



RESEARCH PAPER

Operating at the very low end of the crassulacean acid metabolism spectrum: *Sesuvium portulacastrum* (Aizoaceae)

Klaus Winter^{1,*}, Milton Garcia¹, Aurelio Virgo¹ and Joseph A. M. Holtum^{1,2}

¹ Smithsonian Tropical Research Institute, PO Box 0843-03092, Balboa, Ancón, Republic of Panama

² Centre for Tropical Biodiversity and Climate Change, College of Science and Engineering, James Cook University, Townsville 4811, Queensland, Australia

* Correspondence: winterk@si.edu

Received 23 May 2018; Editorial decision 28 November 2018; Accepted 30 November 2018

Editor: Christine Raines, University of Essex, UK

Abstract

Demonstration of crassulacean acid metabolism (CAM) in species with low usage of this system relative to C₃-photosynthetic CO₂ assimilation can be challenging experimentally but provides crucial information on the early steps of CAM evolution. Here, weakly expressed CAM was detected in the well-known pantropical coastal, leaf-succulent herb *Sesuvium portulacastrum*, demonstrating that CAM is present in the Sesuvioideae, the only sub-family of the Aizoaceae in which it had not yet been shown conclusively. In outdoor plots in Panama, leaves and stems of *S. portulacastrum* consistently exhibited a small degree of nocturnal acidification which, in leaves, increased during the dry season. In potted plants, nocturnal acidification was mainly facultative, as levels of acidification increased in a reversible manner following the imposition of short-term water-stress. In drought-stressed plants, nocturnal net CO₂ exchange approached the CO₂-compensation point, consistent with low rates of CO₂ dark fixation sufficient to eliminate respiratory carbon loss. Detection of low-level CAM in *S. portulacastrum* adds to the growing number of species that cannot be considered C₃ plants *sensu stricto*, although they obtain CO₂ principally via the C₃ pathway. Knowledge about the presence/absence of low-level CAM is critical when assessing trajectories of CAM evolution in lineages. The genus *Sesuvium* is of particular interest because it also contains C₄ species.

Keywords: Crassulacean acid metabolism, CO₂ assimilation, C₄ photosynthesis, facultative CAM, salt tolerance, *Sesuvium portulacastrum*, succulence.

Introduction

The Aizoaceae, a family of about 1600 species in the order Caryophyllales (Hartmann, 2017), contains more species with succulent leaves than any other eudicot family (Hernández-Ledesma *et al.*, 2015). In three of the four sub-families of the Aizoaceae, the Aizooideae, Mesembryanthemoideae, and the Ruschioideae, succulence is associated with multiple lineages of crassulacean acid metabolism (CAM), a water-use efficient pathway of photosynthesis (von Willert *et al.*, 1977, 1992; Smith and Winter, 1996; Arakaki *et al.*, 2011).

The Sesuvioideae, the earliest diverging sub-family (Klak *et al.*, 2003; Bohley *et al.*, 2015), is unique in the Aizoaceae in that it is the only sub-family within which CAM has not yet been demonstrated unequivocally (Bohley *et al.*, 2015) and it is the only sub-family that contains species with C₄ photosynthesis, which is present in the genera *Sesuvium* (Bittrich, 1990; Bohley *et al.*, 2015), *Trianthema*, and *Zaleya* (Carolin *et al.*, 1978). Within *Sesuvium*, two clades have been recognized, one composed of species with C₄-type anatomy, C₄ biochemistry,

and carbon isotope values of between -13.3 and -11.2‰ [*S. congeste*, *S. crithmoides*, *S. humifusum* (formerly *Cypselea humifusum*), *S. mesembryanthemoides*, and *S. sesuoides*] (Bohley *et al.*, 2015, 2017), and one with isotope values of between -27.7 and -25.3‰ that are characteristic of plants with C_3 photosynthesis (*S. ayresii*, *S. dystylum*, *S. maritimum*, *S. microphyllum*, *S. portulacastrum*, *S. sessile*, and *S. verrucosum*) (Bohley *et al.*, 2015). For one species, *S. edmonstonei*, native to the Galapagos, a reported isotope value of -21.5‰ is intermediate between those characteristic of C_3 plants and those of plants with C_4 photosynthesis or full CAM (Winter and Holtum, 2002; Bohley *et al.*, 2015; Alonso-Cantabrana and von Caemmerer, 2016), although for plants of a northern Venezuelan population the $\delta^{13}\text{C}$ value is -24‰ (Medina *et al.*, 2008).

Despite a lack of evidence for CAM-type CO_2 dark fixation in the Sesuvioideae, the ~ 50 species in this small subfamily [*Sesuvium*, 14 spp. (Bohley *et al.*, 2017); *Trianthema*, ~ 20 spp. (Hartmann, 2017); *Tribulocarpus*, two spp. (Thulin *et al.*, 2012); *Zaleya*, six spp. (Gilbert *et al.*, 2000)] are mostly succulent-leaved and drought-tolerant, traits common in CAM plants (Ogburn and Edwards, 2010). Bearing in mind recent reports of low-level CAM in other small succulent-leaved herbs within the Caryophyllales that exhibit C_3 - or C_4 -type isotopic signatures [e.g. *Calandrinia* (Montiaceae), *Portulaca* (Portulacaceae), and in the succulent-leaved vine *Anredera baselloides* (Basellaceae)] (Holtum *et al.*, 2017a, 2017b, 2018; Winter and Holtum, 2017), we decided to test whether the ostensibly C_3 or C_4 isotope values reported for *Sesuvium* might mask the presence of low-level CAM. We were further encouraged to do so by a single observation of small nocturnal acidification in field-growing *S. maritimum* (Walter) Britton, Sterns & Poggenb. that exhibited a C_3 -type isotope value of -26‰ (Martin *et al.*, 1982).

A plant exhibits the CAM photosynthetic pathway when, in chloroplast-containing cells, CO_2 is incorporated at night into malic acid that is stored in vacuoles (Osmond, 1978). In the subsequent light-period, the CO_2 is retrieved from malic acid and is used for growth. The manifestation of CAM is therefore defined as an ability of a plant to fix CO_2 in the dark and to accumulate acid at night. Nocturnal acidification is typically determined by measuring the differences in titratable acidity in extracts of tissue harvested at the beginning and the end of the night.

In the majority of plants with CAM, it is expressed alongside C_3 photosynthesis or, in a few species, C_4 photosynthesis (Koch and Kennedy, 1980; Winter and Holtum, 2017). The contribution of CAM to net daily carbon gain varies considerably between species. Nocturnal acidification reported for plants with CAM ranges over two orders of magnitude from $3\text{--}4 \mu\text{mol H}^+ \text{g}^{-1}$ fresh mass (FM; close to the lower limit of detectability) to over $400 \mu\text{mol H}^+ \text{g}^{-1}$ FM. At one extreme of the continuum, CAM may be the principal carbon-contributing pathway in a plant (Winter and Holtum, 2014), whereas at the other extreme the contribution of CAM to carbon gain may be extremely small [e.g. *Platyserium veitchii* (Holtum and Winter, 1999); *Jatropha curcas* (Winter and Holtum, 2015); *Yucca gloriosa* (Heyduk *et al.*, 2016)]. In the latter group, the machinery for CAM is nonetheless present and the pathway is operational,

although nocturnal CO_2 fixation may be masked by dark respiratory CO_2 loss.

The expression of CAM may be constitutive, i.e. non-optional in that expression is part of the pre-set processes of development and growth, or it may be facultative, i.e. optional, in that it is not always present (Winter and Holtum, 2014; Winter *et al.*, 2015). Facultative CAM involves an induction or up-regulation of CAM in response to a stress (typically water stress) that is fully or largely lost when the stress is removed. Facultative and constitutive CAM are not necessarily mutually exclusive in a given photosynthetic organ. In a plant in which CAM is constitutively expressed, transient stress may transiently up-regulate CAM (Winter *et al.*, 2008). Because the contribution of dark CO_2 fixation to lifetime carbon gain can be small in some facultative CAM plants and in species that exhibit low levels of constitutive CAM, such plants may exhibit isotope values similar to C_3 or C_4 plants and the detection of CAM in these species is best measured by quantifying both leaf acidification and nocturnal CO_2 exchange during a wet-dry-wet sequence.

