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# Linearized Siderophore Products Secreted via MacAB Efflux Pump Protect *Salmonella enterica* Serovar Typhimurium from Oxidative Stress

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**ABSTRACT** Nontyphoidal salmonellae (NTS) are exposed to reactive oxygen species (ROS) during their residency in the gut. To survive oxidative stress encountered during infection, salmonellae employ several mechanisms. One of these mechanisms involves the multidrug efflux pump MacAB, although the natural substrate of this pump has not been identified. MacAB homologs in pseudomonads secrete products of nonribosomal peptide synthesis (NRPS). In *Salmonella enterica* serovar Typhimurium, the siderophore enterobactin is produced by NRPS in response to iron starvation and this molecule can be processed into salmochelin and several linear metabolites. We found that *Salmonella* mutants lacking the key NRPS enzyme EntF are sensitive to peroxide mediated killing and cannot detoxify extracellular H<sub>2</sub>O<sub>2</sub>. Moreover, EntF and MacAB function in a common pathway to promote survival of *Salmonella* during oxidative stress. We further demonstrated that *S. Typhimurium* secretes siderophores in iron-rich media when peroxide is present and that these MacAB-secreted metabolites participate in protection of bacteria against H<sub>2</sub>O<sub>2</sub>. We showed that secretion of anti-H<sub>2</sub>O<sub>2</sub> molecules is independent of the presence of the known siderophore efflux pumps EntS and IroC, well-described efflux systems involved in secretion of enterobactin and salmochelin. Both salmochelin and enterobactin are dispensable for *S. Typhimurium* protection against ROS; however, linear metabolites of enterobactin produced by esterases IroE and Fes are needed for bacterial survival in peroxide-containing media. We determined that linearized enterobactin trimer protects *S. Typhimurium* against peroxide-mediated killing in a MacAB-dependent fashion. Thus, we suggest that linearized enterobactin trimer is a natural substrate of MacAB and that its purpose is to detoxify extracellular reactive oxygen species.

**IMPORTANCE** Nontyphoidal *Salmonella* bacteria induce a classic inflammatory diarrhea by eliciting a large influx of neutrophils, producing a robust oxidative burst. Despite substantial progress understanding the benefits to the host of the inflammatory response to *Salmonella*, little is known regarding how *Salmonella* can simultaneously resist the damaging effects of the oxidative burst. The multidrug efflux pump MacAB is important for survival of oxidative stress both *in vitro* and during infection. We describe a new pathway used by *Salmonella Typhimurium* to detoxify extracellular reactive oxygen species using a multidrug efflux pump (MacAB) to secrete a linear siderophore, a metabolite of enterobactin. The natural substrates of many multidrug efflux pumps are unknown, and functional roles of the linear metabolites of enterobactin are unknown. We bring two novel discoveries together to highlight an important mechanism used by *Salmonella* to survive under the oxidative stress con-

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ditions that this organism encounters during the classic inflammatory diarrhea that it also induces.

**KEYWORDS** MacAB, *Salmonella*, multidrug efflux pump, siderophores

**S**almonellae are the most common cause of bacterial foodborne disease in the United States and cause hundreds of millions of cases of foodborne disease, invasive nontyphoidal salmonellosis (iNTS), and enteric fever worldwide each year. Nontyphoidal salmonellae (NTS) are the cause of approximately 1.4 million cases/year of gastrointestinal disease in the United States, in the form of a classic inflammatory diarrhea. This inflammatory diarrhea is characterized by an abundance of neutrophils, elicited by CXC chemokines secreted in response to secretion of effectors of type III secretion system 1 (T3SS1) from NTS (1, 2). Thus, NTS must survive the arsenal of bactericidal mechanisms of neutrophils during the intestinal stage of salmonellosis. During systemic infection, *Salmonella* can be found in lesions in the liver and spleen. Inflammatory cells, including neutrophils and macrophages, are present in these lesions in abundance, and *Salmonella* replicate inside macrophages in a “*Salmonella*-containing vacuole” (SCV) (3, 4).

Salmonellae encode at least 11 multidrug efflux systems (ES) with partially characterized specificity for a number of antibiotics and biocides (5, 6). One of these, the ABC-type multidrug efflux system encoded by MacAB, is widely distributed among bacterial species (7, 8). MacAB systems are tripartite efflux systems (TES) that have been previously shown to be involved in the efflux of antibiotics, including macrolides (9). Although the structure and mechanism by which the MacAB pump functions have been carefully examined previously (10, 11), very few natural substrates of MacAB have been identified (heat-stable enterotoxin II and protoporphyrin PPIX [12, 13]). Deletion of the MacAB efflux system in *Salmonella enterica* serovar Typhimurium renders these organisms avirulent in BALB/c mice, suggesting that this efflux system plays an essential role in virulence (14). Further dissection of the role of MacAB during *Salmonella* infection suggests that this pump participates in the detoxification of extracellular peroxide and in survival of *S. Typhimurium* during exposure to reactive oxygen species (ROS) (15). We presume that the natural substrate of MacAB in *Salmonella* is required for resistance to oxidative stress, but the nature of this substrate has not yet been determined.

Reactive oxygen species generated by NADPH membrane oxidase complex 2 (Nox2) represent a key defense deployed by innate immune cells against bacterial infection. This enzyme complex generates reactive oxygen species that are released from phagocytes both into the extracellular environment and into phagosomes. Hypochlorous acid, superoxide, and hydrogen peroxide are all produced by phagocytes and function to kill microorganisms, although their mechanisms of killing have not been completely elucidated (16, 17). Hydrogen peroxide, produced from superoxide, can cross bacterial membranes and is bactericidal. In the Fenton reaction,  $H_2O_2$  reacts with intracellular iron sulfur clusters to produce damaging free radicals, including (among others) hydroxyl radicals (18), that can damage cellular components, including proteins, lipids, and DNA.

Bacterial pathogens possess numerous mechanisms to avoid and overcome the damaging effects of ROS. These mechanisms include direct detoxification of ROS by enzymes, including superoxide dismutases (SODs), catalases, and peroxidases. *S. Typhimurium* expresses cytosolic superoxide dismutases SodA and SodB, also present in *Escherichia coli*, as well as two periplasmic copper-zinc SODs, SodCI and SodCII (19, 20). SODs convert superoxide anion into  $H_2O_2$ . Enzymatic detoxification of hydrogen peroxide, in turn, depends on the activity of catalases and peroxidases. *S. Typhimurium* encodes three catalase enzymes, KatE, KatG, and KatN, and two peroxidases, AhpC and TsaA. These enzymes are highly redundant in their ability to degrade intracellular peroxide (21, 22). In addition, catecholate siderophores, metabolites produced by bacteria in response to iron limitation, were recently reported to play a role in

protection of *S. Typhimurium* from oxidative stress (23). In previous studies, *E. coli* mutants lacking *entE*, a subunit of enterobactin synthase, were more sensitive to oxidative killing, leading to the conclusion that enterobactin is required for protection from ROS (24, 25).

Here, we show that the antibiotic efflux system encoded by *macAB* is required for optimal *S. Typhimurium* colonization in the inflamed intestine and we identify the natural substrate of this efflux system. We show that mutants lacking *entF*, which are unable to perform nonribosomal peptide synthesis (NRPS) to generate enterobactin and its derivatives, are also sensitive to peroxide-mediated killing and cannot detoxify extracellular peroxide. In contrast to the dogma (26–28), siderophores were produced by *Salmonella* under iron-rich conditions when peroxide was included in the media. Our genetic dissection of the enterobactin processing pathways revealed that glucosylated products of enterobactin (salmochelin and its linearized forms) are not required for protection from peroxide, while esterases IroE and Fes are both required for this protection. Finally, we determined that the linearized enterobactin trimer (Ent-TRI) dihydroxybenzoylserine (DHBS)<sub>3</sub> protects *Salmonella* from peroxide-mediated killing when added exogenously or from conditioned media of the wild-type organism containing peroxide. We conclude that (DHBS)<sub>3</sub> is a natural substrate of MacAB and is a novel antioxidant responsible for enhancing *Salmonella* survival under inflammatory conditions during infection as well as during ROS exposure *in vitro*.

