

A new method for the direct tracking of *in vivo* lignin nanocapsules in *Eragrostis tef* (Poaceae) tissues

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ABSTRACT

Environmental concerns have driven scientists to research new eco-friendly approaches for the preparation of nanosystems. For this purpose, novel bio-polymers have been selected. Among these, one of the most promising is lignin, which is biodegradable and biocompatible. Additionally, lignin is one of the main by-products of the paper industry and can be re-used in nanosystems building. Lignin-based nanosystems could be used in agriculture, to improve the uptake of bioactive compounds, thus avoiding soil pollution. However, the mechanism of penetration in the plant and the route of transportation within the internal plant tissues are unknown and need to be clearly elucidated. Here we present a method of lignin nanocapsules staining and tracking by fluorochrome: Fluoral Yellow 088, which is a well-suited dye for the tracking of lipids and other oil phases. Two different applications were applied: in the first one fourteen-day plants were soaked with fluorescent nanocapsules (fNCs) pure solution and in the second one, *Eragrostis tef* plants were laid down on Bibula paper and soaked with diluted fNCs solution. Wetting the roots of Teff plantlets with the pure fNCs solution resulted in the most efficient way of nanocapsule entrance. The dyeing of lignin nanocapsules allowed us to track them in *Eragrostis tef* plant tissues through microscopic observations. In particular, fNCs were proven to be able to permeate roots, reaching xylem vessels where through water pressure, they reached the leaf.

Key words: Nanocapsules tracking; Fluoral Yellow 088; *Eragrostis tef*; nanocapsule histochemistry; nanotechnology; phytonanotechnology.

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Introduction

Nanotechnology has the potential to be used in a wide range of applications, among which medicine, engineering and energy. The opportunity to modify the physicochemical properties of nanoassemblies (size, shape, surface structure, *etc.*) makes them suitable for such different applications. More recently, nanomaterials have found their use also in agriculture, where they have been designed for the delivery of active biomolecules such as pesticides, hormones and herbicides.¹⁻⁴ However, the growing interest of nanotechnology applications in agriculture^{5,6} has opened a new conception that some authors called “Phytonanotechnology”.⁷ This new application is aiming at the development of nanotechnology at large-scale production, although the perspective to extend the use of nanovectors has risen many concerns in the scientific community about their eventual release in the environment. In fact, once applied in the agricultural field, nanosystems may pollute the soil and diffuse in neighboring areas, possibly penetrating the groundwater. Besides, the long-term fate of nanomaterials is difficult to forecast, mainly due to their size, which makes tracking of nanoparticles in the environment extremely difficult.^{8,9} Therefore, new sustainable approaches are needed to overcome the possible contamination of water and soil, especially if nanovectors are planned to be used at large scale. For this reason, industries are developing eco-friendly strategies to reduce the impact not only during nanomaterial production processes,¹⁰ but also in a perspective of their release in the environment.^{11,12} Moreover, to avoid nanomaterial pollution, scientists have investigated new materials and chemical compounds which are biodegradable and eco-compatible, such as some biopolymers (*i.e.*, lignin,^{13,14} chitosan,¹⁵ and alginate¹⁶). These biopolymers have been already used for nanosystem applications in several sectors of agriculture. In particular, lignin seems to be particularly promising for nanometer containers, since it can be produced as byproduct by several industrial activities and is not toxic.^{17,18} In addition, the opportunity to tag biopolymer nanovectors with optically traceable markers could allow monitoring nanomaterials and associated chemicals in plant tissues,^{19,20} as well as their tracking in the ecosystem. This could represent a relevant further advancement and, at the same time, a powerful tool to explain the mechanism of nanovector entrance in plants, as well as its distribution in their different organs, which is still unclear. Developing new nanosystems marked with an appropriate dye would make it possible to track the pathway of nanovectors in plants from the roots to the leaves, through the stem.²¹ In this paper, a new methodology of lignin nanocapsules (NCs) traceability is described in *Eragrostis tef* (Zuccagni) Trotter seedlings (Teff) (Poaceae). The choice of *Eragrostis tef* use in the experiments depends on two reasons. The first one is that *E. tef* represents a model plant easy to cultivate and extremely resistant to different conditions. The second one is that this species has an important commercial interest since it represents one of the most cultivated cereals in Ethiopia considering that its production represents the 20% (2 10⁶ t) of the total.^{22,23} The tracking method, here described, shows for the first time the use of the fluorochrome Fluorol Yellow 088 (FY088), which was loaded in NCs (fNCs), for tracking lignin nanocapsules in plants. This fluorochrome enabled NC tracking in plantlets through optical microscope. Moreover, two administration methodologies were tested in order to understand the most efficient way of fNCs entrance in plant tissues.