To further investigate the possibility of CAM in the Sesuvioideae we chose as a test species *S. portulacastrum*. Typically regarded as a C_3 species even when growing under conditions of stress (Lüttge *et al.*, 1989; Voznesenskaya *et al.*, 2010; Bohley *et al.*, 2015), a single value of overnight H^+ increase of field-growing *S. portulacastrum* has been reported (Ting, 1989). The species is a pan-tropical succulent-leaved coastal herb with an extensive distribution that spans coastlines of Africa, North and South America, tropical and temperate Asia, Australia, and the Pacific islands between 35°N and 42°S (Gonçalves, 1978; Lonard and Judd, 1997; Bohley *et al.*, 2017; Hartmann, 2017). In some regions, the salt-tolerating halophyte is cultivated for food and fodder, and used for dune stabilization and phytoremediation (Lokhande *et al.*, 2013a, 2013b).

Here, we provide results from a detailed study of gas exchange and nocturnal acidification in plants from a Panamanian population of *S. portulacastrum* either cultivated in pots or growing in outdoor experimental plots throughout tropical wet-season/dry-season transitions. Conclusive evidence is provided for the presence of low-level CAM in both the stems and leaves of *S. portulacastrum*.

Materials and methods

Plant material

Cuttings were obtained from five plants of *Sesuvium portulacastrum* (L.) L. collected at Sarigua National Park, Azuero Peninsula, Republic of Panama (8.013348N , 80.485658W) and further propagated at the Smithsonian Tropical Research Institute. A voucher specimen was deposited at the herbarium of the National University of Panama (PMA) (J. Aranda, A. Virgo, M. García, K. Winter 4324; 25 October 2018).

Outdoor experiments

Two experiments were performed. In the first, at the beginning of September, 2016 (middle of the wet season), 10 cuttings were planted in forest soil in a $1.5 \times 1.5 \times 0.3$ m raised garden bed surrounded by 3.5-cm thick wood panels at the Smithsonian Tropical Research Institute, Santa Cruz Experimental Research Facility, Gamboa, Republic of Panama

(9.120085N, 79.701894W). Plants were exposed to full natural solar radiation and received natural rainfall. Plants were grown for 186 d. In the second experiment, which began on 1 June, 2017 (early wet season), plants derived from 25 cuttings were grown for 333 d in a mixture of 50:25:25% (v/v/v) of forest soil: potting mix (Miracle-Gro Lawn Products, Marysville, OH): sea-sand (Noveys, Panama) in a raised garden bed as described above (Fig. 1). From July to December 2017, in addition to natural rainfall, the plants received at monthly intervals 10 l of 10% seawater prepared from 3.5 g l⁻¹ Ocean Salt (Instant Ocean, Blacksburg, VA (e.g. Reef *et al.*, 2016). Leaves and stems were sampled at dusk and dawn at weekly or 2-weekly intervals, unless specified otherwise. Samples were weighed for fresh mass determination and stored in liquid nitrogen to be processed further for measurements of titratable acidity, as described below.

Pot experiments

Throughout 2016 and 2017, several experiments were conducted to study titratable acidity changes and gas-exchange responses during wet-dry-wet cycles. Plants were grown in terracotta pots (ranging from 0.5–3.5 l) containing potting mix (Miracle-Gro). Plants were maintained at the Tupper Centre of the Smithsonian Tropical Research Institute, Panama City (8.962568N, 79.543911W) under either full solar radiation or underneath rain shelters at 70% natural light. Growth conditions are specified in the relevant figure legends. As in the outdoor experiments, for acidity measurements mature leaves and stems excised from plants at dusk and dawn were weighed for determination of fresh mass and frozen in liquid nitrogen. In some experiments, prior to freezing the leaf area was determined using a LI-3100 leaf area meter (Li-Cor, Lincoln, NE).

Titratable acidity and dry mass

Frozen tissue was either extracted sequentially in boiling 50% ethanol and in water (Winter and Holtum, 2017), or was freeze-dried for 72 h (Labonco, Freezone 4.5, Kansas City, MO) and reweighed for dry mass determination before extraction. All extracts were titrated with 5 mM KOH to pH 6.5.

Net CO₂ exchange

To determine daily CO₂ exchange patterns during wet-dry-wet cycles whole plants, attached branches, or individual attached leaves were studied. In one of the experiments, the shoot of a small plant was enclosed inside a Perspex cuvette (internal dimensions 11×11×10 cm) that rested on the 0.5-l terracotta pot in which the plant grew in potting mix (Miracle-Gro). The roots and the pot remained outside the cuvette. In another experiment, an individual leaf attached to a plant growing in a 1-l terracotta pot containing potting mix was enclosed in a 4.5-cm diameter PMK-10 leaf cuvette (porometer head; Walz, Effeltrich, FRG). After initial daily irrigation with water or a solution equivalent to 10% seawater, a drought



Fig. 1. *Sesuvium portulacastrum* growing outside in a 1.5×1.5×0.3-m raised garden bed at the Smithsonian Tropical Research Institute, Republic of Panama in January 2018.

treatment was imposed by withholding irrigation until net CO₂ uptake during the light period approached zero, after which plants were rewatered daily.

The gas-exchange cuvettes were located inside a controlled environment chamber (GC8, EGC, Chagrin Falls, OH) operating under 12/12 h light/dark, 28/22 °C cycles. Illumination was supplied by a LED grow light (GroPro300, Model LL4L-GP300, www.LL4L.com). Photon flux densities are specified in the relevant figure legends. The cuvette was supplied with air containing 400 ppm CO₂ at a flow rate of 1.26 l min⁻¹. Net CO₂ exchange was measured in a flow-through gas-exchange system consisting of Walz components and a LI-6252 CO₂ Analyzer (Li-Cor) (Holtum and Winter, 2003).

Results

Outdoor experiments in garden beds

Sesuvium portulacastrum experienced two distinct climatic phases when grown outside for 186 d in forest soil (Fig. 2). During the first 125 d (i.e. from September 2016 to early January 2017) it rained on many days, whereas for the final 61 d rainfall was essentially absent (Fig. 2B). As rainfall ceased, the average daily irradiance increased (Fig. 2A). By the end of March, all plants had died. During the rainy period, leaves contained low levels of titratable acidity, when expressed on a fresh mass basis, but the values were significantly greater at dawn than at dusk for 9 of 15 weekly measurements (Fig. 2C). Stem acidity measurements began in October. In all wet-season samples, stem acidity increased significantly overnight, with the dawn-to-dusk difference being greater in stems than in leaves (Fig. 2C, D).

Soon after initiating the experiment, we noted that leaves slightly wilted during the daytime and recovered overnight. This observation prompted us to determine fresh:dry mass ratios from December onwards, and to express leaf and stem acidities also on a dry mass basis (Fig. 2E, F).

Nocturnal acidification in leaves, on both a fresh mass (FM) and a dry mass (DM) basis, was greater during the dry season than in the rainy season (Fig. 2C, E). ΔH^+ was 1.3±0.9 $\mu\text{mol g}^{-1}$ FM (mean±SD, $n=15$ d) during the wet season and 7.7±3.2 $\mu\text{mol g}^{-1}$ FM ($n=8$ d) during the dry season ($P\leq 0.001$). Corresponding values on a dry mass basis were 38±10 $\mu\text{mol g}^{-1}$ DM ($n=4$ d; wet season) and 95±26 $\mu\text{mol g}^{-1}$ DM ($n=9$ d; dry season) ($P\leq 0.001$). In stems, acidification was also higher during the dry season, but only when values were expressed on a fresh mass basis (5.1±2.2 $\mu\text{mol g}^{-1}$ FM, $n=8$ d, wet season versus 9.1±1.8 $\mu\text{mol g}^{-1}$ FM, $n=9$ d, dry season; $P\leq 0.001$) and not when expressed on a dry mass basis (Fig. 2D, F). As the soil dried out during the progression between seasons, the FM to DM ratios of both leaves and stems decreased (Fig. 2G, H). Diurnal reductions in leaf FM:DM were large during the wet season and decreased during the dry season (Fig. 2G). Diurnal variations in FM:DM were largely absent in stems.

