International Journal of Molecular Sciences
MDPI

Article

Dairy-inspired coatings for bone implants from whey protein isolate-derived self-assembled fibrils

Rebecca Rabe [1], Ute Hempel [2], Laurine Martocq [3,*], Julia K. Keppler [1,4], Jenny Aveyard [5] and Timothy E.L. Douglas [3,6]

[1] Division of Food Technology, Kiel University, Germany; RebeccaRabe1@gmx.de; julia.keppler@wur.nl
[2] Institute of Physiological Chemistry, Technische Universität Dresden, Germany; hempel-u@mail.zih.tu-dresden.de
[3] Engineering Department, Lancaster University, United Kingdom; l.martocq@lancaster.ac.uk; t.douglas@lancaster.ac.uk
[4] Laboratory of Food Process Engineering, Wageningen University & Research AFSG, The Netherlands
[5] School of Engineering, University of Liverpool, United Kingdom; zippy78@liverpool.ac.uk
[6] Materials Science Institute (MSI), Lancaster University, United Kingdom
* Correspondence: l.martocq@lancaster.ac.uk

Received: date; Accepted: date; Published: date

Abstract: To improve integration of a biomaterial with surrounding tissue, its surface properties may be modified by adsorption of biomacromolecules, e.g. fibrils. Whey protein isolate (WPI), a dairy industry by-product, supports osteoblastic cell growth. WPI's main component, β-lactoglobulin, forms fibrils in acidic solutions. In this study, aiming to develop coatings for biomaterials for bone contact, substrates were coated with WPI fibrils obtained at pH 2 or 3.5. Importantly, WPI fibrils coatings withstood autoclave sterilization and appeared to promote human bone marrow stromal cells (hBMSC) spreading and differentiation. In the future, WPI fibrils coatings could facilitate immobilization of biomolecules with growth stimulating or antimicrobial properties.

Keywords: coating, stem cell, whey protein isolate, bone, fibril

1. Introduction

Whey protein isolate (WPI) is a dairy industry by-product which contains > 95% protein, of which 75% is β-lactoglobulin [1]. Previous studies showed that WPI enhances cell proliferation and osteogenic differentiation and displays antibacterial properties [1–3]. Upon heating under acidic conditions, β-lactoglobulin degrades into smaller peptides which undergo self-assembly to form fibrils several micrometers long and a few nm thick [4], whose morphology is pH-dependent (long semi-flexible fibers at pH 2, worm-like structures at pH 3.5) [5–7].

Better cell-biomaterial interactions, and biomaterial integration into host tissue can be achieved by improving surface properties, e.g. by coatings. Fibrillar coatings have advantages including high surface/volume ratio, promoting fibril adhesion to substrates. Biologically active molecules can be immobilized on fibrils [8,9], which can form aligned superstructure scaffolds [10], improve cell line attachment and act as biomimetic cell culture platforms [11–13].

A commonly used fibrillar molecule used as implant coating materials is collagen, which is known to promote cell adhesion, spreading and proliferation [14–18]. Fibronectin is another commonly used molecule to improve cell adhesion, also in fibrillar form [19,20]. One advantage of WPI is its low cost, as it is a by-product of the dairy industry.

With the intention of developing coatings for biomaterials for bone contact, WPI fibrillar coatings were formed to support and enhance spreading, attachment and differentiation of hBMSC, which have greater clinical relevance than cell lines. WPI fibrils were hypothesized to withstand

autoclaving as WPI hydrogels do [21]. Autoclaving was preferred due to its ubiquity, clinical acceptance and low cost.

WPI fibrils were formed in solutions at two different pH values, 2 and 3.5. A WPI solution concentration of 2.5 wt% was used. WPI concentrations between 2.5 % and 5 % WPI were found to result in a high fibril yield [3,4], and especially 2.5 % WPI is often used in WPI or beta-Lactoglobulin fibril studies due to the lower sample viscosity which improves handling. Because the protein concentration also affects the aggregation kinetics and morphology of the resulting aggregates [22–24], deviations from these ideal values can also reduce the comparability to other studies. and then adsorbed onto substrates and imaged by Scanning Electron Microscopy (SEM). Finally, autoclaved coatings were characterized using hBMSC.