Materials and Methods

Materials

Kraft Lignin (alkali lignin, low sulphur, CAS number 8068-05-1), Acetone and NaOH were purchased by Sigma Aldrich and used as received. Common olive oil was used for nanocapsule preparation. Fluorol Yellow 088 (2,8-dimethylnaphtho[3,2,1-k1]xanthene) was provided by Sigma. Milli-Q filtered water was obtained from a Millipore system (20 MΩ cm at 25°C). *Eragrostis tef* seeds were purchased in the Tigray Region of Ethiopia.

Nanocapsule preparation

NaOH was dissolved in MilliQ water to obtain a solution with a pH of 12.5. The obtained alkali solution was used to dissolve the lignin powder at a concentration of 1% w/v. The final pH after lignin dissolution was 10.5. Fluorol Yellow 088 (FY088) was diluted in olive oil (0.1% w/v) and then added to acetone, 1:1 (v/v) ratio. Once obtained an olive oil/acetone emulsion, 300 μL of this volume was added to 3mL of lignin alkali water. Then, high-intensity ultrasound technology was used to emulsify the oil phase in the aqueous medium and to facilitate the fluorochrome encapsulation inside the nanocapsules (Figure 1). For this purpose, the oil/water emulsions were sonicated by Branson 450 Digital Sonifier at the 50% of the total power (400 W), for 100s (1s pulse on and 1s pulse off, to avoid overheating). After ultrasound application, the lignin nanocapsules labeled with FY088 (fNCs) were put in contact with *Eragrostis tef* seedlings.

Nanocapsule treatment

Eragrostis tef seeds were laid down in germination dishes on bibula paper soaked by water. On the 14th day, the seedlings were transferred in Petri dishes to be treated with fNCs for 24h. Two different treatments were performed as it is shown in Figure 2: Teff plantlets were soaked with pure nanocapsules solution as it was prepared or laid down on the bibula paper soaked with fNC solution diluted 1:1 with water.

Microscope analysis

After 24h, plantlets were sectioned with Cryo-Cut Microtome (-20°C) to obtain longitudinal and transverse 20-30 μm thick sections. The sections were mounted on microscope slides. A Wild Heerbrugg stereomicroscope and a Leitz DMRB light and fluorescence microscopy with an excitation wavelength of 450-480 nm, equipped with a Nikon DS-5M digital camera were used for microscopic examinations.

Results

It is generally acknowledged, in laboratory practices that FY088 is an excellent fluorescent marker for lipids staining in fresh tissues.²⁴⁻²⁶ This specific property is the reason why FY088 was chosen in the present study. Indeed, by dissolving it in olive oil, the fluorochrome efficiently dyed the lipid phase. This phase was added to the hydrophilic component in which lignin powder had been previously dissolved at pH=12.5. Ultrasound sonication promoted the formation of an oil/water emulsion, thus improving the entrance of the FY088 present in the lipid phase and stabilizing the fluorochrome inside the lignin shell by hydrophobic repulsion. The resulting lignin NCs loaded with FY088 are shown in Figure 3. These lignin NCs were characterized and found to be a fairly homogenous distribution in size.¹⁷ The average diameter of loaded lignin NCs was about 200-250 nm while it was in the range of

