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## Monograph

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# Revision of the poorly known Neotropical butterfly genus *Zischkaia* Forster, 1964 (Lepidoptera, Nymphalidae, Satyrinae), with descriptions of nine new species

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**Abstract.** We provide the first comprehensive taxonomic revision of the poorly known South American butterfly genus *Zischkaia* Forster, 1964, hitherto regarded as including three described species. A phylogenetic analysis based on DNA sequence data shows that *Zischkaia* is monophyletic and consists of two morphologically diagnosable clades. Morphological characters and DNA ‘barcodes’ support the recognition of twelve species in the genus, a significant increase even for the relatively poorly studied subtribe Euptychiina. Consequently, nine new species are described and named herein, including *Z. arctoa* Nakahara, sp. nov., *Z. chullachaki* Nakahara & Zacca, sp. nov., *Z. baku* Zacca, Dolibaina & Dias, sp. nov., *Z. arenisca* Nakahara, Willmott & Hall, sp. nov., *Z. argyrosflecha* Nakahara, L. Miller & Huertas, sp. nov., *Z. abanico* Nakahara & Petit, sp. nov., *Z. josti* Nakahara & Kleckner, sp. nov., *Z. mielkeorum* Dolibaina, Dias & Zacca, sp. nov. and *Z. warreni* Dias, Zacca & Dolibaina, sp. nov. In addition, a neotype is designated for *Satyrus pacarus* Godart, [1824], and lectotypes are designated for *Euptychia amalda* Weymer, 1911, *Euptychia fumata* Butler, 1867 and *Euptychia saundersii* Butler, 1867.

**Key words.** DNA “barcodes”, Euptychiina, monophyletic, Neotropical region, systematics, taxonomy.

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## Introduction

*Zischkaia* Forster, 1964 has hitherto been regarded as a relatively small genus of three species, placed in the satyrine subtribe Euptychiina Reuter (Lamas 2004; Freitas *et al.* 2018). Among the 33 euptychiine genera that he described in the same paper, Forster (1964) described *Zischkaia* and designated *Euptychia fumata* Butler, 1867 (the date was erroneously given as 1857 by Forster) as the type species, recognizing two other species in this genus, namely *Euptychia amalda* Weymer, 1911 and *Euptychia saundersii* Butler, 1867. Subsequently, Lamas (2004) also included *Satyrus pacarus* Godart, [1824] and *Euptychia mima* Butler, 1867, in addition to recognizing one undescribed species and placing *E. fumata* (the type species of the genus) as a junior subjective synonym of *Z. pacarus*. However, Lamas (2004) did not provide any justification for the placement of these taxa in *Zischkaia*, this decision presumably being based on their similar ventral wing patterns. *Euptychia mima* was recently transferred to *Nhambikuara* Freitas, Barbosa & Zacca, 2018 (Freitas *et al.* 2018). Members of *Zischkaia* have an entirely Cis-Andean distribution and all of them are medium-sized (forewing length roughly 20–30 mm), uniformly brown butterflies marked with relatively large ventral hindwing ocelli, which therefore appear to be typical, drab euptychiine butterflies. However, compared to many other common euptychiine genera, species of *Zischkaia* are poorly represented in almost all museum collections, hindering taxonomic study and resulting in the genus remaining poorly understood. Although a number of other euptychiine genera have been the focus of taxonomic and/or systematic studies after the checklist of Lamas (2004), especially in recent years (e.g., Barbosa *et al.* 2015; Zacca *et al.* 2017; Nakahara *et al.* 2018c), partly as a result of the lack of material deposited in collections, *Zischkaia* has received no attention by butterfly researchers until now. A taxonomic revision for this group is therefore in urgent need, given the similarities among

the species of *Zischkaia* and consequent difficulties in identification, and the potential significance of the group for biogeographic and conservation research as a result of the highly restricted geographic ranges of some species.

This paper is part of a planned series of publications by some of the authors (SN, TZ and KW) to revise a number of taxonomically problematic genera in the nymphalid subtribe Euptychiina. The past few years have seen an explosion of interest in euptychiine systematics (e.g., Cong & Grishin 2014; Barbosa *et al.* 2015; Zacca *et al.* 2017, 2018; Nakahara *et al.* 2015a, 2015b, 2018b, 2018c, 2019; Willmott *et al.* 2019; see also <http://www.flmnh.ufl.edu/museum-voices/euptychiina/>), where the need for generic revisions and revisionary systematic study in this subtribe has been discussed in several publications (e.g., Marín *et al.* 2011). This study towards revising the genus *Zischkaia* was originally initiated by FD and DD, although this plan was halted due to the inaccessibility of the types of some taxa. Recently, SN initiated a complete revision of the genus, partly to contribute towards a better overall understanding of the systematics and diversity of Euptychiina. This collaborative study has enabled us to satisfactorily review what is known about the genus, as a first step towards understanding the true species diversity of the group. As a result, we regard *Zischkaia* as containing twelve species, remarkably with nine of them described and named herein. A neotype is designated for *Satyrus pacarus* and lectotypes are designated for *Euptychia amalda*, *E. fumata* and *E. saundersii*.

## Material and methods

### Morphological study

Legs, labial palpi, and abdomens were soaked in hot KOH solution for 5–10 minutes prior to dissection, dissected, and subsequently stored in glycerine. Dissection information regarding genitalia of *Zischkaia* can be found in Table 1. Drawings of external morphology were done using a camera lucida attached to either Leica MZ 16 stereoscopic microscope (at MGCL) or Nikon SMZ2800 (at MUSM). Images of genitalia taken at MGCL were taken with an Auto-montage Pro 5.01 system (Synoptics Ltd.) using a JVC digital camera (model KY-F65U) and Leica Z16APO lens, except for female genitalia of *Z. arenisca* Nakahara, Willmott & Hall, sp. nov. which were photographed using Helicon Focus 6.7.1 using Canon EOS 6D, subsequently stacked using Helicon Remote (ver. 3.8.7 W); photos of male and female genitalia were obtained at DZUP in Leica LAS 3D view and LAS montage version 4.7 with the aid of a video camera Leica DFC 500 attached to a stereoscopic microscope Leica MZ16. Illustrations of venation were prepared with the aid of a camera lucida attached to a stereoscopic microscope and subsequently vectorized by using the software GIMP version 2.8.10 (GIMP team 2016) (for female *Z. pacarus* (Godart, 1824)) and CorelDRAW X5 (for *Z. saundersii* (Butler, 1867)), in addition to tracing line drawing by ink and subsequently modified using Adobe Photoshop CC 2017 (for male *Z. pacarus*). A JEOL JSM-5510LV Scanning Electron Microscope (SEM) was used to take images of androconial scales.

Terminology for wing venation follows the Comstock & Needham (1898) system described in Miller (1970: 46), and terminology for wing pattern elements follows Zacca *et al.* (2018). Nomenclature of genitalia mostly follows Klots (1956), although we also follow Muschamp (1915) for the term “brachia”; we follow Austin & Mielke (2008) in referring to the part of the genitalia typically termed the “vinculum” as the “combined ventral arms of tegumen and dorsal arms of saccus” (see Nakahara *et al.* 2015a, for more details); Nakahara *et al.* (2018b) is followed for those terms related to phallus. Finally, we apply the term ‘sclerotized tube’ to the sclerotized portion of ductus bursae, which appears somewhat as a continuation of the lamella antevaginalis, since this character is useful in distinguishing species in the “*pacarus* clade” and we hope that this term will permit clearer understanding of species diagnoses. The term antrum (*sensu* Klots 1956) might apply to this particular structure, but further discussion of terminology is beyond the scope of this article.

**Table 1** (continued on next page). A list of dissected specimens of *Zischkaia* Forster, 1964 for this study.

Taxon	Sex	Genitalia dissection ID	Voucher No.	Repository	Locality (Country)
<i>Z. amalda</i>	♂	SN-17-21	N/A (LT)	ZSM	Bolivia
<i>Z. amalda</i>	♂	SN-17-8	1036040	FLMNH/MGCL	Peru
<i>Z. arctoa</i>	♂	SN-17-9	1036041	FLMNH/MGCL	Venezuela
<i>Z. arctoa</i>	♂	SN-17-73	1036042	FLMNH/MGCL	Venezuela
<i>Z. chullachaki</i>	♂	SN-17-5	1036033	FLMNH/MGCL	Peru
<i>Z. chullachaki</i>	♂	SN-17-75	1036037	FLMNH/MGCL	Peru
<i>Z. chullachaki</i>	♂	SN-17-66	1036039	FLMNH/MGCL	Peru
<i>Z. chullachaki</i>	♂	N/A	21627	DZUP	Brazil
<i>Z. chullachaki</i>	♀	N/A	37003	DZUP	Brazil
<i>Z. chullachaki</i>	♀	N/A	36508	DZUP	Brazil
<i>Z. chullachaki</i>	♂	N/A	21620	DZUP	Brazil
<i>Z. chullachaki</i>	♂	N/A	21228	DZUP	Peru
<i>Z. baku</i>	♂	N/A	MGCL-LOAN 201	ZUEC	Brazil
<i>Z. baku</i>	♂	N/A	MGCL-LOAN 292	ZUEC	Brazil
<i>Z. baku</i>	♂	SN-17-10	1036015	FLMNH/MGCL	Brazil
<i>Z. baku</i>	♂	SN-17-64	1036018	FLMNH/MGCL	Brazil
<i>Z. baku</i>	♂	SN-17-63	1036022	FLMNH/MGCL	Brazil
<i>Z. baku</i>	♀	SN-17-65	1036029	FLMNH/MGCL	Brazil
<i>Z. baku</i>	♂	N/A	5573	DZUP	Brazil
<i>Z. baku</i>	♂	N/A	5572	DZUP	Brazil
<i>Z. baku</i>	♀	N/A	5571	DZUP	Brazil
<i>Z. baku</i>	♂	M-2239 J.Y. Miller	1036032	FLMNH/MGCL	Brazil
<i>Z. arenisca</i>	♂	SN-17-16	1036290	FLMNH/MGCL	Ecuador
<i>Z. arenisca</i>	♂	SN-17-74	1036291	FLMNH/MGCL	Ecuador
<i>Z. arenisca</i>	♀	SN-17-174	149623	FLMNH/MGCL	Ecuador
<i>Z. arenisca</i>	♀	SN-16-73	105658	MUSM	Peru
<i>Z. arenisca</i>	♀	SN-19-32	297321	FLMNH/MGCL	Ecuador
<i>Z. argyrosflecha</i>	♂	M-9142 Lee D. Miller	N/A	MNHU	Peru
<i>Z. argyrosflecha</i>	♂	N/A	1718076	NHMUK	Peru
<i>Z. pacarus</i>	♂	N/A	21207	DZUP	Brazil
<i>Z. pacarus</i>	♂	N/A	5578	DZUP	Brazil
<i>Z. pacarus</i>	♂	N/A	36989	DZUP	Brazil
<i>Z. pacarus</i>	♀	N/A	36841	DZUP	Brazil
<i>Z. pacarus</i>	♀	N/A	5577	DZUP	Brazil
<i>Z. pacarus</i>	♀	N/A	36881	DZUP	Brazil
<i>Z. pacarus</i>	♂	SN-17-6	1036004	FLMNH/MGCL	Brazil
<i>Z. pacarus</i>	♀	SN-17-72	1036005	FLMNH/MGCL	Brazil
<i>Z. abanico</i>	♂	prep. genit. 575 19.07.2017/J.Lorenc	11926	MZUJ	Ecuador
<i>Z. abanico</i>	♂	N/A	10430711	NHMUK	Colombia
<i>Z. saundersii</i>	♀	N/A	36969	DZUP	Peru

**Table 1** (continued).

Taxon	Sex	Genitalia dissection ID	Voucher No.	Repository	Locality (Country)
<i>Z. saundersii</i>	♂	N/A	21221	DZUP	Peru
<i>Z. saundersii</i>	♀	N/A	5580	DZUP	Peru
<i>Z. saundersii</i>	♂	N/A	5579	DZUP	Peru
<i>Z. saundersii</i>	♂	N/A	38159	DZUP	Brazil
<i>Z. josti</i>	♀	N/A	N/A (HT)	NMBE	Venezuela
<i>Z. josti</i>	♀	2018-002 D.J.Harvey	233844	USNM	Guyana
<i>Z. mielkeorum</i>	♂	N/A	36599	DZUP	Brazil
<i>Z. mielkeorum</i>	♂	N/A	21200	DZUP	Brazil
<i>Z. mielkeorum</i>	♀	N/A	36688	DZUP	Brazil
<i>Z. mielkeorum</i>	♀	N/A	36711	DZUP	Brazil
<i>Z. warreni</i>	♂	N/A	21214	DZUP	Brazil
<i>Z. warreni</i>	♀	N/A	36731	DZUP	Brazil
<i>Z. warreni</i>	♀	N/A	36669	DZUP	Brazil
<i>Z. warreni</i>	♀	N/A	36689	DZUP	Brazil
<i>Z. warreni</i>	♂	SN-17-7	1036043	FLMNH/MGCL	Brazil
<i>Z. warreni</i>	♀	SN-17-172	1036044	FLMNH/MGCL	Brazil

A total of 384 specimens were studied and the following 22 collection acronyms are used throughout this article.

#### Institutional acronyms

- ADW = Andrew David Warren collection, Castle Rock, USA  
 AMNH = American Museum of Natural History, New York, USA  
 BEJO = Bernhard Jost collection, Münsingen, Switzerland  
 DATR = David Trembath collection, Dorking, UK  
 DD = Diego Rodrigo Dolibaina collection, Curitiba, Brazil  
 DZUP = Coleção Pe. Jesus Santiago Moure, Departamento de Zoologia, Universidade Federal do Paraná, Curitiba, Brazil  
 FLMNH = McGuire Center for Lepidoptera and Biodiversity (MGCL), Florida Museum of Natural History, Gainesville, USA  
 HAWA = Haydon Warren-Gash collection, London, UK  
 INABIO = Instituto Nacional de Biodiversidad, Quito, Ecuador (formerly MECN)  
 JEPE = Jean-Claude Petit collection, Ducy, France  
 MECN = Museo Ecuatoriano de Ciencias Naturales, Quito, Ecuador  
 MNHN = Muséum national d'Histoire naturelle, Paris, France  
 MNHU = Museum für Naturkunde, Leibniz-Institut für Evolutions- und Biodiversitätsforschung an der Humboldt Universität, Berlin, Germany  
 MUSM = Museo de Historia Natural, Universidad Nacional Mayor de San Marcos, Lima, Peru  
 MZSP = Museu de Zoologia, Universidade de São Paulo, São Paulo, Brazil  
 MZUJ = Zoological Museum, Jagellonian University, Krakow, Poland.  
 NHMUK = Natural History Museum, London, UK (formerly BMNH)  
 NMBE = Naturhistorisches Museum der Bürgergemeinde Bern, Bern, Switzerland  
 RFC = Romero Family collection, Maracay, Venezuela  
 USNM = National Museum of Natural History, Smithsonian Institution, Washington, DC, USA  
 ZSM = Zoologische Staatssammlung München, München, Germany  
 ZUEC = Museu de Zoologia da Universidade Estadual de Campinas, Campinas, Brazil

**Morphological abbreviations:**

DFW	=	Dorsal forewing
DHW	=	Dorsal hindwing
LT	=	Lectotype
PLT	=	Paralectotype
PT	=	Paratype
VFW	=	Ventral forewing
VHW	=	Ventral hindwing

Appropriate type labels will be added to all types designated here.

**Molecular study**

DNA extraction methods, PCR conditions and primers used for amplification of cytochrome c oxidase I (COI) largely follow Nakahara *et al.* (2018a), except for several pairs of primers designed to amplify smaller fragments of COI; information on all primers used in this study is provided in Table 2. Sequences generated through this study were uploaded to GenBank and sequence voucher information is provided in Table 3. The dataset (646 bp), including 28 ingroup taxa representing eight species and five outgroup taxa (see Table 3), was aligned using MUSCLE (ver. 3.8.425) (Edgar 2004) in Geneious ver. 11.1.5 (Biomatters Ltd.), the best-fit model of nucleotide evolution (TIM2+F+I+G4) was obtained using ModelFinder (Kalyaanamoorthy *et al.* 2017), and the likelihood tree with the highest log-likelihood score given the aforementioned dataset was inferred by conducting Nearest Neighbor Interchange (NNI) search strategy based on the twenty best initial trees in IQ-TREE ver. 1.6.9 (Nguyen *et al.* 2015). The tree was rooted with *Pindis squamistriga* R. Felder, 1869 based on the much larger dataset including >2000 euptychiine individuals representing >420 euptychiine species (unpubl. data). Branch support was calculated using ultrafast bootstrap (UFBoot) with 1000 replications (Hoang *et al.* 2018), in addition to assessments of node support through 1000 replications of the Shimodaira Hasegawa approximate Likelihood Ratio Test (SH-aLRT) (Guindon *et al.* 2010). The genetic distances among species of *Zischkaia* presented in Table 4 were calculated by the Tamura-Nei distance model based on the aforementioned COI alignment in Geneious ver. 11.1.5 (Biomatters Ltd.).

**Databasing and georeferencing**

Label data of examined specimens were entered into a Microsoft Access database, which was also used to generate data for the ‘Examined specimens’ sections. If coordinate data were not provided on the label, we georeferenced the locality information and coordinates are provided in square brackets in the relevant section. For historical type specimens, information on the labels was written verbatim and information for each label is separated by double forward slashes ‘//’; if additional information is found on the other side of the label, a single forward slash ‘/’ is used to reflect this information. Square brackets ‘[]’ are used to enclose supplementary or inferred information.

**Table 2.** Information regarding COI primers used in this study.

Sequence 5'-3'	Primer name	Annealing temp. (°C)	Direction	References
CCAGGATWTTTAATTGGDGAATGA	COI_bc_EuF2	51	forward	modified based on Strutzenberger <i>et al.</i> (2012)
GGATTTGGWAATTGATTARTYCC	COI_bc_EuF3	51	forward	modified based on Strutzenberger <i>et al.</i> (2012)
AGTATYGTAGAAAATGGAGCTGG	COI_bc_EuF4	56	forward	modified based on Strutzenberger <i>et al.</i> (2012)
TTTGAGCTGTHGGAATTACAGC	COI_bc_EuF6	56	forward	modified based on Strutzenberger <i>et al.</i> (2012)
TATTATTATACGVGGRAAAGCTA	COI_bc_EuR2X	51	reverse	modified based on Strutzenberger <i>et al.</i> (2012)
GTAATAGCTCCRGCTAAAACAG	COI_bc_EuR5X	51	reverse	modified based on Strutzenberger <i>et al.</i> (2012)
AAAAATTATAATAAAAAGCATGRGC	COI_bc_TegR1	51	reverse	modified based on Strutzenberger <i>et al.</i> (2012)
ATTGTRGTAATAAAAATTAAATAGCTCC	COI_bc_TegR4X	56	reverse	modified based on Strutzenberger <i>et al.</i> (2012)
TAAACTTCAGGATGACCAAAAA	HCO_nym	56 (45 for 1 and 2 part)	reverse	Nakahara <i>et al.</i> (2015b)
WGGGGGTAAACTGTTCATCC	K699	56 (45 for 2 part)	reverse	Elias <i>et al.</i> (2007)
TTTCTACAAATCATAAAGATAATTGG	LCO_nym	56 (45 for 1 and 2 part)	forward	Nakahara <i>et al.</i> (2015b)
CCTGGTAAAATTAAAAATATAAACTTC	Nancy	45	reverse	Monteiro & Pierce (2001)
GGATCACCTGATATAGCAITCCC	Ron	45	forward	Monteiro & Pierce (2001)



**Table 3.** GenBank accession numbers for specimens used for molecular analysis in this study.

Voucher code	Genus	Species	GenBank Acc. No.
LEP-68763	<i>Zischkaia</i>	<i>josti</i>	MK880122
BC-DZ-227	<i>Zischkaia</i>	<i>warreni</i>	MK880099
BC-DZ-231	<i>Zischkaia</i>	<i>chullachaki</i>	MK880097
BC-DZ-247	<i>Zischkaia</i>	<i>pacarus</i>	MK880098
LEP-37388	<i>Zischkaia</i>	<i>abanico</i>	MK880100
BC-DZ-226	<i>Zischkaia</i>	<i>warreni</i>	MK880101
BC-DZ-249	<i>Zischkaia</i>	<i>mielkeorum</i>	MK880102
CP14-02	<i>Zischkaia</i>	<i>pacarus</i>	GQ864819
BC-DZ-246	<i>Zischkaia</i>	<i>pacarus</i>	MK880103
MGCL-LOAN-266	<i>Zischkaia</i>	<i>chullachaki</i>	MK880109
LEP-64863	<i>Zischkaia</i>	<i>arenisca</i>	MK880110
LEP-64860	<i>Zischkaia</i>	<i>arenisca</i>	MK880111
LEP-64859	<i>Zischkaia</i>	<i>arenisca</i>	MK880112
LEP-64862	<i>Zischkaia</i>	<i>arenisca</i>	MK880113
LEP-64857	<i>Zischkaia</i>	<i>arenisca</i>	MK880114
LEP-64861	<i>Zischkaia</i>	<i>arenisca</i>	MK880115
LEP-64858	<i>Zischkaia</i>	<i>arenisca</i>	MK880116
LEP-64864	<i>Zischkaia</i>	<i>arenisca</i>	MK880116
MGCL-LOAN-518	<i>Zischkaia</i>	<i>pacarus</i>	MK880104
BC-DZ-229	<i>Zischkaia</i>	<i>chullachaki</i>	MK880105
BC-DZ-232	<i>Zischkaia</i>	<i>chullachaki</i>	MK880106
LEP-08939	<i>Zischkaia</i>	<i>arenophilia</i>	MK880107
BC-DZ-230	<i>Zischkaia</i>	<i>chullachaki</i>	MK880108
MGCL-LOAN-548	<i>Zischkaia</i>	<i>saundersii</i>	MK880118
MGCL-LOAN-541	<i>Zischkaia</i>	<i>saundersii</i>	MK880119
LEP-18704	<i>Zischkaia</i>	<i>josti</i>	MK880120
BC-DZ-225	<i>Zischkaia</i>	<i>mielkeorum</i>	MK880123
BC-DZ-248	<i>Zischkaia</i>	<i>mielkeorum</i>	MK880122
NW165-5	<i>Pindis</i>	<i>squamistriga</i>	GQ357211
CP01-23	<i>Amphidecta</i>	<i>clio</i>	DQ338810
CP02-48	<i>Splendeptychia</i>	<i>boliviensis</i>	GU205866
CP02-44	<i>Splendeptychia</i>	<i>itonis</i>	DQ338811
CP12-06	<i>Pharneuptychia</i>	<i>innocentia</i>	DQ338808

**Table 4** (continued on next page). Genetic distances calculated based on COI using the Tamura-Nei distance model.

LEP-68763	<i>Zischkaia josti</i> _Bolivar_Venezuela	0.086	0.089	0.097	0.083	0.086	0.011	0.092	0.084	0.072	0.074	0.073	0.073	0.072	0.077	0.074	0.072	0.073	0.083	0.085	0.083	0.073	0.081	0.062	0.062	0.003	0.011	0.01	
BC-DZ_Willmott-227	<i>Zischkaia warreni</i> _Brazil_Parana	0.086	0.089	0.097	0.083	0.086	0.011	0.092	0.084	0.072	0.074	0.073	0.073	0.072	0.077	0.074	0.072	0.073	0.083	0.085	0.083	0.073	0.081	0.062	0.062	0.003	0.011	0.01	
BC-DZ_Willmott-231	<i>Zischkaia chullachaki</i> _Brazil_Acre	0.089	0.11	0.11	0.122	0	0.086	0.119	0.114	0.104	0.113	0.11	0.111	0.11	0.116	0.11	0.11	0.11	0.11	0.108	0.111	0.108	0.111	0.09	0.092	0.079	0.095	0.092	
BC-DZ_Willmott-247	<i>Zischkaia pacarus</i> _Brazil_Parana	0.097	0.11	0.036	0.071	0.106	0.093	0.01	0.002	0.042	0.039	0.036	0.036	0.036	0.042	0.036	0.036	0.01	0.038	0.038	0.035	0.038	0.101	0.103	0.101	0.1	0.097	0.097	
LEP-37388	<i>Zischkaia abanico</i> _Ecuador_Morona-Santiago	0.083	0.122	0.067	0.071	0.101	0.098	0.052	0.046	0.039	0.047	0.047	0.047	0.046	0.05	0.046	0.044	0.046	0.05	0.052	0.05	0.045	0.047	0.093	0.093	0.09	0.09	0.088	
BC-DZ_Willmott-226	<i>Zischkaia warreni</i> _Brazil_Santa_Catarina	0.086	0	0.109	0.106	0.101	0.088	0.098	0.096	0.086	0.096	0.093	0.093	0.093	0.097	0.093	0.093	0.093	0.093	0.094	0.091	0.091	0.091	0.091	0.076	0.078	0.082	0.095	0.093
BC-DZ_Willmott-249	<i>Zischkaia mielkeorum</i> _Brazil_Espirito_Santo	0.011	0.086	0.094	0.093	0.098	0.088	0.098	0.089	0.085	0.086	0.085	0.085	0.085	0.089	0.085	0.085	0.085	0.085	0.092	0.092	0.083	0.092	0.063	0.065	0.011	0.005	0.004	
CP14-02	<i>Zischkaia pacarus</i> _unknown	0.092	0.119	0.046	0.01	0.052	0.098	0.098	0.008	0.039	0.038	0.036	0.036	0.035	0.037	0.035	0.033	0.035	0.015	0.041	0.04	0.034	0.037	0.095	0.095	0.097	0.098	0.096	
BC-DZ_Willmott-246	<i>Zischkaia pacarus</i> _Brazil_Parana	0.084	0.114	0.039	0.002	0.046	0.096	0.089	0.008	0.035	0.033	0.031	0.031	0.031	0.031	0.029	0.031	0.01	0.037	0.035	0.03	0.033	0.092	0.092	0.093	0.088	0.088	0.088	
MGCL-LOAN-266	<i>Zischkaia chullachaki</i> _Brazil_Acre	0.072	0.104	0.005	0.042	0.039	0.086	0.085	0.039	0.035	0.01	0.01	0.01	0.01	0.01	0.013	0.01	0.008	0.01	0.032	0.005	0.008	0.008	0.002	0.074	0.074	0.08	0.081	0.079
LEP-64863	<i>Zischkaia arenisca</i> _Ecuador_Zamora-Chinchiipe	0.074	0.113	0.007	0.039	0.047	0.096	0.086	0.038	0.033	0.01	0.002	0.002	0.002	0.005	0.002	0.002	0.002	0.002	0.031	0.008	0.01	0.003	0.01	0.08	0.082	0.081	0.081	0.079
LEP-64860	<i>Zischkaia arenisca</i> _Ecuador_Zamora-Chinchiipe	0.073	0.11	0.005	0.036	0.047	0.093	0.085	0.036	0.031	0.01	0.002	0	0	0.003	0	0	0.029	0.008	0.01	0.002	0.008	0.077	0.079	0.08	0.08	0.08	0.078	
LEP-64859	<i>Zischkaia arenisca</i> _Ecuador_Zamora-Chinchiipe	0.073	0.111	0.005	0.036	0.047	0.093	0.085	0.036	0.031	0.01	0.002	0	0	0	0	0	0.029	0.008	0.01	0.002	0.008	0.079	0.079	0.08	0.08	0.08	0.078	
LEP-64862	<i>Zischkaia arenisca</i> _Ecuador_Zamora-Chinchiipe	0.072	0.11	0.005	0.036	0.046	0.093	0.085	0.035	0.031	0.01	0.002	0	0	0.003	0	0	0.029	0.008	0.01	0.002	0.008	0.078	0.078	0.079	0.081	0.08	0.08	

**Table 4** (continued). Genetic distances calculated based on COI using the Tamura-Nei distance model.

LEP-64857	<i>Zischkaia-arenisca</i> _Ecuador_Zamora-Chinchipe	0.077	0.116	0.01	0.042	0.05	0.097	0.089	0.037	0.031	0.013	0.005	0.003	0	0.003	0.003	0.003	0.028	0.008	0.01	0.002	0.008	0.08	0.08	0.082	0.081	0.083
LEP-64861	<i>Zischkaia-pacarus</i> _Ecuador_Zamora-Chinchipe	0.074	0.11	0.005	0.036	0.046	0.093	0.085	0.035	0.031	0.01	0.002	0	0	0.003	0	0	0.028	0.008	0.01	0.002	0.008	0.078	0.08	0.083	0.081	0.079
LEP-64858	<i>Zischkaia-arenisca</i> _Ecuador_Zamora-Chinchipe	0.072	0.11	0.005	0.036	0.044	0.093	0.085	0.033	0.029	0.008	0.002	0	0	0.003	0	0	0.027	0.008	0.008	0	0.006	0.078	0.078	0.08	0.081	0.079
LEP-64864	<i>Zischkaia-arenisca</i> _Ecuador_Zamora-Chinchipe	0.073	0.11	0.005	0.036	0.046	0.093	0.085	0.035	0.031	0.01	0.002	0	0	0.003	0	0	0.028	0.008	0.01	0.002	0.008	0.078	0.079	0.082	0.08	0.079
MGCL-LOAN-518	<i>Zischkaia_pacarus</i> _Brasil_Bahia	0.083	0.108	0.032	0.01	0.05	0.094	0.085	0.015	0.01	0.032	0.031	0.029	0.029	0.028	0.028	0.027	0.028	0.033	0.032	0.027	0.03	0.089	0.091	0.09	0.085	0.085
BC-DZ_Willmott-229	<i>Zischkaia_chullachaki</i> _Brasil_Acre	0.085	0.111	0	0.038	0.052	0.091	0.092	0.041	0.037	0.005	0.008	0.008	0.008	0.008	0.008	0.008	0.033	0.002	0.01	0.003	0.086	0.087	0.092	0.091	0.091	
BC-DZ_Willmott-232	<i>Zischkaia_chullachaki</i> _Brasil_Acre	0.083	0.111	0	0.038	0.05	0.091	0.092	0.04	0.035	0.003	0.01	0.01	0.01	0.01	0.01	0.01	0.008	0.008	0.002	0.002	0.006	0.079	0.079	0.08	0.081	
LEP-08939	<i>Zischkaia_arenisca</i> _Ecuador_Zamora-Chinchipe	0.073	0.108	0.003	0.035	0.045	0.091	0.083	0.034	0.03	0.008	0.003	0.002	0.002	0.002	0	0.002	0.027	0.01	0.008	0.006	0.079	0.079	0.08	0.08	0.081	
BC-DZ_Willmott-230	<i>Zischkaia_chullachaki</i> _Brasil_Acre	0.081	0.111	0	0.038	0.047	0.091	0.092	0.037	0.033	0.002	0.01	0.008	0.008	0.008	0.008	0.008	0.03	0.003	0.003	0.006	0.083	0.083	0.088	0.088	0.089	
MGCL-LOAN-548	<i>Zischkaia_saundersii</i> _Brasil_Rondonia	0.062	0.09	0.095	0.101	0.093	0.076	0.063	0.095	0.092	0.074	0.08	0.077	0.079	0.078	0.08	0.078	0.078	0.078	0.089	0.086	0.084	0.079	0.083	0.002	0.068	0.062
MGCL-LOAN-541	<i>Zischkaia_saundersii</i> _Brasil_Rondonia	0.062	0.092	0.098	0.103	0.093	0.078	0.065	0.095	0.092	0.074	0.082	0.079	0.079	0.078	0.08	0.08	0.078	0.079	0.091	0.087	0.086	0.079	0.083	0.002	0.068	0.062
LEP-18704	<i>Zischkaia_josti</i> _Guyana_Upper_Takutu-Upper_Essequibo	0.003	0.079	0.092	0.101	0.09	0.082	0.011	0.097	0.093	0.08	0.081	0.08	0.079	0.079	0.082	0.083	0.08	0.082	0.09	0.092	0.091	0.08	0.088	0.068	0.016	0.016
BC-DZ_Willmott-225	<i>Zischkaia_mielkeorum</i> _Brasil_Minus_Gerais	0.011	0.095	0.097	0.1	0.09	0.095	0.005	0.098	0.088	0.081	0.081	0.08	0.08	0.081	0.081	0.081	0.081	0.081	0.089	0.091	0.089	0.08	0.088	0.062	0.062	0.016
BC-DZ_Willmott-248	<i>Zischkaia_mielkeorum</i> _Brasil_Espirito_Santo	0.01	0.092	0.095	0.097	0.088	0.093	0.004	0.096	0.088	0.079	0.079	0.078	0.08	0.083	0.079	0.079	0.079	0.079	0.085	0.091	0.089	0.081	0.089	0.062	0.062	0.016

## Results

Order Lepidoptera Linnaeus, 1758  
 Superfamily Papilionoidea Latreille, [1802]  
 Family Nymphalidae Rafinesque, 1815  
 Subfamily Satyrinae Boisduval, 1833  
 Tribe Satyrini Boisduval, 1833  
 Subtribe Euptychiina Reuter, 1896

Genus *Zischkaia* Forster, 1964

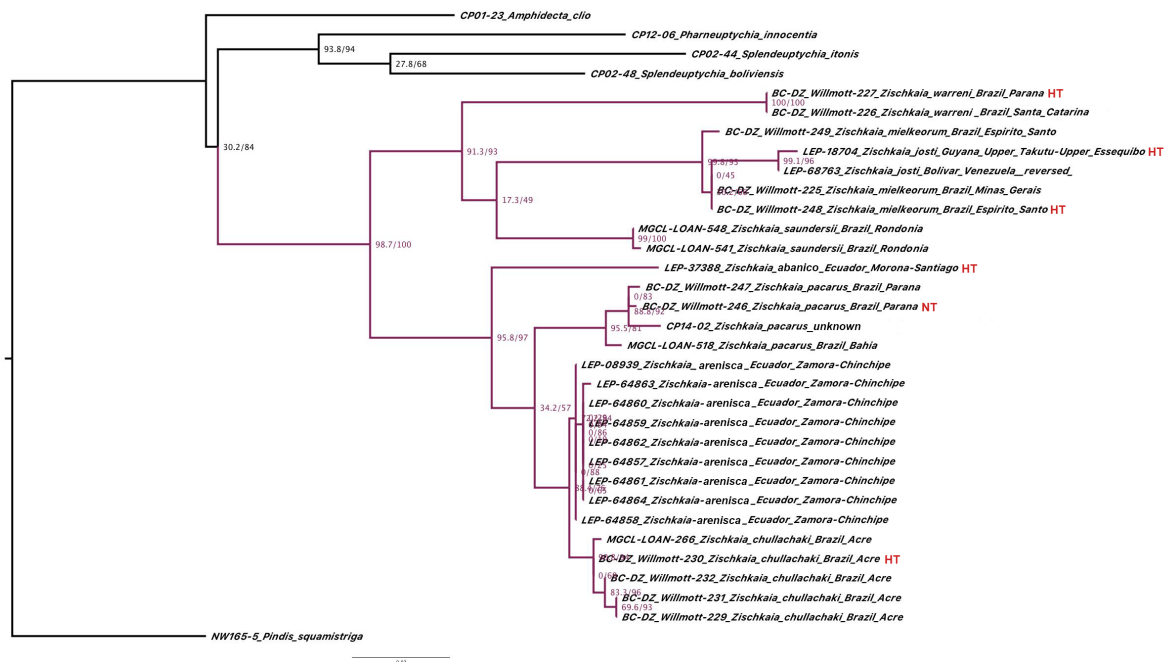
*Zischkaia* Forster, 1964: 116.