2. Results and discussion

WPI fibril formation (shown schematically in Figure 1a) was influenced by pH, as measurements of fibrillar yield at pH 2 (approximately 25%) and pH 3.5 (> 40%) demonstrated (Figure 1b). Similar observations were reported previously [5–7]. Differences in yield are attributed to differences in the fibril building blocks, which are specific acid hydrolysed peptides at pH 2 [4], but unspecific non-hydrolyzed proteins at pH 3.5 [5].

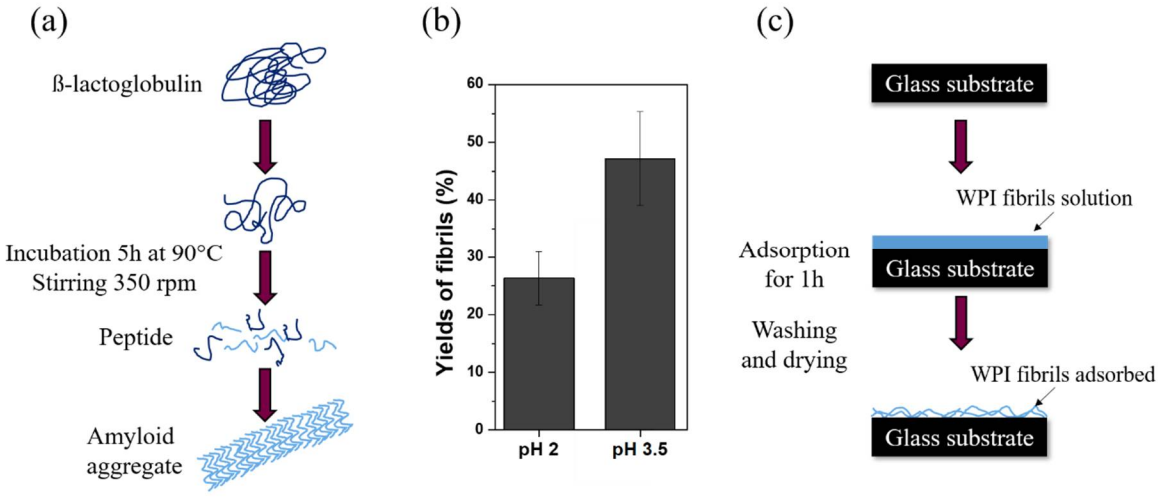

Figure 1. (a) Process of fibrils formation at pH 2 in solution; at pH 2, β-lactoglobulin denatures and hydrolyses at 90°C. Specific peptides self-associate into the amyloid aggregates, which can consist of approximately three intertwined protofibrils. At pH 3.5, acid hydrolysis is reduced; therefore non-hydrolyzed β-lactoglobulin assembles into worm-like aggregates, which are not amyloid but amyloid-like and of different shape and morphology than at pH. (b) Fibrillar yield in solutions of different pH and (c) adsorption of WPI fibrils on glass substrates.

Contact angle (CA) measurements demonstrated a significant increase from 20 to approximately 55° on both coated sample types (Figure 2a). Similar increases were reported for edible WPI fibril coatings on fruits [25]. Advantageously for cell growth, CA remains lower than 100° [26]. A hydrophilic surface is beneficial for cell adhesion to make sure that proteins from cell culture medium adsorb to the surface in the desired conformation, so that binding sites on the proteins are recognized by the cells. Furthermore, a lower contact angle and higher wettability would facilitate coating of rough or porous biomaterial surfaces.

SEM images confirmed that fibril coatings withstood washing and drying. Fibrils prepared at pH 3.5 (Figure 2c) appeared to be shorter and less straight than those formed at pH 2 (Figure 2b), in agreement with previous studies [5]. Fibrils were detected by SEM after autoclaving (supplementary

information, figure S1); hence, they withstand sterilization. Adhesion of the fibrils to substrates would hinder fibril aggregation and degradation as in previous studies [27,28]; hence the coating is estimated to be one fibril thick.

