



# Journal of Advanced Veterinary Research

<http://advetresearch.com/index.php/avr/index>



## Prevalence, Electron Microscopy and Molecular Characterization of *Cryptosporidium* species Infecting Sheep in Egypt

Reham S. Elmadawy<sup>1\*</sup>, Mohamed S. Diab<sup>2</sup>, Yasser F. Elnaker<sup>3</sup>

<sup>1</sup>Parasitology Department, Faculty of Veterinary Medicine, Benha University, Moshtohor, Toukh 13736, Egypt.

<sup>2</sup>Animal Hygiene and Zoonoses Diseases Department, Faculty of Veterinary Medicine, New Valley, Assiut University, Egypt.

<sup>3</sup>Infectious Diseases, Animal Medicine Department, Faculty of Veterinary Medicine, New Valley, Assiut University, Egypt.

### ARTICLE INFO

#### Original Research

#### Received:

24 March 2017

#### Accepted:

02 April 2017

#### Keywords:

PCR  
*Cryptosporidium*  
sheep  
RFLP  
Egypt

### ABSTRACT

*Cryptosporidium* sp. is predominant universally and sheep are an imperative zoonotic supply of the disease. Owing to the little information presented with respect to *Cryptosporidium* sp. infecting sheep, this study was directed to survey the predominance and molecular characterization of *Cryptosporidium* sp. among sheep of different ages and sexes in Qalyubia governorate, Egypt. The fecal specimens were gathered from 432 sheep of various ages ( $\leq 1$  to  $< 6$ , 6-12 and  $> 12$  months) and sexes. The samples were microscopically examined after staining by modified Zeihl-Neelsen technique and the intestinal mucosa was scanned by electron microscopy. A nested PCR was connected to amplify a 830 bp of 18S rRNA sequence of *Cryptosporidium*. RFLP (restriction fragment length polymorphism) technique using *SspI* and *VspI* enzymes for digestion of the secondary product of PCR for species identification was applied. The total infection rate was 25.93%. The parasite was more prevalent in males than females of different age groups. Two zoonotic *Cryptosporidium* species were distinguished after RFLP-PCR sequencing: *C. parvum* and *C. ubiquitum* (identified previously as Cervine genotype). The finding recommends that sheep must be considered as a noteworthy potential source of human cryptosporidiosis. A strict reconnaissance of zoonotic cryptosporidiosis must be set up to counteract human infection and to assess forthcoming disease when applying control programs.

— J. Adv. Vet. Res. (2017), 7 (2), 47-52

### Introduction

Cryptosporidiosis is considered as an important enteric diseases causing severe diarrhea and inefficient weight gains with a great hazard among small ruminant population (Foreyt, 1990). Molecular techniques could identify *Cryptosporidium* sp. in sheep feces in different countries (Soltane *et al.*, 2007; Geurden *et al.*, 2008; Quilez *et al.*, 2008; Fayer and Santin, 2009; Féres *et al.*, 2009; Paoletti *et al.*, 2009; Yang *et al.*, 2009; Wang *et al.*, 2010; ). *C. parvum*, *C. xiaoi* and *C. ubiquitum* are mainly responsible for *Cryptosporidium* infections in sheep, whereas *C. hominis*, *C. andersoni*, *C. fayeri*, *C. suis* and pig genotype II have already been recognized in a low number of animals (Majewska *et al.*, 2000). Sheep can harbor the zoonotic *Cryptosporidium* sp. causing clinical manifestations in human and they are considered as a main source of *Cryptosporidium* infection either by direct infection or by pollution of the environment (Castro-Hermida *et al.*, 2007; Geurden *et al.*, 2008;

Paoletti *et al.*, 2009). In Egypt, owing to the incessant human contact with sheep which usually excrete the most number *Cryptosporidium* oocysts during diarrhea (Bukhari and Smith, 1997) and due to the less information about molecular characterization and zoonotic impact of sheep cryptosporidiosis, so the present study was conducted to determine the prevalence of *Cryptosporidium* among different age and sex groups of sheep to identify the species infecting them using RFLP-PCR molecular technique, in addition to electron microscopic study of the parasite

### Materials and methods

#### Samples

The identification of the oocysts of *Cryptosporidium* was done on haphazardly gathered fecal specimens from sheep (124 males 308 females) obtained from different localities in Qalyubia governorate, Egypt. The samples were gathered directly from the rectum into plastic packs that were promptly topped, marked, put in a protected holder and transported to the parasitology laboratory in the Faculty of Veterinary Medicine, Mohstohor, Egypt, to be prepared and examined within

\*Corresponding author: Reham S. Elmadawy  
E-mail address: rehamelmadawy@hotmail.com