### Type species

*Euptychia fumata* Butler, 1867: 109, pl. 12, fig. 14 (by original designation).

### Systematics

*Zischkaia* is a member of a clade consisting of several species of *Splendeuptychia* Forster, 1964 (e.g., *S. itonis* (Hewitson, 1862), *S. clementia* (Butler, 1877)), *Amphidecta* Butler, 1867 and *Rareuptychia* Forster, 1964, based on the aforementioned large dataset including >2000 individuals representing > 420 species (unpubl. data). Although *Zischkaia* was not included in the analysis of Espeland *et al.* (2019), the clade containing these related species (the “*Amphidecta* clade”) was well supported based on hybrid enrichment data incorporating 368 loci (Espeland *et al.* 2019). The “*Amphidecta* clade” is sister to a large clade including some major euptychiine clades, such as the “*Pareuptychia* clade”, “*Taygetis* clade” and “*Splendeuptychia* clade” (all *sensu* Peña *et al.* 2010), but the support for this relationship is low. Our molecular data based on the COI barcoding region strongly support *Zischkaia* itself as monophyletic (Fig. 1; SH-aLRT/UFBoot = 98.7/100), with the removal by Freitas *et al.* (2018)



**Fig. 1.** Likelihood tree with the best log-likelihood score ( $-\ln L = -2536.1613$ ) given our data based on DNA ‘barcodes’, depicting *Zischkaia* as a well-supported clade. Support values are represented by SH-aLRT/UF Bootstrap. Abbreviations: HT = Holotype; NT = Neotype.

of *Euptychia mima* Butler, 1867 to *Nhambikuara*, a genus in the “*Pareuptychia* clade”. *Zischkaia* can be divided into two moderate to well-supported clades based on our molecular data (Fig. 1; SH-aLRT/UFBoot = 95.8/97; 91.3/93), which are also diagnosable by several morphological characters discussed below under the ‘Diagnosis’ section. Despite the morphological differences between the two clades, which are also highly supported based on molecular data, the overall wing patterns and habitats appear to be rather similar among species in both of these clades. Considering that generic names should ideally be informative for the recognition of monophyletic groups of similar appearing species, in addition to having potential value in predicting unknown biological traits, we here treat *Zischkaia* as representing both clades rather than treating them as distinct genera. The morphological diversity within *Zischkaia*, such as the presence or absence of wing androconial scales, is also seen within other euptychiine genera of similar species diversity, such as *Taygetina* Forster, 1964.

### Diagnosis

Species of *Zischkaia* can be distinguished from all other genera of Euptychiina by the combination of the following characters: 1) absence of ocelli on the DFW, DHW and VFW (also true of some other euptychiines); 2) five or six ocelli on the VHW from Sc+R<sub>1</sub> or M<sub>1</sub> to 2A with the white pupils (often diffuse scales rather than a single spot) displaced distally from the center (*Splendeuptychia doxes* (Godart, [1824]) and relatives possess a superficially similar pupil, but it is displaced basally instead of distally); 3) VFW with submedian line restricted to discal cell or absent (also true of a number of other euptychiine genera); 4) inter-segmental membrane between seventh and eighth sternites not pleated, but folded posteriorly of ostium bursae with its sclerotized region forming a ‘scoop-like’ structure below lamella antevaginalis (projection more apparent in species of the “*pacarus* clade”). This form of inter-segmental membrane is rather unique among euptychiines, which often have this membrane pleated and expandable, although some species in the genus *Euptychia* Hübner, 1818 are somewhat similar in this respect.

Two clades are recognized in *Zischkaia*, which we call the “*pacarus* clade” and the “*saundersii* clade”; the former can be distinguished from the latter by: 1) presence of androconial scales on DFW of males (absent in “*saundersii* clade”) (Figs 2–3); 2) presence of a ‘tusk’-like projection (Fig. 4A) from the posterior region of the tegumen above the uncus (the projection of the tegumen appears as a ‘bulb’ in the “*saundersii* clade” (Fig. 5A)); 3) uncus that is narrow and long, somewhat curving down, terminating in a small ‘bulb’ (rather than being straight and broad as in the “*saundersii* clade”); 4) brachia curved dorsally (rather than straight as in the “*saundersii* clade”); 5) valva being rather short in lateral view, with the apical point not extending beyond that of the uncus (the apical point extends beyond that of the uncus in the “*saundersii* clade”); 6) tip of the anterior projection of the saccus extending further than the tegumen in lateral view (the tip of the anterior projection of the saccus does not extend beyond the tegumen in lateral view in the “*saundersii* clade”); 7) fultura inferior (i.e., juxta) appearing as a thin strip in posterior view (Fig. 8A) (it appears as a well-developed plate in posterior view in the “*saundersii* clade” (Fig. 8B)); 8) median region of the ductus bursae with a well-developed sclerotized half-ring (Fig. 6A) (absent in the “*saundersii* clade” (Fig. 7A), or very reduced as in *Z. saundersii* and *Z. josti* sp. nov.).

The diagnostic characters provided above for the “*pacarus* clade” can be used to separate all species in this clade from species in the “*saundersii* clade”, and vice-versa. Therefore, species diagnoses focus on comparing similar species within the respective clade. Note that females of *Z. amalda* (Weymer, 1911), *Z. argyrosflecha* Nakahara, L. Miller & Huertas, sp. nov., *Z. abanico* Nakahara & Petit, sp. nov. and *Z. arctoa* Nakahara, sp. nov. are still unknown or unrecognized, thus the diagnostic characters provided for females of related species might not be applicable to these four species, and discovery of female specimens of these taxa would be extremely valuable. Similarly, the male is unknown or unrecognized for *Z. josti* Nakahara & Kleckner, sp. nov.



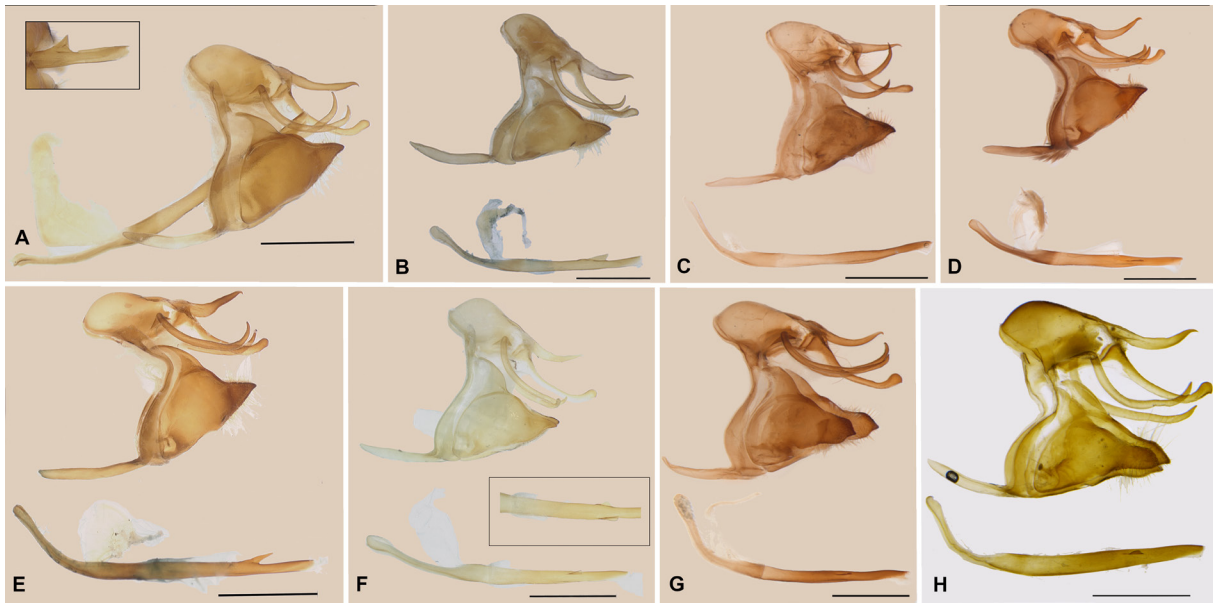
**Fig. 2.** *Zischkaia* “*pacarus* clade”, wing plate. **A–B.** *Z. amalda* (Weymer, 1911), lectotype, ♂, dorsal and ventral views. **C–D.** *Z. arctoa* Nakahara, sp. nov., paratype, ♂ (in RFC, collected in 1975), dorsal and ventral views. **E–F.** *Z. chullachaki* Nakahara & Zacca, sp. nov., holotype, ♂, dorsal and ventral views. **G–H.** *Z. chullachaki* sp. nov., paratype, ♀ (DZ 36. 508), dorsal and ventral views. **I–J.** *Z. baku* Zacca, Dolibaina & Dias, sp. nov., holotype, ♂, dorsal and ventral views. **K–L.** *Z. baku* sp. nov., paratype, ♀ (DZ 36. 404), dorsal and ventral views. **M–N.** *Z. arenisca* Nakahara, Willmott & Hall, sp. nov., holotype, ♂, dorsal and ventral views. **O–P.** *Z. arenisca* sp. nov., paratype, ♀ (FLMNH-MGCL 297320), dorsal and ventral views. **Q–R.** *Z. argyrosflecha* Nakahara, L. Miller & Huertas, sp. nov., holotype, ♂, dorsal and ventral views. **S–T.** *Z. pacarus* (Godart, [1824]), neotype, ♂, dorsal and ventral views. **U–V.** *Z. pacarus*, ♀ (DZ 36. 909), dorsal and ventral views. **W–X.** *Z. abanico* Nakahara & Petit, sp. nov., holotype, ♂, dorsal and ventral views. Scale bar: 1 cm.

### History of classification

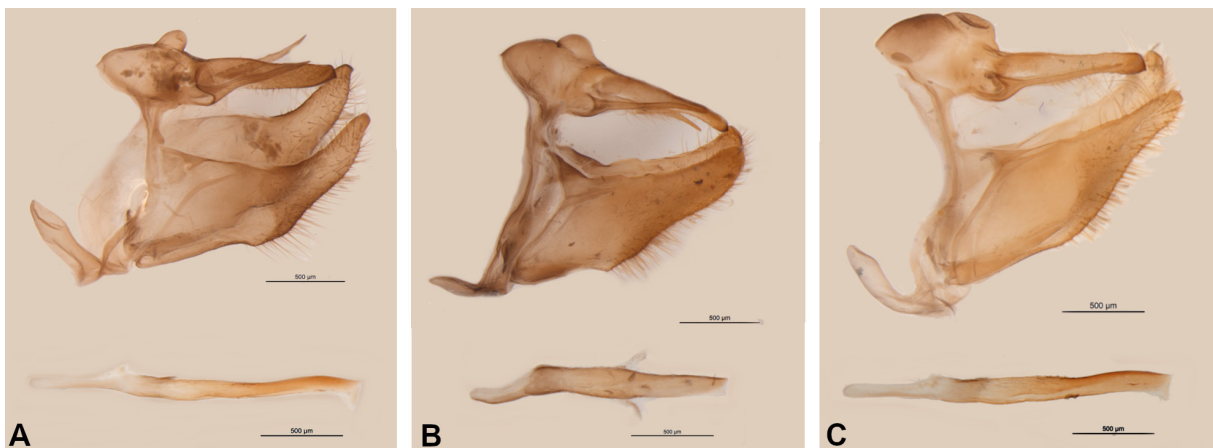
Butler (1867a) was the first to propose a systematic classification for *Zischkaia*. In his monograph of *Euptychia* (then used as a catch-all genus to include most euptychiine species), Butler divided the genus into seven groups (Division I to VII), and Division VI was the group relevant to *Zischkaia*. This division included *E. saundersii* Butler, 1867, *E. mima* Butler, 1867, *E. fumata* nomen nudum, *E. pacarus* (Godart, [1824]) and *E. insignis* Butler, 1867. The following diagnosis was given for Butler’s Division VI: “Upper surfaces brown, without marks; ventral forewings generally unmarked; [ventral] hindwings frequently with oval-shaped black ocelli, pupilled with silvery spots”. Subsequently, Butler (1877) proposed the “*E. pacarus* group” and included *E. saundersii*, *E. mima*, *E. fumata* Butler, 1867, *E. pacarus*, *E. insignis*,



**Fig. 3.** *Zischkaia* “*saundersii* clade”, wing plate. **A–B.** *Z. saundersii* (Butler, 1867), ♂ (DZ 37. 029), dorsal and ventral views. **C–D.** *Z. saundersii*, ♀ (DZ 36. 969), dorsal and ventral views. **E–F.** *Z. josti* Nakahara & Kleckner, sp. nov., holotype, ♀, dorsal and ventral views. **G–H.** *Z. josti* sp. nov., paratype, ♀ (USNM ENT 00233844), dorsal and ventral views. **I–J.** *Z. mielkeorum* Dolibaina, Dias & Zacca, sp. nov., holotype, ♂, dorsal and ventral views. **K–L.** *Z. mielkeorum* sp. nov., paratype, ♀ (DZ 36. 618), dorsal and ventral views. **M–N.** *Z. warreni* Dias, Zacca & Dolibaina, sp. nov., holotype, ♂, dorsal and ventral views. **O–P.** *Z. warreni* sp. nov., paratype, ♀ (DZ 36. 508), dorsal and ventral views. Scale bar: 1 cm.

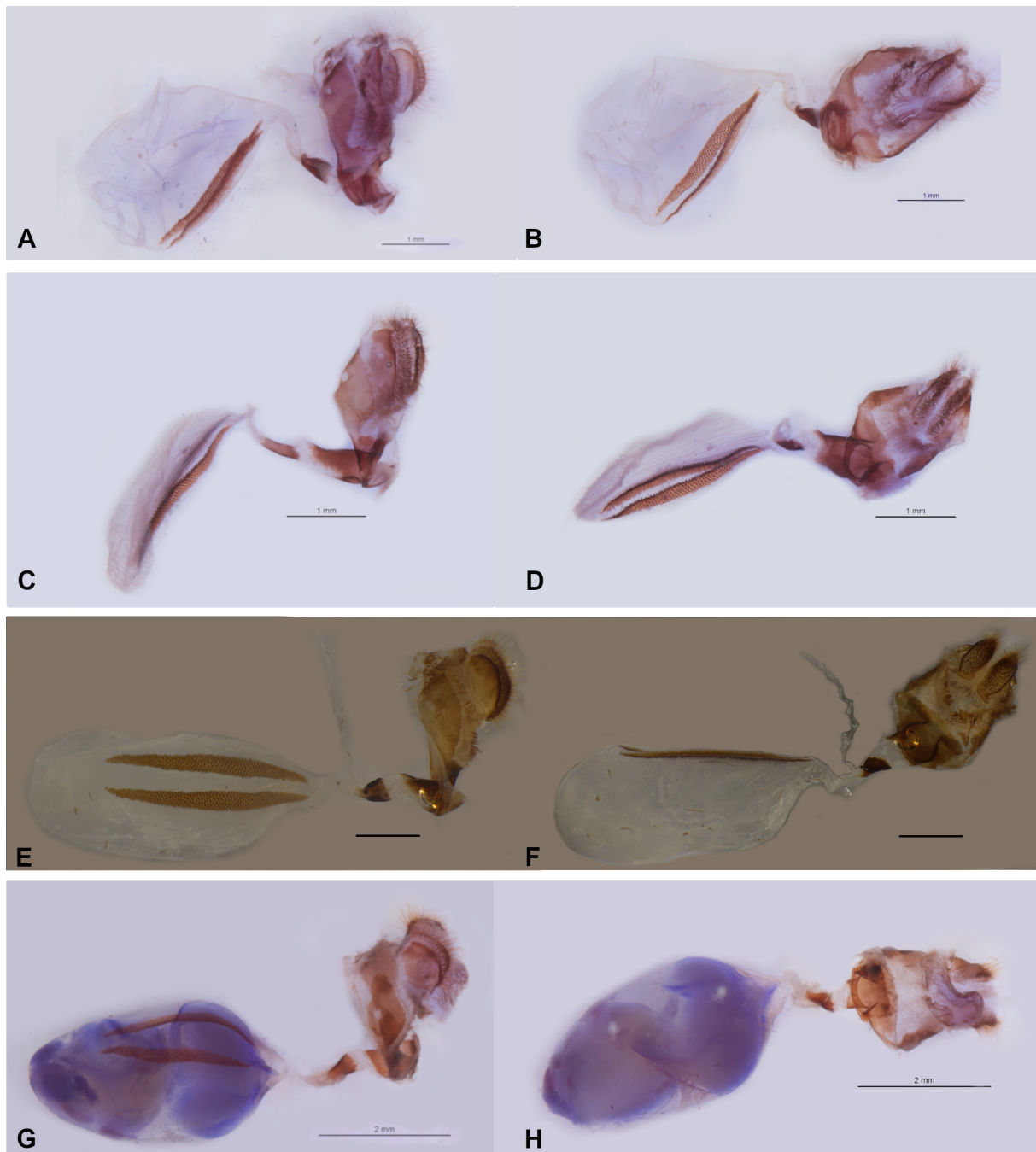


**Fig. 4.** *Zischkaia* “*pacarus* clade”, male genitalia. **A.** *Z. amalda* (Weymer, 1911), lectotype, ♂, lateral view, with phallus separated (winglet on aedeagus shown inside rectangle) (Genitalic vial: SN-17-21). **B.** *Z. arctoa* Nakahara, sp. nov., ♂, lateral view, with phallus separated (Genitalic vial: SN-17-9). **C.** *Z. chullachaki* Nakahara & Zacca, sp. nov., ♂, lateral view, with phallus separated (Genitalic vial: DZ 21. 235). **D.** *Z. baku* Zacca, Dolibaina & Dias, sp. nov., ♂, lateral view, with phallus separated (Genitalic vial: DZ 5. 573). **E.** *Z. arenisca* Nakahara, Willmott & Hall, sp. nov., ♂, lateral view, with phallus separated (Genitalic vial: SN-17-16). **F.** *Z. argyrosflecha* Nakahara, L. Miller & Huertas, sp. nov. ♂, lateral view, with phallus separated (two winglets on aedeagus shown inside rectangle) (Genitalic vial: M-9142 ♂, Lee D. Miller). **G.** *Z. pacarus* (Godart, [1824]), ♂, lateral view, with phallus separated (Genitalic vial: DZ 5. 578). **H.** *Z. abanico* Nakahara & Petit, sp. nov., ♂, lateral view, with phallus separated (Genitalic vial: 575/19.07.2017, J. Lorenc-Brudecka, MZUJ).

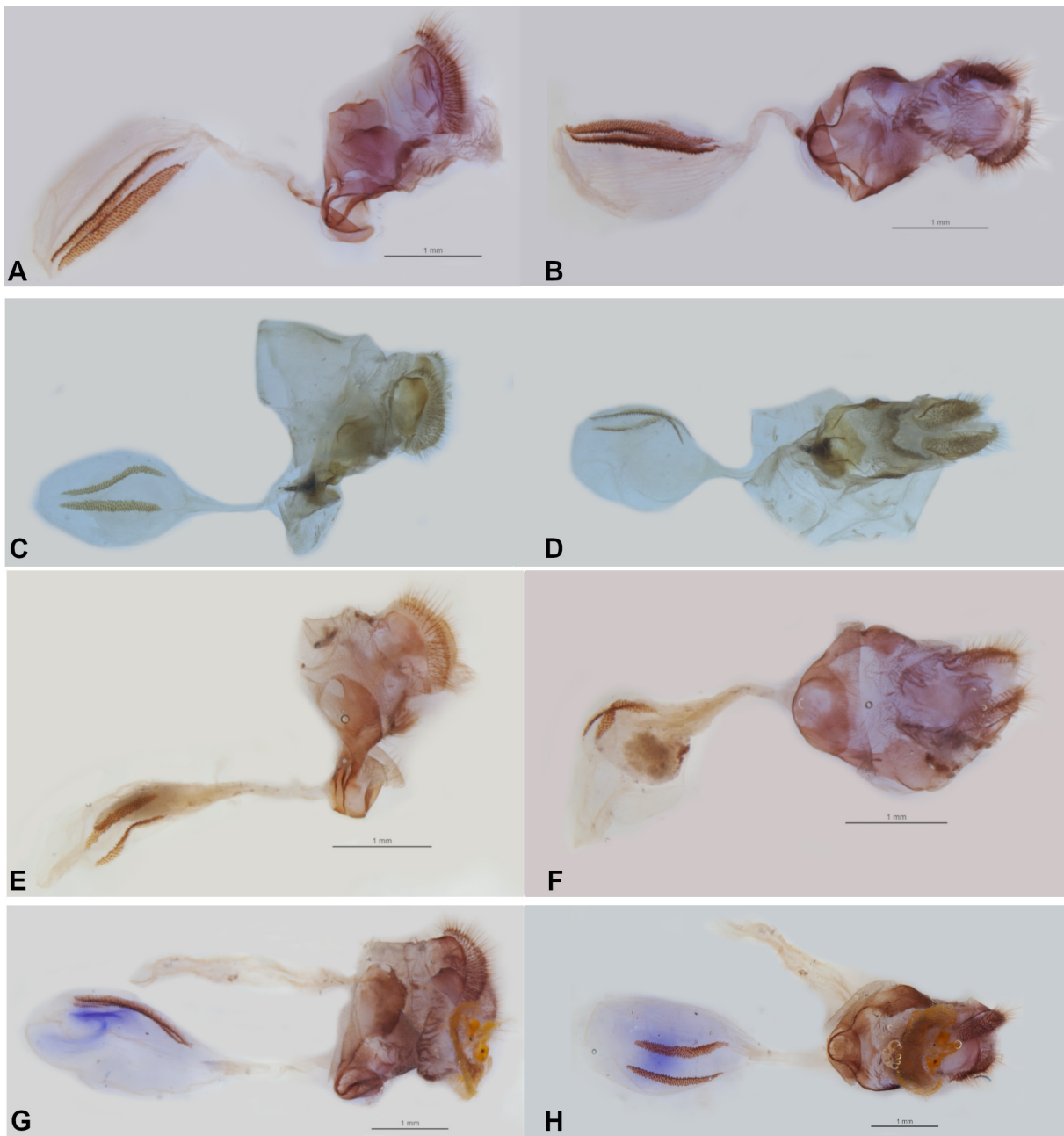


**Fig. 5.** *Zischkaia* “*saundersii* clade”, male genitalia. **A.** *Z. saundersii* (Butler, 1867), ♂, lateral view, with phallus separated (Genitalic vial: DZ 21. 221). **B.** *Z. mielkeorum* Dolibaina, Dias & Zacca, sp. nov., ♂, lateral view, with phallus separated (Genitalic vial: DZ 21. 200). **C.** *Z. warreni* Dias, Zacca & Dolibaina, sp. nov., ♂, lateral view, with phallus separated (Genitalic vial DZ 21. 214).





**Fig. 6.** *Zischkaia* “*pacarus* clade”, female genitalia. **A–B.** *Z. chullachaki* Nakahara & Zacca, sp. nov., ♀, lateral and ventral views (Genitalic vial DZ 36. 508). **C–D.** *Z. baku* Zacca, Dolibaina & Dias, sp. nov., ♀, lateral and ventral views (Genitalic vial: DZ 5. 571). **E–F.** *Z. arenisca* Nakahara, Willmott & Hall, sp. nov., ♀, lateral and ventral views (Genitalic vial: SN-17-174). **G–H.** *Z. pacarus* (Godart, [1824]), ♀, lateral and ventral views (Genitalic vial: DZ 36. 731).



**Fig. 7.** *Zischkaia* “*saundersii* clade”, female genitalia. **A–B.** *Z. saundersii* (Butler, 1867), ♀, lateral and ventral views (Genitalic vial: DZ 36. 969). **C–D.** *Z. josti* Nakahara & Kleckner, sp. nov., ♀, lateral and ventral views (Genitalic vial: 2018-002, D.J. Harvey). **E–F.** *Z. mielkeorum* Dolibaina, Dias & Zacca, sp. nov., ♀, lateral and ventral views (Genitalic vial: DZ 36. 688). **G–H.** *Z. warreni* Dias, Zacca & Dolibaina, sp. nov., ♀, lateral and ventral views (Genitalic vial: DZ 36. 731).

*E. peculiaris* Butler, 1874 and *E. erichtho* Butler, 1867. The last taxon was described and placed in Division VII in Butler's (1867a) classification, but the name is currently regarded as a junior subjective synonym of *Neonympha antonina* C. Felder & R. Felder, 1867 (now placed in *Erichthodes* Forster, 1964) (Lamas 2004). Weymer (1911) proposed a similar group, named the “*pacarus* group”, in which he included *E. saundersii*, *E. mima*, *E. fumata*, *E. pacarus*, *E. peculiaris*, *E. insignis*, *E. erichtho* and his *E. amalda*. Forster (1964) erected the genus *Zischkaia* by designating *Euptychia fumata* as the type species and stated: “In this newly erected genus I include some very similar in appearance, large, dorsally unicolorous brown species, which regarding the anatomical structure of the male Genitalia (figs 123–125) correspond well with one another and are differentiated from all other ‘*Euptychia*’ species by the long, slender subunci and unique, unpaired outgrowths of the tegumen, which are located dorsally over the uncus”. Based on these diagnostic characters, in addition to the type species, Forster (1964) recognized *Z. amalda* and *Z. saundersii* in the genus, and figured the male genitalia for all three species. Lamas (2004) included four species, namely *Z. amalda*, *Z. pacarus*, *Z. saundersii*, and *Z. mima*, in addition to recognizing one undescribed species (described here as *Z. arenisca* sp. nov.). The checklist of Lamas (2004) also treated *Euptychia fumata* (described by Butler 1867b: 109) as a junior subjective synonym of *E. pacarus*, and *E. fumata* (cited by Butler 1867a: 501) as a nomen nudum. *Euptychia mima* Butler, 1867, a taxon formerly placed in *Zischkaia*, was recently made the type species of *Nhambikuara* by Freitas *et al.* (2018).

### Distribution and natural history

All confirmed records for *Zischkaia* are exclusively east of the Andes (but see also discussion under *Z. abanico* sp. nov.), where species are found from sea level up to about 1600 m. The known distributions of a number of species are highly restricted, with typically only a single species occurring in a particular locality, although label data suggest local sympatry between *Z. warreni* Dias, Zacca & Dolibaina, sp. nov. and *Z. pacarus* in southeastern Brazil, and *Zischkaia chullachaki* Nakahara & Zacca, sp. nov. and *Z. saundersii* in southeastern Peru. Perhaps notably, these two cases concern species in the two different clades, with no known cases of sympatry in members of the same clade. There are a few published observations on the behavior of *Zischkaia*, which are typically found in forest in close proximity to patches of bamboo (Poaceae: Bambusoideae), their likely hostplant; Brown (1992) reported the larvae of *Z. pacarus* using “bamboo”, as did Freitas in Beccaloni *et al.* (2008) (see also natural history notes under *Z. arenisca* sp. nov.).



**Fig. 8.** Fultura inferior (i.e., juxta). **A.** *Zischkaia pacarus* (Godart, [1824]) (DZ 5. 578), in ventral view. **B.** *Z. saundersii* (Butler, 1867) (DZ 5. 580), in posterior view.

