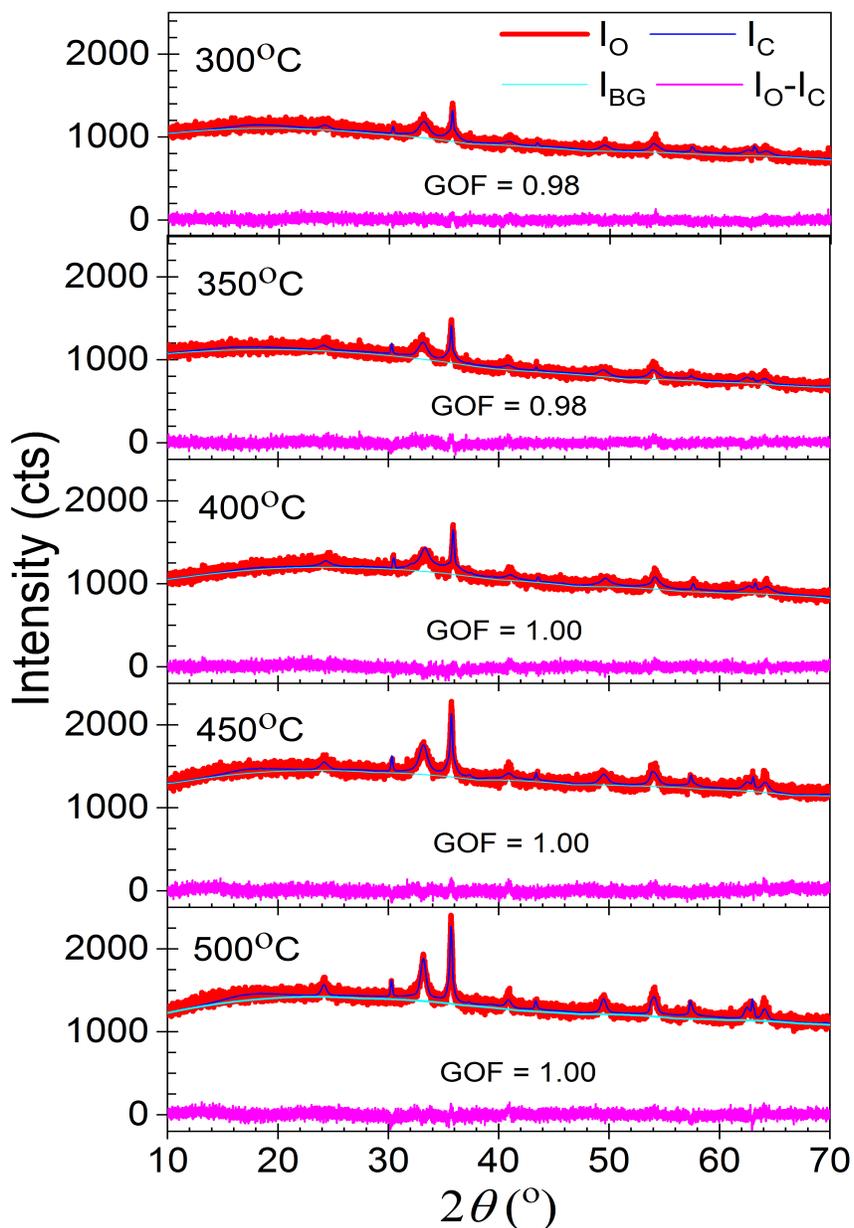


Fig. 20: XRD spectra of the sludge byproducts derived from the thermal treatment. Red: XRD spectrum, Blue: Rietveld fit, Cyan: Background, Pink: Difference between the experimental and Rietveld refinement

5.3.1.2 Material phase analysis

The Rietveld refinement was performed on the XRD spectrum by considering four phases of iron minerals α -Fe₂O₃ (hematite), γ -Fe₂O₃ (maghemite), Fe₃O₄ (magnetite), and FeS₂ (pyrite). The phase weight percentages (%) did not show any particular trend with the temperature (Fig. 21). In all cases, magnetite and hematite were found to be the most dominant phases in the materials (Fig. 21). Magnetite phase ranged from 14 to 39%, with the highest percentage observed at 350°C. The weight percentage of the hematite phase ranged from 41 to 76%, with its highest percentage, observed at 400°C. Another, ferromagnetic phase observed in this study was maghemite (γ -Fe₂O₃). Maghemite is likely an intermediate phase between the transformation of magnetite to hematite and has a structure and ferromagnetic properties similar to magnetite (Khan et al., 2015; Nikiforov et al., 2014). The presence of both magnetite and maghemite phases in the byproducts indicates successful conversion of the sludge materials to magnetic particles. More magnetite extraction might have been possible if the thermal treatment was conducted at low oxygen concentrations (1.0-3.0 kPa) (Thorpe et al., 1984), as the magnetite phase has lower stability at high oxygen concentrations. In this study, strict partial pressure or low oxygen concentration were not maintained, only a simple way of thermal treatment process was used.

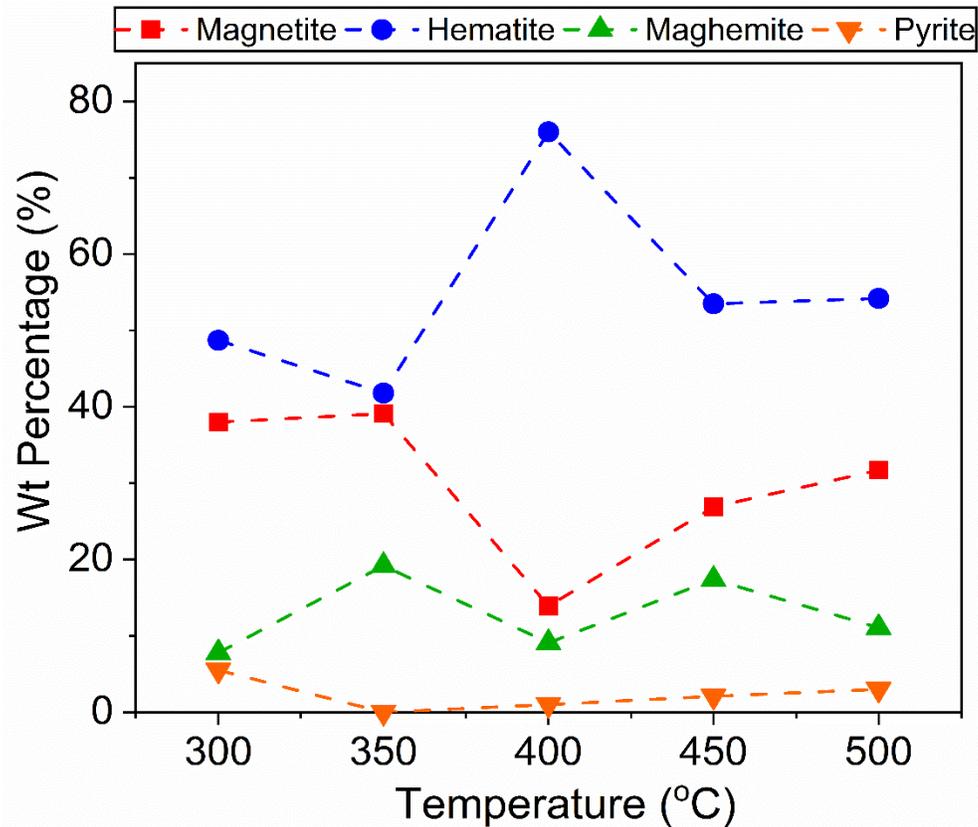


Fig. 21: Weight percentages of different phases of iron minerals derived from the sludge materials at different temperatures

5.3.2 Magnetic Measurements

High-resolution hysteresis measurements on the sludge byproducts taken at room temperature for magnetic field $H = -20$ to 20 kOe are shown in Fig. 22. The coercivity, H_c (field necessary to completely demagnetize a fully magnetized ferromagnet), remanent magnetization, M_r (residual magnetization in a ferromagnetic material after the external magnetic field is removed), and saturation magnetization, M_s (highest magnetization in a ferromagnetic material) were estimated from these hysteresis measurements. The values of M_s ranged from 6.3 to 10.9 emu/g and M_r ranged from 0.7 to 2.0 emu/g (Supplementary Materials, Table 1), which are lower than the reported magnetization values for pure magnetite and maghemite (Aziziha et al., 2019b; Bao et al., 2011; Dar and Shivashankar, 2014; Lee et al., 2005; Özdemir and Dunlop, 2014). These smaller values are not

surprising because these magnetic properties vary considerably depending on many details, such as crystallite size, strain, purity, and defects.

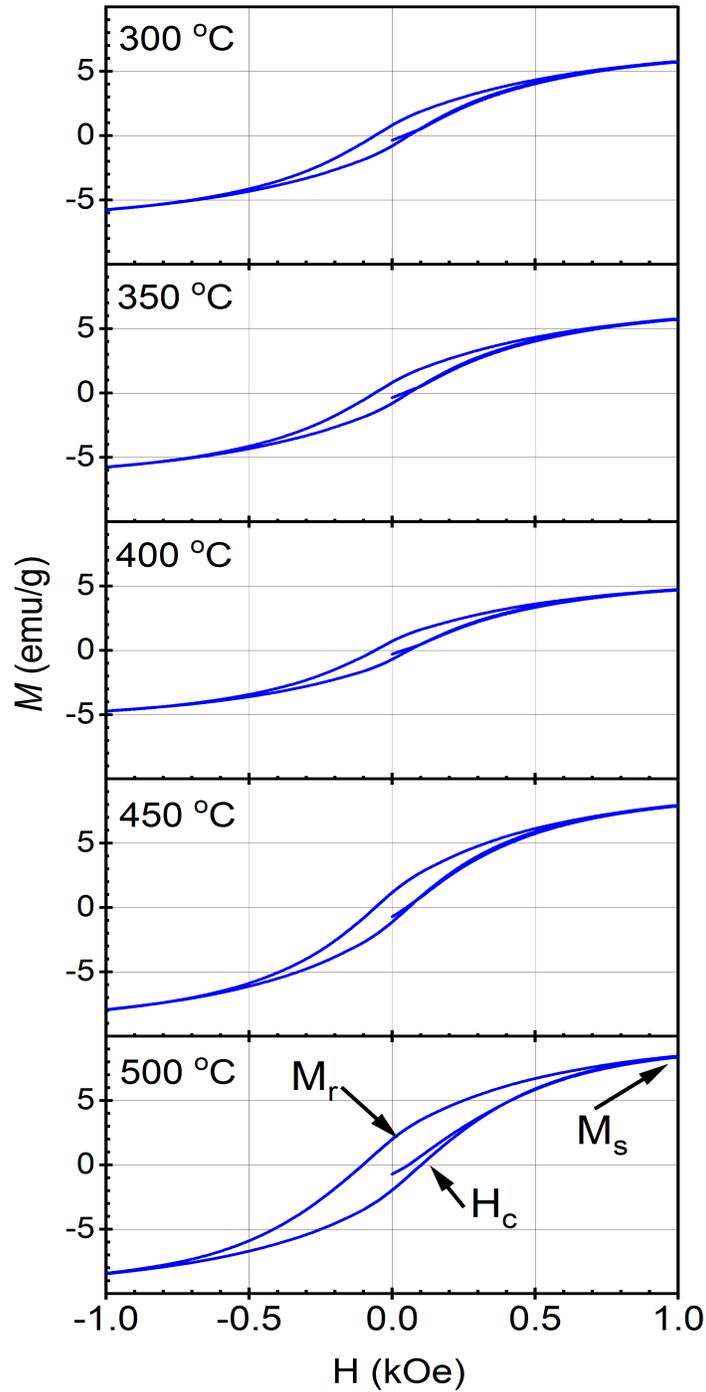


Fig. 22: High-resolution hysteresis measurements of the sludge byproducts derived at different temperatures

There was no significant change in remanent magnetization (M_r) for bake temperatures from 300 to 400°C, but M_r increased significantly above 400°C (Fig. 23). The highest magnetization (both remanent and saturation) was observed for the 500°C baked sample. This increased magnetization is probably due to the change in particle size. The magnetic properties of nanocrystals were reported to be dependent on the shape and size of the particles with magnetization increasing with the crystallite size (Bao et al., 2011; Byrne et al., 2011; Lee et al., 2005; Wu et al., 2008; Xu et al., 2007). With the increasing bake temperatures, the crystallite size is observed to increase, which is consistent with the observed increased magnetization. The substantial magnetite phase (32%) observed for the 500°C sample in large part explains the high magnetization for this S500 sample. Given the highest % of magnetite was observed at 350 °C whereas highest magnetization was observed at 500°C, for this study, we used both the S350 and S500 samples to evaluate the adsorption capabilities.