300nm for the empty nanoparticles. Fluorochrome nanocapsules were tested on *Eragrostis tef* seedlings to track their entrance in plant tissues. *Eragrostis tef* seedlings were grown in germination chambers for 14 days until they presented a well-developed leaf (Figure 4A) and root (Figure 4B). The seedlings were treated with two different methodologies: the first sample plantlets were soaked with a pure fNC solution and the second group of plantlets was ordered on bibula paper and wet with a diluted fNC solution in water. After 24h, the entrance of fNCs was detected through optical microscope observations and compared with the controls (Figure 4 C,D). After 24h, fNCs entered inside Teff plantlets through the roots, reaching the xylem vessels (Figure 4F) and they were able to move inside the seedlings, without any forcing from the outside (*i.e.*, magnets).²⁷ Moreover, they rapidly arrived to leaf tissues driven by water flow, as it is shown in Figure 4 G,E). Using FY088, it was possible to track the nanocapsules path inside Teff tissues, from the roots, through the stem to the leaf (Figure 4 G,E). When the fNCs reached the xylem vessels, they were observed inside the vessels and, in large quantity, lining along the vessels walls (Figure 4F). Comparing the two methodologies of administration, the most effective one was found the procedure featuring the immersion of Teff roots in fNC solution, since the number of fNCs into the xylem vessels of the stem was higher than that found in the second groups of plantlets disposed on bibula paper soaked with fNC diluted in water 1:1.

Discussion

One of the main questions about drug administration to plants via nanocapsules is the route of entrance of the nanocapsules themselves. In this regard, relevant topics are both the penetration route through the plant tegumental tissues towards the internal tissues and the permeation through the wall and eventually plasma membranes towards the cell cytoplasm. To solve this problem, we have designed a new method for tracking lignin NCs *in vivo* with a fluorescent dye.

Both methods (roots directly in contact with NCs and roots in contact with bibula paper soaked with NCs) were found to be effective, even if the direct exposure to NCs appeared to yield quantitatively better results: more NCs were observed inside. The number of penetrated NCs could be easily detected thanks to the use of the fluorochrome. Concerning the penetration of NCs, several aspects must be considered: the penetration through the external teguments may occur either directly through the walls in the apical roots/root hairs or through stomata in the aerial part of the plant. The direct crossing of the cuticle may be possible for hydrophobic NCs.¹⁷

Penetration in internal tissues may occur through the apoplastic route (pecto-cellulosic cell walls) and/or the symplastic route through plasmodesmata, while the final entering of the cells implies a crossing of both the pectocellulosic wall and the plasma membrane. The Size exclusion limits (SEL) of plasmodesmata does not appear compatible with the penetration of NCs of the dimension employed here (200-500 nm) since typical SEL are about 1 nm^{28,29} even if SEL is a dynamic property³⁰ and viruses are able to cause an increase of the SEL itself³¹ while callose may reduce it. The crossing of the pecto-cellulosic cell walls can be done by larger particles. For instance Poly(lactic-co-glycolic) up to 50 nm nanoparticles can cross the walls, while even Nps up to 5-600 nm can cross the plasma membranes.³²

The penetration of NCs inside the xylem vessels and their translocation upwards also implies that NCs are able to cross a lignin wall and that they can be transported by the water flux inside the xylem. We observed the abundant presence of the NCs at the

level of the lignified walls of the xylem vessels and even their translocation inside the xylem. In the leaves, the NCs appeared widely spread in the parenchyma.

Previously, other stainings have been used for tracking nanoparticles, such as Alcian Blue in hyaluronic-acid-based nanoparticles in muscle cells to be visualized at Transmission Electron Microscope,³³ while our method appears to be the first used in plants for tracking lignin nanocapsules with a fluorescent microscope in plants.