## RESULTS

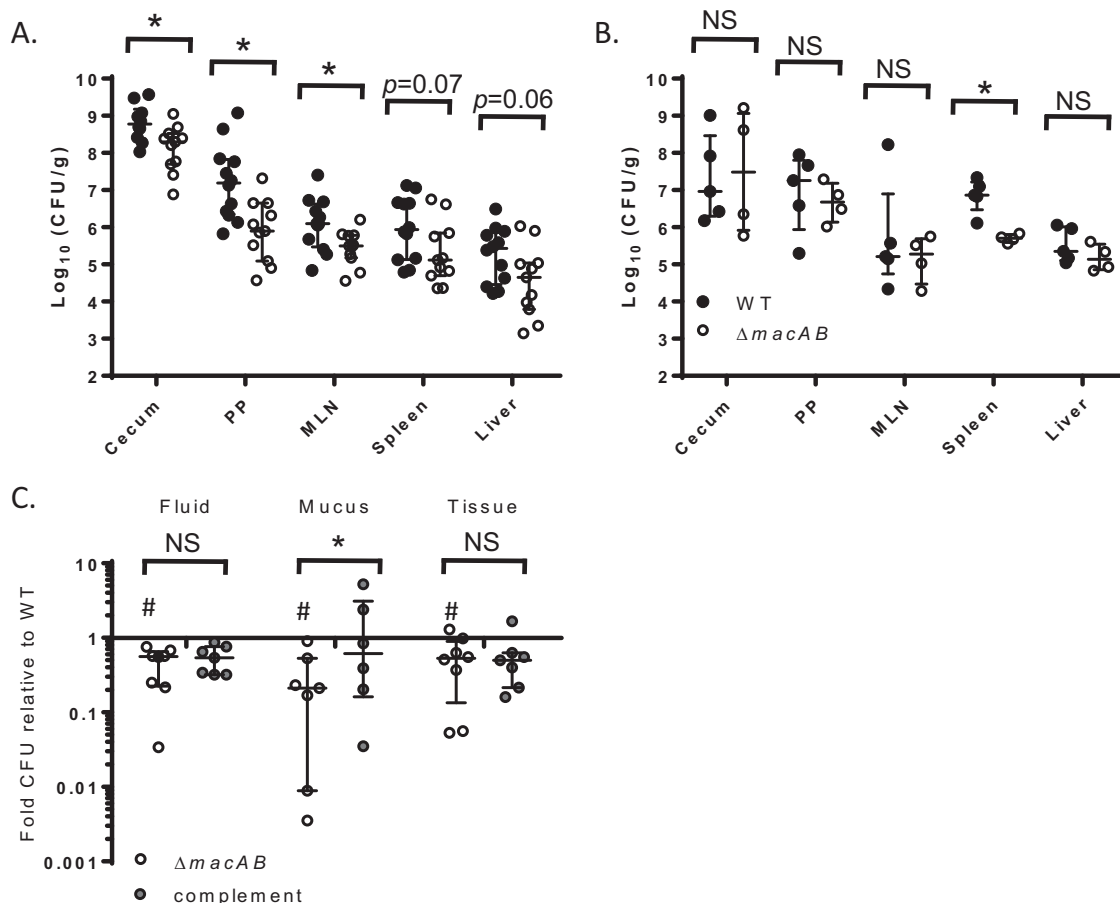
### The MacAB efflux pump is required for survival in the inflamed intestines.

Streptomycin treatment and subsequent *Salmonella* infection induce a strong neutrophilic influx into the intestinal tissue and lumen, mimicking the classical inflammatory diarrhea caused by NTS in humans (29). We have previously shown the importance of the MacAB efflux pump for *Salmonella* fitness in streptomycin-treated mice (15) (Fig. 1A), and the intestinal colonization defects of *macAB* mutants in this model are reversible by complementation in *trans* (Fig. S1). Furthermore, in C57Bl6 mice that were not pretreated with streptomycin, the MacAB efflux pump was dispensable for intestinal colonization (Fig. 1B). In untreated C57Bl6 mice,  $\Delta macAB$  mutants poorly colonized the spleen, a niche where salmonellae reside within macrophages (Fig. 1B). These data support the hypothesis that MacAB is necessary for surviving in murine models that develop an intestinal inflammatory response or in cells where reactive oxygen species play a prominent role.

In contrast to mice, calves are natural hosts for *Salmonella* that exhibit similar clinical and pathological signatures upon infection with *S. Typhimurium*, including neutrophilic infiltrates in the intestine, without any prior treatment (30). To further link the role of MacAB with survival of *Salmonella* during the inflammatory response in a host with an intact intestinal microbiota, we tested the fitness of a  $\Delta macAB$  mutant in bovine ligated ileal loops. The  $\Delta macAB$  mutant had a significantly reduced ability to replicate and/or survive in the luminal fluid, the mucus, and the intestinal tissue compared to the otherwise isogenic wild type (Fig. 1C). Complementation of the  $\Delta macAB$  mutant with intact copies of the *macAB* genes restored the ability of the resulting strain to survive in the intestinal mucus (Fig. 1C). These data further support the hypothesis suggesting the necessity for MacAB in surviving oxidative stress during infection.

### Production of enterobactin but not salmochelin is required for protection against an oxidative stress.

MacA and MacB in salmonellae share 34% to 47% identity and 51% to 62% similarity to the corresponding proteins in *Pseudomonas aeruginosa* PAO-1, where they are linked to the export of products of nonribosomal peptide synthesis (NRPS) (31). On this basis, we hypothesized that the function of MacAB in *Salmonella* could be similar to its function in *Pseudomonas*, to export products of NRPS. In *Salmonella*, the siderophore enterobactin is synthesized via NRPS. Enterobactin is a cyclic trimer of 2,3-hydroxybenzoyl serine, and the NRPS enzyme encoded by *entF* performs the last step in the synthesis of this molecule (32, 33). Enterobactin can be



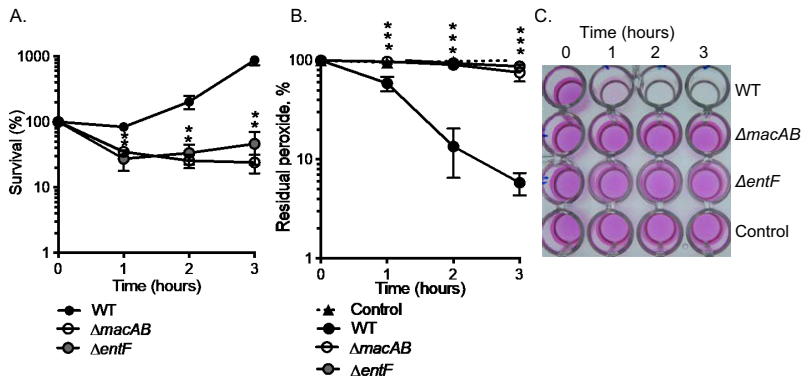
**FIG 1** MacAB is required for survival in the inflamed intestines. (A and B) C57BL/6 mice were treated with streptomycin 24 h before infection (A) or left untreated (B). Animals in both groups were orally infected with approximately  $10^8$  CFU of the wild-type strain (closed circles) or the  $\Delta macAB$  mutant (open circles). Cecum, Peyer's patches (PP), mesenteric lymph nodes (MLN), spleen, and liver were collected after 4 days of infection and analyzed for CFU enumeration. The asterisks indicate significance in Student's *t* test with *P* values of  $<0.05$ . NS, not significant. (C) Calf ligated ileal loops were inoculated with approximately  $10^7$  to  $10^8$  CFU of the wild-type strain or the  $\Delta macAB$  or  $\Delta macAB$  pWSK29-*macAB* mutant strain. Fluid, mucus, and ileal tissue were collected from each loop after 12 h of infection and analyzed for CFU enumeration. Data are expressed as fold growth relative to the wild type. The crosshatch symbol (#) indicates significance in Student's *t* test with *P* values of  $<0.05$ . The asterisk indicates statistical significance in 2-way analysis of variance (ANOVA) with Bonferroni's multiple-comparison test.

glucosylated into a second siderophore called salmochelin by IroB and/or further processed into several linear products (34, 35).

To determine whether the products of NRPS are linked to survival of *Salmonella* in the presence of reactive oxygen species (ROS), we evaluated the growth of an  $\Delta entF$  mutant in medium containing hydrogen peroxide (1 mM). Mutants lacking EntF cannot produce enterobactin or any of its downstream products. Under these growth conditions, 70% of the bacteria in the  $\Delta entF$  mutant culture were killed, a reduction in viability very similar to that seen with the  $\Delta macAB$  mutant under similar conditions (Fig. 2A). In contrast, the wild-type organism resumed growth after 2 h of exposure to peroxide (Fig. 2A).

Because bacteria lacking the MacAB transporter are unable to decrease the concentration of  $H_2O_2$  in the growth medium (15), we were interested in determining whether a mutant lacking *entF* would have a similar phenotype. Using the Amplex red assay, we determined that mutants lacking *entF* cannot detoxify extracellular peroxide (Fig. 2B and C). Collectively, these data indicate that *Salmonella* is sensitive to hydrogen peroxide in the absence of siderophore production and cannot reduce the amount of  $H_2O_2$  in the growth media.

Finally, enterobactin can be glycosylated in *S. Typhimurium* by the glycosyltransferase IroB to form a second siderophore called salmochelin (34). In order to determine



**FIG 2** Production of siderophores is required for protection against an oxidative stress. (A) Overnight cultures of the wild-type strain (WT) (black circles),  $\Delta macAB$  mutant (open circles), and  $\Delta entF$  mutant (gray circles) were subcultured at 1/100 in LB containing 1 mM  $H_2O_2$  and incubated at 37°C with aeration. Aliquots were collected hourly, serially diluted, and plated. The supernatants were retained for analysis in the experiments whose results are shown in panels B and C. Data are expressed as the survival means of results from at least three independent experiments with standard errors. (B and C) Supernatants collected from bacteria grown in peroxide-containing media as described in the panel A legend were tested for the residual  $H_2O_2$  concentration using Amplex red reagent (Invitrogen). Sterile  $H_2O_2$ -containing medium (black triangle, dotted line) was included as a control. Data are expressed as means of levels of residual peroxide in the media from at least three independent experiments with standard errors (B). (C) Image representative of the results of the Amplex red assay. The concentration of peroxide is directly proportional to the pink color development as a result of Amplex red oxidation. The asterisks in panels A and B indicate significance in Student's *t* test with *P* values of <0.05.

whether salmochelin plays a role in the protection of *Salmonella* from peroxide-mediated stress, we tested the ability of a mutant lacking *iroB* to resist peroxide-mediated killing and to detoxify extracellular peroxide. We found that the level of resistance of the salmochelin-deficient  $\Delta iroB$  mutant to peroxide-mediated killing was similar to that seen with the wild-type strain grown under similar conditions (Table 1 and Fig. S2A). In the Amplex red assay, the  $\Delta iroB$  mutant was able to efficiently reduce the concentration of peroxide in the growth medium to a level similar to that seen with wild-type strain (Table 1 and Fig. S3A). Collectively, our results support the hypothesis that it is enterobactin, or metabolites thereof, and not salmochelin or its metabolites which is involved in protection of *S. Typhimurium* against oxidative stress.

