

Influence of type 2 diabetes on muscle deoxygenation during ramp incremental cycle exercise

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The published article is available from doi:
<https://doi.org/10.1016/j.resp.2019.103258>

1 **TITLE**

2 Influence of type 2 diabetes on muscle deoxygenation during ramp incremental cycle exercise

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23 **ABSTRACT**

24 We tested the hypothesis that type 2 diabetes (T2D) alters the profile of muscle fractional
25 oxygen (O₂) extraction (near-infrared spectroscopy) during incremental cycle exercise.
26 Seventeen middle-aged individuals with uncomplicated T2D and 17 controls performed an
27 upright ramp test to exhaustion. The rate of muscle deoxygenation (i.e. deoxygenated
28 haemoglobin and myoglobin concentration, $\Delta[\text{HHb}+\text{Mb}]$) profiles of the vastus lateralis muscle
29 were normalised to 100% of the response, plotted against % power output (PO) and fitted with
30 a double linear regression model. Peak oxygen uptake was significantly ($P<0.05$) reduced in
31 individuals with T2D. The $\% \Delta[\text{HHb}+\text{Mb}]/\% \text{PO}$ slope of the first linear segment of the double
32 linear regression function was significantly ($P<0.05$) steeper in T2D than controls (1.81 ± 0.61
33 vs 1.35 ± 0.43). Both groups displayed a near-plateau in $\Delta[\text{HHb}+\text{Mb}]$ at an exercise intensity
34 (**%PO) not different among them**. Such findings suggest that a reduced O₂ delivery to active
35 muscles is an important underlying cause of exercise intolerance during a maximum graded test
36 in middle-aged individuals with T2D.

37

38 **Keywords:** near-infrared spectroscopy, oxygen extraction, cycling, exercise tolerance, type 2
39 diabetes

40

41 **1. Introduction**

42 Individuals with uncomplicated type 2 diabetes mellitus (T2D) demonstrate impairments in
43 peak exercise capacity ($\dot{V}O_{2\text{peak}}$), an established clinical predictor of cardiovascular and all-
44 cause mortality (Kodama et al., 2009; Swift et al., 2013), in the region of 20% (Baldi et al.,
45 2003; Kiely et al., 2015; Mac Ananey et al., 2011; O'Connor et al., 2015; O'Connor et al., 2012;
46 Regensteiner et al., 1998). Importantly, this impairment is independent of obesity and age, and
47 present in the absence of clinically apparent cardiovascular disease (Green et al., 2015). Whilst
48 the precise mechanisms for this diminished exercise capacity remain to be elucidated, it is likely
49 the consequence of a complex array of pathophysiological changes at a central and/or peripheral
50 level (Green et al., 2015; Poitras et al., 2018). Maximum $\dot{V}O_2$, representative of the integration
51 of the pulmonary, cardiovascular and muscular systems to uptake, transport and utilise O_2
52 respectively, is governed by the oxygen cascade from the environment to the muscle
53 mitochondria (Poole, 1997; Wagner et al., 1997), and is thus, consequent to the product of
54 whole-body perfusive and diffusive O_2 conductance. However, **most commonly**, the Fick
55 relationship is determined **either at the pulmonary level, or across the exercising limb(s)**, and is
56 representative of **pooled fractional O_2 extraction across multiple compartments which** may not
57 necessarily reflect the discrete adjustments of O_2 exchange within the microvasculature of the
58 active muscle (Iannetta et al., 2017; Okushima et al., 2016; Spencer et al., 2012). As such,
59 considering the **matching of O_2 delivery (QO_2)-to- $\dot{V}O_2$ and diffusive O_2 conductance** at the
60 level of the active muscle vasculature during exercise is of great relevance when exploring the
61 mechanistic bases for the decreased exercise tolerance observed in T2D.

62

63 Substantial evidence exists to suggest that peripheral O_2 delivery in the lower limbs is impaired
64 in individuals with uncomplicated T2D. For instance, the maximum leg haemodynamic and
65 vasodilatory responses during an incremental calf plantar-flexion exercise (Kiely et al., 2014)
66 as well as steady-state femoral artery blood flow measurements during cycling (Kingwell et al.,

67 2003) and knee extension exercise (Lalande et al., 2008) are reduced in men and women with
68 uncomplicated T2D. Additionally, leg vascular conductance kinetics at the onset of heavy-
69 intensity plantar-flexion exercise (Kiely et al., 2014; MacAnaney et al., 2011), and quadriceps
70 muscle microvascular blood flow kinetics during moderate cycling (Bauer et al., 2007) are
71 impaired (i.e. slowed/blunted) in individuals with T2D free from cardiovascular disease. In
72 contrast, Poitras et al. (2015) recently reported unaffected leg blood flow kinetics during knee
73 extension/flexion exercise in individuals with T2D; although participants had a more advanced
74 diabetes and history of cardiovascular disease, with their control group also having a similar
75 history of cardiovascular disease/comorbidities (Poitras et al., 2015). In agreement with Poitras
76 et al. (2015), Copp et al. (2010) found that locomotory muscle(s) blood flow during running
77 was not decreased in the rat GK model of type 2 diabetes (Copp et al., 2010) despite grossly
78 impaired microvascular perfusion at rest (Padilla et al., 2006).

79

80 It is therefore plausible that the maldistribution of active muscle blood flow in individuals with
81 uncomplicated T2D (Kiely et al., 2014; MacAnaney et al., 2011), and subsequently a decreased
82 microvascular partial pressure of O₂ (P_{mvo_2}) (Padilla et al., 2007), would mandate an increased
83 reliance on fractional O₂ extraction in the exercising muscle in an effort to achieve a given
84 increase in $\dot{V}O_2$. The use of near-infrared spectroscopy (NIRS) during exercise permits a non-
85 invasive assessment of microvascular O₂ extraction (DeLorey et al., 2003). By measuring the
86 concentration changes in deoxygenated haemoglobin and myoglobin ($\Delta[HHb+Mb]$), an
87 estimate of fractional O₂ extraction is possible. NIRS, therefore, provides insights into the
88 dynamic balance between regional QO_2 and $\dot{V}O_2$ at the level of the microvasculature (Spencer
89 et al., 2012), the determining factor for P_{mvo_2} . Accordingly, investigating the dynamic response
90 of $[HHb+Mb]$ within the microcirculation of the exercising muscles during a ramp incremental
91 test may offer insight into pathophysiological mechanisms potentially implicated in the reduced
92 exercise capacity in T2D. In the present study the profile of $\% \Delta[HHb+Mb]$ during a ramp

93 incremental test was characterized using a function including two linear segments; the ‘double-
94 linear model’ (Vieth, 1989) as it has been proffered to best characterise this profile (Spencer et
95 al., 2012). In the first segment, a linear increase in $\% \Delta[\text{HHb}+\text{Mb}]$ relative to changes in work
96 rate occurs, representing the increasing reliance on O_2 extraction relative to metabolic demand.
97 This culminates at a ‘breakpoint’ ($\Delta[\text{HHb}+\text{Mb}] - \text{BP}$), from which a “plateau-like” response
98 ensues despite the continued increase in work rate. The breakbpoint has been associated with
99 transitions in exercise intensity domains between heavy to severe-intensity exercise (Bellotti et
100 al., 2013; Keir et al., 2015). This plateau in the $[\text{HHb}+\text{Mb}]$ signal does not indicate the upper
101 limit of O_2 extraction during incremental tests, and it seems to be connected to the re-
102 distribution of blood flow towards the active tissues once this upper boundary of exercise is
103 achieved (Inglis et al., 2017).

104

105 The aim of the present study was to explore the influence of T2D on the profile of local muscle
106 fractional O_2 extraction, as indicated by the NIRS-derived $\Delta[\text{HHb}+\text{Mb}]$ response. We
107 hypothesized that individuals with T2D would display an accelerated muscle deoxygenation
108 response throughout the ramp incremental exercise bout. This would be depicted by a steeper
109 primary slope of the double linear equation, thereby signifying an increased dependence on O_2
110 extraction for providing adequate $\dot{V}\text{O}_2$ at a given work rate. To avoid the potential effects of
111 aging on the T2D-related impairments on exercise tolerance previously established in men
112 (O'Connor et al., 2015; Wilkerson et al., 2011) we limited the age of participants to < 55 yr.