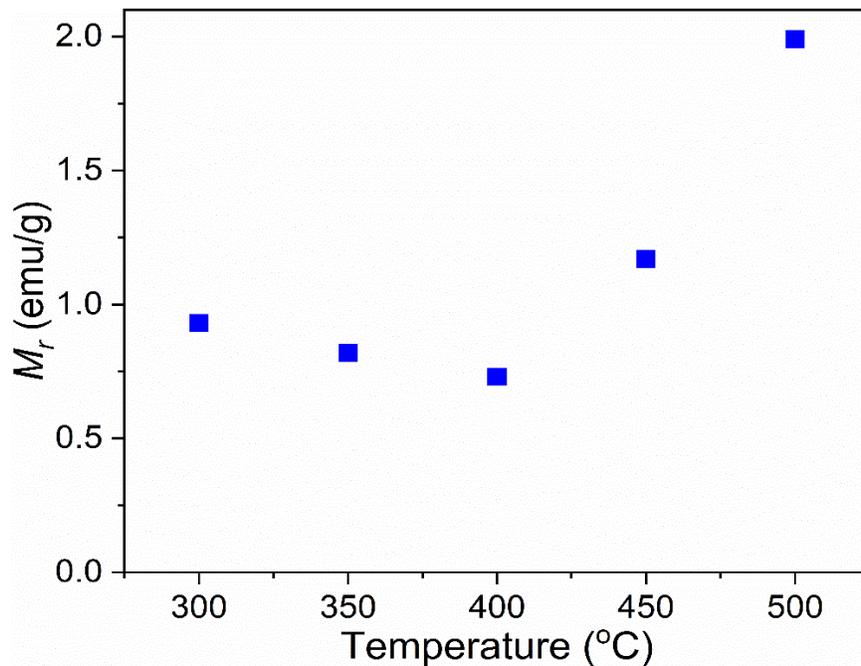


Fig. 23: Remanent magnetization (M_r) of the sludge byproducts derived at different temperatures

5.3.3 Adsorption Performance

5.3.3.1 Adsorption Kinetics

Adsorption kinetics of S350 and S500 for $\text{PO}_4^{3-}\text{-P}$ showed rapid adsorption that reached 98% and 90% of the equilibrium capacities at pH 7.0 within 3 hours for S350 and S500 respectively (Fig. 24). Adsorption equilibrium was reached within 24 hours for both samples. For both the adsorbents, pseudo-second-order model characterized the adsorption kinetics well ($R^2= 0.9999$ for S350 and $R^2= 0.9982$ for S500). The estimated equilibrium adsorption capacity (q_e) was lower for S500 than for S350 (40 vs. 44 mg $\text{PO}_4^{3-}\text{-P/g-adsorbent}$), similarly the rate constant was also lower for S500 than S350 (0.001 vs. 0.006 g/ (mg \times min)). With increasing temperature, samples became more crystalline with larger particle size and thus less specific surface area available for adsorption. This reduction in specific surface area probably leads to the lower equilibrium adsorption capacity for S500. The higher rate constant of S350 than S500 indicates S350 sample reached to equilibrium adsorption capacity faster. The rate constants observed in this study are slightly better than the rate constants observed for pure iron oxide particles, which indicate good adsorption capacity of the treated sludge materials (Ajmal et al., 2018; Yoon et al., 2014).

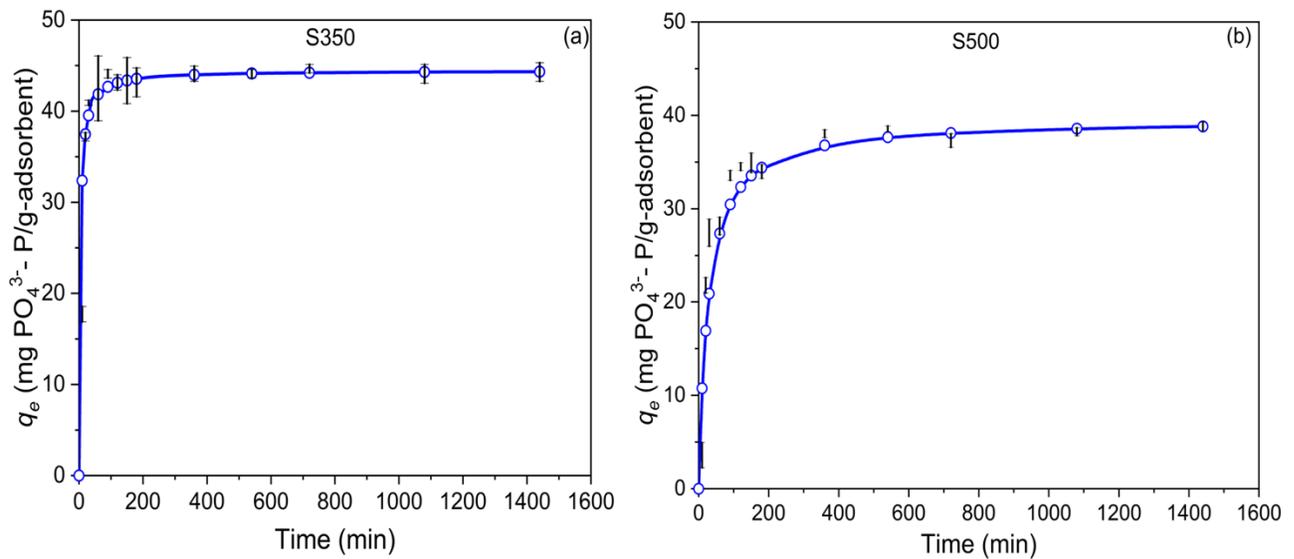


Fig. 24: Adsorption kinetic data and pseudo-second-order kinetic models for (a) S350 and (b) S500

5.3.3.2 Effects of pH

The effects of pH on S350 was examined using initial $\text{PO}_4^{3-}\text{-P}$ concentration of 100 mg/L. The adsorption capacity ranged from 31 to 35 mg $\text{PO}_4^{3-}\text{-P/g}$ adsorbent for pH from 3 to 6. The adsorption capacity decreased considerably at pHs above 6, leading to about 50% reduction at pH 10.5 from the acidic conditions (Fig. 25). This pH dependent adsorption of P can be explained by the point of zero charges (pH_{PZC}) of magnetite and hematite, which were the dominant minerals in the sludge byproduct. The pH_{PZC} values of magnetite and hematite were reported to be from 6.5 to 7 in different studies (Čerović et al., 2009; Milonjić et al., 1983; Preočanin et al., 2011). At low pHs (3-6), the adsorbent surface was dominated by positive charges, exhibiting higher electrostatic interactions with anionic phosphate (H_2PO_4^- and HPO_4^{2-}) and resulting in better adsorption performance. At pH above pH_{PZC} , overall negative surface charges caused adsorption to decline.

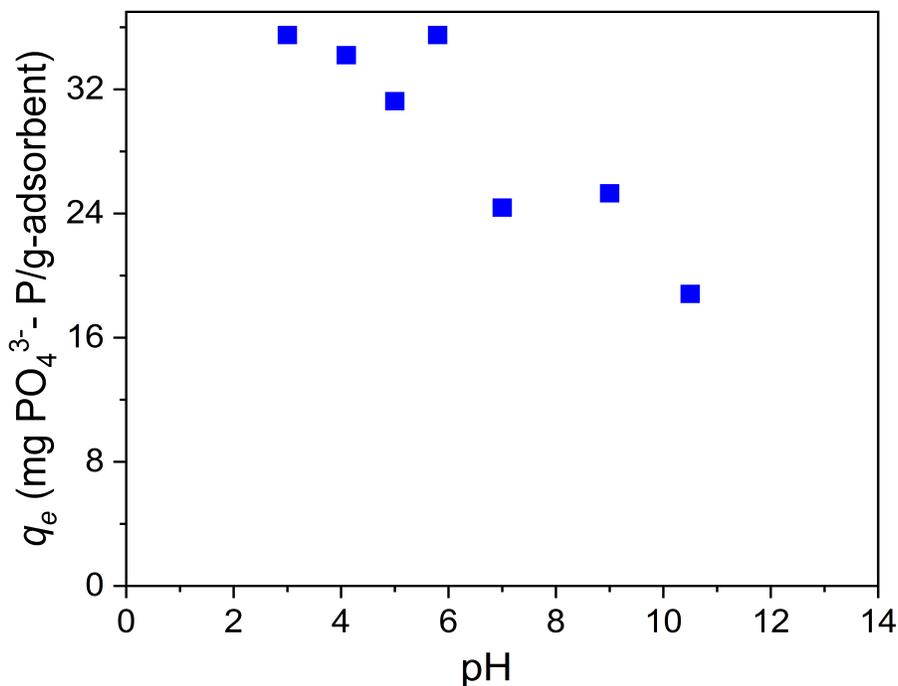


Fig. 25: Adsorption capacity of S350 for phosphate ions

5.3.3.3 Adsorption Isotherms

The adsorption capacities were higher for S350 than S500 at different equilibrium concentrations (C_e). High crystallinity, larger crystals in S500 resulted in lower surface area for adsorption versus low crystalline, smaller crystals in S350. For S350 and S500,

the adsorption capacities were similar at lower equilibrium concentrations, but showed significant differences at high equilibrium concentrations (Fig. 26). The high adsorption capacities of S350 and S500 at high $\text{PO}_4^{3-}\text{-P}$ concentrations suggest that these treated sludge materials can be excellent adsorbent for $\text{PO}_4^{3-}\text{-P}$ removal from nutrient rich wastewater, e.g., agricultural drainage, urban runoff, industrial wastewater.

