



This document is a postprint version of an article published in *Animal Production Science* © CSIRO after peer review. To access the final edited and published work see <https://doi.org/10.1071/AN16641>

Document downloaded from:



1 **Neurophysiological development of newborn pigs: effect of the sow's parity number in eutocic**
2 **farrowings**

3

4 P. Roldan-Santiago^a, D. Mota-Rojas^{b,g}, J. Martfnez-Burnes^c, A. Velarde^d, R. Muns^e and A. Lopez-Mayagoitia^f

5

6 ^aPh.D. Program in Biological and Health Sciences, Universidad Autonoma

7 Metropolitana Campus Iztapalapa-Xochimilco-Cuajimalpa, Mexico D. F. 04960,

8 Mexico.

9 ^bUniversidad Autonoma Metropolitana, Stress Physiology and Farm Animal Welfare,

10 Department of Animal Production and Agriculture, Mexico D.F., Mexico.

11 ^cUniversidad Autonoma de Tamaulipas, Graduate Division and Research, Veterinary

12 Medicine Faculty, Ciudad Victoria, Tamaulipas, Mexico.

13 ^dIRTA, Animal Welfare Subprogram, Veinat Sies S-N, Monells 17121, Spain.

14 ^eDepartment of Obstetrics, Gynaecology and Reproduction, Faculty of Veterinary

15 Science, Chulalongkorn University, Pathumwan, Bangkok, 10330, Thailand.

16 ^fAtlantic Veterinary College, University of Prince Edward Island, Charlottetown, Canada.

17 ^gCorresponding author. Email: dmota100@yahoo.com.mx, dmota@correo.xoc.uam.mx

18 **Abstract**

19 The objective of the present study was to determine the effect of sow parity on neonatal piglet health and
20 vitality at birth. We evaluated 1505 neonate piglets, which were born of York-Landrace sows with the
21 following parity distribution: primiparous ($n = 202$), second ($n = 207$), third ($n = 211$), fourth ($n = 222$), fifth
22 ($n = 225$), sixth ($n = 218$) and seventh parity ($n = 220$). Piglets born to primiparous and seventh-parity sows

23 had the lowest and highest birthweights respectively, and showed the most marked imbalances in blood
24 gas exchanges, acid-base balance and energy profiles, as well as the highest percentage of severe
25 meconium staining of the skin and the lowest vitality scores ($P < 0.05$). In contrast, the neonates from the
26 fourth-parity sows had the highest vitality scores, required less time to reach the mother's teat, and had
27 the highest percentage of adhered umbilical cords and newborns with dyspnoea, apnoea and abnormal
28 heartbeat ($P < 0.05$). The results of this study suggest that during eutocic farrowings, the sow's parity
29 number has the following effects on newborn piglets: reduced vigour and longer latencies to begin
30 breathing, stand and take the teat. These effects are due to the presence of imbalances in gas exchanges,
31 the acid-base balance and energy profiles that occurred when the mother was a primiparous or older sow.
32 These signs indicate that the newborn piglet survived a process of intrapartum asphyxia.

33 **Introduction**

34 It is currently believed that the sow's parity number (and sow's age) is closely related to processes of neonatal
35 hypoxia (Olmos- Hernandez *et al.* 2008; Mota-Rojas *et al.* 2012a, 2012b; Muns *et al.* 2013). Also, some authors
36 postulate that the neonatal period is the most critical stage in the production cycle of farm animals, ~14% of all
37 live-born pigs have low postnatal viability due to decreased fetal blood flow and oxygen during birth (Mota-
38 Rojas *et al.* 2002,2007; Rootwelt *et al.* 2013). For these reasons, various studies have been conducted to
39 determine the principle causes of perinatal hypoxia (Mota-Rojas *et al.* 2005, 2016). Maternal factors associated
40 with neonatal hypoxia include the duration of labour, placental blood flow and stress, be that environmental
41 (such as heat stress) or stemming from altered health or endocrine status that may reduce oxygenation
42 (Alonso-Spilsbury *et al.* 2005; Mota-Rojas *et al.* 2006; Orozco-Gregorio *et al.* 2008, 2010). Another risk factor for
43 intrapartum hypoxia is the birthweight of the neonate, as newborns with low birthweights are more likely to
44 suffer oxygen restriction and the secondary effects of hypoxemia (Herpin *et al.* 1996; Marchant *et al.* 2000;
45 Orozco-Gregorio *et al.* 2008, 2010). With similar objectives in mind, other studies have determined that
46 assessing neonatal vitality is a valid clinical approach to the study of the neurofunctional status of newborns
47 and short-term survival in piglets, as low scores could be indirect indicators of intrapartum asphyxia (Mota-

48 Rojas *et al.* 2005, 2006; Gonzalez-Lozano *et al.* 2009a; Orozco-Gregorio *et al.* 2011). Other important factors
49 that predispose neonatal piglets to hypoxia are birth order, more extensive meconium staining, laceration or
50 rupture of the umbilical cord by effects of successive contractions, prolonged farrowing and dystocia (Martinez-
51 Rodriguez *et al.* 2011; Mota- Rojas *et al.* 2012a; Rootwelt *et al.* 2013). Analysing these alterations has made it
52 possible to determine the severity of asphyxia in neonate piglets (van Dijk *et al.* 2006; Trujillo *et al.* 2011),
53 puppies (Crissiuma *et al.* 2010), colts (Castagnetti *et al.* 2010) and humans (Low *et al.* 1994; da Silva *et al.* 2000).
54 Furthermore, these analyses have provided useful information that has improved our understanding of the
55 neurophysiological status of newborns (Nodwell *et al.* 2005; Orozco-Gregorio *et al.* 2008; Mota-Rojas *et al.*
56 2016). Various studies have determined the main causes of neonatal hypoxia in piglets, but without exploring
57 how the age and parity of the sow affect the physiological and metabolic profiles and survival of newborn
58 piglets, while setting aside the problematic cases of dystocia. Therefore, the objective of the present study was
59 to comprehensively evaluate the effect of the sow's parity number on the key biological parameters of
60 neonatal pigs in eutocic farrowings, including birthweight, morphology of the umbilical cord, degree of
61 meconium staining of the skin, vitality scores, gas exchanges, acid-base balance and blood energy profiles.

62

63 **Materials and methods**

64 *Ethics*

65 The experimental protocol of the study was approved by the Doctoral Commission of Biological Sciences at the
66 Universidad Autonoma Metropolitana Iztapalapa-Xochimilco in Mexico City, Mexico, in accordance with The
67 Code of Ethics of the World Medical Association (Declaration of Helsinki). All work was conducted in
68 accordance with the guidelines for the ethical use of animals in applied ethological studies, described by
69 Sherwin *et al.* (2003). Research was conducted on a commercial pig farm in central Mexico with ~1250 sows.
70 Telephones, televisions and other sources of noise were not allowed in the production and farrowing areas to
71 decrease potential sources of stress.