24 h. of collection. The specimens were categorized into three groups consistent with age: group 1 ( $\leq 1$  to  $< 6$  months of age, 124 animals), group 2 (6-12 months of age, 124 animals) and group 3 ( $> 12$  months, 184 animals). Direct microscopic examination of fecal smears prepared from fresh or concentrated fecal samples, followed by modifying Ziehl-Neelsen staining were performed for accurate oocysts detection (Casemore et al., 1985). The clinical symptoms and death cases accompanied cryptosporidiosis were registered. Small parts of the intestinal mucosa of dead lambs due to cryptosporidiosis were kept in glutaraldehyde (2.5%) at 4°C for scanning electron microscope. The specimens were rinsed in distilled water, dehydrated in acetone alcohol series (25%, 50%, 75%, and 100% for 10 min each). The critical point drying was done using CO<sub>2</sub> to complete the drying process. Finally, the samples were coated by gold using a spraying tool. Finally, the samples were visualized with SEM (Kuo, 2007).

#### Molecular diagnosis

It was applied on *Cryptosporidium* oocysts obtained from 112 positive animals. The whole technique was done according to Xiao et al. (1999) with modification, where a nested PCR- was applied using the following primers (Promega, USA): Primary PCR primers were 5'-TTCTAGAGCTAATACATGCG-3' (SSU-F2) and 5'-CCCATTCCTTCGAAACAGGA-3' (SSU-R2) and secondary primers were 5'-GGAAGGGTTGTATTATTAGATAAAG-3' (SSU-F3) and 5-AAGGAGTAAGGAACAA-CCTCCA-3' (SSU-R4). The amplification was done in Perkin Elmer Gene Amp, PCR Thermocycler 2400. The primers were used to amplify 830 bp of the 18S rRNA gene. Primary PCR mixture contained 3 mM MgCl<sub>2</sub>, 1× PCR buffer, 0.2 mM each dNTP, 1U of BSA (0.1 g/10 ml), 2.5 U Taq and 1M for forward and reverse primers. Total of 35 cycles for 45s at 94°C, for 45s at 55°C and for 60s at 72°C. Initial heating step for 3 min at 95°C and a final extension for 7 min at 72°C. The secondary step was done as follow: Total of 40 cycles for 30s at 94°C, for 90s at

58°C, and for 2 min at 72°C. Initial heat for 3 min at 94°C and a final extension for 7 min at 72°C. PCR products were investigated on agarose gel electrophoresis (1.5%). A PCR- RFLP assay of the secondary PCR products was applied as follow: a total volume of 50 µl reaction mixture contained 20U of *VspI*, 20U of *SspI* (Promega, USA), and 5 µl of the restriction buffer to digest 20µl of each secondary PCR product for 1hour at 37°C. Digested products were additionally fractionated on agarose gel (1.5%) and envisioned by ethidium bromide staining. *Cryptosporidium* sp. genotypes were identified by their band designs. Gel free PCR products were sequenced in both directions utilizing a big dye terminator cycle (Applied Biosystem), loaded on an AbiPrism 3100 and GeneScan Analysis v3.1 Software (Applied Biosystem, USA) as previously indicated (Bensch et al., 2000). The obtained sequence were compared with the accessible sequences in the database of the GenBank of National Center for Biotechnology Information. The alignment of the sequence and the determination of the phylogenetic association among various *Cryptosporidium* sp. were done using Clustal W (ver. 2) and Mega (Ver. 6) programs.

#### Statistical analysis

It was done utilizing ANOVA with two factors under significance level of 0.05 for the entire results using SPSS (Ver. 19). (Steel et al., 1997) Data were dealt with as a complete randomization outline and different examinations were done applying LSD.

#### Results

Out of 432 fecal specimens of sheep collected from various localities in Qalyubia governorate, *Cryptosporidium* oocysts could be identified in 25.93% after staining by modified Zeihl- Neelsen technique (Fig.1 A, B). Diarrhea, dehydration were the common symptoms of cryptosporidiosis