**Secretion of the antioxidant molecule is independent of EntS and IroC efflux pumps.** Secretion of enterobactin and salmochelin in *Salmonella* is mediated by two efflux pumps, EntS and IroC, respectively (36–38). We investigated whether (i) secretion of an antioxidant molecule during peroxide exposure relied on the presence of EntS and IroC and (ii) whether EntS and IroC efflux pumps act with MacAB in a common

**TABLE 1** Genetic analysis of siderophore-related mutants<sup>a</sup>

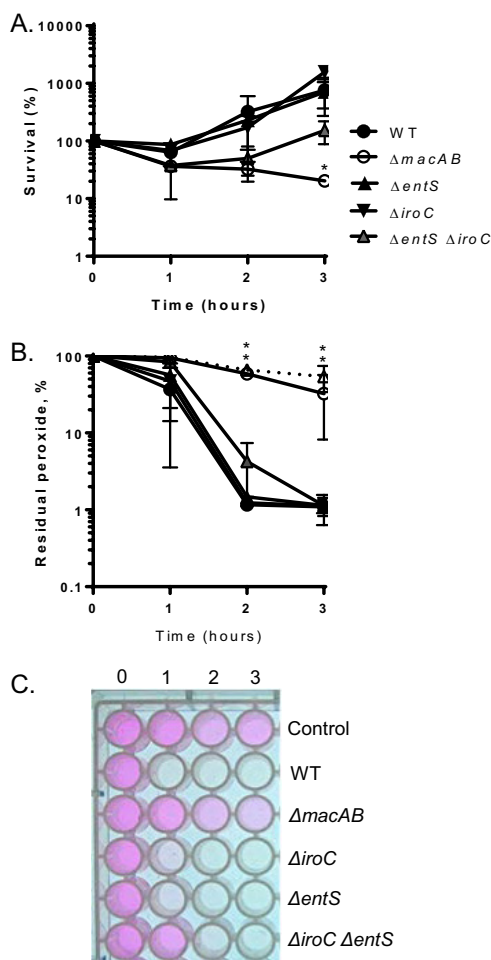
Strain	Biological function	$H_2O_2$ sensitivity <sup>c</sup>	Residual $[H_2O_2]$ <sup>d</sup>
<i>S. Typhimurium</i> WT		167.7 ± 37.4	5.5 ± 1.7
<i>S. Typhimurium</i> $\Delta macAB$	ABC-type efflux pump, substrate unknown	29.8 ± 6.3	81.6 ± 10.0
<i>S. Typhimurium</i> $\Delta entF$	Required for synthesis of (E), (S), and all linear metabolites <sup>b</sup>	27.3 ± 9.4	83.6 ± 5.8
<i>S. Typhimurium</i> $\Delta iroB$	Glucosyltransferase; converts (E) to (S)	83.3 ± 11.8	10.0 ± 8.1
<i>S. Typhimurium</i> $\Delta entS$	ABC-type efflux pump for (E) export	230.6 ± 27.5	1.2 ± 0.2
<i>S. Typhimurium</i> $\Delta iroC$	ABC-type efflux pump for (S) export	171.4 ± 65.6	1.5 ± 0.1
<i>S. Typhimurium</i> $\Delta iroD$	Esterase; converts (E) and (S) to linear metabolites	128.2 ± 39.7	6.9 ± 2.9
<i>S. Typhimurium</i> $\Delta iroE$	Esterase; converts (E) and (S) to linear metabolites	53.5 ± 13.6	28.9 ± 14.5
<i>S. Typhimurium</i> $\Delta fes$	Esterase; converts (E) and (S) to linear metabolites	58.6 ± 3.5	55.5 ± 14.0
<i>S. Typhimurium</i> $\Delta iroDE$		51.1 ± 9.0	23.4 ± 7.4
<i>S. Typhimurium</i> $\Delta iroDE \Delta fes$		21.0 ± 12.7	73.8 ± 16.9

<sup>a</sup>(E), enterobactin; (S), salmochelin; WT, wild type.

<sup>b</sup>Linear metabolites; see Fig. 6.

<sup>c</sup>After 2 h of growth in 1 mM  $H_2O_2$ , percent survival relative to *t*(0).

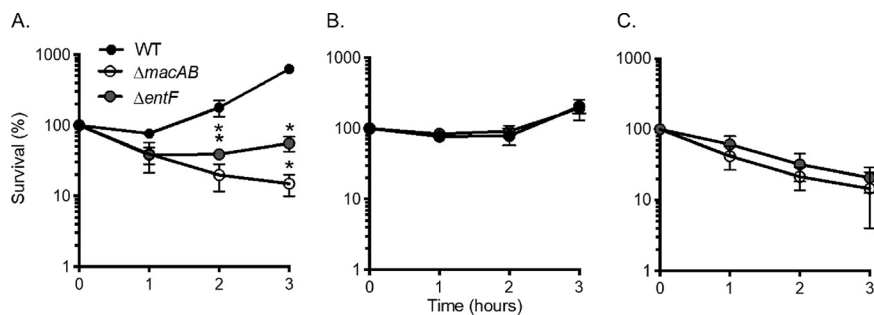
<sup>d</sup>After 2 h of growth in 1 mM  $H_2O_2$ , percent peroxide relative to *t*(0).



**FIG 3** Secretion of antioxidant molecule is independent of EntS and IroC efflux pumps. (A) Overnight cultures of the wild-type strain (black circles) and the  $\Delta macAB$  (open circles),  $\Delta entS$  (black upward-pointing triangle),  $\Delta iroC$  (black downward-pointing triangle), and  $\Delta entS \Delta iroC$  (gray upward-pointing triangle) mutants were subcultured at 1/100 in LB containing 1 mM  $H_2O_2$  and incubated at 37°C with aeration. Aliquots were collected hourly, serially diluted, and plated. The supernatants were retained for analysis in the experiments whose results are presented in panels B and C. Data are expressed as the survival means of results from at least three independent experiments with standard errors. (B and C) Supernatants collected from bacteria grown in peroxide-containing media as described in the panel A legend were tested for the residual  $H_2O_2$  concentration using Amplex red reagent (Invitrogen). Sterile  $H_2O_2$ -containing medium (black triangle, dotted line) was included as a control. Data are expressed as means of levels of residual peroxide in the media from at least three independent experiments with standard errors (B). Data corresponding to the y axis in panel B represent percent residual peroxide, and the data corresponding to the x axis represent time in hours. (C) Representative image of the Amplex red assay. The asterisks in panels A and B indicate significance in Student's *t* test with *P* values of <0.05.

pathway. We found that deletion of  $\Delta entS$  or  $\Delta iroC$  did not change the hydrogen peroxide sensitivity of the mutant strains compared to that of the wild type (Fig. 3A). Double mutants lacking both efflux pumps were not statistically different in peroxide sensitivity relative to the wild-type organism (Fig. 3A). In the Amplex red assay, both the single mutants with a mutation in  $\Delta entS$  or  $\Delta iroC$  and the double mutant  $\Delta entS \Delta iroC$  reduced the extracellular peroxide concentration with kinetics similar to the results seen with the wild-type organism (Fig. 3B and C).

To validate our findings, we cocultured the  $\Delta macAB$  mutant with the single or double mutations in siderophore efflux pumps in peroxide-containing media. We found that, similarly to the results seen with coculture experiments performed previously with wild-type *S. Typhimurium* (15), coculture with the  $\Delta entS$  (Fig. S4A and B),  $\Delta iroC$  (Fig. S4C and D), or  $\Delta entS \Delta iroC$  mutant (Fig. S4E and F) protected growth of the peroxide-sensitive  $\Delta macAB$  mutant strain in the medium containing 1 mM  $H_2O_2$ . These results



**FIG 4** *entF* and *macAB* function in the same genetic pathway. (A) Overnight cultures of the wild-type strain (black circles) and the  $\Delta macAB$  (open circles) and  $\Delta entF$  (gray circles) mutants were subcultured at 1/100 in LB containing 1 mM H<sub>2</sub>O<sub>2</sub> and incubated at 37°C with aeration. Aliquots were collected hourly, serially diluted, and plated for CFU enumeration. Data are presented as mean survival results from at least three independent experiments. (B and C) Individually grown overnight cultures of the wild-type strain and the  $\Delta entF$  mutant (B) or  $\Delta macAB$  and  $\Delta entF$  mutants (C) were mixed in equal ratios, and the resulting mixtures were subcultured in peroxide-containing LB and grown at 37°C with aeration. Aliquots were processed as described for panel A. The asterisk indicates significance in Student's *t* test with *P* values of <0.05.

indicate that secretion of an antioxidant molecule was taking place in the absence of EntS and IroC efflux pumps. Thus, we conclude that the MacAB efflux pump functions independently of known siderophore efflux systems EntS and IroC.