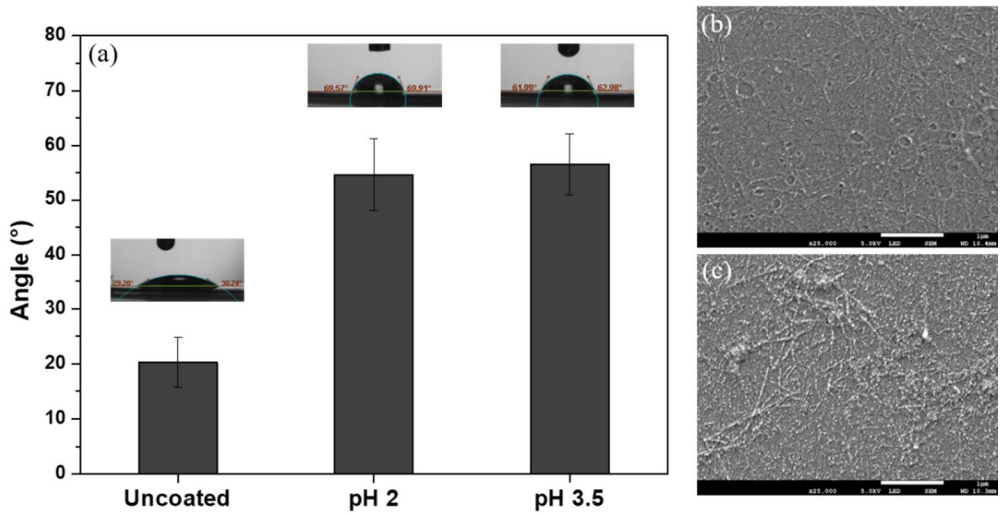

Figure 2. (a) CA measurements of uncoated and fibrillar coated samples with solution at pH 2 and pH 3.5 and SEM images of fibrillar coatings obtained at (b) pH 2 and (c) pH 3.5 (scale bar: 1 μm)

Adhesion of hBMSC was confirmed on the uncoated (Figure 3a) and coated samples (Figure 3b,c). Spreading was superior and tissue non-specific alkaline phosphatase (TNAP) activity (a marker of osteogenic differentiation and linked to the calcium deposition as shown previously [9,29]) was higher on coated samples (Figure 3g). The cells became confluent over the WPI coatings.

hBMSC on coated samples showed 2 h after seeding clear focal adhesion contacts and a well-organized cytoskeleton. A possible reason for that could be that diverse proteins from the serum and cellular in situ-formed proteins adsorbed on the WPI layer and thus promoted initial adhesion. The cell number on coated samples at day 2 and day 4 after seeding (MTS formazan formation is a measure of cell number and can be used as an index of cell proliferation) did not differ substantially from that on glass.

For hBMSC, proper adhesion, spreading and re-organization of cytoskeleton is an essential prerequisite for proliferation and differentiation of the cells. The fibrillar coating promoted the adhesion and re-organization of the cytoskeleton of hBMSC, did not influence the number of adherent cells, but obviously improved their "quality" as evidenced by higher TNAP activity.

Hence, we suspect that the coatings induced a superior start of the differentiation program of cells. Analysis concerning the molecular mechanisms are planned for future studies. In the present study, we wished to ascertain whether such a fibrillar coating has advantages for hBMSC adhesion and promotes osteogenic parameters.

Apparently differences in yields and morphologies of straight fibrils observed at pH 2 and the worm-like aggregates observed at pH 3.5 do not affect hBMSC. hBMSC metabolic activity increased slightly from day 2 to 4 (Figure 3h,i), but not significantly.

