

1 **Population genetic structure of the bank vole *Myodes glareolus***
2 **within its glacial refugium in peninsular Italy**

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4 **Running title:** Bank vole genetic structure in Italy

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23 structure.

24 **Abstract**

25 It is now well established that Southern European peninsulas have been major glacial refugia
26 for temperate species during Pleistocene climatic oscillations. However, substantial
27 environmental changes occurred also within these peninsulas throughout the Pleistocene, rising
28 questions about the role and interplay of various of micro-evolutionary processes in shaping
29 patterns of intraspecific diversity within these areas. Here, we investigate the patterns of genetic
30 variation in the bank vole *Myodes glareolus* within the Italian peninsula. By using a panel of
31 13 microsatellite loci, we found more intra-specific variation than expected based on previous
32 assessments. Indeed, both Bayesian and ordination-based clustering analyses of variation
33 recovered five main geographic/genetic clusters along the peninsula, with three clusters
34 geographically restricted to the southern portion of the peninsula. This clustering is supported
35 by previous evidences of some morphological distinctiveness among these populations. This
36 pattern can be explained by a refugia-within-refugia scenario, with the occurrence of multiple
37 sub-refugia for the bank vole within the Italian, likely promoted by the major paleo-
38 environmental changes which affected forested habitats within this area during the Pleistocene.
39 Moreover, our results support a scenario whereby the high levels of intraspecific diversity
40 observed within major Pleistocene refugia are better explained by dynamic micro-evolutionary
41 processes occurred within these areas, rather than by long-term demographic stability of
42 refugial population. Finally, the narrow and isolated distribution of some of the identified
43 lineages, suggest the need for future assessments of their conservation and taxonomic status.

44 **Introduction**

45 Southern European peninsulas have provided an excellent research ground to investigate
46 how past climate changes and topographic features influenced species' evolutionary histories
47 (Hewitt, 2011). Plenty of studies in the last thirty years highlighted the role of these peninsulas
48 as climatic refugia for temperate species during Pleistocene glacials (Bennet & Provan, 2008;
49 Comes & Kadereit, 1998; Feliner, 2011; Hewitt, 1996; 2004; Schmitt, 2007; Weiss & Ferrand,
50 2007; Stewart, Lister, Barnes & Dalén, 2010). Due to the strong topographic complexity of
51 these peninsulas, species underwent extreme population fragmentation within refugia (Gomez
52 & Lunt, 2007; Hofreiter & Stewart, 2009). As a consequences, these areas have been found to
53 be particularly rich of intraspecific genetic lineages – sometimes highly divergent from the
54 closest relatives (e.g. Canestrelli, Cimmaruta, Costantini & Nascetti, 2006), often with narrow
55 distribution (e.g. Bisconti et al., 2018) – detecting which is crucial to understanding species
56 genetic structure and to defining evolutionary and management units for conservation planning
57 (Avise, 2008; Frankham, 2010; Groves et al., 2017; Palsbøll, Berube & Allendorf, 2007).
58 However, despite their disproportionate importance as both cradles and reservoir for Western
59 Palearctic biodiversity, current knowledge of historical biogeographic patterns and processes
60 within these areas is still far from satisfactory. For many taxa, the genetic structure is still little
61 known, and the available knowledge if often flawed by limited sampling or limited number of
62 genetic markers.

63 The bank vole *Myodes glareolus* (Schreber, 1780) is a small woodland-dwelling rodent,
64 widespread throughout temperate and boreal forests of most of Europe (Amori, Contoli &
65 Nappi, 2008a), which has been a key species in the study of the European fauna response to the
66 Pleistocene climate changes. It has been one of the most convincing examples of a woodland
67 species surviving glaciations within a cryptic northern refugium in Europe, i.e. a refugium
68 located further north of the traditionally recognized refugia in the Southern European
69 peninsulas (Bhagwat & Willis, 2008; Bilton, Mirol, Mascheretti, Fredga, Zima & Searle, 1998;
70 Deffontaine et al., 2005; Filipi, Marková, Searle & Kotlík, 2015; Kotlík, Deffontaine,
71 Mascheretti, Zima, Michaux, & Searle, 2006). However, bank vole populations survived
72 Pleistocene glaciations also in southern refugia. There were identified distinct evolutionary
73 lineages in either Balkan, Iberian and Italian peninsulas (Colangelo, Aloise, Franchini, Annesi
74 & Amori, 2012; Deffontaine et al., 2005; Filipi et al., 2015). Within the Italian peninsula, four
75 distinct evolutionary lineages have been characterized by mean of mitochondrial DNA
76 variation: one widespread across Alps and northern Italy, one distributed mainly throughout