According to the R^2 values in Table 5, the Langmuir and Freundlich models characterize the isotherm data well for S350. But, for S500, the Freundlich model ($R^2 = 0.89$) does not fit as well as Langmuir model ($R^2 = 0.95$). For the Langmuir model, the maximum adsorption capacity at pH 7.0 was estimated as 310 and 190 mg $\text{PO}_4^{3-}\text{-P/g}$ -adsorbent for S350 and S500, respectively (Fig. 26a). These adsorption capacities are slightly higher than the reported values of iron oxides used as an adsorbent in previous studies (Ajmal et al., 2018; Sousa et al., 2012; Trazzi et al., 2016). The feasibility of PO_4^{3-} adsorption can also be measured with a dimensionless parameter R_L , expressed as $1/(1+K_L \cdot C_0)$. Adsorption is considered favorable if R_L is between 0 and 1, unfavorable if greater than 1. The average R_L was 0.55 and 0.52 for S350 and S500, respectively, indicating their favorable adsorption of $\text{PO}_4^{3-}\text{-P}$ from the aqueous solution.

Table 5: Langmuir and Freundlich Isotherm model parameters for S350 and S500

Isotherm Model						
Temp. (°C)	Langmuir			Freundlich		
	q_{max} (mg $\text{PO}_4^{3-}\text{-P/g}$ adsorbed)	K_L (L/mg)	R^2	K_f (mg/g) (L/mg) ^{1/n}	n	R^2
350	310	0.004	0.94	9.8	2.09	0.97
500	190	0.004	0.95	5.7	2.04	0.89

K_F values were estimated to be 9.8 and 5.7 (mg/g) (L/mg)^{1/n} for S350 and S500, respectively. These values are relatively higher than the reported values of iron oxide and iron coated adsorbents (Ajmal et al., 2018; Boujelben et al., 2008). The high K_F values suggest easy uptake of PO_4^{3-} from solution with a high adsorptive capacity of the

adsorbent. The K_F value decreased with increasing baking temperature, which is consistent with results obtained from the Langmuir model fitting. The n values obtained from the Freundlich models lie between one and ten, indicating a favorable sorption process (Krishna and Swamy, 2012). The results obtained from both isotherm models suggest that both the sludge byproducts derived from 350 and 500°C thermal treatment can be used to efficiently remove $\text{PO}_4^{3-}\text{-P}$ from aqueous solutions.

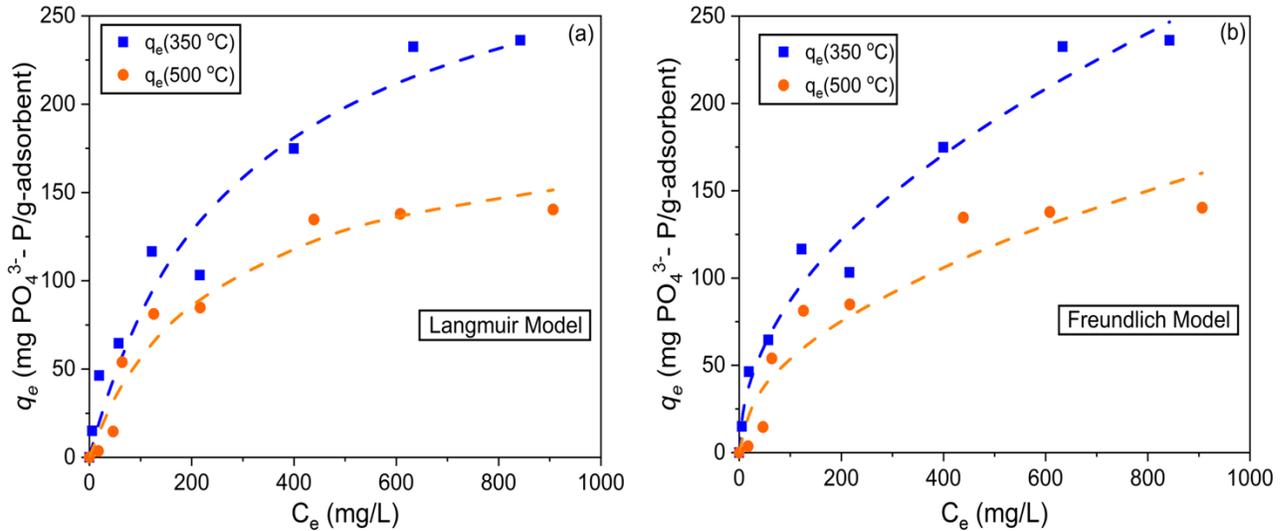


Fig. 26: Adsorption isotherm data, and (a) Langmuir and (b) Freundlich models for S350 and S500 at pH 7.0

5.4 Conclusion

In this study, a simple thermal treatment method was developed to produce magnetic particles from the iron sulfide sludge of a newly developed iron-dosed anaerobic biological treatment system. This innovative approach has demonstrated a potential pathway of recovering useful sludge byproducts while alleviating sludge disposal or management. Extracting sludge and its conversion to magnetic byproducts, and using them for beneficial applications can improve the sustainability of the wastewater treatment system. For practical applications, dewatering of the sludge before the thermal treatment would significantly reduce the energy required for generating the byproducts. There are opportunities of using the magnetic iron oxides (magnetite, maghemite) within the Fe(III)-dosed treatment system for process intensification. For examples, they can be used as a

coagulant in the primary clarifier or adsorptive phosphate removal from wastewater effluent as a polishing treatment. Alternatively, they can be used as a supplement for iron dosing as an electron acceptor for organics oxidation facilitated by iron reducers. This will reduce the demand of external iron source as well as reduce the environmental footprint of the treatment system. Further studies are also warranted to transform the sludge under controlled conditions (e.g., O₂ partial pressure) for better control of the byproduct characteristics (e.g., size, surface magnetic phases) and purification to generate products for targeted applications, such as those in the biomedical field.

5.5 Supplementary Materials

Table S1: Saturation magnetization (M_s), Remanent magnetization (M_r) and Coercive field (H_c) at each temperature

Baking Temperature, T (°C)	Coercive Field, H_c (Oe)	Remanent Magnetization, M_r (emu/g)	Saturation Magnetization, M_s (emu/g)
300	75	0.9	7.2
350	50	0.8	7.8
400	50	0.7	6.3
450	50	1.2	10.4
500	100	2.0	10.9

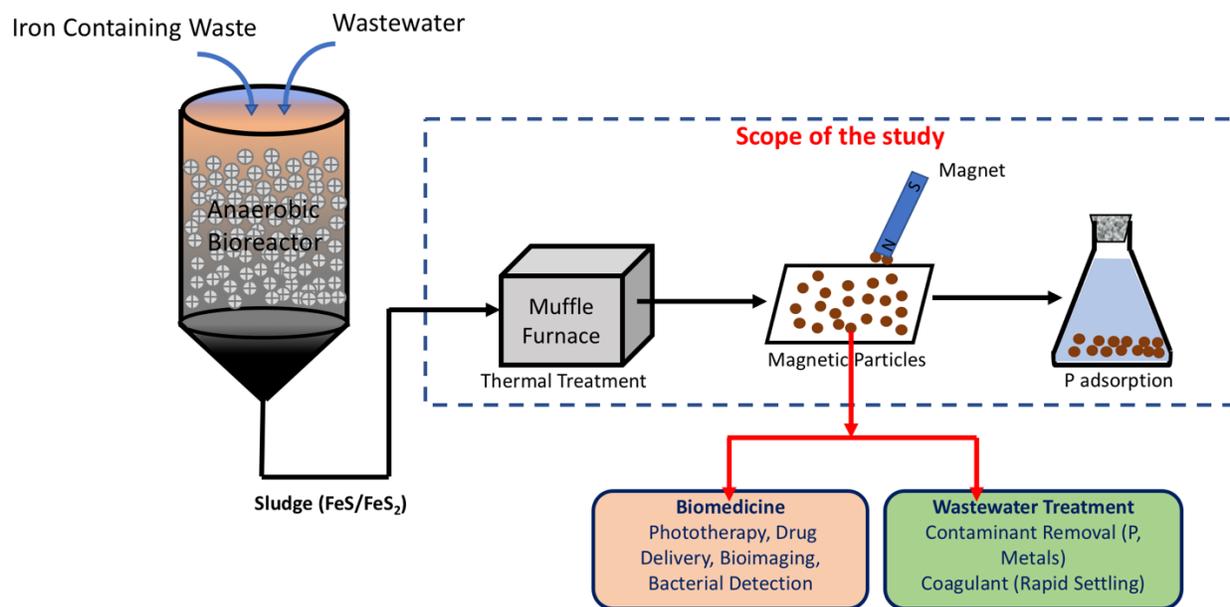


Fig. S9: Schematic diagram depicting the scope of this study

References

- Ahmadi, A., Heidarzadeh, S., Mokhtari, A.R., Darezereshki, E., Harouni, H.A., 2014. Optimization of heavy metal removal from aqueous solutions by maghemite (γ -Fe₂O₃) nanoparticles using response surface methodology. *J. Geochemical Explor.* 147, 151–158.
- Ahmed, M., Hasan, C.K., Rahman, H., Ali Hossain, M., Uddin, S.A., 2015. Prospects of using wastewater as a resource-nutrient recovery and energy generation. *Am. J. Environ. Sci.* <https://doi.org/10.3844/ajessp.2015.99.114>
- Ahmed, M., Lin, O., Saup, C.M., Wilkins, M.J., Lin, L.S., 2019. Effects of Fe/S ratio on the kinetics and microbial ecology of an Fe(III)-dosed anaerobic wastewater treatment system. *J. Hazard. Mater.* 369, 593–600. <https://doi.org/10.1016/j.jhazmat.2019.02.062>
- Ahmed, M., Saup, C.M., Wilkins, M.J., Lin, L.-S., 2020. Continuous ferric iron-dosed anaerobic wastewater treatment: treatment performance, sludge characteristics, and microbial composition. *J. Environ. Chem. Eng.* 8. <https://doi.org/10.1016/j.jece.2019.103537>
- Ahmed, M.A., Ali, S.M., El-Dek, S.I., Galal, A., 2013. Magnetite-hematite nanoparticles prepared by green methods for heavy metal ions removal from water. *Mater. Sci. Eng. B* 178, 744–751. <https://doi.org/10.1016/j.mseb.2013.03.011>
- Ajmal, Z., Muhmood, A., Usman, M., Kizito, S., Lu, J., Dong, R., Wu, S., 2018. Phosphate removal from aqueous solution using iron oxides: Adsorption, desorption and regeneration characteristics. *J. Colloid Interface Sci.* 528, 145–155. <https://doi.org/10.1016/j.jcis.2018.05.084>
- Ali, A., Zafar, H., Zia, M., ul Haq, I., Phull, A.R., Ali, J.S., Hussain, A., 2016. Synthesis, characterization, applications, and challenges of iron oxide nanoparticles.