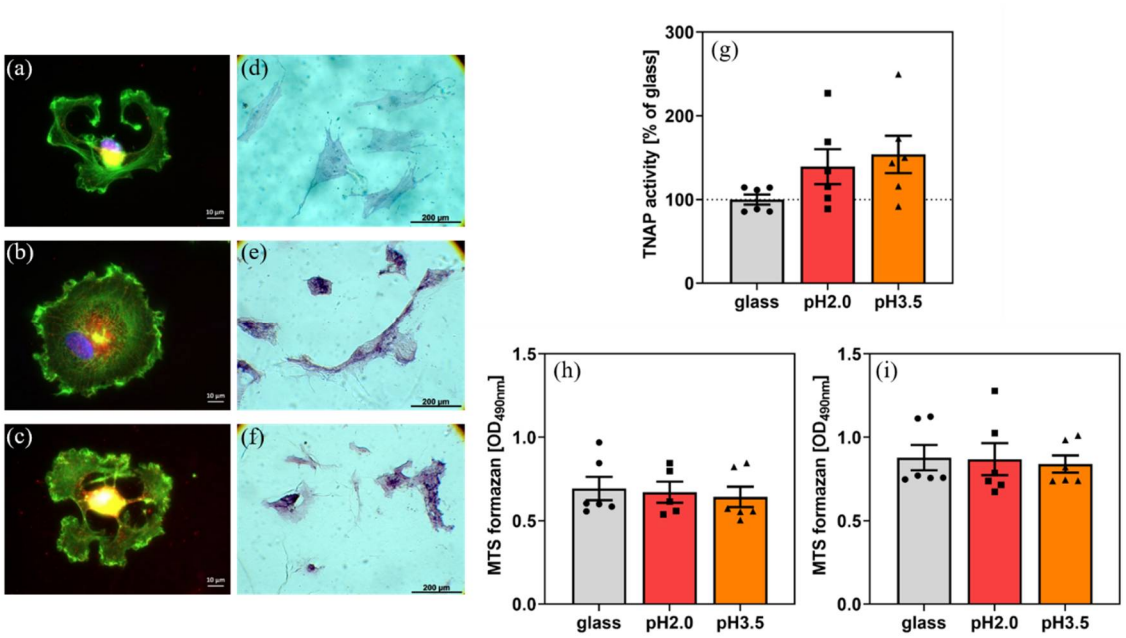

Figure 3. (a) Morphology of hBMSC on (a) glass, (b) fibrillar coating (pH=2), (c) fibrillar coating (pH=3.5), 2 h after plating, and (d), (e), (f) TNAP staining at day 11 after plating, respectively. (g) TNAP activity on different substrates (day 11) and metabolic activity results at (h) day 2 and (i) day 4 after plating.

Future work will focus on preparation of coatings from fibrils obtained at other pH values. It is well known that the typical fibrils with amyloid structure only occur at pH 2, while worm-like aggregates can be observed at pH 3.5 [6]. Spherical aggregates emerge at pH 4 to 5 [7] and otherwise there are also smaller aggregates observed at neutral pH. Besides pH-induced changes, the addition of sodium chloride or the protein concentration can affect the morphology, as well as addition of solvents [22,30]. The structures have different yields (i.e., portion of amyloid to non-assembled material) but also different processing stabilities. Thus, there is a whole range of conditions that can be used to alter the morphology and to study the correlation between structure and cell behaviour in the future.

Another focus of future work will be extension of the cell biological characterization of the coatings to elucidate the exact mechanism by which coatings may promote differentiation and to include cell-induced mineralization. From previous investigations, it is known that an increase of TNAP activity leads to an increase in released phosphate ions into the conditioned medium and in consequence to enhanced mineralization [9], which should be studied in future.

WPI fibrillar coatings can be enhanced by incorporation of molecules with growth-stimulatory or antimicrobial properties into the coatings, coating thickness and mechanical measurements and substrates more appropriate for bone contact (e.g. titanium alloy). Furthermore, WPI fibrillar coatings should be compared to more commonly used fibrillar coatings of fibronectin and collagen, which are known to promote cell adhesion [14–20].