72 *Animals and housing conditions*

73 A total of 105 York-Landrace sows (first to seventh parity) and their litters (1505 piglets) were included. Fifteen
74 sows of each farrowing were used, and the distribution of the piglets by parity number was as follows: first (n =
75 202 piglets), second (n = 207 piglets), third (n = 211 piglets), fourth (n = 222 piglets), fifth (n = 225 piglets), sixth
76 (n = 218 piglets) and seventh (n = 220 piglets). The parameters evaluated were: neurological function, vitality
77 score, blood gas exchanges, acid-base equilibrium and energy balance. The day of insemination was considered
78 day 1 of gestation. Throughout gestation, all sows were housed in individual stalls (3.7 m²) with plasticised
79 slatted-floors and water available *ad libitum*. Sows were fed 2.5-3.0 kg/day of a corn-soybean gestation diet
80 that provided 14% crude protein, 0.70% lysine, 4.0% crude fibre and 14 MJ ME/, sufficient to meet or exceed
81 their nutritional requirements. This remained unchanged throughout pregnancy up to the 4th day before the
82 expected farrowing. Afterwards, the sows were fed a corn- soybean lactation diet (1.25% Lys, 5.0% crude fibre
83 and 20% CP) up to postpartum day 21. On day 109 of gestation, the sows were moved to a climate-controlled
84 (27°C, 60% relative humidity) farrowing room and housed in individual farrowing crates with the following
85 dimensions: 1.45-m long x 0.8-m wide x 0.57-m high. Sows and piglets had *ad libitum* access to water in
86 separate nipple drinkers. Farrowing time was induced by an intramuscular injection of prostaglandins
87 (dinoprost trometamin; Lutalyse® Pharmacia Animal Health and Upjohn, Inc., New York, NY, USA; 10 mg of
88 prostaglandin analogue IM) administered 36 h before the expected farrowing date on day 114 of gestation.

89 **Treatment**

90 To avoid interference with our measurements, no birth management or assistance was offered to the sows or
91 piglets during farrowing or at birth. The exclusion criteria applied included: sows that required obstetric
92 handling due to a delay longer than 90 min between the expulsion of piglets, sows with duration of expulsion
93 >200 min and sows that required an application of oxytocin before contractions ended. Extremely nervous or
94 obese sows with >28 mm of back fat at farrowing (determined by ultrasound Renco Preg-Alert, Minneapolis,
95 MN, USA) were excluded from electronic uterus monitoring in accordance with the methodology outlined by
96 Gonzalez- Lozano *et al.* (2009b). In addition, distocic sows that had uterine contractions >30 mm/Hg, compared
97 with 30 mm/Hg for eutocic sows, and showed uterine atony for at least 40 min were excluded; according to

98 Gonzalez-Lozano *et al.* (2010), maternal-fetal dystocia is considered when at least one of the first four piglets
99 expelled is an intrapartum stillbirth.

100 **Measurements**

101 *Electronic uterus monitoring*

102 Electronic uterus monitoring covered the time from expulsion of the first piglet to the birth of the last one, and
103 included all sows. Prior to farrowing, the uterus transducer was placed on the sow's abdomen with abundant
104 obstetric gel (Farmaceuticos Altamirano de Mexico SA de CV, Mexico City, Mexico). Electronic uterus
105 monitoring was carried out using an electronic digital cardiotocograph (Fetal Monitor Medical Systems, Inc.,
106 Wallingford, CT, USA). Patterns of uterine contractions were recorded on thermal paper according to the
107 information processed by the transducer, where every centimetre measured by the device represented a 1-min
108 record. Electronic uterus monitoring included uterus activity, and the number, duration, interval and frequency
109 of contractions, following a methodology used previously by other authors (Trujillo *et al.* 2007; Orozco-Gregorio
110 *etal.* 2008; Gonzalez-Lozano *etal.* 2009b; Mota-Rojas *et al.* 2011). Additionally, duration of farrowing was
111 registered as the time elapsed (min) from the rupture of the amniotic membrane to the expulsion of the
112 placenta.

113 *Physiological and metabolic profiles*

114 In order to assess acid-base balance and gas exchange, all blood samples of the neonates were gently collected
115 by a trained researcher. Blood was taken immediately after the expulsion of each piglet that presented regular
116 respiratory movements. The average sampling time was <20 s. Sampling was performed by retro-orbital
117 bleeding with a 100- μ L microcapillary tube in the medial canthus of the eye at a 30-45° angle towards the back
118 of the eye. The tube was inserted into the venous suborbital sinus, following the technique described by
119 Orozco-Gregorio *et al.* (2008). All blood samples were placed in glass tubes containing lithium heparin. Blood
120 pH, glucose (mg/dL), electrolytes (Na^+ , K^+ and Ca^{2+} (mEq/L)), bicarbonate (HCO_3^-) and lactate (mg/dL) levels,
121 and partial carbon dioxide (pCO₂ (mm Hg)) and oxygen pressure (pO₂ (mm Hg)) were measured with a blood

122 gas and electrolyte analyser (GEM Premier 3000; Instrumentation Laboratory Co., Lexington, MA, USA;
123 Instrumentation Laboratory SpA, Milano, Italy). All personnel involved in the sampling procedure had received
124 prior instruction and training.

125 *Vitality assessment*

126 Immediately after the blood samples were obtained, a viability score for each piglet was calculated according to
127 the scale described by Zaleski and Hacker (1993), and modified by Mota-Rojas *et al.* (2005). The following
128 variables were scored individually on a scale from 0-2: (1) heart rate (<110, 121-160 or >161 beats/min), (2) the
129 time interval from birth to first breath (>1 min, 16 s-1 min, or <15 s), (3) snout skin colour (pale, cyanotic or
130 pink), (4) the interval from birth to first standing (>5 min, 1-5 min or <1 min), and (5) meconium staining of the
131 skin (severe, mild or absent). The time to first breath was recorded as the time interval between birth and the
132 moment when thoracic movements were visible and air exhalation from the snout was noted. The degree of
133 meconium staining of the skin was also evaluated. This parameter was considered severe (2) when >50% of the
134 body surface was stained; moderate (1) when <50% was stained; and absent (0) when there was no visible
135 staining. The scores of the 5 variables were aggregated to obtain a viability score between 0-10. It is important
136 to mention that heart rate was not measured until the neonates were capable of maintaining a standing
137 posture. Additionally, the time required to connect to the teat was recorded. As the piglets were removed from
138 their pens to draw the blood samples for the purposes of this study, the time recorded for the piglet to stand
139 up and the time when it made first contact with the teat were both determined after they were returned to
140 their dams.

141 Umbilical cord integrity was evaluated at birth and classified as adhered (normal) or broken, using the
142 criteria described by Mota-Rojas *et al.* (2002). Immediately after assessing the umbilical cord, piglets were
143 individually weighed in a digital bascule (Salter Weight-Tronix Ltd, West Bromwich, UK), and their temperature
144 was taken using a tympanic membrane thermometer (ThermoScan Braun GMBH, Kronberg, Germany).
145 Neonates were considered motionless when they made no attempt to stand within 5 min after expulsion.

146 Evaluations of vitality and physiological performance were conducted by two researchers with experience in
147 handling newborn piglets. Finally, all the procedures performed with each piglet were completed in an average
148 time of 5 min.

149 *Statistical analyses*

150 All statistical analyses were carried out using JMP 8.0 (JMP Institute, Marlow, Buckinghamshire, UK). All data
151 were explored to determine distributions using the JMP univariate procedure. To study the effect of the sow's
152 parity number on the parametric variables analysed (i.e. blood metabolites, temperature, birthweight, vitality
153 scale, latency to standing, latency to take the teat), an analysis of variance was performed using general linear
154 models. In these models, the sow's parity number was included as the main effect, birthweight and duration of
155 parturition were introduced as the covariates to be considered in the model for temperature, vitality, attempts
156 to stand and latency to take the teat. When significant differences were found, a Tukey multiple comparison of
157 means test was conducted. As blood pH is a logarithmic scale, the endpoint was analysed with a Kruskal-Wallis
158 test ($P < 0.05$). To study the effect of the sow's parity number on the non-parametric variables, the Chi-squared
159 test (for number of motionless neonates, number of newborn piglets with dyspnoea, meconium staining of the
160 skin and umbilical cord integrity) was conducted ($P < 0.05$), followed by the Fisher's exact test to locate
161 differences. In addition, a linear regression analysis was carried out for the groups of neonates from the gilts
162 and seventh-parity sows. The sex was not included in statistical analyses. The significance level was set for two
163 tails in all cases ($P < 0.05$). The researchers who performed the assessment and collected the study outcomes
164 were not aware of the treatments and did not participate in selecting the animals or data analysis.