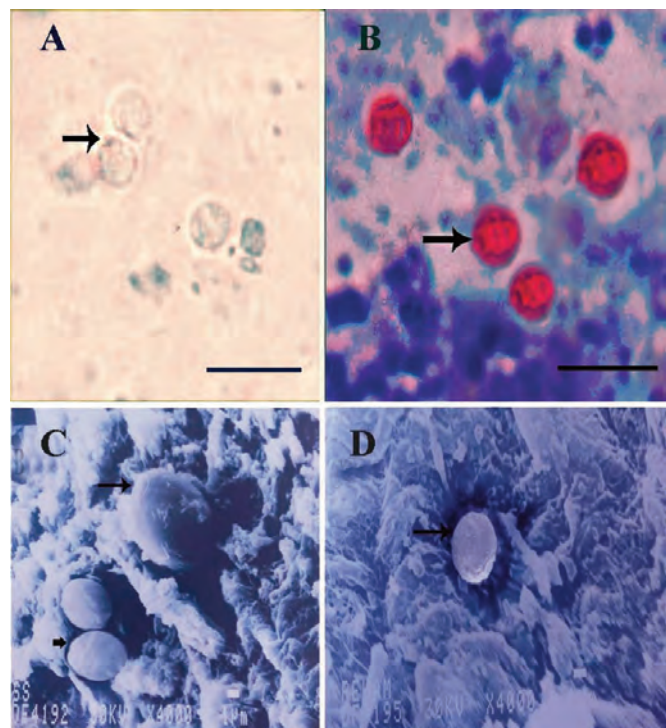


Fig. 1. A: *Cryptosporidium* oocysts in fresh fecal sample X400 (arrow; scale bar= 10µm), B: *Cryptosporidium* oocysts stained by modified Zeihl- Neelsen stain X1000 (arrow; Scale bar=10 µm).C: Scanning electron microscope showing large gamont of *Cryptosporidium* (long arrow), thin wall *Cryptosporidium* oocysts (short arrow), D: A thick wall oocyst of *Cryptosporidium* (arrow) (Scale bar =1µm).

specially in lambs <1 month of age. Four cases were found in a state of hypothermia followed by death. The results were displayed according to different age and sex groups (Table 1). Statistical analysis didn't show any substantial effect of sex on the rate of *Cryptosporidium* infection, where the total infection rates among males and females were not significantly different ( $P>0.05$ ). Though, the prevalence was significantly higher among males of  $\leq 1$ -6 month age than females of the same age. Moreover, there was a significant effect of age on the infection rate of *Cryptosporidium*, with the noteworthy prevalence in lambs of  $\leq 1$  to <6 months of age (45.16%).

PCR could amplify 830 bp region of 18 SrRNA gene (Fig. 2). PCR-RFLP analysis of 830 bp region using restriction enzymes *SspI* of nested PCR products uncovered three distinct bands at 440, 267 and 106 bp regions proposing that the species was either *C. parvum* or *C. hominis*, while the band patterns of 454, 384 bp were predictable with *C. ubiquitum* (Fig. 3). To discriminate between *C. hominis* and *C. parvum*, the nested PCR products of all species were further digested with *VspI*. The band's profiles were predictable with *C. parvum* (629, 115bp) and PCR products of 461, 169, 115 bp were consistent to *C. ubiquitum* patterns (Fig. 4). The correlation of PCR-RFLP sequence (sequence 1 and 2) with the database in the Gen. Bank and phylogenetic analysis permitted recognizing them as *C. parvum* and *C. ubiquitum* (Fig. 5). The detected sequences were submitted in the gene bank under accession numbers KU715980 and KU715981.

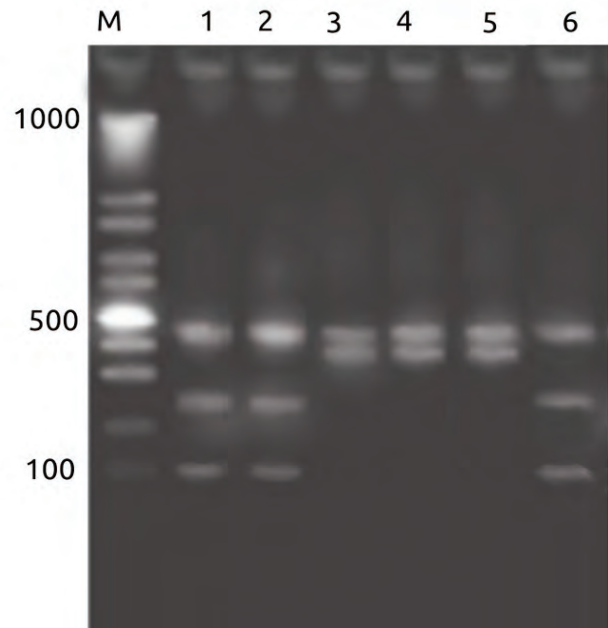


Fig.3. PCR- RFLP of *Cryptosporidium* species using *SspI* enzyme; Lane: 1, 2, 6 *C. parvum* / *C. hominis*, Lane 3,4,5: *C. ubiquitum*.

