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A Review

Trianthema portulacastrum L.: Traditional medicine in healthcare and biology

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Medicinal plants are the major folk and traditional medicine for the prevention of diseases worldwide. *Trianthema portulacastrum* L. (family: Aizoaceae), a small perennial weed, found in the America, Africa, India, and other regions of the world, and are extensively used not only as medicine but also as vegetable for its various health benefits. Phytochemical analysis of *T. portulacastrum* reveals the presence of alkaloids, phytosterols, terpenoids, saponins, flavonoids and phenolic compounds. *In vitro* and *in vivo* studies have demonstrated its pharmacological and biological activities. Different parts of *T. portulacastrum* L. are conventionally being used as analgesic, anti-pyretic, lipid lowering and microbicide agent; and protect liver and kidney from carcinogen, inflammation and oxidant chemicals.

Keywords: Antioxidant, Anti-inflammation, Hepatoprotective, Phytochemicals, Trianthema portulacastrum L.

Introduction

Medicinal plants are the major folk and traditional medicine for the prevention of diseases worldwide, especially in underdeveloped and developing countries, where the modern scientific treatment and therapies are challenging or expensive¹⁻³. The global demand for herbal medicine is increasing. Compounds isolated from different herbs are used as key components to treat various diseases. A major share of FDA approved drugs has been reported directly or indirectly is based on natural products¹. Many synthetic molecules resemble structural homology with various natural products that serve as leads⁴. India is

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Abbreviations: ACE, Angiotensin Converting Enzyme; AFB1, Aflatoxin B1; CA, caffeic acid; CAPE, Caffeic acid phenethyl ester; CCl₄, Carbon tetrachloride; DENA, Diethylnitrosamine; DMBA, 7,12-dimethylbenz[a]anthracene; DPPH, 1,1-diphenyl-2-picryl hydrazyl; FA, Ferulic acid; FLT3, Fms-like Tyrosine Kinase 3; GLUT, Glucose Transporter; HCC, Hepatocellular Carcinoma; HO-1, heme oxygenase-1; MDR, Multidrug resistance; NAFLD, Non-alcoholic fatty liver disease; NAPQI, N-acetyl-p-benzoquinone; NF-κB, Nuclear factor-kappa B; Nrf2, Nuclear factor (erythroid-derived 2)-like 2; PDGFR, Platelet-Derived Growth Factor Receptor; PHBA, p-hydroxy benzoic acid; ROS, Reactive oxygen species; RTK, Receptor Tyrosine Kinase family; STZ, Streptozotocin; TA, Thioacetamide; TASO, Thioacetamide S-oxide; TASO₂, Thioacetamide S,S-dioxide; TNF-α, tumor necrosis factor-α; VA, Vanillic acid

the rich source of traditional medicinal systems, where out of 2500 species of medicinal plants, 150 species are harvested for commercial use on a grand scale^{5,6}.

Trianthema porulacastrum L.

Trianthema porulacastrum L. is a well-known medicinal plant used from ancient time to treat several diseases. T. portulacastrum L. (also called Trianthema monogyna L.)⁷, (family of Aizoaceae, also known as horse purslane, Bishkhapra, carpetweed, Punarnava, Gadabani and Labuni) has historically been valued by Indian and African cultures for its numerous medicinal properties^{8,9}. The herb is found worldwide e.g. Southeast Asia (India, Bangladesh, Sri Lanka, Pakistan, etc.), Africa (like Ghana and Tanzania) and America. It grows in sunny desert as well as in irrigated and tropical rainfall areas. It's over growing nature is common in agricultural field especially in rainy seasons^{10,11}.

Botany

T. portulacastrum is considered as annual or perennial depending on geographical area and the plant is propagated by seeds but the fragments of the stem can be spread by cuttings very easily. The plant is often succulent, branched, annual terrestrial and prostrate herb that produces colored flower (red or white color). Two varieties of the plant are found, one is red-flowered variety known as 'Rakta Punarnava'

or 'Lal Sabuni' and white-flowered one is known as 'Shwet Punarnava' or 'Svet Sabuni'; and the former is more abundant⁸. The flowers are small, solitary, bisexual, pale pink or white in color and have stamens and white filaments. Both types are grown best under partial shade and flourish in neutral to alkaline soil. Few vernacular names are listed in (Table 1)¹²⁻¹⁴.

The roots are thin, tortuous, slender, lateral branching fibrous. The leaves are succulent, green, opposite, oval shaped and unequal in size. The stems are branched, cylindrical, fleshy and angular to some extent and prostrate in nature. Fruit is circumsessile capsule like shaped that partly exerted from partial perianth containing 2-8 seeds. The seeds are hairless, kidney-shaped 1.5-2.5 mm long and black in color^{7,12,15}. Pictorial form of the plant is shown in (Fig. 1).

Nutritional value

Trianthema portulacastrum L. is commonly used as vegetable in East Asian countries including India and in African countries especially, in Ghana, Cameroon and Tanzania⁹. This edible wild plant is a good source of carbohydrates, protein and minerals¹⁶⁻¹⁸. Trianthema portulacastrum L. contain approximately 9% crude protein¹⁸, 3% carbohydrate¹⁶, and supply nearly 76 kcal energy/ 100 g¹⁶. This plant is easily digestable due to its simple structural carbohydrate in cell wall¹⁹. Storage form of energy (lipid) is about 2%¹⁶. The leaves of *T. portulacastrum* contain nearly 43% of crude fiber²⁰.

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Table 1 — Ve	ernacular names:[Anonymous 2003; Shanmugam 2007; Zihad <i>et al.</i> 2019] [12-14]
Language	Name of Trianthema portulacastrum L.
English	Horse Purslane
Bengali	Gadabani, Swet punarnova, Kulpasag
Hindi	Santhi, Sabuni, Vishakhapara, Lalsabuni, Svetsabuni
Sanskrit	Shvetapunarnava, Chiratika, Dhanapatra, Shvetamula, Upothaki
Tamil	Sharunnai, Shavalai
Telegu	Ambatimadhu, Atikamamidi, Galijeru
Bombay	Svetapunarnava
Punjab	Bishkapra, itsit
Kan.	Muchchugoni
Mal.	Sharunnau
Madras	Mukkarattai
Marathi	Pundharighetntuli
Urdu	Narma

Vitamin A (\sim 0.8 mg/g), vitamin B₂ (\sim 2.02 mg/g)¹⁶, Vitamin B₃, vitamin C²¹, sodium (\sim 44 mg /g), potassium (\sim 51.6 mg/g), copper (\sim 20 mg/kg), zinc (\sim 200 mg/kg), nickel (\sim 30 mg/kg), iron (\sim 6.44 mg/g) and manganese (40 mg/kg) were found in *T. portulacastrum*¹⁶.

Different parts of T. portulacastrum are used traditionally as valuable source for pharmacological components that is used to treat alcohol poisoning, liver ailments, bronchitis, heart diseases, asthma, ascites, anemia, beri-beri, dropsy, corneal ulcers, edema, inflammation, migraine, rheumatism, piles and night blindness^{17,21,22}. Both adverse and beneficial effects of T. portulacastrum ingestion have also been reported particularly in consumption of old leaves. It is reported that the consumption of old leaves causes diarrhea and paralysis both in humans and also in domestic animals. Seeds are also reported to have harmful effects⁹. But old leaves of *T. portulacastrum* are used to treat gonorrhea in Nigeria²³. On the contrary, the root is used for diseases of the liver, spleen, and malignancy, while its leaves have been used for the treatment for diuretics diseases like edema and ascites in India, Africa and Asia⁷. The root is applied for the treatment of eye disorders like itching, corneal ulcer, night blindness and dimness of sight²².

Phytochemicals

Trianthema portulacastrum L. contains a wide range of secondary metabolites like carbohydrates, fats, tannins, terpenoides, flavonoides, steroids, alkaloids, saponins, cinnnamic acid derivatives and benzoic acid derivatives $^{24-27}$. The main constituent tetraprenoid (trianthenol; 15-hydroxymethyl-2, 6, 10, 18, 22, 26, 30-heptamethyl-14-methylene-17-hentriacontene) was isolated from chloroform extract having anti-fungal property 24 . Four more compounds such as 3-acetyl aleuritolic acid, 5-hydroxy-2-methoxy benzaldehyde, p-propoxy benzoic acid and p-methoxy benzoic acid were also reported 24 . Other tetraterpene and β-carotene were also reported 21 .

Hydrocarbons were isolated from fresh leaves surface wax using gas liquid chromatography²⁸. Total phenolic content varied within 50-98 mg gallic acid equivalents/g of dry weight. The flavonoids include 5,2'-dihydroxy-7-methoxy- 6,8-dimethylflavone (*C*-methylflavone) and 5,7- dihydroxy-6,8-dimethyl chromone (leptorumol)²⁷. Few plant sterols, such as stigmasterol, β-sitosterol and β-glucopyranosides were isolated from dried plant²⁷. Alkaloids like trianthemine and punarnavine also found in this plant²⁹.

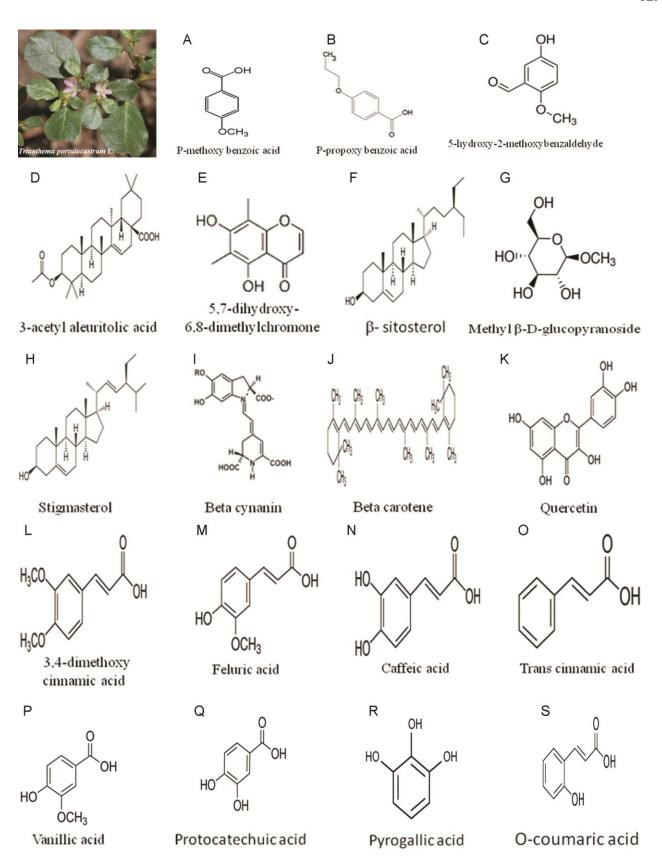


Fig. 1 — Phytochemicals available from *Trianthema portulacastrum* L.

Ecdysterone a plant steroid has been isolated from the whole plant³⁰. The red pigment β -Cyanin and 3,4-dimethoxy cinnamic acid have been reported from this plant³¹. Others phytochemicals isolated from different parts of *T. portulacastrum* include vanillic acid, ferulic acid, p-hydroxybenzoic acid, protocatechuic acid, caffeic acid, pyrogallic acid and o-coumaric acid³².

Biological activities of different phytochemicals available from *T. portulacastrum*:

- a. **p-Methoxybenzoic acid** exhibited hepatoprotective activity against carbon tetrachloride (CCl₄) and paracetamol induced hepatotoxicity *in vivo*, and thioacetamide and galactosamine-induced hepatotoxicity in isolated rat hepatocytes^{33, 34}.
- b. p-hydroxy benzoic acid (PHBA) has antimicrobial, antialgal, antimutagenic, antiestrogenic, anti-inflammatory, anti-platelet aggregate, nematicidal, antiviral, antioxidant and hypoglycemic activities. It finds use in cosmetic products, pharmaceuticals, drugs preservative and food and beverages industry³⁵⁻³⁷. Derivatives of PHBA inhibit oedema induced by acetic acid and is used in sickle cell disease³⁵.
- c. **5-hydroxy-2-methoxybenzaldehyde** acts as an essential compound for the formation of bis (benzo[b]furan-2-yl)methanones that inhibit FLT3 (Fms-like tyrosine kinase 3) and PDGFR (platelet-derived growth factor receptor) which have been implicated in numerous pathological conditions like cancer and are the members of receptor tyrosine kinase (RTK) family³⁸.
- d. **3-acetyl aleuritolic acid:** Acetyl aleuritolic acids showed effect against *S. aureus* and *S. typhimurium*³⁹. It also showed tumor-inhibitory properties toward the P-388 lymphocytic leukemia test system⁴⁰. 3-O-acetylaleuritolic acid inhibited the proliferation and migration of cancer cell lines as well as contributed to autophagy induction showing some anticancer properties⁴¹, and exhibited significant inhibitory activities on the vitality of adult male worms of *O. gutturosa*⁴².
- e. Anti-proliferative activity of 5,7-dihydroxy-6,8-dimethyl flavanone was evaluated on human colon cancer (HCT 116) cell line⁴³.
- f. **Phytosterols** (β-Sitosterol, Stigmasterol) are widely present in vegetable oils, nuts, cereal products, fruits, and berries. Phytosterol compounds reduce the inflammatory reaction in LPS-induced

macrophage models; and also inhibit the expression and activity of pro-inflammatory mediators⁴⁴.