- Nanotechnol. Sci. Appl. 9, 49–67. <https://doi.org/10.2147/NSA.S99986>
- Anderson, N.J., Blesing, N. V., Bolto, B.A., Jackson, M.B., 1987. The role of polyelectrolytes in a magnetic process for water clarification. *React. Polym. ion Exch. Sorbents* 7, 47–55.
- Ansar, M.Z., Atiq, S., Riaz, S., Naseem, S., 2015. Magnetite Nano-crystallites for Anti-cancer Drug Delivery, in: *International Conference on Solid State Physics 2013*. Elsevier Ltd., pp. 5410–5414. <https://doi.org/10.1016/j.matpr.2015.11.060>
- Ansari, S.A.M.K., Ficiarà, E., Ruffinatti, F.A., Stura, I., Argenziano, M., Abollino, O., Cavalli, R., Guiot, C., D'Agata, F., 2019. Magnetic iron oxide nanoparticles: Synthesis, characterization and functionalization for biomedical applications in the Central Nervous System. *Materials (Basel)*. 12, 1–24. <https://doi.org/10.3390/ma12030465>
- Antakyali, D., Meyer, C., Preyl, V., Maier, W., Steinmetz, H., 2013. Large-scale application of nutrient recovery from digested sludge as struvite. *Water Pract. Technol.* 8, 256–262. <https://doi.org/10.2166/wpt.2013.027>
- APHA, AWWA, WEF, 2005. *Standard methods for the examination of water and wastewater*, 21st ed. American Public Health Association, American water Works Association, and Water Environment Federation.
- Aziziha, M., Beesley, R., Magers, J.R., Mottaghi, N., Holcomb, M.B., Lewis, J.P., Seehra, M.S., Johnson, M.B., 2019a. Electronic state and concentration of Fe³⁺ in CuAl_{1-x}Fe_xO₂ determined by magnetic measurements. *J. Magn. Mater.* 471, 495–500.
- Aziziha, M., Byard, S.A., Beesely, R., Lewis, J.P., Seehra, M.S., Johnson, M.B., 2019b. Magnetic properties of Fe-doped CuAlO₂ and role of impurities. *AIP Adv.* 9. <https://doi.org/10.1063/1.5080099>
- Bao, L., Yang, H., Wang, X., Zhang, F., Shi, R., Liu, B., Wang, L., Zhao, H., 2011. Synthesis and size-dependent magnetic properties of single-crystalline hematite nanodiscs. *J. Cryst. Growth* 328, 62–69. <https://doi.org/10.1016/j.jcrysgro.2011.05.030>
- Berner, R.A., 1973. Phosphate removal from sea water by adsorption on volcanogenic ferric oxides. *Earth Planet. Sci. Lett.* 18, 77–86. [https://doi.org/10.1016/0012-821X\(73\)90037-X](https://doi.org/10.1016/0012-821X(73)90037-X)
- Bish, D.L., Howard, S.A., 1988. Quantitative phase analysis using the Rietveld method. *J. Appl. Crystallogr.* 21, 86–91. <https://doi.org/10.1107/S0021889887009415>
- Booker, N.A., Keir, D., Priestley, A.J., Ritchie, C.B., Sudarmana, D.L., Woods, M.A., 1991. Sewage clarification with magnetite particles. *Water Sci. Technol.* 23, 1703–1712. <https://doi.org/10.2166/wst.1991.0625>
- Booker, N.A., Priestley, A.J., Fraser, I.H., 1999. Struvite formation in wastewater treatment plants: Opportunities for nutrient recovery. *Environ. Technol.* 20, 777–782.
- Boujelben, N., Bouzid, J., Elouear, Z., Feki, M., Jamoussi, F., Montiel, A., 2008. Phosphorus removal from aqueous solution using iron coated natural and engineered sorbents. *J. Hazard. Mater.* 151, 103–110. <https://doi.org/10.1016/j.jhazmat.2007.05.057>
- Byrne, J.M., Telling, N.D., Coker, V.S., Patrick, R.A.D., Van Der Laan, G., Arenholz, E., Tuna, F., Lloyd, J.R., 2011. Control of nanoparticle size, reactivity and magnetic properties during the bioproduction of magnetite by *Geobacter sulfurreducens*.

- Nanotechnology 22. <https://doi.org/10.1088/0957-4484/22/45/455709>
- Campbell, H.W., 2000. Sludge management - Future issues and trends. *Water Sci. Technol.* 41, 1–8.
- Cao, Y., Pawłowski, A., 2012. Sewage sludge-to-energy approaches based on anaerobic digestion and pyrolysis: Brief overview and energy efficiency assessment. *Renew. Sustain. Energy Rev.* 16, 1657–1665. <https://doi.org/10.1016/j.rser.2011.12.014>
- Čerović, L., Lefèvre, G., Jaubertie, A., Fédoroff, M., Milonjić, S., 2009. Deposition of hematite particles on polypropylene walls in dynamic conditions. *J. Colloid Interface Sci.* 330, 284–291. <https://doi.org/10.1016/j.jcis.2008.10.079>
- Chaubey, G.S., Nandwana, V., Poudyal, N., Rong, C.B., Ping Liu, J., 2008. Synthesis and characterization of bimagnetic bricklike nanoparticles. *Chem. Mater.* 20, 475–478. <https://doi.org/10.1021/cm7028068>
- Chiavola, A., D'Amato, E., Stoller, M., Chianese, A., Boni, M.R., 2016. Application of iron based nanoparticles as adsorbents for Arsenic removal from water. *Chem. Eng. Trans.* 47, 325–330. <https://doi.org/10.3303/CET1647055>
- Ciešlik, B.M., Namiešnik, J., Konieczka, P., 2015. Review of sewage sludge management: Standards, regulations and analytical methods. *J. Clean. Prod.* 90, 1–15. <https://doi.org/10.1016/j.jclepro.2014.11.031>
- Dada, A.O., Olalekan, A.P., Olatunya, A.M., Dada, O., 2012. Langmuir, Freundlich, Temkin and Dubinin–Radushkevich Isotherms Studies of Equilibrium Sorption of Zn 2+ Unto Phosphoric Acid Modified Rice Husk. *IOSR J. Appl. Chem.* 3, 38–45. <https://doi.org/10.9790/5736-0313845>
- Dar, M.I., Shivashankar, S.A., 2014. Single crystalline magnetite, maghemite, and hematite nanoparticles with rich coercivity. *RSC Adv.* 4, 4105–4113. <https://doi.org/10.1039/c3ra45457f>
- Dave, P.N., Chopda, L. V., 2014. Application of iron oxide nanomaterials for the removal of heavy metals. *J. Nanotechnol.* 2014. <https://doi.org/10.1155/2014/398569>
- De La Torre, A.G., Bruque, S., Aranda, M.A.G., 2001. Rietveld quantitative amorphous content analysis. *J. Appl. Crystallogr.* 34, 196–202. <https://doi.org/10.1107/S0021889801002485>
- Demortière, A., Panissod, P., Pichon, B.P., Pourroy, G., Guillon, D., Donnio, B., Bégin-Colin, S., 2011. Size-dependent properties of magnetic iron oxide nanocrystals. *Nanoscale* 3, 225–232. <https://doi.org/10.1039/c0nr00521e>
- Desta, M.B., 2013. Batch Sorption Experiments: Langmuir and Freundlich Isotherm Studies for the Adsorption of Textile Metal Ions onto Teff Straw (*Eragrostis tef*) Agricultural Waste Mulu. *J. Thermodyn.* 61, 1–6. <https://doi.org/10.1080/00380768.2015.1020506>
- El-Dib, F.I., Mohamed, D.E., El-Shamy, O.A.A., Mishrif, M.R., 2019. Study the adsorption properties of magnetite nanoparticles in the presence of different synthesized surfactants for heavy metal ions removal. *Egypt. J. Pet.* 0–6. <https://doi.org/10.1016/j.ejpe.2019.08.004>
- EPA, 2000. Land Application of Biosolids, in: *Biosolids Technology Fact Sheet*. pp. 1–9.
- Farhanian, D., De Crescenzo, G., Tavares, J.R., 2018. Large-Scale Encapsulation of Magnetic Iron Oxide Nanoparticles via Syngas Photo-Initiated Chemical Vapor Deposition. *Sci. Rep.* 8, 1–11. <https://doi.org/10.1038/s41598-018-30802-1>
- Giraldo, L., Erto, A., Moreno-Piraján, J.C., 2013. Magnetite nanoparticles for removal of

- heavy metals from aqueous solutions: Synthesis and characterization. *Adsorption* 19, 465–474. <https://doi.org/10.1007/s10450-012-9468-1>
- Gong, Y., Tang, J., Zhao, D., 2016. Application of iron sulfide particles for groundwater and soil remediation: A review. *Water Res.* 89, 309–320. <https://doi.org/10.1016/j.watres.2015.11.063>
- Hogemann, D., Josephson, L., Weissleder, R., Basilion, J.P., 2000. Improvement of MRI probes to allow efficient detection of gene expression. *Bioconjug. Chem.* 11, 941–946. <https://doi.org/10.1021/bc000079x>
- Hu, H., Li, X., Nguyen, A.D., Kavan, P., 2015. A critical evaluation of waste incineration plants in Wuhan (China) based on site selection, environmental influence, public health and public participation. *Int. J. Environ. Res. Public Health* 12, 7593–7614. <https://doi.org/10.3390/ijerph120707593>
- Hu, J., Chen, G., Lo, I.M.C., 2006. Selective Removal of Heavy Metals from Industrial Wastewater Using Maghemite Nanoparticle: Performance and Mechanisms. *J. Environ. Eng.* 132, 709–715. [https://doi.org/10.1061/\(ASCE\)0733-9372\(2006\)132](https://doi.org/10.1061/(ASCE)0733-9372(2006)132)
- Josephson, L., Perez, J.M., Weissleder, R., 2001. Magnetic Nanosensors for the Detection of Oligonucleotide Sequences. *Angew. Chemie* 113, 3204–3206.
- Karataz, D., Lancia, A., Prisciandaro, M., Musmarra, D., Mazziotti Di Celso, G., 2013. Influence of oxygen on adsorption of elemental mercury vapors onto activated carbon. *Fuel* 111, 485–491. <https://doi.org/10.1016/j.fuel.2013.03.068>
- Khan, U.S., Amanullah, Manan, A., Khan, N., Mahmood, A., Rahim, A., 2015. Transformation mechanism of magnetite nanoparticles. *Mater. Sci. Pol.* 33, 278–285. <https://doi.org/10.1515/msp-2015-0037>
- Krishna, R.H., Swamy, A.V.V.S., 2012. Physico-Chemical Key Parameters, Langmuir and Freundlich isotherm and Lagergren Rate Constant Studies on the removal of divalent nickel from the aqueous solutions onto powder of calcined brick. *Int. J. Eng. Res. Dev.* 4, 2278–67.
- Lee, Y., Lee, J., Bae, C.J., Park, J.G., Noh, H.J., Park, J.H., Hyeon, T., 2005. Large-scale synthesis of uniform and crystalline magnetite nanoparticles using reverse micelles as nanoreactors under reflux conditions. *Adv. Funct. Mater.* 15, 503–509. <https://doi.org/10.1002/adfm.200400187>
- Liu, J., Wu, Z., Tian, Q., Wu, W., Xiao, X., 2016. Shape-controlled iron oxide nanocrystals: Synthesis, magnetic properties and energy conversion applications. *CrystEngComm* 18, 6303–6326. <https://doi.org/10.1039/c6ce01307d>
- Marusak, L., Walker, P., Mulay, L., 1976. The magnetokinetics of oxidation of pyrite (FeS₂). *IEEE Trans. Magn.* 12, 889–891.
- Milonjić, S.K., Kopečni, M.M., Ilić, Z.E., 1983. The point of zero charge and adsorption properties of natural magnetite. *J. Radioanal. Chem.* 78, 15–24. <https://doi.org/10.1007/BF02519745>
- Molino, A., Erto, A., Natale, F. Di, Donatelli, A., Iovane, P., Musmarra, D., 2013. Gasification of Granulated Scrap Tires for the Production of Syngas and a Low-Cost Adsorbent for Cd(II) Removal from Wastewaters. *Ind. Eng. Chem. Res.* 52, 12154–12160.
- Nassar, N.N., 2012. Iron Oxide Nanoadsorbents for Removal of Various Pollutants from Wastewater: An Overview. *Appl. Adsorbents Water Pollut. Control* 81–118. <https://doi.org/10.2174/978160805269111201010081>