113

114 **2. Methods**

115 *2.1. Participants*

116 Thirty four individuals, 17 with uncomplicated T2D (12 males, 5 females), and 17 age- and
117 BMI-matched controls (ND) (12 males, 5 females) volunteered to participate in this study. The
118 age range of all participants was between 36 and 55 yr. (Table 1). Participants in the control

119 group (ND) were recruited from the general population, whilst participants with T2D were
120 recruited from the diabetes outpatient clinics of St. Columcille's Hospital (Loughlinstown, Co.
121 Dublin) and St. Vincent's University Hospital (SVUH, Dublin 4), following chart review. Five
122 female participants were premenopausal (2 T2D, 3 ND) and 5 were postmenopausal (3 T2D, 2
123 ND) not undergoing hormone replacement therapy. All participants were non-smokers and had
124 not smoked during the 12-month period preceding the study. Individuals with T2D had a
125 clinical history of diabetes of between 2 to 9.5 years, with adequately controlled HbA_{1c} levels
126 (<10%) (Table 1) and were not taking insulin or beta-blockers. Two of the controls were on
127 prescriptive medications (statins, $n = 2$), and with the exception of one participant, all
128 participants with T2D were taking oral ($n = 15$) and/or subcutaneous ($n = 1$) hypoglycaemic
129 prescription medications (metformin monotherapy, $n = 9$; metformin & sulphonylurea, $n = 3$;
130 metformin & thiazolidinedione, $n = 1$; glucagon-like peptide 1, $n = 1$; sodium glucose
131 cotransporter 2 inhibitors, $n = 4$). In addition, a subgroup of individuals with T2D were taking
132 antihypertensive prescription drugs (angiotensin converting enzyme inhibitor, $n = 4$;
133 angiotensin II receptor blocker, $n = 2$; calcium channel blocker, $n = 5$) and statins ($n = 6$).

134

135 At the commencement of the present study, individuals with T2D displayed no clinical evidence
136 of ischemic heart disease (normal ECG during treadmill stress test following the Bruce
137 protocol), peripheral arterial disease ($0.9 < \text{ABI} < 1.3$), kidney dysfunction (urine protein <
138 200mg/dl), or liver dysfunction (urine creatinine levels < 2.2 mg/dl). Participants were
139 classified as physically inactive by self-report (≤ 1.5 h.week⁻¹ of moderate-intensity exercise in
140 the preceding 6 months), which was confirmed by the use of 5-day RT3 triaxial accelerometry
141 (Stayhealthy Inc, CA) in a subset of participants (Table 1) (Rowlands et al., 2004). All
142 participants provided written informed consent before commencement, and the study was
143 approved by the Faculty of Health Sciences' Research Ethics Committee, Trinity College

144 Dublin, and St Vincent's Healthcare Ethics and Medical Research Committee, and conducted
145 in accordance with the Declaration of Helsinki (2008).

146

147 2.2. Study Protocol

148 2.2.1. Overview.

149 Following a satisfactory completion of the 12-lead ECG stress test, participants were tested on
150 one occasion either at St. Columcille's Hospital or the cardiovascular laboratory in Trinity
151 College Dublin. Premenopausal participants were tested during the mid-follicular phase (days
152 5-12) of the menstrual cycle. All participants refrained from consuming alcohol, caffeine and
153 non-prescribed nutritional supplements in the 24 hours prior to testing and constrained their
154 exercise to normal activities of daily living. All participants performed a **ramp incremental**
155 cycling test to exhaustion to determine $\dot{V}O_{2peak}$.

156

157 2.2.2. *Ramp incremental cycling tests to exhaustion.* The **ramp incremental** cycling test to
158 exhaustion was performed in an upright position on an electrically braked cycle ergometer
159 (Excalibur Sport; Lode B.V., Groningen, The Netherlands). Exercise was performed at an initial
160 workload of 10 W for 2 min. This was followed by 10-15 W.min⁻¹ increments in PO in females
161 or 10-25 W.min⁻¹ increments in males (depending on **stated** activity levels), until volitional
162 exhaustion. Pedal frequency was held constant at an individually selected cadence between 60-
163 75 revolutions per minute (rpm). Failure in a test was determined as a drop in cadence exceeding
164 10 rpm for >5 s. Peak workload was determined according to the point of termination of the
165 test. $\dot{V}O_{2peak}$ was determined by identifying the highest 15-s mean $\dot{V}O_2$ value recorded before
166 the participant's volitional termination of the test. The ventilatory threshold (VT) was
167 determined as the exercise level at which $\dot{V}_E/\dot{V}O_2$ exhibited a systematic exponential increase
168 without a concomitant increase in $\dot{V}_E/\dot{V}CO_2$ (Wasserman et al., 1973), and the deflection point
169 of carbon dioxide output ($\dot{V}CO_2$) versus O₂ uptake ($\dot{V}O_2$; V-slope method) (Amann et al., 2006;

170 Beaver et al., 1986). The respiratory compensation point (RCP) was estimated by identifying
171 the second non-linear increase of \dot{V}_E and $\dot{V}CO_2$, whereby an increase in $\dot{V}_E/\dot{V}O_2$ was
172 accompanied by an increase of $\dot{V}_E/\dot{V}CO_2$ (Wasserman and McIlroy, 1964).

173

174 2.3. Measurements

175 During exercise participants wore a facemask to continuously collect expired air using an online
176 metabolic system (Innocor, Innovision A/S, Odense, Denmark). Analysis of expired air allowed
177 determination of pulmonary O_2 uptake ($\dot{V}O_2$), carbon dioxide output ($\dot{V}CO_2$), minute
178 ventilation (\dot{V}_E) and the respiratory exchange ratio (RER) breath by breath. Heart rate was
179 recorded every 5 s (Polar S610i, Polar Ltd, Finland), with peak HR defined as the highest heart
180 rate attained within the last 15 s of the point of termination of the test. Beat-to-beat systolic and
181 diastolic blood pressure was continuously monitored throughout the exercise protocol using the
182 volume clamp method at the level of the finger (Finometer, Finepress Medical Systems B.V.
183 the Netherlands). MAP was calculated from systolic and diastolic pressures (MAP: 0.33
184 systolic BP + 0.66 diastolic BP). Peak BP was expressed as the highest 15-second mean
185 pressure obtained before the participant's volitional termination of the test.

186

187 A continuous wave NIRS system (Hamamatsu Niro 200Nx; Hamamatsu Photonics,
188 Hamamatsu, Japan), was used to non-invasively determine the oxygenation status of the right
189 quadriceps' *vastus lateralis* (VL) muscle. This was determined using the spatially resolved
190 spectroscopy (SRS) technique and modified Beer-Lambert (MBL) principle with three
191 wavelengths of emitting light ($\lambda = 735, 810, \text{ and } 850 \text{ nm}$). The theoretical basis of NIRS and its
192 use in exercise measurements have been described in detail elsewhere (Ferrari et al., 2011).
193 **Briefly**, this technique estimates the optical density changes of deoxygenated haemoglobin and
194 myoglobin (HHb+Mb) based on the O_2 dependency of absorption changes for near-infrared
195 light in these proteins. As the VL muscle is a dominant locomotor muscle during cycling

196 (Laplaud et al., 2006), the present study examined the Δ [HHb+Mb] profiles of the right VL
197 muscle. After shaving the skin, the probes were placed on the belly of the muscle (5-8 cm above
198 the lateral femoral condyle), parallel to the major axis of the thigh with a 3 cm spacing between
199 the emitter and receiver. The probes were housed in a black rubber holder and secured on the
200 skin surface with bi-adhesive tape and then covered with a dark elastic bandage, which
201 minimised extraneous movement and the intrusion of stray light throughout the exercise
202 protocol. Since the depth of the measured area is estimated to be between one-half and one-
203 third of the distance between the emitter and the receiver (~1.5 cm) (Ferrari et al., 2004; Van
204 Beekvelt et al., 2001), the thickness of the skin and adipose tissue at the site of the probe
205 placement was measured via 2D ultrasound operating in B-mode (Zonare Ultra Smart Cart,
206 Software version 4.7, USA). This was to ensure that data largely represented absorption of near-
207 infrared light in muscle tissue and not in subcutaneous fat.