B-sitosterol. the most common dietary phytosterol, lowers the cholesterol levels, enhances the production of plasminogen activators, and exhibits anticancer and antiatherogenic effects⁴⁵. It enhances glycemic control by increasing the activation of insulin receptor and glucose transporter 4 (GLUT4) proteins in adipose tissue. In Silico analysis showed that β-sitosterol possesses the greater binding affinity with β-arrestin-2, c-Src, and IRS-1 as well as Akt proteins and attenuate insulin resistance. It also attenuates high fat diet-induced detrimental changes in adipose tissue⁴⁶. It has been associated with cardiovascular protection by increasing the antioxidant defense system and effectively lowering the serum cholesterol level. It inhibits vascular adhesion molecule 1 and intracellular adhesion molecule 1 expression in TNF-α-stimulated human aortic endothelial cells⁴⁷.

β-sitosterol shows anti-inflammatory activity⁴⁸. It can modulate the functions of macrophages and might be a promising agent for rheumatoid arthritis therapy⁴⁹. It mediates the p53/NF-κB/breast cancer resistance protein signaling axis and regulates the response of colorectal cancer to chemotherapy⁵⁰. β-Sitosterol can prospectively be used to mitigate diet-induced non-alcoholic fatty liver disease (NAFLD)⁵¹. It is useful for prevention of Alzheimer's disease, ameliorates memory and learning impairment in APP/PS1 mice and possibly decreases Aβ deposition⁵². It also contributes to the development of the compounds as anti-aging ingredients⁵³.

Stigmasterol showed antioxidant⁵⁴, anti-inflammatory, hypoglycemic effect⁴⁸ and antimicrobial activity⁵⁵. It promoted transintestinal cholesterol secretion⁵⁶. It has neuro-protective effect against the ischemic/reperfusion (I/R) injury, possibly associated with reduction of oxidative stress and inactivation of autophagy *via* AMPK/mTOR and JNK pathways⁵⁷. It also inhibited growth of gastric cancer cells⁵⁸. It acts as a precursor in the synthesis of progesterone and acts as an intermediate in the biosynthesis of androgens, estrogens, corticoids and in the synthesis of vitamin D₃^{59,60}.

g. β -D-glucopyranoside derivative has been reported against influenza virus $(H1N1)^{61}$.

- h. Nitrogen-containing β-cyanins (red-violet) has been used as colorant in cosmetics pharmaceuticals⁶². It is a scavenger of reactive oxygen species and exhibits gene-regulatory activity partly via nuclear factor (erythroid-derived 2)-like 2-(Nrf2) dependent signaling pathways. This may induce phase II enzymes and antioxidant defense mechanisms⁶².
- i. β-carotene, important precursors of retinol (vitamin A), quench highly reactive singlet oxygen under certain conditions and can block free radicalmediated reactions^{63,64}.
- j. Quercetin and its main derivatives, such as rhamnetin, rutin, hyperoside, etc., are the major polyphenolic flavonoid found in food products, including berries, apples, cauliflower, tea, cabbage, nuts, and onions that have traditionally been treated as anticancer and antiviral, and used for the treatment of allergic, metabolic, and inflammatory disorders, eye and cardiovascular diseases. and arthritis. It has been examined against several pathogenic bacteria, viruses and parasites. It has shown beneficial effects against Alzheimer's disease, due to its inhibitory effect against acetylcholinesterase⁶⁵. It is known for its free radical scavenging activity, anti-inflammatory, anti-hypertensive, vasodilator, anti-obesity, antihypercholesterolemic and anti-atherosclerotic activities^{66,67}. These critical properties of quercetin are responsible for anti-diabetic effect⁶⁸ controlling the pathogenesis of NAFLD⁶⁹. addition, its effect on proliferation, angiogenesis, or apoptosis, are considered as anti-tumor property to enhance breast cancer treatment⁷⁰. Quercetin also demonstrated a significant protective effect on metronidazole-induced neuronal toxicity⁷¹.
- k. **3.4-dimethoxy cinnamic** acid has been reported to have anti-oxidant activity³⁴.
- 1. Plant derived Ferulic acid (FA) is an antioxidant phenolic compound. Ferulic acid phenoxyl radical is considered as stable and unreactive, which contribute it's overall antioxidant activity⁷². It prevented methotrexate-induced hepatotoxicity by activating Nrf2/HO-1 (heme oxygenase-1) signaling and PPARy, and attenuating oxidative stress, inflammation and cell death⁷³. FA and derivatives acted as platelet aggregation inhibitor, tyrosinase-inhibitor, angiotensin converting enzyme (ACE) inhibitor, and superoxide dismutase like

activities, and are involved in repair of blood vessel injury like thrombosis⁷⁴.

Ferulic acid causes cell cytotoxicity and apoptosis of HeLa and Caski cells, and the PI3K/Akt signaling pathway is down-regulated in Caski cells⁷⁵. It preserves self-renewal in embryo stem cells, and contributes to adipose-derived mesenchymal stem cells self-renewal and effective weight control in obesity⁷⁶. FA suppressed benzo(a)pyrene -induced toxicity in microglia, and exert neuroprotective effects by inhibiting microglia-mediated pro-inflammatory response⁷⁷.

Symptoms of osteoporosis include a reduction in bone strength, osteopenia, and damage to the bone microstructure. FA suppresses the fusion and apoptosis of mature osteoclasts, increase the mRNA and protein levels of SIRT1, reduced expression of nuclear factor-kappa B (NF-κB), and increase bone mineral density⁷⁸

m. Caffeic acid (3,4-dihydroxycinnamic acid) showed anti-ischemia antioxidant. reperfusion, thrombosis, antihypertension, anti-fibrosis, antivirus and antitumor properties⁷⁹. Caffeic acid (CA)-treated mice exhibited significantly lower levels of 4-hydroxynonenal, an oxidative stress marker in the hippocampus, but no effect on the expression levels of neurotrophic factors and inflammatory or anti-inflammatory cytokines, as well as significantly fewer activated microglia⁸⁰. It can be used to treat folliculitis, usually caused by a bacterial or fungal infection, due to its antioxidant potential and antimicrobial properties⁸¹. CA can prevent and delay the advanced glycation end products-induced vascular dysfunction in diabetes⁸². It can down regulate the miR-636 expression level, which is involved in development of diabetic nephropathy⁸³. CA stimulates the expression of detoxification enzymes such as regulates HO-1, and glutamatecysteine ligase through the ERK/Nrf2 pathway, and it may be an effective chemoprotective agent for protecting liver damage against oxidative damage⁸⁴. Caffeic acid has induced toxic effects and morphological changes in breast cancer cells via apoptosis⁸⁵.

Multidrug resistance (MDR) is a complicated ever-changing problem in cancer treatment, and P-glycoprotein (P-gp), a drug efflux pump, is regarded as the major cause. Caffeic acid is a promising candidate for P-gp inhibition and cancer MDR attenuation⁸⁶. It can alleviate the cell damage

induced by overexpressing A53T α -synuclein and that CA reduced A53T α -synuclein by activating the JNK/Bcl-2-mediated autophagy pathway⁸⁷.

Caffeic acid phenethyl ester (CAPE) has various biological activities including antioxidant and antiinflammatory effect. CAPE decreases the bone resorption, enhances the bone healing, prevent alveolar bone loss and stimulate periodontal tissue healing. CAPE significantly induces mRNA expression and production of VEGF in rat clonal odontoblast-like KN-3 cells cultured in normal medium or osteogenic induction medium. CAPE treatment enhances NF-kB transcription factor activation; up regulates the expression of VEGF receptor-2 and increase mineralization activity in KN-3 cells, and might be useful for the dental pulp conservative therapy.⁸⁹

Androgen receptor (AR) plays important role in the development, progression, and metastasis of prostate cancer and CAPE treatment reduces AR stability and AR transcriptional activity in PCa cells⁹⁰. Caffeic acid esters are potent bactericidal compounds against *Paenibacillus* larvae and eliminate bacterial growth through an oxidative stress mechanism⁹¹. It is also a promising agent for the prevention of skin photoaging⁹².

- n. **Cinnamic acid** and derivatives have multipurpose functions, such as drugs for anti-tuberclosis, antidiabetic, antioxidant, antimicrobial, hepatoprotective, CNS depressant, anti-cholesterolemic, antifungal, fungitoxic, anti-hyperglycemic, antimalerial, antiviral, anxiolytic, cytotoxic, anti-inflammatory and UV rays absorbent⁹³. Transcinnamic acid showed effect against colon cancer in xenograft nude mice. Trans-cinnamic acid inhibit histone deacetylases in cancer cells⁹⁴.
- o. Vanillic acid (VA), an oxidized form of vanilla, is a flavoring agent. VA improves oxidative stress in endothelial cells stimulated by palmitic acid by activating AMPK and its downstream proteins, and protect from diabetic vascular complications⁹⁵. VA exerts cardioprotective effects against Doxorubicininduced cardiotoxicity by decreasing oxidative stress, suppressing TLR4 signaling and consequently inflammation pathway⁹⁶. Anti-inflammatory and antioxidative properties of vanillic acid are associated with neuroprotective effects, resulting from Akt or ERK signaling activation. The activation of the mammalian target of rapamycin (mTOR), a key downstream target of Akt and ERK signaling, is a

crucial therapeutic target for treating depression⁹⁷. It alleviates osteoarthritis progression in a rat model by suppressing the IL-1β induced activation of MAPK and PI3K/AKT/NF-κB pathways⁹⁸.

Vanillic acid has a potent antibacterial and antibiofilm activity against carbapenem-resistant Enterobacter hormaechei and potential to be used in the food industry as a food preservative and surface disinfectant⁹⁹. VA and derivatives have antihelmintic and antisickling activities, and suppress hepatic fibrosis in chronic liver injury^{100,101}.

p. **Protocatechuic acid (PCA)**, a complex polyphenols of anthocyanins and procyanidins, possesses antioxidant, anti-inflammatory as well as antihyperglycemic and neuroprotective activities ¹⁰², antibacterial, anticancer, anti-ageing, anti-atherogenic, anti-tumoral, anti-asthma, antiulcer, antispasmodic and neurological properties ¹⁰³. PCA also have chemopreventive activity as it can inhibit antiproliferative and pro-apoptotic effects induced by chemical carcinogenes ¹⁰².

Protocatechuic acid and its alkyl esters ethyl protocatechuate are promising candidates for the prevention and treatment of UVB-induced skin photodamage and photoaging caused by generation of reactive oxygen species (ROS)¹⁰⁴. PCA-stimulated miR-219a-5p expression mitigates alcoholic liver disease by reducing p66shc-mediated ROS formation¹⁰⁵.

SIRT1 exhibits inhibitory effects on microglial activation-induced neurodegeneration. PCA inhibited the release of inflammatory mediators in LPS-activated BV2 microglia *via* the SIRT1/NF-κB pathway and thereby attenuated microglial activation-induced PC12 cell apoptosis¹⁰⁶. It has potential to prevent Alzheimer's disease¹⁰⁷.

Protocatechuic acid attenuated anterior cruciate ligament transection-induced osteoarthritis by suppressing osteoclastogenesis by inhibiting the MAPK, ATK and NF-κB signaling pathways¹⁰⁸. It shows protection against anticancer drug methotrexate-induced hepatorenal toxicity *via* antioxidant, anti-inflammatory, and antiapoptotic mechanisms¹⁰⁹. Another study suggested that phytochemicals having antioxidant efficacy is responsible for detoxification¹¹⁰.

q. **Pyrogallic acid** has anti-bacterial (cyanobacteria) activity^{111,112} and can also regulate the bacterial gene expression¹¹². It is used in various industrial and consumer products. PA is autoxidized to

purpurogallin (PG), which is further autoxidized to other polyphenolic compounds. PA and PG might participate a futile redox cycle, which mediated ROS-induced toxicity in M. aeruginosa through oxidative damage to DNA strands and cell membrane¹¹³.

r. O-coumaric acid has limited biological activity. It anticarcinogenic and chemopreventive activity in human breast cancer cell (MCF-7) line. It showed regulatory role on p53 protein, Caspase-3 protein and Bax and Bcl-2 protein and mRNA levels¹¹⁴.