**Species accounts**

*Zischkaia* Forster, 1964

- amalda* (Weymer, 1911)
- arctoa* Nakahara, sp. nov.
- chullachaki* Nakahara & Zacca, sp. nov.
- baku* Zacca, Dolibaina & Dias, sp. nov.
- arenisca* Nakahara, Willmott & Hall, sp. nov.
- argyrosflecha* Nakahara, L. Miller & Huertas, sp. nov.
- pacarus* (Godart, [1824])
  - = *fumata* (Butler, 1867a), nom. nud.
  - = *fumata* (Butler, 1867b)
- abanico* Nakahara & Petit, sp. nov.
- saundersii* (Butler, 1867)
- josti* Nakahara & Kleckner, sp. nov.
- mielkeorum* Dolibaina, Dias & Zacca, sp. nov.
- warreni* Dias, Zacca & Dolibaina, sp. nov.

**Key to species of *Zischkaia***

- DFW and DHW androconial scales present in males; projection from posterior region of tegumen above uncus ‘tusk’-like; uncus narrow and long, somewhat curving down, terminating in a small ‘bulb’ in lateral view; brachia curved dorsally; valva rather short in lateral view, apical point does not extend beyond that of uncus; tip of anterior projection of saccus extends further than tegumen in lateral view; fultura inferior (i.e., juxta) appears as a thin strip in posterior view; developed sclerotized half-ring in median region of ductus bursae present ..... ***Z. pacarus*** clade
- DFW and DHW androconial scales absent in males; projection from posterior region of tegumen above uncus ‘bulb’-like; uncus appearing straight in lateral view; valva apical point extends beyond that of uncus; tip of anterior projection of saccus does not extend beyond tegumen in lateral view; fultura inferior (i.e., juxta) appears as a developed plate in posterior view; developed sclerotized half-ring in median region of ductus bursae absent or reduced..... ***Z. saundersii*** clade

**“*pacarus* clade”:**

1. VHW ocelli elongated (ocelli in VHW cells M<sub>2</sub>-M<sub>3</sub>, M<sub>3</sub>-Cu<sub>1</sub>, and Cu<sub>1</sub>-Cu<sub>2</sub> occupying more than half space between submedian line and submarginal line) ..... 2
  - VHW ocelli not elongated, rather circular (ocelli in VHW cells M<sub>2</sub>-M<sub>3</sub>, M<sub>3</sub>-Cu<sub>1</sub> and Cu<sub>1</sub>-Cu<sub>2</sub> occupying less than half space between submedian line and submarginal line) ..... 3
2. Prominent ocellus absent in VHW Rs-M<sub>1</sub>; lack of silverish-purple scales on the basal side of VHW ocelli (distad of submedian line); one winglet on aedeagus; aedeagus winglet obtuse-angled; ventral margin of apical process of valva not concave; saccus longer than ventral margin of valva..... ***Z. arenisca*** Nakahara, Willmott & Hall, sp. nov.
  - Prominent ocellus present in VHW Rs-M<sub>1</sub>; two winglets on aedeagus; aedeagus winglet an acute-angled triangle; ventral margin of apical process of valva concave; saccus shorter than ventral margin of valva ..... ***Z. argyrosflecha*** Nakahara, L. Miller & Huertas, sp. nov.
3. Apical process of valva somewhat rectangular with a convex distal margin ..... 7
  - Apical process of valva somewhat subtriangular ..... 4
4. Adults small (forewing length 21–23 mm); male dorsal androconial scales rather indistinct; from Andean foothills..... 5

- Adults large (forewing length around 25 mm); male dorsal androconial scales distinct; from Amazon Basin ..... 6
- 5. Apical process of valva somewhat elongate; ventral margin of apical process of valva convex; from Cordillera de la Costa (Venezuela) ..... *Z. arctoa* Nakahara, sp. nov.
  - Apical process of valva not elongate; ventral margin of apical process of valva almost straight or concave; from Andean foothills of Peru and Bolivia..... *Z. amalda* (Weymer, 1911)
- 6. Posterior projection of tegumen rather straight; winglet of aedeagus reduced, almost absent; lamella antevaginalis developed as a tube around ductus bursae.. *Z. baku* Zacca, Dolibaina & Dias, sp. nov.
  - Posterior projection of tegumen curved; winglet of aedeagus prominent, clearly visible; lamella antevaginalis not developed as a tube around ductus bursae..... *Z. chullachaki* Nakahara & Zacca, sp. nov.
- 7. Southern and southwestern Brazil to northeastern Argentina..... *Z. pacarus* (Godart, [1824])
  - Tropical Andes of Colombia and Ecuador..... *Z. abanico* Nakahara & Petit, sp. nov.

“*saundersii* clade”:

- 1. VHW marked with prominent blue/grayish shading between submedian line and marginal line; signa long, extending almost entire length of corpus bursae..... *Z. saundersii* (Butler, 1867)
  - VHW not marked with prominent blue/grayish shading between submedian line and marginal line; signa not extending across the entire corpus bursae..... 2
- 2. DHW submarginal and marginal line clearly visible; ocellus in VHW  $M_1$ - $M_2$  often less than half size of that in VHW  $M_2$ - $M_3$ ; median line and submarginal line fused immediately after 2A in males ..... *Z. warreni* Dias, Zacca & Dolibaina, sp. nov.
  - DHW submarginal and marginal line almost invisible; ocellus in VHW  $M_1$ - $M_2$  often about half size (or larger) of that in VHW  $M_2$ - $M_3$ ; median line and submarginal line not fused immediately after 2A in males..... 3
- 3. VHW ocelli in  $M_1$ - $M_2$  and/or  $Cu_2$ -2A appearing as incomplete ocelli with black central area and/or pupil indistinct; small sclerotized region absent at one-third distance from ostium bursae to corpus bursae; from southeastern Brazil ..... *Z. mielkeorum* Dolibaina, Dias & Zacca, sp. nov.
  - VHW ocelli in  $M_1$ - $M_2$  and/or  $CuP$ -2A appearing as complete ocellus with black central area and/or pupil rather distinct and clearly visible; small sclerotized region present at one-third distance from ostium bursae to corpus bursae; from Guianas..... *Z. josti* Nakahara & Kleckner, sp. nov.

*Zischkaia amalda* (Weymer, 1911)

Figs 2A–B, 4A, 14

*Euptychia amalda* Weymer, 1911: 213, pl. 48, fig. f[2] (Type Locality: Mapiri [La Paz, Bolivia]).

*Euptychia amalda* – Gaede 1931: 437. — D’Abrera 1988: 789.

*Zischkaia amalda* – Forster 1964: 116–117, fig. 124. — Lamas 2004: 223.

**Material examined**

**Lectotype** (here designated)

BOLIVIA • ♂; “//♂Typus Nr. *Euptychia amalda* (Stgr)Weymer Zoologische Staatssammlung München.// *amalda* Weym[er].// Mapiri [La Paz, Bolivia]// 521.// 261.// Original [i.e., Type]// Genitalic vial SN-17-21 S. Nakahara//”; ZSM.

**Other material** (12 ♂♂)

BOLIVIA – **La Paz** • 1 ♂; Coroico; [16°10' S, 67°44' W]; 1200 m; Fassl A.H. leg.; ZSM • 1 ♂; Río Zongo; [16°3'40" S, 68°1'2" W]; 1200 m; 1895–1896; Garlepp leg.; MNHU • 1 ♂; Zongo [= Río Zongo]; [16°7' S, 68°2' W]; Garlepp leg.; MNHU • 1 ♂; same collection data as for preceding; BMNH(E)-10247952; NHMUK.

PERU – **Huánuco** • 1 ♂; Tingo María; [9°18' S, 76°0' W]; 800 m; 7 Aug. 1979; FLMNH-MGCL-1036040; FLMNH. – **Junín** • 1 ♂; Chanchamayo; [11°4' S, 75°19' W]; 1000–1400 m; Jun.–Aug. 1901; W. Hoffmanns leg.; BMNH(E)-1718080; NHMUK. – **Not located** • 6 ♂♂; “Peru” [most likely from Bolivia]; USNM.

**Identification and taxonomy**

The male of *Zischkaia amalda* is distinguished from *Z. baku* sp. nov. and *Z. chullachaki* sp. nov. by the VFW submarginal line not undulating (the VFW submarginal line is undulating in *Z. baku* sp. nov. and *Z. chullachaki* sp. nov.), with the area of dorsal androconia rather reduced and the androconia not as prominent as in those two species. Furthermore, the male genitalia can be used to distinguish this species from *Z. baku* sp. nov., with the winglet of the aedeagus being developed (Fig. 4A) (reduced in *Z. baku* sp. nov.), and the anterior margin of the tegumen being concave in dorsal view in *Z. amalda*, whereas it is rather straight or convex in *Z. baku* sp. nov. and *Z. chullachaki* sp. nov. The male of *Z. amalda* is distinguished from that of *Z. pacarus* by the dorsal androconia being more prominent (dorsal androconia somewhat inconspicuous in *Z. pacarus*), in addition to its smaller adult size (*Z. amalda*: FW length 22–23 mm (n = 3); *Z. pacarus*: 25 mm (n = 5)). The male genitalia of *Z. amalda* is distinguished from that of *Z. pacarus* by having a subtriangular apical process of the valva (apical process of valva somewhat rectangular with a convex distal margin in *Z. pacarus*). *Zischkaia amalda* is perhaps most similar to *Z. arctoa* sp. nov., but see the relevant section of *Z. arctoa* sp. nov. for diagnostic characters to separate these two species. Also, see the relevant section of *Z. arenisca* sp. nov. for diagnostic characters to distinguish *Z. amalda* from *Z. arenisca* sp. nov. and *Z. chullachaki* sp. nov.

*Euptychia amalda* was described by Weymer (1911) based on an unstated number of specimens from Mapiri, La Paz, Bolivia. The original description is accompanied by a ventral surface illustration (Weymer 1911: pl. 48, fig. f[2]), but the description and illustration alone cannot be used to confidently identify this species due to the fact that wing patterns of species of *Zischkaia* are often similar. The syntype male, located in the ZSM, has androconia on the DFW and DHW, which is implied in the original description (“The basal and median areas of the forewing above are velvety dark brown, the costal and distal margins lighter brown”), indicating that the description did indeed apply to a species in the “*pacarus* clade” of *Zischkaia*. Forster (1964), examined the abovementioned syntype, in addition to another male specimen from Coroico, La Paz, Bolivia (collected by A.H. Fassl), and stated that “this species is apparently widespread in the Yungas, this rare species is similar to the common southern Brazil [*Z.*] *fumata* [i.e., *Z. pacarus*] and it is so similar that it can be seen as subspecies of a single species. In genital apparatus no differences can be observed, however, [*Z.*] *amalda* is smaller with lighter brown color of the dorsal side of the wings so that the androconial spot of the males emerges more clearly”. Forster (1964: 114, fig. 124) correctly identified *Z. pacarus* (referred to as *Z. fumata*), but we disagree with his claim of a lack of genitalic differences between *Z. amalda* and *Z. pacarus*, having observed slight differences between the male genitalia of these two species (see Fig. 4A, G). Nevertheless, the specimens of *Z. amalda* deposited at the ZSM examined by Forster were not dissected and the origin of the specimen he dissected is unknown. It is therefore not possible to verify whether he dissected *Z. amalda* or some other similar species, such as *Z. chullachaki* sp. nov., which has similar genitalia. Due to resemblance of this taxon to congeners described in this study, we designate as lectotype of *Euptychia amalda* Weymer 1911 the male syntype, illustrated by Warren *et al.* (2018) and deposited at the ZSM, in order to settle the nomenclature (**lectotype designation**).

### Variation

The curvature of the VHW median line below  $M_3$  is apparently variable; the curvature of the phallobase is also apparently variable.

### Distribution (Fig. 14)

This species is currently known from the foothills of the eastern Andes, from Tingo María, Huánuco, Peru to Coroico, La Paz, Bolivia.

*Zischkaia arctoa* Nakahara, sp. nov.

[urn:lsid:zoobank.org:act:AF533240-EA7F-41D3-8100-F9675291FF95](https://zoobank.org/urn:lsid:zoobank.org:act:AF533240-EA7F-41D3-8100-F9675291FF95)

Figs 2C–D, 4B, 14

### Diagnosis

*Zischkaia arctoa* sp. nov. is similar to *Z. amalda*, but is distinguished by the concavity of the ventral margin of the apical process of the valva, in addition to the distal margin of the winglet of the aedeagus being shorter compared to the basal margin. The ventral margin of the apical process of the valva is less concave than in *Z. amalda*, and the length of the distal margin of the winglet of the aedeagus is similar to that of the basal margin in *Z. amalda*. However, given the limited number of specimens of both species available for examination, we suggest diagnosing these two species based on their locality; *Z. arctoa* sp. nov. is currently known from the Cordillera de la Costa, Venezuela and *Z. amalda* is known to date from the east Andean foothills from central Peru to Bolivia. We do not know of any putative female specimen for this species.

### Etymology

This specific epithet is based on the feminine Latin adjective ‘*arctoa*’, in accordance with the feminine generic name, meaning ‘northern’, ‘of the far north’, in reference to the fact that this species is the most northerly occurring species of *Zischkaia*.

### Type material examined

#### Holotype

VENEZUELA • ♂; “// VENEZUELA: DIST. FEDERAL Massif du Naiguata 720–800 m. 28.vii 1943; R. Lichy// Allyn Museum Acc. 1986-5// Genitalic vial SN-17-73 S. Nakahara// UF FLMNH MGCL 1036042//”; FLMNH.

#### Paratypes (3 ♂♂)

VENEZUELA – **Aragua** • 1 ♂; Choróní; [10°29' N, 67°37' W]; 1250 m; 1975; RFC • 1 ♂; Choróní; [10°29' N, 67°37' W]; 850 m; May 1979; RFC. – **Miranda** • 1 ♂; Massif du Naiguatá; [10°31' N, 66°49' W]; 3 Jun. 1940; R. Lichy leg.; FLMNH-MGCL-1036041; FLMNH.

### Description

#### Male

FOREWING LENGTH. 21–23 mm (n = 4).

HEAD. Eyes naked, with grayish scales at base; frons brownish; post-genal area with lightly colored long hair-like scales and grayish scales; labial palpi with first segment mostly with brownish long hair-like scales; second segment length almost twice as great as eye depth and adorned with brown scales laterally, dorsally mostly with light brownish long hair-like scales, ventrally adorned with brownish and white hair-like scales, about 3–4 times as long as segment width; third segment about two-fifth of second

segment in length and covered with brownish scales, ventrally with brownish hair-like scales, with slight patch of creamy-white scales laterally; antennae approximately two-fifth of forewing length, with ca 39 segments ( $n = 1$ ), distal 13–15 segments composing club, club not prominent.

THORAX. Dorsally, laterally and ventrally scattered with grayish scales with long multi-colored hair-like scales.

LEGS. Foreleg brownish, foretarsus slightly shorter than tibia, femur similar to tarsus in length; midleg and hindleg with femur creamy white ventrally, tibia and tarsus grayish dorsally, whitish ventrally, tarsus and tibia spined ventrally, and a pair of tibial spurs present at distal end of tibia.

ABDOMEN. Eighth tergite as stripe at base of eighth abdominal segment, in addition to presence of distal broader patch; eighth sternite divided into two patches.

WING VENATION. Basal half of forewing Subcosta swollen; base of Cubitus swollen; forewing recurrent vein absent; origin of  $M_2$  towards  $M_1$  than  $M_3$ ; hindwing humeral vein developed.

WING SHAPE. Forewing subtriangular, apex rounded, costal margin convex, outer margin slightly convex (almost straight), inner margin straight, but rounded towards thorax near base; hindwing slightly elongate, rounded, costal margin almost straight, angled towards thorax near base, outer margin slightly undulating, inner margin slightly concave near tornus, anal lobe convex, slightly round.

DORSAL FOREWING. Ground color light brownish, distally slightly paler; black androconial scales, not prominent, present in middle of DFW, from base to submedian area; trace of submarginal band almost invisible.

DORSAL HINDWING. Ground color similar to forewing, black androconial scales present in discal and adjacent area, not as prominent as DFW; trace of submarginal band almost invisible (clearly visible in one specimen).

VENTRAL FOREWING. Ground color light chestnut brown; submedian line invisible; dark brown narrow median line extends from near costa to  $Cu_2-2A$ , in outward diagonal direction, concolorous slightly sinuate submarginal line extending from apex towards tornus, but terminates around 2A; concolorous marginal line, narrower than submarginal line, extending from apex towards tornus, but terminates around 2A; fringe dark brownish.

VENTRAL HINDWING. Ground color similar to forewing; regular dark-brown submedian line almost straight, extending from costal to inner margin; median line almost parallel to submedian line, concolorous, similar in width, passing origin of  $M_3$ , curved inwards after passing origin of  $M_3$  and posterior end bent inwards in 2A-A; submarginal line extending from apex towards tornus, anterior end occasionally fused with submedian line near costa, undulating, posterior end slightly broadening and occasionally fused to submedian line in 2A-3A; marginal line, concolorous, slightly undulating along outer margin, thinner than submarginal line; submarginal ocelli from  $M_1$  to 2A (tiny ocellus in  $Rs-M_1$  in three specimens), rounded, pupil appear as scattered silverish scales placed distally, black central spot ringed with orangish ring then with thin dark brownish indistinct ring, ocellus in  $Cu_2-2A$  smallest; bright silvery purple ground color visible between submedian line and submarginal line when seen under light; fringe dark brownish.

GENITALIA (Fig. 4B). Tegumen rounded in lateral view, elongated posterior projection of tegumen developed, apparently slightly shorter than uncus, tapering posteriorly and hooked at terminal point; combination of ventral arms of tegumen and dorsal arms of saccus sinuous, broadening towards saccus;



appendices angulares present, but somewhat reduced; saccus anterior half slightly curved upwards, similar to uncus in length; uncus long and narrow, sparsely with hair-like setae, curved in lateral view, rounded at terminal point, posterior end of ventral margin appearing as small projection; either side of base of uncus with hair-like setae; brachia similar to uncus in length, slightly narrower, curved in lateral view, tapering posteriorly and crossing over each other near terminal point; fultura inferior present; valva subtriangular in lateral view, apical process subtriangular, dorsal and ventral margin almost equally convexed, scarcely covered by hairy-like setae; costa developed and triangular, dorsal margin slightly sinuous; phallobase slightly shorter than half of phallus in length, slightly curved; ductus ejaculatorius coming out higher than anterior end of coecum; aedeagus straight with manica covering approximately one-fifth, winglet present, distal opening located ventrally where vesica is visible.

### Female

Unknown or unrecognized.

### Variation

The VFW submedian line traverses in an outward diagonal direction below  $M_3$  in one examined specimen, whereas it appears rather straight in the other three examined specimens; the size of ocellus in VHW Rs- $M_1$  is variable, ranging from being almost absent to being clearly visible; the curvature of the VHW submedian line below  $M_3$  is apparently variable.

### Distribution (Fig. 14)

This species is known to date from the Cordillera de la Costa, a mountain range extending along the north Venezuelan coast.

### Remarks

*Zischkaia arctoa* sp. nov. is similar to *Z. amalda*, and one might argue their conspecificity due to their morphological resemblance. However, these two taxa are treated as different species here mainly due to their geographical isolation and the presence of allopatrically occurring *Z. abanico* sp. nov. between the range of the two species. *Zischkaia arctoa* sp. nov. occurs in the Cordillera de la Costa, which is isolated from the adjacent Cordillera de la Mérida (the northeastern extension of the tropical Andes) by the flat and scrubby lowlands of the Depression of Unare; *Z. amalda* is known from the eastern slopes of tropical Andes in Bolivia and Peru, which is over 2500 km (straight line distance) from the known range of *Z. arctoa* sp. nov., and with no records in the intervening Andes despite intensive sampling in at least some regions. Unfortunately, we were unable to obtain DNA data for either of these two taxa to help further clarify the taxonomy, and obviously such data would be very valuable.

*Zischkaia chullachaki* Nakahara & Zacca, sp. nov.

[urn:lsid:zoobank.org:act:EBDD3929-4E74-48BE-B0F9-2FE17A9E0567](https://zoobank.org/urn:lsid:zoobank.org:act:EBDD3929-4E74-48BE-B0F9-2FE17A9E0567)

Figs 1, 2E–H, 4C, 6A–B, 12E–F, 14

*Zischkaia amalda* – Lamas *et al.* 1991: 10 [misidentification]. — Lamas 1994: 165 [misidentification].  
— Robbins *et al.* 1996: 232 [misidentification]. — Mielke *et al.* 2010: 291 [misidentification].

### Diagnosis

*Zischkaia chullachaki* sp. nov. is similar to *Z. baku* sp. nov., but is distinguishable based on posterior projection of tegumen being curved in lateral view (posterior projection of tegumen is rather straight in *Z. baku* sp. nov.); posterior projection of tegumen narrow in dorsal view (posterior projection of tegumen rather broad in dorsal view in *Z. baku* sp. nov.); winglet of aedeagus reduced, almost absent (winglet of aedeagus prominent, clearly visible in *Z. baku* sp. nov.); sclerotized tube reduced, almost

invisible in lateral view in *Z. chullachaki* sp. nov. (sclerotized tube about half the length of ductus bursae in *Z. baku* sp. nov.).

### Etymology

The specific epithet is derived from a mythological creature, Chullachaki, a 'forest guardian' known from the Peruvian and Brazilian jungle, in reference to this species known from the Peruvian and Brazilian Amazon. This specific epithet is regarded as a Latinized masculine noun in apposition.

### Type material examined

#### Holotype

BRAZIL • ♂; “// 8-10-IX-2004, Reserva Humaitá, Porto Acre, Acre, 200 m, O. Mielke & Casagrande leg. //DZ 37.009 //BC-DZ Willmott 230 //”; DZUP.

#### Paratypes (46 ♂♂, 3 ♀♀)

BRAZIL – **Acre** • 1 ♂; 50 km NW of Bujari; [9°32'53" S, 68°18'9" W]; 200 m; O.H.H. Mielke and M.M. Casagrande leg.; 10–12 Sep. 2004; DZ-36758; DZUP • 1 ♂; 6.4 km E Santa Rosa do Purus; [9°28'39" S, 70°26'58" W]; 3–4 Aug. 2008; O.H.H. Mielke and E. Carneiro leg.; DZ-37043; DZUP • 1 ♂; Assis Brasil, Estação Ecológica do Alto Acre; [11°3' S, 70°16' W]; 300 m; 26 Aug. 2005; K.S. Brown leg.; ZUEC • 4 ♂♂; Porto Acre, Reserva Humaitá; [9°45'18" S, 67°36'50" W]; 200 m; 8–10 Sep. 2004; O.H.H. Mielke and M.M. Casagrande leg.; DZ-21235, DZ-21620, DZ-36578, DZ-36848; DZUP.

PERU – **Madre de Dios** • 2 ♂♂; 0–2 km W of Puerto Maldonado; [12°36'12" S, 69°12'32" W]; 14 Aug. 1981; L.D. Miller; FLMNH-MGCL-1036033, FLMNH-MGCL-1036034; FLMNH • 1 ♂; same collection data as for preceding but 16 Aug. 1981; FLMNH-MGCL-1036035; FLMNH • 1 ♂; Boca Río La Torre; [12°50' S, 69°17' W]; 300 m; 24 Oct. 1983; G. Lamas leg.; MUSM-LEP-105641; MUSM • 1 ♀; same collection data as for preceding but 24 Sep. 1981; MUSM-LEP-105651; MUSM • 1 ♂; same collection data as for preceding but 26 Sep. 1981; MUSM-LEP-105642; MUSM • 1 ♂; same collection data as for preceding but 27 Sep. 1981; MUSM-LEP-105640; MUSM • 1 ♂; same collection data as for preceding but 30 Nov. 1979; MUSM-LEP-105643; MUSM • 1 ♂; Cerro Pantiacolla, E slope, 5–6 km ENE of Shintuya; [12°39'53" S, 71°13'8" W]; 15 Aug. 1980; J.F. Douglass leg.; FLMNH-MGCL-1036036; FLMNH • 1 ♂; same collection data as for preceding but 19 Aug. 1980; FLMNH-MGCL-1036037; FLMNH • 1 ♂; Los Amigos Biological Station; [12°34'2" S, 70°5'56" W]; 250 m; 23 Oct. 2005; C. Peña; MUSM-LEP-105649; MUSM • 1 ♂; same collection data as for preceding but 3 Oct. 2005; MUSM-LEP-105650; MUSM • 1 ♂; Parque Manu, Pakitza; [11°55'48" S, 71°15'18" W]; 340 m; 10 Oct. 1991; R.K. Robbins leg.; MUSM-LEP-105648; MUSM • 1 ♀; Parque Manu, Pakitza; [11°55'48" S, 71°15'18" W]; 340–400 m; 1 Oct. 1991; M. Casagrande leg.; DZ-37003; DZUP • 1 ♂; same collection data as for preceding but 11 Oct. 1991; DZ-36733; DZUP • 1 ♂; same collection data as for preceding but 13 Oct. 1991; DZ-21228; DZUP • 1 ♂; same collection data as for preceding but 28 Sep. 1991; DZ-37023; DZUP • 1 ♀; same collection data as for preceding but 30 Sep. 1991; DZ-36508; DZUP • 1 ♂; (same collection data as for preceding but 30 Sep. 1991; Häuser leg.; DZ-36783; DZUP • 1 ♂; same collection data as for preceding but 14 Oct. 1991; G. Lamas leg.; DZ-37033; DZUP • 1 ♂; same collection data as for preceding but 27 Sep. 1991; DZ-36983; DZUP • 1 ♂; same collection data as for preceding but 28 Sep. 1991; DZ-36993; DZUP • 2 ♂♂ same collection data as for preceding but 10 Oct. 1991, Mielke leg.; DZ-36608, DZ-36793; DZUP • 1 ♂; same collection data as for preceding but 26 Sep. 1991; DZ-36763; DZUP • 1 ♂; same collection data as for preceding but 27 Sep. 1991; DZ-21627; DZUP • 4 ♂♂; same collection data as for preceding but 4 Oct. 1991; DZ-36019, DZ-36753, DZ-36803, DZ-37013; DZUP • 1 ♂; same collection data as for preceding but 26 Sep. 1991; R.K. Robbins leg.; DZ-36743; DZUP • 1 ♂; Parque Manu, Pakitza; [11°55'48" S, 71°15'18" W]; 400 m; 23 Sep. 1989; D.J. Harvey leg.; USNM • 1 ♂; same collection data as for preceding but 2 Oct. 1990; G. Lamas leg.; USNM • 1 ♂ same collection data as for preceding but 2 Oct. 1990; G. Lamas leg.; MUSM-LEP-105646; MUSM • 1 ♂; same collection data as for preceding but 8 Oct. 1990; MUSM-LEP-105647; MUSM • 1 ♂; same collection data as for preceding but 8–14 Sep. 1989; MUSM-LEP-105644; MUSM • 1 ♂; same

collection data as for preceding but 14 Sep. 1989; M.G. Pogue leg.; MUSM-LEP-105645; MUSM • 1 ♂; Albergue Pantiacolla, 12°39' S, 71°14' W, 400 m, 1 Nov. 2018; D. Bolt leg.; MUSM. – **Puno** • 2 ♂♂; Río Tambopata; [12°36' S, 69°11' W]; 270 m; 15 Jul. 1979; FLMNH-MGCL-1036038 to FLMNH-MGCL-1036039; FLMNH • 1 ♂; Río Tambopata, Z.R. Tambopata-Candamo; [13°22' S, 69°34' W]; 270 m; 1 Jan.; FLMNH-MGCL-1036045; FLMNH • 1 ♂; same collection data as for preceding but 26 Aug. 1995; Cambridge University leg.; BMNH(E)-1718103; NHMUK • 1 ♂; same collection data as for preceding but 3 Sep. 1995; BMNH(E)-1718102; NHMUK.

## **Description**

### **Male**

FOREWING LENGTH. 23–25 mm (n = 28).

HEAD. Eyes naked, with grayish scales at base; frons brownish; post-genal area with lightly colored long hair-like scales and grayish scales; labial palpi first segment with whitish scales and light brown long hair-like scales ventrally; second segment length almost twice as great as eye depth and with brown and white scales laterally, dorsally mostly with brownish and whitish hair-like scales, ventrally with brownish long hair-like scales and white hair-like scales, about 3–4 times as long as segment width; third segment almost half of second segment in length and covered with brownish scales, ventrally with brownish hair-like scales, with slight patch of creamy-white scales laterally; antennae approximately two-fifth of forewing length, with ca 39–47 segments (n = 2), distal 13–15 segments composing club, club not prominent.

THORAX. Dorsally, laterally and ventrally scattered with grayish scales, and long multi-colored hair-like scales.

LEGS. Foreleg brownish, foretarsus longer than tibia and femur, tibia shorter than femur in length; midleg and hindleg with femur creamy white ventrally, tibia and tarsus grayish dorsally, whitish to ocher ventrally, tarsus and tibia spined ventrally, and a pair of tibial spurs at distal end of tibia.

ABDOMEN. Eighth tergite as stripe at base of eighth abdominal segment, in addition to presence of distal broader patch; eighth sternite divided into two patches.

WING VENATION. Basal half of forewing Subcosta swollen; base of Cubitus swollen; forewing recurrent vein absent; origin of  $M_2$  towards  $M_1$  than  $M_3$ ; hindwing humeral developed.

WING SHAPE. Forewing subtriangular and appear elongated, apex angular, costal margin convex, outer margin slightly convex (almost straight), inner margin straight, but rounded towards thorax near base; hindwing slightly elongate, rounded, costal margin almost straight, angled towards thorax near base, outer margin slightly undulating, inner margin slightly concave near tornus, anal lobe convex, slightly round.

DORSAL FOREWING. Ground color grayish brown; prominent black androconial scales present in middle of DFW, from base to submedian region occupying the space between Radius and inner margin; trace of submarginal and marginal line visible.

DORSAL HINDWING. Ground color similar to forewing, black androconial scales present in discal cell and adjacent area, from  $M_1$  to 2A; trace of submarginal and marginal line visible.

VENTRAL FOREWING. Ground color grayish chestnut brown; submedian line invisible; dark brown narrow median line extends from near costa to  $Cu_2$ -2A, terminates within this cell, in slightly outward diagonal direction; concolorous undulating submarginal line extending from apex towards tornus, terminates

around 2A; concolorous marginal line, narrower than submarginal line, extending from apex towards tornus, terminates around 2A; fringe dark brown.

**VENTRAL HINDWING.** Ground color similar to forewing; regular dark-brown submedian line almost straight, extending from costal to inner margin, curves inwards in 2A-3A; median line almost parallel to submedian line, concolorous, similar in width, passing origin of  $M_3$ , curved inwards after passing origin of  $M_3$ , and posterior end bent inwards in 2A-3A; undulating submarginal line extending from apex towards tornus, anterior end occasionally fused with submedian line near costa, posterior end slightly broadening and occasionally fused to submedian line in 2A-3A; marginal line, concolorous, slightly undulating along outer margin, narrower than submarginal line; submarginal ocelli from Rs to 2A, rounded, pupil as scattered silverish scales placed distally, black central spot ringed with orangish ring then with thin dark brownish indistinct ring; light purplish lilac coloration visible between submedian line and submarginal line when seen under lighting; fringe dark brownish.