77 northern and central Apennines, one restricted to the Gargano promontory (Apulia), and one
78 found only in Calabria (Colangelo et al., 2012). This differentiation is supported also by some
79 slight morphological distinctiveness (Amori et al., 2008a). Interestingly, the Calabrian clade
80 showed strong and ancient (Early Pleistocene) genetic divergence from all other *M. glareolus*
81 lineages, resulting as the basal clade of the entire bank vole phylogeny, whereas the Gargano
82 clade does not cluster with any of the other lineages (Colangelo et al., 2012; Filipi et al., 2015).
83 Nevertheless, all phylogenetic and phylogeographic inferences were based only on
84 mitochondrial data, which have several limitations in inferring population genetic structure and
85 patterns of gene flow among populations (Ballard & Whitlock, 2004). Indeed, despite Calabrian
86 and Apulian populations resulted genetically differentiated from the other peninsular
87 populations, and virtually geographically isolated by the rarefaction/fragmentation of species
88 habitat, the lack of a multi-marker analysis of species genetic structure does not allow
89 inferences on genetic isolation.

90 In this study, we further investigate the genetic structure of the bank vole in the Italian
91 peninsula. We employ a set of thirteen microsatellite loci in order to complement previously
92 published mitochondrial data (Colangelo et al., 2012), with the aim of better understanding the
93 geographic structure of genetic variation and to shed more light on the bank vole evolutionary
94 history. Moreover, considering the narrow ranges of the Calabrian and Apulian lineages and
95 the ongoing reduction of the forest habitat (Scarascia-Mugnozza, Oswald, Piussi & Radoglou,
96 2000), a better understanding of the pattern of genetic isolation is mandatory to evaluate the
97 need for conservation actions concerning southern bank vole populations.

98

99 **Materials and methods**

100 We collected 76 *Myodes glareolus* individuals from 15 localities spanning the Italian
101 peninsula; collecting sites and sample sizes are given in Table 1 and Figure 1; voucher numbers
102 are listed in Appendix I. Tissue samples were obtained from an auricle biopsy on live-trapped
103 animals during field sessions, or from museum specimens (Museum of Comparative Anatomy
104 G.B. Grassi of the University of Rome ‘La Sapienza’).

105 DNA extractions were performed by using the standard cetyltrimethylammonium bromide
106 protocol (Doyle and Doyle, 1987). We analysed genetic variation at thirteen microsatellite loci:
107 *Cg14E1*, *Cg15F7*, *Cg17A7*, *Cg3A8*, *Cg3F12*, *Cg4F9*, *Cg12A7*, *Cg13C12*, *Cg17E9*, *Cg10A11*,
108 *Cg10H1*, *Cg13G2*, *Cg6A1* (Guivier et al., 2011; Rikalainen, Grapputo, Knott, Koskela &
109 Mappes, 2008) following protocols published in Guivier et al. (2011). We chose a subset of

110 available loci after excluding those that exhibited reaction inconsistency in over 30% of the
111 samples analysed. The thirteen loci were assembled in three multiplex as described in Table 2.
112 Forward primers were fluorescently labelled, and PCR products were electrophoresed by
113 MacroGen Inc. on an ABI 3730xl genetic analyser (Applied Biosystems) with a 400-HD-size
114 standard.

115 The microsatellite data were analysed using GeneMapper[®] 4.1. Micro-Checker 2.2.3 (Van
116 Oosterhout, Hutchinson, Wills & Shipley, 2004) was used to test for null alleles and large-allele
117 dropout influences. Allelic frequencies were computed by using GENETIX 4.05 (Belkhir,
118 Borsa, Chikhi, Raufaste & Bonhomme, 1996), while FSTAT (Goudet, 1995) was used to test
119 for deviations from the expected Hardy-Weinberg and linkage equilibria. Estimates of genetic
120 diversity, based on the mean allelic richness and the mean observed and expected
121 heterozygosity were computed by the DivRsity R package (Keenan, McGinnity, Cross, Crozier
122 & Prodöhl, 2013), after excluding populations with $n < 4$; allelic richness was computed using
123 the rarefaction method (Petit, Mousadik & Pons, 1998).