Toxicity study

No toxicity was observed at dose of 3g/kg in mice model^{25, 31}. No mortality was observed at dose of 4 g/kg with methanolic extract of whole plant in rat model¹¹⁵. No atypical behavior was observed after intraperitonial administration of ethanolic extract at a dose of 250 mg/ kg in Swiss albino mice¹³.

Pharmacological Activities

Hypolipidemic activity

Dyslipidemia is associated with cardiovascular diseases, obesity and other metabolic disorders¹¹⁶. T. portulacastrum exhibited the hypolipidemic activity by controlling triglyceride, total cholesterol, LDL-cholesterol and HDL-cholesterol in blood 117 (Table 2).

Antimicrobial properties

Plant extracts are reported to exhibit antibacterial activities 118. T. portulacastrum showed anti-bacterial and anti-fungal activity¹¹⁹. Ethanolic extract of T. portulacastrum whole plant acted against gram positive bacteria¹²⁰. Aqueous, methanolic and chloroform extracts showed antibacterial properties against Escherichia Salmonella coli, Pseudomonas aeruginosa, Staphylococcus aureus, Klebsiella pneumoniae, and Shigella flexneri²⁶. Root extract of T. portulacastrum exhibited antibacterial activity against Proteus vulgaris, Bacillus subtilis, Staphylococcus aureus, Escherichia coli, Pseudomonas aeruginosa 121.

Trianthenol-1 isolated from chloroform extract of T. portulacastrum acted moderately against fungal pathogens²⁴. The flavonoid fraction of chloroform and methanolic extracts of T. portulacastrum showed antifungal activity against human pathogens such as Candida albicans, Aspergillus fumigates, A. niger,

A. flavus and Rhizopus oryzae, but showed no effect against $Mucor \text{ spp}^{26}$.

Anthelmintic, larvicidal, hormonal and chemosterilant activity

Antihelmintic activity against Haemonchus contortus (female) and their eggs were observed by aqueous methanolic crude extract of T. portulacastrum in vitro and in sheep model after infecting with nematodes (roundworms) species in vivo¹²². Aqueous and acetone extract also showed larvacidal activity against Culex quinquefasciatus, Anopheles stephensi and Aedes aegypti¹²³.

Hormonal regulation of T. portulacastrum was reported in in vivo insect model 124,125. Ecdysterone (phytoecdysone) from *T. portulacastrum* (whole plant) demonstrated chemosterilant activity through molting hormonal activity in larvae of house fly (Musca domestica)¹²⁴.

Antifertility activity

Chloroform, aqueous and alcoholic extract of T. portulacastrum leaves, stem and roots acts as a potential pregnancy interceptive, at dose of 100, 200 and 400 mg/kg body weight²³.

Analgesic, antipyretic and antinociceptive activity

Ethanolic extract of *T. portulacastrum* whole plant elicited considerable reduction on writhing response in acetic acid induced mice. 250 mg/kg of such extract demonstrated similar effect like aspirin¹³. Podo dolorimeter measurement (measure voltage threshold) showed significant result in mice; however thermal caudal immersion and mechanical tail clip techniques failed to reveal any fruitful result 120. The extract also had antinocicepative activity equivalent to drug aspirin in mice determined by hot-plate reaction time model¹³. Antipyretic activity of T. portulacastrum was observed by whole plant ethanolic extract (50 mg/kg, i.p.) in rat induced by veast pyrexia¹²⁰.

Nephroprotective, Diuretic and Antilithiatic activity

Ethanolic leaf extract of *T. portulacastrum* showed protection against nephrotic syndrome induced by adriamycin¹²⁶ (generic name doxorubicine, a drug used to treat cancer patients) and gentamicin¹²⁷ (drug known as aminoglycoside, used to treat wide range of bacterial infection) in rat by decreasing blood urine nitrogen, serum chloresterol and creatinin level, and by increasing serum protein and albumin level. Methanolic extract of T. Portulacastrum showed nephroprotective

		Ta	able 2 — Pha	rmaceutica	al Activity		
Plant extract type	Animal model & Microorganism	Dose (body weight)	Study time	Route	Toxic control	Result	Reference
Ethanolic	Female Sprage Dawley Rat	50,100, and 200 mg/kg	16 week	p.o.	7,12-dimethylbenz (a) anthracene	Chemopreventive activity against brest cancer	204
Ethanolic	Male wistar rat	100 mg/kg	7 day	p.o.	Aflatoxin	Hepatoprotective	181
Ethanolic	Male wistar rat	100, 200 mg/kg	21 day	p.o.	Aflatoxin	Antihepatotoxic	180
Ethanolic	Male and female wistar rat	100, 200 mg/kg	10 days	p.o.	Thioacetamide and paracetamol	Hepatoprotective	161
Ethanolic	Male Swiss albino mice	150 mg/kg	13 week	p.o.	CCl4	Hepatoprotective	171
Ethanolic	Male Swiss albino mice	50, 100, 150 mg/kg	3 days	p.o.	CCl4	Hepatoprotective	169
Ethanolic	Male Swiss albino mice	100, 150 mg/kg	7 week	p.o.	CCl4	Antihepatotoxic effect	150, 151
Ethanolic	Male and female wistar rat	100, 200 mg/kg	2 week	p.o.	Thioacetamide (150 mg/kg) and paracetamol (3 g/kg)	Antioxidant	162
Ethanolic	Male wistar rat	100, 200 mg/kg	10 days	p.o.	Atherosclerotic diet	Renoprotective and Hepatoprotective	128
Ethanolic	Mice	-	7 days	-	Aspirin and Acetic	Analgesic activity and antin ociceptive activity	13
Metanolic	In vitro	10 μL		-		Free radical Scavenging	134
Methanolic	In vitro	1,10,100,1000, 2000,5000 µg/mL				Radical scavenging and anti-oxidant activity	135
Methanolic	Wistar albino rats	100, 200, 300 mg/kg	7 days	i.p.	Alloxen	Hypoglycemic and hypolipidemic	117
Methanolic	Male wistar albino rat	100, 200 mg/kg	7 days	p.o.	Streptozocin	Antihyperglycemic	31
Aqueous methanolic	Male & female sheep	1,4 & 8 g	15 days	p.o.	-	Anthelmintic	122
Aqueous	Albino rat	10, 30& 50 mg/kg	-	i.p.	-	Antidiuretic	25
Aqueous, alcoholic &chloroform	Female wistar rat	100,200 & 400 mg/kg	5 day	p.o.	-	Antifertility activity	23
Aqueous, ethanolic and chloroform	Male Sprage Dawley Rat	100, 200 mg/kg	22 week	p.o.	Diethynitroso-amine	Anticarcinogenic	127, 207, 20
Aqueous and ethanolic	In vitro (paper disc diffusion)	20 μL				Antibacterial activities	209
Chloroform	In vitro (Candida Albicans, A. fumigatus	20 μL	2 days	-		Antifungal	24
Aqueous & acetone	Culex Quinquefasciatus, Anopheles stephensi and Aedes aegypti	1, 0.75, 0.75 and 1%	4 week	-	-	Larvacidal	123
Whole plant	In vitro	-	-	-	-	Fodder potential & nutritive value	19
							(Conto

Table 2 — Pharmaceutical Activity										
Plant extract type	Animal model & Microorganism	Dose (body weight)	Study time	Route	Toxic control	Result	Reference			
Aqueous, methanolic and chloroform	In vitro (bacterial and fungal)	20 μL	48 h	-		Antifungal & antibacterial activity	26			
n-butanol, hexane, chloroform and ethy acetate	In vitro (bacterial and fungal)	20 μL				Antifungal & antibacterial activity	119			

effect in renal injury, damged by artherosclerotic diet in rat¹²⁸.

Aqueous extract of T. portulacastrum showed significant diuretic activity against non-treated and furosemide treated rats at dose of 10 mg/kg²⁶. Hydro alcoholic leaf extract of T. portulacastrum also reported natriuretic effects in rat model¹²⁹.

Ethylene glycol is metabolised to glycolate by alcohol dehydrogenase, causing acidosis precipitates crystal of calcium oxalate monohydratein. Renal, kidney injury, nervous system depression and cardiopulmonary failure is the cause of ethylene glycol poisoning 130,131. The ethanolic extract of T. portulacastrum showed antilithiatic effect in ethylene glycol induced urolithiasis in male wistar rats at dose of 200 and 400 mg/kg b.w and found that urine output, phosphate, calcium, oxalate and magnesium level in urine, urea, creatinine and uric acid in blood level reestablished at near normal value after treatment 132.

Anti-inflammatory activities

Ethanolic extract of *T. portulacastrum* whole plant at dose of 100 mg/kg (i.p.) showed significant antiinflammatory response against formaldehyde induced arthritis 120,133

Antioxidant activities

T. portulacastrum extract showed free radical scavenging activity against hydrogen peroxide and DPPH (1,1-diphenyl-2-picryl hydrazyl)¹³⁴, ferricreducing power and reversed the action of linoleic acid peroxidation activity¹³⁵. Root extract showed greater inhibition effect against linoleic acid peroxidation compare to other parts 135. Ethanolic extract of T. portulacastrum showed protection against radiation-induced oxidative damage in red blood cells membrane. It optimizes the radiationinduced elevated level of TBARS and inhibits ATPase activity in RBC membrane ghosts¹³⁶. The presence of phenols, flavonoids and tannins, in particular, attenuates ROS¹³⁷.

Antihyperglycemic

Alloxan and streptozotocin are used to induce diabetes in animal through destruction of pancreatic B cell and DNA damage, respectively. Alloxan (2,4,5,6tetraoxypyrimidine; 5,6-dioxyuracil)-induced diabetes has been commonly utilized as an animal model of insuln-independent diabetes mellitus. Alloxan and reduction product dialuric acid establish a redox cycle. Superoxide radicals, hydrogen peroxide and hydroxyl radicals are produced like a chain reaction. The action of ROS with a simultaneous massive increase in cytosolic calcium concentration causes rapid destruction of B cells¹³⁸.

Streptozotocin (STZ), an antibiotic with broadspectrum activity, was isolated from a soil microorganism Streptomyces achromogenes¹³⁹. STZ has been widely used to generate an animal model of T1DM, because STZ-induced diabetes structurally, functionally, and biochemically resembles human T1DM^{140,141}. Low affinity glucose transporter 2 (GLUT2) mediates the entry of STZ into the cells. It is known that GLUT2 is highly expressed in the liver, intestine, basolateral surface of kidney, pancreatic β-cells, and central nervous systems ^{142, 143}. Cells that express GLUT2 are sensitive to STZ¹⁴⁴. STZ has diverse cytotoxic effects such as aberrant DNA alkylation, protein methylation and generation of reactive oxygen and nitrogen species (RONS)145. Excess RONS trigger cell death during early stages 146, impairs mitochondrial respiratory complex, inhibits aconitase activity, and transforms mitochondrial membrane potential, resulting in the disturbance in mitochondrial bioenergetics 147. The pancreas is more vulnerable to oxidative stress than any other tissues the low level because of of enzymes 145,148,149. These effects of STZ are responsible for necrosis of pancreatic β -cells¹³⁸. Methanolic extract of *T. portulacastrum* whole plant yield anti-hyperglycemic activity against STZ (streptozotocin) induced diabetic rat³¹, as well as alloxan stimulated hyperglycemic rat¹¹⁷. These results are comparable to oral hypoglycemic drug.

Hepatoprotective effects

The hepatoprotective activity of *T. portulacastrum* is maintained by regulation of immunity and erythropoiesis¹⁵⁰, antioxidant enzymatic activity¹⁵¹.