**GENITALIA** (Fig. 4C). Tegumen rounded in lateral view, elongated posterior projection of tegumen developed, curved upwards in lateral view, apparently shorter than uncus, tapering posteriorly and hooked at terminal point, posterior end of ventral margin appearing as small projection; combination of ventral arms of tegumen and dorsal arms of saccus sinuous, broadens towards saccus; appendices angulares present, but somewhat reduced; saccus straight, similar to uncus in length; uncus long and narrow, sparsely with hair-like setae, curved in lateral view, rounded at terminal point forming small ‘bulb’; either side of base of uncus with hair-like setae; brachia similar to uncus in length and width, curved in lateral view, tapering posteriorly and crossing over each other near terminal point; fultura inferior (i.e., juxta) present; valva subtriangular in lateral view, apical process subtriangular, dorsal margin convexed, ventral margin concave, scarcely covered by hairy-like setae, costa developed and triangular, dorsal margin slightly sinuous; phallobase about two-fifth of phallus, rather straight; ductus ejaculatorius coming out higher than anterior end of coecum; aedeagus straight with manica covering about half, winglet present, distal opening located ventrally where vesica is visible.

### **Female**

FW length 23 mm (n = 2).

Similar to male except as follows: foreleg whitish, foretarsus first and second subsegments fused; forewing appears slightly more rounded and broad, ground color of both wing surfaces paler; dorsal androconia absent.

**FEMALE ABDOMEN AND GENITALIA** (Fig. 6A–B): inter-segmental membrane between seventh and eighth tergite not pleated, but folded posteriorly of ostium bursae with its sclerotized region forming a ‘scoop-like’ structure below lamella antevaginalis; lamella antevaginalis fused to lateral sclerotized plate of eighth abdominal segment; lateral plate of eighth abdominal segment expanding laterally with small spiracle at superior third; ductus bursae membranous with a half-ring strongly sclerotized at the median region; corpus bursae three times longer than ductus bursae, with paired signa ventrally occupying  $\frac{2}{3}$  its length.

### **Variation**

The VHW ocellus in Rs- $M_1$  is variable in terms of size and presence.

### **Distribution** (Fig. 14)

This species is known from the western Amazon basin (Junín, Puno and Madre de Dios, Peru, and Acre, Brazil).

## Remarks

See relevant section of *Z. arenisca* sp. nov. regarding discussion on taxonomic status of *Z. chullachaki* sp. nov.

*Zischkaia baku* Zacca, Dolibaina & Dias, sp. nov.

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Figs 1, 2I–L, 4D, 6C–D, 14

## Diagnosis

See relevant section of *Z. chullachaki* sp. nov.

## Etymology

The specific epithet is derived from the nickname ‘Baku’ of our friend and Brazilian butterfly researcher, André V.L. Freitas, who has made significant contributions to our understanding of Neotropical butterflies, especially satyrines. His nickname is derived from the Portuguese common name for a nightjar (Caprimulgidae), ‘Bacurau’, and was given in recognition of his faithful imitations of the song of this bird while at college. The specific epithet is regarded as a Latinized masculine noun in apposition.

## Type material examined

### Holotype

BRAZIL • ♂; “// Imperatriz, MA [Maranhão, Brazil], 18-VI-[19]74, Exc[ursão]. Dep[artamen]to. Zool[ogia, Universidade Federal do Paraná] . //DZ 21.242 //”; DZUP.

### Paratypes (53 ♂♂, 6 ♀♀)

BRAZIL – **Goiás** • 1 ♂; Goianésia; 700 m; 5 Sep. 1969; Ebert leg.; DZ-36661; DZUP • 1 ♂, 1 ♀; Goianésia; 900 m; 5 Sep. 1969; Ebert leg.; DZ-5572, DZ-5571; DZUP • 1 ♀; Goiás Velho; [15°34′30″ S, 50°18′ W]; 9 Feb. 1980; Gifford leg.; DZ-36943; DZUP. – **Maranhão** • 1 ♂, 1 ♀; Imperatriz; [5°31′ S, 47°28′ W]; 12 Aug. 1974; Dep. Zool. UFPR leg.; DZ-36733, DZ-36913; DZUP • 1 ♀; same collection data as for preceding but 20 Jul. 1974; DZ-36933; DZUP • 1 ♀; same collection data as for preceding but 25 Aug. 1974; DZ-36404; DZUP • 1 ♂; same collection data as for preceding but 6 Aug. 1974; DZ-36923; DZUP. – **Mato Grosso** • 1 ♂; Aragão, Jacareacanga; 14 Feb. 2009; ZUEC • 1 ♂; Barra do Garça; 19 Apr. 1978; Gifford leg.; DZ-36673; DZUP • 1 ♂; same collection data as for preceding but 21 Apr. 1978; DZ-36712; DZUP • 2 ♂♂; same collection data as for preceding but 26 Apr. 1978; DZ-36623; DZUP • 2 ♂♂; Cáceres; 13 Nov. 1984; Buzzi, M. Casagrande, C. Elias and O.H.H. Mielke leg.; DZ-36593, DZ-5573; DZUP • 2 ♂♂; same collection data as for preceding but 15 Nov. 1984; DZ-36703, DZ-36979; DZUP • 1 ♂; same collection data as for preceding but 18 Nov. 1984; DZ-36721; DZUP • 1 ♂; Diamantino, Alto Rio Arinos, Faz.[enda] S.[ão] João; [14°21′18″ S, 56°9′2″ W]; 14 Jan. 1978; O.H.H. Mielke and M.M. Casagrande leg.; DZ-36903; DZUP • 2 ♂♂; same collection data as for preceding but 17 Jan. 1978; DZ-36893, DZ-36953; DZUP • 1 ♂; same collection data as for preceding but 21 Nov. 1984; DZ-36681; DZUP • 1 ♂; same collection data as for preceding but 6 Sep. 1978; DZ-36683; DZUP • 1 ♂; same collection data as for preceding but 12 Jan. 1978; O.H.H. Mielke and Furtado leg.; DZ-36603; DZUP • 1 ♂; same collection data as for preceding but 14 Jan. 1978; DZ-36701; DZUP • 1 ♂; same collection data as for preceding but 22 Jan. 1978; DZ-36691; DZUP • 1 ♂; Diamantino, Alto Rio Arinos, Faz.[enda] S.[ão] João, [14°21′18″ S, 56°9′2″ W]; 300–400 m; 2 Sep. 1975; H.-H. Ebert leg.; DZ-36723; DZUP • 1 ♂; same collection data as for preceding but 26 Jul. 1975; DZ-36593; DZUP • 1 ♂; same collection data as for preceding but 2 Sep. 1975; E. Furtado leg.; DZ-36583; DZUP • 1 ♂; Jaciara; 9 Feb. 1967; K.S. Brown leg.; FLMNH-MGCL-1036032; FLMNH • 1 ♂; nr. Melguira, Alto Rio Paraguai, Barra do Bugres; [15°4′ S, 57°10′ W]; 150 m; 26–29 Jul. 1974; H.-H. Ebert leg.; DZ-36693; DZUP • 1 ♂; Rio Teles Pires, Cachoeira Sete Quedas; [9°20′15″ S,

56°46'39" W]; 170 m, 11 Apr. 2009; ZUEC. – **Rondônia** • 1 ♂; Fazenda Rancho Grande, Cacaulândia (62 km S Ariquemes, 5 km S of Cacaulândia on linea C-10); [10°17'51" S, 62°32'07" W]; 1 Apr. 1995; O. Gomes leg.; FLMNH-MGCL-1036022; FLMNH • 1 ♂; same collection data as for preceding but 10 Apr. 1994; FLMNH-MGCL-1036023; FLMNH • 1 ♂; same collection data as for preceding but 11 Feb. 1995; FLMNH-MGCL-1036028; FLMNH • 1 ♂; same collection data as for preceding but 15 Apr. 1995; FLMNH-MGCL-1036012; FLMNH • 1 ♂; same collection data as for preceding but 17 Dec. 1994; FLMNH-MGCL-1036021; FLMNH • 1 ♂; same collection data as for preceding but 18 Sept 1993; FLMNH-MGCL-1036014; FLMNH • 2 ♂♂; same collection data as for preceding but 19 Nov. 1994; FLMNH-MGCL-1036017, FLMNH-MGCL-1036027; FLMNH • 1 ♂; same collection data as for preceding but 2 Apr. 1994; FLMNH-MGCL-1036009; FLMNH • 1 ♂; same collection data as for preceding but 2 Aug. 1993; FLMNH-MGCL-1036016; FLMNH • 1 ♂; same collection data as for preceding but 21 Jan. 1995; FLMNH-MGCL-1036015; FLMNH • 1 ♂; same collection data as for preceding but 22 Apr. 1995; FLMNH-MGCL-1036007; FLMNH • 1 ♂; same collection data as for preceding but 24 Jul. 1994; FLMNH-MGCL-1036026; FLMNH • 1 ♂; same collection data as for preceding but 24 Nov. 1994; FLMNH-MGCL-1036008; FLMNH • 2 ♂♂; same collection data as for preceding but 25 Mar. 1995; FLMNH-MGCL-1036011, FLMNH-MGCL-1036013; FLMNH • 1 ♀; same collection data as for preceding but 27 May 1995; FLMNH-MGCL-1036029; FLMNH • 2 ♂♂; same collection data as for preceding but 28 Jan. 1995; FLMNH-MGCL-1036018, FLMNH-MGCL-1036020; FLMNH • 1 ♂; same collection data as for preceding but 30 Jan. 1994; FLMNH-MGCL-1036019; FLMNH • 2 ♂♂; same collection data as for preceding but 30 Mar. 1994; FLMNH-MGCL-1036024, FLMNH-MGCL-1036025; FLMNH • 1 ♂; same collection data as for preceding but 7 Jan. 1995; FLMNH-MGCL-1036006; FLMNH • 1 ♂; same collection data as for preceding but 8 Jan. 1996; FLMNH-MGCL-1036010; FLMNH • 1 ♂; [10°32' S, 62°48' W]; 15 Apr. 1995; O. Gomes leg.; FLMNH-MGCL-1036030; FLMNH • 1 ♂; same collection data as for preceding but 7 Jul. 1995; FLMNH-MGCL-1036031; FLMNH.

## Description

### Male

FOREWING LENGTH. 21–24 mm (n = 21).

HEAD. Eyes naked, with grayish scales at base; frons brownish with grayish scales and hair-like scales; post-genal area with brownish long hair-like scales and grayish scales; labial palpi first segment with whitish scales laterally and dorsally, and light brown long hair-like scales and whitish long hair-like scales ventrally; second segment length almost twice as great as eye depth and with brown and white scales laterally, dorsally with brownish and whitish hair-like scales, ventrally with brownish and white hair-like scales, about 3–4 times as long as segment width; third segment almost half of second segment in length and covered with brownish scales, ventrally with brownish and whitish hair-like scales, and slight patch of creamy-white scales laterally; antennae approximately two-fifth of forewing length, with ca 45–50 segments (n = 2), distal 12–14 segments composing club, club not prominent.

THORAX. Dorsally, laterally and ventrally scattered with grayish scales with long multi-colored hair-like scales.

LEGS. Foreleg brownish, foretarsus shorter than tibia and femur, tibia and femur similar in length; midleg and hindleg with femur creamy white ventrally, tibia and tarsus grayish dorsally, whitish to ocher ventrally, tarsus and tibia spined ventrally, and a pair of tibial spurs present at distal end.

ABDOMEN. Eighth tergite as stripe at base of eighth abdominal segment, in addition to presence of distal broader patch; eighth sternite divided into two patches.

WING VENATION. Basal half of forewing Subcosta swollen; base of Cubitus swollen; forewing recurrent vein absent; origin of  $M_2$  towards  $M_1$  than  $M_3$ ; hindwing humeral developed.

WING SHAPE. Forewing subtriangular and elongated, apex angular, costal margin convex, outer margin slightly convex (almost straight), inner margin straight, but rounded towards thorax near base; hindwing slightly elongate, rounded, costal margin almost straight, angled towards thorax near base, outer margin slightly undulating, inner margin slightly concave near tornus, anal lobe convex, slightly round.

DORSAL FOREWING. Ground color grayish brown; prominent black androconial scales present in middle of DFW, from base to submedian region occupying the space between Radius and inner margin; trace of submarginal and marginal line visible.

DORSAL HINDWING. Ground color similar to forewing, black androconial scales present in discal cell and adjacent area (not reaching the submarginal line), from  $R_s$  to 2A; trace of submarginal and marginal line visible.

VENTRAL FOREWING. Ground color grayish chestnut brown; submedian line invisible; dark brown narrow median line extends from near costa to  $Cu_2$ -2A, terminates within this cell, in slightly outward diagonal direction, concolorous undulating submarginal line extending from apex towards tornus, terminates around 2A; concolorous marginal line, narrower than submarginal line, extending from apex towards tornus, terminates around 2A; fringe dark brown.

VENTRAL HINDWING. Ground color similar to forewing; regular dark-brown submedian line almost straight, extending from costal to inner margin, curves inwards in 2A-3A; median line almost parallel to submedian line, concolorous, similar in width, passing origin of  $M_3$ , curved inwards after passing origin of  $M_3$ , and posterior end bent inwards in 2A-3A; undulating submarginal line extending from apex towards tornus, anterior end occasionally fused with median line near costa, posterior end slightly broadening and occasionally fused to submedian line in 2A-3A, space between the marginal line and outer margin ochre; marginal line, concolorous, slightly undulating along outer margin, narrower than submarginal line; submarginal ocelli from  $M_1$  to 2A, rounded (but elliptical in  $M_2$ - $M_3$ ), pupil appear as scattered silverish scales placed distally, black central spot ringed with orangish ring then with thin dark brownish indistinct ring; light purplish lilac coloration visible between submedian line and submarginal line when seen under lighting; fringe dark brownish.

GENITALIA (Fig. 4D). Tegumen subtriangular in lateral view, somewhat elongated, elongated posterior projection of tegumen developed, straight in lateral view, shorter than uncus, tapering posteriorly and hooked at terminal point, posterior end of ventral margin appearing as small projection; combination of ventral arms of tegumen and dorsal arms of saccus slightly sinuous, broadens towards saccus; appendices angulares present, but somewhat reduced; saccus straight, similar to uncus in length; uncus long and narrow, with sparse hair-like setae, well-curved in lateral view, rounded at terminal point forming a 'bulb'; either side of base of uncus with hair-like setae; brachia smaller than uncus in length, but similar in width, hooked-like in lateral view, tapering posteriorly and not crossing over each other near terminal point; futura inferior present; valva subtriangular in lateral view, apical process subtriangular, dorsal margin convexed, ventral margin concave, scarcely covered by hair-like setae; costa developed and triangular, dorsal margin slightly sinuous; phallobase about one-third of phallus, curved upwards; ductus ejaculatorius visible; aedeagus straight with manica covering more than half, reduced winglet, distal opening located ventrally where vesica is visible.

#### **Female**

FW length 21–23 mm (n = 7).

Similar to male except as follows: foreleg whitish, foretarsus variable in number of tarsomers (see below); forewing appears slightly more rounded and broad, ground color of both wing surfaces paler; DFW androconia absent.

FEMALE ABDOMEN AND GENITALIA (Fig. 6C–D): inter-segmental membrane between seventh and eighth tergite not pleated, but folded posteriorly of ostium bursae with its sclerotized region forming a ‘scoop-like’ structure below lamella antevaginalis; lamella antevaginalis fused to lateral sclerotized plate of eighth abdominal segment; lateral plate of eighth abdominal segment expanding laterally with small spiracle at superior third; ductus bursae basally sclerotized, median region until the connection with corpus bursae membranous, but with three times longer than wide and strongly sclerotized half-ring at the median region; corpus bursae  $\frac{2}{3}$  longer than ductus bursae, with paired signa ventrally almost entirely occupying the corpus bursae.

### Variation

Width of ventral lines of the wings is variable, especially those from Rondônia which seem to have broader lines; number of female fore tarsomers variably fused.

### Distribution (Fig. 14)

This species is known to date from northeastern and midwestern Brazil, namely states of Rondônia, Maranhão, Mato Grosso and Goiás, occurring in forested areas of the western Amazon and within the Cerrado biome.

### Remarks

We were unable to obtain DNA data for this taxon to assess its taxonomic status, especially in relation to *Z. chullachaki* sp. nov., which is phenotypically similar and might arguably be considered conspecific. However, the external morphological differences between these two taxa documented under the ‘Diagnosis’ section of *Z. chullachaki* sp. nov. support our decision to treat this taxon as a species until further evidence becomes available.

*Zischkaia arenisca* Nakahara, Willmott & Hall, sp. nov.

[urn:lsid:zoobank.org:act:83701B3C-3A99-4BFC-9E8E-18F03C47CE0B](https://zoobank.org/act:83701B3C-3A99-4BFC-9E8E-18F03C47CE0B)

Figs 1, 2M–P, 4E, 6E–F, 9, 14–15A–B

*Zischkaia* sp. n. – Lamas 1997: 217; 2004: 223.

### Diagnosis

*Zischkaia arenisca* sp. nov. is readily distinguished from all species of *Zischkaia*, except for *Z. argyrosflecha* sp. nov., by its elongate, oval VHW ocelli, whereas those ocelli are more circular and smaller in *Z. pacarus*, *Z. amalda*, *Z. baku* sp. nov. and *Z. chullachaki* sp. nov. Species in the “*saundersii* clade” also possess somewhat elongate VHW ocelli, but the genitalia differences summarized above (see diagnosis for *Zischkaia*), in addition to the lack of dorsal androconial scales in males, easily distinguish this species from species in the “*saundersii* clade”. The male DFW and DHW androconial scales in *Z. arenisca* sp. nov. appear somewhat reduced and less prominent compared to those of *Z. baku* sp. nov. and *Z. chullachaki* sp. nov. Furthermore, the forewing apex is somewhat rounded in *Z. arenisca* sp. nov., whereas it is more angled in *Z. pacarus*, *Z. amalda* and *Z. baku* sp. nov. In the female genitalia, the sclerotized tube of the ductus bursae in lateral view appears to be longer in *Z. arenisca* than in *Z. chullachaki* sp. nov. and *Z. pacarus*.

*Zischkaia arenisca* sp. nov. is likely most closely related to *Z. argyrosflecha* sp. nov., given their similar elongate VHW ocelli, but *Z. arenisca* sp. nov. can be distinguished from *Z. argyrosflecha* sp. nov. by the



following characters: the lack of silvery-purple scales on the basal side of the VHW ocelli (distad of the submedian line); the lack of or very small VHW ocellus in Rs-M<sub>1</sub>, whereas this ocellus is well developed in VHW Rs-M<sub>1</sub> in *Z. argyrosflecha* sp. nov.; only one winglet being present on the aedeagus, whereas *Z. argyrosflecha* sp. nov. possesses two winglets; the shape of the aedeagus winglet is that of an obtuse-angled triangle in *Z. arenisca* sp. nov., whereas it is an acute-angled triangle in *Z. argyrosflecha* sp. nov.; the saccus is longer than the ventral margin of the valva in *Z. arenisca* sp. nov., whereas it is shorter than ventral margin of the valva in *Z. argyrosflecha* sp. nov.; the ventral margin of the apical process of the valva is not concave in *Z. arenisca* sp. nov., whereas it is concave in *Z. argyrosflecha* sp. nov. The female of *Z. argyrosflecha* sp. nov. is unknown and thus it is not possible to compare that sex.

### Etymology

This specific epithet is derived from the Spanish word ‘arenisca’, meaning ‘sandstone’, and is treated as a feminine noun in apposition. The name alludes to the apparent strong preference of this species to forest growing on sandstone substrates.

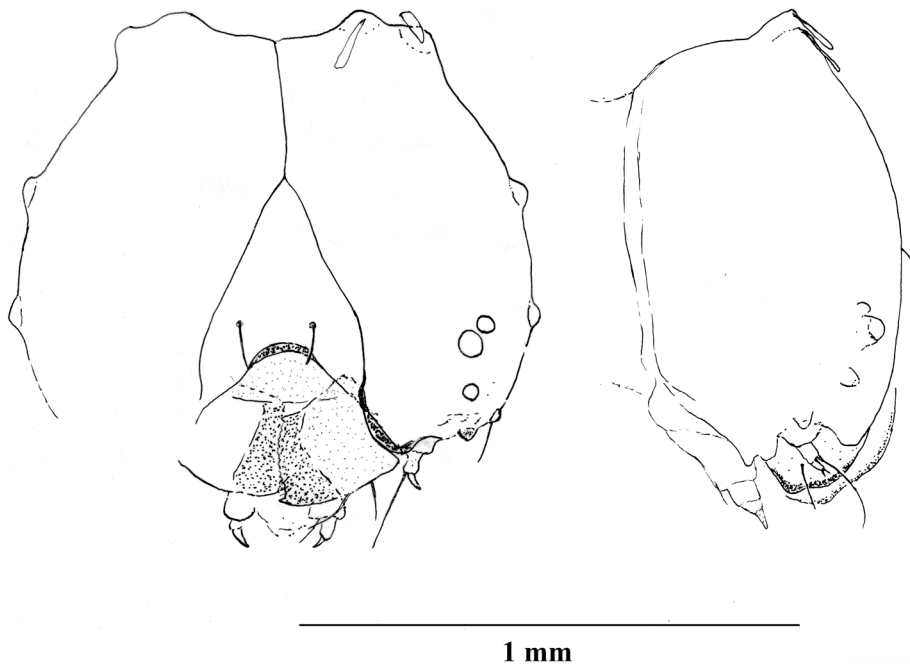
### Type material examined

#### Holotype

ECUADOR • ♂; “//ECUADOR: *Zamora-Chinchipe* Río Nangaritzza, Shaima, 4°19’S, 78°40’W 900m, xi.1997 Jiggins, C.D.// DNA voucher LEP-67941// Genitalic vial SN-17-74 S. Nakahara// Photographed by K. R. Willmott 2004// UF FLMNH MGCL-1036291//”; FLMNH, to be deposited in INABIO.

#### Paratypes (20 ♂♂, 4 ♀♀)

ECUADOR – **Zamora-Chinchipe** • 19 ♂♂; 5 km S of Zurmi, Sumak Yaku mine rd.; [4°8’38” S, 78°38’57” W]; 1000 m; 26, 28–29 Jul. 2018; J.P.W. Hall, K.R. Willmott, J.C.R. and J.I.R. Willmott leg.; FLMNH-MGCL-297322 to FLMNH-MGCL-297340; FLMNH • 1 ♀; same collection data as for preceding; FLMNH-MGCL-297321; FLMNH • 2 ♀♀; km 10 Los Encuentros-El Panguí, ridge



**Fig. 9.** *Zischkaia arenisca* Nakahara, Willmott & Hall, sp. nov., first instar head capsule in anterior and lateral views.

E San Roque; [3°42'11" S, 78°35'36" W]; 1050 m; 18–19, 23 Jul. 2018; J.P.W. Hall, K.R. Willmott, J.C.R. and J.I.R. Willmott leg.; FLMNH-MGCL-297319 to FLMNH-MGCL-297320; FLMNH • 1 ♀; same collection data as for preceding but 4 Aug. 2009; K.R. Willmott and J.P.W. Hall leg.; FLMNH-MGCL-149623; FLMNH • 2 ♂♂; Río Nangaritza, Shaime; [4°20' S, 78°40' W]; 1000 m; Sep. 1997; C.D. Jiggins leg.; FLMNH-MGCL-1036290 to FLMNH-MGCL-1036291; FLMNH.

## Description

### Male

FOREWING LENGTH. 26–27 mm (n = 2).

HEAD. Eyes naked, with grayish scales at base; frons brownish; post-genal area with lightly colored long hair-like scales and grayish scales; labial palpi with first segment covered by white and brownish long hair-like scales, in addition to some whitish scales at base; second segment almost twice as long as eye depth and covered with brown scales and hair-like scales laterally, with partially white scales and hair-like scales, dorsally with brownish long hair-like scales, ventrally with black and white hair-like scales, 3–4 times as long as segment width; third segment about one-third of second segment in length, covered with brownish scales, with small patch of creamy-white scales laterally; antennae approximately two-fifths forewing length, with ca 44 segments (n = 1), distal 16–17 segments composing club, club not prominent.

THORAX. Covered with light brownish hair-like scales.

LEGS. Foreleg brownish, foretarsus slightly shorter than tibia, femur similar to tarsus in length; midleg and hindleg with femur creamy white ventrally, tibia and tarsus grayish dorsally, ocher ventrally, tarsus and tibia spined ventrally, and a pair of tibial spurs present at distal end of tibia.

ABDOMEN. Eighth sternite divided into two sclerotized patches: a thin band at base of eighth abdominal segment, and a distal broader patch.

WING VENATION. Basal half of forewing Subcosta swollen; base of Cubitus swollen; forewing recurrent vein absent; origin of  $M_2$  nearer  $M_1$  than  $M_3$ ; hindwing humeral vein well developed.

WING SHAPE. Forewing subtriangular, apex rounded, costal margin convex, outer margin slightly convex (almost straight), inner margin straight, but rounded towards thorax near base; hindwing slightly elongate, rounded, costal margin almost straight, angled towards thorax near base, outer margin slightly undulating, inner margin slightly concave near tornus, anal lobe convex, slightly round.

DORSAL FOREWING. Ground color light brownish, distally slightly paler; black androconial scales present in middle of DFW from base to submedian area, except for immediately adjacent area around swollen Subcosta and Cubitus, in addition to area anterior to Radial about distal one-fifth of DFW; trace of submarginal band visible.

DORSAL HINDWING. Ground color similar to forewing, black androconial scales present around distal half of discal cell, extending to adjacent area, trace of submarginal band visible.

VENTRAL FOREWING. Ground color chestnut brown; submedian line invisible; dark brown narrow median line, somewhat indistinct, extending from near costa to  $Cu_2$ , slightly curved inwards, concolorous slightly sinuate submarginal line extending from apex towards tornus, but terminating half way between  $Cu_2$  and 2A; concolorous marginal line, narrower than submarginal line, extending from apex towards tornus, but terminating half way between  $Cu_2$  and 2A; fringe dark brownish.

**VENTRAL HINDWING.** Ground color similar to forewing; regular dark-brown submedian line almost straight, extending from costal to inner margin; median band almost parallel to submedian band, concolorous, similar in width, passing origin of  $M_3$  and posterior end bent inwards in 2A-3A; submarginal band extending from apex towards tornus, anterior end occasionally fused with submedian line near costa, undulating, posterior end slightly broadening and occasionally fused to submedian line in 2A-3A; marginal line, concolorous, slightly undulating along outer margin, thinner than submarginal line; submarginal ocelli from  $M_1$  to 2A, roughly ‘egg’-shaped with pointed end basal, black central spot ringed with orangish ring then with thin dark brownish indistinct ring, pupil consisting of scattered silver scales displaced distally from center, ocelli from  $M_2$ - $M_3$ ,  $M_3$ - $Cu_1$  and  $Cu_1$ - $Cu_2$  similar in size, somewhat elongate, those in  $M_1$ - $M_2$  and  $Cu_2$ -2A about half size of these three ocelli, ocellus in  $Rs$ - $M_1$  a tiny orangish spot; fringe dark brownish.

**GENITALIA** (Fig. 4E). Tegumen rounded in lateral view, elongated posterior projection of tegumen developed, slightly shorter than uncus, basal half somewhat inflated, tapering posteriorly and hooked at terminal point, posterior end of ventral margin as small projection; combination of ventral arms of tegumen and dorsal arms of saccus sinuous, bent at right angle below appendices angulares, broadens towards saccus; appendices angulares present, but somewhat reduced; saccus straight, similar to uncus in length; uncus long and narrow, with sparse hair-like setae, curved ventrally, rounded at terminal point; either side of base of uncus with short hair-like setae; brachia similar to uncus in length and width, curved in lateral view, tapering posteriorly and crossing over each other near terminal point; fultura inferior (i.e., juxta) present; valva subtriangular in lateral view, apical process triangular, ventral and dorsal margin both almost straight, scarcely covered by hairy-like setae, costa developed and triangular, dorsal margin slightly sinuous; phallobase slightly shorter than half of phallus in length, curved upwards; ductus ejaculatorius visible; aedeagus straight with manica covering approximately half, winglet present on dorsal surface, distal opening located ventrally where vesica is visible.

### **Female**

Similar to male except as follows: FW length 27 mm ( $n = 1$ ): Foreleg whitish, foretarsus divided into five distinct subsegments, fifth subsegment elongate; forewing slightly more rounded, outer margin slightly convex, ground color of both wing surfaces paler.

**FEMALE ABDOMEN AND GENITALIA** (Fig. 6E–F): inter-segmental membrane between seventh and eighth tergite not pleated, but folded posteriorly of ostium bursae with its sclerotized region forming a ‘scoop-like’ structure below lamella antevaginalis; lamella antevaginalis sclerotized; posterior approximate three-fourths of ductus bursae sclerotized, remainder membranous; posterior end as a circular hole in posterior view, with edge darker (presumably reflecting degree of sclerotization), surrounding sclerotized area somewhat semi-circular in posterior view; origin of ductus seminalis close to sclerotized portion of ductus bursae; eighth tergite with lateral side sclerotized and fused to lamella antevaginalis; corpus bursae roughly oval in dorsal view, extending across entire abdomen, with two signa located in middle, extending across almost entire corpus bursae, parallel to each other.

### **Variation**

Most individuals lack an ocellus in VHW  $Rs$ - $M_1$ , or if it is present, it is very small (much smaller than that in 2A- $Cu_2$ ). However, in one male (FLMNH-MGCL-297332) and one female (FLMNH-MGCL-297320), both barcoded (MUSM-LEP-64859 and MUSM-LEP-64857, respectively), it is rather more developed, approximately half the size of that in cell 2A- $Cu_2$ . The size of the VHW ocelli is also slightly variable, with smaller ocelli being more rounded. The ventral ground color is darker in some specimens, in which the paler brown rings surrounding the ocelli are also more conspicuously paler. The combination of the ventral arms of the tegumen and dorsal arms of the saccus is sinuous and bent

at a right angle below the appendices angulares in the male paratype, whereas it is rather straight in the holotype; the curvature of the phallobase is variable between the two dissected males.

### Distribution (Fig. 14)

This species is known to date from a highly restricted range (ca 70 km between the most northerly and southerly localities) in the Cordillera del Cóndor, an eastern outlier of the tropical Andes that lies along the border of Ecuador and Peru, and several ridges in the adjacent Río Zamora Valley in southeastern Ecuador.

### Remarks

A first instar was found in one dissected female (SN-17-174) and its head capsule is illustrated in Fig. 13. A female specimen from Alfonso Ugarte, Amazonas, Peru in MUSM may represent this species, but the sclerotized portion of ductus bursae extends about three-fourths the distance from ostium bursae to corpus bursae, which contrasts with the rather short ‘sclerotized tube’ of the two barcoded females from Zamora-Chinchipec, Ecuador (MUSM-LEP-08939 and MUSM-LEP-64858; see Fig. 1). Based on this inconsistency, a female specimen in MUSM [Alfonso Ugarte, 2–3 km N PV3; [3°54'S, 78°26'W]; 1600–1750 m; 16 Jul. 1994; G. Lamas leg.; MUSM-LEP-105658] cannot be identified reliably as *Z. arenisca* sp. nov. and is, therefore, excluded from the type series.