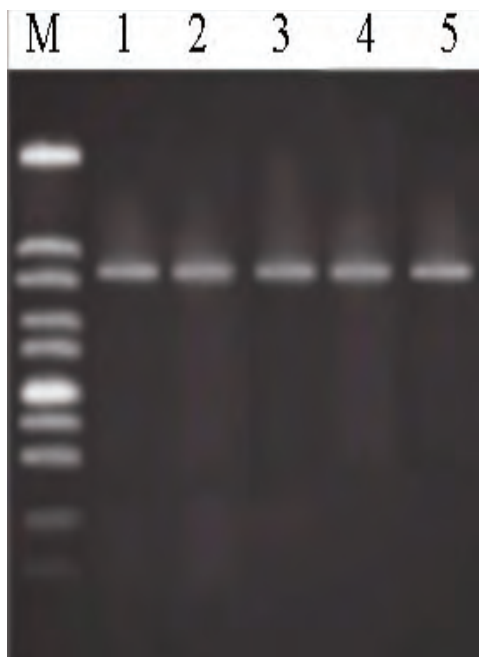


Fig .2. Lane 1-5, Amplified 830 bp of 18s rRNA gene for *Cryptosporidium*.

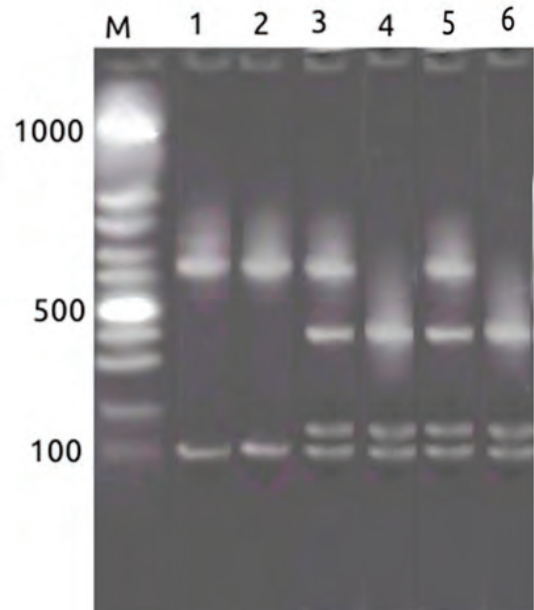


Fig. 4. PCR- RFLP of *Cryptosporidium* species using *VspI* enzyme; Lane 1,2: *C. parvum*, Lane 3,5: *C. parvum* + *C. ubiquitum*, Lane 4,6: *C. ubiquitum*.

Table 1. Frequency of *Cryptosporidium* oocysts detection in feces of sheep of different sex and age

| Groups  | Age (month)           | NO. of examined animals |      |        | Positive                 |                          | Mean (%)           |
|---------|-----------------------|-------------------------|------|--------|--------------------------|--------------------------|--------------------|
|         |                       | Total                   | Male | Female | Male                     | Female                   |                    |
| Group1  | $\leq 1$ to <6 months | 124                     | 32   | 92     | 16 (50.00) <sup>cB</sup> | 40 (43.48) <sup>cA</sup> | 45.16 <sup>c</sup> |
| Group2  | 6-12                  | 124                     | 44   | 80     | 12 (27.27) <sup>bA</sup> | 20 (25.00) <sup>bA</sup> | 25.81 <sup>b</sup> |
| Group 3 | >12                   | 184                     | 48   | 136    | 8 (16.67) <sup>aA</sup>  | 16 (11.76) <sup>aA</sup> | 13.04 <sup>a</sup> |
| Total   | -                     | 432                     | 124  | 308    | 36 (29.03) <sup>A</sup>  | 76 (24.68) <sup>A</sup>  | -                  |

<sup>a,b,c</sup> Values containing the same superscript are not significant ( $P>0.5$ ) within the same column.

<sup>A,B,C</sup> Values containing the same superscript are not significant ( $P>0.5$ ) within the same row.