Overdose (>4 g/day) of antipyretic drug paracetamol/ acetaminophen causes hepatotoxicity. Paracetamol excreted as glucoronide and sulphate conjugates through urine. Electrophilic intermediate N-acetyl-pbenzoguinone imine (NAPOI) formed by isoenzyme of cytochrome CYP2E1 is responsible for hepatotoxicity caused due to excess intake of paracetamol. Trace amount of NAPOI can be inactivated by glutathione, but excess amount of NAPQI deplete glutathione. Higher amount of NAPQI bind with hepatic protein covalently that causes cell death 152. In nutshell, mechanism of liver injury (necrosis) is dependent on the accumulation of acetaminophen metabolites, NAPQI, NAPQI protein adducts, glutathione depletion, oxidative stress, and mitochondrial damage 153,154. The inflammatory cytokines, such as TNF-α, IFN-γ, and IL-1β are also crucial for the development of acetaminophen hepatisis¹⁵⁵. The NK and NKT cells play a detrimental role^{156,157}. The underlying liver injury was mediated by production of IFN-y, chemokines, and up-regulation of FasL expression in the liver 158. Thioacetamide (TA) causes necrosis that lead to hepatotoxicity. Highly reactive thioacetamide S,S-dioxide (TASO₂) produced from intermediate thioacetamide S-oxide (TASO) formed by oxidative bioactivation of thioacetamide. TASO2 alter protein structure and amine lipids causing hepatic damage 159. TA also decreases the GSH level in hepatocyte that increases the ROS production including lipoperoxidation level, leading to mitochondrial injury and cell death ¹⁶⁰. Ethanolic leaf extract of *T. portulacastrum* demonstrated hepatoprotective activity against toxicity induced by thioacetamide and paracetamol¹⁶¹ due to its antioxidant potential¹⁶².

Carbon tetrachloride (CCl₄) is a potent hepatotoxin, induces acute and chronic hepatitis¹⁶³. CCl₄ activates cytochromes (CYP2E1, CYP2B1 or CYP2B2) to form trichloromethyl (CCl₃) radical that reacts with oxygen to form a highly reactive oxygen species (ROS) CCl₃OO* that initiate lipid peroxidation, denaturation of polyunsaturated fatty acids, mitochondria

dependent liver injury and fatty degeneration 164. As a result, the mitochondrial, endoplasmic reticulum and plasma membrane permeability is lost with deregulation of Ca²⁺ in the cells leading to cellular demise¹⁶⁴. Additionally, CCl₄ toxicity leads to hypomethylation of cellular components and liver damage 164. The increased influx of cytokines, chemokines and immune cells like neutrophils, following CCl₄-induced liver injury result in hepatocyte damage (necrosis)¹⁶⁵. The Kupffer cells (KCs) also play a vital role in CCl₄-mediated hepatitis in mice as depletion of KCs protects CCl4-induced liver necrosis and IL-6 production 166. Another study demonstrated that CCl₄-mediated hepatitis was dependent upon the activity of KCs via TNF-α and FasL¹⁶⁷. It causes fat degeneration and lipid accumulation in the liver causing loss of enzyme functions like glucose-6-phosphatases and cytochrome P-450 monooxygenase. CCl₄ is also responsible for reversible blocking of intracellular gap junction immediately after intoxication, leading to impaired movement of calcium and consequently cell death 168. Ethanolic extract of T. portulacastrum showed protective effect against CCl₄ induced hepatotoxicity in mice¹⁶⁹. This protective activity is comparable with the standard hepato-protective silymarin drug¹⁷⁰. It showed protective effect on early DNA damage and chromosomal anomaly in mouse liver damaged by CCl₄¹⁷¹. Free radical scavenging activity and antioxidant property of ethanolic extract of T. portulacastrum reduces lipid peroxidation in CCl₄ induced mice model¹⁵¹, as observed its hepatic destruction by histopathological, hematological and biochemical parameters in liver after oral administration of the T. portulacastrum extract on Swiss albino mice 150.

Aflatoxins (mycotoxins) are mostly produced by *Aspergillus flavus*, *Aspergillus parasiticus* and *Aspergillus nomius*¹⁷². Few other species of *Aspergillus* and *Emericella* are also reported to produce aflatoxins¹⁷³. It has been reported that more than 18 different types of aflatoxins occur in nature, among them B1, B2, G1 and G2 mostly affect animals and humans. These aflatoxins cause toxic effects leading to mutagenicity, carcinogenicity and hepatotoxicity¹⁷⁴. Aflatoxin B1 (AFB1) has been considered to be more toxic than other aflatoxins¹⁷⁴. The liver is a primary target for AFB1, along with the heart, kidney, lungs, testis and bone marrow¹⁷⁴.

Orally treated AFB1 is absorbed in the small intestines and metabolised in the liver. In the liver, AFB1 is biotransformed by microsomal cytochrome

P450 to a highly reactive intermediate, AFB1-8, 9-epoxide, which binds to nucleic acids to form adducts¹⁷⁵. These adducts could block transcription and translation, thereby affecting the regulation of functional gene expression and ultimately causing hepatotoxicity¹⁷⁵. AFB1-induced hepatotoxicity also results from accumulation of ROS, interact with DNA and lead to mutations¹⁷⁵. Acute aflatoxicosis resulting from exposure to high doses of AFB1 through the diet over a short period causes hepatotoxicity while chronic aflatoxicosis resulting from exposure to low doses of AFB1 through the diet over a long period of time has been implicated in hepatocellular carcinoma¹⁷⁶.

Caspases are critical mediators of apoptotic cell death where caspase-3 is activated by mitochondriadependent (intrinsic) and independent (extrinsic) cell death pathways¹⁷⁷. Bax, a pro-apoptotic protein induces the transport of cytochrome c from the outer membrane of mitochondrial to the cytosol, and the anti-apoptotic protein Bcl-2 is involved in releasing cytochrome c¹⁷⁸. AFB1 was included in the 1st class human carcinogen group by the International Agency for Research on Cancer in 1993¹⁷⁹. Pretreatment of mouse with ethanolic leaves extract of T. portulacastrum (administered orally) showed aflatoxin B1 (AFB1, a hepatotoxicity¹⁸⁰. hepatocarcinogenic) stimulated Another study showed that administration of extract at dose 50-800 mg/kg four times in 3 h interval exerted hepatoprotection¹⁸¹.

Methanolic extract of *T. portulacastrum* reduced lipid and cholesterol level in serum and protected against hepatocellular damage induced by atherosclerotic diet (0.5% thiouracil, 1% cholic acid and 4% cholesterol) observed in rat¹²⁸.

Antioxidant and Protective Mechanism

A basal level of activity by these defensive systems of cells appears to be sufficient to protect cells against various oxidative stresses under normal conditions. Redox-responsive transcription factor, Nrf2 is the chief regulator of cellular homeostasis. Nrf2 mediates the expression of numerous oxidative stress related genes, including antioxidant proteins, phase I and II detoxification enzymes, transport proteins, proteasome subunits, chaperones, growth factors and their receptors, and some transcription factors 182-184.

Under basal conditions, Nrf2 is primarily regulated by the Kelch-like ECH-associated protein1 (Keap1), an adaptor protein of the Cullin3 (Cul3) based E3-ligase. This Cul3-E3 ubiquitin ligase complex mediates the proteasomal degradation of Nrf2. Under normal physiological conditions, Keap1 constitutively targets Nrf2 for ubiquitin-dependent proteasomal degradation. Keap1 is inactivated during oxidative stress, and the ubiquitination of Nrf2 stops, which leads to conformational changes in the Nrf2-Keap1-Cul3 complex, and activate Nrf2. Activated Nrf2 translocate into the nucleus and binds to the antioxidant response element (ARE) located in the promoter region of Nrf2 target genes. Consequently, it induces the transcription of cytoprotective genes, leading to the activation of the defensive system 185, 186. Nrf2 is primarily expressed in metabolically active organs such as the liver¹⁸⁷. The phytochemicals present in fruits and vegetables have been shown to specifically react with the cysteine residues of Keap1 leading to a conformational change, which results in diminished tagging of Nrf2 for proteolysis. Thus, phytochemicals from T. portulacastrum activated Nrf2, which might be beneficial in protecting against liver injury 188-190.

Anticarcinogenic activity

Diethylnitrosamine (DENA) is a carcinogenic agent and plays an important role in cell cycle regulation. DENA induces hepatocellular carcinoma (HCC) through over expression of regulatory protein of G₁/S phase in rat¹⁹¹. It has been proposed that DENA-induced HCC in mice models has closer histologic and genetic features to those observed in human HCC of poor prognosis than others HCC models¹⁹². DENA is biotransformed by cytochrome P450 enzymes into an ethyldiazonium ion, an electrophile that forms DENA-DNA adducts, with concomitant release of ROS¹⁹³⁻¹⁹⁵. Both oxidative stress and DNA adduct formation contribute to DENA carcinogenicity. T. portulacastrum extract showed chemo preventive activity in Sprague-Dawley rat model exposed by DENA 196. Chloroform extract also exhibited protection against hepatocarcinogenesis initiated by DENA and Phenobarbital ¹⁹⁷.

7,12-dimethylbenz[a]anthracene or DMBA, a polycyclic aromatic hydrocarbon, is one of the major carcinogenic components of car and industrial exhaust and char-broiled food 198. It initiates and promotes tumorigenesis, especially in the breast, ovary and skin depending on the route of exposure 199. It has been reported that DMBA activates the aryl hydrocarbon receptor (AhR) transcription factor that regulates a number of genes involved in cellular metabolism²⁰⁰. AhR upregulate the cytochrome P450 enzymes and

metabolizes DMBA into a mutagenic intermediate that causes DNA damage and is responsible for the initiation of tumorigenesis^{201,202}. DMBA up regulate the expression of Cyclin D1 and c-Myc, possibly through NF-κB and Wnt pathways, and play critical role in carcinogenesis²⁰³. Ethanolic extract of *T. portulacastrum* showed chemopreventive activity against DMBA-induced tumorigenic mammary gland in female Sprague-Dawley rats²⁰⁴.

Oxidative stress and Microenvironment in Cancer

Oxidative stress is involved in different stages of carcinogenesis; initiation, promotion and progression. Free radicals activate numerous mechanisms at the initiation step and contributing to mutations. In the promotion step initiated cells either enhances the proliferation and/or inhibits the cell death. During the progression step, free radicals cause uncontrolled growth of tumor cells, resistance to chemotherapy, genomic instability, metastasis and invasion²⁰⁵. Antioxidants from phytochemicals, such as *T. portulacastrum*, detoxify free radicals and ROS directly or indirectly, and thus may protect against cancer²⁰⁶.

Conclusion

Trianthema portulacastrum L. has shown its potential as nutritional plant as well for the remedial purposes in different medical purposes, such as inflammation, microbial infection, antioxidant, hyperglycemia, nephropathy and cancer etc. The phytochemicals present in T. portulacastrum are responsible for health benefits. Available evidences suggest that antioxidant property of this plant is one of the major factors responsible for its beneficial effects. On the contrary, understanding of the detailed protective mechanisms in several medical conditions is yet to be elucidated. Extensive study on the available phytochemicals from T. portulacastrum and elucidation of their mechanism on action will help to develop new drug(s) in several ailment conditions at a low cost due to its abundant availability.

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References

- Harvey AL, Natural products in drug discovery. *Drug Discov Today*, 13 (2008) 894.
- 2 Butler MS, Natural products to drugs: natural product derived compounds in clinical trials. *Nat Prod Rep*, 25 (2008) 475.