4. Materials and Methods

WPI (BiPro, Davisco Foods International Inc., USA) was dissolved in Milli-Q (2.5 wt%). pH was set to 2 and 3.5 by adding HCl. 15 mL WPI solution was heated (90°C. 5h, stirring speed 350 rpm) to induce fibril formation resulting in a fibrillar suspension. Glass substrates (chosen as an inexpensive substrate for this pilot study) were coated with fibrils by adsorption from the suspension. Substrates were rinsed with Milli-Q to remove non-adhered fibrils, air-dried and autoclaved (121°C, 15 minutes). SEM, CA measurements and fibrillar yield quantification were performed as described previously [5,7,31,32].

144 hBMSC were isolated from bone marrow aspirates from donors (males, average 27 ± 5 yrs.) who
145 gave full informed consent (local ethics commission (ethic vote No. EK466112016)), at the Bone
146 Marrow Transplantation Center, University Hospital Dresden, characterized and plated onto
147 samples (5,555 hBMSC/cm^2) as described previously [29,33].
148 After 2h, hBMSC morphology was monitored by fluorescence staining of F-actin and
149 phosphorylated focal adhesion kinase, as described previously [8]. Metabolic activity of hBMSC was
150 determined by the standard MTS assay (Cell Titer96 AQueous One Solution Proliferation Assay)
151 (Promega, Mannheim, Germany). Cell culture medium was replaced by fresh medium containing
152 10% of MTS dye solution. After incubation in a humidified CO_2 incubator (2h, 37°C), 80 µL medium
153 was removed and absorbance was measured photometrically at 490 nm.
154 At day 11 after seeding hBMSC were stained for tissue non-specific alkaline phosphatase
155 (TNAP) enzyme activity with a commercial staining kit (86-R, Sigma). Images were obtained and
156 TNAP enzyme activity was determined, as described previously [8].
157 Experiments were performed with cells from three different donors (n=3) each in triplicate.
158 Results are presented as mean ± standard error of the mean. Statistical significance was analysed with
159 GraphPad Prism 8.4 software (Statcon, Witzenhausen, Germany) by ANOVA analysis with
160 Bonferroni's post-test.
161

162 ## 5. Conclusions

163 Formation of WPI fibrils in solution was strongly pH-dependant; fibrillar yield increased when
164 pH was increased from 2 to 3.5. WPI fibrillar coatings resisted autoclave sterilization and supported
165 the attachment, spreading and differentiation of hBMSC. The pH 2 and pH 3.5 fibrils had an equally
166 positive effect on cell differentiation.

167 **Supplementary Materials:** Supplementary materials can be found at www.mdpi.com/xxx/s1.

168 **Author Contributions:** Conceptualization, T.E.L.D. and J.K.K.; methodology, T.E.L.D., J.K.K., U.H. and J.A.;
169 formal analysis, R.R., U.H., J.A. and T.E.L.D.; investigation, R.R., U.H., J.A. and T.E.L.D.; resources, J.K.K., U.H.
170 and T.E.L.D.; data curation, R.R., U.H. and J.A.; writing—original draft preparation, L.M., J.K.K. and T.E.L.D.;
171 writing—review and editing, L.M. and T.E.L.D.; visualization, U.H., R.R. and T.E.L.D.; supervision, T.E.L.D. and
172 J.K.K.; project administration, T.E.L.D.; funding acquisition, T.E.L.D. and L.M. All authors have read and agreed
173 to the published version of the manuscript.

174 **Funding:** This research was funded by N8 Agrifood "Food2Bone" (T.E.L.D.), and EPSRC "A novel coating
175 technology based upon polyatomic ions from plasma" grant number EP/S004505/1 (L.M.).

176 **Conflicts of Interest:** The authors declare no conflict of interest. The funders had no role in the design of the
177 study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to
178 publish the results.