- Nikiforov, V.N., Goldt, A.E., Gudilin, E.A., Sredin, V.G., Irhin, V.Y., 2014. Magnetic properties of maghemite nanoparticles. *Bull. Russ. Acad. Sci. Phys.* 78, 1075–1080. <https://doi.org/10.3103/S1062873814100141>
- Nishihara, K., Kondo, Y., 1959. Studies of the oxidation of Pyrite I. *Mem. Fac. Eng. Kyoto Univ.* 20, 285–306.
- Oliveira, L.C.A., Rios, R.V.R.A., Fabris, J.D., Garg, V., Sapag, K., Lago, R.M., 2002. Activated carbon/iron oxide magnetic composites for the adsorption of contaminants in water. *Carbon N. Y.* 40, 2177–2183. [https://doi.org/10.1016/S0008-6223\(02\)00076-3](https://doi.org/10.1016/S0008-6223(02)00076-3)
- Olugasa, T.T., Odesola, I.F., Oyewola, M.O., 2014. Energy production from biogas: A conceptual review for use in Nigeria. *Renew. Sustain. Energy Rev.* 32, 770–776. <https://doi.org/10.1016/j.rser.2013.12.013>
- Özdemir, Ö., Dunlop, D.J., 2014. Hysteresis and coercivity of hematite. *J. Geophys. Res. Solid Earth* 119, 2582–2594. <https://doi.org/10.1002/2014JB011376>. Received
- Patterson, A.L., 1939. The scherrer formula for X-ray particle size determination. *Phys. Rev.* 56, 978–982. <https://doi.org/10.1103/PhysRev.56.978>
- Preočanin, T., Stipić, F., Selmani, A., Kallay, N., 2011. Surface Potential of Polycrystalline Hematite in Aqueous Medium. *Int. J. Electrochem.* 2011, 1–6. <https://doi.org/10.4061/2011/412731>
- Roberts, R.J., Chen, M., 2006. Waste incineration - How big is the health risk? A quantitative method to allow comparison with other health risks. *J. Public Health (Bangkok)*. 28, 261–266. <https://doi.org/10.1093/pubmed/fdl037>
- Saeed, M., Ren, W., Wu, A., 2018. Therapeutic applications of iron oxide based nanoparticles in cancer: basic concepts and recent advances. *Biomater. Sci.* 6, 708–725.
- Scherrer, P., 1912. Determination of the internal structure and size of colloid particles using X-rays, in: *Colloid Chemistry A Textbook*. Springer, Berlin, Heidelberg. https://doi.org/10.1007/978-3-662-33915-2_7
- Shahid, M.K., Kim, Y., Choi, Y.-G., 2019. Adsorption of phosphate on magnetite-enriched particles (MEP) separated from the mill scale. *Front. Environ. Sci. Eng.* 13.
- Sousa, A.F. de, Braga, T.P., Gomes, E.C.C., Valentini, A., Longhinotti, E., 2012. Adsorption of phosphate using mesoporous spheres containing iron and aluminum oxide. *Chem. Eng. J.* 210, 143–149. <https://doi.org/10.1016/j.cej.2012.08.080>
- Teja, A.S., Koh, P.-Y., 2009. Synthesis, properties, and applications of magnetic iron oxide nanoparticles. *Prog. Cryst. Growth Charact. Mater.* 55, 22–45.
- Thorpe, A.N., Senftle, F.E., Alexander, C.C., Dulong, F.T., 1984. Oxidation of pyrite in coal to magnetite. *Fuel* 63, 662–668. [https://doi.org/10.1016/0016-2361\(84\)90163-7](https://doi.org/10.1016/0016-2361(84)90163-7)
- Tian, N., Tian, X., Ma, L., Yang, C., Wang, Y., Wang, Z., Zhang, L., 2015. Well-dispersed magnetic iron oxide nanocrystals on sepiolite nanofibers for arsenic removal. *RSC Adv.* 5, 25236–25243.
- Trazzi, P.A., Leahy, J.J., Hayes, M.H.B., Kwapinski, W., 2016. Adsorption and desorption of phosphate on biochars. *J. Environ. Chem. Eng.* 4, 37–46. <https://doi.org/10.1016/J.JECE.2015.11.005>
- Uggetti, E., Ferrer, I., Llorens, E., García, J., 2010. Sludge treatment wetlands: A review on the state of the art. *Bioresour. Technol.* 101, 2905–2912. <https://doi.org/10.1016/j.biortech.2009.11.102>

- Van, D.P., Fujiwara, T., Leu Tho, B., Song Toan, P.P., Hoang Minh, G., 2019. A review of anaerobic digestion systems for biodegradable waste: Configurations, operating parameters, and current trends. *Environ. Eng. Res.* 25, 1–17. <https://doi.org/10.4491/eer.2018.334>
- Vrieze, J. De, Smet, D., Klok, J., Colsen, J., Angenent, L.T., Vlaemnick, S.E., 2016. Thermophilic sludge digestion improves energy balance and nutrient recovery potential in full-scale municipal wastewater treatment plants. *Bioresour. Technol.* 218, 1237–1245.
- Wang, L., Pan, Y.X., Li, J.H., Qin, H.F., 2008. Magnetic properties related to thermal treatment of pyrite. *Sci. China, Ser. D Earth Sci.* 51, 1144–1153. <https://doi.org/10.1007/s11430-008-0083-7>
- Waters, K.E., Rowson, N.A., Greenwood, R.W., Williams, A.J., 2008. The effect of heat treatment on the magnetic properties of pyrite. *Miner. Eng.* 21, 679–682. <https://doi.org/10.1016/j.mineng.2008.01.008>
- Weller, D., McDaniel, T., 2006. Media for Extremely High Density Recording, in: Sellmyer, D., Skomski, R. (Eds.), *Advanced Magnetic Nanostructures*. Springer, Boston, MA.
- Willis, A.L., Turro, N.J., O'Brien, S., 2005. Spectroscopic characterization of the surface of iron oxide nanocrystals. *Chem. Mater.* 17, 5970–5975. <https://doi.org/10.1021/cm051370v>
- Wu, W., He, Q., Jiang, C., 2008. Magnetic iron oxide nanoparticles: Synthesis and surface functionalization strategies. *Nanoscale Res. Lett.* 3, 397–415. <https://doi.org/10.1007/s11671-008-9174-9>
- Xu, C., Akakuru, O.U., Zheng, J., Wu, A., 2019. Applications of iron oxide-based magnetic nanoparticles in the diagnosis and treatment of bacterial infections. *Front. Bioeng. Biotechnol.* 7, 1–15. <https://doi.org/10.3389/fbioe.2019.00141>
- Xu, J., Yang, H., Fu, W., Du, K., Sui, Y., Chen, J., Zeng, Y., Li, M., Zou, G., 2007. Preparation and magnetic properties of magnetite nanoparticles by sol-gel method. *J. Magn. Mater.* 309, 307–311. <https://doi.org/10.1016/j.jmmm.2006.07.037>
- Yang, G., Zhang, G., Wang, H., 2015. Current state of sludge production, management, treatment and disposal in China. *Water Res.* 78, 60–73. <https://doi.org/10.1016/j.watres.2015.04.002>
- Yang, Y., Chen, T., Sumona, M., Gupta, B. Sen, Sun, Y., Hu, Z., Zhan, X., 2017. Utilization of iron sulfides for wastewater treatment: a critical review. *Rev. Environ. Sci. Biotechnol.* 16, 289–308. <https://doi.org/10.1007/s11157-017-9432-3>
- Yoon, S.Y., Lee, C.G., Park, J.A., Kim, J.H., Kim, S.B., Lee, S.H., Choi, J.W., 2014. Kinetic, equilibrium and thermodynamic studies for phosphate adsorption to magnetic iron oxide nanoparticles. *Chem. Eng. J.* 236, 341–347. <https://doi.org/10.1016/j.cej.2013.09.053>

Chapter 6: Chemical Fate of Nutrients and Their Recovery Potential Through the Fe(III)-dosed Wastewater Treatment

6.1 Introduction

The previous studies have demonstrated feasibility of organics removal from wastewater using ferric iron, Fe(III) as an electron acceptor in an innovative anaerobic wastewater treatment (Ahmed et al., 2020; 2019). This has opened a new avenue for value creation through using iron containing wastes in managing wastewater. Given that iron is a redox active element and its wide range of biogeochemical reactions, this iron-dosed treatment method has also potential benefits of removing and recovering nutrients from wastewater. Nitrogen (N) and phosphorus (P) are critical pollutants to the environment and their excess loadings have caused widespread eutrophication in receiving waterbodies worldwide (Ærtebjerg et al., 2003; De Jonge et al., 2002). Excessive amount of N and P can cause serious impact on biodiversity, species composition and water as well as air quality (Olajire and Imeokparia, 2001; Wang et al., 2016).

Typically, wastewater contains N as primarily ammonium (NH_4^+) form with concentration ranging from 12 to 50 mg/L, and P as phosphate (PO_4^{3-}) form with concentration ranging from 4 to 12 mg/L (Metcalf & Eddy et al., 2014). Various chemical and biological methods have been established for nutrient removal from wastewater. Ammonium stripping and ion-exchange were the most common physiochemical methods for N removal, but have not gained much attention due to higher sludge production than biological method and low efficiency in N removal (Cooper et al., 1994; Lahav and Green, 2000). Biological nitrification/denitrification method has shown comparatively better ammonium removal efficiency. However, maintaining enough presence of organics in the denitrification step, recirculation of sewage sludge or addition of synthetic carbon is required, which makes it an expensive approach (Farazaki and Gikas, 2019; Reboleiro-Rivas et al., 2015). Application of soluble salts (e.g. aluminum sulfate, ferric chloride, calcium hydroxide) were found effective in P removal, but the approach is not feasible due to expensive chemical dosing and P-rich sludge management (Pratt et al., 2012). Enhanced biological phosphorous removal (EBPR) requires an additional anaerobic stage prior to the aeration tank of the wastewater treatment plant. In this process, selective strains of bacteria such

as *Acinetobacter* store large amount of P as polyphosphate within the bacteria cell. These bacteria are responsible for the high P content of EBPR sludge. Due to the fluctuating performance and high dependency on skilled operators, EBPR is also recognized as a complex process and unreliable at community level (Blackall et al., 2002; Bunce et al., 2018).