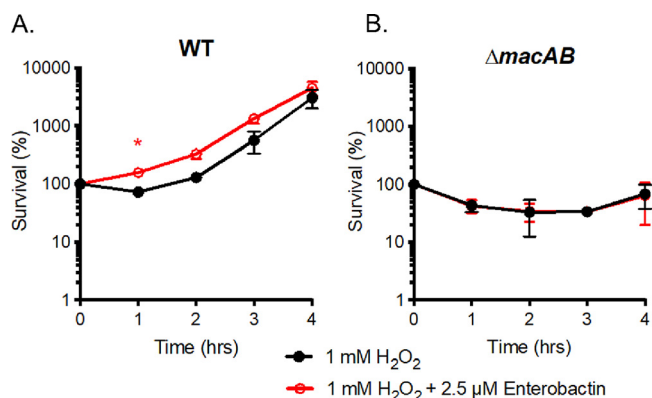
**For growth in and detoxification of extracellular peroxide, *entF* and *macAB* function in the same genetic pathway.** We investigated whether *entF* and *macAB* function in the same pathway or in parallel pathways to mitigate the effects of peroxide-mediated stress. Coculture with wild-type *Salmonella* protects  $\Delta macAB$  mutants from peroxide-mediated killing (15). In order to determine whether an *entF* mutant could be similarly protected by the presence of wild-type bacteria, we mixed individually grown overnight cultures of the wild type and the  $\Delta entF$  mutant in equal ratios and subcultured the mixture into fresh medium containing 1 mM H<sub>2</sub>O<sub>2</sub>.

In contrast to individual cultures grown in the presence of peroxide (Fig. 4A), the presence of the wild-type organism protected the  $\Delta entF$  mutant from peroxide-mediated killing (Fig. 4B). To rule out the possibility that *entF* and *macAB* function in parallel pathways, we mixed individually grown overnight cultures of  $\Delta macAB$  and  $\Delta entF$  mutant strains and tested the survival of both organisms in peroxide as described above. Coculture of the siderophore-deficient mutant with the  $\Delta macAB$  mutant failed to protect either strain from peroxide-mediated killing (Fig. 4C). Taken together, these experimental results indicate that EntF and MacAB function in a common pathway to mitigate the effects of peroxide-mediated stress.

**Exogenous enterobactin cannot rescue  $\Delta macAB$  mutant cells from peroxide-mediated death.** EntF is essential for the production of enterobactin, and, based on the results described above, the simplest hypothesis is that enterobactin itself might be responsible for mitigating peroxide-mediated effects on *S. Typhimurium*. In order to test this idea, we supplemented peroxide-containing growth medium with exogenous enterobactin and used this medium to test for survival and growth of various isolates. Supplementation of peroxide-containing medium with exogenous enterobactin promoted growth of wild-type *S. Typhimurium* (Fig. 5A). However, exogenous enterobactin added in a range of concentrations from 2.5 to 25  $\mu$ M failed to rescue the growth of the  $\Delta macAB$  mutant in the presence of 1 mM H<sub>2</sub>O<sub>2</sub> (Fig. 5B and data not shown). This result suggested that enterobactin itself is not protective during peroxide-mediated stress and is not the substrate of the MacAB efflux pump.

**Esterases Fes and IroE, but not IroD, are involved in protection of *S. Typhimurium* against hydrogen peroxide.** Enterobactin can be further hydrolyzed to linear metabolites by the esterases Fes, IroE, and IroD (Fig. 6). To determine which of these enzymes and, potentially, which linear metabolites play a role in protection of *Salmonella* against an oxidative stress, we generated deletion mutants in *fes*, *iroE*, and *iroD*.



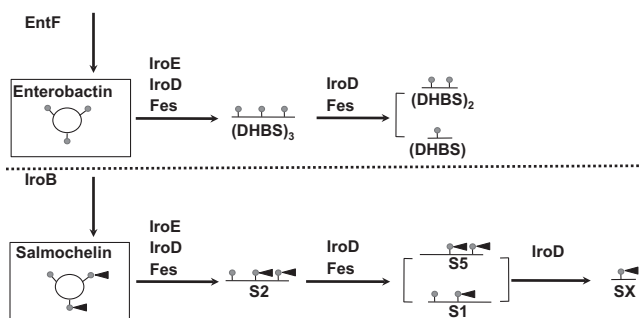


**FIG 5** Enterobactin cannot rescue the  $\Delta macAB$  mutant from peroxide-mediated death. (A and B) Overnight cultures of the wild-type strain (A) or the  $\Delta macAB$  mutant (B) were subcultured at 1/100 in peroxide-containing LB or in 1 mM  $H_2O_2$  with LB additionally supplemented with 2.5  $\mu M$  enterobactin and incubated at 37°C with aeration. Aliquots were collected hourly, serially diluted, and plated for CFU enumeration. Data are presented as mean survival results from at least three independent experiments. The asterisk indicates significance in Student’s *t* test with *P* values of <0.05.

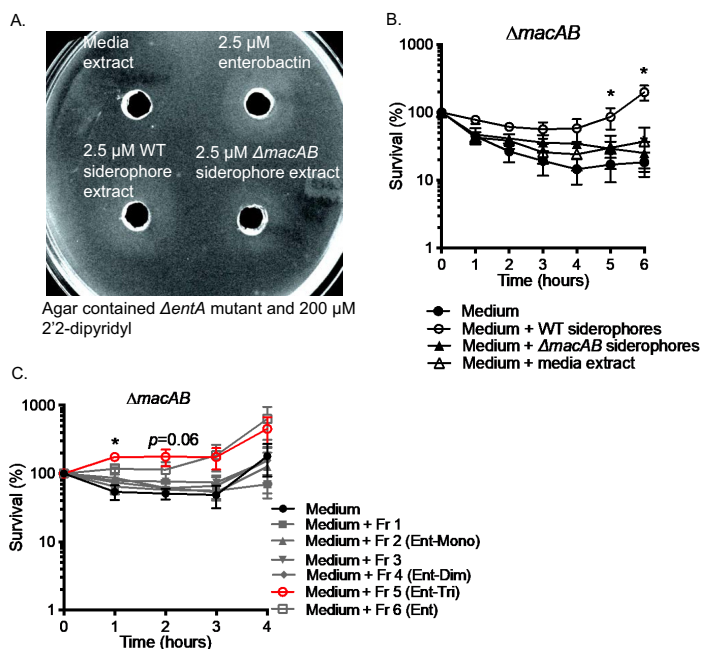
Each mutant strain was tested for two phenotypes associated with the loss of MacAB efflux pump: (i) sensitivity to hydrogen peroxide and (ii) the ability to degrade extracellular  $H_2O_2$  (15). First, as expected, the loss of the esterase IroD, which preferably hydrolyses iron-loaded salmochelin (39), did not affect *S. Typhimurium* sensitivity to peroxide or the ability to reduce concentration of  $H_2O_2$  in the medium (Table 1 and Fig. S2B and S3B). In contrast, deletion of *iroE* both moderately increased sensitivity to peroxide and slowed the degradation of  $H_2O_2$  in the medium in Amplex red assay (Table 1 and Fig. S2C and S3C). Similarly to the loss of IroE, the loss of Fes resulted in a moderate increase in peroxide sensitivity and led to a substantial delay in  $H_2O_2$  degradation in Amplex red assay (Table 1 and Fig. S2D and S3D). A double mutant lacking the ability to produce both IroE and Fes recapitulated the phenotype of a *macAB* mutant for both peroxide sensitivity and peroxide degradation (Table 1 and Fig. S2E and S3E). Taken together, these experiments indicate that esterases IroE and Fes, but not IroD, are involved the production of an enterobactin metabolite that has a function in protection of *S. Typhimurium* against hydrogen peroxide.

**Siderophores and their derivatives are produced in iron-rich medium, and enterobactin trimer (Ent-TRI) protects *S. Typhimurium* against oxidative stress.**

Siderophores are secreted by bacteria under iron-restricted conditions to acquire Fe(III) from the environment. Siderophores are also involved in protection of bacteria against oxidative stress (23) (Fig. 2). Luria-Bertani (LB) broth contains approximately 5.32 to 7.55  $\mu M$  iron (40, 41), an amount sufficient to maintain growth of siderophore-deficient



**FIG 6** Schematic overview of the siderophore pathway. DHBS, 2,3-dihydroxybenzoylserine, enterobactin monomer (Ent-Mono); (DHBS)<sub>2</sub>, enterobactin dimer (Ent-Dim); (DHBS)<sub>3</sub>, linear enterobactin trimer (Ent-TRI); SX, glucosylated 2,3-dihydroxybenzoylserine (pacifarin acid); S1 and S5, glucosylated enterobactin dimers; S2, glucosylated linear enterobactin trimer.