124 In order to investigate the extent of ordination in microsatellite data attributable to population
125 genetic structure, without using previous information on the origin of each individual, a
126 Discriminant Analysis of Principal Components (DAPC, Jombart et al., 2010) was performed
127 using Adegenet R package (Jombart et al., 2008). DAPC optimizes variation between clusters
128 and minimizes variation within them, and it is free of assumptions such as Hardy-Weinberg and
129 linkage equilibria (Jombart, Devillard & Balloux, 2010). At first data are transformed using a
130 Principal Component Analysis (PCA), and then clusters are identified using discriminant
131 analysis. The number of clusters (K) was identified by the *find.clusters* function using the “K-
132 means” algorithm, and the Bayesian information criterion (BIC) was used to choose the most
133 relevant K values for population structure. The discriminant analysis was then performed using
134 the optimal number of principal components identified by a spline interpolation of the a-scores
135 (i.e. the difference between the proportion of successful reassignment of the analysis and the
136 values obtained using random groups).

137 The population genetic structure across the study area was also investigated using the
138 Bayesian clustering algorithm implemented in TESS 2.3.1 and the geographical location of
139 individuals as prior information (Chen, Durand, Forbes & François, 2007; François & Durand
140 2010). The analysis was performed by modelling admixture using a conditional autoregressive
141 model (CAR). Preliminary analyses were carried out to assess model performance, with 20 000
142 steps (the first 5 000 were discarded as burn-in) and 10 replicates for each K value (i.e. the
143 number of clusters) between 2 and 10. The final analysis contained 100 replicates for each K

144 value, with $K = 2-10$; each run consisted of 80 000 steps, with the first 30 000 discarded as
145 burn-in. The spatial interaction parameter was initially kept at the default value (0.6), and the
146 updating option was activated. The model that best fitted the data was selected using the
147 deviance information criterion (DIC). DIC values were averaged over the 100 replicates for
148 each K value, and the most probable K value was selected as the one at which the average DIC
149 reached a plateau. For the selected K value, the estimated admixture proportions of the 10 runs
150 with the lowest DIC were averaged using CLUMPP 1.1.2 (Jakobsson & Rosemberg, 2007).

151

152 **Results**

153 The final dataset consisted of a multi-locus genotype for 76 individuals at thirteen
154 microsatellite loci, with 9.8% of missing data. Micro-Checker detected the possible occurrence
155 of null alleles at locus *Cg14E1* in population 13 and at locus *Cg15F7* in population 11. Except
156 for these two populations, no significant deviation from the Hardy-Weinberg and linkage
157 equilibria was found after the Bonferroni correction was applied. Allelic richness and mean
158 expected heterozygosity estimates for each population are shown in Table 1. Population 13
159 (Sila Massif, central Calabria) and population 4 (Foresta della Lama, Tuscan-Emilian
160 Apennines) showed the highest values of genetic diversity, whereas the lowest values of
161 heterozygosity and allelic richness were observed in population 12 (Gargano, N Apulia).

162 DAPC identified $K = 5$ as the best clustering option, being the one with the lowest BIC value.
163 The optimization of the spline interpolation of the a-scores suggests to use only the first 11
164 principal components (accounting for 54,4% of the total variance) as the more informative ones
165 for the discriminant analysis. The inspection of the scatterplot resulting by the DAPC analysis
166 (Fig. 2) clearly identified five main genetic clusters, including individuals from: i) the Western
167 Alps (pops. 1-2), ii) the Northern and Central Apennines (pops. 3-11), iii) the Gargano
168 Promontory (pop. 12), iv) the Sila Plateau and Catena Costiera massif (central Calabria, pops.
169 13-14), and v) the Aspromonte Massif (southern Calabria, pop. 15).