- 3 Maurya R, Srivastava S, Kulshreshta DK & Gupta CM, Traditional remedies for fertility regulation. *Curr Med Chem*, 11 (2004) 1431.
- 4 Ganeshpurkar A & Saluja A, The pharmacological potential of hesperidin. *Indian J Biochem Biophys*, 56 (2019) 287.
- Modak M, Dixit P, Londhe J, Ghaskadbi S & Devasagayam TP, Indian herbs and herbal drugs used for the treatment of diabetes. J Clin Biochem Nutr, 40 (2007) 163.
- 6 Seth SD & Sharma B, Medicinal plants in India. *Indian J Med Res*, 120 (2004) 9.
- 7 Shivhare KM, Singour PK, Chaurasiya PK & Pawar RS, Trianthema portulacastrum L. (Bishkhapra). Pharmacogn Rev, 6 (2012) 132.
- 8 Prasad S, Pharmacognostical studies of Punarnava; stem and leaf characteristics of *Boerhaavia diffusa* L. And *Trianthema* portulacastrum L. J Am Pharm Assoc Am Pharm Assoc, 37 (1948) 103.
- 9 Jansen PCM, *Trianthema portulacastrum* L. [Internet] Record from PROTA4U. tropicale) Wageningen, 2004.
- Balyan RS & Bhan VM, Emergence, growth and reproduction of horse purslane (*Trianthema portulacastrum L.*) as influenced by environmental conditions. *Weed Sci*, 34 (1986) 516.
- 11 Randhawa MA, Khan MA & Khan NH, Influence of Trianthema portulacastrum L. infestation and plant spacing on the yield and quality of maize grain. Int J Agric Biol, 11 (2009) 225.
- 12 Anonymous, In: Quality standards of Indian medicinal plants (Indian Council of Medical Research, New Delhi), 2003, 261.
- 13 Shanmugam SK, Bama S, Kiruthiga N, Kumar RS, Sivakumar T & Dhanabal P, Investigation of analgesic activity of leaves part of the *Trianthema portulacastrum* (L.) in standard experimental animal models. *Int J Green Pharm*, 1 (2007) 39.
- 14 Zihad SMNK, Gupt Y, Uddin SJ, Islam MT, Alam MR, Aziz S, Hossain M, Shilpi JA, Nahar L & Sarker SD, Nutritional value, micronutrient and antioxidant capacity of some green leafy vegetables commonly used by southern coastal people of Bangladesh. *Heliyon*, 5 (2019) e02768.
- Yamaki J, Venkata NKC, Mandal A, Bhattacharyya P & Bishayee A, Health-promoting and disease-preventive potential of *Trianthema portulacastrum* L. (Gadabani) An Indian medicinal and dietary plant. *J Integr Med*, 14 (2016) 84.
- 16 Khan N, Sultana A, Tahir N & Jamila N, Nutritional composition, vitamins, minerals and toxic heavy metals analysis of *Trianthema portulacastrum* L., a wild edible plant from Peshawar, Khyber Pakhtunkhwa. *Pak Afr J Biotechnol*, 12 (2013) 6079.
- 17 Yadav E, Singh D, Yadav P & Verma A, Attenuation of dermal wounds via downregulating oxidative stress and inflammatory markers by protocatechuic acid rich n-butanol fraction of *Trianthema portulacastrum* L. in wistar albino rats. Biomed Pharmacother, 96 (2017) 86.
- 18 Imran M, Talpur F N, Jan M I, Khan A & Khan I, Analysis of nutritional components of some wild edible plants. *J Chem Soc Pak*, 29 (2007) 500.
- 19 Bharathidhasan S Jr, Ganesh BNS & Balakrishnan V, In vitro evaluation of the nutritive value of Trianthema portulacastrum L. as a source of fodder for ruminants. Malays J Nutr, 13 (2007) 179.

- 20 Akindahunsi AA & Salawu SO, Phytochemical screening and nutritional, anti nutritional composition of leafy vegetables. Afr J Biotechnol, 4 (2005) 496.
- 21 Khare C. In: Indian medicinal plants, an illustrated dictionary. (Springer-Verlag, New York), 2007.
- 22 Falade T, Ishola IO, Akinleye MO, Oladimeji-Salami JA & Adevemi OO. Antinociceptive and anti-arthritic effects of aqueous whole plant extract of Trianthema portulacastrum L. in rodents: Possible mechanisms of action. J Ethnopharmacol, 238 (2019) 111831.
- 23 Pare S & Dabhadkar D, Evaluation of potential antifertility activity of plant Trianthema portulacastrum L. in female albino rat. Int J Appl Pharm Sci Biomed Sci, 2 (2013) 7.
- 24 Nawaz HR, Malik A & Ali MS, Trianthenol: an antifungal tetraterpenoid from Trianthema portulacastrum (Aizoaceae). Phytochemistry, 56 (2001), 99.
- 25 Asif M, Atif M, Malik ASA, Dan ZC, Ahmad I & Ahmad A, Diuretic activity of Trianthema portulacastrum L. crude extract in albino rats. Trop J Pharm Res, 12 (2013) 967.
- Kavitha D, Parvatham R & Padma PR, Assessment of Trianthema portulacastrum L. for its antimicrobial potential and investigation of their phytochemicals using HPTLC, GC-MS, and IR. Int J Pharm Pharm Sci, 6 (2016) 675.
- 27 Kokpol U, Wannachet-Isara N, Tip-Yang S, Chavasiri W, Veerachota G, Simpson J & Weavers RT, A Cmethyl flavone from Trianthema portulacastrum L. Phytochemistry, 44 (1997) 719.
- Singh BP, Singh RP & Jha OP, Flavonoids of some aizoaceae and molluginaceae of bhagalpur. Biol Bull India, 4
- 29 Chopra RN, Chatterjee CN & Ghosh S, A comparative study of Boerhaavia diffusa L, and the white and red flowered varieties of Trianthema portulacastrum L. J Med Res, 28
- 30 Banerji A, Chintalwar GJ, Joshi NK & Chadha MS, Isolation of ecdysterone from Indian plants. Phytochemistry, 10 (1971)
- Sunder AS, Rajalakshmi G, Bharath A & Rajeshwar Y, Antihyperglycemic activity of *Trianthema portulacastrum* L. streptozotocin induced diabetic in Pharmacologyonline, 1 (2009) 1006.
- Sherif EAA & Gharieb HR, Allelochemical effect of Trianthema portulacastrum L. on Amaranthus viridis L. supports the ecological importance of allelopathy. Afr J Agric Res, 6 (2011) 6690.
- Gadgoli C & Mishra SH, Antihepatotoxic activity of p-methoxy benzoic acid from Capparisspinosa. J Ethnopharmacol, 66 (1999) 187.
- 34 Vikas VV, Jaydeep NG, Sachin SP & Maharudra BK, Simultaneous quantification of p-methoxybenzoic Acid, 3, 4dimethoxycinnamic acid and ecdysteron in the extract of Trianthema portulacastrum L. and its marketed polyherbal formulation using HPLC. Int J Innov Res Dev, 4 (2015) 369.
- Manuja R, Sachdeva S, Jain A & Chaudhary J, A comprehensive review on biological activities of p-hydroxy benzoic acid and its derivatives. Int J Pharm Sci Rev Res, 22 (2013) 109.
- Charnock C & Finsrud T, Combining esters of para-hydroxy benzoic acid (parabens) to achieve increased antimicrobial activity. J Clin Pharm Ther, 32 (2007) 567.

- 37 Yu S, Plan MR, Winter G & Krömer JO, Metabolic engineering of Pseudomonas putida KT2440 for the production of para-hydroxy benzoic acid. Front Bioeng Biotechnol, 4 (2016) 90.
- Mahboobi S, Uecker A, Ce'nac C, Sellmer A, Eichhorn E, Elz S, Bo"hmer FD & Dove S, Inhibition of FLT3 and PDGFR tyrosine kinase activity by bis(benzo[b]furan-2-yl) methanones. Bioorg Med Chem, 15 (2007) 2187.
- Peres MT, DelleMonache F, Cruz AB, Pizzolatti MG & Yunes RA, Chemical composition and antimicrobial activity of Croton urucurana Baillon (Euphorbiaceae). J Ethnopharmacol, 56 (1997) 223.
- Torrance SJ, Wiedhopf RM & Cole JR, Antitumor agents Jatropha macrorhiza (Euphorbiaceae) acetylaleuritolic acid. J Pharm Sci, 66 (1977) 1348.
- Toton E, Kedziora I, Romaniuk-Drapala A, Konieczna N, Kaczmarek M, Lisiak N, Paszel-Jaworska A, Rybska A, Duszynska W, Budzianowski J, Rybczynska M & Rubis B, Effect of 3-O-acetylaleuritolic acid from in vitro-cultured Drosera spatulata on cancer cells survival and migration. Pharmacol Rep, 72 (2020) 166.
- 42 Nyasse B, Ngantchou I, Nono JJ & Schneider B, Antifilarial activity in vitro of polycarpol and 3-O-acetyl aleuritolic acid from cameroonian medicinal plants against Onchocerca gutturosa. Nat Prod Res, 20 (2006) 391.
- Memon AH, Ismail Z, Al-Suede FS, Aisha AF, Hamil MS, Saeed MA, Laghari M & Majid AM, Isolation, Characterization, Crystal Structure Elucidation of Two Flavanones and Simultaneous RP-HPLC Determination of Five Major Compounds from Syzygium campanulatum Korth. Molecules, 20 (2015) 14212.
- Yuan L, Zhang F, Shen M, Jia S & Xie J, Phytosterols suppress phagocytosis and inhibiti inflammatory mediators via ERK pathway on LPS-triggered inflammatory responses in RAW264.7 macrophages and the correlation with their structure. Foods, 8 (2019). pii: E582.
- Jiang YH, Li X, Niu W, Wang D, Wu B & Yang CH, β-Sitosterol regulated microRNAs in endothelial cells against an oxidized low-density lipoprotein. Food Funct, 11 (2020) 1881.
- Babu S, Krishnan M, Rajagopal P, Periyasamy V, Veeraraghavan V, Govindan R & Jayaraman S. β-sitosterol attenuates insulin resistance in adipose tissue via IRS-1/Akt mediated insulin signaling in high fat diet and sucrose induced type-2 diabetic rats. Eur J Pharmacol, 873 (2020) 173004.
- 47 Loizou S, Lekakis I, Chrousos GP & Moutsatsou P, b-sitosterol exhibits anti-inflammatory activity in human aortic endothelial cells. Mol Nutr Food Res, 54 (2010) 551.
- Kaur N, Chaudhary J, Jain A & Kishore L, Stigmasterol: A Comprehensive Review. Int J Pharm Sci Res, 2 (2011) 2259.
- Liu R, Hao D, Xu W, Li J, Li X, Shen D, Sheng K, Zhao L, Xu W, Gao Z, Zhao X, Liu Q & Zhang Y, β-Sitosterol macrophage polarization and attenuates modulates rheumatoid inflammation in mice. Pharm Biol, 57 (2019)
- Wang Z, Zhan Y, Xu J, Wang Y, Sun M, Chen J, Liang T, Wu L & Xu K, β-Sitosterol Reverses Multidrug Resistance via BCRP Suppression by Inhibiting the p53-MDM2 Interaction in Colorectal Cancer. J Agric Food Chem, 68 (2020) 3850.

- 51 Gumede NM, Lembede BW, Nkomozepi P, Brooksbank RL, Erlwanger KH & Chivandi E, β-Sitosterol mitigates the development of high-fructose diet-induced non-alcoholic fatty liver disease in growing male Sprague-Dawley rats. *Can J Physiol Pharmacol*, 98 (2020) 44.
- 52 Ye JY, Li L, Hao QM, Qin Y & Ma CS, β-Sitosterol treatment attenuate cognitive deficits and prevent amyloid plaque deposition in amyloid protein precursor/presenilin 1 mice. *Korean J Physiol Pharmacol*, 24 (2020) 39.
- 53 Haiyuan YU, Shen X, Liu D, Hong M & Lu Y, The protective effects of β-sitosterol and vermicularin from *Thamnolia vermicularis* (Sw.) Ach. against skin aging *in vitro*. *An Acad Bras Cienc*, 91 (2019) e20181088.
- 54 Panda S, Jafri M, Kar A & Meheta BK, Thyroid inhibitory, antiperoxidative and hypoglycemic effects of stigmasterol isolated from Buteamonosperma. *Fitoterapia*, 80 (2009) 123.
- 55 Yinusa I, George NI, Shuaibu UOA & Ayo RG, Bioactivity of stigmasterol isolated from the aerial part of Spillanthesacmella (Murr) on selected microorganism. *Int J Curr Microbiol App Sci*, 3 (2014), 475.
- 56 Lifsey HC, Kaur R, Thompson BH, Bennett L, Temel RE & Graf GA, Stigmasterol stimulates transintestinal cholesterol excretion independent of liver X receptor activation in the small intestine. *J Nutr Biochem*, 76 (2020) 108263.
- 57 Sun J, Li X, Liu J, Pan X & Zhao Q, Stigmasterol exerts neuro-protective effect against ischemic/ reperfusion injury through reduction of oxidative stress and inactivation of autophagy. *Neuropsychiatr Dis Treat*, 15 (2019) 2991.
- 58 Li K, Yuan D, Yan R, Meng L, Zhang Y & Zhu K, Stigmasterol exhibits potent antitumor effects in human gastric cancer cells mediated *via* inhibition of cell migration, cell cycle arrest, mitochondrial mediated apoptosis and inhibition of JAK/STAT signalling pathway. *J Buon*, 23 (2018) 1420.
- 59 Sundararaman P & Djerassi C, A convenient synthesis of progesterone from stigmasterol. *J Org Chem*, 42 (1977) 3633.
- 60 Kametani T & Furuyama H, Synthesis of vitamin D3 and related compounds. Med Res Rev, 7 (1987) 147.
- 61 Parhira S, Yang ZF, Zhu GY, Chen QL, Zhou BX, Wang YT, Liu L, Bai LP & Jiang ZH, *In vitro* anti-influenza virus activities of a new lignan glycoside from the latex of Calotropis gigantea. *PLoS One*, 9 (2014) e104544.
- 62 Esatbeyoglu T, Wagner AE, Schini-Kerth VB, Rimbach G, Betanin--a food colorant with biological activity. *Mol Nutr Food Res*, 59 (2015) 36.
- 63 Bendich A & Olson JA, Biological actions of carotenoids. *FASEB J*, 3 (1989) 1927.
- 64 Fiedor J & Burda K, Potential Role of Carotenoids as Antioxidants in Human Health and Disease. *Nutrients*, 6 (2014) 466.
- 65 Batiha GE, Beshbishy AM, Ikram M, Mulla ZS, El-Hack MEA, Taha AE, Algammal AM & Elewa YHA, The pharmacological activity, biochemical properties, and pharmacokinetics of the major natural polyphenolic flavonoid: quercetin. *Foods*, 9 (2020). pii: E374.
- 66 Salvamani S, Gunasekaran B, Shaharuddin NA, Ahmad SA & Shukor MY, Antiartherosclerotic effects of plant flavonoids. *Biomed Res Int*, 2014 (2014), 480258.