208

209 2.4. Data analysis

210 2.4.1. *Muscle deoxygenation.* The NIRS-derived signal was normalised whereby the unloaded
211 exercise baseline value was adjusted to zero ('zero set'). Thus the NIRS data are presented as a
212 relative change from the baseline- to the end-exercise values. As such 0% represents the mean
213 steady-state value of the last 30 s of the unloaded cycling and 100% represents the highest mean
214 value of the last 30 s of any work rate. This was done given the uncertainty of the optical path
215 length in the VL at rest and during exercise, so, data are presented as normalised delta units
216 Δ [HHb+Mb]. Prior to analysis, NIRS data were averaged to give 1 s intervals. The second-by-
217 second [HHb+Mb] data was averaged by applying a five-point moving average and then
218 normalised to the peak amplitude of the response ($\% \Delta$ [HHb+Mb]). The [HHb+Mb] response
219 dynamics were expressed in relation to relative power output (%PO) prior to curve fitting.
220 Therefore, individual profiles were plotted as a function of %PO and characterised by a linear
221 function with two terms to establish the slope of increase of deoxygenation ($Slope_1$), plateau as

222 maximal exercise was approached ($Slope_2$), and the break point (BP) located between the
223 increasing deoxygenation and its plateau. The double linear function was applied using
224 TableCurve 2D (Systat Software, USA) as:

225

$$226 \quad y = a + b * x - c * (x-d)*f$$

227

$$228 \quad f = \text{if}(x < d, 0, 1)$$

229 where a and b represent the y-intercept and slope of the first linear function ($slope_1$), d is the
230 time delay or BP where the segments intersect, with the slope of the second linear function
231 ($slope_2$) being calculated from the parameter estimates of b and c ($slope_2 = b - c$).

232

233 2.4.2. $\Delta\dot{V}O_2/\Delta PO$

234 The rate of change $\dot{V}O_2$ relative to PO during **ramp incremental** exercise reflects the capacity
235 of aerobic metabolism to adjust to the non-steady state conditions incurred during a **ramp**
236 **incremental** protocol. **Initially, the mean response time (MRT) of ramp incremental exercise**
237 **was estimated using the approach recently described by Iannetta et al. (2019). Briefly, we**
238 **determined the average steady-state $\dot{V}O_2$ corresponding to three separate bouts of moderate-**
239 **intensity constant-power outputs (performed on a separate visit), and we then compared the**
240 **ramp-derived power output associated with that $\dot{V}O_2$ to the constant-power output that elicited**
241 **that $\dot{V}O_2$ (Iannetta et al., 2019). The difference between these power outputs was converted to**
242 **the time to retrieve the time-interval corresponding to MRT.** The breath by breath $\dot{V}O_2$ data
243 were averaged over 15 s intervals and plotted as a function of work rate **after applying the MRT**
244 to reflect the increase in aerobic metabolism ($\Delta\dot{V}O_2$) for each increase in power output (ΔPO).
245 From this the $\Delta\dot{V}O_2/\Delta PO$ slope was calculated over the same range of PO as used to determine
246 the first $\% \Delta[HHb+Mb]/\% PO$ slope (i.e parameter b or $slope_1$) as described above.

247

248 *2.5. Statistical analyses*

249 Statistical analysis was performed using the software SigmaPlot version 12.5 (Systat Software,
250 Point Richmond, CA). Prior to analysis, normal Gaussian distribution of the data was assessed
251 using the Shapiro-Wilk's test. Physical characteristics and NIRS-derived muscle deoxygenation
252 responses between groups were compared using unpaired 2-tailed Student's t-test for
253 parametric analyses, or the Mann-Whitney U test for non-parametric analyses. Based on *a*
254 *priori* evidence on the pre-determined reduced functional exercise capacity in individuals with
255 uncomplicated T2D, the peak physiological responses between groups were compared using
256 unpaired 1-tailed Student's t-test for parametric analyses, or the Mann-Whitney U test for non-
257 parametric analyses. Correlations between variables were established using the Pearson
258 product-moment correlation coefficient (Pearson *r*). Statistical significance was accepted at a *P*
259 ≤ 0.05 . All values are expressed as means \pm standard deviation (SD) or as median and
260 interquartile ranges for data that were deemed not normally distributed.

261

262 **3. Results**

263 *3.1. Physical characteristics and activity levels.*

264 Participants' physical characteristics and activity levels are shown in Table 1. Both groups were
265 well matched according to sex, age, body mass and BMI. Inactivity levels did not differ between
266 groups, but individuals with T2D recorded higher light intensity activity levels. As expected,
267 participants with T2D displayed higher HbA_{1c} and fasting plasma glucose levels. They also had
268 higher total cholesterol than the controls.

269

270 *3.2. Performance data from ramp incremental cycling test*

271 Relative $\dot{V}O_{2peak}$ (mean difference = 6.14 mL.kg⁻¹.min⁻¹), absolute $\dot{V}O_{2peak}$ (mean difference =
272 0.42 L.min⁻¹) and peak PO were significantly (*P* < 0.05) reduced in individuals with T2D

273 compared with controls (Table 2). In addition, $\dot{V}O_2$ at VT and $\dot{V}O_2$ at RCP were also
274 significantly lower in T2D ($P < 0.05$) compared with controls (Table 2).

275

276 3.3. NIRS-derived [HHb+Mb] response dynamics *and correlations*

277 Group mean parameter estimates from the double linear model of the $\% \Delta$ [HHb+Mb] profile as
278 a function of normalised power output (%PO) are displayed in Table 3. **Individual**
279 representative profiles of the modelled [HHb+Mb] response dynamics as a function of %PO
280 are displayed in Fig 1, **while group mean responses are shown in Fig 2**. Due to a technical error
281 with the NIRS data (i.e. the entire [HHb+Mb] responses were negative instead of positive), data
282 from 6 participants (3 controls: 2 males, 1 female; and 3 participants with T2D: 2 males, 1
283 female). were excluded from the analyses. The slope of the first linear regression function
284 ($slope_1$) used to establish the dynamic adjustment of [HHb+Mb] was significantly steeper ($P <$
285 0.05) in participants with T2D than the controls (Table 3, **Fig 3**). **In addition, in T2D $slope_1$ was**
286 **significantly correlated with absolute $\dot{V}O_{2peak}$ ($r = -0.67$, $P = 0.009$), relative $\dot{V}O_{2peak}$ ($r = -0.64$;**
287 **$P = 0.013$) and peak PO ($r = -0.74$; $P = 0.003$); whereas $slope_1$ was not correlated with these**
288 **variables in ND controls ($r = 0.132$, $P = 0.65$; $r = 0.01$, $P = 0.97$; $r = 0.155$, $P = 0.60$;**
289 **respectively). Correlations between $slope_1$ and absolute $\dot{V}O_{2peak}$ for both groups are shown in**
290 **Fig 4**. The exclusion of these 6 participants did not affect the physical characteristics of each
291 group (i.e. they were matched in terms of age, body composition and activity levels) or the peak
292 exercise responses between groups.

293

294 3.4. $\Delta \dot{V}O_2 / \Delta PO$

295 The rate of change in $\dot{V}O_2 / PO$ was not significantly different during the **ramp incremental**
296 exercise between the T2D and ND groups with no observed differences in slopes (**9.3 ± 3.4 vs.**
297 **$9.6 \pm 1.1 \text{ mL} \cdot \text{min}^{-1} \cdot \text{W}^{-1}$ respectively, $P = 0.23$).**

298

299 4. Discussion

300 The principal original finding of the present investigation was that individual with T2D
301 demonstrated a significantly steeper primary slope of the bi-linear regression used to establish
302 the dynamic adjustment of [HHb+Mb] during a ramp incremental exercise compared with
303 controls. Concomitant with the reduced ($\sim 21\%$) $\dot{V}O_{2\text{peak}}$ responses observed in individuals with
304 T2D compared with controls herein and previously (Baldi et al., 2003; Kiely et al., 2015; Mac
305 Ananey et al., 2011; O'Connor et al., 2015; O'Connor et al., 2012; Regensteiner et al., 1998),
306 such adjustment of [HHb+Mb] provides further insight into pathophysiological mechanisms
307 potentially responsible for the reduced functional capacity in this clinical population. Given that
308 overall, the objectively measured physical activity levels did not differ between groups, the
309 exaggerated exercise intolerance is likely not affected by differences in activity levels.
310 Therefore, in agreement with our hypothesis, the present study suggests that T2D alters the
311 profile of muscle fractional O_2 extraction during ramp incremental cycle exercise. Specifically,
312 T2D induced a greater reliance on normalized O_2 extraction for a given normalized PO up to
313 the [HHb+Mb]-BP (i.e. larger slope_1), and importantly, slope_1 was inversely correlated with
314 peak exercise capacity in participants with T2D.