165 **Results**

166 All the piglets showed changes in vitality, neurofunctional performance, gas exchanges, acid-base balance,
167 mineral balance and energy profiles. The newborn piglets in the seventh parity group showed significant
168 changes in vitality, neurofunctional performance, gas exchange, acid-base balance, mineral balance and energy
169 profiles compared with the piglets of the other parities ($P < 0.05$). The neonates born to the second, third, fifth

170 and sixth-parity sows had a tendency toward imbalances in glucose, lactate and calcium, but the newborns
171 born to the gilts and the seventh-parity sows showed the largest imbalances for all variables. Piglets from the
172 fourth-parity sows had the lowest indexes of neurophysiological imbalances ($P < 0.05$). The neonates born to
173 the seventh-parity sows showed a mean decrease of 0.5°C in body temperature compared with the other
174 groups ($P < 0.05$). In addition, this group had the highest average birthweight, at 1682.45 ± 11.95 g, which
175 exceeded the mean

176 birthweights of all other groups ($P < 0.05$) (Table 1). With regard to neonate performance, there were
177 significant differences in vitality scores and neurofunctional performance at the time of birth among the
178 different parity groups ($P < 0.05$). However, there was no sex effect on the blood physiological, metabolic
179 variables and vitality scores in relation to parity number ($P > 0.05$).

180 *Acid-base balance*

181 Piglets born to gilts and seventh-parity sows had the lowest pO_2 (18.1 ± 0.27 and 17.75 ± 0.26 mmHg
182 respectively) compared with fourth-parity sows (27.79 ± 0.39 ; $P < 0.05$). In contrast, piglets from the third-,
183 fourth- and fifth-parity sows presented an increase in pO_2 that averaged 8.95 mmHg above that of the
184 newborns from the gilts and sixth-parity sows ($P < 0.05$). With regard to pCO_2 , the piglets born to the gilts and
185 seventh-parity sows showed an increase ($P < 0.05$) of 47 mmHg above the neonates of the second-, third-,
186 fourth-, fifth- and sixth-parity groups. However, the neonates from the fourth-parity sows had the lowest pCO_2
187 concentration (44.0 ± 0.73 mmHg) of all groups ($P < 0.05$) (Table 2). The blood pH values of the piglets from the
188 fourth- (7.33 ± 0.009), fifth- (7.27 ± 0.02) and sixth-parity sows (7.26 ± 0.02) were higher ($P < 0.05$) than those
189 of the neonates born to the gilts (7.10 ± 0.01) and the second- (7.16 ± 0.009), third- (7.22 ± 0.02) and seventh-
190 parity sows (7.04 ± 0.006), but the neonates from the latter had the lowest blood pH values of all groups ($P <$
191 0.05). Blood HCO_3^- levels were lower in neonates from the sixth- and seventh-parity sows by a mean of 3.5
192 mmol/L compared with the levels observed in neonates from the gilts and the third-, fourth- and fifth-parity
193 sows ($P < 0.05$). Finally, the piglets from the seventh-parity sows had the lowest blood bicarbonate levels (16.80
194 ± 0.21 mmol/L) of all groups ($P < 0.05$; Table 2). In the linear regression analysis, the variables pCO_2 and pO_2

195 were significantly correlated with the variables Ca²⁺ and pH in the neonates born to the gilts and seventh-
196 parity sows (P < 0.05). For the neonates born to the gilts (Table 3), pO₂ concentrations were inversely related
197 to pCO₂ (r = -0.245) and glucose concentrations (r = -0.138), but directly related to Ca²⁺ concentrations (r =
198 0.426) and pH (r = 0.162). For the neonates in the seventh-parity group (Table 4), pCO₂ concentrations were
199 inversely correlated with pO₂ concentrations (r = -0.25) and pH (r = -0.238).

200 *Gas exchange*

201 Data revealed a pO₂ decrease (P < 0.05) in the piglets born to gilts (18.11 ± 0.27 mmHg) and the seventh-parity
202 sows (17.75 ± 0.26 mmHg), compared with fourth-parity sows (P < 0.05). In contrast, piglets from the third-,
203 fourth- and fifth-parity sows presented an increase in pO₂ that averaged 8.95 mmHg above that of the
204 newborns from the gilts and sixth-parity sows (P < 0.05; Fig. 1). With regard to pCO₂, piglets born to gilts and
205 seventh- parity sows showed an increase (P < 0.05) of 47 mmHg above the neonates of the second-, third-,
206 fourth-, fifth- and sixth-parity groups. However, the neonates from the fourth-parity sows had the lowest pCO₂
207 concentration (44.0 ± 0.73 mmHg) than all other groups (P < 0.05) (Table 2).

208 *Energy metabolism*

209 Energy metabolism showed an increase of 26.91 mg/dL in the blood glucose levels of the piglets born to the
210 seventh-parity sows compared with the blood values for all groups of neonate piglets (P < 0.05). Blood lactate
211 levels, meanwhile, showed a mean increase of 34 mg/dL in the piglets born to the seventh-parity sows above
212 those determined for all groups (P < 0.05). Also, the piglets born to the third- (48.56 ± 1.29 mg/dL) and fourth-
213 parity sows (46.52 ± 1.82 mg/dL) showed blood lactate concentrations above those (P < 0.05) of the piglets
214 born to the gilts and the second-, fifth-, sixth- and seventh-parity sows (Table 2).

215 *Acid-base balance*

216 Results obtained for acid-base balance showed that the blood pH values of the piglets from the fourth- (7.33 ±
217 0.009), fifth- (7.27 ± 0.02) and sixth-parity sows (7.26 ± 0.02) were higher (P < 0.05) than those born to gilts
218 (7.10 ± 0.01) and second- (7.16 ± 0.009), third- (7.22 ± 0.02) and seventh-parity sows (7.04 ± 0.006). However,
219 the neonates from the seventh parity sows had the lowest blood pH values compared with all groups (P < 0.05).

220 Blood HCO₃⁻ levels decreased in the neonates from the sixth- and seventh-parity sows by a mean of 3.5
221 mmol/L compared with the levels observed in neonates from gilts and third-, fourth- and fifth- parity sows (P <
222 0.05). Finally, piglets from the seventh-parity sows had the lowest blood bicarbonate levels (16.80 ± 0.21
223 mmol/L) among all groups (P < 0.05) (Table 2).

224 *Mineral balance*

225 This study found an increase (P < 0.05) in Ca²⁺ concentrations in the piglets born to the gilts and seventh-parity
226 sows (1.97 ± 0.008 mmol/L, respectively, above the levels for the other groups). Blood K⁺ concentrations in the
227 neonates born to the gilts (6.22 ± 0.04 mmol/L) and seventh-parity sows (6.23 ± 0.03 mmol/L) were significantly
228 lower (P < 0.05) compared with the other groups of piglets (Table 2).

229 *Neurophysiological performance*

230 The neonates born to the seventh-parity sows showed a mean decrease of 0.5°C in body temperature
231 compared with the other groups (P < 0.05). In addition, this group also had the higher average birthweight of
232 1682.45 ± 11.95 g and exceeded the mean birthweights of all other groups (P < 0.05). Table 1 shows the effect of
233 the sow's parity number on the neurophysiological parameters measured. The parameters presented include
234 the vitality scale, latency to reach the teat, birthweight and body temperature (Table 1).