**FIG 7** Siderophores and their derivatives are produced in iron-rich media, and Ent-TRI protects *Salmonella* against oxidative stress. (A) Siderophores extracted from the conditioned broth with the wild-type strain and the  $\Delta macAB$  mutant exposed to peroxide were tested in cross-feeding assays for their ability to support growth of siderophore-deficient  $\Delta entA$  mutant cells on the plate containing iron chelator 2'-2-dipyridyl (200  $\mu M$ ). Purified enterobactin (2.5  $\mu M$ ) and mock siderophore extract were used as the positive control and negative control, respectively. A halo around a well indicates the presence of siderophore in the test sample. The picture was taken after 24 h of incubation at 37°C. (B) An overnight culture of the  $\Delta macAB$  mutant was subcultured at 1/100 in peroxide-containing LB (black circles) or in the same media additionally supplemented with siderophore extract from the media containing the wild type (open circles), from  $\Delta macAB$ -conditioned media (black triangles), and from mock extract (open triangles) and incubated at 37°C with aeration. Aliquots were collected hourly, serially diluted, and plated for CFU enumeration. (C) An overnight culture of  $\Delta macAB$  mutant was subcultured at 1/100 in peroxide-containing LB (black circles) or in the same media additionally supplemented with HPLC-separated fractions (Fr) of siderophore extract from wild-type-conditioned media (as described in Materials and Methods). The resulting cultures were incubated at 37°C with aeration. Aliquots were processed as described for panel B. Data in panels B and C are presented as mean survival results from at least three independent experiments. The asterisk indicates the significance in Student's *t* test with *P* values of <0.05.

mutants (42). To show that siderophores are indeed secreted under iron-rich conditions and under conditions of oxidative stress, we grew wild-type *Salmonella* or the  $\Delta macAB$  mutant strain in LB broth containing 0.5 mM H<sub>2</sub>O<sub>2</sub> and isolated siderophores from the resulting conditioned media, which had been filtered to remove bacteria. The concentration of catecholate siderophores in the conditioned media was determined by Arnov assay and adjusted to 2.5  $\mu M$  (43).

Cross-feeding assays were used to determine whether extracts of the conditioned medium described above contained siderophores in concentrations sufficient to rescue the growth of siderophore-deficient  $\Delta entA$  mutants. Siderophore extracts prepared from both the wild-type-conditioned media and the  $\Delta macAB$  mutant-conditioned media, but not a “mock” extract from uninoculated LB broth, supported the growth of siderophore-deficient  $\Delta entA$  mutants on agarized medium containing the iron chelator 2'-2-dipyridyl (200  $\mu M$ ) (Fig. 7A). This result suggests that siderophores are secreted by *Salmonella* when peroxide is present in the medium, even under iron-rich conditions.

Next, we investigated whether siderophores in these extracts would be able protect an *S. Typhimurium*  $\Delta macAB$  mutant against hydrogen peroxide-mediated death. First, we confirmed that the extracts were not toxic to *Salmonella* (Fig. S5). Despite the presence of siderophores in extracts prepared from both wild-type-conditioned and  $\Delta macAB$  mutant-conditioned media (Fig. 7A), only the extract from the wild type had

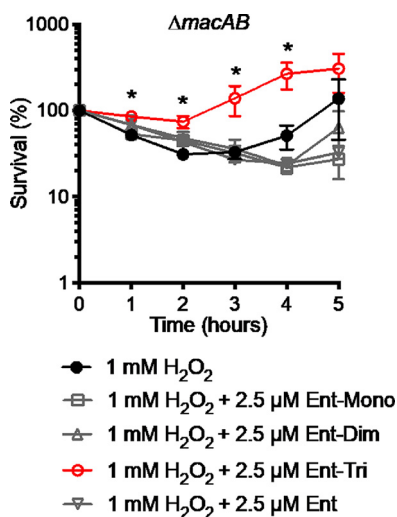
a protective effect on the growth of the  $\Delta macAB$  mutant in peroxide-containing media (Fig. 7B). This finding suggests that enterobactin and salmochelin, secreted via the ABC-type transporters IroC and EntS, are not themselves the critical molecules for protection that are being secreted by *Salmonella* against oxidative stress. We hypothesized that derivatives of enterobactin could be playing a role in the protection of *Salmonella* against oxidative stress.

To identify the enterobactin derivatives responsible for protection against reactive oxygen species, we separated siderophore extracts by high-pressure liquid chromatography (HPLC) and collected fractions expected to contain the following enterobactin derivatives: enterobactin monomer (Ent-Mono; 2,3-dihydroxybenzoylserine [DHBS]) (fraction 2); enterobactin dimer [Ent-Dim; (DHBS)<sub>2</sub>] (fraction 4); enterobactin trimer [Ent-TRI; (DHBS)<sub>3</sub>] (fraction 5), and enterobactin cyclic trimer (fraction 6). The enterobactin derivatives expected in each fraction were determined in advance by running purified commercially available derivatives of enterobactin on HPLC to determine their retention times. For negative controls, we collected two fractions (fractions 1 and 3) that were not expected to contain any siderophores. Each fraction was tested for its ability to rescue the hydrogen peroxide sensitivity of the  $\Delta macAB$  mutant. Only fraction 5 prepared from the wild-type siderophore extract, expected to contain Ent-TRI, had a statistically significant protective effect on the survival of the  $\Delta macAB$  mutant grown in the presence of peroxide (Fig. 7C). In contrast, the corresponding fraction prepared from  $\Delta macAB$  siderophore extract failed to support growth of the  $\Delta macAB$  mutant in peroxide-containing medium (data not shown).

**Exogenous linearized enterobactin protects the *S. Typhimurium*  $\Delta macAB$  mutant from hydrogen peroxide-mediated killing.** To confirm whether purified enterobactin derivatives could protect a *macAB* mutant from peroxide-mediated killing, we used commercially available purified enterobactin derivatives, i.e., Ent-Mono, Ent-Dim, Ent-TRI (EMC Microcollections), and Ent (Sigma), in a peroxide survival assay. Each siderophore derivative was used at 2.5  $\mu$ M. In agreement with our previous observations (Fig. 7C), neither Ent-Mono nor Ent-Dim was able to rescue growth of the peroxide sensitive  $\Delta macAB$  mutant in medium containing 1 mM H<sub>2</sub>O<sub>2</sub>. In contrast, supplementation of peroxide-containing medium with 2.5  $\mu$ M Ent-TRI significantly improved the survival of the  $\Delta macAB$  mutant (Fig. 8). The results of this experiment suggest that linearized enterobactin trimer is the substrate of MacAB efflux pump and that this molecule can protect *Salmonella* from damaging reactive oxygen species.

**Linearized enterobactin protects *S. Typhimurium*  $\Delta iroE \Delta fes$  esterase mutants from hydrogen peroxide.** Given that active esterases IroE and Fes are needed for protection of *S. Typhimurium* against peroxide-mediated damage (Table 1 and Fig. S2E and S3E), we investigated whether they work in the same pathway with the MacAB efflux pump. As expected, the  $\Delta iroE \Delta fes$  mutant was more sensitive to peroxide-mediated killing than the wild type (Fig. S6A). Coculture of the  $\Delta iroE \Delta fes$  mutant with the wild type in the presence of H<sub>2</sub>O<sub>2</sub> resulted in improved survival of the mutant (Fig. S6B). In contrast, coculture of the  $\Delta iroE \Delta fes$  and the  $\Delta macAB$  mutants in the presence of peroxide did not improve the survival of esterase mutant (Fig. S6C). These results indicate that IroE and Fes function in the same pathway as MacAB.

We hypothesize that the linearized enterobactin, Ent-TRI, a compound that protects the  $\Delta macAB$  mutant from peroxide-mediated killing (Fig. 8), would also protect the  $\Delta iroE \Delta fes$  esterase mutant. Similarly to the  $\Delta macAB$  mutant (Fig. 9C), supplementation of peroxide-containing LB broth with 2.5  $\mu$ M Ent-TRI significantly improved the survival of the  $\Delta iroE \Delta fes$  double mutant (Fig. 9B). To ensure that Ent-TRI selectively protects mutants with mutations in genes that belong to MacAB pathway, we tested the effect of linearized enterobactin on the unrelated peroxide-sensitive  $\Delta recA$  mutant. Mutation in *recA* abrogates the ability of *Salmonella* to repair peroxide-damaged DNA, and this eventually results in bacterial death. Supplementation of peroxide-containing media with Ent-TRI did not improve the survival of the  $\Delta recA$  mutant (Fig. 9D). Taken together, these experiments demonstrated that linearized enterobactin trimer, secreted by *Sal-*



**FIG 8** Exogenous linearized enterobactin protects  $\Delta macAB$  mutant cells from hydrogen peroxide-mediated killing. An overnight culture of the  $\Delta macAB$  mutant was subcultured at 1/100 in peroxide-containing LB (black circles) or in the same media additionally supplemented with enterobactin monomer (gray open squares), enterobactin dimer (gray open upward-pointing triangles), enterobactin trimer (open circles), and enterobactin (gray open downward-pointing triangles) and incubated at 37°C with aeration. Aliquots were collected hourly, serially diluted, and plated for CFU enumeration. Data are presented as mean survival results from at least three independent experiments. Asterisks indicate significance in Student's *t* test with *P* values of <0.05.