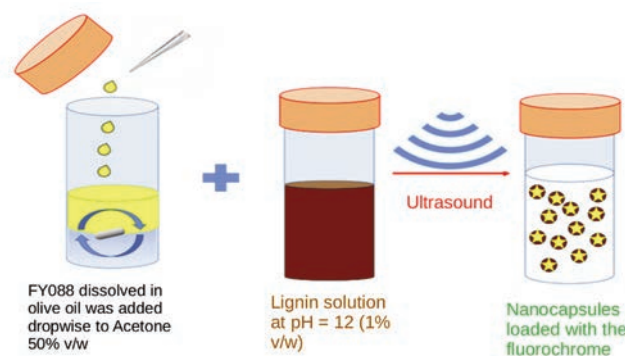


Figure 1. Scheme of fluorescent nanocapsule preparation.

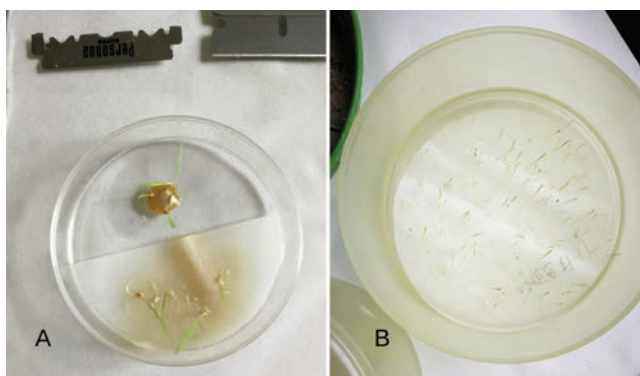


Figure 2. A) *Eragrostis tef* seedlings in germination dish. B) The two methods of fNC administration to *Eragrostis tef* seedlings.

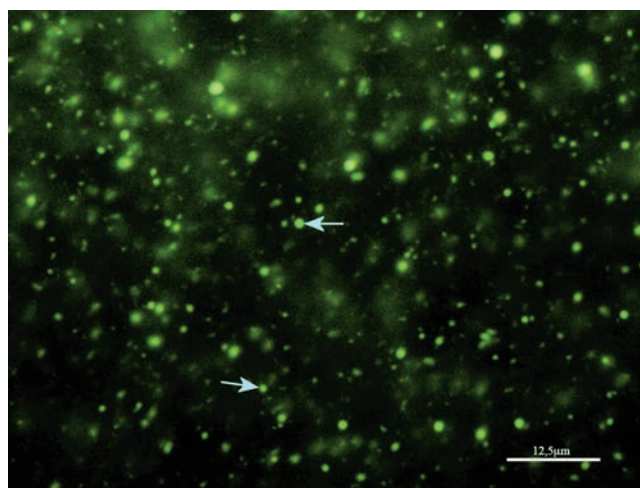


Figure 3. Fluorescence images of lignin nanocapsules loaded with FY088.

The use of the fluorochrome charged lignin NCs made it possible to assess the penetration of nanocapsules inside the plant and to identify the best way of administration. It was also possible to show that the NCs reached the xylem vessels in the root and were translocated upwards towards the leaves. In the xylem, a high number of NCs lined the xylem cells at the wall level, hence we can hypothesize that the NCs preferentially follow the walls during their penetration and are also apparently able to fol-

low the lignified walls present in the xylem vessels. The use of the fluorochrome for tracking the NCs penetration route may be extended also to NCs composed by matrixes different from lignin, provided that they are able to store hydrophobic dyes as Fluorol Yellow 088. Further development will include the investigation of lignin nanocapsules in plants with transmission electron microscope, in order to understand their interactions with the cell walls and organules.