- 67 Sultana B & Anwar F, Flavonols (Kaempeferol, quercetin, myricetin) contents of selected fruits, vegetables and medicinal plants. *Food Chem*, 108 (2008), 879.
- 68 Abdelkader NF, Eitah HE, Maklad YA, Gamaleldin AA, Badawi MA & Kenawy SA, New combination therapy of gliclazide and quercetin for protection against STZ-induced diabetic rats. *Life Sci*, 247 (2020) 117458.
- 69 Pasdar Y, Oubari F, Zarif MN, Abbasi M, Pourmahmoudi A & Hosseinikia M, Effects of quercetin supplementation on hematological parameters in non-alcoholic fatty liver disease: a randomized, double-blind, placebo-controlled pilot study. Clin Nutr Res, 9 (2020) 11.
- 70 Ezzati M, Yousefi B, Velaei K & Safa A, A review on anticancer properties of quercetin in breast cancer. Life Sci, 248 (2020) 117463.
- 71 Chaturvedi S, Malik MY, Rashid M, Singh S, Tiwari V, Gupta P, Shukla S, Singh S & Wahajuddin M, Mechanistic exploration of quercetin against metronidazole induced neurotoxicity in rats: possible role of nitric oxide isoforms and inflammatory cytokines. Neurotoxicology, 79 (2020) 1.
- 72 Amić A, Marković Z, Dimitrić Marković JM, Milenković D & Stepanić V, Antioxidative potential of ferulic acid phenoxyl radical. *Phytochemistry*, 170 (2020) 112218.
- 73 Mahmoud AM, Hussein OE, Hozayen WG, Bin-Jumah M & Abd El-Twab SM, Ferulic acid prevents oxidative stress, inflammation, and liver injury via upregulation of Nrf2/HO-1 signaling in methotrexate-induced rats. Environ Sci Pollut Res Int, 27 (2020) 7910.
- 74 Kayahara H, Miao Z & Fujiwara G, Synthesis and biological activities of ferulic acid derivatives. *Anticancer Res*, 19 (1999), 3763.
- 75 Luo L, Zhu S, Tong Y & Peng S, Ferulic Acid Induces Apoptosis of HeLa and Caski Cervical Carcinoma Cells by Down-Regulating the Phosphatidylinositol 3-Kinase (PI3K)/Akt Signaling Pathway. *Med Sci Monit*, 26 (2020) e920095.
- 76 Cho J & Park E. Ferulic acid maintains the self-renewal capacity of embryo stem cells and adipose-derived mesenchymal stem cells in high fat diet-induced obese mice. *J Nutr Biochem*, 77 (2020) 108327.
- 77 Bao Y, Chen Q, Xie Y, Tao Z, Jin K, Chen S, Bai Y, Yang J & Shan S, Ferulic acid attenuates oxidative DNA damage and inflammatory responses in microglia induced by benzo(a)pyrene. *Int Immunopharmacol*, 77 (2019) 105980.
- 78 Hou T, Zhang L & Yang X, Ferulic acid, a natural polyphenol, protects against osteoporosis by activating SIRT1 and NF-κB in neonatal rats with glucocorticoid-induced osteoporosis. *Biomed Pharmacother*, 120 (2019) 109205.
- 79 Ren-Wang J, Kit-Man L, Po-Ming H, Mak TCW, Kam-Sang W & Kwok-Pui F, Chemistry and Biological Activities of Caffeic Acid Derivatives from Salvia miltiorrhiza. Curr Med Chem, 12 (2005) 237.
- 80 Koga M, Nakagawa S, Kato A & Kusumi I, Caffeic acid reduces oxidative stress and microglial activation in the mouse hippocampus. *Tissue Cell*, 60 (2019) 14.
- 81 Carolina Oliveira Dos Santos L, Spagnol CM, Guillot AJ, Melero A & Corrêa MA, Caffeic acid skin absorption: delivery of microparticles to hair follicles. *Saudi Pharm J*, 27 (2019) 791.

- 82 Cao X, Xia Y, Zeng M, Wang W, He Y & Liu J, Caffeic acid inhibits the formation of advanced glycation end products (AGEs) and mitigates the AGEs-induced oxidative stress and inflammation reaction in human umbilical vein endothelial cells (HUVECs). Chem Biodivers, 16 (2019) e1900174.
- Salem AM, Ragheb AS, Hegazy MGA, Matboli M & Eissa S, Caffeic acid modulates miR-636 expression in diabetic nephropathy rats. Indian J Clin Biochem, 34 (2019)
- Yang SY, Pyo MC, Nam MH & Lee KW, ERK/Nrf2 pathway activation by caffeic acid in HepG2 cells alleviates its hepatocellular damage caused by t-butylhydroperoxideinduced oxidative stress. BMC Complement Altern Med, 19 (2019) 139.
- 85 Rezaei-Seresht H, Cheshomi H, Falanji F, Movahedi-Motlagh F, Hashemian M & Mireskandari E, Cytotoxic activity of caffeic acid and gallic acid against MCF-7 human breast cancer cells: An in silico and in vitro study. Avicenna J Phytomed, 9 (2019) 574.
- 86 Teng YN, Wang CCN, Liao WC, Lan YH & Hung CC, Caffeic acid attenuates multi-drug resistance in cancer cells by inhibiting efflux function of human P-glycoprotein. Molecules, 25 (2020) pii: E247.
- 87 Zhang Y, Wu Q, Zhang L, Wang Q, Yang Z, Liu J & Feng L, Caffeic acid reduces A53T α-synuclein by activating JNK/Bcl-2-mediated autophagy in vitro and improves behaviour and protects dopaminergic neurons in a mouse model of Parkinson's disease. Pharmacol Res, 150 (2019) 104538.
- Kızıldağ A, Arabacı T, Albayrak M, Taşdemir U, Şenel E, Dalyanoglu M & Demirci E, Therapeutic effects of caffeic acid phenethyl ester on alveolar bone loss in rats with endotoxin-induced periodontitis. J Dent Sci, 14 (2019) 339.
- 89 Kuramoto H, Hirao K, Yumoto H, Hosokawa Y, Nakanishi T, Takegawa D, Washio A, Kitamura C & Matsuo T, Caffeic Acid Phenethyl Ester (CAPE) induces VEGF expression and production in rat odontoblastic cells. Biomed Res Int, 2019 (2019) 5390720.
- 90 Kuo YY, Huo C, Lin CY, Lin HP, Liu JS, Wang WC, Chang CR & Chuu CP, Caffeic acid phenethyl ester suppresses androgen receptor signaling and stability via inhibition of phosphorylation on Ser81 and Ser213. Cell Commun Signal, 17 (2019)100.
- 91 Collins W, Lowen N & Blake DJ, Caffeic acid esters are effective bactericidal compounds against Paenibacillus larvae by altering intracellular oxidant and antioxidant levels. Biomolecules, 9 (2019) pii: E312.
- 92 Shin EJ, Jo S, Choi HK, Choi S, Byun S & Lim TG, Caffeic acid phenethyl ester inhibits UV-induced MMP-1 expression by targeting histone acetyltransferases in human skin. Int *J Mol Sci*, 20 (2019) pii: E3055.
- Sharma P, Cinnamic acid derivatives: A new chapter of various pharmacological activities. J Chem Pharm, 3 (2011)
- Zhu B, Shang B, Li Y & Zhen Y, Inhibition of histone deacetylases by trans-cinnamic acid and its antitumor effect against colon cancer xenografts in athymic mice. Mol Med Rep, 13 (2016) 4159.
- Ma WF, Duan XC, Han L, Zhang LL, Meng XM, Li YL & Wang M, Vanillic acid alleviates palmitic acid-induced

- oxidative stress in human umbilical vein endothelial cells via Adenosine Monophosphate-Activated Protein Kinase signaling pathway. J Food Biochem, 43 (2019) e12893.
- Baniahmad B, Safaeian L, Vaseghi G, Rabbani M & Mohammadi B, Cardioprotective effect of vanillic acid against doxorubicin-induced cardiotoxicity in rat. Res Pharm Sci, 15 (2020) 87.
- Chuang HW, Wei IH, Lin FY, Li CT, Chen KT, Tsai MH & Huang CC, Roles of Akt and ERK in mTOR-Dependent Antidepressant Effects of Vanillic Acid. ACS Omega, 5 (2020) 3709.
- Huang X, Xi Y, Mao Z, Chu X, Zhang R, Ma X, Ni B, Cheng H & You H, Vanillic acid attenuates cartilage degeneration by regulating the MAPK and PI3K/AKT/ NF-κB pathways. Eur J Pharmacol, 859 (2019) 172481.
- Qian W, Yang M, Wang T, Sun Z, Liu M, Zhang J, Zeng Q, Cai C & Li Y, Antibacterial Mechanism of Vanillic Acid on Physiological, Morphological, and Biofilm Properties of Carbapenem-Resistant Enterobacter hormaechei. J Food Prot, 83 (2020) 576.
- 100 Itoh A, Isoda K, Kondoh M, Kawase M, Kobayashi M, Tamesada M & Yagi K, Hepatoprotective Effect of Syringic Acid and Vanillic Acid on Concanavalin A-Induced Liver Injury. Biol Pharm Bull, 32 (2009) 1215.
- 101 Itoh A, Isoda K, Kondoh M, Kawase M, Watari A, Kobayashi M, Tamesada M & Yagi K, Hepatoprotective Effect of Syringic Acid and Vanillic Acid on CCl4-Induced Liver Injury. Biol Pharm Bull, 33 (2010) 983.
- 102 MasellaR, Santangelo C, D'Archivio M, LiVolti G, Giovannini C & Galvano F, Protocatechuic Acid and Human Disease Prevention: Biological Activities and Molecular Mechanisms. Curr Med Chem, 19 (2012) 2901.
- 103 Khan AK, Rashid R, Fatima N, Mahmood S, Mir S, Khan S, Jabeen N & Murtaza G, Pharmacological activities of protocatechuic acid. Acta Polonia e Pharmaceutica ñ Drug Res, 72 (2015) 643.
- 104 Daré RG, Oliveira MM, Truiti MCT, Nakamura CV, Ximenes VF & Lautenschlager SOS, Abilities of protocatechuic acid and its alkyl esters, ethyl and heptyl protocatechuates, to counteract UVB-induced oxidative injuries and photoaging in fibroblasts L929 cell line. J Photochem Photobiol B, 203 (2020) 111771.
- 105 Fu R, Zhou J, Wang R, Sun R, Feng D, Wang Z, Zhao Y, Lv L, Tian X & Yao J. Protocatechuic Acid-Mediated miR-219a-5p Activation Inhibits the p66shc Oxidant Pathway to Alleviate Alcoholic Liver Injury. Oxid Med Cell Longev, 2019 (2019) 3527809.
- 106 Kaewmool C, Kongtawelert P, Phitak T, Pothacharoen P & Udomruk S, Protocatechuic acid inhibits inflammatory responses in LPS-activated BV2 microglia via regulating SIRT1/NF-κB pathway contributed to the suppression of microglial activation-induced PC12 cell apoptosis. J Neuroimmunol, 341 (2020) 577164.
- 107 Huang L, Zhong X, Qin S & Deng M, Protocatechuic acid attenuates \(\beta \)-secretase activity and okadaic acid-induced autophagy via the Akt/GSK-3β/MEF2D pathway in PC12 cells. Mol Med Rep, 21 (2020) 1328.
- 108 Zhang J, Fu B, Chen X, Chen D & Yang H, Protocatechuic acid attenuates anterior cruciate ligament transection-induced osteoarthritis by suppressing osteoclastogenesis. Exp Ther Med, 19 (2020) 232.