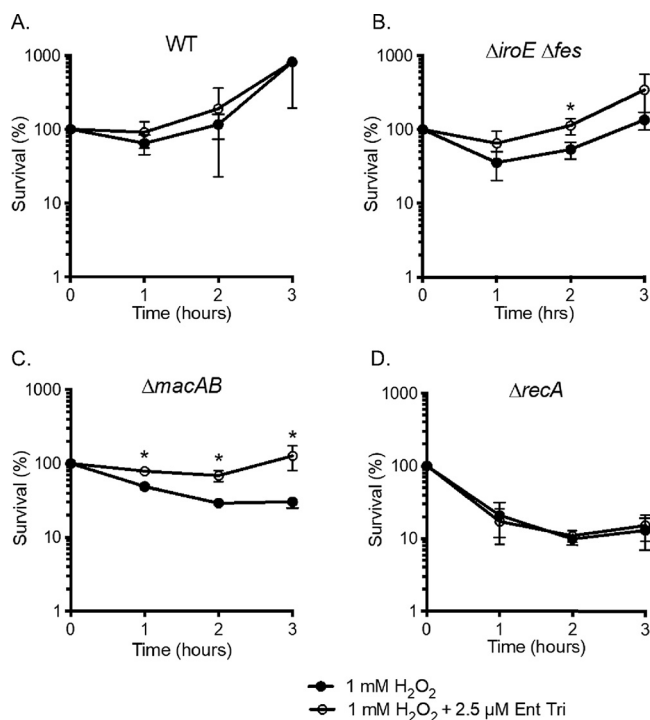
*monella* under iron-rich conditions when peroxide is present, protects *S. Typhimurium* against peroxide-mediated killing in a MacAB-dependent fashion.

## DISCUSSION

Multidrug efflux pumps play an important role in survival of bacteria during antibiotic therapy (44). These pumps are usually chromosomally encoded membrane protein complexes that belong to the ancient families of proteins (45). Multiple efflux pumps with overlapping specificities to drugs are usually present in the single bacterial genome, which suggests that these protein complexes may have additional natural functions unrelated to the role in efflux of antimicrobials.

We showed previously that MacAB, an ABC-type efflux pump, originally linked to macrolide resistance and later to virulence in mice (9, 14), is involved in protection of *Salmonella* Typhimurium against oxidative stress (15). Reactive oxygen species are generated by the mammalian host in response to bacterial infection (46). To model human enteritis in response to *Salmonella* infection, we utilized two animal models: streptomycin-treated mice that develop colitis in response to infection and ligated ileal loops in calves, a model that closely mimics the clinical and pathological changes occurring during the early phase of salmonellosis. In agreement with earlier work (15), the mutant lacking MacAB was attenuated in both models in the inflamed intestines, indicating that this pump facilitates survival of *Salmonella* during infection.

The MacAB efflux pump was previously linked to secretion of heat-stable enterotoxin II produced by enterotoxigenic *E. coli* (12) and was suggested to be involved in export of the heme precursor protoporphyrin PPIX in *E. coli* when bacterial heme homeostasis is disrupted by iron starvation (13). MacAB homologs in *Pseudomonas* species were previously shown to be involved in secretion of lipopeptides (arthrofactin, putisolvins I and II, xantholysin, syringomycin, and syringopeptin) (47–50) and the siderophore pyoverdinin (31). Both lipopeptide and siderophore molecules are synthesized nonribosomally through the function of multimodular nonribosomal peptide synthetases (NRPS). In the genomes of these bacteria, *macAB* genes are located downstream of the NRPS cluster. The *Salmonella* genome encodes a single NRP synthetase (EntF) that is necessary for siderophore synthesis (51). The genes that



**FIG 9** Linearized enterobactin protects  $\Delta$ *iroE*  $\Delta$ *fes* mutant cells from hydrogen peroxide-mediated killing. Overnight cultures of the wild-type strain (A) and the  $\Delta$ *iroE*  $\Delta$ *fes* (B),  $\Delta$ *macAB* (C), and  $\Delta$ *recA* (D) mutants were subcultured at 1/100 in peroxide-containing LB (black circles) or in the same media additionally supplemented with enterobactin trimer (open circles) and incubated at 37°C with aeration. Aliquots were collected hourly, serially diluted, and plated for CFU enumeration. Data are presented as mean survival results from at least three independent experiments. Asterisks indicate significance in Student's *t* test with *P* values of <0.05.

encode MacAB efflux pump, however, are located in an independent region of the genome. Nevertheless, the loss of EntF led to a drastic increase in hydrogen peroxide sensitivity and, similarly to the loss of MacAB efflux pump (15), to an inability to degrade extracellular H<sub>2</sub>O<sub>2</sub>. Under iron-limiting conditions, *Salmonella* produces two siderophores, enterobactin and its glycosylated derivative, salmochelin (35).

Catecholates siderophores were recently proposed to have an antioxidant role in addition to their well-established role in iron chelation (23–25), and it was also proposed that there is a link between the components of oxidative stress response and siderophore biosynthesis (52). In agreement with previous observations (23), the production of salmochelin was dispensable for *Salmonella* survival in peroxide. Surprisingly, the addition of exogenous enterobactin did not protect  $\Delta$ *macAB* mutants against peroxide-mediated killing. This finding allowed us to conclude that enterobactin is not the natural substrate of MacAB, supported by the fact that the pathway for enterobactin export via EntS is well established (37).

Enterobactin can be further processed by esterases IroD, IroE, and Fes to produce shorter linear products (39). Even though these proteins are homologous to each other, they differ in their substrate specificities and in their cellular locations (39, 53). IroD is located in the cytoplasm and cleaves iron-loaded salmochelin (39). We found that IroB, the enzyme that glycosylates enterobactin to form salmochelin (54), was not required for protection of *Salmonella* against peroxide. We also found that the dedicated transporter for salmochelin, IroC (36), is not required for protection from ROS. Finally, the loss of IroD did not affect *S. Typhimurium* survival in the presence of peroxide. Thus, neither salmochelin nor its degradation products (S2, S1, and SX) could be a substrate of MacAB.

Fes and IroE, located in the cytoplasm and periplasm, respectively, hydrolyze both enterobactin and salmochelin (39). In addition, IroE hydrolyses iron-free siderophores to

produce linearized trimer (DHBS)<sub>3</sub>, while Fes prefers iron-loaded siderophores to sequentially produce linearized trimer, dimer, and monomer degradation products. We found that both IroE and Fes are necessary to provide protection against hydrogen peroxide similar to the protection provided by organisms with intact MacAB and EntF systems. Consistent with these findings, *entE* and *fes* are also required for protection from oxidative killing in *E. coli*, although *E. coli* does not encode a homolog of IroE (24). Thus, both synthesis of enterobactin and its processing to a linear product are required for protection against ROS. Finally, a considerable amount of the linear products produced by the action of these esterases had long been known to be secreted from the bacterial cell, and yet the machinery required for their export remained unknown (36).

One cannot abrogate the production of endogenously produced enterobactin in *S. Typhimurium* to define the importance of this metabolite during infection, because it is a precursor to salmochelin and other linear products. However, several lines of evidence suggest that the primary purpose of enterobactin production by *Salmonella* during infection may not be its siderophore activity. First, enterobactin is produced by many *Enterobacteriaceae* (55) and is present in the murine intestine, where iron is not limited (55, 56). Second, deletion of *entS*, a gene encoding a key protein involved in the export of enterobactin, does not significantly reduce *Salmonella* virulence in murine models (36). Third, lipocalin-2, a host molecule secreted by several cell types, including neutrophils, binds iron-loaded enterobactin to sequester it from bacteria but is unable to bind salmochelin (57). Finally, while IroN and FepA, receptors on the cell surface that bind enterobactin prior to import, reduce *Salmonella* virulence when deleted (58), these receptors may also be involved in siderophore piracy of enterobactin produced by other enterobacteriaceae in the intestine and other siderophores, including corynebactin and myxochelin (59). Our data suggest that enterobactin serves not only as a precursor for salmochelin but also as a precursor for the production of linear products of enterobactin that are secreted via MacAB and whose main purpose may be neutralizing reactive oxygen species outside the bacterial cell during infection.

While the precise mechanism of this anti-H<sub>2</sub>O<sub>2</sub> protection is not yet defined, there are at least two possible scenarios: linearized enterobactin trimer can either exhibit catalase-like activity or participate in ROS scavenging. An unrelated siderophore, yersiniabactin, complexed with copper, Cu(II)-Ybt, was recently shown to have superoxide dismutase (SOD)-like activity, supporting survival of uropathogenic *E. coli* within macrophages (60). Another example of SOD mimic compounds includes metalloporphyrins and salens that also possess catalase-like activity (61–63). Among all the compounds mentioned above, catalytic activity depends on the presence of redox-active metal: copper, manganese, or iron. The requirement for iron in enterobactin trimer-mediated peroxide detoxification is currently unknown. The second potential mechanism for Ent-TRI detoxification of reactive oxygen species is based on the well-known antioxidant activity of catechol moieties (64–68). These moieties are present and exposed that in the linearized enterobactin trimer. Elucidation of the detailed mechanism of the antiperoxide action of this molecule will be a fascinating area of future work.