315

316 Accordingly, the accelerated muscle deoxygenation revealed by the steeper primary
317 $\% \Delta[\text{HHb+Mb}] / \% \text{PO}$ slope of the bi-linear regression indicates a reduced capacity to increase
318 peripheral O_2 delivery to meet increasing O_2 demands. The expression of this response in
319 relation to the absolute workload may provide misleading conclusions given a diseased
320 population with an established exercise intolerance (i.e. lower peak PO) is being compared to
321 a healthy, albeit obese, population. A steeper adjustment of $\Delta[\text{HHb+Mb}]$ would be expected in
322 participants with T2D given their lower peak PO during the ramp incremental test. Thus, it is
323 warranted to make comparisons amongst these populations in the context of relative intensity
324 (i.e. as a function of PO%) (Murias et al., 2013). A reduced $\text{Pmv}O_2$ and intracellular PO_2 will

325 radically impact muscle metabolism by reducing [phosphocreatine] and elevating [ADP]_{free},
326 [Pi], [H⁺] and [NADH]. This increased glycolysis will rely on finite glycogen stores
327 culminating in premature muscular fatigue and ultimately increased exercise intolerance in this
328 clinical population. It should be noted that owing to the generation of noisy $\dot{V}O_2$ data in some
329 of the participants, in the present study we were unable to assess the relationship of
330 $\% \Delta[\text{HHb}+\text{Mb}]$ with $\% \dot{V}O_2$ responses.

331

332 These findings are in accordance with studies whereby O₂ availability during incremental
333 exercise is deliberately compromised. Specifically, where O₂ delivery was manipulated via
334 exercising in the supine posture and subsequently reducing perfusion pressure (DiMenna et al.,
335 2010; Egaña et al., 2013). In particular, DiMenna et al. (2010) demonstrated a significantly
336 steeper slope of the $\% \Delta[\text{HHb}+\text{Mb}]/\% \text{PO}$ sigmoidal response profile in the supine compared
337 with upright posture during a ramp incremental exercise, implying a greater reliance on O₂
338 extraction for the same PO (DiMenna et al., 2010). Similarly, Behnke et al. (2002) reported
339 *P_{mvo2}* reductions at a given muscle stimulation intensity in the GK rodent model of T2D
340 compared with healthy controls predicating either a reduced O₂ diffusion across the capillary-
341 myocyte space to the mitochondria or a lowered intramyocyte PO₂ which would impair muscle
342 metabolism and function (Behnke et al., 2002). Thus, the findings of the present study combined
343 with the previously reported blunted microvascular blood flow responses at the onset of
344 moderate-intensity cycling exercise in individuals with uncomplicated T2D (Bauer et al., 2007),
345 strengthens the argument for reduced O₂ delivery as a likely source of impairment in $\dot{V}O_2$
346 control in this population.

347

348 With evidence of an imbalance in QO₂ relative to PO within the microvasculature during ramp
349 incremental exercise in T2D, and the resultant lowered *P_{mvo2}*, an impaired haemodynamic
350 response can be posited as a potential mechanistic basis for the diminished exercise capacity

351 herein. Indeed, the significant correlations observed between the initial slope of muscle
352 deoxygenation with $\dot{V}O_{2peak}$ and peak PO in the group with T2D support this notion. In this
353 regard, the attenuated hyperaemic and haemodynamic response during maximum graded calf
354 plantar flexion exercise demonstrated by this clinical population (Kiely et al., 2014) is of
355 relevance. Specifically, Kiely et al. (2014) demonstrated that peak leg blood flow and the slope
356 of leg blood flow relative to percentage peak force during an incremental calf exercise were
357 significantly blunted in men and women with T2D. These reductions were accompanied by
358 significantly lower (magnitude of ~15%) peak force relative to MVC during the calf graded
359 test, which also coincided with a significant (~15%) reduction in $\dot{V}O_{2peak}$ during a graded
360 cycling test in the same participants (Kiely et al., 2014). Therefore, the demonstration in the
361 present study of a faster rise in the primary linear $\% \Delta[\text{HHb}+\text{Mb}]/\% \text{PO}$ signal (*slope₁*) in T2D
362 compared with controls, combined with a similar rate of increase in $\dot{V}O_2$ relative to PO (i.e.
363 $\Delta \dot{V}O_2/\Delta \text{PO}$) extends the findings of a dampened hyperaemic response previously observed in
364 isolated muscle groups to that of whole body exercise in uncomplicated T2D.

365

366 Although the mechanisms responsible for the altered profile of muscle fractional O_2 extraction
367 observed in individuals with T2D were not directly explored in this study, the impaired vascular
368 function extensively evidenced in T2D is a likely culprit. For instance, attenuated endothelium-
369 dependent vasodilation of resistance vessels in both, the resting forearm (McVeigh et al., 1992;
370 Williams et al., 1996), and the lower limb during cycle exercise (Kingwell et al., 2003) have
371 been reported in individuals with uncomplicated T2D compared to controls. In addition
372 tempered vasodilator responses of the vascular smooth muscle elicited subsequent to
373 exogenous, direct-acting nitric oxide (NO) donors in the form of glyceryl trinitrate (McVeigh
374 et al., 1992) and sodium nitroprusside (Kingwell et al., 2003; Williams et al., 1996) have also
375 been reported in the respective T2D cohorts. It is pertinent to acknowledge, however, that in
376 the absence of cardiac output (CO) data, we cannot exclude the possibility that impairments in

377 cardiac function (Joshi et al., 2010; Regensteiner et al., 2009; Wilson et al., 2017a; Wilson et
378 al., 2017b) could induce subsequent regional O₂ delivery impediments; although peak CO is
379 not significantly reduced in uncomplicated T2D (Baldi et al., 2003; Regensteiner et al., 2009).
380 Moreover, factors beyond convective and diffusive O₂ delivery may also be involved given that
381 structural changes in the skeletal muscles of individuals with T2D have been observed.
382 Specifically, reductions in mitochondrial content (~30%) (Boushel et al., 2007; Ritov et al.,
383 2005) and functional capacity (~40%) (Kelley et al., 2002; Ritov et al., 2005), as well as
384 alterations in muscle fibre type (Marin et al., 1994), having an approximate 2-fold increase in
385 type IIb fibres relative to type I (Mogensen et al., 2007) have been reported in T2D, although
386 the functional evidence for this notion is unclear (Rabol et al., 2006).

387

388 Limitations of the present study should be acknowledged. Firstly, given the functional
389 limitations of the NIRS technology utilised herein, we were unable to make direct comparisons
390 of absolute concentration and changes in Δ [HHb+Mb] between individuals with and without
391 T2D. However, [HHb+Mb] possesses a time course similar to fractional O₂ extraction (Koga et
392 al., 2012). Secondly, the present findings relate to the evaluation of a single muscle, the VL,
393 and as such, cannot wholly represent the skeletal muscle blood flow response to exercise. Also,
394 the heterogeneity within an individual muscle is recognised; structurally, pertaining to
395 vascularity and fibre type (Johnson et al., 1973), and functionally, relating to fibre recruitment,
396 vascular control and blood flow (Behnke et al., 2003; Koga et al., 2011; McDonough et al.,
397 2005). Thirdly, adipose tissue thickness at the site of measurement has the potential to influence
398 NIRS measurements through its effect on the scattering properties of the tissue. As such, the
399 thickness of the skin and adipose tissue was measured at the site of the interrogation via 2D
400 ultrasound operating in B-mode, with no differences revealed between groups. The current
401 findings are applicable to individuals <55 yr, so, future studies should assess if these effects are
402 also apparent in older people with T2D.

403

404 **5. Conclusions**

405 The findings from the present study offer an insight into potential contributory mechanisms for
406 the consistently observed reduction in exercise capacity in T2D. The demonstration of a greater
407 rate of O₂ extraction for a given increase in PO suggests that a reduced O₂ delivery within the
408 microvasculature is an important underlying cause of exercise intolerance during a maximum
409 graded test in T2D. This observation strengthens the notion that factors beyond central control
410 also contribute to the diminished exercise tolerance of this clinical population. Such factors are
411 most likely attributed to impairments in active muscle microvascular perfusion. Thus, exercise
412 training interventions designed to benefit exercise tolerance in T2D should also focus on
413 microvascular O₂ delivery.