*Zischkaia arenisca* sp. nov. is recovered as sister to *Z. chullachaki* sp. nov. based on COI barcode data (see Fig. 1), but the genetic distance between these two taxa is atypically low for divergence between species (Table 4). Such low divergence might suggest conspecificity of these taxa, but the two species are recovered as moderately supported clades, with a number of consistent morphological differences (see above). In particular, the elongate, ‘egg’-shaped VHW ocelli and reduced, less prominent dorsal androconial scales of *Z. arenisca* sp. nov. support regarding these taxa as distinct species rather than as conspecific.

In southeastern Ecuador, this species is known from three localities, all of which support a species of climbing bamboo. Two of these sites are sandstone ‘tepui’ that have been mined for their sand, evidently because of its purity, and the poor soils on the slopes and ridges apparently support only stunted forest. An unidentified species of *Chusquea* Kunth (Poaceae: Bambusoideae) (identified by Lynn Clark) was common at one site, the ridge east of San Roque, where it grew abundantly over remnant trees and bushes along the edges of the dirt road to the mine site on the ridge top. A similar, or the same, bamboo species was likewise common along the Sumak Yaku mine road (see Fig. 15A), and was observed along the edges of forest and inside disturbed forest beside the village of Shaime, although it was absent along most other forest trails and roads in the region. Two females were collected at the edge of a large bamboo patch on the ridgetop at San Roque (see Fig. 15B), flying and resting within 2 m of the ground from 09:00 to 12:30, while two other females were flying along the edge of a dirt road near bamboo from 11:30–12:30. A number of males were observed and collected flying rapidly 1–4 m above the ground, from 10:25 to 11:20, in a sunny but sheltered corridor between two patches of vegetation covered with bamboo near a ridgetop at Sumak Yaku. The presence of multiple males and repeated flight paths suggests territorial behavior. Numerous males were also collected at the same site in traps baited with rotting fish, hung 1 m above the ground in the understorey of secondary forest with abundant bamboo. Despite a reasonable amount of effort collecting and trapping at other forest sites in the region, *Z. arenisca* sp. nov. was not recorded anywhere else, highlighting its apparent dependence on the climbing bamboo with which the adult butterflies were always closely associated.

*Zischkaia argyrosflecha* Nakahara, L. Miller & Huertas, sp. nov.  
[urn:lsid:zoobank.org:act:D67B48E9-3E2F-42BE-B6AC-01AA0EBD2C48](https://zoobank.org/act:D67B48E9-3E2F-42BE-B6AC-01AA0EBD2C48)

Figs 2Q–R, 4F, 14

*Euptychia saundersi* [sic] – D’Abrera 1988: 781, fig. [11] [misidentification].

*Euptychia mima* – D’Abrera 1988: 78, fig. [12] [misidentification].

### Diagnosis

This species is most similar phenotypically to *Z. arenisca* sp. nov., and characters that distinguish the two species are discussed in the Diagnosis of that taxon.

### Etymology

This specific epithet is based on the Greek word ‘*argyros*’, meaning ‘silver’, appended to the Spanish word ‘*flecha*’, meaning ‘arrow’, alluding to the arrow-shaped silvery markings along the basal side of the VHW ocelli. This specific epithet is considered to be a Latinized feminine noun.

### Type material examined

#### Holotype

PERU • ♂; “// [Peru], Pebas Amazonas M. de Mathan fin X<sup>bre</sup>& 1<sup>er</sup> Tr. 1880// Ex Oberthür Coll. Brit. Mus. 1927-3.// BMNH (E) 1718075//”; NHMUK.

#### Paratypes (7 ♂♂)

PERU – Loreto • 1 ♂; ‘Cavallo Cocha’ [= Caballococha]; [3°55’ S, 70°31’ W]; 90 m, May–Jul. 1884; BMNH(E)-1718078; NHMUK • 1 ♂; Iquitos; [3°45’ S, 73°15’ W]; 100 m, H. Whitely leg.; BMNH(E)-1718079; NHMUK • 3 ♂♂; Pebas; [3°19’ S, 71°51’ W]; 120 m; Dec. 1879–Mar. 1880; M. de Mathan leg.; BMNH(E)-1205410, BMNH(E)-1718076, BMNH(E)-1718077; NHMUK • 1 ♂; Río Napo ‘Ecuador’ [error]; H. Whitely leg.; BMNH(E)-1718074; NHMUK • 1 ♂; Yurimaguas; [5°54’ S, 76°6’ W]; 120 m; P. Hahnel leg.; MNHU.

### Description

#### Male

FOREWING LENGTH. 25.5–28 mm (n = 3).

HEAD. Eyes naked, frons brownish; labial palpi covered by brown and scattered white large scales, long hair-like whitish scales at base; second segment with a mixture of brown and white hair-like thin scales; third segment about one-third of second segment in length, covered with brown and white scales; antennae approximately two-thirds of FW length, with ca 44 segments (n = 1), distal 16–17 segments composing club, club not prominent and with dark ocher color at the base.

THORAX. Covered with bright iridescent brown scales.

LEGS. Foreleg brown with whitish scales, foretarsus slightly shorter than tibia, femur similar to tarsus in length; midleg and hindleg covered with whitish scales at the base, tarsus and tibia brown and a pair of tibial spurs present at distal end of tibia.

ABDOMEN. Not examined.

WING VENATION. Basal half of forewing Subcostal swollen; base of Cubitus swollen; forewing recurrent vein absent; origin of M<sub>2</sub> nearer M<sub>1</sub> than M<sub>3</sub>; hindwing humeral vein well-developed.

WING SHAPE. Forewing subtriangular and elongated towards apex, apex somewhat angular, costal margin convex, outer margin slightly convex (almost straight), inner margin straight, but rounded towards thorax near base; hindwing slightly elongate, rounded, costal margin almost straight, angled towards thorax near base, outer margin slightly undulating, inner margin slightly concave near tornus, anal lobe convex, slightly round.

DORSAL FOREWING. Ground color brownish, distally slightly paler; black androconial scales present in middle of DFW from base to submedian area, except for immediately adjacent area around swollen Subcostal and Cubitus, in addition to area anterior to Radius about distal one-fifth of DFW; trace of submarginal band invisible.

DORSAL HINDWING. Ground color similar to forewing, black androconial scales present around distal half of discal cell, extending to adjacent area, trace of submarginal band invisible.

VENTRAL FOREWING. Ground color chestnut brown; submedian line invisible; dark brown narrow median line, somewhat indistinct, extends from near costa to  $Cu_2-2A$ , bent outwards and fading after passing  $Cu_2$ , concolorous slightly sinuate submarginal line extending from apex towards tornus, terminates at 2A; concolorous marginal line, narrower than submarginal line, extending from apex towards tornus, terminates at 2A; fringe dark brownish.

VENTRAL HINDWING. Ground color similar to forewing; regular dark-brown submedian line almost straight, extending from costal to inner margin; median line almost parallel to submedian line, concolorous, similar in width, passing origin of  $M_3$ ; submarginal line extending from apex towards tornus, anterior end fused with submedian line in  $Rs-M_1$ , slightly jagged, posterior end fused to median line in 2A-3A; marginal line, concolorous, slightly undulating along outer margin, thinner than submarginal line; submarginal ocelli from  $Rs$  to 2A, roughly oval but somewhat screwed distally, pupil appear as scattered silver scales placed distally, black central spot ringed with orangish ring then with thin dark brownish indistinct ring; silverish-purple scales along basal margin and distal margin of ocelli in  $M_1-M_2$ ,  $M_3-Cu_1$  and  $Cu_1-Cu_2$ , in addition to distal side of marginal line; fringe dark brownish.

GENITALIA (Fig. 4F). Tegumen rounded in lateral view, elongated posterior projection of tegumen developed, apparently slightly shorter than uncus, tapering posteriorly and hooked at terminal point; combination of ventral arms of tegumen and dorsal arms of saccus sinuous, broadens towards saccus; appendices angulares present, but somewhat reduced; saccus straight, similar to uncus in length; fultura inferior (i.e., juxta) present; uncus long and narrow, sparsely with hair-like setae, curved ventrally, rounded and slightly inflated at terminal point, posterior end of ventral margin appearing as small projection; either side of base of uncus with hair-like setae; brachia similar to uncus in length and width, curved in lateral view, tapering posteriorly and crossing over each other near terminal point; valva subtriangular, apical process triangular, dorsal margin almost straight, ventral margin concave, scarcely covered by hair-like setae, costa developed and triangular, dorsal margin slightly sinuous; phallobase about half of phallus in length, almost straight, ductus ejaculatorius coming out higher than anterior end of coecum; aedeagus straight with manica covering approximately one-fifth, two winglets present, distal opening located ventrally where vesica is visible.

### Female

Unknown or unrecognized.

### Variation

The posterior end of the submarginal line is completely fused to the median line in 2A-3A in some specimens, whereas it is detached in others; the intensity of the silvery-purple scales around the VHW ocelli and distal margin of the marginal line is variable.

### Distribution (Fig. 14)

This species is known from the western Amazon basin in northern Peru.

### Remarks

Although we do not have DNA data for this taxon, morphological differences between this species and *Z. arenisca* sp. nov. documented above support its specific status.

### *Zischkaia pacarus* (Godart, [1824])

Figs 1, 2S–V, 4G, 6G–H, 8A, 10A–C, 11, 12A–D, 14

*Satyrus pacarus* Godart, [1824]: 465, 495 (type locality: Brazil).

*Euptychia fumata* Butler, 1867b: 109, pl. 12, fig. 14 (type locality: “Rio Grande”).

*Neonympha pacarus* – Westwood 1851: 375.

*Euptychia pacarus* – Butler 1867a: 501; 1868: 38; 1877: 120. — Kirby 1871: 55. — Gaede 1931: 458. — Weymer 1911: 213. — D’Abrera 1988: 789.

*Euptychia fumata* – Butler 1867a: 501, nomen nudum.

*Euptychia fumata* – Butler 1868: 38; 1877: 120. — Kirby 1871: 55; 1879: 134. — Weymer 1911: 213. — Riley & Gabriel 1924: 22. — Gaede 1931: 446. — Miller 1968: 95. — D’Abrera 1988: 789.

*Zischkaia fumata* – Forster 1964: 116–118, fig. 123 (the description date of the name was mistakenly given as “1857”). — Brown 1992: 152, fig. 49. — Mielke & Casagrande 1998: 976. — Brown & Freitas 2000: 105.

*Zischkaia pacarus* – Mielke 1995: 772. — Lamas 2004: 223, syn.: *fumata* Butler, 1867. — Emery *et al.* 2006: 90. — Beccaloni *et al.* 2008: 346. — Carneiro *et al.* 2008: 264. — Peña *et al.* 2010: 248, 250–251, 253. — Dolibaina *et al.* 2011: 350. — Silva *et al.* 2015: 6.

### Type material examined

#### Neotype (here designated)

BRAZIL • ♂; “//BRASIL, PARANÁ, TUNEIRAS DO OESTE, REBIO [Reserva Biológica] DAS PEROBAS, 23° 50’ 49”S 52° 44’ 18”W, 8-X-2012 LABELP [Laboratório de Estudos de Lepidoptera Neotropical, Departamento de Zoologia, Universidade Federal do Paraná] LEG.// BC-DZ Willmott 246// DZ 36.998//”; DZUP.

#### Lectotype (here designated)

BRAZIL • ♂; “(//Rio Grande [do Sul, Brazil] Hewitson coll[ection]. 79-69 Euptychia Fumata, Butl[er]. 2.// BM TYPE No. Rh 3251 Euptychia fumata ♂ Butl[er].// Type// BMNH(E) 1267093//”; NHMUK.

#### Other material examined (87 ♂♂, 50 ♀♀)

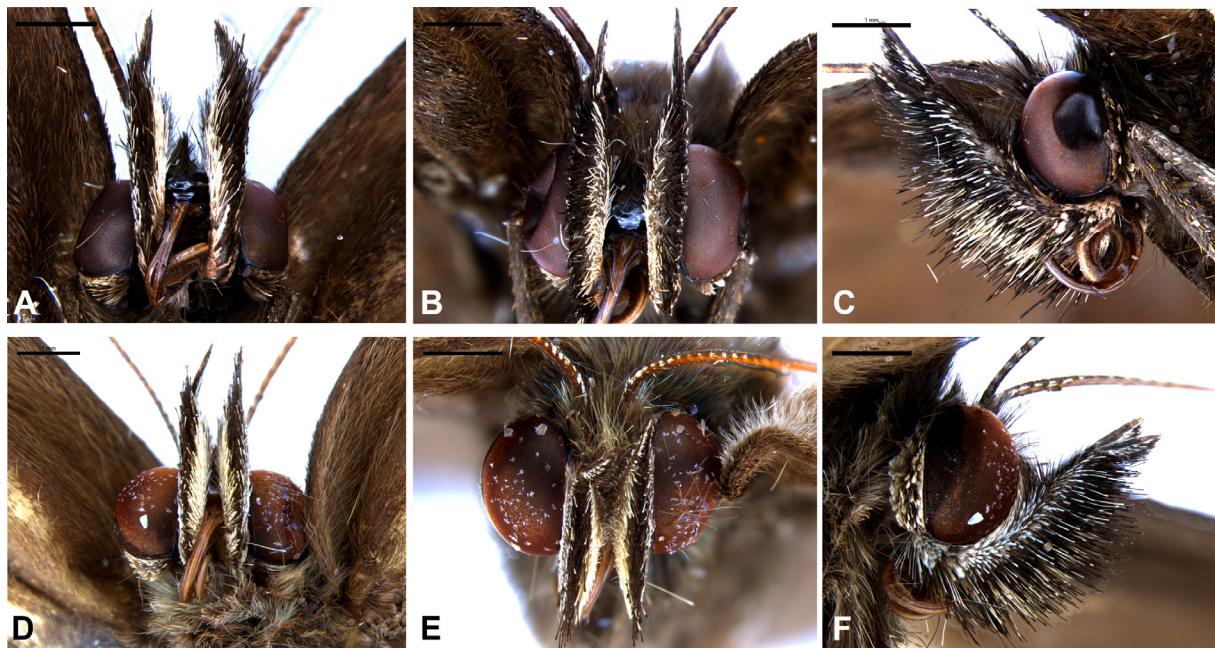
ARGENTINA–Misiones • 2 ♂♂; Apr. 1927; I. Heider leg.; BMNH(E)-10247985, BMNH(E)-10430700; NHMUK • 1 ♀; El Dorado; [26°23’ S, 54°40’ W]; ZSM • 1 ♂; Loreto; [27°19’ S, 55°32’ W]; Oct. 1956; W. Forster leg.; ZSM.

BOLIVIA – **Not located** • 1 ♂; “Torochila” [perhaps representing “Jorochito”]; Garlepp leg.; 1890; MNHU.

BRAZIL – **Bahia** • 1 ♂; Una, Reserva Biológica de Una; [15°10'38" S, 39°6'20" W]; G.M. Accácio leg.; ZUEC. – **Espírito Santo** • 1 ♂; Fruhstorfer leg.; BMNH(E)-1205412; NHMUK. – **Minas Gerais** • 1 ♀; S of Minas Gerais; BMNH(E)-1718052; NHMUK • 2 ♀♀; Carangola, Serra Rio Glória, Sítio Boa Vista; [20°43'44" S, 42°3'27" W]; 600–800 m; Dec. 1950; Ebert leg.; DZ-36529, DZ-36549; DZUP • 4 ♂♂; Porto Real; [20°11' S, 45°43' W]; BMNH(E)-1718043 to BMNH(E)-1718045, BMNH(E)-1718056; NHMUK • 2 ♀♀; same collection data as for preceding; BMNH(E)-1718050 to BMNH(E)-1718051; NHMUK • 1 ♂; San [São] Jacinto; [17°51' S, 41°30' W]; Nov. 1907; F. Birch leg.; BMNH(E)-1718042; NHMUK. – **Paraná** • 1 ♂; BMNH(E)-1205411; NHMUK • 1 ♀; BMNH(E)-1718062; NHMUK • 1 ♂; Balsa Nova, São Luís do Purunã; 1000 m; 8 Apr. 2006; Beltrami and Selusniaki leg.; DZ-36643; DZUP • 1 ♂; Campo Mourão, (Justus); Jan. 1952; DZ-36861; DZUP • 5 ♂♂; Candói, Santa Clara; [25°37' S, 51°59' W]; 21 Nov. 1986; O.H.H. Mielke and M.M. Casagrande leg.; DZ-36589, DZ-36633, DZ-36831, DZ-36891, DZ-5575; DZUP • 2 ♀♀; same collection data as for preceding; DZ-36851, DZ-36991; DZUP • 1 ♂, 1 ♀; Castro; [24°47' S, 50°1' W]; 950 m; E.D. Jones leg.; BMNH(E)-1718058, BMNH(E)-1718055; NHMUK • 1 ♂; same collection data as for preceding but collector unknown; BMNH(E)-1718041; NHMUK • 2 ♂♂; Chopinzinho; [25°51' S, 52°30' W]; 1 Mar. 1970; E. Furtado leg.; DZ-36971, DZ-37031; DZUP • 1 ♂; Foz do Iguaçu; [25°30' S, 53°48' W]; Mar. 1952; Justus leg.; DZ-37021; DZUP • 1 ♂; Iguazu; 24 Jan. 1922; BMNH(E)-1718057; NHMUK • 1 ♀; Londrina; [23°19' S, 51°10' W]; 25 Oct. 1985; O.H.H. Mielke and M.M. Casagrande leg.; DZ-37041; DZUP • 1 ♀; Ponta Grossa; [25°6' S, 50°10' W]; Nov. 1956; Justus leg.; DZ-36821; DZUP • 1 ♀; Vila Velha; [25°8' S, 49°58' W]; 900 m; 16 Mar. 1972; Mielke leg.; DZ-36841; DZUP • 1 ♀; 7 Mar. 1973; DZ-36981; DZUP • 1 ♀; Lapa, Rio de Várzea; [25°46' S, 49°43' W]; Feb. 1938; Pohl leg.; MZSP • 1 ♀; Tibagi, Parque Estadual do Guartelá, [24°33'59" S, 50°15'25" W]; 1000 m; 11–12 Feb. 2016; D. Dolibaina, E. Carneiro and A. Warren leg.; DD-374 • 6 ♂♂, 4 ♀♀; Tuneiras do Oeste, Reserva Biológica das Perobas; [23°50'49" S, 52°44'18" W]; (‘LELN UFPR’); 6 Oct. 2012; DZ-36498, DZ-36728, DZ-36858, DZ-36918, DZ-36939, DZ-36949, DZ-36778, DZ-36909, DZ-36919, DZ-36959; DZUP • 4 ♂♂, 2 ♀♀; same collection data as for preceding but 8 Oct. 2012; DZ-36708, DZ-36929, DZ-36989, DZ-37039, DZ-36588, DZ-36999; DZUP • 1 ♂; [23°53' S, 52°44' W]; 578 m; 6 Oct. 2012; G. Lamas leg.; MUSM • 1 ♂; same collection data as for preceding but 8 Oct. 2012; MUSM • 1 ♂; Turvo, Britador; [25°2'11" S, 51°32'8" W]; 1000 m; 19 Feb. 2010; D. Dolibaina and A. Warren leg.; DD-303 • 1 ♂, 1 ♀; Turvo, Salto do Paulinho Rickli; [25°3'59" S, 51°31'8" W]; 1050 m; 15 Mar. 2009; D. Dolibaina leg.; DD-301 to DD-302 • 4 ♂♂, 1 ♀; same collection data as for preceding but 7 Feb. 2016; D. Dolibaina and Warren leg.; DD-369 to DD-373. – **Rio Grande do Sul** • 1 ♀; 1890; Mabilde leg.; MNHU • 1 ♀; Elsenau [Panambi]; Dec. 1904; ZSM • 3 ♂♂, 2 ♀♀; Campo Novo; 9 Nov. 1985; Araújo, O.H.H. Mielke and M.M. Casagrande leg.; DZ-36781, DZ-36941, DZ-36951, DZ-36741, DZ-36751; DZUP • 2 ♀♀; Derrubadas, Parque Florestal Estadual do Turvo; [27°13'58" S, 53°51'5" W]; 10 Nov. 1985; O.H.H. Mielke, Araújo, and M.M. Casagrande leg.; DZ-36761, DZ-36811; DZUP • 1 ♀; Pelotas; [31°45' S, 52°22' W]; 15 Oct. 1953; C.M. Biezanko leg.; DZ-36931; DZUP • 1 ♂; 28 Feb. 1961; AMNH • 1 ♂, 2 ♀♀; 11 Mar. 1961; USNM • 1 ♂; 25 Nov. 1961; AMNH • 1 ♂; 12 Dec. 1961; USNM • 1 ♀; 22 Mar. 1964; AMNH • 1 ♂; 13 Feb. 1966; V. Becker leg.; DZ-36801; DZUP • 1 ♀; 6 Nov. 1961; C.M. Biezanko leg.; ZSM • 1 ♂; 7 Nov. 1966; ZSM • 1 ♂; 7 Nov. 1966; DZ-36971; DZUP • 1 ♂; 3 Mar. 1967; DZ-36791; DZUP • 1 ♂; 7 Mar. 1970; DZ-21207; DZUP. – **Santa Catarina** • 1 ♂; Wernicke leg.; ZSM • 1 ♂; Rio Capivary [Capivari]; 1883; Fruhstorfer leg.; BMNH(E)-1718059; NHMUK • 1 ♀; ridge between Joinville and Itajaí, Tajú; [26°34'26" S, 48°44'51" W]; Apr.; Schmid leg.; DZ-36881; DZUP • 1 ♂; same collection data as for preceding but Feb.; DZ-37001; DZUP • 1 ♂; Águas Mornas, Teresópolis; [27°44' S, 48°56' W]; BMNH(E)-1718048; NHMUK • 1 ♀; same collection data as for preceding; Michalis leg.; BMNH(E)-1718063; NHMUK • 1 ♂; Blumenau; [26°55' S, 49°4' W]; Fruhstorfer leg.;



BMNH(E)-1205413; NHMUK • 1 ♀; Florianópolis, Naufragados; 11 May 2003; E. Carneiro leg.; DZ-37011; DZUP • 5 ♂♂, 1 ♀; Seara, Nova Teutonia; [27°3' S, 52°23' W]; 300–500 m; Feb. 1973; F. Plaumann leg.; FLMNH-MGCL-1036000 to FLMNH-MGCL-1036005; FLMNH • 1 ♂; 1 Feb. 1972; F. Plaumann and O.H.H. Mielke leg.; DZ-36658; DZUP • 1 ♂; Taió, Rio Taió; [27°6' S, 49°59' W]; 2 Oct. 1935; Pohl leg.; MZSP • 1 ♀; Timbó; [26°49' S, 49°17' W]; Apr. 1936; Pohl leg.; MZSP. – **São Paulo** • 1 ♀; Araras; [22°19' S, 47°58' W]; 600 m; 17 Apr. 1966; Ebert leg.; DZ-36519; DZUP • 1 ♂, 1 ♀; same collection data as for preceding but 6 Jan. 1966; DZ-36499, DZ-5577; DZUP • 1 ♂; Casa Branca; [21°47' S, 47°5' W]; G[arbe] leg.; MNHU • 1 ♂ [conf]; 1893–1895; Garbe leg.; MNHU • 1 ♂; Cotia, Morro Grande; 22 Nov. 2000; K.S. Brown and A.V.L. Freitas leg.; ZUEC • 1 ♂; Guararapes, Figueira; 5 Apr. 1940; R.F. d'Almeida leg.; DZ-36509; DZUP • 1 ♀; Paranapiacaba, Alto da Serra; Feb. 1922; Spitz leg.; MZUSP-56158; MZSP • 1 ♂; Rio Claro, [22°24' S, 47°33' W]; 600 m; 13 Apr. 1967; Ebert leg.; DZ-36911; DZUP • 1 ♀; same collection data as for preceding but 26 May 1963; DZ-5576; DZUP • 1 ♂, 1 ♀; same collection data as for preceding but 31 Dec. 1966; DZ-5578, DZ-36559; DZUP • 1 ♂; Rio das Cobras; [23°7'13" S, 46°5'44" W]; Feb. 1942; Widerski leg.; DZ-36539; DZUP • 1 ♀; Salesópolis, Estação Biológica da Boracéia; [23°40' S, 45°53' W]; 850 m; 20 Mar. 1968; Lopes leg.; MZSP • 1 ♂; same collection data as for preceding but 27 Feb. 1968; Oliveira Santos leg.; MZSP • 1 ♂; São Paulo; [23°32' S, 46°37' W]; 790 m; BMNH(E)-1718047; NHMUK • 3 ♂♂, 1 ♀; same collection data as for preceding; Garbe leg.; MZUSP-56159 to MZUSP-56162; MZSP • 1 ♂, 1 ♀; Teodoro Sampaio, Parque Estadual Morro do Diabo; [22°30' S, 52°20' W]; 250–500 m; 15–18 Mar. 1990; O.H.H. Mielke and M.M. Casagrande leg.; DZ-36569, DZ-36579; DZUP • 1 ♂; same collection data as for preceding but 15–18 Mar. 1991; DZ-36901; DZUP • 1 ♂, 1 ♀; same collection data as for preceding but 20–23 Feb. 1990; DZ-36921, DZ-36613; DZUP • 1 ♂; same collection data as for preceding but 22–23 May 1990; DZ-36771; DZUP. – No specific locality • 1 ♂; H. Rogers leg.; *PLT fumata*; BMNH(E)-1718068; NHMUK – **Not located** • 1 ♂; 'Brazil'; BMNH(E)-1718046; NHMUK • 1 ♂; 'Brazil'; *PLT fumata*; BMNH(E)-1718067; NHMUK • 1 ♀; 'Brazil'; *PLT fumata*; BMNH(E)-1718066; NHMUK.



**Fig. 10.** Head morphology. **A–C.** *Zischkaia pacarus* (Godart, 1824) (DZ 36.728). **A.** Ventral view. **B.** Frontal view. **C.** Lateral view. **D–F.** *Z. saundersii* (Butler, 1867) (DZ 36.029). **D.** Ventral view. **E.** Frontal view. **F.** Lateral view.

PARAGUAY – Caaguazú • 1 ♂; Caaguazú; [25°26' S, 56°2' W]; Apr. 1960; ZSM.

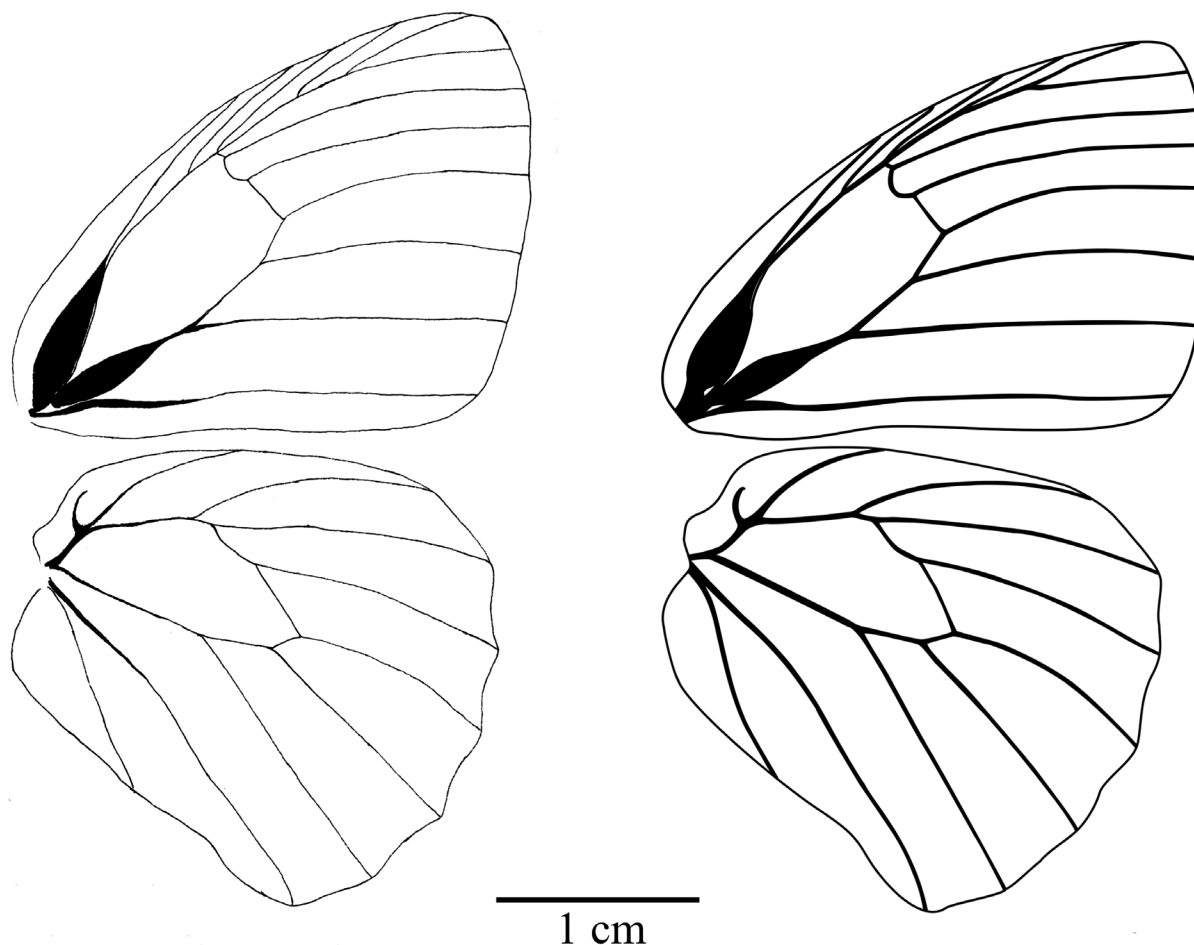
VENEZUELA – Not located • 1 ♂; ‘Venezuela’, BMNH(E)-1718065; NHMUK.

COUNTRY UNKNOWN – Not located • 1 ♂; no data; BMNH(E)-1718049; NHMUK.