- 109 Owumi SE, Ajijola IJ & Agbeti OM, Hepatorenal protective effects of protocatechuic acid in rats administered with anticancer drug methotrexate. *Hum Exp Toxicol*, 38 (2019) 1254.
- 110 Agrawal ND, Nirala SK, Bhadauria M, Srivastava S & Shukla S, Protective potential of *Moringa oleifera* Lam. along with curcumin and piperine against beryllium-induced alterations in hepatorenal biochemistry and ultramorphology in rats. *Indian J Biochem Biophys*, 56 (2019) 70.
- 111 He Z, Wang H & Zhang T, Inhibitory effect of pyrogallic acid on Microcystis aeruginosa and the model analysis. *Wei Sheng Yan Jiu*, 42 (2013) 134.
- 112 Wu Z, Shi J & Yang S, The effect of pyrogallic acid on growth, oxidative stress, and gene expression in *Cylindrospermopsis raciborskii* (Cyanobacteria). *Ecotoxicology*, 22 (2013) 271.
- 113 Lu Z, Zhang Y, Gao Y, Liu B, Sun X, He F, Zhou Q & Wu Z, Effects of pyrogallic acid on Microcystis aeruginosa: oxidative stress related toxicity. *Ecotoxicol Environ Saf*, 132 (2016) 413.
- 114 Sen A, Atmaca P, Terzioglu G & Arslan S, Anticarcinogenic effect and carcinogenic potential of the dietary phenolic acid: o-coumaric acid. *Nat Prod Commun.* 8 (2013) 1269.
- 115 Tripathi KD, In: Essentials of Medical Pharmacology (Jaypee Brothers Medical Publishers (P) Ltd, New Delhi), 2004, 575.
- 116 Bhat B, Habib B, Bhagat N & Bajaj BK, Cholesterol lowering and antioxidant potential of probiotic bacteria isolated from locally fermented milk product kalarei. *Indian J Biochem Biophys*, 56 (2019) 363.
- 117 Anreddy RNR, Porika M, Yellu NR & Devarakonda RK, Hypoglycemic and hypolipidemic activities of *Trianthema* portulacastrum L. plant in normal and alloxan induced diabetic rats. *Int J Pharmacol*, 6 (2010) 129.
- 118 Namasivayam SKR, Shankar KG, Vivek JM, Nizar M & Sudarsan AV, *In silico* and *in vitro* analysis of quorum quenching active phytochemicals from the ethanolic extract of medicinal plants against quorum sensing mediated virulence factors of *Acinetobacter baumannii*. *Indian J Biochem Biophys*, 56 (2019) 276.
- 119 Mohammed R, El-Hawary SS & Abo-youssef AM, Biological investigation of some wild Aizoaceae and Chenopediaceae species growing in Egypt. J Nat Prod, 5 (2012) 193.
- 120 Vohora SB, Shah SA, Naqvi SA, Ahmad S & Khan MS, Studies on *Trianthema portulacastrum L. Planta Med*, 47 (1983) 106.
- 121 Kumar SS, Rajesh R & Siddiqui AA, Pharmacognostical and antibacterial activity of *Trianthema portulacastrum L. Orient J Chem*, 2 (2006) 641.
- 122 Hussain A, Khan M N, Iqbal Z, Sajid MS & Khan MK, Anthelmintic activity of *Trianthema portulacastrum L*. and *Musa paradisiaca L*. against gastrointestinal nematodes of sheep. *Vet Parasitol*, 179 (2011) 92.
- 123 Singh SP, Raghavendra K & Thomas TG, Mosquito larvicidal properties of aqueous and acetone extracts of *Trianthema portulacastrum* L. (family: Aizoaceae) against vector species of mosquitoes. *J Commun Dis*, 43 (2011) 237.
- 124 Dinan L, Phytoecdysteroids: biological aspects. *Phytochemistry*, 57 (2001) 325.

- 125 Ravishankar GA & Mehta AR, Control of ecdysterone biogenesis in tissue culture of *Trianthema portulacastrum L.* J Nat Prod, 42 (1979) 152.
- 126 Karim MS, Asraf N, Kalam A, Jahan N, Jafri MA & Ahmad G, Effects of Biskhapra (*Trianthema portulacastrum L.*) leaves extract in adriamycin induced nephrotic syndrome. *Int J Green Pharm*, 5 (2011) 329.
- 127 Balamurugan G, Jagan Mohan CM & Muthusamy P, Protective effect of *Trianthema portulacastrum L.* leaves on gentamicin induced nephrotoxicity in rats. *J Nat Remedies*, 9 (2009) 165.
- 128 Sunder SA, Reddy RNA, Rajeshwar Y, Kira NG, Prasad KD, Baburao B, Thirumurugu S & Karthik A, Protective effect of methanolic extract of *Trianthema portulacastrum* L. in atherosclerotic diet induced renal and hepatic changes in rats. *Der Pharm Lett*, 2 (2010) 540.
- 129 Karim MS, Kalam MA, Jahan N, Ahmad G & Jafri MA, Evaluation of diuretic activity of hydro alcoholic extract of Biskhapra leaves (*Trianthema portulacastrum* L.) in rat. *Hippocratic J Unani Med*, 6 (2011) 81.
- 130 Jacobsen D & McMartin KE, Methanol and ethylene glycol poisonings. Mechanism of toxicity, clinical course, diagnosis and treatment. *Med Toxicol*, 1 (1986) 309.
- 131 Seo JW, Lee JH, Son IS, Kim YJ, Kim DY, Hwang Y, Chung HA, Choi HS & Lim SD, Acute oxalate nephropathy caused by ethylene glycol poisoning. *Kidney Res Clin Pract*, 31 (2012) 249.
- 132 Lakshmi SK, Prabhakaran V, Mallikarjuna G & Gowthami A, Antilithiatic activity of *Trianthema portulacastrum* L. and Gymnema sylvestre R.Br against ethylene glycol induced urolithiasis. *Int J Pharm Sci Rev Res*, 25 (2014) 16.
- 133 Kendri SS & Wari UG, Screening of the anti-inflammatory activity of "Trianthema portulacastrum L." in acute models of inflammation. J Evol Med Dental Sci, 4 (2015) 5185.
- 134 Sunder AS, Reddy RNA, Prasad KD, Chander PK & Vemula S, Free radical scavenging activity of methanolic whole plant extract of *Trianthema portulacastrum* L. (Aizoaceae). *Int* J Pharm Sci, 2 (2010) 589.
- 135 Yaqoob S, Sultana B & Mushtaq M, In vitro antioxidant activities of Trianthema portulacastrum L. hydrolysates. Prev Nutr Food Sci, 19 (2014) 27.
- 136 Das U, Saha T & Das SK, Trianthema portulacastrum L. extract protects against gamma radiation-induced human red blood cell membrane damage in vitro. Indian J Biochem Biophys, 55 (2018) 321.
- 137 Begum SM, Padmavathi P, Saradamma B, Maturu P, Vardhan AH, Varadacharyulu NC & Reddy DV, Effect of green tea consumption on RBC morphology, membrane properties and antioxidant status in chronic cigarette smokers. *Indian J Biochem Biophys*, 55 (2018) 256
- 138 Szkudelski T, The mechanism of alloxan and streptozotocin action in B Cells of the rat pancreas. *Physiol Res*, 50 (2001) 536
- 139 Reusser F, Mode of action of streptozotocin. *J Bacteriol*, 105 (1971) 580.
- 140 Eleazu CO, Eleazu KC, Chukwuma S & Essien UN, Review of the mechanism of cell death resulting from streptozotocin challenge in experimental animals, its practical use and potential risk to humans. *J Diabetes Metab Disord*, 12 (2013) 60.

- 141 Jasmine R, Kumar GA & Rajaram R, Probing the mechanism of the anti-diabetic potential of a terpenoid from Elephantopus scaber L., an Indian ethnomedicinal plant in STZ diabetic rats- in vivo and in silico analysis. Indian J Biochem Biophys, 55 (2018) 384
- 142 Karim S, Adams DH & Lalor PF, Hepatic expression and cellular distribution of the glucose transporter family. World J Gastroenterol, 18 (2012) 6771.
- 143 Thorens B, GLUT2, glucose sensing and glucose homeostasis. Diabetologia, 58 (2015) 221.
- 144 Schnedl WJ, Ferber S, Johnson JH & Newgard CB, STZ transport and cytotoxicity. Specific enhancement in GLUT2expressing cells, Diabetes, 43 (1994) 1326.
- 145 Lee HA, Lee E, Do GY, Moon EK, Quan FS & Kim I, Histone deacetylase inhibitor MGCD0103 protects the pancreas from streptozotocin-induced oxidative stress and β-cell death. Biomed Pharmacother, 109 (2019) 921.
- 146 Gonzalez E, Rosello-Catafau J, Jawerbaum A, Sinner D, Pustovrh C, Vela J, White V, Xaus C, Peralta C & Gimeno M, Pancreatic nitric oxide and oxygen free radicals in the early stages of streptozotocin-induced diabetes mellitus in the rat, Braz J Med Biol Res, 33 (2000) 1335.
- 147 Nahdi A, John A &, Raza H, Elucidation of molecular mechanisms of streptozotocin-induced oxidative stress, apoptosis, and mitochondrial dysfunction in Rin-5F pancreatic β-Cells. Oxid Med Cell Longev, 2017 (2017)
- 148 Lenzen S, Drinkgern J & Tiedge M, Low antioxidant enzyme gene expression in pancreatic islets compared with various other mouse tissues, Free Radic Biol Med, 20 (1996) 463.
- 149 Miki A, Ricordi C, Sakuma Y, Yamamoto T, Misawa R, Mita A, Molano RD, Vaziri ND, Pileggi A & Ichii H, Divergent antioxidant capacity of human islet cell subsets: a potential cause of β-cell vulnerability in diabetes and islet transplantation. PLoS One, 13 (2018) e0196570.
- 150 Mandal A, Karmakar R, Bandyopadhyay S & Chatterjee M, Antihepatotoxic potential of Trianthema portulacastrum L. in carbon tetrachloride-induced chronic hepatocellular injury in mice: Reflection in haematological, histological and biochemical characteristics. Arch Pharm Res, 21 (1998) 223.
- 151 Mandal A, Bandyopadhyay S & Chatterjee M, Trianthema portulacastrum L. reverses hepatic lipid peroxidation, glutathione status and activities of related antioxidant enzymes in carbon tetrachloride-induced chronic liver damage in mice. Phytomedicine, 4 (1997) 239.
- 152 Vitols S, Paracetamol hepatotoxicity at therapeutic doses. J Intern Med, 253 (2003) 95.
- 153 Larson AM, Acetaminophen hepatotoxicity. Clin Liver Dis, 11 (2007) 525.
- 154 Bantel H & Schulze-Osthoff K, Mechanisms of cell death in acute liver failure. Front Physiol, 3 (2012) 79.
- 155 Ishida Y, Kondo T, Ohshima T, Fujiwara H, Iwakura Y & Mukaida N, A pivotal involvement of IFN-gamma in the pathogenesis of acetaminophen-induced acute liver injury. FASEB J, 16 (2002) 1227.
- 156 Liu ZX, Govindarajan S & Kaplowitz N, Innate immune system plays a critical role in determining the progression and severity of acetaminophen hepatotoxicity. Gastroenterology, 127 (2004) 1760.
- 157 Cover C, Liu J, Farhood A, Malle E, Waalkes MP, Bajt ML & Jaeschke H, Pathophysiological role of the acute