Nutrients can be recovered from wastewater and sewage sludge by various physiochemical, chemical and biological techniques which include ammonia stripping, ion exchange, bioelectrochemical systems, struvite precipitation, constructed wetlands, nanofiltration, and chemical reduction (Cornel and Schaum, 2009; Miles and Ellis, 2001; Sengupta et al., 2015). Some of these methods (e.g. struvite precipitation, ammonia stripping and adsorption, air scrubbing, membrane filtration) were used for nutrient recovery from anaerobic digestate to be used as fertilizers (Vaneckhaute et al., 2017). Using digestate for nutrient recovery has high potential in nutrient-rich regions, but, in most of the cases, they cannot ensure high quality end-product that can be used as fertilizers. Due to this reason, the use of recovered bio-based fertilizers are not encouraged by the environmental legislators. Moreover, these nutrient recovery techniques have several limitations which include scale formation for struvite precipitation, low recovery efficiency for ammonia adsorption and bioelectrochemical systems, and high operational costs for air stripping (Ahmed et al., 2015; Vaneckhaute et al., 2017). Thus, novel technologies with better applicability need to be explored for nutrient recovery from wastewater.

A few previous studies have investigated NH_4^+ oxidation under iron reducing condition by a bacterial-mediated process, in which ferric reduction is coupled to ammonium (NH_4^+) oxidation (denoted as Feammox). In this process, NH_4^+ was oxidized to either nitrite (NO_2^-), nitrate (NO_3^-) or nitrogen (N_2) when ferric, Fe(III) is reduced to ferrous, Fe(II) (Clément et al., 2005; Huang and Jaffé, 2015; Yang et al., 2012) (Fig. 27). Until now, most of the studies related to Feammox were reported in wetland soils (Li et al., 2015; Shrestha et al., 2009), tropical forest soil (Yang et al., 2012), paddy soil (Ding et al., 2014), and sediments (Yao et al., 2019). In the Fe(III)-dosed anaerobic biological treatment of wastewater, Feammox could be a microbial process for N removal. Such a nitrogen

removal mechanism is expected to be greatly affected by competition between heterotrophic iron reducing bacteria (IRB) and autotrophic Feammox bacteria. Heterotrophic IRB can outcompete Feammox bacteria for ferric iron and suppress the autotrophs when organics is not limited given that organic carbon is thermodynamically more favorable than NH_4^+ as an electron donor. This is supported by a previous study showing decreasing Feammox rate with increasing organic carbon in a soil system (Clément et al., 2005). In a previous study on riparian wetland soils, *Acidimicrobiaceae bacterium* A6, an autotrophic bacterium utilizing inorganic carbon as carbon source, was found to be the predominant bacterial species responsible for Feammox reaction with continuous NH_4^+ supply (Huang and Jaffé, 2018, 2015). Presence of *Geobacter*, a known IRB, was observed in these studies at the initial inoculation period, but their abundance gradually decreased without additional supply of organic carbon. In the Fe(III)-dosed anaerobic wastewater treatment, enough Fe^{3+} concentration would be a crucial factor so that NH_4^+ oxidation can occur after sufficient organics removal by heterotrophic IRB.

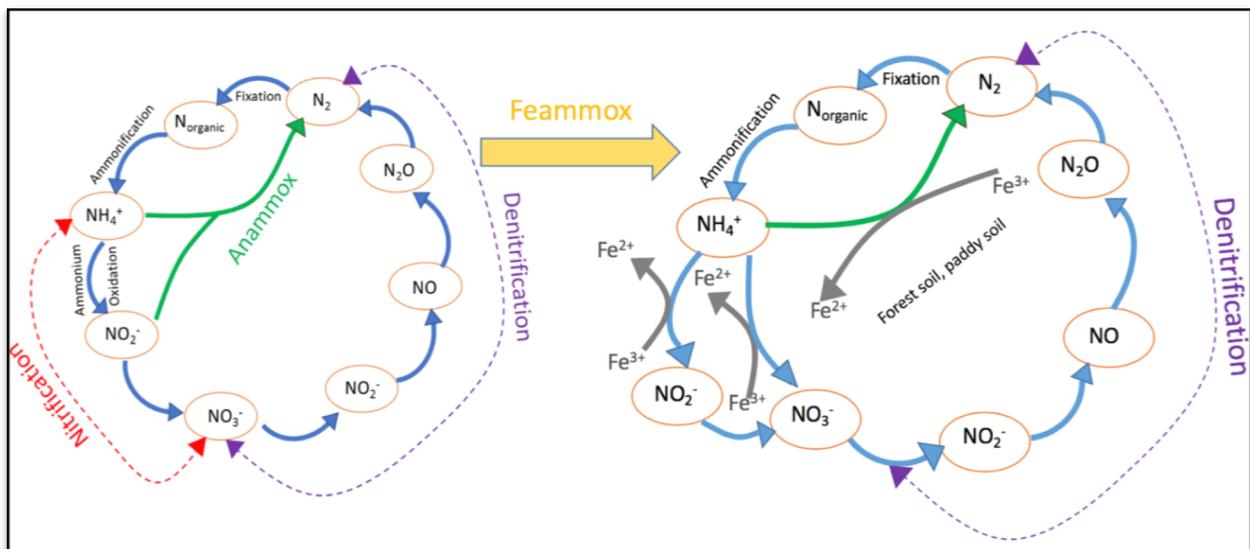


Fig. 27: Adaptation of the N- cycle in the Feammox reaction for NH_4^+ oxidation

Iron has commonly been used as a coagulant for P removal. Continuous ferric iron dosing in the bioreactor, P is expected to form FePO_4 precipitate as a mechanism of P removal from wastewater. A potential mechanism in the bioreactor is the formation of ferrous ammonium phosphate (FAP) given the presence of biogenic ferrous ion, ammonium, and

phosphate. FAP, an insoluble compound at circumneutral pH, is mainly used as food fortificant (Walczyk et al., 2013), and has the potential to use as a form of fertilizer given its nutrient content. The main objective of this study is to investigate the chemical fate of nutrients (N and P) along with organics and sulfate (SO_4^{2-}) transformations in the Fe(III)-dosed anaerobic bioreactor used for wastewater treatment. This study is very relevant to developing innovative and energy-efficient treatment option for nutrients and their recovery.

6.2 Material and Methods

6.2.1 Experimental Set-up

In this study, a 2.5 L bioreactor was used to treat a synthetic wastewater with continuous feeding of a ferric chloride solution. A 4-L tank was used as wastewater reservoir and a solution containing sodium acetate anhydrous ($\text{C}_2\text{H}_3\text{NaO}_2$, 3.2 mM), ethanol ($\text{C}_2\text{H}_6\text{O}$, 1.93 mM), lactose monohydrate ($\text{C}_{12}\text{H}_{22}\text{O}_{11}\cdot\text{H}_2\text{O}$, 0.36 mM), sodium sulfate (Na_2SO_4 , 0.55 M), sodium bicarbonate (NaHCO_3 , 1.67 mM), ammonium chloride (NH_4Cl , 3.74 mM), sodium phosphate monobasic dehydrate ($\text{NaH}_2\text{PO}_4\cdot 2\text{H}_2\text{O}$, 0.32 mM) and trace elements (4.75 mL/L) was prepared as the synthetic wastewater. The solution was prepared to maintain a specific concentration of COD (420 mg/L), SO_4^{2-} (50 mg/L), $\text{NH}_4^+\text{-N}$ (50 mg/L) and $\text{PO}_4^{3-}\text{-P}$ (10 mg/L). A 4-L tank containing a ferric chloride solution ($\text{FeCl}_3\cdot 6\text{H}_2\text{O}$, 10.25 mM) was used as a ferric source for the bioreactor and pH of the ferric solution was adjusted to 4-4.2 using a sodium hydroxide solution (NaOH , 5N). The influent and effluent samples were collected daily and later intermittently to measure the concentrations of NH_4^+ , COD, SO_4^{2-} , iron, P to examine the treatment performance. Additionally, microscopic analyses were performed to characterize the elemental composition of the produced sludge materials.

6.2.2 Analytical Methods

All the chemical analyses were done following the Standard Methods (APHA et al., 2005). Briefly, COD concentration was measured by following a closed reflux, colorimetric method (Standard Method 5220 D) using a spectrophotometer (HACH, DR 2800). Soluble sulfate concentration was measured by a turbidimetric method (USEPA method 375.4) using a spectrophotometer (Thermo Scientific, GENESYS 10UV). Total $\text{NH}_4^+\text{-N}$

concentration was measured using the Phenate method (4500 F). P concentrations in the filtrate samples were measured by Ascorbic Acid method (Standard Method 4500 E). Total iron concentration was measured using an atomic absorption spectrophotometer (Perkin Elmer 3100) after the samples were acidified with a 70% nitric acid (HNO₃) solution. Ferrous iron concentration was determined using 1, 10 phenanthroline method (Standard Method 3500 B) and ferric iron concentration was calculated by the difference between the total and ferrous iron concentrations.

6.2.3 Sludge Characterization

A scanning electron microscope (SEM) equipped with an energy dispersion spectroscopy (SEM-EDS, Hitachi S 4700) was used to examine the morphological and elemental composition of the sludge. The sludge samples were first dried in a closed desiccator filled with calcium sulfate and flushed with N₂ gas to prevent sludge oxidation. The SEM scanning was performed under an accelerating voltage of 10-20 kV and qualitative elemental analysis of the sludge samples was conducted by EDS spectrometry under an accelerating voltage of 10 kV. The powdered sludge samples were also used to determine the chemical states of Fe, N and P to investigate the presence of FAP by X-ray photoelectron spectroscopy (XPS) analysis (PHI 5000 Versaprobe). The sample powder was mounted on a sample holder with a zero reflective quartz plate (MTI corporation, CA) located underneath. XPS spectra were obtained with a monochromatized Al K α X-ray source (1487 eV) while base pressure of the analytical chamber was on the order of 10⁻⁷ Pa. A pass energy of 23.5 eV was used to conduct elemental scans of Fe, N and P.

6.3 Results and Discussion

6.3.1 Treatment Performance

The water quality analyses on the influent and effluent samples showed fairly consistent removal of organics, N (as ammonium), P (as phosphate) and SO₄²⁻ from the wastewater (Fig. 28). Average removal efficiencies of COD, PO₄³⁻-P, SO₄²⁻ and NH₄⁺-N was 97 ± 2%, 99.7 ± 0.5%, 87.1 ± 3% and 20.3 ± 9.0%, respectively. The high removal efficiency of COD is attributed to organics oxidation coupled to ferric reduction as well as sulfate reduction. Approximate 87% sulfate reduction and 89% iron removal indicated that both ferric iron and sulfate reduction played a significant role in COD oxidation. Significant P

removal was achieved due to precipitation as FePO_4 and FAP. Removal of N may be associated with the Feammox process or FAP precipitation. Mean total iron and ferrous (Fe^{2+}) loading of the effluent samples were 21 ± 9 mg/d and 6 ± 4 mg/d, respectively. The residual ferrous iron in the effluent represents an oxygen demand and can be oxidized by chlorine in a disinfection unit. There was not any significant presence of nitrate and nitrite in the effluent.