## MATERIALS AND METHODS

**Bacterial strains and media.** All of the *Salmonella enterica* serovar Typhimurium strains used for this study are listed in Table 2 and were derived from ATCC 14028. Mutants were generated by lambda-Red recombinase-mediated homologous recombination (69, 70). HA420 is a fully virulent, spontaneously nalidixic acid-resistant (Nal<sup>r</sup>) derivative of ATCC 14028 (71). Deletion mutants used in this study were moved into a clean genetic background using P22 transduction (72).

Strains were routinely cultured in Luria-Bertani (LB) broth and plates, supplemented with antibiotics as needed at the following concentrations: 50 mg/liter nalidixic acid, 100 mg/liter carbenicillin, 50 mg/liter kanamycin, or 20 mg/liter chloramphenicol. For calf ligated ileal loop infections, strains were grown overnight at 37°C with aeration (200 rpm) in LB broth with appropriate antibiotic, subcultured 1/100 in fresh media, and grown for additional 3 h at 37°C. Bacteria were pelleted by centrifugation and washed twice in sterile LB, and the bacterial concentration was adjusted to approximately 10<sup>7</sup> to 10<sup>8</sup> CFU in 3 ml LB. For infection of *Salmonella*-susceptible C57BL/6 mice, strains were grown aerobically at 37°C to stationary phase in LB broth with an appropriate antibiotic.

**TABLE 2** Strain and plasmid list

Plasmid	Strain <sup>a</sup>	Reference or source
HA420	ATCC 14028, spontaneously Nal <sup>r</sup>	71
HA995	ATCC 14028 $\Delta macAB::Cm^r$	14
LB321	ATCC 14028 $\Delta macAB::Cm^r$ pWSK29- <i>macAB</i> , Amp <sup>r</sup>	This study
LB329	HA420 $\Delta entF::Kan^r$	This study
JE384	HA420 $\Delta iroB::Kan^r$	This study
JE374	HA420 $\Delta entS::Kan^r$	This study
JE418	HA420 $\Delta iroC::Cm^r$	This study
JE419	HA420 $\Delta entS::Kan^r$ $\Delta iroC::Cm^r$	This study
JE430	HA420 $\Delta iroD::Kan^r$	This study
JE424	HA420 $\Delta fes::Cm^r$	This study
LB437	HA420 $\Delta iroE::Cm^r$	This study
LB444	HA420 $\Delta iroDE::Kan^r$	This study
LB649	HA420 $\Delta iroE::Kan^r$ $\Delta fes::Cm^r$	This study
LB500	HA420 $\Delta iroDE::Kan^r$ $\Delta fes::Cm^r$	This study
LB786	ATCC 14028 $\Delta entA::Kan^r$	This study
LB250	HA420 $\Delta recA-hydN::Kan^r$	15

<sup>a</sup>Amp<sup>r</sup>, ampicillin resistance; Cm<sup>r</sup>, chloramphenicol resistance; Kan<sup>r</sup>, kanamycin resistance; Nal<sup>r</sup>, nalidixic acid resistance.

**Plasmid construction.** A complementing plasmid carrying intact *macAB* genes was generated as follows. A DNA fragment containing the full-length open reading frames with approximately 300 bp upstream and downstream of *macAB* was amplified by PCR using *macAB*-EcoRI-Fwd (5'-GCGAATTCAGTAACGTATTTAACTCC-3') as a forward primer and *macAB*-KpnI-Rev (5'-ATGGTACCAAGTGGTTCAACAATGCC-3') as a reverse primer. PCR products were digested with EcoRI and KpnI (New England Biolabs) and ligated into pWSK29 vector (73) that had been previously digested with the same enzymes. Clones with the correct insertion were confirmed by restriction digestion and sequencing.

**Calf ligated ileal loop infection model.** All experiments involving animals described in this work were carried out in accordance with the recommendations in the Guide for the Care and Use of Laboratory Animals of the National Institutes of Health and were approved by the Institutional Animal Care and Use Committee at Texas A&M University or at North Carolina State University. *Salmonella* infections were performed as previously described (74). Briefly, Angus cross calves were separated from the dam at 1 day of age and fed milk replacement twice a day and were provided with access to water and grass hay. Fecal samples were collected weekly and were tested for the presence of *Salmonella* as described previously (30, 75). When they reached 3 to 6 weeks of age, the calves were anesthetized for ligated ileal loop surgery and ileal loops were tied and injected with 3 ml of approximately 10<sup>7</sup> to 10<sup>8</sup> CFU of wild-type or  $\Delta macAB$  mutant *Salmonella* Typhimurium or with 3 ml of sterile LB broth. Ligated loops were returned to the abdominal cavity, incisions were closed, and calves were kept under anesthesia for the duration of the experiment. At the end of incubation period, individual loops were excised. Calves were humanely euthanized after all loops were excised as described previously (74). Intestinal fluid, mucus, and tissue samples were collected from each loop. Fluid volume, which correlates with the level of inflammation in the intestines (76), was determined by weighing each loop before and after fluid collection. Mucus was scraped from the luminal surface of the intestine and placed in 3 ml of sterile phosphate-buffered saline (PBS). The remaining tissue was placed in 5 ml PBS. All specimens were homogenized (IKA T25 Basic S1), serially diluted in PBS, and plated for CFU enumeration. To compare the bacterial burden in the loops infected with wild-type *Salmonella* to the CFU from the loops infected with mutant strain, data were expressed as fold growth as follows:  $[CFU(t_{12})/CFU(t_0)]_{mutant}/[CFU(t_{12})/CFU(t_0)]_{wild\ type}$ . Statistical significance was determined using Student's two-tailed *t* test with significance set at a *P* value of <0.05.

**Oral infection of *Salmonella*-susceptible mice.** A *macAB* null mutant and a virulent HA420 strain (spontaneously Nal<sup>r</sup>) were tested for their ability to colonize 8-to-10-week-old female C57BL/6 mice (Jackson Laboratory) using the following protocol. Overnight cultures to be used as inocula were grown at 37°C with aeration, serially diluted, and plated for CFU to determine the exact titer.

Groups of five mice were infected by gavage with approximately 1 × 10<sup>8</sup> bacteria of either the wild-type strain or the  $\Delta macAB$  mutant strain in 100 μl of LB broth. At 4 days postinfection, mice were humanely euthanized and livers, spleens, Peyer's patches, mesenteric lymph nodes, and ceca of infected mice were excised and homogenized in 3 ml PBS. Organ homogenates were weighed, serially diluted, and plated to determine bacterial burden in the infected tissues. Data are expressed as CFU counts per gram of tissue converted to a logarithmic scale. Statistical significance was determined using a Student's *t* test and *P* values set to less than 0.05.

**Oral infections in the murine colitis model.** Female C57BL/6 mice (Jackson Laboratories) (8 to 10 weeks of age) were treated with 20 mg of streptomycin by gavage 24 h prior to infection. The wild-type strain (HA420), the  $\Delta macAB$  mutant, and the  $\Delta macAB$  mutant complemented with an intact copy of *macAB* were grown to stationary phase at 37°C with aeration. Cultures were serially diluted and plated to determine the exact titer of each strain used to inoculate animals.

Groups of five mice were inoculated by gavage with approximately 1 × 10<sup>8</sup> bacteria in 100 μl. At 4 days postinfection, mice were humanely euthanized and livers, spleens, Peyer's patches, mesenteric

lymph nodes, and ceca of infected mice were excised and homogenized in 3 ml of PBS and serially diluted, and bacteria were enumerated. Data are expressed as CFU counts per gram of tissue, converted logarithmically. Statistical significance was determined using a Student's *t* test and *P* values set less than 0.05.

**Sensitivity of individual isolates to hydrogen peroxide.** Overnight cultures were subcultured at 1/100 in LB with appropriate antibiotics, with or without 1 mM H<sub>2</sub>O<sub>2</sub> (VWR). Subcultures were incubated at 37°C with aeration. Aliquots were collected every hour for CFU determination. Results were expressed as percent survival calculated as [CFU(*t<sub>n</sub>*)/CFU(*t<sub>0</sub>*)] \* 100 over time. Experiments were performed on at least three separate occasions. Statistical significance was determined using a Student's *t* test.

**Hydrogen peroxide detection with Amplex red.** Overnight cultures were subcultured at 1/100 in LB broth with appropriate antibiotics containing 1 mM H<sub>2</sub>O<sub>2</sub> (VWR). Uninoculated LB broth containing 1 mM H<sub>2</sub>O<sub>2</sub> was used as a control and was treated in parallel with other samples. Subcultures were incubated at 37°C with aeration, and aliquots for hydrogen peroxide detection were collected every hour. Bacteria were collected by centrifugation at maximum speed (Eppendorf 5415D) for 3 min. Cleared supernatants were used for hydrogen peroxide detection using an Amplex red hydrogen peroxide/peroxidase kit according to the manufacturer's protocol (Invitrogen). Hydrogen peroxide concentrations correlated with production of resorufin, and fluorescence was measured at 530/585 nm. Results were expressed as percent hydrogen peroxide degradation calculated as [fluorescence<sub>530/585</sub>(*t<sub>n</sub>*)/fluorescence<sub>530/585</sub>(*t<sub>0</sub>*)] \* 100 over time. Each experiment was performed on at least three separate occasions. Statistical significance was determined using a Student's *t* test.