- inflammatory response during acetaminophen hepatotoxicity. Toxicol Appl Pharmacol, 216 (2006) 98.
- 158 Masson MJ, Carpenter LD, Graf ML & Pohl LR, Pathogenic role of natural killer T and natural killer cells in acetaminopheninduced liver injury in mice is dependent on the presence of dimethyl sulfoxide. Hepatology, 48 (2008) 889.
- 159 Hajovsky H, Hu G, Koen Y, Sarma D, Cui W, Moore DS, Staudinger JL & Hanzlik RP, Metabolism and toxicity of thioacetamide and thioacetamide S-oxide in rat hepatocytes. Chem Res Toxicol, 25 (2012) 1955.
- 160 Staňková P, Kučera O, Lotková H, Roušar T, Endlicher R & Cervinková Z. The toxic effect of thioacetamide on rat liver in vitro. Toxicol In Vitro, 24 (2010) 2097.
- 161 Kumar G, Banu SG, VanithaPappa P, Sundararajan M & Pandian RM, Hepatoprotective activity of Trianthema portulacastrum L. against paracetamol and thioacetamide intoxication in albino rats. J Ethnopharmacol, 92 (2004) 37.
- 162 Kumar G, Banu GS & Pandian MR, Evaluation of the antioxidant activity of Trianthema portulacastrum L. Indian J Pharmacol, 37 (2015) 331.
- 163 Uysal T, Uğurtan MH, Sezer ENŞ, Vatansev H, Bozkurt M & Evliyaoğlu N, The in vivo investigation of apoptotic effects of Nigella sativa on carbon tetrachloride-induced hepatotoxicity. Indian J Biochem Biophys, 55 (2018) 245.
- 164 Weber LW, Boll M & Stampfl A, Hepatotoxicity and mechanism of action of haloalkanes: carbon tetrachloride as a toxicological model. Crit Rev Toxicol, 33 (2003) 105.
- 165 Ramaiah SK & Jaeschke H, Role of neutrophils in the pathogenesis of acute inflammatory liver injury. Toxicol Pathol, 35 (2007) 757.
- 166 Kiso K, Ueno S, Fukuda M, Ichi I, Kobayashi K, Sakai T, Fukui K & Kojo S, The role of Kupffer cells in carbon tetrachloride intoxication in mice. Biol Pharm Bull, 35 (2012) 980.
- 167 Sato A, Nakashima H, Nakashima M, Ikarashi M, Nishiyama K, Kinoshita M & Seki S, Involvement of the TNF and FasL produced by CD11b Kupffer cells/macrophages in CCl4induced acute hepatic injury. PLoS One, 9 (2014) e92515.
- 168 Recknagel RO, Glende EA Jr, Dolak JA & Waller RL, Mechanisms of carbon tetrachloride toxicity. Pharmacol Ther, 43 (1989) 139.
- 169 Bishayee A, Mandal A & Chatterjee M, Prevention of alcohol-carbon tetrachloride-induced signs of early hepatotoxicity in mice by Trianthema portulacastrum L. Phytomedicine, 3 (1996) 155.
- 170 Mandal A, Bishayee A & Chatterjee M, Trianthema portulacastrum L. affords antihepatotoxic activity against carbon tetrachloride-induced chronic liver damage in mice: Reflections in subcellular levels. Phytother Res, 11 (1997), 216.
- 171 Sarkar A, Pradhan S, Mukhopadhyay I, Bose SK, Roy S, & Chatterjee M, Inhibition of early DNA-damage and chromosomal aberrations by Trianthema portulacastrum L. in carbon tetrachloride-induced mouse liver damage. Cell Biol Int. 23 (1999) 703.
- 172 Varga J, Frisvad JC & Samson R, Two new aflatoxin producing species, and an overview of Aspergillus section Flavi. Stud Mycol, 69 (2011) 57.
- 173 Reiter E, Zentek J & Razzazi E, 2009. Review on sample preparation strategies and methods used for the analysis of aflatoxins in food and feed. Mol Nutr Food Res, 53 (2009) 508.

- 174 Singh C, Prakash C, Mishraa P, Tiwaria KN, Mishra SK, Moree RS, Kumar V & Singh J, Hepatoprotective efficacy of *Premna integrifolia* L. leaves against aflatoxin B1-induced toxicity in mice. *Toxin*, 166 (2019) 88.
- 175 Rotimi OA, Rotimi SO, Duru CU, Ebebeinwe OJ, Abiodun AO, Oyeniyi BO, Faduyile FA. Acute aflatoxin B1 –induced hepatotoxicity alters gene expression and disrupts lipid and lipoprotein metabolism in rats. *Toxicology Reports*, 4 (2017) 408.
- 176 Gong YY, Watson S & Routledge MN, Aflatoxin exposure and associated human health effects, a review of epidemiological studies. *Food Saf*, 4 (2016) 14.
- 177 Bender CE, Fitzgerald P, Tait SW, Llambi F, McStay GP, Tupper DO, Pellettieri J, Alvarado AS, Salvesen GS & Green, D.R. 2012. Mitochondrial pathway of apoptosis is ancestral in metazoans. *Proc Natl Acad Sci U S A*, 109 (2012) 4904.
- 178 Tait SW & Green DR, Mitochondrial regulation of cell death. Cold Spring Harb. Perspect. Biol, 5 (2013) pii: a008706.
- 179 International Agency for Research on Cancer (IARC), Some naturally occurring substances: food items and constituents, heterocyclic aromatic amines and mycotoxins. *IARC monographs on the evaluation of carcinogenic risks to humans*, 6 (1999) 1.
- 180 Banu SG, Kumar G & Murugesan AG, Effect of ethanolic leaf extract of *Trianthema portulacastrum* L. on aflatoxin induced hepatic damage in rats. *Indian J Clin Biochem*, 24 (2009) 414.
- 181 Banu SG, Kumar G & Murugesan AG, Ethanolic leaves extract of *Trianthema portulacastrum* L. ameliorates aflatoxin B1 induced hepatic damage in rats. *Indian J Clin Biochem*, 24 (2009) 250.
- 182 Kobayashi M & Yamamoto M, Molecular mechanisms activating the Nrf2-Keap1 pathway of antioxidant gene regulation. *Antioxid Redox Signal*, 7 (2005) 385.
- 183 Stepkowski TM & Kruszewski MK, Molecular cross-talk between theNRF2/KEAP1 signaling pathway, autophagy, and apoptosis. *Free Radic Biol Med*, 50 (2011) 1186.
- 184 Taguchi K, Motohashi H & Yamamoto M, Molecular mechanisms of the Keap1-Nrf2 pathway in stress response and cancer evolution. Genes Cells, 16 (2011) 123.
- 185 Niture SK & Jaiswal AK, Nrf2 protein up-regulates antiapoptotic protein Bcl-2and prevents cellular apoptosis. *J Biol Chem*, 287 (2012) 9873.
- 186 Duan X, Li J, Li W, Xing X, Zhang Y, Zhao L, Sun G, Gao X & Li B, Antioxidant tert-butylhydroquinone ameliorates arsenic-induced intracellular damages and apoptosis through induction of Nrf2-dependent antioxidant responses aswell as stabilization of anti-apoptotic factor Bcl-2 in human keratinocytes. Free Radic Biol Med, 94 (2016) 74.
- 187 Lee JS & Surh YJ, Nrf2 as a novel molecular target for chemoprevention. *Cancer Lett*, 224 (2005) 171.
- 188 Costa S, Utan A, Speroni E, Cervellati R, Piva G, Prandini A & Guerra MC, Carnosic acid from rosemary extracts: a potential chemoprotective agent against aflatoxin B1. An in vitro study. J Appl Toxicol, 27 (2007) 152.
- 189 Cavin C, Marin-Kuan M, Langouët S, Bezençon C, Guignard G, Verguet C, Piguet D, Holzhäuser D, Cornaz R & Schilter B,

- Induction of Nrf2-mediated cellular defenses and alteration of phase I activities as mechanisms of chemoprotective effects of coffee in the liver. *Food Chem Toxicol*, 46 (2008) 1239
- 190 Hayes JD, McMahon M, Chowdhry S & Dinkova-Kostova AT, Cancer chemoprevention mechanisms mediated through the Keap1–Nrf2 pathway. *Antioxid. Redox Signal*, 13 (2010) 1713.
- 191 Park DH, Shin JW, Park SK, Seo JN, Li L, Jang JJ & Lee MJ, Diethylnitrosamine (DEN) induces irreversible hepatocellular carcinogenesis through overexpression of G1/S-phase regulatory proteins in rat. *Toxicol Lett*, 191 (2-3) (2009) 321.
- 192 Lee JS, Chu IS, Mikaelyan A, Calvisi DF, Heo J, Reddy JK & Thorgeirsson SS, Application of comparative functional genomics to identify best-fit mouse models to study human cancer. *Nat Genet*, 36 (2004) 1306.
- 193 Park JE, Seo JE, Lee JY & Kwon H. Distribution of seven N-Nitrosamines in food. *Toxicol Res*, 31 (2015) 279.
- 194 Kang JS, Wanibuchi H, Morimura K, Gonzalez FJ & Fukushima S, Role of CYP2E1 in diethylnitrosamine-induced hepatocarcinogenesis *in vivo. Cancer Res*, 67 (2007) 11141.
- 195 Yang FC, Zheng SS & Jiang TA, A modified rat model for hepatocellular carcinoma. *Hepatobiliary pancreat Dis Int*, 3 (2004) 585.
- 196 Bhattacharya S & Chatterjee M, Protective role of Trianthema portulacastrum L. against diethylnitrosamine induced experimental hepatocarcinogenesis. Cancer Lett, 129 (1998) 7.
- 197 Bhattacharya S & Chatterjee M, Inhibitory effect of Trianthema portulacastrum L. diethylnitroso-amine-induced phenobarbital promoted hepatocarcinogenesis. Neoplasma, 46 (1999) 105.
- 198 Kleiner HE, Vulimiri SV, Hatten WB, Reed MJ, Nebert DW, Jefcoate CR & DiGiovanni J, Role of cytochrome p4501 family members in the metabolic activation of polycyclic aromatic hydrocarbons in mouse epidermis. *Chem Res Toxicol*, 17 (2004) 1667.
- 199 Xue W & Warshawsky D, Metabolic activation of polycyclic and heterocyclic aromatic hydrocarbons and DNA damage: a review. *Toxicol Appl Pharmacol*, 206 (2005) 73.
- 200 Singhal R, Shankar K, Badger TM & Ronis MJ, Estrogenic status modulates aryl hydrocarbon receptor-mediated hepatic gene expression and carcinogenicity. *Carcinogenesis*, 29 (2008) 227.
- 201 Nebert DW, Petersen DD & Fornace AJ, Jr: Cellular responses to oxidative stress: The [Ah] gene battery as a paradigm. Environ Health Perspect, 88 (1990) 13.
- 202 Rundle A, Tang D, Hibshoosh H, Estabrook A, Schnabel F, Cao W, Grumet S & Perera FP, The relationship between genetic damage from polycyclic aromatic hydrocarbons in breast tissue and breast cancer. *Carcinogenesis*, 21 (2000) 1281
- 203 Papaconstantinou AD, Shanmugam I, Shan L, Schroeder IS, Qiu C, Yu M & Snyderwine EG, Gene expression profiling in the mammary gland of rats treated with 7,12-dimethylbenz[a] anthracene. *Int J Cancer*, 118 (2006) 17.
- 204 Bishayee A & Mandal A, *Trianthema portulacastrum* L. exerts chemoprevention of 7,12-dimethylbenz(a)anthracene-induced mammary tumorigenesis in rats. *Mutat Res*, 768 (2014) 107.

- 205 Jezierska-Drutel A, Rosenzweig SA & Neumann CA, Role of oxidative stress and the microenvironment in breast cancer development and progression. Adv Cancer Res, 119 (2013) 107.
- 206 Shrivastava A, Aggarwal LM, Mishra SP, Khanna HD, Shahi UP & Pradhan S, Free radicals and antioxidants in normal vs cancerous cells — An overview. Indian J Biochem Biophys, 56 (2019) 7.
- 207 Mandal A & Bishayee A, Trianthema portulacastrum L. displays anti-inflammatory responses during chemically induced rat mammary tumorigenesis through simultaneous
- and differential regulation of NF-kB and Nrf2 signaling pathways. Int J Mol Sci, 16 (2015) 2426.
- 208 Currier N, Solomon SE, Demicco EG, Chang DL, Farago M, Ying H, Dominguez I, Sonenshein GE, Cardiff RD, Xiao ZX, Sherr DH & Seldin DC, Oncogenic signalling pathways activated in DMBA-induced mouse mammary tumors. Toxicol Pathol, 33 (2005) 726.
- 209 Rattanata N, Daduang S, Phaetchanla S, Bunyatratchata W, Promraksa B, Tavichakorntrakool R, Uthaiwat P, Boonsiri P & Daduang J, Anti-oxidant and antibacterial properties of selected Thai weed extracts. Asian Pac J Trop Biomed, 4 (2014) 890.