235 In terms of the vitality scale, piglets born to gilts (6.27 ± 0.11) and the seventh-parity sows (6.4 ± 0.07)
236 achieved lower scores than those of second-, third-, fourth-, fifth- and sixth-parity sows (P < 0.05); whereas the
237 newborns from the fourth-parity sows (8.28 ± 0.07) showed the highest scores (P < 0.05) of all parity groups.
238 Regarding latency to reach the teat, the study revealed that neonates born to gilts and (54.55 ± 0.98 min)
239 seventh-parity sows (48.85 ± 0.57 min) took an average 21.1 s more to reach the teat and begin suckling than
240 piglets born to the second-, third-, fourth-, fifth- and sixth-parity sows (P < 0.05). The piglets born to the fourth-
241 parity sows (23.9 ± 0.53 min) required the lowest time to connect to the mother's teat (P < 0.05).

242 *Meconium staining and umbilical cord morphology*

243 The piglets born to the gilts and seventh-parity sows had the highest percentages of severe meconium staining

244 (9.9% and 10.45%, respectively) compared with the other groups ($P < 0.05$). The neonates born to the fourth-
245 parity sows had the highest percentage of adhered umbilical cords (98.2%) compared with all groups. In
246 contrast, the piglets born to the seven-parity sows had the highest percentage of broken umbilical cords
247 (14.09%) compared with all groups (Table 5).

248 Discussion

249 An important finding in this study is that the neonates born to primiparous and seventh-parity sows in eutocic
250 farrowings suffered the most pronounced alterations of their vitality, physiological and metabolic profiles. The
251 association of these alterations in the piglets with the sow's parity indicates the existence of different factors
252 inherent in the neonates that could influence their individual performance during the birthing process. In
253 contrast, the physiological changes that the piglets experience at the moment of birth, including increased
254 blood $p\text{CO}_2$, lactate and Ca^{2+} levels, together with the low levels of pH, $p\text{O}_2$ and K^+ , may indicate a higher level
255 of intrapartum hypoxia (Ferriero 2004); however, these alterations of some aspects of the acid-base balance
256 (glucose, lactate, pH, $p\text{CO}_2$), as asphyxia indicators, are much more common in newborn piglets born from
257 dystocic farrowings (Gonzalez-Lozano *et al.* 2012), in contrast to our findings from sows with eutocic
258 farrowings, which is why we suggest that the peripartum period can be especially stressful for gilts because of
259 the new environment, the farrowing process itself (Tummaruk *et al.* 2010; Baxter *et al.* 2012; Muns *et al.* 2014).
260 In addition, the narrowness of the gilt's pelvis and the stress that primiparous sows experience compared with
261 multiparous sows could explain the high levels of hypoxia that their offspring suffered, and the increase in the
262 number and intensity of uterine contractions, two factors that might increase the risk of intrapartum hypoxia
263 (Olmos-Hernandez *et al.* 2008; Gonzalez- Lozano *et al.* 2009a). With respect to piglets of sows of the seventh
264 parity, it may be that the physiological imbalances that the piglets in our study presented are more closely
265 related to high birthweights, as this condition in neonates has been associated with respiratory imbalances in
266 blood $p\text{CO}_2$ and $p\text{O}_2$ concentrations and with acid-base imbalances (Martinez- Rodriguez *et al.* 2011). These
267 conditions are accompanied by a decrease in the number and intensity of uterine contractions (Mota-Rojas *et*
268 *al.* 2015), which may complicate the process of expelling the neonates and, consequently, generate muscular

269 fatigue in the mother. However, the findings from our study suggest that weight was more closely associated
270 with the sows' parity, since as each sow's number of parities increased, the average weight of the piglets also
271 increased. Thus, even though the heavier piglets presented imbalances in their vitality, physiological and
272 metabolic profiles, weight was not found to be a determining factor in those disequilibriums, as the neonates
273 from first-parity mothers also manifested them and, moreover, some authors have associated this variable with
274 poor thermoregulatory ability due to increased heat loss and reduced vitality, all of which affect the piglet's
275 ability to compete for the teat (Tummaruk and Sang-Gassanee 2013; Kirkden *et al.* 2013). In contrast, the
276 increase in lactate levels and decrease in blood pH were identified earlier by Orozco-Gregorio *et al.* (2008), who
277 attributed them to changes in the blood variables associated with a state of metabolic acidosis, which might
278 reflect the severity of hypoxia at birth in piglets (van Dijk *et al.* 2008). In our study, the condition of acidosis that
279 the piglets born to the gilts and sixth- parity mothers presented may also be associated with the long time
280 required to connect to their mothers' teat and lower vitality scores reported by some authors (Zaleski and
281 Hacker 1993; Herpin *et al.* 1996; Casellas *et al.* 2004; Trujillo *et al.* 2007). Another source of lactate is anaerobic
282 glycolysis: in the absence of oxygen, pyruvate is reduced to lactate through lactate dehydrogenase (Kaneko
283 1997). Therefore, the results of the present study suggest that the neonates born to the seventh- parity sows
284 experienced more acute fetal suffering. In contrast, the piglets born to the fourth-parity sows showed the
285 lowest imbalances of physiological and metabolic blood profiles, indicating the lowest level of intrapartum
286 hypoxia among the parity groups. In addition, the increase in blood Ca^{2+} concentrations observed in the piglets
287 born to the gilts and seventh-parity sows might also be the result of the process of hypoxia, because during
288 delivery the level of muscular activity increases, thus favouring the mobilisation of calcium from the newborns'
289 bones (Knol *et al.* 2002; Rydhmer *et al.* 2008; Mota-Rojas *et al.* 2015), possibly as a consequence of the high
290 muscular activity required of piglets during the expulsion process, which could be related with the incidence of
291 broken umbilical cords, severe meconium staining and low vitality of the neonates (Mota-Rojas *et al.* 2006). It
292 has been reported that older sows have reduced uterine muscle tone (Zaleski and Hacker 1993; Mota-Rojas *et*
293 *al.* 2005; Rootwelt *et al.* 2012, 2013), which is finally reflected in the degree of meconium staining of the skin

294 and intrauterine fetal distress (Mota-Rojas *et al.* 2002; Mota-Rojas *et al.* 2006; Trujillo *et al.* 2007; Mota-Rojas
295 *et al.* 2016). The increase in blood glucose ascertained for the neonates from the seventh-parity sows could
296 indicate greater neonatal distress, reflecting the piglets' efforts to compensate for the hypoxic process (Randall
297 1972; Gonzalez-Lozano *et al.* 2010; Mota-Rojas *et al.* 2011, 2015). The high levels of blood glucose found in the
298 present study are similar to those reported by Herpin *et al.* (1996) and Martinez-Rodriguez *et al.* (2011), who
299 determined glucose concentrations of 102.3 ± 6.5 mg/dL in neonate piglets with intrapartum asphyxia and high
300 birthweight. However, Mota-Rojas *et al.* (2005) suggested that high neonatal glucose concentrations could be
301 characteristic of a shorter period of asphyxia during which piglets succeed in conserving their energy
302 substrates, whereas low glucose concentrations would be associated with a prolonged period of hypoxia, and
303 greater utilisation and consumption of energy reserves. In our results, blood glucose levels could be an
304 indicator of acute stress that may have increased due to the energy requirements that exist as a consequence
305 of the lack of oxygenation of the fetus. This is justified by the high percentage of piglets that were born with
306 broken umbilical cords at the moment of expulsion, coupled with apnoea, severe meconium staining, low
307 mobility and, as a result, low scores on the vitality scale. In this regard, Herpin *et al.* (1996) mention that
308 glucose is a good marker of neonatal distress and may reflect the capability of the piglets to compensate the
309 process. At birth, the increments in glucose plasma levels may be explained by a release of catecholamines and
310 stimulation of liver glycogenolysis secondary to the intrapartum asphyxia. All the findings presented above
311 indicate that intrapartum hypoxia was intensified in the neonates born to the gilts and seventh-parity sows,
312 reducing their vitality scores and their capacity to search for the maternal teat, despite the fact that they were
313 products of eutocic births with no clinical evidence of the obstetric problems associated with distocic births, as
314 can be appreciated in the present study.