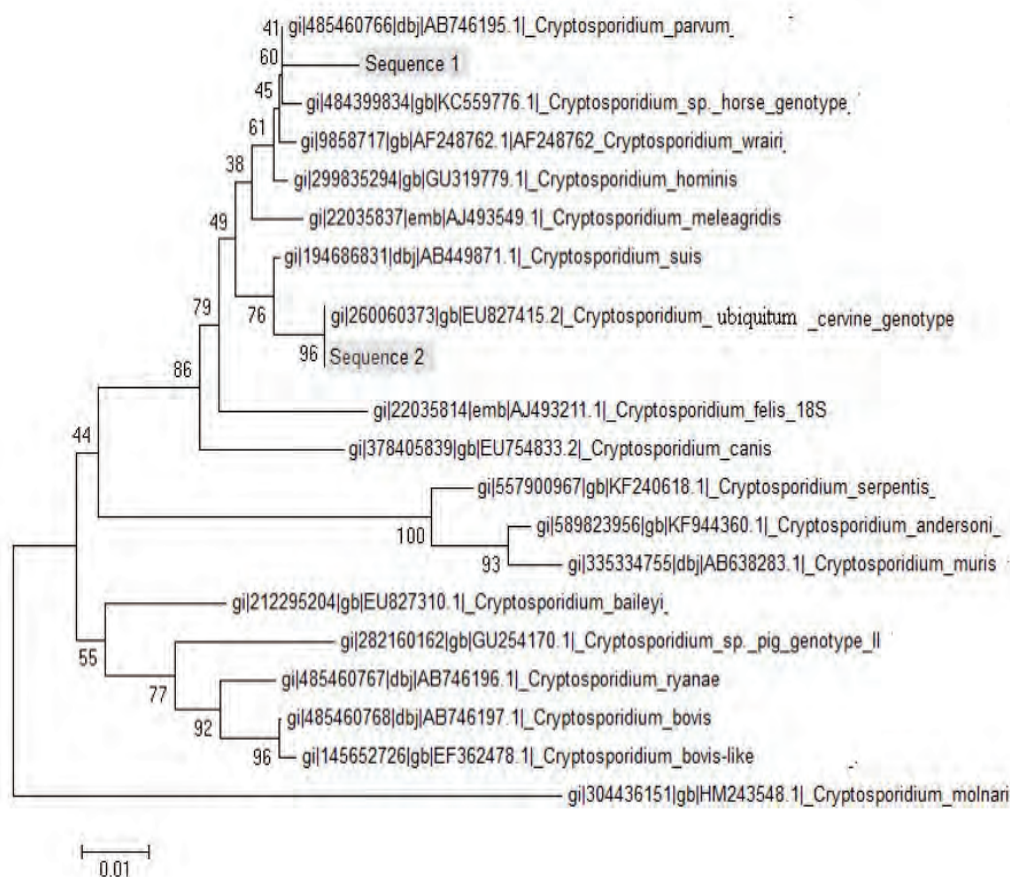


Fig. 5. A Phylogenetic relationship estimated by the Maximum Likelihood with Tamura-Nei model shows the phylogenetic comparison of the available 18S rRNA sequences from respective *Cryptosporidium* isolates with two species detected in this study (sequence 1 and sequence 2).

Table 2: *Cryptosporidium* species identified in sheep with regard to age

| <i>Cryptosporidium</i> sp.             | Group1                   | Group2                   | Group3                  | Mean               |
|--|--------------------------|--------------------------|-------------------------|--------------------|
|  | (≤1 to <6 months)        | (6-12 months)            | (>12 months)            |                    |
|  | Infected /124            | infected/124             | Infected/184            |                    |
| <i>C. parvum</i>                       | 20 (16.13) <sup>bB</sup> | 8 (6.45) <sup>aA</sup>   | 8 (4.35) <sup>abA</sup> | 8.98 <sup>b</sup>  |
| <i>C. ubiquitum</i>                    | 28 (22.58) <sup>cC</sup> | 20 (16.13) <sup>bB</sup> | 16 (8.70) <sup>bA</sup> | 15.80 <sup>c</sup> |
| <i>C. parvum</i> + <i>C. ubiquitum</i> | 8 (6.45) <sup>aB</sup>   | 4 (3.23) <sup>aAB</sup>  | 0 (0.00) <sup>aA</sup>  | 3.23 <sup>a</sup>  |
| <b>Mean</b>                            | <b>15.05<sup>B</sup></b> | <b>8.60<sup>A</sup></b>  | <b>4.35<sup>A</sup></b> | <b>-</b>           |

<sup>a,b,c</sup> Values containing the same superscript are not significant (P>0.5) within the same column.

<sup>A,B,C</sup> Values containing the same superscript are not significant (P>0.5) within the same row.

The infection rate varied significantly (P<0.05) among the examined groups and *C. ubiquitum* was the most prevailing species. Mostly, *Cryptosporidium* sp. prevailed at age group of ≤1-6 months when contrasted with the other two age groups (6-12 and >12 months). A significant variation was recorded between 1<sup>st</sup> and 2<sup>nd</sup> age group and also between the 1<sup>st</sup> and 3<sup>rd</sup> age group for the prevalence of *C. parvum* (P<0.05). *C. ubiquitum* was more pervasive at age group of ≤1-6 months (22.58%). Mixed infection with *C. parvum* and *C. ubiquitum* showed no significant variation between 1<sup>st</sup> and 2<sup>nd</sup> age group, but an efficient significance was found between 1<sup>st</sup> and 3<sup>rd</sup> age groups (P<0.05) (Table 2).

Scanning electron microscopy of the intestine showed the presence of gamont like stage measuring 6.5X5 μm and fully

developed thin wall oocysts measuring 2-5 μm in diameter, located in the intestine and attached to the microvillus brush border of the epithelium of the small intestine with velvet appearance of the intestinal microvilli surrounding the parasite (Fig. 1.C). Thick wall oocysts were also seen with increasing of the globule secretion of the goblet cell due to distortion in the intestinal glands and villi (Fig. 1. D).