Since *S. portulacastrum* typically grows in coastal saline environments, a second outdoor experiment was conducted during the following 2017/2018 season in which plants were irrigated monthly with 10% seawater during the wet period. The input of extra moisture was small, equivalent to 4.4 mm precipitation per month. The salt was supplied to fulfil a hypothetical requirement for optimal growth, not to stress the plants *per se*.

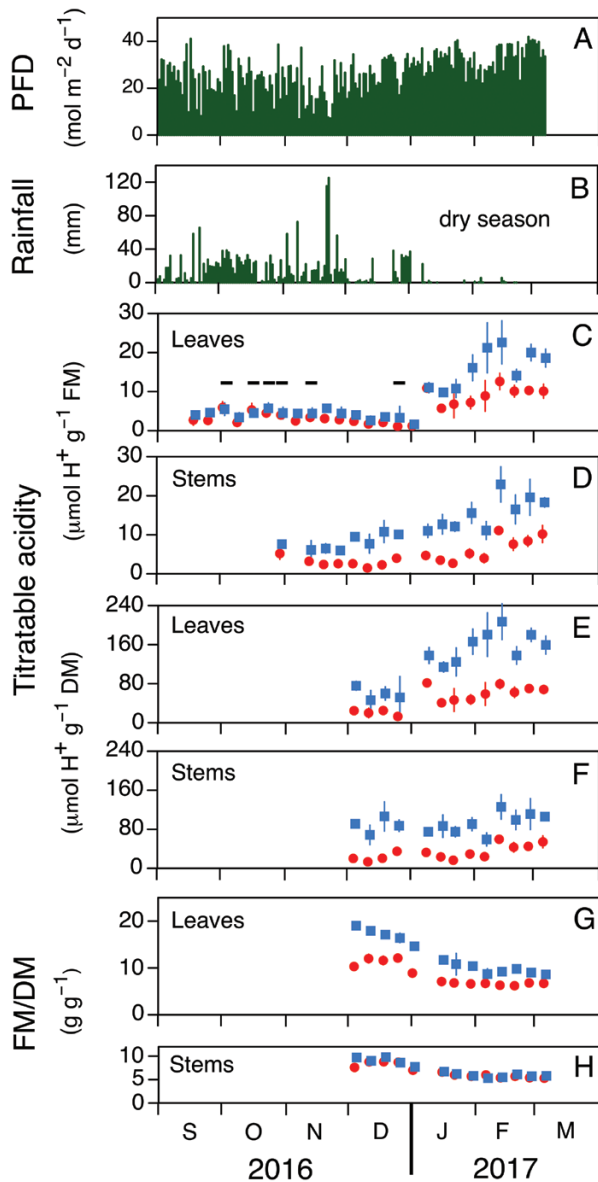


Fig. 2. Seasonal changes in (A) photon flux density (PFD), (B) rainfall, (C–F) variation of titratable acidity at dusk (circles) and dawn (squares), and (G, H) variation in fresh:dry mass ratio (FM/DM) at dusk and dawn in leaves and stems of *Sesuvium portulacastrum*. Plants were grown in non-saline soil during the latter half of the 2016 wet season (September–December) and as they transitioned into the 2017 dry season. Acidity levels are expressed per unit fresh mass for leaves (C) and stems (D) and per unit dry mass for leaves (E) and stems (F). Acidity and FM/DM values are means (\pm SD) ($n=5$; for leaves, each sample comprised eight leaves). In (C–F), horizontal bars indicate non-significant differences between dusk and dawn values (one-tailed t -test, $P \leq 0.05$).

The rainfall period lasted longer during the 2017/2018 season than during the 2016/2017 season of the first experiment (Fig. 3). Significant rainfall events extended into the middle of January 2018 (the wettest January since rainfall measurements started at the site 14 years previously). January 2018 rainfall was 80 mm, whereas January 2017 rainfall was 28 mm. In contrast to the previous experiment, plants survived the 2018 dry season.

As occurred in plants grown outdoors in non-saline soil, nocturnal acidification in *S. portulacastrum* was observed in both leaves

and stems in plants grown outdoors under rainfall supplemented with 10% seawater (Fig. 3). During the rainy period, acidification in stems was consistently greater than in leaves, about 2-fold on a DM basis and 3-fold on a FM basis. Nevertheless, the absolute levels of nocturnal acidification in leaves and stems, irrespective of what basis they were expressed, were similar to those observed in plants grown during the previous year.

Again, during the wet season, leaves exhibited low levels of acidity on a FM basis that were significantly higher at dawn than at dusk on 22 of 28 wet-season days. On a DM basis, dawn values were higher on all wet-season days. Nocturnal acidification in leaves significantly increased during the dry season (collection dates from 27 January to 22 April 2018), from $1.4 \pm 0.8 \mu\text{mol H}^+ \text{g}^{-1} \text{FM}$ ($n=28$ d) during the wet season to $3.8 \pm 3.0 \mu\text{mol H}^+ \text{g}^{-1} \text{FM}$ ($n=11$ d) during the dry season ($P \leq 0.05$), and from $41 \pm 17 \mu\text{mol H}^+ \text{g}^{-1} \text{DM}$ ($n=28$ d) during the wet season to $68 \pm 27 \mu\text{mol H}^+ \text{g}^{-1} \text{DM}$ ($n=11$ d) during the dry season ($P \leq 0.01$). No significant dry-season increase in nocturnal acidification was observed in stems, either on a FM or a DM basis.

Experiments with potted plants

Several pot experiments were conducted to study nocturnal acidification patterns in response to short-term drought stress. Leaves of well-irrigated plants supplied with either water or 10% seawater exhibited either no or a small nocturnal acidification. In the case of the experiment shown in Fig. 4, nocturnal acidification was below $2 \mu\text{mol H}^+ \text{g}^{-1} \text{FM}$ under high initial soil moisture conditions. Following the imposition of water stress, the acid accumulation at night was stimulated ~ 19 -fold. Upon rewatering, acidification was markedly reduced but was not eliminated, remaining at $\sim 4 \mu\text{mol H}^+ \text{g}^{-1} \text{FM}$.

In a separate experiment, stems of well-watered *S. portulacastrum* exhibited low levels of nocturnal acidification of less than $2 \mu\text{mol H}^+ \text{g}^{-1} \text{FM}$ (Fig. 5). Following the cessation of watering, nocturnal acidification increased ~ 7 -fold on a FM basis. When the plants were rewatered, the nocturnal accumulation was reduced markedly but was still present at a level that was ~ 3 -fold greater than the values at the beginning of the experiment. The wet-dry-wet acidification patterns were similar when H^+ was expressed on a DM or area basis for leaves or on a DM basis for stems. In all cases, a strong drought-induced up-regulation of nocturnal acid accumulation was observed, which was largely, albeit not fully, reversible.