414

415 **Author contribution statement**

416 NG, JR, ME, DO'S and SG designed the study. NG, JR and AMcD contributed to data
417 collection. NG, JR, SG and ME performed the data analysis. NG, JR and ME performed the
418 statistical analyses. NG and ME wrote the manuscript. All authors commented on the
419 manuscript and approved the final version of the manuscript.

420

421 **Declarations of interest**

422 None.

423

424 **Funding**

425 This publication has emanated from research conducted with the financial support of the Health
426 Research Board (Grant No HRA_POR/2073/274).

429 **References**

- 430 Amann, M., Subudhi, A.W., Foster, C., 2006. Predictive validity of ventilatory and lactate thresholds
431 for cycling time trial performance. *Scand J Med Sci Sports* 16 (1), 27-34.
- 432 Baldi, J.C., Aoina, J.L., Oxenham, H.C., Bagg, W., Doughty, R.N., 2003. Reduced exercise
433 arteriovenous O₂ difference in Type 2 diabetes. *J Appl Physiol* 94 (3), 1033-1038.
- 434 Bauer, T.A., Reusch, J.E., Levi, M., Regensteiner, J.G., 2007. Skeletal muscle deoxygenation after the
435 onset of moderate exercise suggests slowed microvascular blood flow kinetics in type 2
436 diabetes. *Diabetes Care* 30 (11), 2880-2885.
- 437 Beaver, W.L., Wasserman, K., Whipp, B.J., 1986. A new method for detecting anaerobic threshold
438 by gas exchange. *J Appl Physiol* 60 (6), 2020-2027.
- 439 Behnke, B.J., Kindig, C.A., McDonough, P., Poole, D.C., Sexton, W.L., 2002. Dynamics of
440 microvascular oxygen pressure during rest-contraction transition in skeletal muscle of
441 diabetic rats. *Am J Physiol Heart Circ Physiol* 283 (3), H926-932.
- 442 Behnke, B.J., McDonough, P., Padilla, D.J., Musch, T.I., Poole, D.C., 2003. Oxygen exchange profile in
443 rat muscles of contrasting fibre types. *J Physiol* 549 (Pt 2), 597-605.
- 444 Bellotti, C., Calabria, E., Capelli, C., Pogliaghi, S., 2013. Determination of maximal lactate steady state
445 in healthy adults: can NIRS help? *Med Sci Sports Exerc* 45 (6), 1208-1216.
- 446 Boushel, R., Gnaiger, E., Schjerling, P., Skovbro, M., Kraunsoe, R., Dela, F., 2007. Patients with type
447 2 diabetes have normal mitochondrial function in skeletal muscle. *Diabetologia* 50 (4), 790-
448 796.
- 449 Copp, S.W., Hageman, K.S., Behnke, B.J., Poole, D.C., Musch, T.I., 2010. Effects of type II diabetes on
450 exercising skeletal muscle blood flow in the rat. *J Appl Physiol* (1985) 109 (5), 1347-1353.
- 451 DeLorey, D.S., Kowalchuk, J.M., Paterson, D.H., 2003. Relationship between pulmonary O₂ uptake
452 kinetics and muscle deoxygenation during moderate-intensity exercise. *J Appl Physiol* (1985)
453 95 (1), 113-120.
- 454 DiMenna, F.J., Bailey, S.J., Jones, A.M., 2010. Influence of body position on muscle deoxy[Hb+Mb]
455 during ramp cycle exercise. *Respir Physiol Neurobiol* 173 (2), 138-145.
- 456 Egaña, M., Columb, D., O'Donnell, S., 2013. Effect of low recumbent angle on cycling performance,
457 fatigue, and V O₂ kinetics. *Med Sci Sports Exerc* 45 (4), 663-673.
- 458 Ferrari, M., Mottola, L., Quaresima, V., 2004. Principles, techniques, and limitations of near infrared
459 spectroscopy. *Can J Appl Physiol* 29 (4), 463-487.
- 460 Ferrari, M., Muthalib, M., Quaresima, V., 2011. The use of near-infrared spectroscopy in
461 understanding skeletal muscle physiology: recent developments. *Philos Trans A Math Phys*
462 *Eng Sci* 369 (1955), 4577-4590.
- 463 Green, S., Egana, M., Baldi, J.C., Lamberts, R., Regensteiner, J.G., 2015. Cardiovascular control during
464 exercise in type 2 diabetes mellitus. *J Diabetes Res* 2015 654204.
- 465 Iannetta, D., Murias, J.M., Keir, D.A., 2019. A Simple Method to Quantify the V O₂ Mean Response
466 Time of Ramp-Incremental Exercise. *Med Sci Sports Exerc* 51 (5), 1080-1086.
- 467 Iannetta, D., Qahtani, A., Millet, G.Y., Murias, J.M., 2017. Quadriceps Muscles O₂ Extraction and
468 EMG Breakpoints during a Ramp Incremental Test. *Front Physiol* 8 686.
- 469 Inglis, E.C., Iannetta, D., Murias, J.M., 2017. The plateau in the NIRS-derived [HHb] signal near the
470 end of a ramp incremental test does not indicate the upper limit of O₂ extraction in the vastus
471 lateralis. *Am J Physiol Regul Integr Comp Physiol* 313 (6), R723-r729.
- 472 Johnson, M.A., Polgar, J., Weightman, D., Appleton, D., 1973. Data on the distribution of fibre types
473 in thirty-six human muscles. An autopsy study. *J Neurol Sci* 18 (1), 111-129.
- 474 Joshi, D., Shiwalkar, A., Cross, M.R., Sharma, S.K., Vachhani, A., Dutt, C., 2010. Continuous, non-
475 invasive measurement of the haemodynamic response to submaximal exercise in patients
476 with diabetes mellitus: evidence of impaired cardiac reserve and peripheral vascular
477 response. *Heart* 96 (1), 36-41.

478 Keir, D.A., Fontana, F.Y., Robertson, T.C., Murias, J.M., Paterson, D.H., Kowalchuk, J.M., Pogliaghi, S.,
479 2015. Exercise Intensity Thresholds: Identifying the Boundaries of Sustainable Performance.
480 *Med Sci Sports Exerc* 47 (9), 1932-1940.

481 Kelley, D.E., He, J., Menshikova, E.V., Ritov, V.B., 2002. Dysfunction of mitochondria in human
482 skeletal muscle in type 2 diabetes. *Diabetes* 51 (10), 2944-2950.

483 Kiely, C., O'Connor, E., O'Shea, D., Green, S., Egaña, M., 2014. Hemodynamic responses during
484 graded and constant-load plantar flexion exercise in middle-aged men and women with type
485 2 diabetes. *J Appl Physiol* (1985) 117 (7), 755-764.

486 Kiely, C., Rocha, J., O'Connor, E., O'Shea, D., Green, S., Egana, M., 2015. Influence of menopause and
487 Type 2 diabetes on pulmonary oxygen uptake kinetics and peak exercise performance during
488 cycling. *Am J Physiol Regul Integr Comp Physiol* 309 (8), R875-883.

489 Kingwell, B.A., Formosa, M., Muhlmann, M., Bradley, S.J., McConell, G.K., 2003. Type 2 diabetic
490 individuals have impaired leg blood flow responses to exercise: role of endothelium-
491 dependent vasodilation. *Diabetes Care* 26 (3), 899-904.

492 Kodama, S., Saito, K., Tanaka, S., Maki, M., Yachi, Y., Asumi, M., Sugawara, A., Totsuka, K., Shimano,
493 H., Ohashi, Y., Yamada, N., Sone, H., 2009. Cardiorespiratory fitness as a quantitative predictor
494 of all-cause mortality and cardiovascular events in healthy men and women: a meta-analysis.
495 *Jama* 301 (19), 2024-2035.

496 Koga, S., Kano, Y., Barstow, T.J., Ferreira, L.F., Ohmae, E., Sudo, M., Poole, D.C., 2012. Kinetics of
497 muscle deoxygenation and microvascular PO₂ during contractions in rat: comparison of
498 optical spectroscopy and phosphorescence-quenching techniques. *J Appl Physiol* (1985) 112
499 (1), 26-32.