315

316 **Conclusions**

317 In the present study, the parity of sows was seen to have effects on the health and vitality of newborn piglets,
318 as the neonates born to first- and seventh-parity dams were shown to have low vitality scores reflected by the

319 presence of severe meconium staining, cyanotic skin, and longer latencies to both stand and take the mother's
320 teat. Those animals presented imbalances in gas exchanges, the acid-base balance and energy profiles, even in
321 births considered eutocic. These results suggest that not only births classified as distocic affect the physiological
322 responses of newborn piglets. Upon evaluating the parity of the sows that had eutocic births, the study was
323 able to determine that the neonates produced by the fourth-parity dams had the fewest physiological
324 alterations and the highest vitality scores, as they required less time to stand and connect to the teat, and had
325 pink skin with adhered umbilical cords. Through these findings, the present study demonstrated that the
326 alterations in the health of newborn piglets and low vitality scores due to the effects of the parity of the sow
327 provide evidence that the newborn piglets survived a process of intrapartum asphyxia.

328

329 **Conflicts of interest**

330 The authors declare no conflicts of interest.

331

332 **Acknowledgements**

333 This manuscript was part of the Ph.D. thesis by Patricia Roldan-Santiago, who is enrolled in the Doctoral
334 Program in Biological Sciences and Health at the Universidad Autonoma Metropolitana in Mexico, and
335 supported by the Consejo Nacional de Ciencia y Tecnologia, Mexico (CONACYT).

336

337 **References**

338 Alonso-Spilsbury M, Mota-Rojas D, Villanueva-Garcia D, Martinez-Bumes J, Orozco H, Ramirez-Necoechea R,

339 Trujillo ME (2005) Perinatal asphyxia pathophysiology in pig and human: a review. *Animal Reproduction*

340 *Science* 90, 1-30. doi:[10.1016/j.anireprosci.2005.01.007](https://doi.org/10.1016/j.anireprosci.2005.01.007)

341 Baxter EM, Jarvis S, Palarea-Albaladejo J, Edwards SA (2012) The Weaker Sex? The Propensity for Male-Biased

342 Piglet Mortality. *PLoS One* 7, e30318. doi:[10.1371/journal.pone.0030318](https://doi.org/10.1371/journal.pone.0030318)

343 Casellas J, Rauw WM, Piedrafita J, Sanchez A, Arque M, Noguera JL (2004) Viability of Iberian x Meishan F-2
344 newborn pigs. I. Analysis of physiological and vitality variables. *Journal of Animal Science* 82, 1919-1924.
345 doi:[10.2527/2004.8271919x](https://doi.org/10.2527/2004.8271919x)

346 Castagnetti C, Pirrone A, Mariella J, Mari G (2010) Venous blood lactate evaluation in equine neonatal intensive
347 care. *Theriogenology* 73, 343-357. doi:[10.1016/j.theriogenology.2009.09.018](https://doi.org/10.1016/j.theriogenology.2009.09.018)

348 Crissiuma AL, Juppa CJ, de Almeida FM, Gershony LC, Labarthe NV (2010) Influence of the order of birth on
349 blood gasometry parameters in the fetal neonatal transitional period of dogs born by elective caesarean
350 parturition. *International Journal of Applied Research in Veterinary Medicine* 8, 7-15.

351 da Silva S, Hennebert N, Denis R, Wayenberg JL (2000) Clinical value of a single postnatal lactate measurement
352 after intrapartum asphyxia. *Acta Paediatrica (Oslo, Norway)* 89, 320-323. doi:[10.1111/j.1651-2227.](https://doi.org/10.1111/j.1651-2227.2000.tb01334.x)
353 [2000.tb01334.x](https://doi.org/10.1111/j.1651-2227.2000.tb01334.x)

354 Ferriero DM (2004) Medical progress: neonatal brain injury. *The New England Journal of Medicine* 351, 1985-
355 1995. doi:[10.1056/NEJM ra041996](https://doi.org/10.1056/NEJM ra041996)

356 Gonzalez-Lozano M, Mota-Rojas D, Velazquez-Armenta EY, Nava- Ocampo AA, Hernandez-Gonzalez R, Becerril-
357 Herrera M, Trujillo- Ortega ME, Alonso-Spilsbury M (2009a) Obstetric and fetal outcomes in dystocic and
358 eutocic sows to an injection of exogenous oxytocin during farrowing. *The Canadian Veterinary Journal. La*
359 *Revue Veterinaire Canadienne* 50, 1273-1277.

360 Gonzalez-Lozano M, Ortega MET, Becerril-Herrera M, Alonso-Spilsbury M, Ramirez-Necoechea R, Hernandez-
361 Gonzalez R, Mota-Rojas D (2009b) Effects of oxytocin on critical blood variables from dystocic sows.
362 *Veterinaria (Mexico)* 40, 231-245.

363 Gonzalez-Lozano M, Ortega MET, Becerril-Herrera M, Alonso-Spilsbury M, Rosales-Torres AM, Mota-Rojas D
364 (2010) Uterine activity and fetal electronic monitoring in parturient sows treated with vetrabutrin
365 chlorhydrate. *Journal of Veterinary Pharmacology and Therapeutics* 33, 28-34. doi:[10.1111/j.1365-](https://doi.org/10.1111/j.1365-2885.2009.01094.x)
366 [2885.2009.01094.x](https://doi.org/10.1111/j.1365-2885.2009.01094.x)

367 Gonzalez-Lozano M, Ortega MET, Alonso-Spilsbury M, Rosales AM, Ramirez-Necoechea R, Gonzalez-Maciél A,

368 Mota-Rojas D (2012) Vetrabutine clorhydrate use in dystotic farrowings minimizes hemodynamic sequels in
369 piglets. *Theriogenology* 78, 455-461. doi:[10.1016/j.theriogenology.2012.02.025](https://doi.org/10.1016/j.theriogenology.2012.02.025)

370 Herpin P, LeDividich J, Hulin JC, Fillaut M, DeMarco F, Bertin R (1996) Effect of the level of asphyxia during
371 delivery on viability at birth and early postnatal vitality of newborn pigs. *Journal of Animal Science* 74, 2067-
372 2075. doi: [10.2527/1996.7492067x](https://doi.org/10.2527/1996.7492067x)

373 Kaneko JJ (1997) Carbohydrate metabolism and its diseases. In 'Clinical biochemistry of domestic animals'. 5 th
374 edn. (Eds JJ Kaneko, JW Harvey, ML Bruss) pp. 45-81. (Academic Press: San Diego, CA)

375 Kirkden RD, Broom DM, Andersen IL (2013) Piglet mortality: the impact of induction of farrowing using
376 prostaglandins and oxytocin. *Animal Reproduction Science* 138, 14-24.
377 doi:[10.1016/j.anireprosci.2013.02.009](https://doi.org/10.1016/j.anireprosci.2013.02.009)

378 Knol EF, Leenhouders JI, van der Lende T (2002) Genetic aspects of piglet survival. *Livestock Production Science*
379 78, 47-55. doi:[10.1016/S0301-6226\(02\)00184-7](https://doi.org/10.1016/S0301-6226(02)00184-7)