Seven gas-exchange experiments were conducted with essentially identical results, two of which are depicted in Figs 6 and 7. For a well-watered plant of *S. portulacastrum*, net CO_2 uptake by its shoot was restricted to the 12-h light period (Fig. 6). CO_2 efflux was relatively constant throughout the night following an initial overshoot and period of equilibration to the night temperature, although a small curvature in the nocturnal CO_2 release pattern was detectable with lower rates in the middle than at the end of the dark period. Following cessation of watering on day 4, CO_2 uptake during the day continued to increase as the shoot continued to grow, utilizing water remaining in the pot. From day 14 onwards there was a progressive reduction in both CO_2 uptake during the day and CO_2 loss at night. Daytime CO_2 exchange developed a prominent

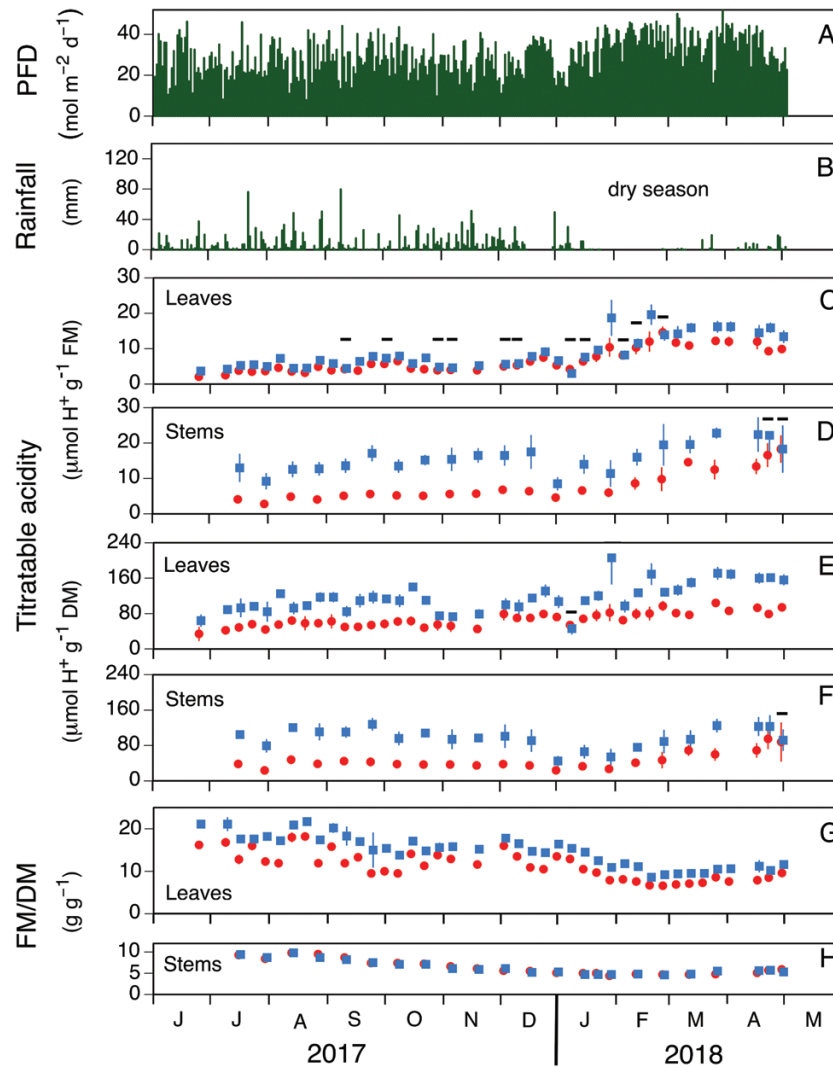


Fig. 3. Seasonal changes in (A) photon flux density (PFD), (B) rainfall, (C–F) variation of titratable acidity at dusk (circles) and dawn (squares), and (G, H) variation in fresh:dry mass ratio (FM/DM) at dusk and dawn in leaves and stems of *Sesuvium portulacastrum* grown in soil supplemented with sea-salt. Plants were grown for most of the 2017 wet season, and as they transitioned into the 2018 dry season. Acidity levels are expressed per unit fresh mass for leaves (C) and stems (D) and per unit dry mass for leaves (E) and stems (F). Acidity and FM/DM values are means (\pm SD) ($n=5$; for leaves, each sample comprised eight leaves). In (C–F), horizontal bars indicate non-significant differences between dusk and dawn values (one-tailed t -test, $P \leq 0.05$).

mid-day depression of uptake. Nocturnal CO_2 exchange approached the CO_2 compensation point but never transitioned to net CO_2 gain.

Following rewatering, a recovery of CO_2 uptake in the light was observed within 4 h and dark respiration increased during the subsequent night. Within 48 h after rewatering, the shoot exhibited a pattern of CO_2 exchange similar to that observed in the original well-watered conditions. The kinetics of nocturnal CO_2 efflux in the shoots of the rewatered plants were more curved than the efflux pattern in the original well-watered plants. The rate of CO_2 loss was also greater. The daytime CO_2 uptake rates following rewatering exceeded those at the onset of the experiment because the plants continued to grow inside the gas-exchange cuvette throughout the experiment, although at a reduced rate during the period of drought. The CO_2 exchange pattern during a watering–droughting–rewatering cycle of a leaf of a plant, in this particular case irrigated with 10% seawater (Fig. 7), was similar to that of the

shoot of a plant that was irrigated with water only (Fig. 6). The leaf of a well-watered plant exhibited net CO_2 uptake during the day and net CO_2 loss at night. At 3 d after watering ceased, daytime net CO_2 exchange began to decrease, exhibiting a progressively larger mid-day depression of uptake. CO_2 efflux at night also decreased, such that the CO_2 compensation point was reached but net nocturnal CO_2 uptake was not detected.

Within a few hours of rewatering on day 8, the rates of daytime CO_2 uptake began to recover. During the following night, nocturnal CO_2 loss gradually increased. By day 11, the patterns and extent of daytime CO_2 uptake and night-time CO_2 loss were similar to the patterns observed on day 1.

Discussion

The purpose of this study was to clarify whether CAM is present or absent in *S. portulacastrum*. To this end, plants grown

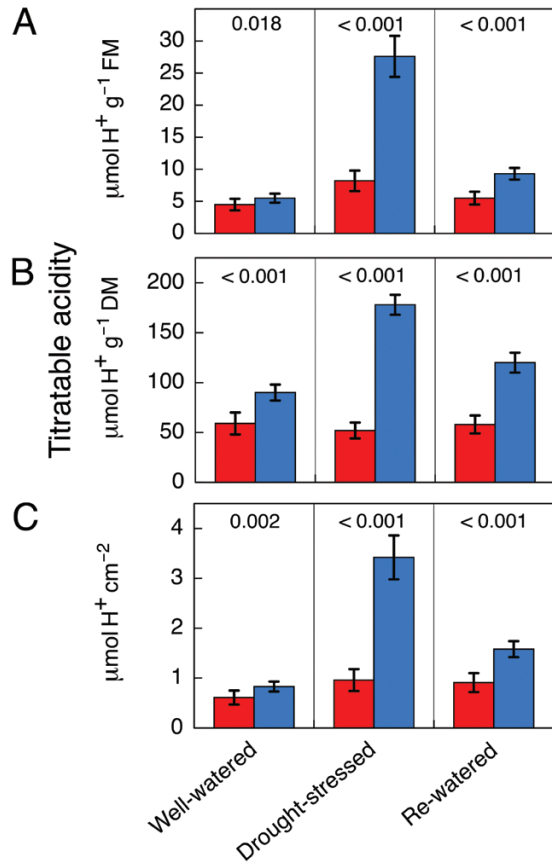


Fig. 4. Titratable acidity at dusk (red) and dawn (blue) in leaves of *Sesuvium portulacastrum* grown in 3.5-l pots in soil supplemented with sea-salt underneath a rain shelter at 70% of natural sunlight. Samples were taken from eight plants. The experiment was performed from the end of October to early December 2017. Data are shown for well-watered plants, droughted plants (20 d without irrigation), and plants that had been droughted and rewatered (20 d with irrigation). The data are expressed on the basis of (A) fresh mass, (B) dry mass, and (C) leaf area, and are means (\pm SD) ($n=8$; each sample comprised eight leaves). P -values from one-tailed t -tests are shown.

outdoors in the ground or potted plants grown under more controlled conditions were subjected to a range of conditions, including supplementation with NaCl and drought stress. Under most conditions, small nocturnal increases in acid content were measured in both leaves and stems, suggesting that CAM is present in *Sesuvium* and thus in the Sesuvioideae, the only sub-family of the Aizoaceae for which CAM had not yet been conclusively documented (Fig. 8). Given the ability to perform low-level CAM, *S. portulacastrum* joins a growing number of species that cannot be considered C_3 plants *sensu strictu*, although they obtain CO_2 principally via the C_3 -pathway.

