

Citation for published version: Gonzalez, JT 2019, 'Symposium overview: Fructose in Physiology', *Journal of Physiology*, vol. 597, no. 14, pp. 3253-3525. https://doi.org/10.1113/JP278267

DOI: 10.1113/JP278267

Publication date: 2019

Document Version Peer reviewed version

Link to publication

This is the peer reviewed version of the following article: Gonzalez, J. T. (2019), Symposium overview: fructose in physiology. J Physiol, 597: 3523-3525., which has been published in final form at https://doi.org/10.1113/JP278267. This article may be used for non-commercial purposes in accordance with Wiley. Torme and Conditions for Self Archiving Wiley Terms and Conditions for Self-Archiving.

University of Bath

General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

1 Symposium overview: Fructose in Physiology

- 2 Author: Javier T. Gonzalez
- 3 Affiliation: Department for Health, University of Bath, Bath, UK.
- 4 Email: <u>J.T.Gonzalez@bath.ac.uk</u>
- 5

One of the first uses of the term "fructose" was in 1857 by William Allen Miller FRS, 6 7 and it was already known then that fructose was a distinctive carbohydrate, 8 characterised by sweet taste (Miller, 1957). In most circumstances dietary free sugars 9 can also be classified as fructose-containing carbohydrates (e.g. sucrose, highfructose corn syrup), and the fructose component of these sugars is thought to be 10 primarily responsible for unique metabolic effects. The role of sugars in the diet could 11 12 be viewed as contentious, with many arguing that dietary sugar is the cause of type 2 13 diabetes and obesity (Bray & Popkin, 2014), whereas others posit that dietary sugars are innocuous to healthy individuals (Archer, 2018). One reason for this confusion and 14 conflict is that much evidence presented is observational and/or based upon self-15 report assessments of dietary intake or physical activity. This is a problem, because 16 observational data can never definitely establish cause-and-effect and can lead to 17 18 spurious, misleading correlations. Furthermore, self-report methods are subject to observation and reporting biases. Herein lies the potential for a physiological approach 19 20 to solve some of this confusion and conflict.

By understanding mechanistic links, we can explain some apparent 21 22 discrepancies in observational data, and also take a step towards establishing 23 causality. Plausible physiological links can increase confidence in causality of a 24 behaviour when randomized controlled trials with hard endpoints would be considered 25 unethical or impossible to perform. For example, it might be deemed unethical to 26 randomise people to high versus low fructose intakes for decades to establish whether 27 fructose intake causes type 2 diabetes or cardiovascular disease. However, by assessing the effects of fructose intake of physiological process that underlie disease 28 risk, shorter-term studies can be used to establish causal effects of fructose intake, 29 without necessarily affecting long-term health risk. 30

31 "Fructose in Physiology: Friend or Foe" was the title of a Symposium delivered 32 at Europhysiology 2018 in London (UK), and was supported by *The Journal of* 33 *Physiology*. The aim of this symposium was to bring together leading researchers 34 across various career stages to discuss the physiology of fructose metabolism in health and disease. In doing so, this symposium highlighted the potential mechanisms
by which fructose may exert metabolic effects within specific populations, thereby
overcoming some of the confusion around fructose and health.

4 Pinnick and Hodson (2019) describe the potential tissue- and sex-specific effects of fructose metabolism. Short-term (<7 days) high-fructose intake can increase 5 plasma triglyceride concentrations and intrahepatic fat content, which are implicated 6 7 in metabolic disease risk (Pinnick & Hodson, 2019). The mechanisms by which high 8 fructose intake increases plasma triglyceride concentrations includes hepatic de novo 9 lipogenesis (DNL) and fatty acid oxidation (Pinnick & Hodson, 2019). A novel focus was the discussion of the potential effects of fructose on adipose tissue metabolism, 10 which could be direct or indirect (Pinnick & Hodson, 2019). The classical view is that 11 the splanchnic tissues are the primary site of fructose metabolism (Gonzalez & Betts, 12 2018), and therefore adipose tissue is unlikely to be directly affected by fructose 13 intake. Nevertheless, fructose could indirectly affect adipose tissue metabolism via 14 increased plasma lactate concentrations following fructose consumption (Liu et al., 15 2009; Gonzalez et al., 2015). Furthermore, there is potential for some direct effects of 16 17 fructose on adipose tissue, since it has been recently estimated that ~15% of a 30-g 18 oral fructose load can escape first-pass splanchnic metabolism and thereby be exposed to peripheral tissues (Francey et al., 2019). In addition to evidence that 19 20 adipose tissue expresses the fructose-specific transporter, GLUT5, it is plausible that 21 fructose could have some direct effects on adipose tissue, and this will be an important 22 avenue for future research (Pinnick & Hodson, 2019). With respect to sex-specific responses, there is some evidence that males may display greater metabolic 23 24 perturbations to high fructose intake when compared to females, including increased 25 incorporation of fructose carbons into very low-density lipoprotein (VLDL)-TAG 26 palmitate (reflective of hepatic DNL), greater suppression of fat oxidation, and 27 increased basal endogenous glucose production (Pinnick & Hodson, 2019). However, other work has shown that females displayed higher hepatic DNL than males, when 28 assessed using deuterium oxide (Low et al., 2018). The discrepancies between 29 studies may be explained by doses of fructose ingested (absolute vs normalised to 30 fat-free mass), the method of assessing hepatic DNL, or participant characteristics 31 and background diet. Accordingly, fructose can clearly stimulate lipogenesis in 32 33 hepatocytes, but there is a need to further understand the sex-specific effects of fructose intake on metabolism and health. 34