500 Koga, S., Poole, D.C., Fukuoka, Y., Ferreira, L.F., Kondo, N., Ohmae, E., Barstow, T.J., 2011.
501 Methodological validation of the dynamic heterogeneity of muscle deoxygenation within the
502 quadriceps during cycle exercise. *Am J Physiol Regul Integr Comp Physiol* 301 (2), R534-541.

503 Lalande, S., Gusso, S., Hofman, P.L., Baldi, J.C., 2008. Reduced leg blood flow during submaximal
504 exercise in type 2 diabetes. *Med Sci Sports Exerc* 40 (4), 612-617.

505 Laplaud, D., Hug, F., Grelot, L., 2006. Reproducibility of eight lower limb muscles activity level in the
506 course of an incremental pedaling exercise. *J Electromyogr Kinesiol* 16 (2), 158-166.

507 Mac Ananey, O., Malone, J., Warmington, S., O'Shea, D., Green, S., Egaña, M., 2011. Cardiac output
508 is not related to the slowed O₂ uptake kinetics in type 2 diabetes. *Med Sci Sports Exerc* 43 (6),
509 935-942.

510 MacAnaney, O., Reilly, H., O'Shea, D., Egaña, M., Green, S., 2011. Effect of type 2 diabetes on the
511 dynamic response characteristics of leg vascular conductance during exercise. *Diab Vasc Dis*
512 *Res* 8 (1), 12-21.

513 Marin, P., Andersson, B., Krotkiewski, M., Bjorntorp, P., 1994. Muscle fiber composition and capillary
514 density in women and men with NIDDM. *Diabetes Care* 17 (5), 382-386.

515 McDonough, P., Behnke, B.J., Padilla, D.J., Musch, T.I., Poole, D.C., 2005. Control of microvascular
516 oxygen pressures in rat muscles comprised of different fibre types. *J Physiol* 563 (Pt 3), 903-
517 913.

518 McVeigh, G.E., Brennan, G.M., Johnston, G.D., McDermott, B.J., McGrath, L.T., Henry, W.R.,
519 Andrews, J.W., Hayes, J.R., 1992. Impaired endothelium-dependent and independent
520 vasodilation in patients with type 2 (non-insulin-dependent) diabetes mellitus. *Diabetologia*
521 35 (8), 771-776.

522 Mogensen, M., Sahlin, K., Fernstrom, M., Glintborg, D., Vind, B.F., Beck-Nielsen, H., Hojlund, K.,
523 2007. Mitochondrial respiration is decreased in skeletal muscle of patients with type 2
524 diabetes. *Diabetes* 56 (6), 1592-1599.

525 Murias, J.M., Keir, D.A., Spencer, M.D., Paterson, D.H., 2013. Sex-related differences in muscle
526 deoxygenation during ramp incremental exercise. *Respir Physiol Neurobiol* 189 (3), 530-536.

527 O'Connor, E., Green, S., Kiely, C., O'Shea, D., Egana, M., 2015. Differential effects of age and type 2
528 diabetes on dynamic vs. peak response of pulmonary oxygen uptake during exercise. *J Appl*
529 *Physiol* (1985) 118 (8), 1031-1039.

530 O'Connor, E., Kiely, C., O'Shea, D., Green, S., Egaña, M., 2012. Similar level of impairment in exercise
531 performance and oxygen uptake kinetics in middle-aged men and women with type 2
532 diabetes. *Am J Physiol Regul Integr Comp Physiol* 303 (1), R70-76.

533 Okushima, D., Poole, D.C., Barstow, T.J., Rossiter, H.B., Kondo, N., Bowen, T.S., Amano, T., Koga, S.,
534 2016. Greater V O₂peak is correlated with greater skeletal muscle deoxygenation amplitude
535 and hemoglobin concentration within individual muscles during ramp-incremental cycle
536 exercise. *Physiol Rep* 4 (23).

537 Padilla, D.J., McDonough, P., Behnke, B.J., Kano, Y., Hageman, K.S., Musch, T.I., Poole, D.C., 2006.
538 Effects of Type II diabetes on capillary hemodynamics in skeletal muscle. *Am J Physiol Heart*
539 *Circ Physiol* 291 (5), H2439-2444.

540 Padilla, D.J., McDonough, P., Behnke, B.J., Kano, Y., Hageman, K.S., Musch, T.I., Poole, D.C., 2007.
541 Effects of Type II diabetes on muscle microvascular oxygen pressures. *Respir Physiol*
542 *Neurobiol* 156 (2), 187-195.

543 Poitras, V.J., Bentley, R.F., Hopkins-Rosseel, D.H., LaHaye, S.A., Tschakovsky, M.E., 2015.
544 Independent effect of type 2 diabetes beyond characteristic comorbidities and medications
545 on immediate but not continued knee extensor exercise hyperemia. *J Appl Physiol* (1985) 119
546 (3), 202-212.

547 Poitras, V.J., Hudson, R.W., Tschakovsky, M.E., 2018. Exercise intolerance in Type 2 diabetes: is there
548 a cardiovascular contribution? *J Appl Physiol* (1985) 124 (5), 1117-1139.

549 Poole, D.C., 1997. Influence of exercise training on skeletal muscle oxygen delivery and utilization.
550 In Crystal RG, West JB, Weibel ER, Barnes PJ, editors. *The Lung: Scientific Foundations*. New
551 York: Raven Press. . 1957-1967.

552 Rabol, R., Boushel, R., Dela, F., 2006. Mitochondrial oxidative function and type 2 diabetes. *Appl*
553 *Physiol Nutr Metab* 31 (6), 675-683.

554 Regensteiner, J.G., Bauer, T.A., Reusch, J.E., Brandenburg, S.L., Sippel, J.M., Vogelsson, A.M., Smith,
555 S., Wolfel, E.E., Eckel, R.H., Hiatt, W.R., 1998. Abnormal oxygen uptake kinetic responses in
556 women with type II diabetes mellitus. *J Appl Physiol* 85 (1), 310-317.

557 Regensteiner, J.G., Bauer, T.A., Reusch, J.E., Quaife, R.A., Chen, M.Y., Smith, S.C., Miller, T.M.,
558 Groves, B.M., Wolfel, E.E., 2009. Cardiac dysfunction during exercise in uncomplicated type 2
559 diabetes. *Med Sci Sports Exerc* 41 (5), 977-984.

560 Ritov, V.B., Menshikova, E.V., He, J., Ferrell, R.E., Goodpaster, B.H., Kelley, D.E., 2005. Deficiency of
561 subsarcolemmal mitochondria in obesity and type 2 diabetes. *Diabetes* 54 (1), 8-14.

562 Rowlands, A.V., Thomas, P.W., Eston, R.G., Topping, R., 2004. Validation of the RT3 triaxial
563 accelerometer for the assessment of physical activity. *Med Sci Sports Exerc* 36 (3), 518-524.

564 Spencer, M.D., Murias, J.M., Paterson, D.H., 2012. Characterizing the profile of muscle
565 deoxygenation during ramp incremental exercise in young men. *Eur J Appl Physiol* 112 (9),
566 3349-3360.

567 Swift, D.L., Lavie, C.J., Johannsen, N.M., Arena, R., Earnest, C.P., O'Keefe, J.H., Milani, R.V., Blair, S.N.,
568 Church, T.S., 2013. Physical activity, cardiorespiratory fitness, and exercise training in primary
569 and secondary coronary prevention. *Circ J* 77 (2), 281-292.

570 Van Beekvelt, M.C., Colier, W.N., Wevers, R.A., Van Engelen, B.G., 2001. Performance of near-
571 infrared spectroscopy in measuring local O₂ consumption and blood flow in skeletal muscle.
572 *J Appl Physiol* (1985) 90 (2), 511-519.

573 Vieth, E., 1989. Fitting piecewise linear regression functions to biological responses. *J Appl Physiol*
574 (1985) 67 (1), 390-396.

575 Wagner, P.D., Hoppeler, H., Saltin, B., 1997. Determinants of maximal oxygen uptake. In Crystal RG,
576 West JB, Weibel ER, Barnes PJ, editors. *The Lung: Scientific Foundations*. New York: Raven
577 Press. 2033-2204.

578 Wasserman, K., McIlroy, M.B., 1964. DETECTING THE THRESHOLD OF ANAEROBIC METABOLISM IN
579 CARDIAC PATIENTS DURING EXERCISE. *Am J Cardiol* 14 844-852.