Whilst CAM photosynthesis is present in *S. portulacastrum*, in comparison to C_3 photosynthesis, the contribution of nocturnal CO_2 assimilation to the carbon gain of the plant is extremely small. Whole-leaf carbon isotope values of plants in the field of generally around -24 to -26‰ (Winter et al., 1981; Lüttge et al., 1989; Medina et al., 2008; Bohley et al., 2015), indicate that, averaged over the life of the plant, the vast majority of carbon is assimilated by C_3 photosynthesis during the light (Winter and Holtum, 2002). In the gas-exchange time-courses for shoots and leaves shown in Figs 6 and 7 (as well as

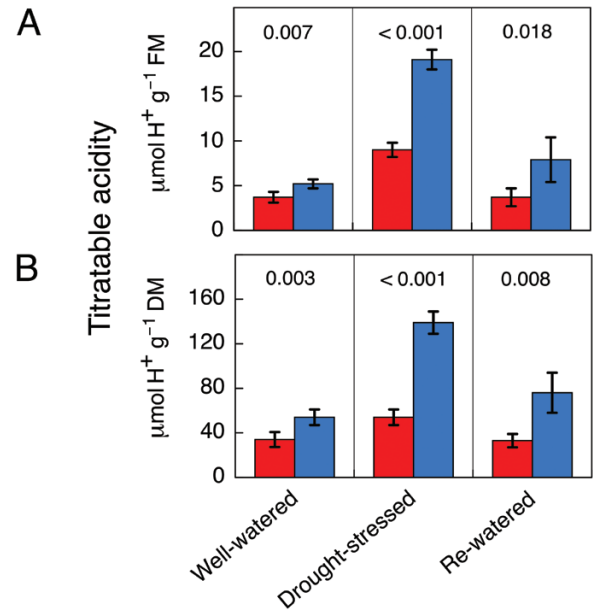


Fig. 5. Titratable acidity at dusk (red) and dawn (blue) in stems of *Sesuvium portulacastrum* grown in 2.5-l pots in non-saline soil under full sunlight. The experiment was performed during January/February 2017. Samples were randomly taken from eight plants. Data are shown for well-watered plants, droughted plants (6 d without irrigation), and plants that had been droughted and rewatered (7 d with irrigation). The data are expressed on the basis of (A) fresh mass, and (B) dry mass, and are means (\pm SD) ($n=4$ stems; at a given time-point each stem was harvested from a different plant). P -values from one-tailed t -tests are shown.

in the additional experiments for which data are not shown), the rates of dark CO_2 fixation were not high enough to compensate fully for respiratory CO_2 loss. CO_2 exchange at night approached but did not exceed the CO_2 compensation point.

Similar levels of nocturnal acidification accumulated in plants grown in outdoor raised garden plots and in plants grown in pots under more controlled conditions. Acidification was detected effectively throughout the life of the plants, even during the wet season, although acidification levels in leaves were greater during the dry season. In contrast to the evidence from the outdoor experiments, examination of the pot experiment acidification data (Figs 4, 5) would suggest that most, if not all, of the CAM expression in leaves was facultative as acidification in non-stressed leaves was extremely low. The evidence for mostly facultative CAM in the pot experiments with controlled irrigation and for longer-term low-level CAM in the outdoor experiments is not necessarily contradictory. Observation of leaves of *S. portulacastrum* on outwardly healthy, fast-growing plants growing in the outdoor plots subject to natural daylight during the wet season indicated that the relatively elastic leaves (low elastic modulus) often appeared flaccid in the afternoons. The resultant leaf water deficit was evidently sufficient to lead to low-level CAM that persisted throughout the experiment. A similar phenomenon has been reported to induce CAM in leaves of well-watered plants of the facultative CAM plant *M. crystallinum* (Winter and Holtum, 2007), especially in plants grown under non-saline conditions.

Observations in the two outdoor experiments of increased nocturnal acidification in leaves during the dry season are

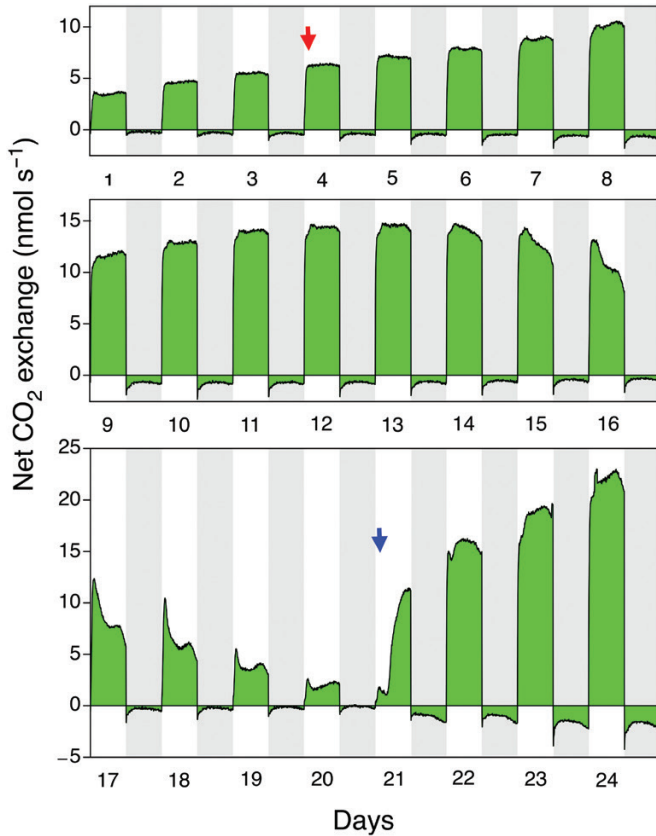


Fig. 6. Net CO₂ exchange over a 24-d period by the shoot of a *Sesuvium portulacastrum* plant growing in a 0.5-l pot containing non-saline soil. Watering was withheld on day 4 (red arrow) and recommenced on day 21 (blue arrow). The grey shaded areas represent the 12-h dark periods. Photon flux density was 700 $\mu\text{mol m}^{-2} \text{s}^{-1}$ at the level of the shoot. On the last day of the experiment, the total leaf area was 6 cm², the leaf dry mass was 0.129 g, and the stem dry mass was 0.024 g.

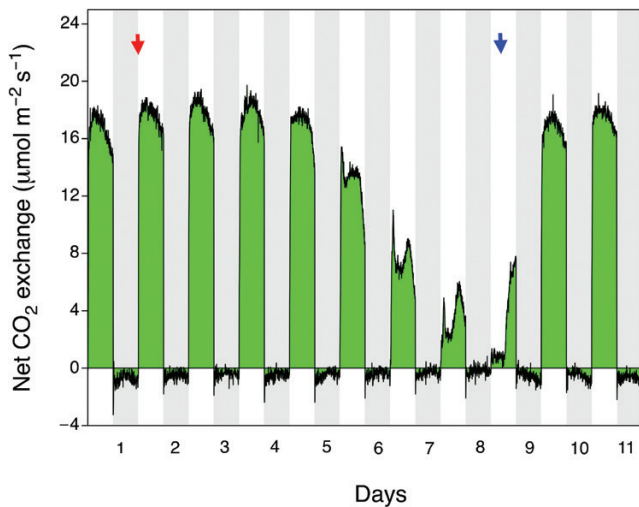


Fig. 7. Net CO₂ exchange over an 11-d period by an individual leaf of a *Sesuvium portulacastrum* plant growing in soil supplemented with sea-salt. Watering was withheld on day 2 (red arrow) and recommenced on day 9 (blue arrow). The grey shaded areas represent the 12-h dark periods. Photon flux density was 480 $\mu\text{mol m}^{-2} \text{s}^{-1}$. Gas exchange was measured on a leaf area of 2.98 cm².