170 The Bayesian clustering analyses carried out with TESS revealed a clear geographic
171 structuring of genetic variation, consistent with results of the DAPC analysis. The plots of DIC
172 values versus K values reached a plateau at $K = 5$ and only a minor decrease in the DIC values
173 was observed at higher K values. The spatial distribution of the five clusters had a clear
174 geographical structure: one is widespread in the Alps and, a lower frequency, in the Northern
175 Apennines; one is found from Northern to Central Apennines; one is restricted to the Gargano
176 Promontory region; one ranges from the Sila Plateau to the Catena Costiera, and one from the

177 Catena Costiera to the Aspromonte Massif. Bar-plots showing the individual admixture
178 proportions and pie-charts showing the average proportion of each cluster within each sampled
179 population are given in Figure 3. Large genetic admixture is observed in individuals from
180 Catena Costiera (pop. 14), as well as in those from northern (pop. 3-5) and central Apennines
181 (pop. 6-11).

182

183 **Discussion**

184 The geographic structure of microsatellite variation supports the existence of five main
185 genetic clusters within *Myodes glareolus* in the Italian peninsula, in contrast to the four lineages
186 identified by the mitochondrial DNA. Indeed, previous studies identified a genetic lineage from
187 Europe widespread in northern Italy, a distinct genetic lineage in central Italy, a slightly
188 differentiated lineage restricted to the Apulian region, and a highly differentiated lineage
189 restricted to the Calabrian region (Colangelo et al., 2012; Filipi et al., 2015). Our data support
190 for the independent evolution of these lineages, and identified further sub-structuring within the
191 Calabrian region. We found evidence for genetic differentiation between populations from
192 Aspromonte and Sila, and for admixture between these two groups within the Catena Costiera.
193 Genetic differentiation is also corroborated by some morphological distinctiveness between
194 these populations, which has led to the description of distinct subspecies: *Myodes glareolus*
195 *curcio* (von Lehmann, 1961) for the Sila Massif and *Myodes glareolus hallucalis* (Thomas,
196 1906) for the Aspromonte Massif (Amori et al., 2008a; Viro & Niethammer, 1982). Moreover,
197 our results suggest strong genetic isolation between southern populations and the other
198 Apennine populations and claim for considering the Calabrian and Gargano lineages as
199 independent evolutionary and conservation units, deserving special attention in conservation
200 planning.

201 Strong genetic differentiation and high intra-specific variation of Calabrian populations is a
202 fairly common pattern in both animal and plant species (Bisconti et al., 2018; Canestrelli et al.,
203 2006; Canestrelli, Cimmaruta & Nascetti, 2008; Canestrelli, Aloise, Cecchetti & Nascetti 2010;
204 Canestrelli, Sacco & Nascetti, 2012; Chiocchio, Bisconti, Zampiglia, Nascetti & Canestrelli,
205 2017; Hewitt, 2011; Vega, Amori, Aloise, Cellini, Loy & Searle, 2010). The Calabrian region
206 is a well-known glacial refugium for temperate species in peninsular Italy, and provides one of
207 the best examples of highly sub-structured refugia, a scenario known as refugia-within-refugia
208 (Gomez & Lunt, 2007). Indeed, for most of the temperate species studied to date in this area,
209 the Calabrian region provided suitable albeit fragmented habitats through most of Pleistocene,

210 allowing long-term survival of relict populations (Bisconti et al., 2018; Senczuk, Colangelo, De
211 Simone, Aloise & Castiglia, 2017). Accordingly, the Early-Middle Pleistocene origin of
212 Calabrian bank vole populations was suggested by both molecular dating and fossil evidence
213 (Colangelo et al., 2012; Sala & Masini 2007). Moreover, palynological data support the
214 expansion of Alpine forests in Calabria during the Early-Middle Pleistocene transition, as a
215 consequence of particularly humid glacial cycles (Capraro et al., 2005; Palombo, Raia &
216 Giovinazzo, 2005). The southward expansion of forest habitats promoted southward
217 colonization of several forest and woodland-dwelling species, and might have promoted the
218 establishment of the bank vole populations in Calabria, which probably remained trapped after
219 the following shrinking of woodlands. The almost complete absence of admixture between the
220 Calabrian cluster and those located more to the north, strongly suggests an ancient isolation of
221 Calabrian populations.