### Discussion

*Cryptosporidium* infections of sheep is widely distributed in several countries, the prevalence rate ranged from 1.6 to 77.4% using microscopy and molecular assay (Santin *et al.*, 2007; Fiuza *et al.*, 2011). Few studies had been carried out in

Egypt and to date; most of *Cryptosporidium* studies in sheep focused on traditional microscopic examination and lacked the more sensitive technique. In the present study, the overall prevalence of *Cryptosporidium* was 25.93% among the analyzed sheep of different age and sex groups. This result was broadly consistent with other studies (Santin et al., 2007 and Connelly et al., 2013). The prevalence of *Cryptosporidium* was higher than that were formerly found in Egypt (2.5% and 32.2%) (Mahfouz et al., 2014; Helmy et al., 2012 respectively), in Iran (11.3%) (Heidari and Gharakhani, 2012), in Iraq (13.3%) (Mahdi and Ali, 2002), in Tunisia (11.2%) (Soltane et al., 2007), in Belgium (13.1%) (Geurden et al., 2008). Higher results were recorded in Mexico (67.5%) (Romero-Salas et al., 2016), in Spain (31-59%) (Causape et al., 2002; Castro-Hermida et al., 2007). However, the comparison of prevalence in different geographical locations must be limited by the animal characters, breeds and rearing condition. PCR-RFLP method recognized two types of *Cryptosporidium* infecting sheep: *C. parvum* and *C. ubiquitum* (Cervine genotype). Those were previously encountered by (Mirhashemi et al., 2016). Nevertheless, *C. xiaoi* was the only species encountered in sheep in Egypt (Mahfouz et al., 2014). This recorded difference may be ascribed to difference in sheep breeds or sample size. In accordance with the herein results, *C. ubiquitum* was the most predominant species among different ages in Iran (Geurden et al., 2008), in China (Wang et al., 2010; Shen et al., 2011). The occurrence of *C. parvum* among sheep population seems to be consistent with those of some reports that review the role of sheep as a zoonotic source of *Cryptosporidium* in human (El-Sherbini and Mohamed, 2006; Paoletti et al., 2009; Helmy et al., 2012; Yang et al., 2014). The zoonotic significance of *C. parvum* and *C. ubiquitum* was in the past perceived, recommending that this animal might assume bigger parts as reservoir species than that already thought, posing a substantial public health danger (Santin and Fayer, 2007). Thus, an important public health control and prevention strategies must be considered for the surveillance of *Cryptosporidium* in sheep. The higher infection rate among males than female sheep recorded in the current study was predictable with that of Heidari and Gharakhani (2012); Jafari et al. (2013) and Romero-Salas et al. (2016). Yet, it is inverse to an alternate investigation (Regassa et al., 2013; Gharekhani et al., 2014) in Iran. The purpose behind this slight uniqueness is not known, in spite of the fact that this result could be credited to the standard routine of having an unequal female: male ratio in a herd and the more contact of males to disease than females. It could likewise be related to host genetics, physiology and immunology or environmental and management practices. The actual mechanism of the influence of these factors are not understood and need clarification (Ayinmode et al., 2010). Concerning age, the present study demonstrates a statistical difference among different age groups for *Cryptosporidium* infection, where the most noteworthy infection rate was in sheep ( $\leq 1$  to  $< 6$  months of age). This corresponded with that recorded by (Ayinmode et al., 2010; Gharekhani et al., 2014; Romero-Salas et al., 2016). The clinical manifestations of cryptosporidiosis agreed that noticed by Angus et al. (1982)

Electron microscopic study showed different stages of *Cryptosporidium* were found mainly attached the border of the intestinal epithelial. The common site the parasite was mainly the small intestine. This was in accordance with Caccio and Widmer (2014), who stated that *C. parvum* and *C. ubiquitum* mainly infecting and producing pathological lesions in the small intestine.

## Conclusion

This study is the first to distinguish *Cryptosporidium* sp.

infecting sheep in Qalyubia governorate, Egypt. The recorded *Cryptosporidium* sp. are *C. parvum* and *C. ubiquitum*, which have critical zoonotic significance. This finding proposes that sheep must be considered as a huge public health risk and as a potential wellspring of human cryptosporidiosis. The reconnaissance of zoonotic cryptosporidiosis must be set up to prevent human disease and to assess prospective infection when applying control programs.

## Acknowledgement

The author appreciates the assistance and backing of Dr. Amal Mohamed El-Asaly, Assistant Professor of Fish diseases and Management and member of the Biotechnology Center in the Faculty of Veterinary Medicine, Benha University, Egypt.