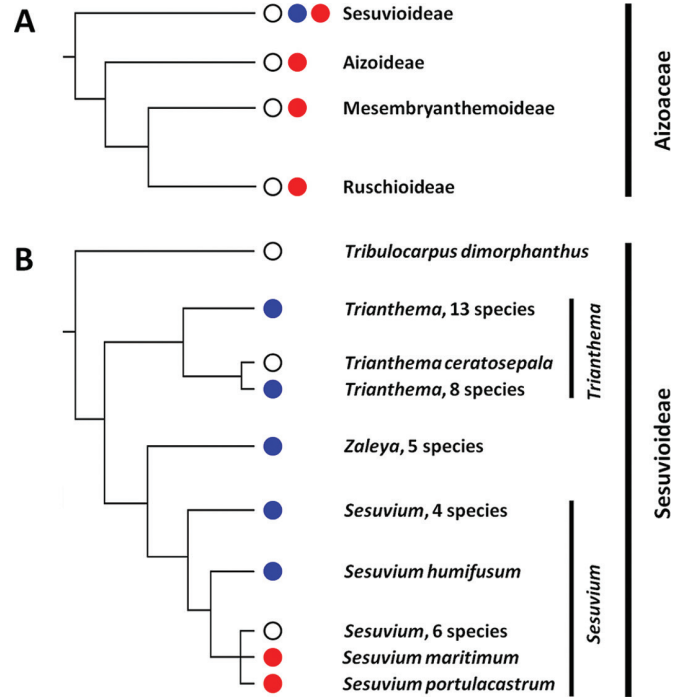


Fig. 8. Distribution of CAM (red) and C₄ (blue) within (A) the four sub-families of the Aizoaceae, and (B) genera of the sub-family Sesuvioideae (adapted from Smith and Winter, 1996; Klak *et al.*, 2003; Muhaidat *et al.*, 2007; Bohley *et al.*, 2015, 2017, and the current study). Phylogenetic relationships in (A) are based upon analysis of three chloroplastic and nuclear loci (Klak *et al.*, 2003), and relationships in (B) are based upon analysis of five chloroplastic and nuclear loci (Bohley *et al.*, 2015). The open circles indicate plants with C₃-type isotope values. Without measurements of acidification and gas exchange during a watering-droughting-rewatering regime, it cannot yet be determined whether such individuals exhibit C₃ *sensu strictu*, low-level CAM, or are C₃-C₄ intermediates. In (B), the presence of CAM in *S. maritimum* is based upon nocturnal acidification reported during a single night for a plant growing in the field (Martin *et al.*, 1982).

consistent with a strong facultative CAM component in leaves, while the absence of a consistent dry-season increase in nocturnal stem acidification suggests that low-level CAM in stems has a substantial constitutive component. The ability of *S. portulacastrum* to maintain nocturnal acidification levels well into the dry season, even as FM/DM ratios fell below 5 for stems and below 10 in succulent leaves, provides an indication of the relative drought-tolerance of the species. Indeed, plants growing on a saline plain in northern Venezuela have been reported to operate at xylem tensions of as low as -5.6 MPa (Lüttge *et al.*, 1989).

CAM is now documented in all four sub-families of the Aizoaceae (Fig. 8A). Pronounced CAM, undoubtedly overwhelmingly constitutive, is present in the sub-families Aizoideae, Mesembryanthemoideae, and Ruschioideae (Smith and Winter, 1996), and facultative CAM is present in the Mesembryanthemoideae [e.g. *Mesembryanthemum crystallinum* (Winter and Holtum, 2007) and *Aptenia cordifolia* (Treichel, 1975)] and now in the Sesuvioideae (*S. portulacastrum*). All of the sub-types of CAM, constitutive, facultative, pronounced and weak, are also known in the only other clade of the Caryophyllales with CAM plants, the sub-order Portulacineae.

In the portullugo (Portulacineae plus Molluginaceae; Ogburn and Edwards, 2010; Edwards and Ogburn, 2012), pronounced CAM predominates in most genera within the Cactaceae and Didiereaceae (*Alluaudia*, *Alluaudiopsis*, *Decarya*, *Didierea*), whereas weak and facultative CAM are more commonly expressed in the Anacampserotaceae (Guralnick and Jackson, 2001; Winter and Holtum, 2017), *Portulacaria* within the Didiereaceae (Ting and Hanscom 1977; Guralnick and Ting, 1986), Montiaceae (Winter and Holtum, 2011; Martin *et al.*, 1988), Talinaceae (Herrera *et al.*, 1991), Basellaceae (Holtum *et al.*, 2018), and in the C₄ and C₃-C₄ Portulacaceae (Koch and Kennedy, 1980; Winter and Holtum, 2017; K. Winter *et al.*, unpublished results). In the thin-leaved Molluginaceae, C₄ and C₃ are present but CAM has not been reported (Christin *et al.*, 2011).

CAM lineages and C₄ lineages tend to cluster in certain regions of the angiosperm phylogenetic tree (Sage *et al.*, 2011a; Edwards and Ogburn, 2012; Christin *et al.*, 2015), with particularly closely related CAM and C₄ lineages identified within the Aizoaceae, Euphorbiaceae, and Portulacineae (Klak *et al.*, 2003, 2017; Sage *et al.*, 2011b; Horn *et al.*, 2012, 2014; Ocampo *et al.*, 2013; Christin *et al.*, 2014). Following the discovery of CAM and C₄ photosynthesis in *Euphorbia* and *Portulaca*, *Sesuvium* becomes only the third genus known to contain both types of carbon-concentrating photosynthesis.

In the Sesuvioideae, the most basal of the sub-families in the Aizoaceae, CAM is reported here in *Sesuvium*, the most derived clade (Fig. 8B). C₄ is present in *Zaleya* and *Trianthema* (Muhaidat *et al.*, 2007; Koteyeva *et al.*, 2015), whereas neither C₄ nor CAM have been reported in the most basal clade, *Tribulocarpus* (Bohley *et al.*, 2015, 2017). Succulence is present in *Sesuvium*, *Trianthema*, and to a lesser extent in *Zaleya* and *Tribulocarpus*.

CAM and C₄ are present in *Sesuvium* but they are yet to be demonstrated in the same species. The genus circumscribes a C₃ clade and a C₄ clade (Bohley *et al.* 2015, 2017). Both *S. portulacastrum* and the closely related *S. maritimum* studied by Martin *et al.* (1982) are in the C₃ clade. It would be informative to explore whether CAM is present within members of C₄ *Sesuvium* clade. In contrast to what is currently known about *Sesuvium*, in *Portulaca* CAM is expressed within at least three clades of C₄ species and thus, in those species with CAM, it is co-expressed with C₄ in the same plant (Koch and Kennedy, 1980; Holtum *et al.*, 2017b; Winter and Holtum, 2017). In *P. cryptopetala*, facultative CAM is present in a C₃-C₄ intermediate species that exhibits a C₂-cycle (Muhaidat *et al.*, 2007; Vozsensky *et al.*, 2010, 2017; K. Winter *et al.*, unpublished results). Within the genus *Euphorbia*, which contains C₃, C₄, and CAM species (Webster *et al.*, 1975; Sage *et al.*, 2011b; Yang and Berry, 2011), there are no reports as yet of CAM within the C₄ clade (subg. *Chamaesyce*), although it appears to have repeatedly evolved alongside succulence from C₃ lineages (Horn *et al.*, 2014). Together, the *Euphorbia*, *Portulaca*, and *Sesuvium* lineages with their differing evolutionary histories of CAM and C₄ photosynthesis provide promise for exploring the biochemical, anatomical, and ecological trajectories that favour the selection of one carbon-concentrating mechanism over the other, or permit the co-existence of both.