### Identification and taxonomy

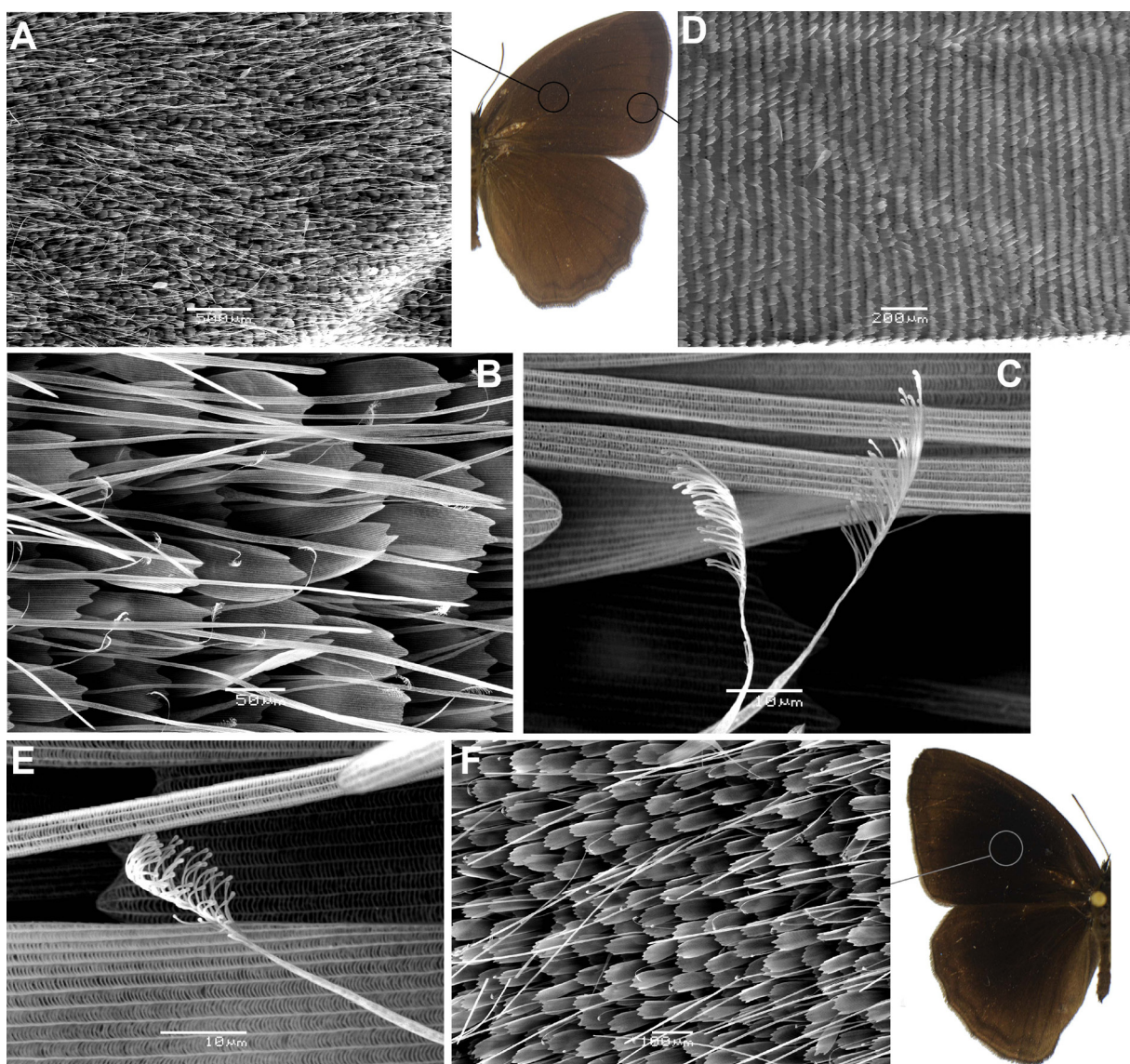
The male of *Zischkaia pacarus* is distinguishable from males of other species of *Zischkaia* by the apical process of the valva being somewhat rectangular with a convex distal margin (Fig. 4G), whereas the apical process of the valva is somewhat triangular in other species of *Zischkaia*. The female of *Z. pacarus* can also be distinguished by the genitalia (Fig. 6G–H), namely by the sclerotized tube being one-fourth the length of the ductus bursae, whereas this sclerotized tube is reduced and restricted to the area around the ostium bursae in *Z. chullachaki* sp. nov., more developed and reaching almost half the length of the ductus bursae in *Z. baku* sp. nov. and *Z. arenisca* sp. nov.

Godart ([1824]: 495) described *Satyris pacarus* based on a single specimen from Brazil, without any mention of the sex of this specimen. He provided a short description of the species (Godart ([1824]: 465), stating “[margin of the] wings entire, dark brown, upper side without spots; forewing under side with two darker wavy lines, hind wing under side with three [darker wavy lines], and five eyes [i.e., ocelli]



**Fig. 11.** *Zischkaia pacarus* (Godart, 1824), wing venation. A. ♂ (FLMHN-MGCL-1036002). B. ♀ (MGCL-LOAN 518).

having the pupil formed by silver dots.” The full description on page 495 is lengthier and more precise: “It has approximately two inches [about 5 cm] of wingspan. Its wings [margins] are entire, and dark brown in both sides. Its [wings] upper sides are without spots. Its [wings] under sides with, towards the extremity, two lines darker than the ground color. The second wings [i.e., hindwings] have a third similar line, placed transversely towards the middle, and separated from the others by a row of five round, very dark, eyes [i.e., ocelli], with a pupil formed by silver dots, and a brownish iris surrounded by a very dark circle. The first wings [forewings] have no eyes [i.e., ocelli].” (translation based on the original description of both French and Latin version). Certain phenotypic characters described by Godart, namely, no mention of the VFW ocellus (or ocelli), no ocellus (or ocelli) on the dorsal surface, the mention of two lines on the VFW and three lines on the VHW (i.e., lack of VFW submedian line), and the fact this species was allied with several other euptychiine species (e.g., *Hermeuptychia sosybius*



**Fig. 12.** Wing scales of *Zischkaia* Forster, 1964. **A–C.** Images of forewing androconial scales of *Z. pacarus* (Godart, [1824]), showing the presence of plumose androconial scales. **D.** Typical wing scales of *Z. pacarus*. **E–F.** Images of forewing androconial scales of *Z. chullachaki* Nakahara & Zacca, sp. nov., showing the presence of plumose androconial scales.

(Fabricius, 1793)), suggest that Godart was describing a species of *Zischkaia*. There is no mention of dorsal surface androconia in Godart's description, one of the putative synapomorphic characters for the “*pacarus* clade” of *Zischkaia*, and we offer two possible explanations: 1) although the specimen he examined was a male, Godart failed to perceive or did not regard as important the presence of this character; 2) the specimen examined by Godart was a female, and therefore without androconia. Given that the dorsal androconia are not well marked in *Z. pacarus*, and considering that females are rarer than males, the former is perhaps the more likely. For example, Godart also did not describe the androconial patch of *Calisto hysius hysius* (Godart, [1824]) (described on page 471) and *Forsterinaria necys* (Godart, [1824]) (described on page 511), although males of these two species also possess dorsal wing androconial patches. There remains a possibility that Godart was describing *Z. warreni* sp. nov., which also lacks a VFW submedian line and occurs in sympatry with *Z. pacarus*. However, *Z. warreni* sp. nov. has a much narrower geographic range and is much scarcer than *Z. pacarus* in museum collections (43 specimens examined of the former species and 138 of the latter). Most likely, the holotype (fixed by monotypy, following Article 73.1.2. of the ICZN (1999)) of *pacarus* should have been collected in Minas Gerais or Rio de Janeiro, where *Z. warreni* sp. nov. is apparently rare (see below). Furthermore, at least some of the species described by Godart are not known or are very rare within the range of *Z. warreni* sp. nov. (e.g., *Godartiana byses* (Godart, [1824]), see Zacca *et al.* (2017), *Adelpha epione* (Godart, [1824]), see Willmott (2003)), suggesting that the type of *Z. pacarus* might have come from further east in southern Brazil. We therefore contend that Godart's description most likely referred to *Z. pacarus* as treated here. The holotype of *S. pacarus* has not been located in any museums where specimens from Godart's collection are usually found (e.g., MNHN; Royal Scottish Museum, Edinburgh (Grimshaw 1897; Horn *et al.* 1990; Bland 2019)). Therefore, a male specimen deposited at the DZUP (Fig. 2), which we also barcoded and included in the phylogeny (BC-DZ 246), is designated as the neotype of *Satyryus pacarus* Godart, [1824] in order to settle the nomenclature (neotype designation).

Butler (1867a) introduced the name *Euptychia fumata*, but in a way that failed to fulfill Article 12.1 of the ICZN (1999), as the name was not accompanied by a description, definition, or indication of the taxon it denotes. Therefore, the name was correctly recognized as a nomen nudum by Lamas (2004). Subsequently, Butler (1867b) properly described *Euptychia fumata* Butler, 1867 based on an unknown number of specimens from “Rio Grande” (likely the state of Rio Grande do Sul, southern Brazil) from Hewitson's collection. Given the presence of other similar *Zischkaia* species in southern Brazil, a male syntype, deposited in the NHMUK, which compares closely to the description and illustration provided by Butler (1867b), is here designated lectotype, in order to settle the nomenclature (lectotype designation). *Euptychia fumata* Butler, 1867 was treated as a junior subjective synonym of *Z. pacarus* by Lamas (2004). Although Lamas (2004) did not provide any justification for this synonymy, the lectotype of *E. fumata*, here designated, compares closely with Godart's description of *S. pacarus* and the neotype male designated above; therefore, we retain *Euptychia fumata* Butler, 1867 as a junior subjective synonym of *Z. pacarus*.

### Variation

The conspicuous dorsal androconia is variable, from being slightly to moderately marked; the VHW ocellus in Rs-M<sub>1</sub> is variable in terms of presence and development; the visibility of the bright silvery purple ground color between the VHW submedian line and submarginal line when seen under direct light is also apparently variable, from absent to present.

### Distribution (Fig. 14)

This species is known from southwestern and southern Brazil to northeastern Argentina, in the states of Minas Gerais, São Paulo, Paraná, Santa Catarina, Rio Grande do Sul (Brazil), in the department of Caaguazú (Paraguay) and the province of Misiones (Argentina). Old records from Amazon Superior, Brazil, Bolívia and Venezuela probably refer to mislabeled specimens.

## Remarks

In *Araucaria* forests in Paraná, adults of *Z. pacarus* are found during the summer either on small and isolated or large and continuous patches of the bamboo *Merostachys* Spreng. (Poaceae: Bambusoideae) (Fig. 16A, C). Males fly fast, up to 5 m above the ground, with several individuals engaged in patrolling flight. Females fly slowly and lower than males. Butterflies are never found away from the bamboo, and they are more active between the middle of the morning and the beginning of the afternoon. The preferred habitat of *Z. pacarus* in *Araucaria* forests suggests that the species likely uses *Merostachys* as the caterpillar host plant. In addition, according to A.V.L Freitas (pers. comm. in Beccaloni *et al.* 2008), larvae feed on *Bambusa* (Poaceae: Bambusoideae).

*Zischkaia abanico* Nakahara & Petit, sp. nov.

[urn:lsid:zoobank.org:act:ECB82262-F5E0-4C0A-9FDB-468065263D73](https://zoobank.org/act:ECB82262-F5E0-4C0A-9FDB-468065263D73)

Figs 1, 2W–X, 4H, 14, 15C

## Diagnosis

*Zischkaia abanico* sp. nov. is readily distinguishable from other species in the “*pacarus* clade” by the shape of the apical portion of the valva, which is somewhat rectangular with a convex (i.e., projecting) distal margin in lateral view, whereas the apical process of the valva is more triangular in other species of *Zischkaia*, except for *Z. pacarus*. *Zischkaia abanico* sp. nov. and *Z. pacarus* are in fact similar in terms of the shape of the valva, as well as other external characters, including wing pattern, and these two taxa cannot be distinguished externally, although our DNA ‘barcodes’ data clearly suggest they are not conspecific (see Fig. 1). In fact, these two taxa can be distinguished by their range, with *Z. abanico* sp. nov. known from the slopes of the Andes from Colombia to Ecuador and *Z. pacarus* known from southern and southeastern Brazil to northeastern Argentina. We do not know of any putative female specimen for this species.

## Etymology

The specific epithet is based on the name of the river, Río Abanico, which flows below the ridge where the holotype was captured; the site is thus referred to as the ‘Abanico ridge’ among lepidopterists. This specific epithet is considered as a latinized masculine noun in apposition.

## Type material examined

### Holotype

ECUADOR • ♂; “//11926 Euptychia sp. 2002, 1213 Ecuador 9 de Octubre, M-Sgo [Morona-Santiago], 1600m JCP// DNA voucher LEP-37388// prep. genit. 575 19.07.2017/J.Lorenc *Zischkaia* sp Nueve do Octubre Ecuador//”; JEPE, to be deposited in MZUJ.

### Other material examined (2 ♂♂)

COLOMBIA – Cauca • 2 ♂♂; Popayán; [2°27' N, 76°36' W]; Lehmann leg.; BMNH(E)-10430710 to BMNH(E)-10430711; NHMUK.

## Description

### Male

FOREWING LENGTH. 23 mm (n = 1).

HEAD. Eyes naked, with grayish scales at base. Frons brownish. Post-genal area with lightly colored long hair-like scales and grayish scales. Labial palpi with first segment mostly with brownish long hair-like scales; second segment length almost twice as great as eye depth and adorned with brown scales laterally, dorsally mostly with light brownish long hair-like scales, ventrally adorned with brownish and

white hair-like scales, about 3-4 times as long as segment width; third segment about two-fifth of second segment in length and covered with brownish scales, ventrally with brownish hair-like scales, with slight patch of creamy-white scales laterally. Antennae approximately two-fifth of forewing length, with ca 39 segments ( $n = 1$ ), distal 13–15 segments composing club, club not prominent.

THORAX. Dorsally, laterally and ventrally scattered with grayish scales with long multi-colored hair-like scales.

LEGS. Foreleg brownish, foretarsus slightly shorter than tibia, femur similar to tarsus in length; midleg and hindleg with femur creamy white ventrally, tibia and tarsus grayish dorsally, whitish ventrally, tarsus and tibia spined ventrally, and a pair of tibial spurs present at distal end of tibia.

ABDOMEN. Eighth tergite as stripe at base of eighth abdominal segment, in addition to presence of distal broader patch.

WING VENATION. Most of forewing Subcostal swollen; base of Cubitus swollen; forewing recurrent vein absent; origin of  $M_2$  towards  $M_1$  than  $M_3$ . Hindwing Humeral developed.

WING SHAPE. Forewing subtriangular, apex rounded, costal margin convex, outer margin slightly convex (almost straight, but see also below), inner margin straight, but rounded towards thorax near base; hindwing slightly elongate, rounded, costal margin almost straight, angled towards thorax near base, outer margin slightly undulating, inner margin slightly concave near tornus, anal lobe convex, slightly round.

DORSAL FOREWING. Ground color light brownish, distally slightly paler; black androconial scales, not prominent, present in middle of DFW, from base to submedian area; trace of submarginal and marginal band almost invisible.

DORSAL HINDWING. Ground color similar to forewing, black androconial scales present in discal cell and adjacent area, not as prominent as DFW; trace of submarginal band almost invisible (clearly absent in one specimen).

VENTRAL FOREWING. Ground color light chestnut brown; submedian line invisible; dark brown narrow median line extends from near costa to  $Cu_2-2A$ , in slightly outward diagonal direction, concolorous slightly sinuate submarginal line extending from apex towards tornus, but terminates around 2A; concolorous marginal line, narrower than submarginal line, extending from apex towards tornus, but terminates around 2A; fringe dark brownish.

VENTRAL HINDWING. Ground color similar to forewing; regular dark-brown submedian line almost straight, extending from costal to inner margin, passing origin of  $M_1$ ; median line almost parallel to submedian line, concolorous, similar in width, passing origin of  $M_3$ , and posterior end bent inwards in 2A-3A; submarginal line extending from apex towards tornus, undulating, posterior end slightly broadening and apparently fused to submedian line in 2A-3A; marginal line, concolorous, slightly undulating along outer margin, thinner than submarginal line; submarginal ocelli from Rs to 2A (ocellus in Rs- $M_1$  absent in one specimen), rounded, pupil appear as scattered silverish scales placed rather distally, black central spot ringed with orangish ring then with thin dark brownish indistinct ring, ocellus in Rs- $M_1$  smallest, if present; bright silvery purple ground color visible between submedian line and submarginal line when seen under light; fringe dark brownish.

GENITALIA (Fig. 4H). Tegumen rounded in lateral view, elongated posterior projection of tegumen developed, apparently slightly shorter than uncus, tapering posteriorly and hooked at terminal point;

combination of ventral arms of tegumen and dorsal arms of saccus sinuous, broadens towards saccus; appendices angulares present, but somewhat reduced; saccus anterior half slightly curved upwards, similar to uncus in length; uncus long and narrow, sparsely with hair-like setae, curved ventrally, rounded at terminal point, posterior end of ventral margin appearing as small projection; either side of base of uncus with hair-like setae; brachia similar to uncus in length, slightly narrower, curved in lateral view, tapering posteriorly and crossing over each other near terminal point; fultura inferior present; valva subtriangular in lateral view, apical process subtriangular, somewhat pointy distal end, scarcely covered by hairy-like setae; costa developed and triangular, dorsal margin slightly sinuous; phallobase slightly shorter than phallus in length, curved; ductus ejaculatorius not examined; aedeagus straight with manica not examined, winglet present, distal opening located ventrally where vesica is visible.

### Female

Unknown or unrecognized.

### Variation

The FW outer margin is more curved in the holotype, whereas it is straighter in the two examined specimens from Colombia; the VHW median line is wavy below the origin of  $M_3$  in the holotype, whereas it is rather straight in the two Colombian specimens.

### Distribution (Fig. 14)

This species is known from two localities only, the eastern Andes of Ecuador (Morona-Santiago) (Fig. 15C) and the Cauca valley in Colombia (Cauca). However, there is some doubt about the reliability of the latter locality, as discussed further below.

### Remarks

We excluded from the type series the two male specimens from Popayán, Colombia, listed above, despite the male genitalia of the dissected specimen (BMNH(E)-10430711) being identical to that of the holotype male, in particular in the apical portion of the valva being somewhat rectangular with a convex (i.e., projecting) distal margin in lateral view. However, given the absence of other specimens of this evidently rare species from Colombia or other localities in northern Ecuador, and the otherwise low overlap in the butterfly fauna between Morona-Santiago and the Cauca valley, we considered it reasonably likely that the Colombian specimens might have been mislabeled. In particular, there are several mislabeled specimens of Ithomiini (Nymphalidae) in the NHMUK also labeled as having been collected by Lehmann in Popayán, Colombia, including *Dircenna loreta loreta* Haensch, 1903 (a lowland west Amazonian taxon, 1 ♂), *Melinaea marsaeus mothone* (Hewitson, 1860) (an east Andean foothill taxon, 1 ♂) and *Hypothyris ninonia daeta* (Boisduval, 1836) (Southeastern Brazil, 2 ♂♂). Notably, the last mentioned taxon occurs within the range of *Z. pacarus*, so it is possible that the two Colombian specimens are actually mislabeled *Z. pacarus*. Thus, although Popayán is a somewhat plausible locality for this species, and certainly this taxon should be searched for in Colombia, we decided to treat the presence of this species in Colombia as requiring confirmation.

### *Zischkaia saundersii* (Butler, 1867)

Figs 1, 3A–D, 5A, 7A–B, 8B, 10D–F, 13–14

*Euptychia saundersii* Butler, 1867a: 500, pl. XL, fig. 17 (type locality: Ega [= Tefé]).

*Euptychia saundersii* – Butler 1868: 38; 1877: 120. — Kirby 1871: 55; 1879: 132. — Weymer 1911: 213. — Riley & Gabriel 1924: 52. — Gaede 1931: 464. — Forster 1964: 117.

*Euptychia saundersi* [sic] – D’Abrera 1988: 781.

*Zischkaia saundersii* – Robbins *et al.* 1996: 232. — Lamas & Grados [1997]: 58. — Lamas 2004: 223.

### Type material examined

#### Lectotype (here designated)

BRAZIL • ♀; “//Ega [Tefê, Amazonas, Brazil], U[pper]. Amazon[a]s [River]. H[enry]. W[alter]. Bates.// Godman-Salvin Coll[ection]. 1904. – 1. *Euptychia saundersi* [sic], Butl[er].// *Euptychia saundersii* Butler Monog[raph]./type// SYN-TYPE// saundersii/ ♀ Ega// BMNH(E) 1718073//”; NHMUK.

#### Paralectotypes

BRAZIL • 1 ♂; “//B.M. TYPE No. Rh. 3248 *Euptychia saundersi* [sic] ♂ Butl[er].// Pernambuco. [Brazil]. Pres[ented]. by Mrs Smith 45 – 70// *Euptychia saundersii* Butler Monog[raph].// Type/ *Euptychia saundersii* Butler ♂//SYN-TYPE// Pernambuco[, Brazil]/ 45 70// BMNH(E): 1267094// NHMUK 010247986//”; NHMUK • 1 ♀; “//Pernambuco[, Brazil]. Pres[ented]. By Mrs Smith 45 – 70// Pernambuco/ 45 70// BMNH(E) 1718070//”; NHMUK.

### Other material examined (13 ♂♂, 4 ♀♀)

BRAZIL – **Maranhão** • 1 ♂; Feira Nova do Maranhão, 26 km E of Fazenda Forquilha dos Brejos; [6°59’12.7”S 46°25’47.4”W]; 13–16 Apr. 2011; Mielke & Casagrande leg.; DZ-38159; DZUP. – **Pará** • 1 ♂, 1 ♀; Santarém; [2°26’ S, 54°43’ W]; Oct. 1884; H.H. Smith leg.; BMNH(E)-1718071 to BMNH(E)-1718072; NHMUK. – **Pernambuco** • 1 ♂; PLT *saundersii*; BMNH(E)-1267094; NHMUK. – **Rondônia** • 1 ♂; Porto Velho, Caiçara; 150 m; 10 Sep. 2012; ZUEC • 1 ♂; same collection data as for preceding but 8 Oct. 2013; ZUEC. – **Not located** • 2 ♂♂; ‘Brazil’; ZSM.

PERU – **Huánuco** • 1 ♂; Cordillera del Sira; [9°25’ S, 74°45’ W]; 1380 m; Sep. 1987–Aug. 1988; Exp. Universidad Viena leg.; MUSM-LEP-105654; MUSM. – **Madre de Dios** • 1 ♀; Parque Manu, Pakitza; [11°55’48” S, 71°15’18” W]; 340 m; 5 Oct. 1991; M.M. Casagrande leg.; MUSM-LEP-105657; MUSM • 1 ♂; same collection data as for preceding but 14 Oct. 1991; H. Lamas leg.; DZ-37029; DZUP • 1 ♀; same collection data as for preceding but 15 Oct. 1991; Mielke leg.; DZ-36969; DZUP • 1 ♂; same collection data as for preceding but 340–400 m; 13 Oct. 1991; M.M. Casagrande leg.; DZ-36649; DZUP • 1 ♀; same collection data as for preceding but 14 Oct. 1991; DZ-5580; DZUP • 1 ♂; same collection data as for preceding but 15 Oct. 1991; DZ-5579; DZUP • 1 ♂; same collection data as for preceding but 9 Oct. 1991; Mielke leg.; DZ-21221; DZUP • 1 ♂; same collection data as for preceding but 15 Oct. 1991; R.K. Robbins; DZ-36619; DZUP • 1 ♂; same collection data as for preceding but 400 m; 18 Oct. 1990; G. Lamas leg.; MUSM-LEP-105656; MUSM • 1 ♂; same collection data as for preceding but 14 Oct. 1990; R.K. Robbins leg.; MUSM-LEP-105655; MUSM.

### Identification and taxonomy

*Zischkaia saundersii* is distinguished from other species in the “*saundersii* clade” by the metallic silvery blue shading between the submedian line and the marginal line on the VHW. However, although not as prominent as in *Z. saundersii*, similar VHW shading, sometimes somewhat purplish, may be seen in the three new species described below. For this reason, in addition to color, which can fade over time, the following genitalic characters enable confident identification of this species with respect to the three other species in the “*saundersii* clade” (see diagnosis of “*pacarus* clade” for genitalic characters to distinguish *Z. saundersii* from species in that clade): 1) the ‘bulb’-like projection of the tegumen is rather small, pointing posteriorly (larger in *Z. mielkeorum* sp. nov. and *Z. warreni* sp. nov.); 2) the apical process of the valva is somewhat narrow and pointing upwards in lateral view (more or less straight in *Z. mielkeorum* sp. nov. and *Z. warreni* sp. nov.); the signa are long, extending almost the entire length of the corpus bursae (shorter in *Z. mielkeorum* sp. nov. and *Z. warreni* sp. nov.).



*Euptychia saundersii* was described by Butler (1867a) based on an unspecified number of specimens from Tefê, Amazonas, Brazil, in the collection of Bates, and an unstated locality in the state of Pernambuco, Brazil. Three likely syntypes were located at the NHMUK, based on the information provided in the original description. The illustration provided in the original description (Butler 1867a: pl. XL, fig. 17) clearly shows a specimen with a silvery blue shading between the VHW submedian line and marginal line. Furthermore, the description by Butler (1867a: 500) of *Z. saundersii* was also based on a specimen with these VHW markings: “Upper wings brown, with two fine and indistinct marginal lines: body grayish-brown, antennae rusty-brown. Lower wings paler, with a violet tinge, the apical area of the HW with a greenish or violet tinge, with two dark brown central lines, diverging near the FW costa, the distalmost [of these two lines] somewhat angled [curved] in the middle of the HW [should be the FW], with the margin narrowly darker, the marginal and submarginal lines narrow and dark brown; HW with five oval ocelli and rarely with one tiny apical [ocellus], the first and fifth the smallest, black, ringed with brown, surrounded by dark brown and with tiny silver pupils; body dark grey-brown”. The description and illustration closely match a female syntype from Tefê, Amazonas, Brazil (BMNH(E) 1718073), and because of the possibility that the type series contains several taxa, as discussed further below, we designate this specimen as the lectotype of *Euptychia saundersii* Butler, 1867 (lectotype designation). The syntype male from Pernambuco figured in Warren *et al.* (2018), which has the characteristic handwritten label “*Euptychia saundersii* Butler Monog[raph].”, does not possess the VHW grayish shading of the lectotype. We examined the distal side of the valva of this syntype, which appears to be more or less straight, as in *Z. mielkeorum* sp. nov. and *Z. warreni* sp. nov. (see Fig. 5). Thus, the male syntype from Pernambuco is most likely not conspecific with the lectotype of *Z. saundersii*, and the taxonomic status of this specimen requires further investigation. If the male specimen from Pernambuco is correctly

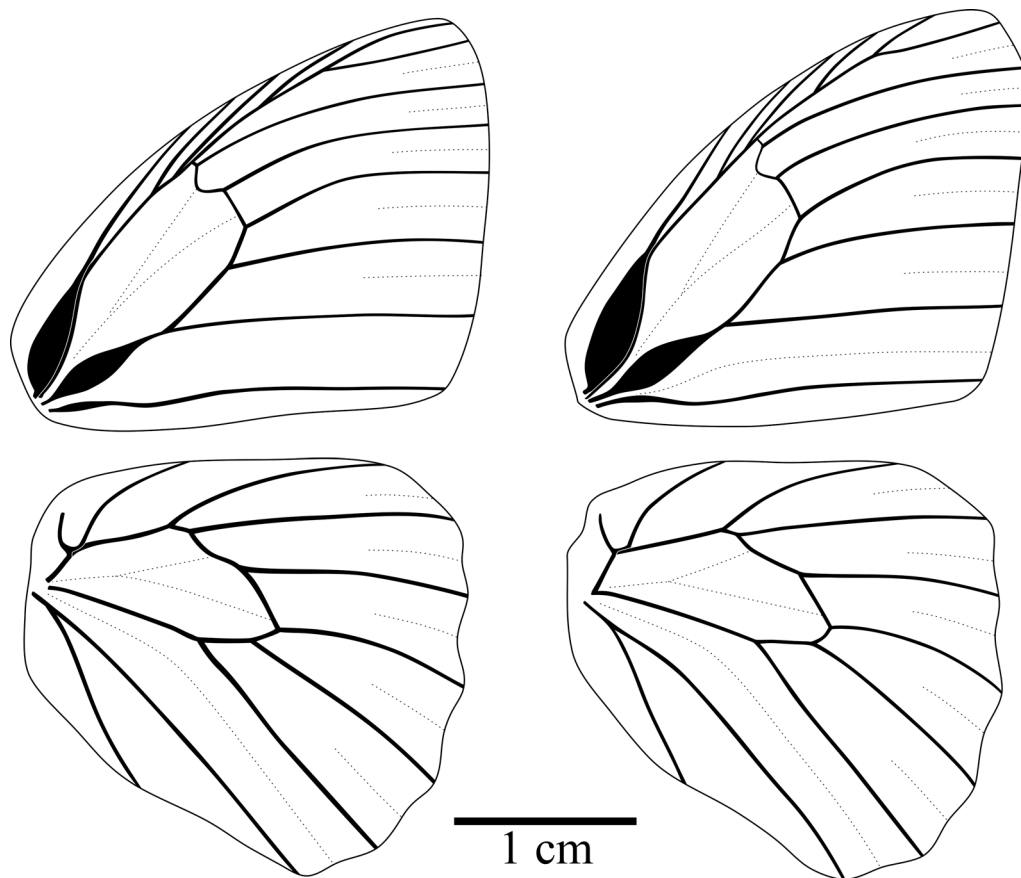


Fig. 13. *Zischkaia saundersii* (Butler, 1867), wing venation. A. ♂ (DZ 37.029). B. ♀ (DZ 36.646).

labeled, then the origin of the female syntype from Pernambuco is also questionable, given that this specimen does have silvery blue shading between the VHW submedian line and marginal line, as in the Amazonian lectotype. Clearly, reliably labeled material from Pernambuco is needed to clarify the presence of this species, or other *Zischkaia* taxa, in that region.

### Variation

The size of the ocellus in VHW Rs-M<sub>1</sub> is variable, ranging from being almost absent to clearly visible.

### Distribution (Fig. 14)

This species is reliably known from the Amazon basin, in the states of Rondônia, Pará and Maranhão, Brazil, and the departments of Huánuco and Madre de Dios, Peru. The apparently conflicting phenotypes of the two known specimens labeled as from Pernambuco, Brazil (discussed further above) makes the presence of this species in that region uncertain and requiring confirmation.

### Remarks

A male specimen from Maranhão, Brazil (DZ 38.159) phenotypically resembles *Z. saundersii* from the Amazon basin except for the VHW metallic silvery blue shading between the submedian line and marginal line being somewhat paler. The genitalia of this male from Maranhão are identical to *Z. saundersii* from the Amazon basin, in particular in the apical process of the valva being somewhat narrow and pointing upwards in lateral view. This character separates *Z. saundersii* from two other closely related species (*Z. mielkeorum* sp. nov. and *Z. warreni* sp. nov.), so we regard this specimen from Maranhão as *Z. saundersii* in the absence of further information.

*Zischkaia josti* Nakahara & Kleckner, sp. nov.

[urn:lsid:zoobank.org:act:572C5B5F-D4A5-45BB-A8DE-3B3C3DED5575](https://zoobank.org/act:572C5B5F-D4A5-45BB-A8DE-3B3C3DED5575)

Figs 1, 3E–H, 7C–D, 14, 16A–B

### Diagnosis

*Zischkaia josti* sp. nov. is closest (see discussion below) and similar to *Z. mielkeorum* sp. nov., but female specimens of these two taxa are distinguishable by the rather elongated, ‘egg’-shaped VHW ocelli of *Z. josti* sp. nov., whereas the VHW ocelli are smaller and more rounded rather than elongated in *Z. mielkeorum* sp. nov. These two species can also be separated by the presence of small sclerotized region at approximately one-third distance from ostium bursae to corpus bursae, whereas this sclerotization is apparently absent in *Z. mielkeorum* sp. nov. The range of these two taxa can also be informative regarding identification, namely *Z. josti* sp. nov. known from Guianas, whereas *Z. mielkeorum* sp. nov. occurring in Southeastern Brazil. See also relevant sections for *Z. saundersii* and *Z. warreni* sp. nov.