## References

- Angus, K.W., Appleyard, W.T., Menzies, J. D., Campbell, I., Sherwood, D., 1982. An outbreak of diarrhoea associated with cryptosporidiosis in naturally reared lambs. *Veterinary Record* 110, 129-130.
- Ayinmode, A., Fagbemi, O.B., Ayinmode, B.A.F., 2010. Prevalence of *Cryptosporidium* infection in cattle from South Nigeria. *Veterinary Archives* 80, 723-731.
- Bensch, S., Stjernman, M., Hasselquist, D., Ostman, O., Hansso, B., Westerdaal H., Pinheiro, R.T., 2000. Host specificity in avian blood parasites: a study of Plasmodium and Haemoproteus mitochondrial DNA amplified from birds. *Proceedings of the Royal Society B: Biological Sciences* 267, 1583-1589.
- Bukhari, Z., Smith, H.V., 1997. *Cryptosporidium parvum*: oocyst excretion and viability patterns in experimentally infected lambs. *Epidemiology and Infection* 119, 105-113.
- Caccio, S.M., Widmer, G., 2014. *Cryptosporidium: Parasite and Disease*. Springer, Wien, Heidelberg, New York, Dordrecht, London. Cama.
- Casemore, D.P., Armstrong M., Sands, R., 1985. Laboratory diagnosis of cryptosporidiosis. *Journal of Clinical Pathology* 38, 1337-13341.
- Castro-Hermida, J.A., González-Warleta, M., Mezo, M., 2007. Natural infection by *Cryptosporidium parvum* and *Giardia duodenalis* in sheep and goats in Galicia (NW Spain). *Small Ruminant Research* 72, 96-100.
- Causape, A.C., Quilez, J., Sanchez-Acedo, C., Del Cacho E., Lopez-Bernad, F., 2002. Prevalence and analysis of potential risk factors for *Cryptosporidium parvum* infection in lambs in Zaragoza (northeastern Spain). *Veterinary Parasitology* 104, 287-98.
- Connelly L., Craig, B.H., Jones B., Alexander, C.L. 2013. Genetic diversity of *Cryptosporidium* spp. within a remote population of Soay Sheep on St. Kilda Islands, Scotland. *Applied and Environmental Microbiology* 79, 2240-22446.
- El-Sherbini, G., Mohammad, K., 2006. Zoonotic cryptosporidiosis in man and animal in farms, Giza Governorate, Egypt. *Journal of the Egyptian Society of Parasitology* 36(2 Suppl), 49-58.
- Fayer, R., Santin, M., 2009. *Cryptosporidium xiaoi* n. sp. (Apicomplexa: Cryptosporidiidae) in sheep (*Ovis aries*). *Veterinary Parasitology* 164, 192-200.
- Féres, F.C., Lombardi, A.L., Carvalho, M.P.P., Mendes, L.C.N., Peiró, J.R., Cadioli, F.A., Meireles, M.V., Perri, S.H.V., Feitosa, F. L.F., 2009. Ocorrência e caracterização molecular de *Cryptosporidium* em cordeiros. *Arquivo Brasileiro de Medicina Veterinária e Zootecnia* 61, 1002-1005.
- Fiuzza, V.R., Cosendey, R.I., Frazão-Teixeira, E., Santín, M., Fayer, R., de Oliveira, F., 2011. Molecular characterization of *Cryptosporidium* in Brazilian sheep. *Veterinary Parasitology* 175, 360-362.
- Foreyt, W.J., 1990. Coccidiosis and cryptosporidiosis in sheep and goats. *Veterinary Clinics of North America: Food Animal Practice* 6, 655-670.
- Geurden, T., Thomas, P., Casaert, S., Verducysse, J., Claerebout, E., 2008. Prevalence and molecular characterisation of *Cryptosporidium* and *Giardia* in lambs and goat kids in Belgium. *Veterinary Parasitology* 155, 142-5.