222 On the other hand, the sub-structuring of the bank vole populations within the Calabrian
223 region appears of more recent origin and it is likely related to the high palaeogeographic
224 instability of this region. According to palaeogeographic reconstructions, the repeated glacio-
225 eusthatic sea level oscillations of the Pleistocene caused repeated marine floods in the lowlands,
226 turning the main mountain massifs into paleo-islands (Bonfiglio et al., 2002; Caloi, Malatesta
227 & Palombo, 1989; Cucci, 2004; Ghisetti, 1979, 1981; Tansi, Muto, Critelli & Iovine, 2007;
228 Tortorici, Monaco, Tansi & Cocina, 1995). The repeated insularization of Sila and Aspromonte
229 massifs heavily affected population structure in most of the terrestrial fauna inhabiting these
230 areas (Canestrelli et al., 2006, 2008, 2010, 2012). This historical process could also have
231 triggered genetic (and morphological) differentiation in the bank vole populations, although
232 caution should be adopted in the present case, due to the lack of molecular dating analyses.

233 The relatively high levels of genetic diversity and genetic admixture observed in these
234 populations could be explained by a more recent secondary contact between the two gene pools.
235 Under this scenario, the high level of genetic variation observed within the Calabrian
236 Pleistocene refugia would be better explained by dynamic microevolutionary processes, which
237 involve cycles of allopatric divergence and secondary contact, rather than by a prolonged
238 demographic stability (Canestrelli et al., 2010).

239 Conversely, the population from the Gargano Promontory - described as a distinct subspecies
240 *Myodes glareolus garganicus* (Hagen, 1958; see Amori et al., 2008a) - showed strong genetic
241 divergence and low genetic diversity, probably as a consequence of a historical isolation and
242 small population size, which favoured genetic erosion by drift. However, a role for the strong

243 anthropogenic impact on the bank vole's habitats, which affected this region during the last
244 decades (Parise & Pascali, 2003; Ladisa, Todorovic, & Liuzzi, 2010), cannot be excluded.

245 Populations from Alps and from north-central Apennine belong to two different genetic
246 clusters. According to previous studies (Deffontaine et al., 2005, Colangelo et al., 2012, Filipi
247 et al., 2015), the genetic cluster widespread in the Alps originated through a post-glacial
248 expansion from a western European refugium. On the contrary, the north-central Apennine
249 cluster is likely derived from an ancient Mediterranean lineage, originally widespread from
250 Balkans up to the Italian and Iberian peninsulas, and more recently isolated in an Apennine
251 refugium. The substantial levels of admixture between these two genetic clusters is consistent
252 with that showed by mtDNA data (Colangelo et al., 2012) and suggests good habitat
253 connectivity and high levels of gene flow throughout Central Apennine and Alps, at least after
254 the last glacial phase. Therefore, the genetic structure of bank vole populations in the Italian
255 peninsula, as inferred by both microsatellites and mitochondrial DNA markers, is consistent
256 with a scenario of independent evolution of multiple genetic lineages within distinct glacial
257 refugia. Interestingly, the postglacial range expansion of the lineages from Northern and Central
258 Italy was more extensive than that showed by the southern lineages, which appear still confined
259 to their refugial areas. However, we suggest caution in considering the geographic and genetic
260 isolation of Calabrian and Apulian populations. The paucity of observations from
261 geographically intermediate populations, as well as the lack of morphological and genetic data
262 does not allow to trace neither geographic nor genetic boundaries among these lineages. Further
263 research should be focused on the intermediate areas, in order to ascertain the presence of bank
264 vole populations and to estimate their genetic structure.

265 Our results have implications for the management of the bank vole populations in Southern
266 Italy. Due to its widespread distribution throughout most of Europe, *Myodes glareolus* is
267 currently categorized as *Least Concern* by the IUCN red list of threatened species, both at the
268 global (Amori et al., 2008b) and national (Rondinini, Battistoni, Peronace, & Teofli, 2013)
269 level. Nevertheless, we identified at least three unique evolutionarily significant units (Moritz,
270 1994) in Southern Italy, with narrow and endemic ranges. Assessments of their demographic
271 consistence, as well as of the current threats to their populations have to be planned in the near
272 future, in order to better integrate these endemic bank vole lineages into the regional strategy
273 of biodiversity conservation.

274 Concluding, this study highlights the importance of investigating species genetic structure
275 with a multi-marker approach, in order to find hidden diversity and fine-scale genetic
276 structuring also in supposedly well-known species. The analysis of bank vole genetic structure

277 in the Italian peninsula revealed more biological diversity than expected (see Colangelo et al.,
278 2012), suggesting the need for thorough research also in other apparently well-known taxa.
279 Finally, the evolutionary history of *Myodes glareolus* provides further evidence supporting the
280 hypothesis that Pleistocene refugia were not so stable as previously thought, and that dynamic
281 micro-evolutionary processes, triggered by the paleoclimatic and palaeogeographic instability
282 of these areas, better explain the high levels of intraspecific diversity they harbour.