1 Von Holstein-Rathlou and Gillum (2019) discuss a key potential regulator of 2 fructose intake, fibroblast growth factor 21 (FGF21). FGF21 is a hepatically-derived 3 hormone that, in mice, can be produced in response to low-protein and ketogenic 4 diets, fructose feeding and ethanol (von Holstein-Rathlou & Gillum, 2019). It has also 5 been shown that FGF21 preferentially inhibits ad libitum consumption of sugars and ethanol in mice, without affecting the intake of other dietary nutrients such as non-6 7 sugar carbohydrates, fat and protein, thereby exerting negative-feedback (von 8 Holstein-Rathlou & Gillum, 2019). In humans, ingestion of sugars and ethanol can also 9 stimulate FGF21 secretion, and genetic variants in the FGF locus have been associated with reported intakes of sweet foods (Søberg et al., 2017). The potential 10 mechanisms by which FGF21 is thought to regulate feeding behaviours is thought to 11 12 involve the activation of the FGF21 receptor complex (comprising FGF receptor 1c 13 and beta-klotho) in the paraventricular nucleus of the hypothalamus (von Holstein-Rathlou & Gillum, 2019). This opens up the intriguing possibility of reducing free-living 14 15 sugar (and ethanol) intakes by treatment with FGF21 or by making use of other strategies that can increase endogenous FGF21 production. 16

17 Fuchs et al. (2019) describe how athletes can exploit some of the metabolic 18 effects of fructose to benefit endurance performance and recovery. Intestinal fructose absorption primarily occurs via GLUT5. This contrasts with glucose, which is primarily 19 20 absorbed via the sodium-dependent glucose transporter, SGLT1 (Fuchs et al., 2019). Since SGLT1 is thought to be saturable at a rate of 1 g/min this can limit the amount 21 22 of exogenous carbohydrate that athletes can ingest and metabolise during exercise. However, by combining fructose with glucose it is possible to make use of both of 23 24 these intestinal transport pathways and thereby deliver more exogenous carbohydrate 25 to the circulation, whilst also decreasing gastrointestinal discomfort associated with 26 ingestion of large amounts of carbohydrate during exercise (Gonzalez et al., 2015; 27 Fuchs et al., 2019). A higher availability of carbohydrates during exercise can have performance benefits in many endurance sports, thereby highlighting a potential 28 29 beneficial role of fructose-containing carbohydrates. Furthermore, rapid restoration of depleted glycogen stores is a key factor dictating recovery time in multi-stage 30 endurance events. Since fructose can potently stimulate hepatic glycogen synthesis 31 (Fuchs et al., 2016) there is potential for fructose-containing carbohydrates to 32 33 accelerate recovery. Indeed, when the total amount of carbohydrate is matched, the ingestion of fructose-glucose mixtures can double the rate of liver glycogen repletion 34

in recovery from exercise, when compared to glucose-based carbohydrates (Fuchs *et al.*, 2019). Furthermore, ingestion of fructose-containing carbohydrates during recovery from exercise can enhance subsequent endurance running capacity, when compared to glucose-based carbohydrates (Maunder *et al.*, 2018). Therefore, at least for specific scenarios, fructose-containing carbohydrate can be useful for athletic performance.