380 Low JA, Panagiotopoulos C, Derrick EJ (1994) Newborn complications after intrapartum asphyxia with
381 metabolic-acidosis in the term fetus. *American Journal of Obstetrics and Gynecology* 170, 1081-1087.
382 doi:[10.1016/S0002-9378\(94\)70101-6](https://doi.org/10.1016/S0002-9378(94)70101-6)

383 Marchant JN, Rudd AR, Mendl MT, Broom DM, Meredith MJ, Corning S, Simmins PH (2000) Timing and causes
384 of piglet mortality in alternative and conventional farrowing systems. *The Veterinary Record* 147, 209-214.
385 doi:[10.1136/vr.147.8.209](https://doi.org/10.1136/vr.147.8.209)

386 Martinez-Rodriguez R, Mota-Rojas D, Ortega MET, Orozco-Gregorio H, Hernandez-Gonzalez R, Roldan-Santiago
387 P, Mora-Medina P, Alonso- Spilsbury M, Rosales-Torres A, Ramirez-Necoechea R (2011) Physiological
388 response to hypoxia in piglets of different birth weight. *Italian Journal of Animal Science* 10, e56.
389 doi:[10.4081/ijas.2011.e56](https://doi.org/10.4081/ijas.2011.e56)

390 Mota-Rojas D, Martinez-Burnes J, Alonso-Spilsbury ML, Ramirez- Necoechea R, Lopez A (2002) Effect of
391 oxytocin treatment in sows on umbilical cord morphology, meconium staining, and neonatal mortality of
392 piglets. *American Journal of Veterinary Research* 63, 1571-1574. doi:[10.2460/ajvr.2002.63.1571](https://doi.org/10.2460/ajvr.2002.63.1571)

393 Mota-Rojas D, Martinez-Burnes J, Ortega MET, Lopez A, Rosales AM, Ramirez R, Orozco H, Merino A, Alonso-
394 Spilsbury M (2005) Uterine and fetal asphyxia monitoring in parturient sows treated with oxytocin. *Animal*
395 *Reproduction Science* 86, 131-141. doi:[10.1016/j.anireprosci.2004.06.004](https://doi.org/10.1016/j.anireprosci.2004.06.004)

396 Mota-Rojas D, Martinez-Burnes J, Alonso-Spilsbury ML, Lopez A, Ramirez-Necoechea R, Trujillo-Ortega ME,
397 Medina-Hernandez FJ, de la Cruz NI, Albores-Torres V, Loredó-Osti J (2006) Meconium staining of the skin
398 and meconium aspiration in porcine intrapartum stillbirths. *Livestock Science* 102, 155-162.
399 doi:[10.1016/j.livsci.2006.01.002](https://doi.org/10.1016/j.livsci.2006.01.002)

400 Mota-Rojas D, Villanueva-Garcia D, Velazquez-Armenta EY, Nava-Ocampo AA, Ramirez-Necoechea R, Alonso-
401 Spilsbury M (2007) Influence of time at which oxytocin is administered during labor on uterine activity and
402 perinatal death in pigs. *Biological Research* 40, 55-63. doi:[10.4067/S0716-97602007000100006](https://doi.org/10.4067/S0716-97602007000100006)

403 Mota-Rojas D, Orozco-Gregorio H, Villanueva-Garcia D, Bonilla-Jaime H, Suarez-Bonilla X, Hernandez-Gonzalez
404 R, Roldan-Santiago P, Trujillo-Ortega ME (2011) Foetal and neonatal energy metabolism in pigs and
405 humans: a review. *Veterinarni Medicina* 56, 215-225.

406 Mota-Rojas D, Martinez-Burnes J, Villanueva-Garcia D, Roldan-Santiago P, Ortega MET, Orozco-Gregorio H,
407 Bonilla-Jaime H, Lopez-Mayagoitia A (2012a) Animal welfare in the newborn piglet: a review. *Veterinarni*
408 *Medicina* 57, 338-349.

409 Mota-Rojas D, Villanueva-Garcia D, Hernandez-Gonzalez R, Roldan-Santiago P, Martinez-Rodriguez R, Gonzalez-
410 Meneses B, Sanchez-Hernandez M, Trujillo-Ortega ME (2012b) Assessment of the vitality of the newborn:
411 an overview. *Scientific Research and Essays* 7, 712-718. doi:[10.5897/SRE11.869](https://doi.org/10.5897/SRE11.869)

412 Mota-Rojas D, Fierro R, Roldan-Santiago P, Orozco-Gregorio H, Gonzalez-Lozano M, Bonilla H, Martinez-
413 Rodriguez R, Garcia-Herrera R, Mora-Medina P, Flores-Peinado S, Sanchez M, Ramirez-Necoechea R (2015)
414 Outcomes of gestation length in relation to farrowing performance in sows and daily weight gain and
415 metabolic profiles in piglets. *Animal Production Science* 55, 93-100. doi:[10.1071/AN13175](https://doi.org/10.1071/AN13175)

416 Mota-Rojas D, Lopez-Mayagoitia A, Muns R, Roldan-Santiago P, Mainau E, Martinez-Burnes J (2016) Piglet
417 welfare. Chapter 5. In 'Animal welfare: a global vision en Spanish America'. 3rd edn. (Eds D Mota-Rojas, A

418 Velarde, SM Huertas-Canen, MN Cajiao-Pachon) pp. 51-62. (Elsevier: Barcelona, Spain)

419 Muns R, Manzanilla EG, Sol C, Manteca X, Gasa J (2013) Piglet behavior as a measure of vitality and its influence
420 on piglet survival and growth during lactation. *Journal of Animal Science* 91,1838-1843. doi:[10.2527/ jas.2012-](https://doi.org/10.2527/jas.2012-5501)
421 [5501](https://doi.org/10.2527/jas.2012-5501)

422 Muns R, Manzanilla EG, Manteca X, Gasa J (2014) Effect of gestation management system on gilt and piglet
423 performance. *Animal Welfare (SouthMimms, England)* 23, 343-351. doi:[10.7120/09627286.23.3.343](https://doi.org/10.7120/09627286.23.3.343)

424 Nodwell A, Carmichael L, Ross M, Richardson B (2005) Placental compared with umbilical cord blood to assess
425 fetal blood gas and acid- base status. *Obstetrics and Gynecology* 105, 129-138. doi:[10.1097/01.](https://doi.org/10.1097/01.AOG.0000146635.51033.9d)
426 [AOG.0000146635.51033.9d](https://doi.org/10.1097/01.AOG.0000146635.51033.9d)

427 Olmos-Hernandez A, Ortega MET, Alonso-Spilsbury M, Sanchez-Aparicio P, Ramirez-Necoechea R, Mota-Rojas A
428 (2008) Foetal monitoring, uterine dynamics and reproductive performance in spontaneous farrowings in
429 sows. *Journal of Applied Animal Research* 33, 181-185. doi:[10.1080/ 09712119.2008.9706923](https://doi.org/10.1080/09712119.2008.9706923)

430 Orozco-Gregorio H, Mota-Rojas D, Alonso-Spilsbury M, Olmos-Hernandez A, Ramirez-Necoechea R, Velazquez-
431 Armenta EY, Nava-Ocampo AA, Hernandez-Gonzalez R, Trujillo-Ortega ME, Villanueva-Garcia D (2008) Short-
432 term neurophysiologic consequences of intrapartum asphyxia in piglets born by spontaneous parturition.
433 *The International Journal of Neuroscience* 118, 1299-1315. doi:[10.1080/00207450701872846](https://doi.org/10.1080/00207450701872846)

434 Orozco-Gregorio H, Mota-Rojas D, Bonilla-Jaime H, Trujillo-Ortega ME, Becerril-Herrera M, Hernandez-Gonzalez
435 R, Villanueva-Garcia D (2010) Effects of administration of caffeine on metabolic variables in neonatal pigs
436 with peripartum asphyxia. *American Journal of Veterinary Research* 71, 1214-1219.
437 doi:[10.2460/ajvr.71.10.1214](https://doi.org/10.2460/ajvr.71.10.1214)

438 Orozco-Gregorio H, Mota-Rojas D, Villanueva D, Bonilla-Jaime H, Suarez-Bonilla X, Torres-Gonzalez L, Bolanos D,
439 Gonzalez RH, Martinez-Rodriguez R (2011) Caffeine therapy for apnoea of prematurity: pharmacological
440 treatment. *African Journal of Pharmacy and Pharmacology* 5, 564-571.