179 ## Abbreviations

CA	Contact angle
hBMSC	Human bone marrow stromal cells
SEM	Scanning electron microscopy
TNAP	Tissue non-specific alkaline phosphatase
WPI	Whey protein isolate

180 ## References

181 1. Douglas, T.E.L.; Vandrovcová, M.; Kročilová, N.; Keppler, J.K.; Zárubová, J.; Skirtach, A.G.; Bačáková,
182 L. Application of whey protein isolate in bone regeneration: Effects on growth and osteogenic
183 differentiation of bone-forming cells. *J. Dairy Sci.* **2018**, *101*, 28–36, doi:10.3168/jds.2017-13119.
184 2. Gupta, D.; Kocot, M.; Tryba, A.M.; Serafim, A.; Stancu, I.C.; Jaegermann, Z.; Pamuła, E.; Reilly, G.C.;
185 Douglas, T.E.L. Novel naturally derived whey protein isolate and aragonite biocomposite hydrogels

have potential for bone regeneration. *Mater. Des.* **2020**, *188*, 108408, doi:10.1016/j.matdes.2019.108408.

3. Keppler, J.K.; Martin, D.; Garamus, V.M.; Berton-Carabin, C.; Nipoti, E.; Coenye, T.; Schwarz, K. Functionality of whey proteins covalently modified by allyl isothiocyanate. Part 1 physicochemical and antibacterial properties of native and modified whey proteins at pH 2 to 7. *Food Hydrocoll.* **2017**, *65*, 130–143, doi:10.1016/j.foodhyd.2016.11.016.
4. Akkermans, C.; Venema, P.; van der Goot, A.J.; Gruppen, H.; Bakx, E.J.; Boom, R.M.; van der Linden, E. Peptides are building blocks of heat-induced fibrillar protein aggregates of β-lactoglobulin formed at pH 2. *Biomacromolecules* **2008**, *9*, 1474–1479, doi:10.1021/bm7014224.
5. Heyn, T.R.; Garamus, V.M.; Neumann, H.R.; Uttinger, M.J.; Guckeisen, T.; Heuer, M.; Selhuber-Unkel, C.; Peukert, W.; Keppler, J.K. Influence of the polydispersity of pH 2 and pH 3.5 beta-lactoglobulin amyloid fibril solutions on analytical methods. *Eur. Polym. J.* **2019**, *120*, doi:10.1016/j.eurpolymj.2019.08.038.
6. Keppler, J.K.; Heyn, T.R.; Meissner, P.M.; Schrader, K.; Schwarz, K. Protein oxidation during temperature-induced amyloid aggregation of beta-lactoglobulin. *Food Chem.* **2019**, *289*, 223–231, doi:10.1016/j.foodchem.2019.02.114.
7. Serfert, Y.; Lamprecht, C.; Tan, C.P.; Keppler, J.K.; Appel, E.; Rossier-Miranda, F.J.; Schroen, K.; Boom, R.M.; Gorb, S.; Selhuber-Unkel, C.; et al. Characterisation and use of β-lactoglobulin fibrils for microencapsulation of lipophilic ingredients and oxidative stability thereof. *J. Food Eng.* **2014**, *143*, 53–61, doi:10.1016/j.jfoodeng.2014.06.026.
8. Hempel, U.; Preissler, C.; Vogel, S.; Möller, S.; Hintze, V.; Becher, J.; Schnabelrauch, M.; Rauner, M.; Hofbauer, L.C.; Dieter, P. Artificial extracellular matrices with oversulfated glycosaminoglycan derivatives promote the differentiation of osteoblast-precursor cells and premature osteoblasts. *Biomed Res. Int.* **2014**, *2014*, doi:10.1155/2014/938368.
9. Hempel, U.; Matthäus, C.; Preissler, C.; Möller, S.; Hintze, V.; Dieter, P. Artificial matrices with high-sulfated glycosaminoglycans and collagen are anti-inflammatory and pro-osteogenic for human mesenchymal stromal cells. *J. Cell. Biochem.* **2014**, *115*, 1561–1571, doi:10.1002/jcb.24814.
10. Knowles, T.P.J.; Oppenheim, T.W.; Buell, A.K.; Chirgadze, D.Y.; Welland, M.E. Nanostructured films from hierarchical self-assembly of amyloidogenic proteins. *Nat. Nanotechnol.* **2010**, *5*, 204–207, doi:10.1038/nnano.2010.26.
11. Reynolds, N.P.; Styan, K.E.; Easton, C.D.; Li, Y.; Waddington, L.; Lara, C.; Forsythe, J.S.; Mezzenga, R.; Hartley, P.G.; Muir, B.W. Nanotopographic surfaces with defined surface chemistries from amyloid fibril networks can control cell attachment. *Biomacromolecules* **2013**, *14*, 2305–2316, doi:10.1021/bm400430t.
12. Reynolds, N.P.; Charnley, M.; Mezzenga, R.; Hartley, P.G. Engineered lysozyme amyloid fibril networks support cellular growth and spreading. *Biomacromolecules* **2014**, *15*, 599–608, doi:10.1021/bm401646x.
13. Reynolds, N.P.; Charnley, M.; Bongiovanni, M.N.; Hartley, P.G.; Gras, S.L. Biomimetic topography and chemistry control cell attachment to amyloid fibrils. *Biomacromolecules* **2015**, *16*, 1556–1565, doi:10.1021/acs.biomac.5b00114.
14. Douglas, T.; Heinemann, S.; Mietrach, C.; Hempel, U.; Bierbaum, S.; Scharnweber, D.; Worch, H. Interactions of collagen types I and II with chondroitin sulfates A-C and their effect on osteoblast adhesion. *Biomacromolecules* **2007**, *8*, 1085–1092, doi:10.1021/bm0609644.
15. Geißler, U.; Hempel, U.; Wolf, C.; Scharnweber, D.; Worch, H.; Wenzel, K.W. Collagen type I-coating of Ti6A14V promotes adhesion of osteoblasts. *J. Biomed. Mater. Res.* **2000**, *51*, 752–760, doi:10.1002/1097-