7 Whilst fructose ingestion may provide a benefit to certain athletic events, a 8 reasonable question to ask is whether such fructose intake is detrimental to the health 9 of athletes. Tappy and Rosset (2019) describe the potential for physical activity to protect against the negative metabolic effects of high-fructose intake, independent 10 from total energy balance (i.e. when controlling for negative energy balance induced 11 by exercise). Whilst high-fructose intake can increase hepatic DNL, intrahepatic fat 12 content, hepatic insulin resistance and plasma triglyceride concentrations in sedentary 13 individuals, all these responses can be prevented under conditions of high physical 14 15 activity (Tappy & Rosset, 2019). Fructose ingested during conditions of high energy output is thought to be directed more to lactate and glucose for utilisation as a fuel by 16 17 skeletal muscle, and less to triglycerides via DNL. The authors therefore speculate 18 that the negative metabolic health consequences of high-fructose intake occur when fructose intake exceeds the capacity of the liver to release lactate and glucose for 19 20 skeletal muscle to utilise (Tappy & Rosset, 2019). This may be more likely to occur 21 under conditions of low energy output, where skeletal muscle utilisation of circulating 22 glucose and lactate is low. The authors propose that this could contribute to regulating hepatic fructose metabolism via a feedback mechanism that is yet to be definitely 23 24 established.

25 Hengist et al. (2019) discuss a further potential mechanism that could explain 26 the protection against fructose-induced metabolic impairments conferred by high 27 levels of physical activity. Hepatic glycogen content plays a key role in regulating hepatic lipid metabolism by acting on both *DNL* and on hepatic fatty acid oxidation 28 (Hengist et al., 2019). Hengist et al. (2019) discuss the evidence that suggests 29 "pushing" glucose into the liver and saturating liver glycogen concentrations increases 30 31 DNL and hypertriglyceridaemia, whereas increasing the inherent capacity for liver 32 glycogen storage does not detrimentally alter plasma triglyceride concentrations. This 33 is consistent with the notion that net lipid synthesis is exacerbated when glycogen stores are saturated. Therefore, under conditions where hepatic glycogen stores are 34

low, or are undergoing an increased rate of turnover, there is likely to be lower rates 1 2 of hepatic DNL and increased rates of hepatic fatty acid oxidation. The net result is 3 less lipid synthesis. Furthermore, this may contribute to the mechanisms explaining why fructose can stimulate lipid synthesis to a greater extent than glucose, since 4 5 fructose potently stimulates hepatic glycogen synthesis at rest and post-exercise (Petersen et al., 2001; Fuchs et al., 2016). Hepatic glycogen status may thereby 6 7 provide a key link between the energy status of an individual and the metabolic 8 responses to fructose intake.

9 In summary, this collection of review articles illuminates several key aspects of fructose metabolism. It is clear that excessive fructose intakes in sedentary individuals 10 can induce a number of metabolic effects that may be detrimental to health. Whether 11 males or females are more sensitive to the effects of fructose intake remains to be 12 established. The hormone FGF21 could hold promise in reducing the levels of fructose 13 intake when a high-fructose intake is undesirable and could therefore contribute to 14 15 improvements in metabolic health. The metabolic effects of fructose ingestion can be utilised to benefit endurance performance and recovery in athletes, and these athletes 16 17 seem to be protected against the negative metabolic effects of high-fructose intake. 18 The mechanisms underlying exercise-induced protection against these metabolic effects remains to be established but may involve the greater conversion of fructose 19 20 into glucose and lactate for oxidation rather than conversion into lipid, and these processes could be regulated by hepatic glycogen content. These physiological 21 22 mechanisms provide a better understanding of why specific populations seem to be 23 more or less vulnerable to high-fructose intakes and can be targeted for improving 24 metabolic health.

25

29

33

26 References

- Archer E. (2018). In defense of sugar: A critique of diet-centrism. *Progress in Cardiovascular Diseases* 61, 10-19.
- Bray GA & Popkin BM. (2014). Dietary sugar and body weight: have we reached a
 crisis in the epidemic of obesity and diabetes?: health be damned! Pour on
 the sugar. *Diabetes Care* 37, 950-956.
- Francey C, Cros J, Rosset R, Crézé C, Rey V, Stefanoni N, Schneiter P, Tappy L &
 Seyssel K. (2019). The extra-splanchnic fructose escape after ingestion of a
 fructose-glucose drink: An exploratory study in healthy humans using a dual
 fructose isotope method. *Clin Nutr ESPEN* 29, 125-132.