- Gharekhani, J., Heidari, H., Youssefi, M., 2014. Prevalence of *Cryptosporidium* infection in sheep in Iran. *Türkiye Parazitoloji Dergisi* 38, 22-5.
- Heidari, H., Gharakhani, J., 2012. Prevalence of *Cryptosporidium* infection in cattle from South Nigeria. *Scientific Journal of Hamadan University of Medical Sciences* 19, 67-74.
- Helmy, A.Y., Krücken, J., Nöckler, K., Zessin, K.H. 2012. Molecular epidemiology of *Cryptosporidium* in livestock animals and humans in the Ismailia province of Egypt. *Veterinary Parasitology* 193 (1-3).
- Jafari, R., Maghsood, A.H., Fallah, M., 2013. Prevalence of *Cryptosporidium* infection among livestock and humans in contact with livestock in Hamadan district, Iran. *Journal of Research in Health Sciences* 13, 86-89.
- Kuo, J., 2007. *Electron Microscopy, methods and protocols*. 2nd Ed., Humana Press Inc. New Jersey, pp 449.
- Mahdi, N.K., Ali, N.H., 2002. Cryptosporidiosis among animal handlers and their livestock in Basrah, Iraq. *East African Medical Journal* 79, 550-553.
- Mahfouz, M.E., Mira, N., Amer, S., 2014. Prevalence and genotyping of *Cryptosporidium* spp. in farm animals in Egypt. *The Journal of Veterinary Medical Science* 76, 1569-1575.
- Majewska, A.C., Werner, A., Sulima, P., Luty, T., 2000. Prevalence of *Cryptosporidium* in sheep and goats bred on five farms in west-central region of Poland. *Veterinary Parasitology* 89, 269-275.
- Mirhashemi, M.E., Zintl, A., Grant, T., Lucy, F., Mulcahy, G., Waal, T.D., 2016. Molecular epidemiology of *Cryptosporidium* species in livestock in Ireland. *Veterinary Parasitology* 216, 18-22.
- Paoletti, B., Giangaspero, A., Gatti, A., Iorio, R., Cembalo, D., Milillo P., Traversa, D., 2009. Immunoenzymatic analysis and genetic detection of *Cryptosporidium parvum* in lambs from Italy. *Experimental Parasitology* 122, 349-52.
- Quilez, J., Torres, E., Chalmers, R.M., Hadfield, S.J., Del E., Cacho-Sanchez-Acedo, C., 2008. *Cryptosporidium* genotypes and subtypes in lambs and goat kids in Spain. *Applied and Environmental Microbiology* 74, 6026-6031.
- Regassa, A., Abunna, F., Abebe, R., Beyene, D., Megersa, B., Debela, E., Asmare, K., Skierve, E., 2013. *Cryptosporidium* in Calves, Lambs and Kids at Haramaya, eastern Ethiopia Alemayehu, Ethiopian Veterinary Journal 17 (1), 81-94.
- Romero-Salas, D., Alvarado-Esquivel, C., Cruz-Romero A., Aguilar-Dominguez, M., Ibarra-Priego, N., Merino-Charrez, O.J., Pérez de León, A.A., Hernández-Tinoco, J., 2016. Prevalence of *Cryptosporidium* in small ruminants from Veracruz, Mexico. *BMC Veterinary Research* 12, 1-6.
- Santin, M., Fayer, R., 2007. Intragenotypic variations in the *Cryptosporidium* sp. cervine genotype from sheep with implications for public health. *Journal of Parasitology* 93, 668-672.
- Santin, M., Trout, J.M., Fayer, R., 2007. Prevalence and molecular characterization of *Cryptosporidium* and *Giardia* species and genotypes in sheep in Maryland. *Veterinary Parasitology* 146, 17-24.
- Shen, Y., Yin, J., Yuan, Z., Lu, W., Xu, Y., Xiao, L., Cao, J., 2011. The identification of the *Cryptosporidium* ubiquitum in pre-weaned Ovines from Aba Tibetan and Qiang autonomous prefecture in China. *Biomedical and Environmental Sciences* 24, 315-320.
- Soltane, R., Guyot, K., Dei-Cas, E., Ayadi A., 2007. Prevalence of *Cryptosporidium* spp. (Eucoccidiorida: Cryptosporiidae) in seven species of farm animals in Tunisia. *Parasite* 14, 335-8.
- Steel, R., Torrie, J., Dickey, D., 1997. *Principles and procedures of Statistics: A Biometrical Approach*, 3rd ed., McGraw-Hill, New York, NY.
- Wang, Y., Feng, Y., Cui, B., Jian, F., Ning, C., Wang, R., Zhang, L., Xiao L., 2010. Cervine genotype is the major *Cryptosporidium* genotype in sheep in China. *Parasitology Research* 106, 341-347.
- Xiao, L., Morgan, U.M., Imor, J.L., Escalante, A., Arrowood, M., Shulaw, W., Thompson, R.C., Fayer R., Lal A.A., 1999. Genetic diversity within *Cryptosporidium parvum* and related *Cryptosporidium*-species. *Applied and Environmental Microbiology* 65, 3386-3391.
- Yang, R., Jacobson, C., Gardner, G., Carmichael, I., Campbell, A.J., Ng-Hublin, J., Ryan, U., 2014. Longitudinal prevalence, oocyst shedding and molecular characterisation of *Cryptosporidium* species in sheep across four states in Australia. *Veterinary Parasitology* 200, 50- 58.
- Yang, R., Jacobson, C., Gordon, C., Ryan, U., 2009. Prevalence and molecular characterisation of *Cryptosporidium* and *Giardia* species in pre-weaned sheep in Australia. *Veterinary Parasitology* 161, 19-24.