4636(20000915)51:4<752::AID-JBM25>3.0.CO;2-7.

16. Vandrovcova, M.; Douglas, T.E.L.; Heinemann, S.; Scharnweber, D.; Dubruel, P.; Bacakova, L. Collagen-lactoferrin fibrillar coatings enhance osteoblast proliferation and differentiation. *J. Biomed. Mater. Res. - Part A* **2015**, *103*, 525–533, doi:10.1002/jbm.a.35199.
17. Kim, H.-W.; Li, L.-H.; Lee, E.-J.; Lee, S.-H.; Kim, H.-E. Fibrillar assembly and stability of collagen coating on titanium for improved osteoblast responses. *J. Biomed. Mater. Res. Part A* **2005**, *75A*, 629–638, doi:10.1002/jbm.a.30463.
18. Douglas, T.; Hempel, U.; Mietrach, C.; Viola, M.; Vigetti, D.; Heinemann, S.; Bierbaum, S.; Scharnweber, D.; Worch, H. Influence of collagen-fibril-based coatings containing decorin and biglycan on osteoblast behavior. *J. Biomed. Mater. Res. Part A* **2008**, *84A*, 805–816, doi:10.1002/jbm.a.31501.
19. Franke, K.; Pompe, T.; Bornhäuser, M.; Werner, C. Engineered matrix coatings to modulate the adhesion of CD133+ human hematopoietic progenitor cells. *Biomaterials* **2007**, *28*, 836–843, doi:10.1016/j.biomaterials.2006.09.031.
20. Gui, L.; Wojciechowski, K.; Gildner, C.D.; Nedelkovska, H.; Hocking, D.C. Identification of the heparin-binding determinants within fibronectin repeat III1: Role in cell spreading and growth. *J. Biol. Chem.* **2006**, *281*, 34816–34825, doi:10.1074/jbc.M608611200.
21. Dziadek, M.; Douglas, T.E.L.; Dziadek, K.; Zagrajczuk, B.; Serafim, A.; Stancu, I.C.; Cholewa-Kowalska, K. Novel whey protein isolate-based highly porous scaffolds modified with therapeutic ion-releasing bioactive glasses. *Mater. Lett.* **2020**, *261*, 127115, doi:10.1016/j.matlet.2019.127115.
22. Heyn, T.R.; Mayer, J.; Neumann, H.R.; Selhuber-Unkel, C.; Kwade, A.; Schwarz, K.; Keppler, J.K. The threshold of amyloid aggregation of beta-lactoglobulin: Relevant factor combinations. *J. Food Eng.* **2020**, *283*, 110005, doi:10.1016/j.jfoodeng.2020.110005.
23. Pellarin, R.; Caflisch, A. Interpreting the Aggregation Kinetics of Amyloid Peptides. *J. Mol. Biol.* **2006**, *360*, 882–892, doi:10.1016/j.jmb.2006.05.033.
24. Ye, X.; Hedenqvist, M.S.; Langton, M.; Lendel, C. On the role of peptide hydrolysis for fibrillation kinetics and amyloid fibril morphology. *RSC Adv.* **2018**, *8*, 6915–6924, doi:10.1039/c7ra10981d.