The ecological significance of the low-level contribution of carbon contributed by CAM to *S. portulacastrum* is as yet unassessed, but as is evident from the outdoor experiments described here, low-level and facultative CAM can be expressed over extensive periods. As a species that is a coloniser of the fringes of saline flats, *S. portulacastrum* probably spends much of its life growing slowly under conditions of salinity, drought stress, high light, and high temperatures, as evidenced by the vividly red-coloured fleshy stems and wilted leaves frequently exhibited by plants in the field. Under such conditions one might expect CO₂ assimilation in the light to be constrained by water limitation. The reductions in water loss and respiratory carbon loss enabled by the low-level CAM, an ability to accumulate compatible solutes, and to accumulate the anion oxalate (Lüttge *et al.* 1989), coupled with other physiological mechanisms (Lokhande *et al.*, 2013b), presumably assist plants to cope with the stresses imposed by the environment it inhabits.

In terms of carbon-isotope composition and gas-exchange pattern, *S. portulacastrum* is C₃-like. Yet our multiple measurements of plants in pots and long-term measurements in the field demonstrate that *S. portulacastrum* consistently exhibits CAM in both leaves and stems. Since the levels of nocturnal acidification are low and easy to overlook, caution is required when assessing such plants for photosynthetic pathway. This study also highlights the care required when selecting C₃ control species in evolutionary studies of lineages in which CAM or C₄ has evolved. CAM and C₄ origins may not always represent completely independent evolutionary phenomena, and may partially share evolutionary trajectories in that one photosynthetic type can be co-opted to evolve the other (Christin *et al.*, 2014).

Acknowledgements

This research was supported by the Smithsonian Tropical Research Institute (STRI) and the Australian Research Council Discovery Project (DP160100098). We acknowledge the assistance of Jorge Aranda who maintained the experimental plots at Gamboa.

References

- Alonso-Cantabrana H, von Caemmerer S. 2016. Carbon isotope discrimination as a diagnostic tool for C₄ photosynthesis in C₃-C₄ intermediate species. *Journal of Experimental Botany* **67**, 3109–3121.
- Arakaki M, Christin P-A, Nyffeler R, Lendel A, Eggli U, Ogburn RM, Spriggs E, Moore MJ, Edwards EJ. 2011. Contemporaneous and recent radiations of the world's major succulent plant lineages. *Proceedings of the National Academy of Sciences, USA* **108**, 8379–8384.
- Bittrich V. 1990. Systematic studies in Aizoaceae. *Mitteilungen aus dem Institut für Allgemeine Botanik in Hamburg* **23b**, 491–507.
- Bohley K, Joos O, Hartmann H, Sage R, Liede-Schumann S, Kadereit G. 2015. Phylogeny of Sesuvioideae (Aizoaceae) – Biogeography, leaf anatomy and the evolution of C₄ photosynthesis. *Perspectives in Plant Ecology, Evolution and Systematics* **17**, 116–130.
- Bohley K, Winter PJD, Kadereit G. 2017. A revision of *Sesuvium* (Aizoaceae, Sesuvioideae). *Systematic Botany* **42**, 124–147.
- Carolin RC, Jacobs SWL, Vesk M. 1978. Kranz cells and mesophyll in the Chenopodiales. *Australian Journal of Botany* **26**, 683–698.
- Christin PA, Arakaki M, Osborne CP, *et al.* 2014. Shared origins of a key enzyme during the evolution of C₄ and CAM metabolism. *Journal of Experimental Botany* **65**, 3609–3621.