### Etymology

This specific epithet is dedicated to Bernhard Jost, a lepidopterist from Switzerland who collected the holotype and kindly allowed us to examine this specimen. The specific epithet is regarded as a Latinized masculine noun in the genitive case.

### Type material examined

#### Holotype

VENEZUELA • ♀; “//Venezuela Bolivar Chivatón, Strasse nach [route to] Kavanayen 1360m N05°37’55”/W061°41’40” 08.Oktober 2007 B. Jost// DNA–voucher 68763//”; BEJO, to be deposited in NMBE.

**Paratype**

GUYANA • 1 ♀; “//GUYANA: Acarai Mts./ridge Sipu R. 2500-3000” 31.X.-10.XI.2000 1°22.2” N 58 57.9”W Leg. S. Fratello et al// USNM ENT 00233844// DNA voucher LEP-18704// Genitalia dissection 2018-002 D.J.Harvey//”; USNM.

**Description**

**Male**

Unknown or unrecognized

**Female**

FOREWING LENGTH. 25–26 mm (n = 2).

HEAD. Eyes naked, with greyish scales at base; frons dark brown, with creamy-white scales and lightly colored long hair-like scales; labial palpi first segment with whitish long hair-like scales and whitish scales, second segment length almost twice as great as eye depth and covered with brownish scales laterally, with partially whitish scales, dorsally with brownish long hair-like scales, ventrally with blackish long hair-like scales and whitish long hair-like scales, about 3–4 times as long as segment width; third segment slightly shorter than half of second segment in length and covered with blackish scales, antennae cannot be examined.

THORAX. Dorsally light brown with lightly colored scales and long hair-like scales; ventrally brownish, with greyish scales lightly colored long hair-like scales.

LEGS. Foretarsus divided into five distinct subsegments; midleg and hindleg tibia and tarsus grayish dorsally, whitish ventrally, tarsus and tibia spined ventrally, and pair of tibial spurs present at distal end of tibia.

ABDOMEN. Eighth tergite uniformly sclerotized.

WING VENATION. Basal half of forewing Subcosta swollen; base of Cubitus swollen; forewing recurrent vein absent; origin of  $M_2$  towards  $M_1$  than  $M_3$ ; hindwing humeral developed.

WING SHAPE. Forewing subtriangular and elongated, apex angular, costal margin convex, outer margin slightly concave, inner margin straight, but rounded towards thorax near base. Hindwing slightly elongate, rounded, costal margin almost straight, angled towards thorax near base, outer margin undulating, inner margin slightly concave near tornus, anal lobe convex, slightly round.

DORSAL FOREWING. Ground color light brownish, apex and distal area slightly darker.

DORSAL HINDWING. Ground color similar to forewing, trace of posterior half of submarginal line visible.

VENTRAL FOREWING. Ground color greyish brown; dark brown narrow median line, almost straight, extends from near costa to origin of  $Cu_2$ , terminates after crossing origin of  $Cu_2$ ; concolorous submedian line, wider than median line, somewhat sinuous, extending from near costa to 2A, traversing in outward direction below  $M_3$ ; concolorous submarginal line, similar to submedian line in width, slightly undulating, traversing from apex towards tornus, terminating around 2A, somewhat undulating; concolorous marginal line, narrower than submarginal line, extending from apex towards tornus; fringe grayish.

VENTRAL HINDWING. Ground color similar to forewing; median line, concolorous with VFW median line, almost straight, extending from costal margin to inner margin, apparently bent inwards near inner margin; concolorous submedian line almost parallel to median line, appear slightly wider, passing origin of  $M_3$ ,

posterior end bent inwards in 2A-3A; undulating submarginal line extending from apex towards tornus, anterior end apparently fused with submedian line near costa, posterior end not fused to submedian line, terminates in 2A-3A; concolorous marginal line, slightly undulating along outer margin, appear wider than submarginal line; submarginal ocelli from  $M_1$  to 2A, oval, pupil appear as scattered silverish scales placed distally, black central spot ringed with orangish ring then with thin dark brownish indistinct ring; fringe grayish.

**FEMALE GENITALIA** (Fig. 7C–D). Inter-segmental membrane between seventh and eighth tergite not pleated, but expanded with posterior edge forming a smooth, curving sclerotized band anterior to ostium bursae that seamlessly borders the broad, sclerotized plate of lamella antevaginalis, which narrows ventally to encircle ostium bursae and is indented throughout with the edges forming raised lip; lamella antevaginalis fused to lateral sclerotized plate of eighth abdominal segment; ductus bursae membranous, small sclerotized region present at one-third distance from ostium bursae to corpus bursae; origin of ductus seminalis located between sclerotized plate and ostium bursae; corpus bursae roughly oval in dorsal view, with two signa located in middle, parallel to each other.

### Variation

The paratype possesses an ocellus in the VHW cell  $Rs-M_1$ , whereas the ocellus is absent in this cell in the holotype female.

### Distribution

 (Fig. 14)

This species is known from the type locality in Bolívar state, Venezuela and a single site in the Acarai Mountains, in southern Guyana.

### Remarks

Despite our initial hypothesis of the holotype and paratype being two different species, the small genetic distance (0.3%) based on DNA ‘barcodes’ between the holotype (MUSM-LEP-68763) and paratype (MUSM-LEP-18704) do not support this hypothesis. The distance between these two Guianan specimens and three barcoded individuals of *Z. mielkeorum* sp. nov. (BC-DZ-225, 248, 249) fall within 0.9–1.6 %, which is in accordance with rather low inter-specific genetic distance of two other taxa of *Zischkaia* regarded as species (*Z. chullachaki* sp. nov. and *Z. arenisca* sp. nov.; see Table 4) compared to many other euptychiine butterflies (pers. obs.). The wing pattern differences between the two Guianan females documented above, combined with their geographical isolation, might merit subspecific status. However, due to the fact that only a single specimen is known from each site, we feel it is premature to draw any conclusion as to the importance of this geographic variation. We thus regard the observed phenotypic difference between these two Guianan females to represent intra-specific variation and treat these specimens as a single taxon. See also corresponding section of *Z. mielkeorum* sp. nov. for relevant discussion for these two species.

*Zischkaia mielkeorum* Dolibaina, Dias & Zacca, sp. nov.

[urn:lsid:zoobank.org:act:3FCE4329-1523-4546-92E7-C7BD15D5DEDF](https://zoobank.org/act:3FCE4329-1523-4546-92E7-C7BD15D5DEDF)

Figs 1, 3I–L, 5B, 7E–F, 14

### Diagnosis

See relevant sections of *Z. saundersii*, *Z. josti* sp. nov., and *Z. warreni* sp. nov.

### Etymology

This species is named to honor three generations of German-Brazilian naturalists: Carl Helmuth Theodor Mielke, Olaf Hermann Hendrik Mielke and Carlos Guilherme Costa Mielke. The specific epithet is a masculine noun in the genitive case.

### Type material examined

#### Holotype

BRAZIL • ♂; “// Brasil, Espírito Santo, [Sooretama,] Res[erva]. Ecológica Sooretama, 19°03'25”S 40°08'50”W, 19-26-II-2013, Mielke & Casagrande leg. // DZ 36.970 // BC-DZ Willmott 248 //”; DZUP.

#### Paratypes (3 ♂♂, 6 ♀♀)

BRAZIL: **Espírito Santo** • 1 ♂; Conceição da Barra; [18°34' S, 39°45' W]; 6 Oct. 1969; C.-C. Elias leg.; DZ-21200; DZUP • 1 ♀; Linhares; [19°23' S, 40°4' W]; Sep. 1981; Elias leg.; DZ-36629; DZUP • 3 ♀♀; Sooretama, Reserva Ecológica de Sooretama; [19°3'25” S, 40°8'50” W]; 100 m; 19–26 Feb. 2013; O.H.H. Mielke and M.M. Casagrande leg.; DZ-36618, DZ-36688, DZ-36968; DZUP • 1 ♀; Santa Teresa, [19°56' S, 40°36' W]; 25–29 Mar. 1970; Ebert leg.; DZ-36711; DZUP. – **Minas Gerais** • 1 ♂; 12 km S of Teofilo Otoni; 600 m; 11 Feb. 2007; O.H.H. Mielke and M.M. Casagrande leg.; DZ-36599; DZUP. – **Rio de Janeiro** • 1 ♀; Nova Iguaçu, Fazenda São Bernardino; 19 Sep. 1937; R.F. d’Almeida leg.; DZ-36671; DZUP. – **Not located** • 1 ♂; ‘Brazil’; BMNH(E)-1205420; NHMUK.

### Description

#### Male

FOREWING LENGTH. 24–25 mm (n = 3).

HEAD. Eyes naked, with whitish scales at base; labial palpi laterally dark reddish brown with whitish scales, ventrally dark brown and whitish lines, first segment about the length of the third segment and as thick as the second, third segment thin, second segment thicker and almost four times the length of the third segment, with long hair-like scales; antennae reddish dark brown dorsally with whitish scales at the base of each segment, ventrally naked, approximately two-fifths the forewing length, with about 46 segments (n = 5), distal segments forming a slender club.

THORAX. Dorsally dark reddish brown; laterally and ventrally dark reddish brown with brownish gray scales.

LEGS. Foreleg brownish gray with whitish scales; mid and hindleg femur brownish gray with whitish scales, tibiae and tarsi mostly brownish gray with whitish scales ventrally, with pair of tibial spurs present at distal end of tibia.

ABDOMEN. Eighth tergite as stripe at base of eighth abdominal segment, in addition to presence of two distal broader patches.

WING VENATION. Basal half of forewing Subcosta swollen; base of Cubitus swollen; forewing recurrent vein absent; origin of  $M_2$  towards  $M_1$  than  $M_3$ ; hindwing humeral developed.

WING SHAPE. Forewing triangular, costal margin rounded, outer margin straight, inner margin more or less straight, apex and tornus rounded; hindwing more or less rounded, humeral area expanded, costal margin slightly rounded, outer margin slightly crenulated, inner margin slightly sinuous, apex and tornus rounded, not very conspicuous.

**DORSAL FOREWING.** Ground color variable, usually dark reddish brown; marginal area and apex conspicuously darker brown.

**DORSAL HINDWING.** Ground color similar to forewing; submarginal and marginal line somewhat visible though transparency along the costal margin, specially near the tornus.

**VENTRAL FOREWING.** Ground color brown, lighter than the dorsal forewing ground color; discal line reddish brown, noticeably fainter than other lines, running more or less obliquely from near the costal margin to 2A; postdiscal line dark reddish brown, variably developed in width, running obliquely from about  $M_1$  to 2A; submarginal line dark reddish brown, variably wavy, from the apex to the tornus along the outer margin, ending at about 2A; marginal line dark reddish brown, straight and narrower than submarginal line, from the apex to the tornus along the outer margin, ending at about 2A; fringe dark brown.

**VENTRAL HINDWING.** Ground color similar to forewing; discal line dark reddish brown, almost straight, running from costal to the inner margin; postdiscal line similar to and more or less parallel to discal line, passing through the end of the discal cell at the origin of  $M_3$ , and posteriorly directed to the base of the wing at 2A-3A; submarginal band wavy, similar in color and width to the discal and postdiscal line, running from apex towards the tornus, occasionally fused with the postdiscal line near the costal margin and occasionally fused to the submedian line in 2A-3A; marginal line similar in color, but thinner than other lines, slightly wavy along the outer margin; submarginal ocelli from  $M_1$  to 2A, roughly oval, dark brown encircled by a marked orange ring and a subtler brown ring; pupil with scattered silver scales, ocelli at  $M_2$ - $M_3$ ,  $M_3$ - $Cu_1$ , and  $Cu_1$ - $Cu_2$  similar in size, somewhat elongate, those in  $M_1$ - $M_2$  and  $Cu_2$ -2A underdeveloped, usually half the size of the other ocelli; scales of the areas between postdiscal and submarginal lines and submarginal and marginal lines with variable light purple to lilac tinge; fringe dark brownish.

**GENITALIA** (Fig. 5B). Tegumen somewhat flattened, anteriorly bulged, with a single dorsal dome-like projection near the base of the uncus; appendices angulares absent; uncus laterally flattened and straight, slightly longer than the length of the tegumen in lateral view; brachia slightly shorter than the uncus, thicker at the base and narrowing to a point posteriorly; combination of ventral arms of the tegumen and dorsal arms of the saccus sinuous; saccus anterior projection tube-like, approximately the length of the tegumen; valva more or less triangular, stretched dorsal and posteriorly; basal half of valva larger, narrowing towards the apex, dorsal margin almost straight, ventral margin convex, apex broadly rounded; futura inferior (i.e., juxta) strap-like; phallobase about one-third of phallus, rather straight; ductus ejaculatorius not discerned; aedeagus straight with manica covering approximately half, winglet absent, distal opening located ventrally where vesica is visible.

### **Female**

**FOREWING LENGTH.** 26–27 mm (n = 6).

Similar to male except as follows: foretarsus divided into five tarsomers; wings ground color generally lighter; forewing more rounded and broader, outer margin usually slightly convex; hindwing outer margin more crenulated; dorsal hindwing submarginal and marginal lines more noticeable seen through transparency.

**FEMALE ABDOMEN AND GENITALIA** (Fig. 7E–F): membranous areas between seventh and eighth sternite folded forming the sclerotized antrum attached to the lamella antevaginalis near the ostium bursae; lamella antevaginalis sclerotized; lamella postvaginalis absent; eighth abdomen segment with a lateral sclerotized area; ductus bursae with a sclerotized area of about a third of its length near the ostium

bursae; ductus seminalis origin close to the ostium bursae; corpus bursae oval, with a pair of parallel and dorsal signa, signa narrow and slightly shorter than the length of the corpus bursae.

### Variation

The silvery blue patch between the median and submarginal lines in VHW can vary in intensity, and the coloration of the ocellar ring is also variable, being more yellowish in some specimens.

### Distribution (Fig. 14)

This species is known to date from southeastern Brazil, from the states of Espírito Santo, Rio de Janeiro and Minas Gerais.

### Remarks

Three barcoded specimens of *Z. mielkeorum* sp. nov. (BC-DZ-225, 248, 249) form a well supported clade (SH-aLRT/UF Boot = 99.8/95), with two individuals of *Z. josti* sp. nov. (MUSM-LEP-68763, 18704) (Fig. 1). Despite the small sampling size ( $n = 5$ ) for these two species, the existence of a ‘barcoding gap’ between these two taxa (see corresponding section of *Z. josti* sp. nov.), combined with the presence of wing pattern and genitalia differences that separate these two species, leads us to consider them as distinct species. The known ranges of these species are approximately 3000 km apart, reinforcing this taxonomic decision. In fact, no known area of sympatry exists between any species pair in the “*saundersii* clade”, which thus appears to have undergone allopatric speciation. However, this taxonomic treatment will result in accepting *Z. mielkeorum* sp. nov. as a paraphyletic group. Although it is not an acceptable taxonomic practice to name non-monophyletic groups, we argue in this case based on the fact that BC-DZ-249 is missing first 80 or so base pairs of its sequence. This missing information might have resulted in separation from the sequence generated from the individual from the same locality (BC-DZ-248, from Res. Ecologica Sooretama, Espírito Santo, Brazil) and, in fact, grouped together with BC-DZ-225, which is a specimen from a different site (Minas Gerais, Brazil). It is also worth noting that a number of butterfly species are recovered as paraphyletic, including species in some of the best studied butterfly genus *Heliconius* Kluk, 1780 (e.g., *H. erato* (Linnaeus, 1758) as in Kozak *et al.* 2015). Clearly, more data based on more individuals is needed in order to draw a firm conclusion as to taxonomic status of these two populations, and our treatment herein is a mere provisional hypothesis based on available information.

*Zischkaia warreni* Dias, Zacca & Dolibaina, sp. nov.

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Figs 1, 3M–P, 5C, 7G–H, 14, 16C

### Diagnosis

*Zischkaia warreni* sp. nov. is distinguished from other species in the “*saundersii* clade” by the absence of the VFW submedian line. Further distinctive characters include the rather prominent DHW submarginal and marginal line, reduced VHW ocellus in  $M_1$ - $M_2$  combined with the ocellus in  $M_2$ - $M_3$  being more than twice as large (in terms of diameter) and the median line and submarginal line being fused immediately after 2A in males. Additional characters are also discussed in the relevant sections of other species.

### Etymology

This specific epithet is in honor of our good friend, Andrew D. Warren, a prominent lepidopterist with a broad knowledge of butterflies. The specific epithet is to be considered a latinized masculine noun in the genitive case.

**Type material examined****Holotype**

BRAZIL • ♂; “// 2-XI-2000, Represa Sanepar, Piraquara, PR [Paraná, Brazil], Bizarro leg. //DZ 36.948 //BC-DZ Willmott 227 //”; DZUP.

**Paratypes (20 ♂♂, 31 ♀♀)**

BRAZIL: **Minas Gerais** • 1 ♀; Itamonte; [22°17'16" S, 44°51'54" W]; Dec. 1957; Ebert leg.; DZ-36731; DZUP. – **Paraná** • 1 ♀; Balsa Nova, São Luís do Purunã; [25°29' S, 49°44' W]; 1000 m; 21 Feb. 2001; G. Lamas leg.; MUSM-LEP-105660; MUSM • 3 ♀♀; Castro; [24°47' S, 50°1' W]; 950 m; BMNH(E)-1718053 to BMNH(E)-1718054, BMNH(E)-1718064; NHMUK • 4 ♀♀, same collection data as for preceding; USNM • 1 ♂, 1 ♀; same collection data as for preceding; E.D. Jones leg.; BMNH(E)-1718069, BMNH(E)-1718061; NHMUK • 1 ♀; same collection data as for preceding but Sep. 1897; BMNH(E)-1718060; NHMUK • 1 ♀; Guarapuava, Serra da Esperança, N side BR 373, 4.3 rd km ENE of Guará; [25°21'49.49"S 51°14'46.07"W]; 1218 m; 4 Feb. 2016; D. Dolibaina and A. Warren leg.; DD-380 • 2 ♂♂, 3 ♀♀; same collection data as for preceding but 6 Feb. 2016; DD-375 to DD-379 • 7 ♂♂, 3 ♀♀, same collection data as for preceding but A. Warren leg.; ADW • 1 ♀; Piraquara, Mananciais da Serra; [25°30'28" S, 49°01'32" W]; 5 Nov. 1972; Mielke leg.; DZ-36739; DZUP • 1 ♂; Quatro Barras, Morro do Anhangava; [25°23'18" S, 49°0'14" W]; 1350 m; 25 Mar. 2009; E. Carneiro leg.; DD-306 • 1 ♂; same collection data as for preceding but D. Dolibaina leg.; DD-305 • 1 ♀; Tijucas do Sul; [25°55' S, 49°12' W]; 850 m; 13 Mar. 2004; G. Lamas leg.; MUSM-LEP-105659; MUSM • 1 ♀; Tijucas do Sul, Rincão; [25°48' S, 49°8' W]; 900 m; 25 Feb. 1969; Mielke leg.; DZ-36719; DZUP • 1 ♂, 1 ♀; Tijucas do Sul, Vossoroca; 850 m; 20 Mar. 1971; Mielke leg.; DZ-21214, DZ-36669; DZUP • 2 ♂♂, 1 ♀; same collection data as for preceding but 8 Mar. 1972; DZ-36659, DZ-36679, DZ-36709; DZUP • 1 ♂; União da Vitória; [26°12' S, 51°5' W]; 610 m; Dec. 1919–Jan. 1920; A. Hall leg.; BMNH(E)-10430703; NHMUK • 1 ♀; same collection data as for preceding; 750 m; 7 Nov. 1985; O.H.H. Mielke and M.M. Casagrande leg.; DZ-36749; DZUP. – **Rio de Janeiro** • 1 ♂; Itatiaia, Serra Itatiaia, vertente SE; [22°19'55" S, 44°36'38" W]; 1300 m; Feb. 1950; Ebert leg.; DZ-36639; DZUP. – **Santa Catarina** • 1 ♀; Campo Alegre, Serra do Quiriri, [26°1'34" S, 48°59'2" W]; 1300 m; 14 Nov. 2009; O.H.H. Mielke, E. Carneiro and Melo leg.; DZ-36538; DZUP • 1 ♀; same collection data as for preceding but 14 Nov. 2009; O. Mielke and E. Carneiro; DD-304 • 1 ♂, 1 ♀; Irani; [27°01'20" S, 51°54'15" W]; 1000 m; 8 Nov. 1985; O.H.H. Mielke and M.M. Casagrande leg.; DZ-36689, DZ-36699; DZUP • 1 ♀; Mafra; [26°07'08" S, 49°48'50" W]; 22–23 Feb. 1982; Mielke leg.; DZ-36729; DZUP • 1 ♂; São Bento do Sul; [26°15' S, 49°23' W]; 850 m; 2 Feb. 1985; H. Miers leg.; FLMNH-MGCL-1036043; FLMNH • 1 ♂; Seara, Nova Teutonia; [27°3' S, 52°23' W]; 300–500 m; Feb. 1973; F. Plaumann leg.; FLMNH-MGCL-1036044; FLMNH. – **São Paulo** • 2 ♀♀; Apiaí, Serra de Paranapiacaba; [24°31' S, 48°51' W]; Feb. 1972; Ebert leg.; DZ-36609, DZ-36653; DZUP. – **Not located** • 1 ♀; ‘Brazil’; BMNH(E)-1205409; NHMUK.

**Description****Male**

FOREWING LENGTH. 24–25 mm (n = 3).

HEAD. Eyes naked, with whitish scales at base; labial palpi first segment with white and brownish long hair-like scales; second segment length almost twice as great as eye depth and covered with brown scales and hair-like scales laterally, with white scales and hair-like scales, dorsally with pale brownish long hair-like scales, ventrally with black and white hair-like scales, about 3–4 times as long as segment width; third segment about one-fourth of second segment in length and covered with black scales and hair-like scales, with slight patch of creamy-white scales laterally; antennae approximately two-fifth of forewing length, with ca 37 segments (n = 1), distal ca 15 composing club, club not prominent.



THORAX. Dorsally, laterally and ventrally scattered with brownish gray scales.

LEGS. Foreleg brownish, foretarsus and tibia similar in length, femur not examined; midleg and hindleg with femur creamy white ventrally, tibia and tarsus grayish dorsally, whitish to ochre ventrally, tarsus and tibia spined ventrally, and a pair of tibial spurs at distal end of tibia.

ABDOMEN. Eighth tergite as stripe at base of eighth abdominal segment, in addition to presence of two distal broader patches.

WING VENATION. Basal half of forewing Subcosta swollen; base of Cubitus swollen; forewing recurrent vein absent; origin of  $M_2$  towards  $M_1$  than  $M_3$ ; hindwing humeral developed.

WING SHAPE. Forewing subtriangular, apex rounded, costal margin convex, outer margin slightly convex (almost straight), inner margin straight, but rounded towards thorax near base; hindwing slightly elongate, rounded, costal margin almost straight, angled towards thorax near base, outer margin slightly undulating, inner margin slightly concave near tornus, anal lobe convex, slightly round.

DORSAL FOREWING. Ground color brownish, distally darker; androconial scales absent; trace of submarginal band indistinct.

DORSAL HINDWING. Ground color similar to forewing, trace of submarginal and marginal band visible.

VENTRAL FOREWING. Ground color light grayish brown, area below 2A paler; submedian line invisible; dark-brown narrow median line, extends from Radius to 2A, traversing wing outward with terminal end of median line almost reaching submarginal line; concolorous submarginal line extending from apex towards tornus, terminating around 2A; concolorous marginal line, narrower than submarginal line, extending from apex towards tornus, but terminates around 2A; fringe dark brownish.

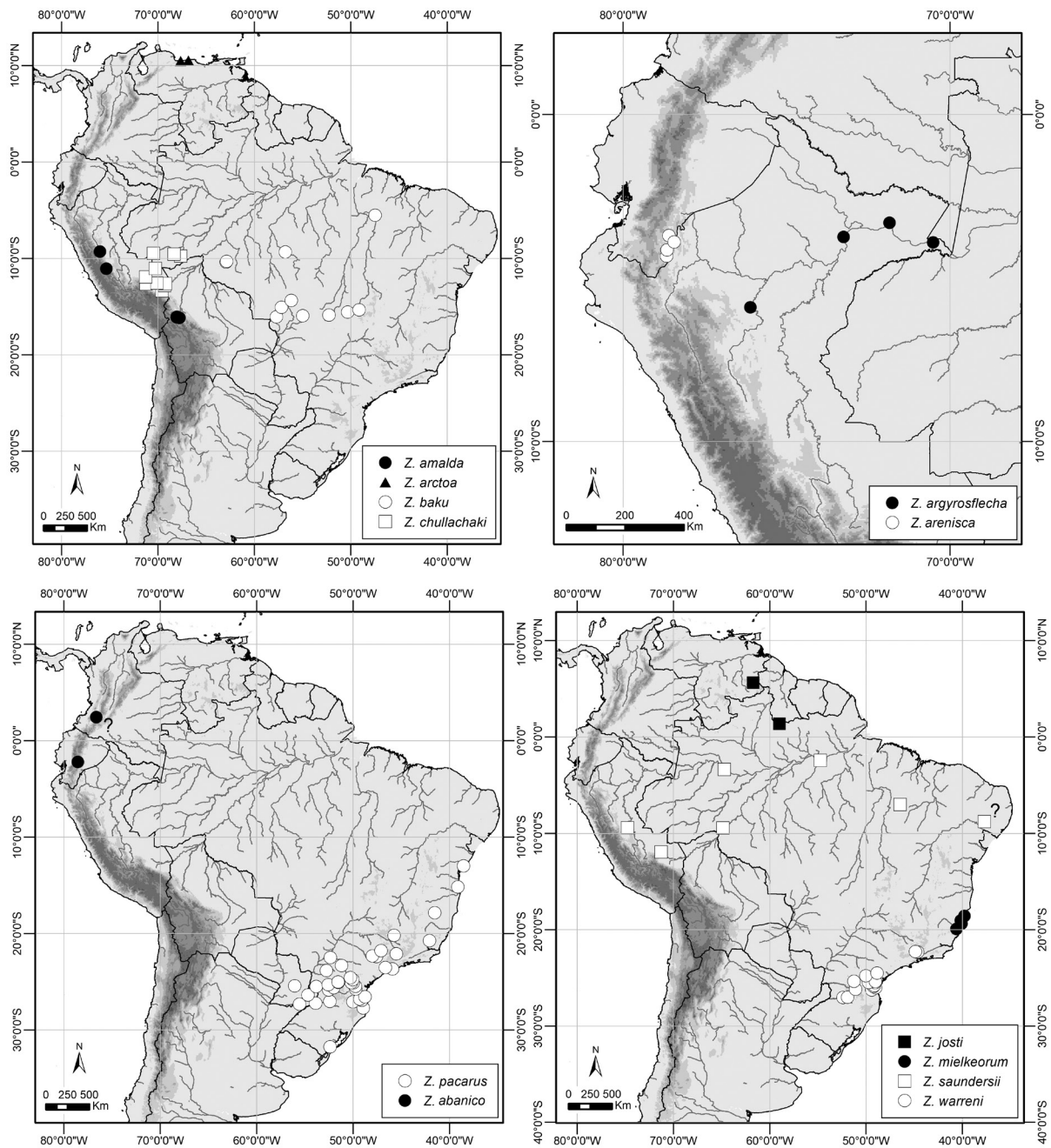
VENTRAL HINDWING. Ground color similar to forewing; regular dark-brown submedian line somewhat sigmoid, extending from costa to inner margin, terminal end curving basally along inner margin; median line almost parallel to submedian line, concolorous, similar in width, passing origin of  $M_3$ , posterior and anterior end fused with submarginal line in 2A-3A and Rs- $M_1$  respectively; submarginal line extending from apex towards tornus, anterior and posterior end fused with median line in Rs- $M_1$ , jagged, posterior end slightly broadening and fused to median line in 2A-3A; marginal line, concolorous, undulate along outer margin, much thinner than submarginal line; submarginal ocelli from  $M_1$  to 2A, roughly oval, pupil as silver scales placed distally, black central spot ringed with orangish ring then with thin dark brownish indistinct ring, those in  $M_2$ - $M_3$  and  $M_3$ - $Cu_1$  are both similar in size, those in  $M_1$ - $M_2$  and  $Cu_2$ -2A smaller, especially ocellus in  $M_1$ - $M_2$  reduced, ocellus in  $Cu_1$ - $Cu_2$  largest; fringe dark brownish.

GENITALIA (Fig. 5C). Tegumen somewhat rounded in lateral view, dorsally curved in lateral view, dome-shaped hump present posteriorly along dorsal margin; combination of ventral arms from tegumen and dorsal arms from saccus almost straight; appendices angulares present but rather inconspicuous; saccus shorter than uncus in length; uncus long and narrow, about twice as long as tegumen in length, with sparse hair-like setae, slightly hooked at posterior end; brachia long and narrow, shorter than uncus in length; fultura inferior (i.e., juxta) present as developed plate in posterior view; valva setose, basal half of valva roughly trapezoidal in lateral view, ventral margin convex in middle, dorsal margin almost straight except for margin basal to costa angles down to meet ventral margin, distal half narrow and somewhat elongate with rounded apex; costa subtriangular in lateral view, projecting towards appendices angulares; phallobase about one-third of phallus, rather straight; ductus ejaculatorius coming out higher than anterior end of coecum; aedeagus straight with manica covering approximately half, winglet absent, distal opening located ventrally where vesica is visible.

**Female**

FOREWING LENGTH. 26–27 mm (n = 6).

Similar to male except as follows: foretarsus first and second subsegments apparently fused; forewing more rounded and broad; ground color of both wing surfaces paler, posterior end of VFW discal line do not come close to submarginal line as in male. VHW area between postdiscal and submarginal line whitish (around ocelli).