580 Wasserman, K., Whipp, B.J., Koyle, S.N., Beaver, W.L., 1973. Anaerobic threshold and respiratory gas
581 exchange during exercise. *J Appl Physiol* 35 (2), 236-243.

582 Wilkerson, D.P., Poole, D.C., Jones, A.M., Fulford, J., Mawson, D.M., Ball, C.I., Shore, A.C., 2011. Older
583 type 2 diabetic males do not exhibit abnormal pulmonary oxygen uptake and muscle oxygen
584 utilization dynamics during submaximal cycling exercise. *Am J Physiol Regul Integr Comp*
585 *Physiol* 300 (3), R685-692.

586 Williams, S.B., Cusco, J.A., Roddy, M.A., Johnstone, M.T., Creager, M.A., 1996. Impaired nitric oxide-
587 mediated vasodilation in patients with non-insulin-dependent diabetes mellitus. *J Am Coll*
588 *Cardiol* 27 (3), 567-574.

589 Wilson, G.A., Wilkins, G.T., Cotter, J.D., Lamberts, R.R., Lal, S., Baldi, J.C., 2017a. Impaired ventricular
590 filling limits cardiac reserve during submaximal exercise in people with type 2 diabetes.
591 *Cardiovasc Diabetol* 16 (1), 160.

592 Wilson, G.A., Wilson, L.C., Lamberts, R.R., Majeed, K., Lal, S., Wilkins, G.T., Baldi, J.C., 2017b. beta-
593 Adrenergic Responsiveness in the Type 2 Diabetic Heart: Effects on Cardiac Reserve. *Med Sci*
594 *Sports Exerc* 49 (5), 907-914.

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597

598 **Figure captions**

599 **Figure 1:** Representative profiles of the modelled [HHb+Mb] response dynamics during ramp
600 incremental exercise for an individual without, and an individual with T2D when expressed as
601 a function of relative power output (PO%). Double-linear regression models are superimposed
602 on the data. The first $\% \Delta[\text{HHb+Mb}]/\% \text{PO}$ slope of the double linear regression is indicated
603 beside each curve. Note the relatively larger slope in the participant with T2D compared with
604 the control participant.

605

606 **Figure 2:** Group mean \pm SD normalised [HHb+Mb] responses as a function of relative power
607 output (PO%). Data are shown at 10% PO intervals. Note the relatively steeper increase in
608 [HHb+Mb] in the group with T2D compared with the control group.

609

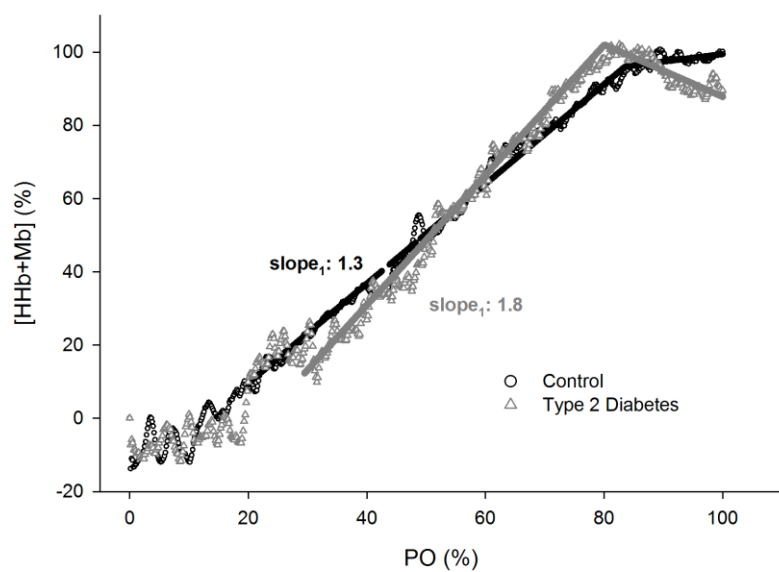
610 **Figure 3:** Individual and mean \pm SD (*bar graph*) responses of the first $\% \Delta[\text{HHb+Mb}]/\% \text{PO}$
611 slope (Slpoe_1) of the double linear regression in the T2D and control groups.

612

613 **Figure 4:** Relationships between first $\% \Delta[\text{HHb+Mb}]/\% \text{PO}$ slope (Slpoe_1) of the double linear
614 regression and $\dot{V}\text{O}_{2\text{peak}}$ ($\text{mL}\cdot\text{kg}^{-1}\cdot\text{min}^{-1}$) in participants with T2D and ND controls.

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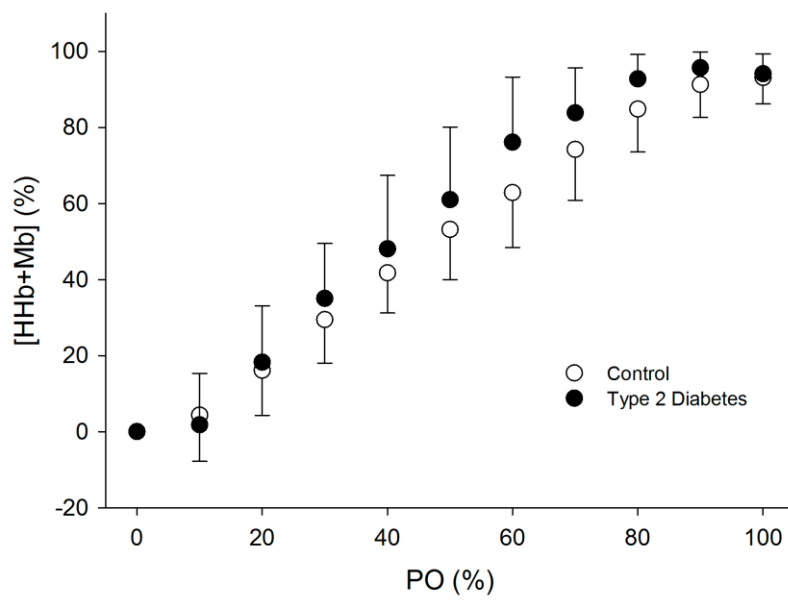
Figure 1



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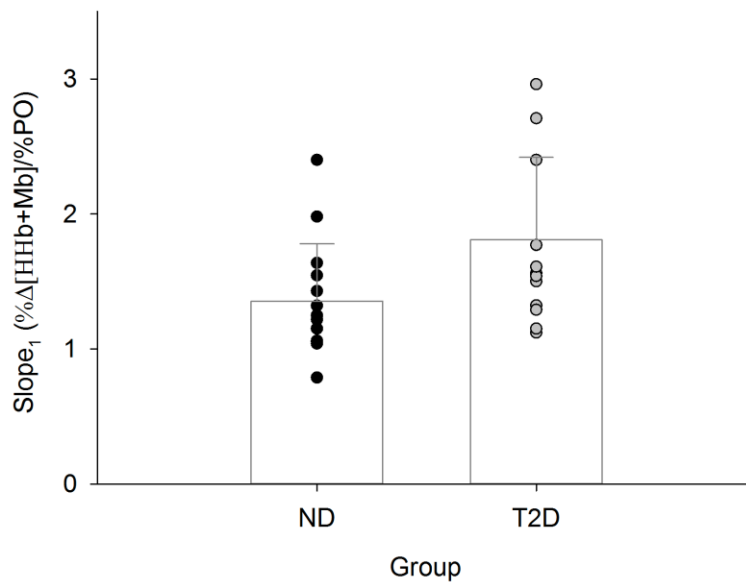
Figure 2