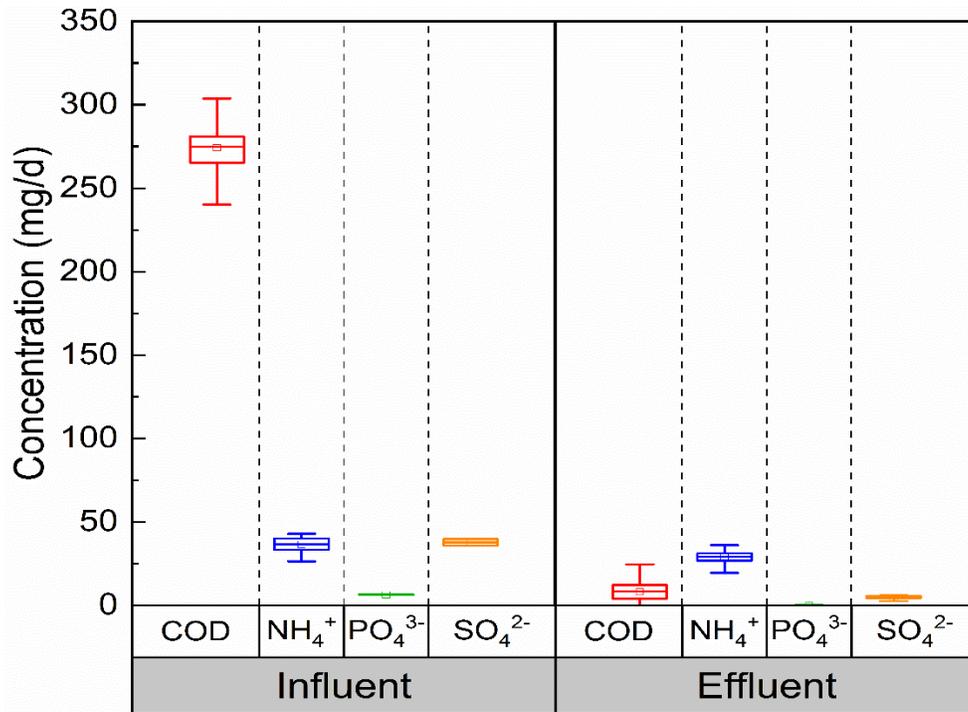


Fig. 28: Influent and effluent concentrations of COD, NH_4^+ -N, PO_4^{3-} -P and SO_4^{2-}

6.3.2 Sludge Characterization

The SEM-EDS analysis showed major elements of the sludge sample as C, O, N, Fe, and S (Fig. 29). The precipitate particles have a very disordered morphology with no specific pattern. Particle sizes were similar to structural descriptions of iron sulfide (≈ 1 -2 μm), FAP (2-6 μm), and ferric phosphate (1-2 μm) presented in previous studies (Bratkova et al., 2018; Csákberényi-Malasics et al., 2012; Li et al., 2014; Vaclavkova et al., 2014; Zhang et al., 2013). The identification of different compounds was not possible with the SEM-EDS analysis.

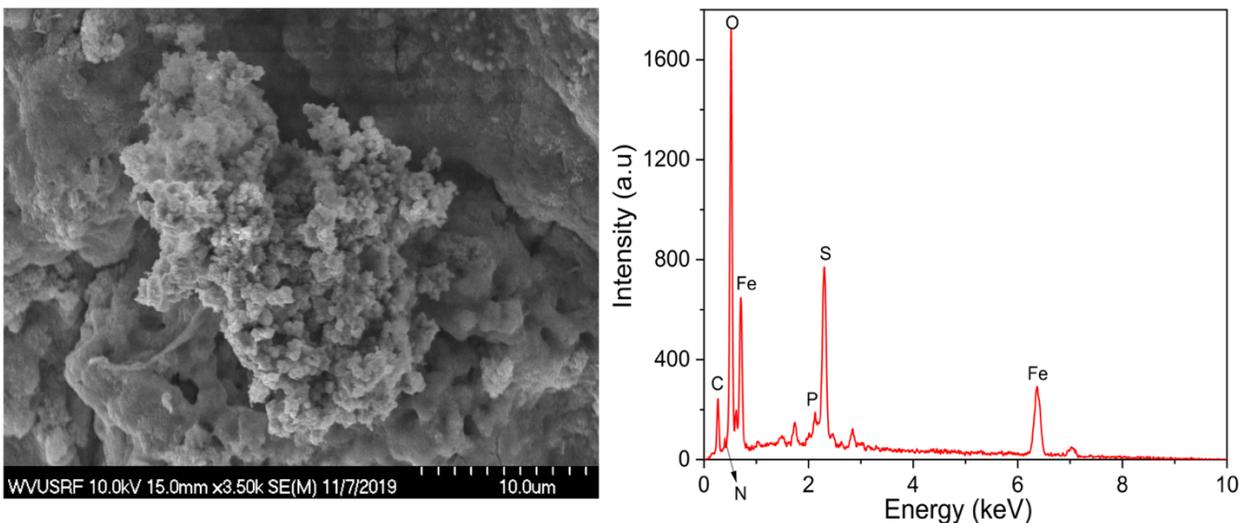


Fig. 29: SEM micrograph and EDS spectrum of the sludge material

To investigate the presence of FAP in the sludge materials, elemental scan of Fe, N and P was conducted using XPS. The narrow region in the spectrum of Fe $2p_{3/2}$ revealed two major peaks at 708.5 eV and 710.5 eV (Fig. 30a), which represent Fe $^{2+}$ and Fe $^{3+}$, respectively (Deng and Lin, 2017). The N1s spectrum was fit with one significant peak at 400.2 eV (Fig. 30b) which represents ammonium (NH $_4^+$) phase and P2 $p_{3/2}$ spectrum was fit well with a peak at 133.1 eV (Fig. 30c) which represents phosphate (PO $_4^{3-}$) phase (Naumkin et al., 2012). Although, there was no specific binding energy of FAP reported in the literature, the presence of individual peaks of Fe $^{2+}$, NH $_4^+$ and PO $_4^{3-}$ suggest probable presence of FAP in the sludge sample.

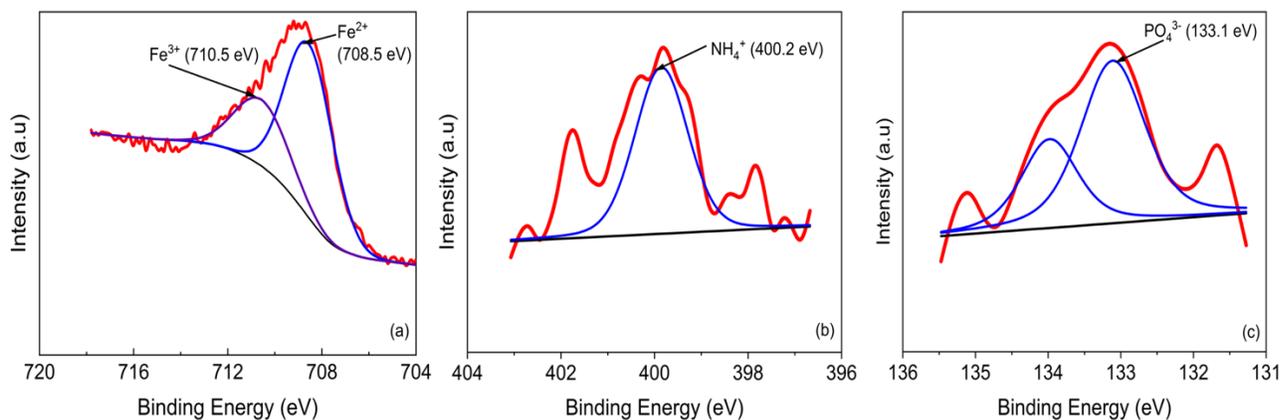


Fig. 30: XPS spectra of the sludge sample for (a) Fe2p_{3/2}, (b) N1s, and (c) P2p_{3/2}

6.4 Conclusion

This study investigated the opportunity of removing and recovering nutrients in the newly developed Fe(III)-dosed anaerobic wastewater treatment system. This treatment method was found to effectively remove organics and phosphorus, and to a significant degree, N from wastewater in the same bioreactor. That reduces the necessity of multiple treatment units as in the existing technologies for N and P removal. For practical implementation of this treatment method, the iron dosing scheme will be a crucial factor to provide sufficient electron acceptor for organics and ammonium oxidation, as well as for ferrous ammonium phosphate precipitation. The beneficial applications of FAP, especially as a fertilizer, has created a new avenue of resource recovery from the sludge that would greatly enhance the sustainability of this treatment method. Future studies related to the microbiological composition and metabolic pathways responsible for the chemical transformations will give better insights into the biogeochemical reactions that facilitate the treatment.

References

- Ærtebjerg, G., Andersen, J. H., A., Hansen, O.S., 2003. Nutrients and Eutrophication in Danish Marine Waters, Ministry of the Environment, Danish Environmental protection Agency & National Environmental Research Institute. <https://doi.org/10.1146/annurev.nutr.25.050304.092633>
- Ahmed, M., Hasan, C.K., Rahman, H., Ali Hossain, M., Uddin, S.A., 2015. Prospects of using wastewater as a resource-nutrient recovery and energy generation. *Am. J. Environ. Sci.* <https://doi.org/10.3844/ajessp.2015.99.114>
- Ahmed, M., Lin, O., Saup, C.M., Wilkins, M.J., Lin, L.S., 2019. Effects of Fe/S ratio on the

- kinetics and microbial ecology of an Fe(III)-dosed anaerobic wastewater treatment system. *J. Hazard. Mater.* 369, 593–600. <https://doi.org/10.1016/j.jhazmat.2019.02.062>
- Ahmed, M., Saup, C.M., Wilkins, M.J., Lin, L.-S., 2020. Continuous ferric iron-dosed anaerobic wastewater treatment: treatment performance, sludge characteristics, and microbial composition. *J. Environ. Chem. Eng.* 8.
- APHA, AWWA, WEF, 2005. Standard methods for the examination of water and wastewater, 21st ed. American Public Health Association, American Water Works Association, and Water Environment Federation.
- Blackall, L.L., Crocetti, G.R., Saunders, A.M., Bond, P.L., 2002. A review and update of the microbiology of enhanced biological phosphorus removal in wastewater treatment plants. *Antonie van Leeuwenhoek, Int. J. Gen. Mol. Microbiol.* 81, 681–691. <https://doi.org/10.1023/A:1020538429009>
- Bratkova, S., Lavrova, S., Angelov, A., Nikolova, K., Ivanov, R., Koumanova, B., 2018. Treatment of wastewaters containing Fe, Cu, Zn and as by microbial hydrogen sulfide and subsequent removal of COD, N and P. *J. Chem. Technol. Metall.* 53, 245–257.
- Bunce, J.T., Ndam, E., Ofiteru, I.D., Moore, A., Graham, D.W., 2018. A review of phosphorus removal technologies and their applicability to small-scale domestic wastewater treatment systems. *Front. Environ. Sci.* 6, 1–15. <https://doi.org/10.3389/fenvs.2018.00008>
- Clément, J.C., Shrestha, J., Ehrenfeld, J.G., Jaffé, P.R., 2005. Ammonium oxidation coupled to dissimilatory reduction of iron under anaerobic conditions in wetland soils. *Soil Biol. Biochem.* 37, 2323–2328. <https://doi.org/10.1016/j.soilbio.2005.03.027>
- Cooper, P., Day, M., Thomas, V., 1994. Process Options for Phosphorus and Nitrogen Removal from Wastewater. *Water Environ. J.* 8, 84–92. <https://doi.org/10.1111/j.1747-6593.1994.tb01096.x>
- Cornel, P., Schaum, C., 2009. Phosphorus recovery from wastewater: Needs, technologies and costs. *Water Sci. Technol.* 59, 1069–1076. <https://doi.org/10.2166/wst.2009.045>
- Csákberényi-Malasics, D., Rodriguez-Blanco, J.D., Kis, V.K., Rečnik, A., Benning, L.G., Pósfai, M., 2012. Structural properties and transformations of precipitated FeS. *Chem. Geol.* 294–295, 249–258. <https://doi.org/10.1016/j.chemgeo.2011.12.009>
- De Jonge, V.N., Elliott, M., Orive, E., 2002. Causes, historical development, effects and future challenges of a common environmental problem: Eutrophication. *Hydrobiologia* 475–476, 1–19. <https://doi.org/10.1023/A:1020366418295>
- Deng, D., Lin, L.S., 2017. Continuous sulfidogenic wastewater treatment with iron sulfide sludge oxidation and recycle. *Water Res.* 114, 210–217. <https://doi.org/10.1016/j.watres.2017.02.048>
- Ding, L.J., An, X.L., Li, S., Zhang, G.L., Zhu, Y.G., 2014. Nitrogen loss through anaerobic ammonium oxidation coupled to iron reduction from paddy soils in a