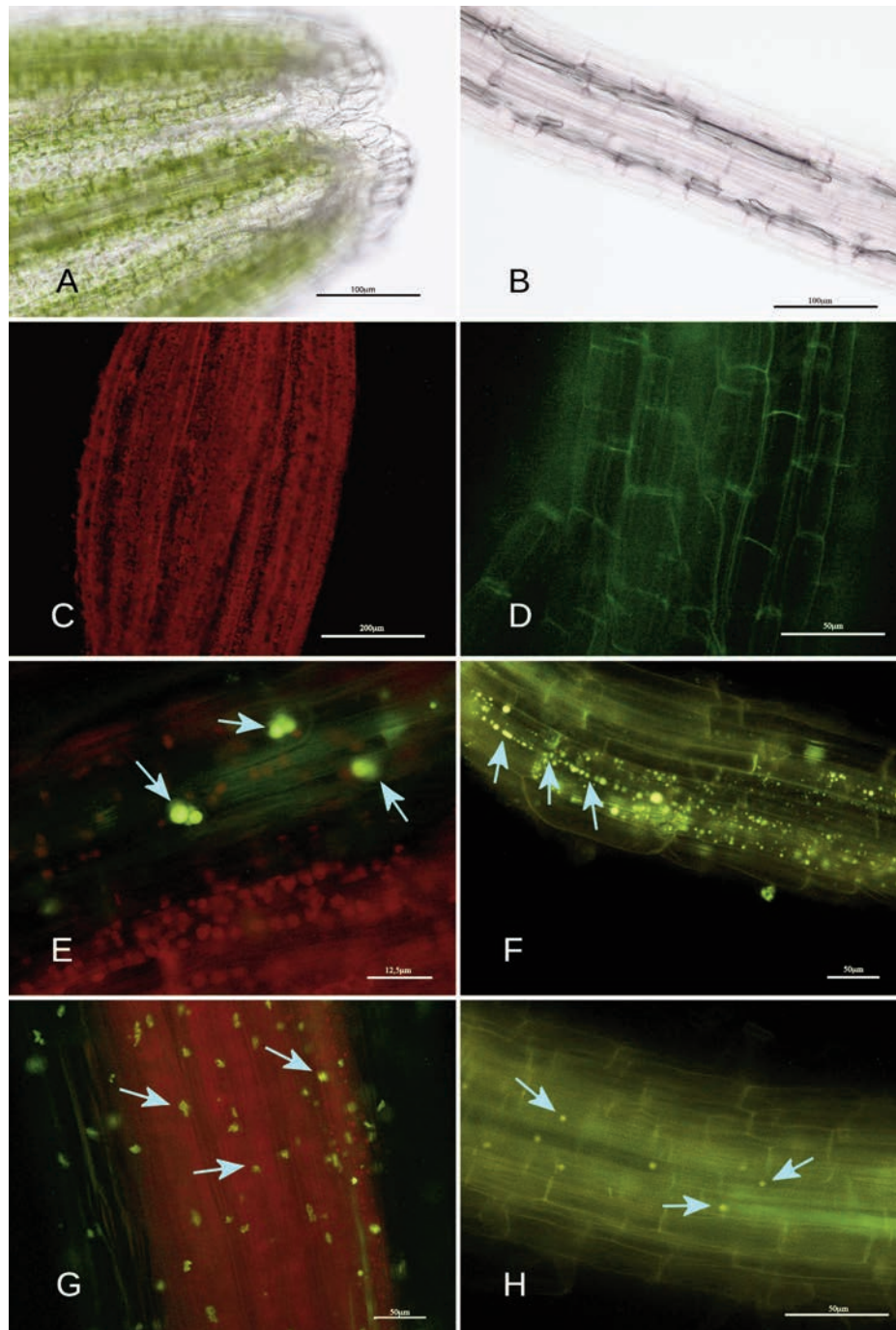


Figure 4. A) Images of a longitudinal sections of Teff leaf. B) Longitudinal section of root. C) Teff leaf without treatment. D) Fluorescence images of longitudinal sections of Teff root; no treatment: only autofluorescence. E) Teff leaf after 24 h treatment in pure NCs (some indicated with arrows) loaded with FY088. F) Teff root after 24 h treatment in pure NCs loaded with FY088; many NCs are lining the xylem walls (arrows). G) Teff leaf after 24 h treatment on blotting paper soaked with NCs (some indicated with arrows) loaded with FY088. H) Teff root after 24 h treatment in blotting paper soaked with NCs (some indicated with arrows) loaded with FY088.

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