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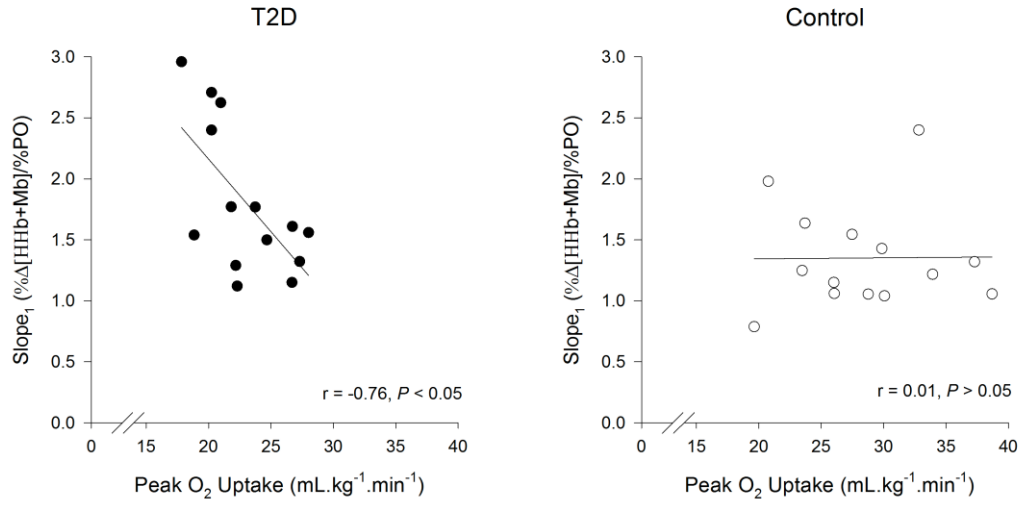
Figure 3



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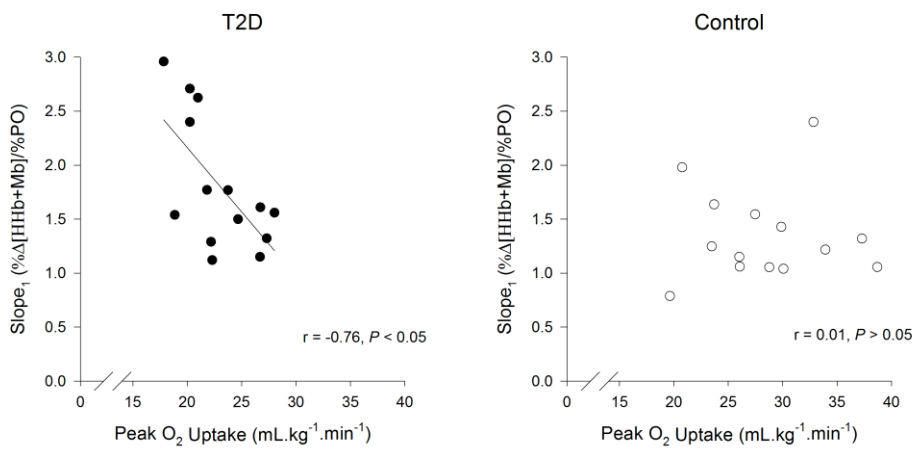
Figure 4



622

623 Alternative Fig 4 (no regression line in control)

Figure 4



624

625

626 Table 1. *Physical characteristics and activity levels.*

	ND	T2D	<i>P</i> value
<i>n</i>	17	17	
Physical characteristics			
Sex (male, female)	12, 5	12, 5	
Age (yr)	44 ± 8	48 ± 7	0.13
BMI (kg.m ⁻²)	30.8 ± 3.5	31.9 ± 4.8	0.46
Body Mass (kg)	91.1 ± 13.8	95.8 ± 18.3	0.40
HbA1c (%) ^a	<i>5.1 (0.5)*</i>	<i>6.8 (0.9)</i>	<0.001
FPG (mmol.L ⁻¹) ^b	<i>4.0 (0.4)*</i>	<i>7.4 (2.9)</i>	<0.001
Fat layer VL (mm) ^c	7.8 ± 4.5	5.9 ± 1.6	0.14
Time since diagnosis (yr)		5.7 ± 3.7	
Total cholesterol (mmol.L ⁻¹) ^d	3.6 ± 0.9*	4.4 ± 0.7	0.03
LDL-C (mmol.L ⁻¹) ^e	2.0 ± 0.7	2.2 ± 0.7	0.50
HDL-C (mmol.L ⁻¹) ^d	1.2 ± 0.2	1.35 ± 0.3	0.62
Triglycerides (mmol.L ⁻¹) ^f	<i>1.1 (0.9)†</i>	<i>1.5 (1.3)</i>	0.08
Habitual physical activity			
Inactive (h.day ⁻¹) ^g	18.9 ± 1.3	18.0 ± 1.0	0.15
Light (h.day ⁻¹) ^g	4.2 ± 1.0*	5.3 ± 1.1	0.05
Moderate (h.day ⁻¹) ^g	0.7 ± 0.4	0.6 ± 0.6	0.69
Vigorous (h.day ⁻¹) ^g	<i>0.2 (0.2)</i>	<i>0.1 (0.1)</i>	0.17

627

628 Mean ± SD values are shown in normal font for variables which were normally
629 distributed; whereas median (and interquartile range) values are shown in italic font for
630 variables which showed significant skewness and were not normally distributed in one
631 or both groups. BMI, body mass index; HbA_{1c}, glycosylated haemoglobin; FPG, fasting
632 plasma glucose; VL, vastus lateralis; LDL-C, low-density lipoprotein cholesterol; HDL-
633 C, high-density lipoprotein cholesterol. Some variables have missing values and the
634 sample sizes with codes are shown below. *Significantly different than T2D ($P \leq 0.05$).

635 †Tendency towards a difference than T2D ($P \leq 0.10$).

636 ^a = 7 (ND) and 15 (T2D); ^b = 10 (ND) and 13 (T2D); ^c = 13 (ND) and 15 (T2D); ^d = 10
637 (ND) and 12 (T2D); ^e = 10 (ND) and 10 (T2D); ^f = 10 (ND) and 13 (T2D); ^g = 13 (ND)
638 and 6 (T2D).

639

640

641 Table 2. *Physiological responses to the ramp incremental test.*

	ND	T2D	<i>P</i> value
<i>n</i>	17	17	
$\dot{V}O_{2peak}$ (mL.kg ⁻¹ .min ⁻¹)	28.62 ± 5.50*	22.48 ± 3.65	<0.001
$\dot{V}O_{2peak}$ (L.min ⁻¹)	2.60 ± 0.58*	2.18 ± 0.65	0.03
Peak PO (W)	<i>196 (108)*</i>	<i>186 (106)</i>	<i>0.04</i>
Peak HR (beats.min ⁻¹)	<i>175 (27)*</i>	<i>165 (26)</i>	0.04
Peak RER (a.u.)	<i>1.2 (0.1)</i>	<i>1.1 (0.1)</i>	<i>0.20</i>
Peak MAP (mmHg) ^a	126 ± 17	137 ± 24	0.14
Peak SBP (mmHg) ^a	170 ± 24*	187 ± 19	0.05
Peak DBP (mmHg) ^a	103 ± 16	103 ± 23	0.48
$\dot{V}O_2$ at VT (W)	1.78 ± 44*	1.55 ± 0.47	0.02
$\dot{V}O_2$ at RCP (W)	2.25 ± 0.49*	1.93 ± 0.57	0.04

642 Mean ± SD values are shown in normal font for variables which were normally distributed;
 643 whereas median (and interquartile range) values are shown in italic font for variables which
 644 showed significant skewness and were not normally distributed in one or both groups. $\dot{V}O_2$,
 645 volume of oxygen uptake; PO, power output; HR, heart rate; RER, respiratory exchange ratio;
 646 MAP, mean arterial pressure; SBP, systolic blood pressure; DBP, diastolic blood pressure; VT,
 647 ventilatory threshold; RCP, respiratory compensation point. Some variables have missing
 648 values and the sample sizes with codes are shown below. *Significantly different than T2D (*P*
 649 ≤ 0.05).

650 ^a = 9 (ND) and 12 (T2D).
 651

652

653 **Table 3.** Parameter estimates for the $\% \Delta[\text{HHb} + \text{Mb}]$ profile for both groups plotted as a
 654 function of normalised PO (%) during the ramp incremental test.
 655

	ND	T2D	<i>P</i> value
<i>n</i>	14	14	
<i>b</i> (<i>slope</i> ₁)	1.35 (0.43)*	1.81 (0.61)	0.02
<i>b - c</i> (<i>slope</i> ₂)	0.15 ± 0.67	-0.21 ± 0.57	0.14
<i>BP</i> (%)	81.2 ± 11.9	75.2 ± 12.5	0.20

656
 657 Mean ± SD values are shown in normal font for variables which were normally distributed;
 658 whereas median (and interquartile range) values are shown in italic font for variables which
 659 showed significant skewness and were not normally distributed in one or both groups. *Slope*₁
 660 and *Slope*₂ of linear regression before and after breakpoint (*BP*) respectively. *Significantly
 661 different than T2D (*P* < 0.05).
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