441 Randall GCB (1972) Observation on parturition in sow. II. Factors influencing stillbirth and perinatal mortality.
442 *The Veterinary Record* 90, 183-186. doi:[10.1136/vr.90.7.183](https://doi.org/10.1136/vr.90.7.183)

443 Rootwelt V, Reksen O, Farstad W, Framstad T (2012) Associations between intrapartum death and piglet,
444 placental, and umbilical characteristics. *Journal of Animal Science* 90, 4289-4296. doi:[10.2527/jas.2012-5238](https://doi.org/10.2527/jas.2012-5238)

445 Rootwelt V, Reksen O, Farstad W, Framstad T (2013) Postpartum deaths: Piglet, placental, and umbilical
446 characteristics. *Journal of Animal Science* 91, 2647-2656. doi:[10.2527/jas.2012-5531](https://doi.org/10.2527/jas.2012-5531)

447 Rydhmer L, Lundeheim N, Canario L (2008) Genetic correlations between gestation length, piglet survival and
448 early growth. *Livestock Science* 115, 287-293. doi:[10.1016/j.livsci.2007.08.014](https://doi.org/10.1016/j.livsci.2007.08.014)

449 Sherwin CM, Christiansen SB, Duncan IJ, Erhard HW, Lay DC, Mench JA, O'Connor CE, Petherick JC (2003)
450 Guidelines for the ethical use of animals in applied ethology studies. *Applied Animal Behaviour Science* 81,
451 291-305. doi:[10.1016/S0168-1591\(02\)00288-5](https://doi.org/10.1016/S0168-1591(02)00288-5)

452 Trujillo OM, Mota-Rojas D, Olmos-Hernandez A, Alonso-Spilsbury M, Gonzalez-Lozano M, Orozco-Gregorio H,
453 Ramirez-Necoechea R, Nava-Ocampo AA (2007) A study of piglets born by spontaneous parturition under
454 uncontrolled conditions: Could this be a naturalistic model for the study of intrapartum asphyxia? *Acta*
455 *Biomedica* 78,29-35.

456 Trujillo OM, Mota-Rojas D, Juarez O, Villanueva-Garcia D, Roldan-Santiago P, Becerril-Herrera M, Hernandez-
457 Gonzalez R, Mora-Medina P, Alonso- Spilsbury M, Rosales AM, Martinez-Rodriguez R, Ramirez-Necoechea R
458 (2011) Porcine neonates failing vitality score: physio-metabolic profile and latency to the first teat contact.
459 *Czech Journal of Animal Science* 56, 499-508.

460 Tummaruk P, Sang-Gassanee K (2013) Effect of farrowing duration, parity number and the type of anti-
461 inflammatory drug on postparturient disorders in sows: a clinical study. *Tropical Animal Health and*
462 *Production* 45, 1071-1077. doi:[10.1007/s11250-012-0315-x](https://doi.org/10.1007/s11250-012-0315-x)

463 Tummaruk P, Tantasuparuk W, Techakumphu M, Kunavongkrit A (2010) Seasonal influences on the litter size at
464 birth of pigs are more pronounced in the gilt than sow litters. *The Journal of Agricultural Science* 148,
465 421-432. doi:[10.1017/S0021859610000110](https://doi.org/10.1017/S0021859610000110)

466 van Dijk AJ, van der Lende T, Taverne MAM (2006) Acid-base balance of umbilical artery blood of liveborn
467 piglets at birth and its relation with factors affecting delivery of individual piglets. *Theriogenology* 66, 1824-

468 1833. doi:[10.1016/j.theriogenology.2006.04.035](https://doi.org/10.1016/j.theriogenology.2006.04.035)

469 van Dijk AJ, van Loon JP, Taverne MA, Jonker FH (2008) Umbilical cord clamping in term piglets: a useful model

470 to study perinatal asphyxia? *Theriogenology* 70, 662-674. doi:[10.1016/j.theriogenology.2008.04.044](https://doi.org/10.1016/j.theriogenology.2008.04.044)

471 Zaleski HM, Hacker RR (1993) Effect of oxygen and neostigmine on stillbirth and pig viability. *Journal of Animal*

472 *Science* 71, 298-305. doi:[10.2527/1993.712298x](https://doi.org/10.2527/1993.712298x)

473

474

Table 1. Effect of the sow's parity number on the neurophysiological parameters
 a–f, Lowercase letters indicate differences among parity number groups within the blood variable ($P < 0.05$); n , number of weaned piglets sampled; s.e., standard error

Variables	Parity 1 ($n = 202$) Mean \pm s.e.	Parity 2 ($n = 207$) Mean \pm s.e.	Parity 3 ($n = 211$) Mean \pm s.e.	Parity 4 ($n = 222$) Mean \pm s.e.	Parity 5 ($n = 225$) Mean \pm s.e.	Parity 6 ($n = 218$) Mean \pm s.e.	Parity 7 ($n = 220$) Mean \pm s.e.
Temperature ($^{\circ}\text{C}$)	37.16 \pm 0.03c	37.33 \pm 0.04c	37.56 \pm 0.04b	37.83 \pm 0.04a	37.64 \pm 0.04b	37.30 \pm 0.04c	36.97 \pm 0.02d
Birthweight (g)	1347.66 \pm 19.01f	1422.2 \pm 13.21ef	1427.33 \pm 12.73de	1489.01 \pm 17.76cd	1537.96 \pm 14.79bc	1594.08 \pm 17.75b	1682.45 \pm 11.95a
Vitality scale	6.27 \pm 0.11e	7.07 \pm 0.07cd	7.32 \pm 0.06c	8.28 \pm 0.07a	7.79 \pm 0.06b	6.9 \pm 0.08d	6.4 \pm 0.07e
Latency to connect to teat (min)	54.55 \pm 0.98a	36.61 \pm 1.05c	28.09 \pm 0.68d	23.9 \pm 0.53e	29.57 \pm 0.92d	34.83 \pm 1.18c	48.85 \pm 0.57b

Table 2. Mean and standard error of the duration of parturition and physiological blood profiles of neonate piglets according to the sow's parity number

a–c, Lowercase letters in the same row indicate differences among groups within the blood variable; Tukey ($P < 0.05$); n , number of weaned piglets sampled; s.e., standard error