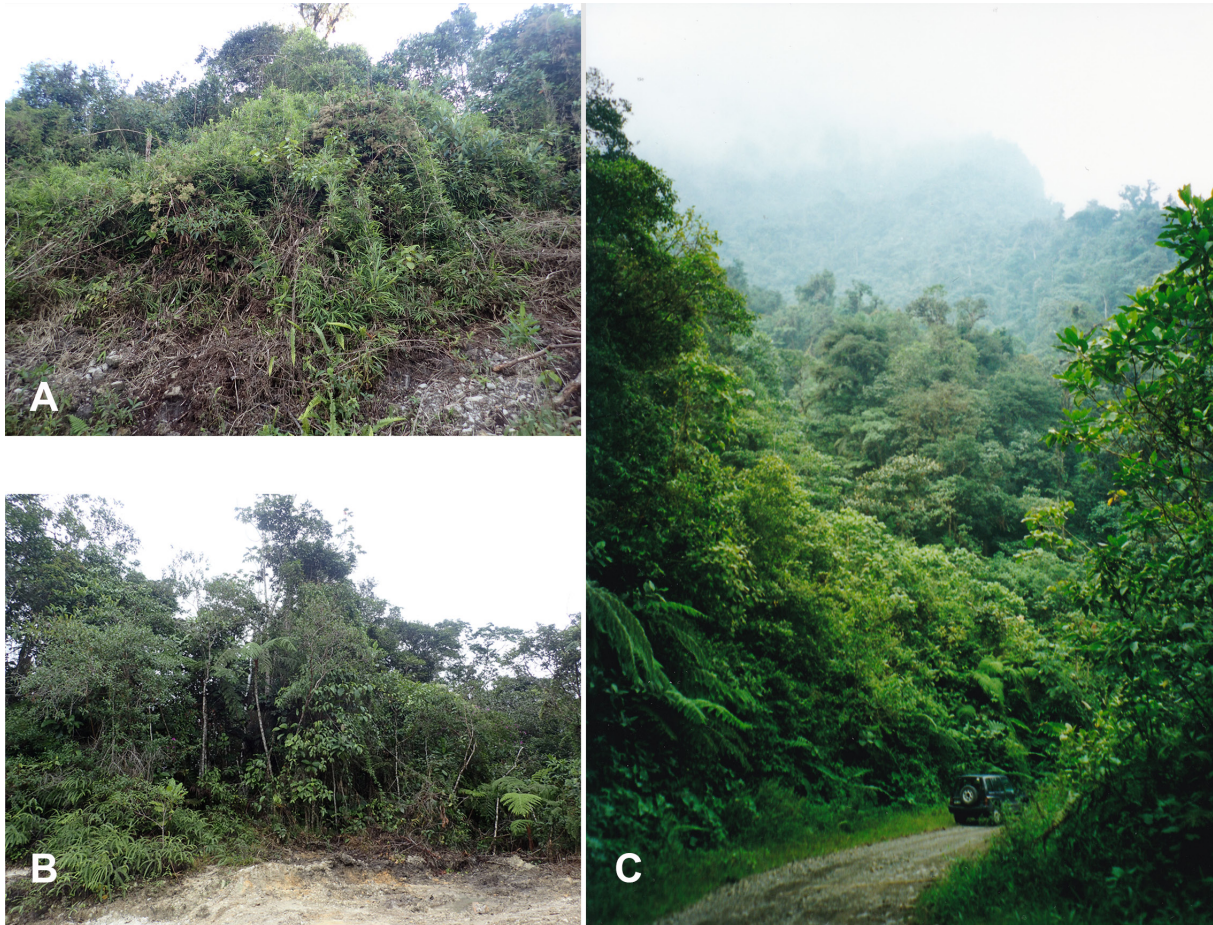


**Fig. 14.** Distribution maps for taxa of *Zischkaia* Forster, 1964.

FEMALE ABDOMEN AND GENITALIA (Fig. 7G–H): inter-segmental membrane between seventh and eighth tergite not pleated, but expanded with posterior edge forming a smooth, curving sclerotized band anterior to ostium bursae that seamlessly borders the broad, sclerotized plate of lamella antevaginalis, which narrows ventrally to encircle ostium bursae and is indented throughout with the edges forming raised lip; eighth abdominal segment lateral side with sclerotized plate and with spiracle near dorsal margin, ventrally connected to lamella antevaginalis as sclerotized stripe, fused below ostium bursae as ‘arm’; ductus bursae membranous; origin of ductus seminalis close to ostium bursae; corpus bursae roughly oval in dorsal view, extending to third abdominal segment, with two signa in middle, parallel to each other.

### Variation

The VFW submarginal and marginal lines are more or less fused in cell  $Cu_2$  in some specimens, whereas they are placed rather further apart in other specimens.



**Fig. 15.** Habitats of species of *Zischkaia* Forster, 1964. **A.** Climbing bamboo species (Poaceae, likely *Chusquea* Kunth) that was common along the Sumak Yaku mine track, Zamora-Chinchi, Ecuador, where *Z. arenisca* Nakahara, Willmott & Hall, sp. nov. was common (photo by Keith Willmott). **B.** Ridge top at San Roque ridge, Zamora-Chinchi, Ecuador, where females of *Z. arenisca* sp. nov. were found (photo by Keith Willmott). **C.** View of the ridge line 500 m west of the Río Abanico (Morona-Santiago, Ecuador) collecting site (photo by Andrew Neild).

**Distribution** (Fig. 14)

This species is known to date from southeastern to southern Brazil, in the states of Rio de Janeiro, Minas Gerais, São Paulo, Paraná and Santa Catarina. The geographic distribution of *Z. warreni* sp. nov. is apparently confined to areas associated with plateaus with *Araucaria* forest in middle to high elevations



**Fig. 16.** Habitats of species of *Zischkaia* Forster, 1964. **A.** Habitat at the type locality of *Z. josti* Nakahara & Kleckner, sp. nov., Kavanayen, Venezuela (photo by Bernhard Jost). **B.** Adult *in situ* at Serra da Esperança, Guarapuava, Paraná, Brazil (photo by Andrew Warren). **C.** Habitat of *Z. warreni* Dias, Zacca & Dolibaina, sp. nov., Serra da Esperança, Guarapuava, Paraná, Brazil (photo by Andrew Warren).

of southern Brazil, not extending beyond Serra Geral in the west. This distributional pattern is distinct from its potentially sympatric species *Z. pacarus*, which is also found in low elevation areas in the Paraná drainage and in the Atlantic coastal zone.

### Remarks

The evidence from molecular data (see Fig. 1 and Table 4) and morphological distinctiveness documented above leaves no doubt as to its specific status. Adults of *Z. warreni* sp. nov. are similar in behavior to *Z. pacarus*, flying in association with the bamboo *Merostachys* (Poaceae: Bambusoideae) (Fig. 16A, C). The males' flight is faster than that of the females, and individuals are commonly found engaged in patrolling flight. Both sexes are never found away from the bamboo and they are more active between the middle of the morning and the beginning of the afternoon. The restricted habitat of *Z. warreni* sp. nov. in *Araucaria* forests suggests that they likely use *Merostachys* as the caterpillar host plant.

### Discussion

Our species-level review of *Zischkaia* based on morphology and molecular data increased the species-richness of the genus by adding nine species named and described above. Despite two distinctive and morphologically diagnosable clades recognized in *Zischkaia* (“*pacarus* clade” and “*saundersii* clade”), species within these two clades are rather similar in terms of wing pattern and genitalia. For example, *Z. pacarus* and *Z. abanico* sp. nov. are easily distinguished from the other species in the “*pacarus* clade” based on the shape of the apical process of the valva, but the male genitalia of these two species (*Z. pacarus* and *Z. abanico* sp. nov.) and the rest of the species in the clade are highly homogenous where interspecific difference either does not exist or rather subtle even if present. A similar case of genitalia homogeneity among species is reported in other euptychiine species-groups, such as *Euptychia* “*audacia* clade” and *Forsterinaria* “*boliviana* clade” (Peña & Lamas 2005; Nakahara *et al.* 2017). This observed morphological homogeneity combined with the lack of DNA ‘barcodes’ data for four species in the “*pacarus* clade” may lead to a discussion as to the conspecificity of some taxa, in particular, *Z. amalda*, *Z. arctoa* sp. nov., *Z. baku* sp. nov. and *Z. chullachaki* sp. nov. (barcoding data available for only *Z. chullachaki* sp. nov.). However, we argue against this by their allopatric distribution and apparent difference in elevational zone where they occur (*Z. amalda* and *Z. arctoa* sp. nov. being montane; *Z. baku* sp. nov. and *Z. chullachaki* sp. nov. being lowland species), in addition to subtle morphological differences between these taxa discussed in relevant sections of those species above. Nevertheless, the lack of DNA sequence data for *Z. amalda*, *Z. arctoa* sp. nov. and *Z. baku* sp. nov. hinders further investigation of their taxonomic status, as well as their relationships to determine pairs of sister taxa. As with species in the “*saundersii* clade”, the known geographic range of taxa in the “*pacarus* clade” suggests they followed allopatric speciation. Thus, determining sister species pairs in the “*pacarus* clade” will enable us to test if speciation across an elevational gradient (i.e., vertical speciation, as in Chapman 1917) occurred in this group or not, to further assess the taxonomic status of these poorly known elements of the Neotropical butterfly fauna.

### Author contributions

SN prepared the first draft of the manuscript, which was read and commented on by all authors. The morphological study and preparation of relevant figures were performed by SN, TZ, FMSD and DRD. The molecular lab work, including the analysis, was conducted by SN, KK and LX. SN, TZ, FMSD, DRD, ME, MMC, OHHM, GL, BH and KW contributed towards compiling specimen data, including gathering tissue samples mainly done by SN, ME and KW. Finally, Jean-Claude Petit, Lee D. Miller (deceased) and Jason Hall co-authored new species described and named in this article thanks to their contributions to collecting key material or to developing species descriptions.

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## References

- Austin G.T. & Mielke O.H.H. 2008. HesperIIDae of Rondônia, Brazil: *Porphyrogenes* Watson (Lepidoptera: Pyrginae: Eudamini), with descriptions of new species from Central and South America. *Insecta Mundi* 44: 1–56.
- Barbosa E.P., Silva A.K., Paluch M., Azeredo-Espin A.M.L. & Freitas A.V.L. 2015. Uncovering the hidden diversity of the Neotropical butterfly genus *Ypthimoides* Forster (Nymphalidae: Satyrinae): description of three new species based on morphological and molecular data. *Organisms Diversity and Evolution* 15 (3): 577–589. <https://doi.org/10.1007/s13127-015-0221-y>
- Beccaloni G.W., Vilorio Á.L., Hall S.K. & Robinson G.S. 2008. *Catalogue of the Hostplants of the Neotropical Butterflies. Catálogo de las Plantas Huésped de las Mariposas Neotropicales*. Monografías del Tercer Milenio 8. Sociedad Entomológica Aragonesa, Zaragoza.
- Bland K.P. 2019. Name-bearing types of butterflies (Lepidoptera, Papilionoidea), in the National Museums of Scotland, Edinburgh. *Zootaxa* 4559 (1): 57–89. <https://doi.org/10.11646/zootaxa.4559.1.2>
- Brown K.S. 1992. *Borboletas da Serra do Japi: Diversidade, habitats, recursos alimentares e variação temporal*. In: Morellato L.P.C. (ed.) *História natural da Serra do Japi. Ecologia e Preservação de uma Área florestal no Sudeste do Brasil*: 142–187. Campinas, Editora da Unicamp/Fapesp.
- Brown K.S. & Freitas A.V.L. 2000. Diversidade de Lepidoptera em Santa Teresa, Espírito Santo. *Boletim do Museu de Biologia Mello Leitão* (N.S.) 11/12: 71–116.
- Butler A.G. 1867a. A monograph of the genus *Euptychia*, a numerous race of butterflies belonging to the family Satyridae; with descriptions of sixty species new to science, and notes to their affinities, etc. *Proceedings of the Zoological Society of London* 1866 (3): 458–504.

- Butler A.G. 1867b. Descriptions of some new species of Satyridae belonging to the genus *Euptychia*. *Proceedings of the Zoological Society of London* 1867 (1): 104–110.
- Butler A.G. 1868. *Catalogue of Diurnal Lepidoptera of the Family Satyridae in the Collection of the British Museum*. Taylor and Francis, London.
- Butler A.G. 1877. On new species of the genus *Euptychia*, with a tabular view of those hitherto recorded. *Journal of the Linnean Society of London (Zoology)* 13 (67): 116–128.  
<https://doi.org/10.1111/j.1096-3642.1877.tb02375.x>
- Carneiro E., Mielke O.H.H. & Casagrande M.M. 2008. Borboletas do sul da ilha de Santa Catarina, Florianópolis, Santa Catarina, Brasil (Lepidoptera: Hesperioidea e Papilionoidea). *Shilap* 36 (142): 261–271.
- Chapman F.M. 1917. The distribution of bird life in Colombia. *Bulletin of the American Museum of Natural History* 31: 1–169.
- Comstock J.H. & Needham J.G. 1898. The wings of insects. *American Naturalist* 32 (373): 43–48.
- Cong Q. & Grishin N.V. 2014. A new *Hermeuptychia* (Lepidoptera, Nymphalidae, Satyrinae) is sympatric and synchronic with *H. sosybius* in southeast US coastal plains, while another new *Hermeuptychia* species - not *hermes* - inhabits south Texas and northeast Mexico. *ZooKeys* 379: 43–91.  
<https://doi.org/10.3897/zookeys.379.6394>
- D’Abrera B. 1988. *Butterflies of the Neotropical Region. Part V. Nymphalidae (Conc.) & Satyridae*: 679–877. Black Rock, Hill House, Victoria.
- Dolibaina D.R., Mielke O.H.H. & Casagrande M.M. 2011. Borboletas (Papilionoidea e Hesperioidea) de Guarapuava e arredores, Paraná, Brasil: um inventário com base em 63 anos de registros. *Biota Neotropica* 11 (1): 341–354. <https://doi.org/10.1590/S1676-06032011000100031>
- Edgar R.C. 2004. MUSCLE: multiple sequence alignment with high accuracy and high throughput. *Nucleic Acids Research* 32 (5): 1792–1797. <https://doi.org/10.1093/nar/gkh340>
- Elias M., Hill R., Willmott K.R., Dasmahapatra K., Brower A., Mallet J. & Jiggins C. 2007. Limited performance of DNA barcoding in a diverse community of tropical butterflies. *Proceedings of the Royal Society of London B* 274: 2881–2889. <https://doi.org/10.1098/rspb.2007.1035>
- Emery E.O., Brown Jr. K.S. & Pinheiro C.E.G. 2006. As borboletas (Lepidoptera, Papilionoidea) do Distrito Federal, Brasil. *Revista Brasileira de Entomologia* 50 (1): 85–92.  
<https://doi.org/10.1590/S0085-56262006000100013>
- Espeland M., Breinholt J., Barbosa E., Casagrande M.M., Huertas B., Lamas G., Marín M. A., Mielke O.H.H., Miller J.Y., Nakahara S., Tan D., Warren A.D., Zacca T., Kawahara A., Freitas A.V.L. & Willmott K.R. 2019. Four hundred shades of brown: higher level phylogeny of the problematic *Euptychiina* (Lepidoptera, Nymphalidae, Satyrinae) based on hybrid enrichment data. *Molecular Phylogenetics and Evolution* 131: 116–124. <https://doi.org/10.1016/j.ympev.2018.10.039>
- Forster W. 1964. Beiträge zur Kenntnis der Insektenfauna Boliviens XIX Lepidoptera III. Satyridae. *Veröffentlichungen der Zoologischen Staatssammlung München* 8: 51–188.
- Freitas A.V.L., Barbosa E.P., Zacca T., Marín M.A., Beirão M.V., Silva A.R.M., Casagrande M.M., Espeland M. & Willmott K.R. 2018. Before it is too late: description of a new genus and species of butterfly from a highly threatened Brazilian biome. *Revista Brasileira de Entomologia* 62: 148–158.  
<https://doi.org/10.1016/j.rbe.2018.02.003>
- Gaede M. 1931. Familia Satyridae. *Lepidopterorum Catalogus* 43: 1–320; 46: 321–544; 48: 545–759.
- GIMP team 2016. GIMP–GNU Image Manipulation Program. Available from <https://www.gimp.org> [accessed Jun. 2016].

- Godart J.B. [1824]. In: Latreille P.A. & Godart J.B. (eds) *Encyclopédie méthodique. Histoire naturelle. Entomologie, ou Histoire naturelle des Crustacés, des Arachnides et des Insectes*: 329–828. Veuve Agasse, Paris.
- Grimshaw P.H. 1897. On some type specimens of Lepidoptera and Coleoptera in the Edinburgh Museum of Science and Art. *Transactions of the Royal Society of Edinburgh* 39 (1): 1–11.
- Guindon S., Dufayard J-F., Lefort V., Anisimova M., Hordijk W. & Gascuel O. 2010. New algorithms and methods to estimate maximum-likelihood phylogenies: assessing the performance of PhyML 3.0. *Systematic Biology* 593: 307–321. <https://doi.org/10.1093/sysbio/syq010>
- Hoang D.T., Chernomor O., von Haeseler A., Minh B.Q. & Vinh L.S. 2018. UFBoot2: Improving the ultrafast bootstrap approximation. *Molecular Biology and Evolution* 35: 518–522. <https://doi.org/10.1093/molbev/msx281>
- Horn W., Kahle I., Friese G. & Gaedike R. 1990. *Collectiones entomologicae. Ein Kompendium über den Verbleib entomologischer Sammlungen der Welt bis 1960. Teil I: A bis K*. Akademie der Landwirtschaftswissenschaften der Deutschen Demokratischen Republik, Berlin.
- ICZN (International Commission on Zoological Nomenclature) 1999. *International Code of Zoological Nomenclature, 4<sup>th</sup> Edition*. International Trust for Zoological Nomenclature, London.
- Kalyaanamoorthy S., Minh B.Q., Wong T.K.F., von Haeseler A. & Jermin L.S. 2017. ModelFinder: Fast Model Selection for Accurate Phylogenetic Estimates. *Nature Methods* 14: 587–589. <https://doi.org/10.1038/nmeth.4285>
- Kirby W.F. 1871. *A Synonymic Catalogue of Diurnal Lepidoptera*. John Van Voorst, London.
- Kirby W.F. 1879. *Catalogue of the Collection of Diurnal Lepidoptera Formed by the Late William Chapman Hewitson of Oatlands, Walton-on-Thames; and Bequeathed by him to the British Museum*. John Van Voorst, London.
- Klots A.B. 1956. Lepidoptera. In: Tuxen S.L. (ed.) *Taxonomist's Glossary of Genitalia in Insects*: 97–111. Munksgaard, Copenhagen.
- Kozak K.M., Wahlberg N., Neild A.F., Dasmahapatra K.K., Mallet J. & Jiggins C.D. 2015. Multilocus species trees show the recent adaptive radiation of the mimetic *Heliconius* butterflies. *Systematic Biology* 64 (3): 505–524. <https://doi.org/10.1093/sysbio/syv007>
- Lamas G. 1994. Butterflies of the Explorer's Inn Reserve. In: Foster R.B., Carr J.L. & Forsyth A.B. (eds) *The Tambopata-Candamo Reserved Zone of Southeastern Perú: A Biological Assessment*: 62–63, 162–177. RAP Working Papers 6, Conservation International.
- Lamas G. 1997. Lepidoptera of the Cordillera del Cóndor. In: Schulenberg T.S. & Awbrey K. (eds) *The Cordillera del Cóndor Region of Ecuador and Peru: A Biological Assessment*: 90–98, 212–230. RAP Working Papers 7, Conservation International.
- Lamas G. 2004. Nymphalidae. Satyrinae. Tribe Satyrini. Subtribe Euptychiina In: Lamas G. (ed.) *Checklist: Part 4A. Hesperioidea - Papilionoidea*. In: Heppner J.B. (ed.), *Atlas of Neotropical Lepidoptera. Volume 5A*: 217–223. Association for Tropical Lepidoptera, Scientific Publishers, Gainesville.
- Lamas G. & Grados J. [1997]. Mariposas de la Cordillera del Sira, Peru. *Revista Peruana de Entomología* 39: 55–61.
- Lamas G., Robbins R.K. & Harvey D.J. 1991. *A preliminary survey of the butterfly fauna of Pakitza, Parque Nacional del Manu, Peru, with an estimate of its species richness*. Publicaciones del Museo de Historia Natural 40, Universidad Nacional Mayor de San Marcos.



- Marín M.A., Peña C., Freitas A.V.L., Wahlberg N. & Uribe S.I. 2011. From the phylogeny of the Satyrinae butterflies to the systematics of Euptychiina (Lepidoptera: Nymphalidae): history, progress and prospects. *Neotropical Entomology* 40: 1–13. <https://doi.org/10.1590/S1519-566X2011000100001>
- Mielke C.G.C. 1995. Papilionoidea e Hesperioidea (Lepidoptera) de Curitiba e seus arrededores, Paraná, Brasil, com notas taxonômicas sobre Hesperiiidae. *Revista Brasileira de Zoologia* 11 (4): 759–776. <https://doi.org/10.1590/S0101-81751994000400018>
- Mielke O.H.H. & Casagrande M.M. 1998. Papilionoidea e Hesperioidea (Lepidoptera) do Parque Estadual do Morro do Diabo, Teodoro Sampaio, São Paulo, Brasil. *Revista Brasileira de Zoologia* 14 (4): 967–1001. <https://doi.org/10.1590/S0101-81751997000400013>
- Mielke O.H.H., Carneiro E. & Casagrande M.M. 2010. Lepidopterofauna (Papilionoidea e Hesperioidea) do Parque Estadual do Chandless e arrededores, Acre, Brasil. *Biota Neotropica* 10 (4): 285–299. <https://doi.org/10.1590/S1676-06032010000400033>
- Miller L.D. 1968. *The Higher Classification, Phylogeny and Zoogeography of the Satyridae (Lepidoptera)*. Memoirs of the American Entomological Society 24. The American Entomological Society, Philadelphia.
- Miller L.D. 1970. Nomenclature of wing veins and cells. *Journal of Research on the Lepidoptera* 8 (2): 37–48.
- Monteiro A. & Pierce N. 2001. Phylogeny of *Bicyclus* (Lepidoptera: Nymphalidae) inferred from COI, COII and EF1a gene sequences. *Molecular Phylogenetics and Evolution* 18: 264–281. <https://doi.org/10.1006/mpev.2000.0872>
- Muschamp P.A.H. 1915. The ci-devant genus *Epinephele*. *Entomologist's Record and Journal of Variation* 27: 152–156.
- Nakahara S., Hall J.P.W., Lamas G. & Willmott K.R. 2015a. Seven new species and one new subspecies of *Euptychia* Hübner, 1818 (Lepidoptera: Nymphalidae: Satyrinae) from the tropical Andes. *Tropical Lepidoptera Research* 25 (2): 63–79.
- Nakahara S., Janzen D.H., Hallwachs W. & Espeland M. 2015b. Description of a new genus for *Euptychia hilara* C. Felder & R. Felder, 1867 (Lepidoptera, Nymphalidae, Satyrinae). *Zootaxa* 4012 (3): 525–541. <https://doi.org/10.11646/zootaxa.4012.3.7>
- Nakahara S., Barbosa E.P. & Freitas A.V.L. 2017. A potentially endangered new species of *Euptychia* Hübner, 1818 (Lepidoptera: Nymphalidae: Satyrinae) from the Atlantic coastal forest of Brazil. *Neotropical Entomology* 46 (3): 302–309. <https://doi.org/10.1007/s13744-016-0466-y>
- Nakahara S., McDonald J., Delgado F. & Padrón S.P. 2018a. Discovery of a rare and striking new pierid butterfly from Panama (Lepidoptera: Pieridae). *Zootaxa* 4527 (2): 281–291. <https://doi.org/10.11646/zootaxa.4527.2.9>
- Nakahara S., Willmott K.R., Mielke O.H.H., Schwartz J., Zacca T., Espeland M. & Lamas G. 2018b. Seven new taxa from the butterfly subtribe Euptychiina (Lepidoptera: Nymphalidae: Satyrinae) with revisional notes on *Harjesia* Forster, 1964 and *Pseudeuptychia* Forster, 1964. *Insecta Mundi* 0639: 1–38.
- Nakahara S., Zacca T., Huertas B., Neild A.F.E., Hall J.P.W., Lamas G., Holian L.A., Espeland M. & Willmott K.R. 2018c. Remarkable sexual dimorphism, rarity and cryptic species: a revision of the “*aegrota* species group” of the Neotropical butterfly genus *Caeruleuptychia* Forster, 1964 with the description of three new species (Lepidoptera, Nymphalidae, Satyrinae). *Insect Systematics and Evolution* 49 (2): 130–182. <https://doi.org/10.1163/1876312X-00002167>
- Nakahara S., Lamas G., Tyler S., Marín M.A., Huertas B., Willmott K.R., Mielke O.H.H. & Espeland M. 2019. A revision of the new genus *Amiga* Nakahara, Willmott & Espeland gen. n., described for

- Papilio arnaca* Fabricius, 1776 (Lepidoptera: Nymphalidae: Satyrinae). *ZooKeys*, 821: 85–152. <https://doi.org/10.3897/zookeys.821.31782>
- Nguyen L-T., Schmidt H.A., von Haeseler A. & Minh B.Q. 2015. IQ-TREE: A fast and effective stochastic algorithm for estimating maximum likelihood phylogenies. *Molecular Biology and Evolution* 32: 268–274. <https://doi.org/10.1093/molbev/msu300>
- Peña C. & Lamas G. 2005. Revision of the butterfly genus *Forsterinaria* Gray, 1973 (Lepidoptera: Nymphalidae, Satyrinae). *Revista peruana de Biología* 12 (1): 5–48. <https://doi.org/10.6084/M9.FIGSHARE.96638>
- Peña C., Nylin S., Freitas A.V.L. & Wahlberg N. 2010. Biogeographic history of the butterfly subtribe Euptychiina (Lepidoptera, Nymphalidae, Satyrinae). *Zoologica Scripta* 39 (3): 243–258. <https://doi.org/10.1111/j.1463-6409.2010.00421.x>
- Riley N.D. & Gabriel A.G. 1924. *Catalogue of the Type Specimens of Lepidoptera Rhopalocera in the British Museum. Part I. Satyridae*. Trustees of the British Museum, London. <https://doi.org/10.5962/bhl.title.118835>
- Robbins R.K., Lamas G., Mielke O.H.H., Harvey D.J. & Casagrande M.M. 1996. Taxonomic composition and ecological structure of the species-rich butterfly community at Pakitza, Parque Nacional del Manu, Perú. In: Wilson D.E. & Sandoval A. (eds) *Manu. The Biodiversity of Southeastern Peru*: 217–252. Smithsonian Institution, Washington DC.
- Silva A.R.M., Pontes D.V., Guimarães M.P., Oliveira M.V., Assis L.T.F. & Uehara-Prado M. 2015. Fruit-feeding butterflies (Lepidoptera: Nymphalidae) of the Área de Proteção Especial Manancial Mutuca, Nova Lima and species list for the Region of Belo Horizonte, Minas Gerais, Brazil. *Biota Neotropica* 15: 1–9. <https://doi.org/10.1590/1676-06032015011814>
- Strutzenberger P., Brehm G. & Fiedler K. 2012. DNA Barcode Sequencing from Old Type Specimens as a Tool in Taxonomy: A Case Study in the Diverse Genus *Eois* (Lepidoptera: Geometridae). *PLoS ONE* 7 (11): e49710. <https://doi.org/10.1371/journal.pone.0049710>
- Warren A.D., Davis K.J., Stangeland E.M., Pelham J.P. & Grishin N.V. 2018. *Illustrated Lists of American Butterflies*. Available from <http://www.butterfliesofamerica.com/> [accessed Mar. 2019].
- Westwood J.O. 1850–1852. The genera of diurnal Lepidoptera: comprising their generic characters, a notice of their habits and transformations, and a catalogue of the species of each genus. In: Doubleday E. *The Genera of Diurnal Lepidoptera: Comprising their Generic Characters, a Notice of their Habits and Transformations, and a Catalogue of the Species of each Genus*. Longman, Brown, Green & Longmans, London. <https://doi.org/10.5962/bhl.title.14057>
- Weymer G. 1911. 4. Familie: Satyridae: 173–283. In: Seitz A. (ed.) *Die Gross-Schmetterlinge der Erde* 5. A. Kernen, Stuttgart.
- Willmott K.R. 2003. *The Genus Adelpha: its Systematics, Biology and Biogeography (Lepidoptera: Nymphalidae: Limenitidini)*. Scientific Publishers, Gainesville.
- Willmott K.R., Marín M.A., Nakahara S., Pomerantz T., Lamas G., Huertas B., Espeland M., Xiao L., Hall J.P.W., Willmott J.I.R. & Freitas A.V.L. 2019. A revision of the new Andean butterfly genus *Optimandes* Marín, Nakahara & Willmott, n. gen., with the description of a new species (Nymphalidae: Satyrinae: Euptychiina). *Tropical Lepidoptera Research* 29 (1): 29–44. <https://doi.org/10.5281/zenodo.2650482>
- Zacca T., Paluch M., Siewert R., Freitas A., Barbosa E., Mielke O.H.H. & Casagrande M.M. 2017. Revision of *Godartiana* Forster (Lepidoptera: Nymphalidae), with the description of a new species from northeastern Brazil. *Austral Entomology* 56: 169–190. <https://doi.org/10.1111/aen.12223>

Zacca T., Casagrande M.M., Mielke O.H.H., Huertas B., Espeland M., Barbosa E., Freitas A.V.L., Magaldi L., Nakahara S. & Willmott K.R. 2018. Systematics of the butterfly genus *Cissia* Doubleday, 1848 (Lepidoptera: Nymphalidae, Satyrinae) using an integrative approach. *Arthropod Systematics & Phylogeny* 76 (2): 349–376.

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## Appendix

**Resumen.** Presentamos la primera revisión taxonómica integral del poco conocido género de mariposas sudamericanas *Zischkaia* Forster, 1964, hasta ahora considerado como incluyendo três especies descritas. Un análisis filogenético con base en los datos de secuencia de ADN muestra que *Zischkaia* es monofilético y consta morfológicamente de dos clados diagnosticables. Los caracteres morfológicos y los “códigos de barras” de ADN respaldan el reconocimiento de doce especies en el género, un aumento significativo para la subtribu Euptychiina que es relativamente poco estudiada. En consecuencia, aquí se describen y nombran nueve especies nuevas: *Z. arctoa* Nakahara, sp. nov., *Z. chullachaki* Nakahara & Zacca, sp. nov., *Z. baku* Zacca, Dolibaina & Dias, sp. nov., *Z. arenisca* Nakahara, Willmott & Hall, sp. nov., *Z. argyrosflecha* Nakahara, L. Miller & Huertas, sp. nov., *Z. abanico* Nakahara & Petit, sp. nov., *Z. josti* Nakahara & Kleckner, sp. nov., *Z. mielkeorum* Dolibaina, Dias & Zacca, sp. nov., y *Z. warreni* Dias, Zacca & Dolibaina, sp. nov.. Además, se designan un neotipo para *Satyrus pacarus* Godart, [1824], y los lectotipos para *Euptychia amalda* Weymer, 1911, *Euptychia fumata* Butler, 1867, y *Euptychia saundersii* Butler, 1867.

**Resumo.** É apresentada a primeira revisão taxonômica de *Zischkaia* Forster, 1964, um gênero de borboletas pouco conhecido da América do Sul composto, até o momento, por três espécies descritas. Uma análise filogenética baseada em sequências de dados de DNA demonstra que *Zischkaia* é monofilético e consiste de dois clados diagnosticáveis morfológicamente. Caracteres morfológicos e DNA “barcodes” suportam o reconhecimento de 12 espécies no gênero, um aumento significativo mesmo para a relativamente pouco estudada subtribo Euptychiina. Consequentemente, nove espécies novas são descritas e nomeadas no presente estudo, a citar: *Z. arctoa* Nakahara, sp. nov., *Z. chullachaki* Nakahara & Zacca, sp. nov., *Z. baku* Zacca, Dolibaina & Dias, sp. nov., *Z. arenisca* Nakahara, Willmott & Hall, sp. nov., *Z. argyrosflecha* Nakahara, L. Miller & Huertas, sp. nov., *Z. abanico* Nakahara & Petit, sp. nov., *Z. josti* Nakahara & Kleckner, sp. nov., *Z. mielkeorum* Dolibaina, Dias & Zacca, sp. nov., y *Z. warreni* Dias, Zacca & Dolibaina, sp. nov.. Adicionalmente, um neótipo é designado para *Satyrus pacarus* Godart, [1824] e lectótipos são designados para *Euptychia amalda* Weymer, 1911, *Euptychia fumata* Butler, 1867 e *Euptychia saundersii* Butler, 1867.