1	
1 2 3 4 5 6	Fuchs CJ, Gonzalez JT, Beelen M, Cermak NM, Smith FE, Thelwall PE, Taylor R, Trenell MI, Stevenson EJ & van Loon LJ. (2016). Sucrose ingestion after exhaustive exercise accelerates liver, but not muscle glycogen repletion compared with glucose ingestion in trained athletes. <i>J Appl Physiol (1985)</i> 120, 1328-1334.
7 8 9 10 11	Fuchs CJ, Gonzalez JT & van Loon LJC. (2019). Fructose co-ingestion to increase carbohydrate availability in athletes <i>Journal of Physiology</i> . DOI: 10.1113/JP277116.
11 12 13 14	Gonzalez JT & Betts JA. (2018). Dietary Fructose Metabolism By Splanchnic Organs: Size Matters. <i>Cell Metab</i> 27, 483-485.
15 16 17 18 19	Gonzalez JT, Fuchs CJ, Smith FE, Thelwall PE, Taylor R, Stevenson EJ, Trenell MI, Cermak NM & van Loon LJ. (2015). Ingestion of glucose or sucrose prevents liver but not muscle glycogen depletion during prolonged endurance-type exercise in trained cyclists. <i>Am J Physiol Endocrinol Metab</i> 309 , E1032-1039.
20 21 22	Hengist A, Koumanov F & Gonzalez JT. (2019). Fructose and metabolic health: governed by hepatic glycogen status? <i>J Physiol</i> . DOI: 10.1113/JP277767.
23 24 25 26 27	Liu C, Wu J, Zhu J, Kuei C, Yu J, Shelton J, Sutton SW, Li X, Yun SJ, Mirzadegan T, Mazur C, Kamme F & Lovenberg TW. (2009). Lactate inhibits lipolysis in fat cells through activation of an orphan G-protein-coupled receptor, GPR81. J Biol Chem 284, 2811-2822.
28 29 30	Low W, Cornfield T, Charlton C, Tomlinson J & Hodson L. (2018). Sex differences in hepatic de novo lipogenesis with acute fructose feeding. <i>Nutrients</i> 10 , E1263.
31 32 33 34	Maunder E, Podlogar T & Wallis GA. (2018). Postexercise Fructose-Maltodextrin Ingestion Enhances Subsequent Endurance Capacity. <i>Med Sci Sports Exerc</i> 50, 1039-1045.
35 36 37	Miller W. (1957). Organic Chemistry. In <i>Elements of Chemistry: Theoretical and Practical, Part III</i> , pp. 52-57. John W. Parker and son, London, England.
38 39 40 41	Petersen KF, Laurent D, Yu C, Cline GW & Shulman GI. (2001). Stimulating effects of low-dose fructose on insulin-stimulated hepatic glycogen synthesis in humans. <i>Diabetes</i> 50 , 1263-1268.
42 43 44	Pinnick K & Hodson L. (2019). Challenging metabolic tissues with fructose: tissue- specific and sex-specific responses. <i>The Journal of Physiology</i> . DOI: 10.1113/JP277115.
45 46 47 48 49 50	Søberg S, Sandholt CH, Jespersen NZ, Toft U, Madsen AL, von Holstein-Rathlou S, Grevengoed TJ, Christensen KB, Bredie WLP, Potthoff MJ, Solomon TPJ, Scheele C, Linneberg A, Jørgensen T, Pedersen O, Hansen T, Gillum MP & Grarup N. (2017). FGF21 Is a Sugar-Induced Hormone Associated with Sweet Intake and Preference in Humans. <i>Cell Metab</i> 25 , 1045-1053.e1046.

1 Tappy L & Rosset R. (2019). Metabolic effects of high fructose intake in humans. 2 Journal of Physiology. DOI: 10.1113/JP278246. 3 4 5 von Holstein-Rathlou S & Gillum MP. (2019). Fibroblast growth factor 21: an endocrine inhibitor of sugar and alcohol appetite. J Physiol. DOI: 6 7 10.1113/JP277117. 8 9 10 **Additional information** 11 The author has received research funding from The European Society for Clinical 12 Nutrition and Metabolism (ESPEN), The Rank Prize Funds, Kenniscentrum Suiker & 13 Voeding, Arla Foods Ingredients, the Medical Research Council, and the 14 Biotechnology and Biological Sciences Research Council. The author has also acted 15 as a consultant to PepsiCo and Lucozade Ribena Suntory. 16 17 18 Acknowledgements 19 The author wishes to thank the speakers for taking the time to share their work, as well as others in the field that have contributed to this evidence-base. In addition, 20

21 thanks to *The Journal of Physiology* for supporting this symposium.

22