25. Feng, Z.; Wu, G.; Liu, C.; Li, D.; Jiang, B.; Zhang, X. Edible coating based on whey protein isolate nanofibrils for antioxidation and inhibition of product browning. *Food Hydrocoll.* **2018**, *79*, 179–188, doi:10.1016/j.foodhyd.2017.12.028.
26. Bacakova, L.; Filova, E.; Parizek, M.; Ruml, T.; Svorcik, V. Modulation of cell adhesion, proliferation and differentiation on materials designed for body implants. *Biotechnol. Adv.* 2011, *29*, 739–767.
27. Gilbert, J.; Campanella, O.; Jones, O.G. Electrostatic stabilization of β-lactoglobulin fibrils at increased pH with cationic polymers. *Biomacromolecules* **2014**, *15*, 3119–3127, doi:10.1021/bm500762u.
28. Li, C.; Born, A.K.; Schweizer, T.; Zenobi-Wong, M.; Cerruti, M.; Mezzenga, R. Amyloid-hydroxyapatite bone biomimetic composites. *Adv. Mater.* **2014**, *26*, 3207–3212, doi:10.1002/adma.201306198.
29. Hempel, U.; Müller, K.; Preissler, C.; Noack, C.; Boxberger, S.; Dieter, P.; Bornhäuser, M.; Wobus, M. Human Bone Marrow Stromal Cells: A Reliable, Challenging Tool for in Vitro Osteogenesis and Bone Tissue Engineering Approaches. *Stem Cells Int.* **2016**, *2016*, doi:10.1155/2016/7842191.
30. Kayser, J.J.; Arnold, P.; Steffen-Heins, A.; Schwarz, K.; Keppler, J.K. Functional ethanol-induced fibrils: Influence of solvents and temperature on amyloid-like aggregation of beta-lactoglobulin. *J. Food Eng.* **2020**, *270*, 109764, doi:10.1016/j.jfoodeng.2019.109764.
31. Douglas, T.E.L.; Hempel, U.; Żydek, J.; Vladescu, A.; Pietryga, K.; Kaeswurm, J.A.H.; Buchweitz, M.; Surmenev, R.A.; Surmeneva, M.A.; Cotrut, C.M.; et al. Pectin coatings on titanium alloy scaffolds

produced by additive manufacturing: Promotion of human bone marrow stromal cell proliferation. *Mater. Lett.* **2018**, *227*, 225–228, doi:10.1016/j.matlet.2018.05.060.

32. Li, M.; Aveyard, J.; Fleming, G.; Curran, J.M.; McBride, F.; Raval, R.; D'Sa, R.A. Nitric Oxide Releasing Titanium Surfaces for Antimicrobial Bone-Integrating Orthopedic Implants. *ACS Appl. Mater. Interfaces* **2020**, *12*, doi:10.1021/acsami.0c00871.

33. Oswald, J.; Boxberger, S.; Joergensen, B.; Bornhaeuser, M.; Ehninger, G.; Werner, C. Mesenchymal Stem Cells (MSC) can be differentiated into endothelial cells in vitro. In Proceedings of the Transactions - 7th World Biomaterials Congress; John Wiley & Sons, Ltd, 2004; Vol. 22, p. 506.