283

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487 **Figure legends**

488 **Figure 1** Geographical localization of the 15 populations of *Myodes glareolus* sampled and
489 analysed. Localities are numbered as in Table 1; dashed lines delimits the main geographic
490 regions named in the text. The map was drawn using the software Canvas 11 (ACD Systems
491 of America, Inc.). Photo: *Myodes glareolus* (from Rudmer Zwerver via Photodune.net).

492

493 **Figure 2** Discriminant Analysis of Principal Components (DAPC). (a) Scatterplot resulting
494 from the DAPC performed on *Myodes glareolus* genotypes from the Italian peninsula, using
495 $K=5$ as clustering option; axes represent the first two principle components; the box shows the
496 relative contribution of the eigenvalues to the discriminant analysis; clusters are named
497 following their geographic distribution. (b) Optimal number of informative principal
498 components, suggested by the optimization of the spline interpolation of the a-scores. (c) line
499 chart showing the BIC values versus the number of genetic clusters (K) ranging from 1 to 15.

500

501 **Figure 3** Genetic structure of Italian populations of *Myodes glareolus* at 13 microsatellite
502 loci estimated using TESS. (a) The bar plot shows the admixture proportions of each
503 individual for the five genetic clusters recovered. (b) The pie diagrams on the maps show the
504 frequency distributions of each cluster among the populations; populations with only one
505 individual were grouped with the nearest population as follow: (1,2), (3,4,5), (6,7),
506 (8,9,10,11); the line chart shows the mean values of the DIC statistics (averaged over 100
507 runs) for the number of genetic clusters (K) ranging from 2 to 10. The map was drawn using
508 the software Canvas 11 (ACD Systems of America, Inc.).

509

510 **Tables**

511 **Table 1** Sample number, collecting locality, sample size (analyzed specimens), geographic
 512 coordinates, allelic richness (Ar), and expected heterozygosity (He).
 513

Sample	Locality	N	Latitude	Longitude	Ar	He
1	Gressoney	1	45.76902	7.82552	-	-
2	Aosta	11	45.73644	7.31298	2.302	0.594
3	Cantagallo	1	44.03176	11.05226	-	-
4	Foresta della Lama	10	43.81710	11.81270	2.869	0.718
5	Passignano sul Trasimeno	1	43.21675	12.13462	-	-
6	Tolfa	1	42.14959	11.93806	-	-
7	Settebagni	1	42.13705	12.53497	-	-
8	Civitella del Tronto	1	42.79336	13.67409	-	-
9	Montereale	3	42.49670	13.19747	2.564	0.564
10	L'Aquila	4	42.35686	13.38911	2.435	0.555
11	Majella Massif	6	42.14480	14.07530	2.584	0.605
12	Gargano Promontory	11	41.75540	16.01180	1.757	0.417
13	Sila Plateau	14	39.37211	16.57618	2.687	0.724
14	Catena Costiera	3	39.27578	16.09198	2.687	0.584
15	Aspromonte Massif	8	38.15910	15.92060	2.357	0.599
		76				

514

515

516 **Table 2** Marker name, repeat motif, and colour dye of the thirteen microsatellite loci
 517 assembled in three multiplex.

518

Multiplex	Marker	Repeat	Dye
A	Cg4F9	(CA) ₂₀	FAM
	Cg14E1	(CT) ₂₀	FAM
	Cg17A7	(ATGT) ₉	HEX
	Cg15F7	(CT) ₂₀	HEX
	Cg3F12	(GT) ₁₆	TAMRA
	Cg3A8	(GT) ₂₁	TAMRA
B	Cg12A7	(GA) ₂₁	FAM
	Cg17E9	(GTAT) ₉	FAM
	Cg13C12	(CT) ₂₁	HEX
C	Cg13G2	(GT) ₁₄	FAM
	Cg10H1	(GACA) ₇	FAM
	Cg10A11	(GT) ₁₅	HEX
	Cg6A1	(CAT) ₁₈	HEX

519