Blood trait	Parity 1 ($n = 202$)	Parity 2 ($n = 207$)	Parity 3 ($n = 211$)	Parity 4 ($n = 222$)	Parity 5 ($n = 225$)	Parity 6 ($n = 218$)	Parity 7 ($n = 220$)
	Mean \pm s.e.	Mean \pm s.e.	Mean \pm s.e.	Mean \pm s.e.	Mean \pm s.e.	Mean \pm s.e.	Mean \pm s.e.
Duration of parturition (min)	184 \pm 16.25a	196 \pm 14.02a	167 \pm 18.3a	194 \pm 15.16a	185 \pm 17.25a	198 \pm 14.76a	187 \pm 16.78a
pH	7.10 \pm 0.01de	7.16 \pm 0.009cd	7.22 \pm 0.02bc	7.33 \pm 0.009a	7.27 \pm 0.02ab	7.26 \pm 0.02b	7.04 \pm 0.006e
Glucose (mg/dL)	99.48 \pm 2.56b	72.23 \pm 1.24d	79.75 \pm 0.96c	80.93 \pm 1.17c	75.69 \pm 1.16cd	70.31 \pm 0.79d	106.65 \pm 2.22a
Lactate (mg/dL)	88.58 \pm 1.26b	59.90 \pm 1.65d	48.56 \pm 1.29e	46.52 \pm 1.82e	58.61 \pm 1.89d	77.62 \pm 1.60c	97.34 \pm 0.76a
HCO ₃ ⁻ (mmol/L)	20.45 \pm 0.16cd	21.05 \pm 0.16c	22.22 \pm 0.15b	23.30 \pm 0.18a	22.42 \pm 0.19b	19.88 \pm 0.22d	16.80 \pm 0.21e
Ca ²⁺ (mmol/L)	1.97 \pm 0.008a	1.69 \pm 0.008b	1.64 \pm 0.01c	1.56 \pm 0.01d	1.61 \pm 0.01c	1.62 \pm 0.008c	1.97 \pm 0.008a
Na ⁺ (mmol/L)	135.20 \pm 0.20ab	134.96 \pm 0.23b	135.24 \pm 0.25ab	136.05 \pm 0.24a	135.78 \pm 0.25ab	135.69 \pm 0.26ab	135.06 \pm 0.18b
K ⁺ (mmol/L)	6.22 \pm 0.04b	6.60 \pm 0.03a	6.72 \pm 0.04a	6.64 \pm 0.03a	6.59 \pm 0.03a	6.61 \pm 0.04a	6.23 \pm 0.03b

Table 3. Correlations of the physiological and metabolic blood variables of the neonate piglets born to the gilts

The independent variables were numbered according to the correlation coefficient (R -value). The final F - and P -values observed in this table are from the analysis of variance of the linear regression analyses ($y = b + mx$). The parameter in the linear equation is shown in relation to the corresponding mean standard error

Dependent variable (y)	Independent variable (x)	B	s.e.	R	R^2	F -value	P -value
pO ₂ (mmHg)	pCO ₂ (mmHg)	-0.073	0.02	-0.245	0.06	12.812	<0.0001
	Lactate (mg/dL)	-0.028	0.015	-0.129	0.017	3.381	0.067
	HCO ₃ ⁻ (mmol/L)	0.022	0.117	0.013	0.0001	0.36	0.85
	Ca ²⁺ (mmol/L)	0.199	0.03	0.426	0.182	44.371	<0.0001
	Glucose (mg/dL)	-0.015	0.008	-0.138	0.019	3.862	0.051
	pH	4.373	1.886	0.162	0.026	5.376	0.021

Table 4. Correlations of the physiological and metabolic blood variables of the neonate piglets born to the seventh-parity sows

The independent variables were numbered according to the correlation coefficient (*R*-value). The final *F*- and *P*-value observed in this table are from the analysis of variance of the linear regression analyses ($y = b + mx$). The parameter in the linear equation is shown in relation to the corresponding mean standard error

Dependent variable (y)	Independent variable (x)	B	s.e.	<i>R</i>	<i>R</i> ²	<i>F</i> -value	<i>P</i> -value
pCO ₂ (mmHg)	pO ₂ (mmHg)	-0.844	0.221	-0.25	0.063	14.56	<0.0001
	Lactate (mg/dL)	0.015	0.08	0.012	0.0001	0.034	0.855
	HCO ₃ ⁻ (mmol/L)	-0.263	0.277	-0.64	0.004	0.903	0.343
	Ca ²⁺ (mmol/L)	20.9	6.614	0.209	0.044	9.99	0.002
	Glucose (mg/dL)	0.03	0.027	0.074	0.005	1.201	0.274
	pH	-30.461	8.423	-0.238	0.057	13.079	<0.0001

Table 5. Assessment of the performance at birth of neonate piglets due to the effect of the sow's parity number

Variables	Pearson's Chi-squared test ($P < 0.05$)							Prob > ChiSq <i>P</i>
	Parity 1 (n = 202) n (%)	Parity 2 (n = 207) n (%)	Parity 3 (n = 211) n (%)	Parity 4 (n = 222) n (%)	Parity 5 (n = 225) n (%)	Parity 6 (n = 218) n (%)	Parity 7 (n = 220) n (%)	
Neonates with adhered umbilical cord	178 (88.11)	191 (92.27)	196 (92.89)	218 (98.2)	210 (93.33)	192 (88.07)	189 (85.91)	<0.05
Neonates with broken umbilical cord	24 (11.88)	14 (6.76)	16(7.58)	4(1.8)	15 (6.67)	26 (11.93)	31 (14.09)	<0.05
Neonates motionless	16 (7.92)	2(1.93)	6 (2.84)	1 (0.45)	7 (3.11)	20 (9.17)	22 (10.0)	<0.05
Neonates with dyspnoea	12 (5.94)	6 (2.89)	3 (1.42)	1 (0.45)	3 (1.33)	10 (4.58)	17 (7.72)	<0.05
Neonates with severe meconium staining	20 (9.90)	10 (4.83)	5 (2.36)	2 (0.90)	7 (3.11)	17 (7.79)	23 (10.45)	<0.05
Neonates with apnoea and heart beat	7 (3.46)	3 (1.44)	2 (0.94)	1 (0.45)	3 (1.33)	6 (2.75)	9 (4.09)	<0.05

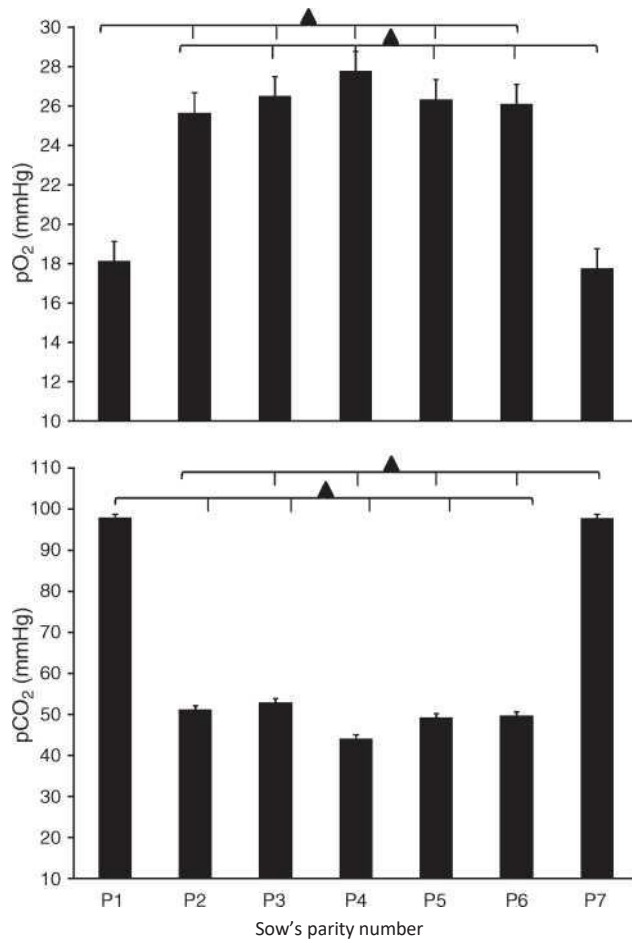


Fig. 1. Assessment of gas exchange in neonate piglets due to the effect of the sow's parity number (mean ± s.e.). Differences among groups within the blood variable (▲; $P < 0.05$; ANOVA followed by Tukey's *post hoc* test). (a) Partial oxygen pressure. (b) Partial carbon dioxide pressure.