- Christin PA, Arakaki M, Osborne CP, Edwards EJ.** 2015. Genetic enablers underlying the clustered evolutionary origins of C_4 photosynthesis in angiosperms. *Molecular Biology and Evolution* **32**, 846–858.
- Christin PA, Sage TL, Edwards EJ, Ogburn RM, Khoshravesh R, Sage RF.** 2011. Complex evolutionary transitions and the significance of C_3 - C_4 intermediate forms of photosynthesis in Molluginaceae. *Evolution* **65**, 643–660.
- Edwards EJ, Ogburn M.** 2012. Angiosperm responses to a low- CO_2 world: CAM and C_4 photosynthesis as parallel evolutionary trajectories. *International Journal of Plant Sciences* **173**, 724–733.
- Gilbert MG, Hartmann HEK, Edwards S.** 2000. Aizoaceae. In: Edwards S, Tadesse S, Demissew S, Hedberg I, eds. *Flora of Ethiopia & Eritrea*. Vol. 2. Addis Ababa and Uppsala: National Herbarium/Uppsala University, 240–248.
- Gonçalves ML.** 1978. Aizoaceae. In: Launert E, ed. *Flora Zambesiaca*. Vol. 4. London: Flora Zambesiaca Managing Committee, 508–521.
- Guralnick LJ, Jackson MD.** 2001. The occurrence and phylogenetics of crassulacean acid metabolism in the Portulacaceae. *International Journal of Plant Sciences* **162**, 257–262.
- Guralnick LJ, Ting IP.** 1986. Seasonal response to drought and rewatering in *Portulacaria afra* (L.) Jacq. *Oecologia* **70**, 85–91.
- Hartmann HEK.** 2017. *Illustrated handbook of succulent plants: Aizoaceae*. 2nd edn. Heidelberg: Springer.
- Hernández-Ledesma P, Berendsohn WG, Borsch T, et al.** 2015. A taxonomic backbone for the global synthesis of species diversity in the angiosperm order Caryophyllales. *Willdenowia* **45**, 281–383.
- Herrera A, Delgado J, Paraguatey I.** 1991. Occurrence of inducible crassulacean acid metabolism in leaves of *Talinum triangulare* (Portulacaceae). *Journal of Experimental Botany* **42**, 493–499.
- Heyduk K, Burrell N, Lalani F, Leebens-Mack J.** 2016. Gas exchange and leaf anatomy of a C_3 -CAM hybrid, *Yucca gloriosa* (Asparagaceae). *Journal of Experimental Botany* **67**, 1369–1379.
- Holtum JAM, Hancock LP, Edwards EJ, Winter K.** 2017a. Facultative CAM photosynthesis (crassulacean acid metabolism) in four species of *Calandrinia*, ephemeral succulents of arid Australia. *Photosynthesis Research* **134**, 17–25.
- Holtum JAM, Hancock LP, Edwards EJ, Winter K.** 2017b. Optional use of CAM photosynthesis in two C_4 species, *Portulaca cyclophylla* and *Portulaca digyna*. *Journal of Plant Physiology* **214**, 91–96.
- Holtum JAM, Hancock LP, Edwards EJ, Winter K.** 2018. Crassulacean acid metabolism (CAM) in the Basellaceae (Caryophyllales). *Plant Biology* **20**, 409–414.
- Holtum JAM, Winter K.** 1999. Degrees of crassulacean acid metabolism in tropical epiphytic ferns. *Australian Journal of Plant Physiology* **26**, 749–757.
- Holtum JA, Winter K.** 2003. Photosynthetic CO_2 uptake in seedlings of two tropical tree species exposed to oscillating elevated concentrations of CO_2 . *Planta* **218**, 152–158.
- Horn JW, van Ee BW, Morawetz JJ, Riina R, Steinmann VW, Berry PE, Wurdack KJ.** 2012. Phylogenetics and the evolution of major structural characters in the giant genus *Euphorbia* L. (Euphorbiaceae). *Molecular Phylogenetics and Evolution* **63**, 305–326.
- Horn JW, Xi Z, Riina R, Peirson JA, Yang Y, Dorsey BL, Berry PE, Davis CC, Wurdack KJ.** 2014. Evolutionary bursts in *Euphorbia* (Euphorbiaceae) are linked with photosynthetic pathway. *Evolution* **68**, 3485–3504.
- Klak C, Hanacek P, Bruyns PV.** 2017. Disentangling the Aizooidae: new generic concepts and a new subfamily in Aizoaceae. *Taxon* **66**, 1147–1170.
- Klak C, Khunou A, Reeves G, Hedderson T.** 2003. A phylogenetic hypothesis for the Aizoaceae (Caryophyllales) based on four plastid DNA regions. *American Journal of Botany* **90**, 1433–1445.
- Koch K, Kennedy RA.** 1980. Characteristics of crassulacean acid metabolism in the succulent C_4 dicot, *Portulaca oleracea* L. *Plant Physiology* **65**, 193–197.
- Koteyeva NK, Voznesenskaya EV, Edwards GE.** 2015. An assessment of the capacity for phosphoenolpyruvate carboxykinase to contribute to C_4 photosynthesis. *Plant Science* **235**, 70–80.
- Lokhande VH, Gor BK, Desai NS, Nikam TD, Suprasanna P.** 2013a. Biochemical and physiological adaptations of the halophyte *Sesuvium portulacastrum* (L.) L. (Aizoaceae) to salinity. *Archives of Agronomy and Soil Science* **59**, 1373–1391.
- Lokhande VH, Gor BK, Desai NS, Nikam TD, Suprasanna P.** 2013b. *Sesuvium portulacastrum*, a plant for drought, salt stress, sand fixation, food and phytoremediation. A review. *Agronomy for Sustainable Development* **33**, 329–348.
- Lonard RI, Judd FW.** 1997. The biological flora of coastal dunes and wetlands. *Sesuvium portulacastrum* (L.) L. *Journal of Coastal Research* **13**, 96–104.
- Lüttge U, Popp M, Medina E, Cram WJ, Diaz M, Griffiths H, Lee HSJ, Schäfer C, Smith JAC, Stimmel KH.** 1989. Ecophysiology of xerophytic and halophytic vegetation of a coastal alluvial plain in northern Venezuela. V. The *Batis maritima*-*Sesuvium portulacastrum* vegetation unit. *New Phytologist* **111**, 283–291.
- Martin CE, Higley M, Wang WZ.** 1988. Ecophysiological significance of CO_2 -recycling via Crassulacean acid metabolism in *Talinum calycinum* Engelm. (Portulacaceae). *Plant Physiology* **86**, 562–568.
- Martin CE, Lubbers AE, Teeri JA.** 1982. Variability in crassulacean acid metabolism: a survey of North Carolina succulent species. *Botanical Gazette* **143**, 491–497.
- Medina E, Francisco AM, Wingfield R, Casañas OL.** 2008. Halofitismo en plantas de la costa caribe de Venezuela: halófitas y halotolerantes. *Acta Botanica Venezuelica* **31**, 49–80.
- Muhaidat R, Sage RF, Dengler NG.** 2007. Diversity of Kranz anatomy and biochemistry in C_4 eudicots. *American Journal of Botany* **94**, 362–381.
- Ocampo G, Koteyeva NK, Voznesenskaya EV, Edwards GE, Sage TL, Sage RF, Columbus JT.** 2013. Evolution of leaf anatomy and photosynthetic pathways in Portulacaceae. *American Journal of Botany* **100**, 2388–2402.
- Ogburn RM, Edwards EJ.** 2010. The ecological water-use strategies of succulent plants. *Advances in Botanical Research* **55**, 179–225.
- Osmond CB.** 1978. Crassulacean acid metabolism: a curiosity in context. *Annual Review of Plant Physiology* **29**, 379–414.
- Reef R, Slot M, Motro U, Motro M, Motro Y, Adame MF, Garcia M, Aranda J, Lovelock CE, Winter K.** 2016. The effects of CO_2 and nutrient fertilisation on the growth and temperature response of the mangrove *Avicennia germinans*. *Photosynthesis Research* **129**, 159–170.
- Sage RF, Christin P-A, Edwards EJ.** 2011a. The C_4 plant lineages of planet Earth. *Journal of Experimental Botany* **62**, 3155–3169.
- Sage TL, Sage RF, Vogan PJ, Rahman B, Johnson DC, Oakley JC, Heckel MA.** 2011b. The occurrence of C_2 photosynthesis in *Euphorbia* subgenus *Chamaesyce* (Euphorbiaceae). *Journal of Experimental Botany* **62**, 3183–3195.
- Smith JAC, Winter K.** 1996. Taxonomic distribution of crassulacean acid metabolism. In: Winter K, Smith JAC, eds. *Crassulacean acid metabolism*. Berlin, Heidelberg: Springer, 427–436.
- Thulin M, Thiede J, Liede-Schumann S.** 2012. Phylogeny and taxonomy of *Tribulocarpus* (Aizoaceae): a paraphyletic species and an adaptive shift from zoochorous trample burrs to anemochorous nuts. *Taxon* **61**, 55–66.
- Ting IP.** 1989. Photosynthesis of arid and subtropical succulent plants. *Aliso* **12**, 387–406.
- Ting IP, Hanscom Z.** 1977. Induction of acid metabolism in *Portulacaria afra*. *Plant Physiology* **59**, 511–514.
- Treichel S.** 1975. Crassulaceansäurestoffwechsel bei einem salztoleranten Vertreter der Aizoaceae: *Aptenia cordifolia*. *Plant Science Letters* **4**, 141–144.
- von Willert DJ, Eller BM, Werger MJA, Brinckmann E, Ihlenfeldt HD.** 1992. Life strategies of succulents in deserts. Cambridge: Cambridge University Press.
- von Willert DJ, Thomas DA, Lobin W, Curdts E.** 1977. Ecophysiological investigations in the family of the Mesembryanthemaceae. *Oecologia* **29**, 67–76.
- Voznesenskaya EV, Koteyeva NK, Edwards GE, Ocampo G.** 2010. Revealing diversity in structural and biochemical forms of C_4 photosynthesis and a C_3 - C_4 intermediate in genus *Portulaca* L. (Portulacaceae). *Journal of Experimental Botany* **61**, 3647–3662.
- Voznesenskaya EV, Koteyeva NK, Edwards GE, Ocampo G.** 2017. Unique photosynthetic phenotypes in *Portulaca* (Portulacaceae): C_3 - C_4

intermediates and NAD-ME C₄ species with Pilosoid-type Kranz anatomy. *Journal of Experimental Botany* **68**, 225–239.

Webster GL, Brown WV, Smith BN. 1975. Systematics of photosynthetic carbon fixation pathways in Euphorbia. *Taxon* **24**, 27–33.

Winter K, Garcia M, Holtum JA. 2008. On the nature of facultative and constitutive CAM: environmental and developmental control of CAM expression during early growth of *Clusia*, *Kalanchoë*, and *Opuntia*. *Journal of Experimental Botany* **59**, 1829–1840.

Winter K, Holtum JA. 2002. How closely do the $\delta^{13}\text{C}$ values of crassulacean acid metabolism plants reflect the proportion of CO₂ fixed during day and night? *Plant Physiology* **129**, 1843–1851.

Winter K, Holtum JA. 2007. Environment or development? Lifetime net CO₂ exchange and control of the expression of crassulacean acid metabolism in *Mesembryanthemum crystallinum*. *Plant Physiology* **143**, 98–107.

Winter K, Holtum JAM. 2011. Induction and reversal of crassulacean acid metabolism in *Calandrinia polyandra*: effects of soil moisture and nutrients. *Functional Plant Biology* **38**, 576–582.

Winter K, Holtum JAM. 2014. Facultative CAM plants: powerful tools for unravelling the functional elements of CAM photosynthesis. *Journal of Experimental Botany* **65**, 3425–3441.

Winter K, Holtum JAM. 2015. Cryptic crassulacean acid metabolism (CAM) in *Jatropha curcas* L. *Functional Plant Biology* **42**, 711–717.

Winter K, Holtum JAM. 2017. CO₂-exchange patterns demonstrate facultative CAM photosynthesis (crassulacean acid metabolism) in four small C₃ and C₄ leaf-succulents. *Australian Journal of Botany* **65**, 103–108.

Winter K, Holtum JA, Smith JA. 2015. Crassulacean acid metabolism: a continuous or discrete trait? *New Phytologist* **208**, 73–78.

Winter K, Osmond CB, Pate JS. 1981. Coping with salinity. In: Pate JS, McComb AJ, eds. *The biology of Australian plants*. Perth: University of Western Australia Press, 88–113.

Yang Y, Berry PE. 2011. Phylogenetics of the Chamaesyce clade (*Euphorbia*, Euphorbiaceae): reticulate evolution and long-distance dispersal in a prominent C₄ lineage. *American Journal of Botany* **98**, 1486–1503.