- chronosequence. *Environ. Sci. Technol.* 48, 10641–10647. <https://doi.org/10.1021/es503113s>
- Farazaki, M., Gikas, P., 2019. Nitrification-denitrification of municipal wastewater without recirculation, using encapsulated microorganisms. *J. Environ. Manage.* 242, 258–265. <https://doi.org/10.1016/j.jenvman.2019.04.054>
- Huang, S., Jaffé, P.R., 2018. Isolation and characterization of an ammonium-oxidizing iron reducer: *Acidimicrobiaceae* sp. A6. *PLoS One* 13, 1–12. [https://doi.org/10.1016/s0378-1097\(97\)00409-6](https://doi.org/10.1016/s0378-1097(97)00409-6)
- Huang, S., Jaffé, P.R., 2015. Characterization of incubation experiments and development of an enrichment culture capable of ammonium oxidation under iron-reducing conditions. *Biogeosciences* 12, 769–779. <https://doi.org/10.5194/bg-12-769-2015>
- Lahav, O., Green, M., 2000. Ammonium removal from primary and secondary effluents using a bioregenerated ion-exchange process. *Water Sci. Technol.* 42, 179–185. <https://doi.org/10.2166/wst.2000.0311>
- Li, P., Du, Y., Li, L., Huang, L., Rudolph, V., Nguyen, A. V, Xu, Z.P., 2014. Preparation and characterisation of manganese and iron compounds as potential control-release foliar fertilizers. *Biointerface Res. Appl. Chem.* 4, 746–753. <https://doi.org/10.1021/ed017p596.1>
- Li, X., Hou, L., Liu, M., Zheng, Y., Yin, G., Lin, X., Cheng, L., Li, Y., Hu, X., 2015. Evidence of Nitrogen Loss from Anaerobic Ammonium Oxidation Coupled with Ferric Iron Reduction in an Intertidal Wetland. *Environ. Sci. Technol.* 49, 11560–11568. <https://doi.org/10.1021/acs.est.5b03419>
- Metcalf & Eddy, I., Tchobanoglous, G., Stensel, H.D., Tsuchihashi, R., Burton, F., 2014. *Wastewater Engineering: Treatment and Resource Recovery*. McGraw-Hill, New York.
- Miles, A., Ellis, T.G., 2001. Struvite precipitation potential for nutrient recovery from anaerobically treated wastes. *Water Sci. Technol.* 43, 259–266. <https://doi.org/10.2166/wst.2001.0690>
- Naumkin, A. V., Kraut-Vass, A., Gaarenstroom, S.W., Powell, C.J., 2012. NIST X-ray Photoelectron Spectroscopy Database [WWW Document]. <https://doi.org/http://dx.doi.org/10.18434/T4T88K>
- Olajire, A.A., Imeokparia, F.E., 2001. Water quality assessment of Osun river: Studies on inorganic nutrients. *Environ. Monit. Assess.* 69, 17–28. <https://doi.org/10.1023/A:1010796410829>
- Pratt, C., Parsons, S.A., Soares, A., Martin, B.D., 2012. Biologically and chemically mediated adsorption and precipitation of phosphorus from wastewater. *Curr. Opin. Biotechnol.* 23, 890–896. <https://doi.org/10.1016/j.copbio.2012.07.003>
- Reboleiro-Rivas, P., Martín-Pascual, J., Juárez-Jiménez, B., Poyatos, J.M., Vílchez-Vargas, R., Vlaeminck, S.E., Rodelas, B., González-López, J., 2015. Nitrogen

- removal in a moving bed membrane bioreactor for municipal sewage treatment: Community differentiation in attached biofilm and suspended biomass. *Chem. Eng. J.* 277, 209–218. <https://doi.org/10.1016/j.cej.2015.04.141>
- Sengupta, S., Nawaz, T., Beaudry, J., 2015. Nitrogen and Phosphorus Recovery from Wastewater. *Curr. Pollut. Reports* 1, 155–166. <https://doi.org/10.1007/s40726-015-0013-1>
- Shrestha, J., Rich, J.J., Ehrenfeld, J.G., Jaffe, P.R., 2009. Oxidation of ammonium to nitrite under iron-reducing conditions in wetland soils: Laboratory, field demonstrations, and push-pull rate determination. *Soil Sci.* 174, 156–164. <https://doi.org/10.1097/SS.0b013e3181988fbf>
- Vaclavkova, S., Jørgensen, C.J., Jacobsen, O.S., Aamand, J., Elberling, B., 2014. The Importance of Microbial Iron Sulfide Oxidation for Nitrate Depletion in Anoxic Danish Sediments. *Aquat. Geochemistry* 20, 419–435. <https://doi.org/10.1007/s10498-014-9227-x>
- Vaneekhaute, C., Lebuf, V., Michels, E., Belia, E., Vanrolleghem, P.A., Tack, F.M.G., Meers, E., 2017. Nutrient Recovery from Digestate: Systematic Technology Review and Product Classification. *Waste and Biomass Valorization* 8, 21–40. <https://doi.org/10.1007/s12649-016-9642-x>
- Walczyk, T., Kastenmayer, P., Storcksdieck Genannt Bonsmann, S., Zeder, C., Grathwohl, D., Hurrell, R.F., 2013. Ferrous ammonium phosphate (FeNH_4PO_4) as a new food fortificant: Iron bioavailability compared to ferrous sulfate and ferric pyrophosphate from an instant milk drink. *Eur. J. Nutr.* 52, 1361–1368. <https://doi.org/10.1007/s00394-012-0445-y>
- Wang, J., Pan, F., Soininen, J., Heino, J., Shen, J., 2016. Nutrient enrichment modifies temperature-biodiversity relationships in large-scale field experiments. *Nat. Commun.* 7, 1–9. <https://doi.org/10.1038/ncomms13960>
- Yang, W.H., Weber, K.A., Silver, W.L., 2012. Nitrogen loss from soil through anaerobic ammonium oxidation coupled to iron reduction. *Nat. Geosci.* 5, 538–541. <https://doi.org/10.1038/ngeo1530>
- Yao, Z., Wang, F., Wang, C., Xu, H., Jiang, H., 2019. Anaerobic ammonium oxidation coupled to ferric iron reduction in the sediment of a eutrophic lake. *Environ. Sci. Pollut. Res.* 1–11. <https://doi.org/10.1007/s11356-019-04907-7>
- Zhang, T.B., Lu, Y.C., Luo, G.S., 2013. Iron phosphate prepared by coupling precipitation and aging: Morphology, crystal structure, and Cr(III) adsorption. *Cryst. Growth Des.* 13, 1099–1109. <https://doi.org/10.1021/cg301441t>

Chapter 7: Conclusions

An innovative Fe(III)-dosed anaerobic wastewater treatment method has been developed in this Ph.D. research by incorporating Fe and S biogeochemical reactions in a laboratory-scale engineered biological treatment system. Overall, this novel treatment method has multiple benefits over the typical aerobic treatment technologies including potential use of iron containing wastes, no need for aeration, significant organics and nutrient removal, and recovery of nutrients and useful sludge byproducts.

Based on a thorough literature review, several critical factors related to applications of ferric reduction in developing such a Fe(III)-dosed treatment system were identified and research objectives were designed accordingly to develop this treatment method. Types of ferric compounds, organics type and concentrations, microorganisms, and ferric bioavailability are identified as the major factors affecting organic oxidation coupled to iron reduction. In this research, typical wastewater constituents (acetate, lactate, ethanol) and conditions (pH, temperature, alkalinity) were used to prepare synthetic wastewaters, which mimicked the real municipal wastewater. Commonly, ferric compounds are precipitated as ferric hydroxide ($\text{Fe}(\text{OH})_3$) in the environment, therefore, $\text{Fe}(\text{OH})_3$ was used as ferric compound. As an ubiquitous constituent of domestic wastewater, sulfate was also considered as an electron acceptor responsible for organics oxidation.

Consistent high removal of organics in this Fe(III)-dosed treatment process indicates the efficient functionality of this process for practical applications. Nutrient removal and potential recovery in one-stage treatment not only reduce the requirement of complex treatment train for full-scale applications but also improve the sustainability of this system. Generation of valuable magnetic particles through a simple thermal treatment method reduces the burden of sludge management and creates opportunities of beneficial applications. Microbiological composition obtained in this research has provided a baseline for the types of microorganisms and their interrelations occurred in this treatment system, which was not described previously. Due to substantial advantages of energy efficiency, excellent pollutant removal, and innovative resource recovery opportunities, this Fe(III)-dosed treatment can be a viable alternative to existing aerobic treatment

technologies. Broad applications of this technology are expected to lead to more sustainable wastewater management and create opportunities for utilizing iron-containing wastes.

Future studies are warranted to further develop this treatment technology in the following areas:

- Additional experiments can be designed and conducted by dosing various ferric compounds (ferric hydroxide, different forms of ferric oxides) to evaluate their effects on the treatment performance as well as sludge production and composition. Additionally, effects of environmental conditions (pH, temperature) need to be investigated.
- A comprehensive sludge treatment method can be developed for production of sludge byproducts with targeted properties (e.g., size, magnetization) and separation of nutrient products (e.g., FAP, vivianite ($\text{Fe}_3(\text{PO}_4)_2 \cdot 8\text{H}_2\text{O}$)).
- Reusing the sludge or the sludge byproducts within the treatment process (e.g., coagulant, iron recycling) can create opportunities for more intensified treatment process (i.e., process intensification). The scope of reusing sludge and its byproducts within the system warrants further investigations.
- Prospects of other metabolic pathways such as Feammox need to be investigated under different iron dosing schemes through comprehensive chemical and microbiological studies on its transformations and chemical fate.
- Iron reducing bacteria such as *Geobacter* and *Shewanella* sp. are observed to generate the highest electrical current density among exoelectrogenic bacteria, given their diverse and efficient electron transfer mechanisms. They can be used in bioelectrochemical systems for electricity generation from wastewater and/or sludge to further enhance energy efficiency of the iron-dosed treatment approach.