**Hydrogen peroxide sensitivity of mixed cultures in cross complementation studies.** Overnight cultures of the wild type and the siderophore or siderophore-processing mutant strain or the *ΔmacAB* mutant and siderophore or siderophore-processing mutant strain were combined in a 1:1 ratio. The resulting mixed cultures were diluted 1/100 in LB broth supplemented with 1 mM H<sub>2</sub>O<sub>2</sub> and incubated at 37°C with aeration. Aliquots were collected hourly, serially diluted, and plated on LB agar supplemented with appropriate antibiotics. Results were expressed as percent survival calculated as [CFU(*t<sub>n</sub>*)/CFU(*t<sub>0</sub>*)] \* 100 over time. Each experiment was performed on at least three separate occasions.

**Siderophore extraction from the conditioned medium.** Overnight cultures of the wild type or the *ΔmacAB* mutant were subcultured at 1/100 in 1 liter LB broth containing 0.5 mM H<sub>2</sub>O<sub>2</sub> (VWR). Subcultures were grown at 37°C with aeration for 3 h. Bacteria were pelleted by centrifugation at 4,000 × *g* for 30 min, and conditioned medium was filtered to remove any remaining bacteria by the use of 0.2-μm-pore-size filters (Millipore). The resulting metabolite-containing medium was acidified to pH 2 with 10 N HCl. An additional liter of LB broth was acidified in a similar fashion as a media control for mock extraction.

Siderophores were extracted from the acidified conditioned media or mock LB control using a protocol previously described by Winkelmann et al. (77). Briefly, siderophores were extracted twice with an equal volume of ethyl acetate. The organic phases were combined and dried over anhydrous sodium sulfate. Solvents were evaporated using a Rotavapor (Büchi). Remaining residues were dissolved in 1 ml methanol. The siderophore concentration in these extracts was evaluated in Arnow assay (43).

**Arnow assay.** The presence of siderophore in the medium extracts was measured using Arnow assay (43). Fifty-microliter volumes of extract (from wild-type-conditioned and *ΔmacAB* mutant-conditioned media) were mixed with an equal volume of 0.5 N HCl, followed by addition of nitrite-molybdate reagent (10 g sodium nitrate and 10 g sodium molybdate in 100 ml of H<sub>2</sub>O) and 1 N NaOH. In the presence of catecholate siderophores, this solution appears pink and has an absorption maximum at 510 nm. The concentration of siderophores in the extracts was calculated based on a calibration curve using enterobactin (Sigma) in a range from 0 to 100 μM.

**Cross-feeding assay.** An overnight culture of the indicator strain *ΔentA* (1 ml) was pelleted by centrifugation, washed twice with PBS, and resuspended in an equivalent volume of PBS. Melted LB agar containing 200 μM 2,2'-dipyridyl (DIP) was cooled to 55°C, inoculated with 25 μl of the *ΔentA* mutant, and used to pour plates. After agar plate solidification, agar plugs were removed from the plate using the wide end of a 200-μl pipette tip to create test wells. The resulting plates were used to detect the presence of siderophores in the siderophore extracts from the conditioned medium. Test wells were filled with 50 μl of extracts from the conditioned medium, and plates were incubated at 37°C. Positive (2.5 μM enterobactin [Sigma]) and negative (double-distilled water [ddH<sub>2</sub>O]) controls were included in each experiment. The *ΔentA* indicator strain cannot grow in the presence of the iron chelator DIP unless exogenous siderophores are present in the extracts added to the test well. The presence of a halo of growth around a test well indicated the presence of siderophores in the extract added to the given test well.

**HPLC analysis of siderophore extracts.** Siderophore extracts were separated on a reversed-phase HPLC column (Supelco) (Nucleosil C<sub>18</sub>, 5-μm pore size, 25 cm by 4 mm) using an acetonitrile gradient in water (6% to 40%) with 0.1% trifluoroacetic acid (TFA) with a flow rate of 1 ml/min and detection at 220 nm. Separation was performed on a Waters HPLC system. L-Tryptophan (100 μM) was added to each sample before injection into HPLC and served as a control for column performance.

An HPLC calibration kit containing enterobactin and its degradation products (enterobactin trimer [ENT-TRI], enterobactin-dimer [ENT-DIM], and enterobactin-monomer [ENT-MONO]) (EMC Microcollections, Germany) was used to determine the retention time for each siderophore derivative.

**HPLC fraction collection.** Fifty-microliter volumes of siderophore extracts from wild-type and *ΔmacAB* mutant-conditioned media as well as the mock extract from LB broth were separated on HPLC as described above. Fractions were collected with the following retention times: 3.07 to 6.47 min (fraction 1 [negative control]); 6.48 to 10.07 min (fraction 2 [expected to contain ENT-MONO]); 10.08 to 13.47 min



(fraction 3 [negative control]); 13.48 to 16.47 min (fraction 4 [expected to contain ENT-DIM]); 16.48 to 20.07 min (fraction 5 [expected to contain ENT-TRI]), and 20.08 to 24.07 (fraction 6 [expected to contain enterobactin]). Collected fractions were air-dried and reconstituted in 100  $\mu$ l methanol. The concentration of siderophores was measured by Arnov assay as described above.

**Hydrogen peroxide sensitivity of the  $\Delta$ macAB mutant in the presence of siderophore-containing HPLC fractions.** An overnight culture of the  $\Delta$ macAB mutant grown in LB broth was diluted 1/100 in the fresh media supplemented with 1 mM H<sub>2</sub>O<sub>2</sub>. Fractions 1 to 6, with retention times corresponding to different siderophores or their metabolites as described above, were collected after HPLC and were added at a concentration of 2.5  $\mu$ M (based on Arnov assay) to individual tubes containing the  $\Delta$ macAB mutant and incubated at 37°C with aeration. A separate sample containing the  $\Delta$ macAB mutant-inoculated LB broth with 1 mM H<sub>2</sub>O<sub>2</sub> was incubated under similar conditions and served as a positive control. Aliquots were collected hourly, serially diluted, and plated on LB agar supplemented with chloramphenicol. Results were expressed as percent survival calculated as [CFU( $t_n$ )/CFU( $t_0$ )] \* 100 over time. Each experiment was performed at least on three separate occasions.

**Hydrogen peroxide sensitivity of the  $\Delta$ macAB mutant in the presence of purified enterobactin and its linearized products.** An overnight culture of the  $\Delta$ macAB mutant grown in LB broth was diluted 1/100 in fresh medium supplemented with 1 mM H<sub>2</sub>O<sub>2</sub>. Enterobactin (Sigma), Ent-TRI, Ent-DIM, and Ent-MONO (all from EMC Microcollections, Germany) were added to individual tubes at 2.5  $\mu$ M and incubated at 37°C with aeration. A separate tube containing the  $\Delta$ macAB mutant-inoculated LB broth supplemented with 1 mM H<sub>2</sub>O<sub>2</sub> was incubated under the same conditions and served as a positive control. Aliquots were collected hourly, serially diluted, and plated on LB agar supplemented with chloramphenicol. Results were expressed as percent survival calculated as [CFU( $t_n$ )/CFU( $t_0$ )] \* 100 over time. Each experiment was performed on at least three separate occasions.

## SUPPLEMENTAL MATERIAL

Supplemental material is available online only.

**FIG S1**, TIF file, 2.7 MB.

**FIG S2**, TIF file, 2.7 MB.

**FIG S3**, TIF file, 2.7 MB.

**FIG S4**, TIF file, 2.7 MB.

**FIG S5**, TIF file, 2.7 MB.

**FIG S6**, TIF file, 2.7 MB.

## REFERENCES

- Coburn B, Grassl GA, Finlay BB. 2007. Salmonella, the host and disease: a brief review. *Immunol Cell Biol* 85:112–118. <https://doi.org/10.1038/sj.icb.7100007>.
- Zhang S, Kingsley RA, Santos RL, Andrews-Polymenis H, Raffatellu M, Figueiredo J, Nunes J, Tsois RM, Adams LG, Bäumlér AJ. 2003. Molecular pathogenesis of Salmonella enterica serotype Typhimurium-induced diarrhea. *Infect Immun* 71:1–12. <https://doi.org/10.1128/iai.71.1.1-12.2003>.
- Salcedo SP, Noursadeghi M, Cohen J, Holden DW. 2001. Intracellular replication of Salmonella typhimurium strains in specific subsets of splenic macrophages in vivo. *Cell Microbiol* 3:587–597. <https://doi.org/10.1046/j.1462-5822.2001.00137.x>.
- Santos RL, Bäumlér AJ. 2004. Cell tropism of Salmonella enterica. *Int J Med Microbiol* 294:225–233. <https://doi.org/10.1016/j.ijmm.2004.06.029>.
- Horiyama T, Yamaguchi A, Nishino K. 2010. TolC dependency of multidrug efflux systems in Salmonella enterica serovar Typhimurium. *J Antimicrob Chemother* 65:1372–1376. <https://doi.org/10.1093/jac/dkq160>.
- Santiviago CA, Fuentes JA, Bueno SM, Trombert AN, Hildago AA, Socias LT, Youderian P, Mora GC. 2002. The Salmonella enterica sv. Typhimurium smvA, yddG and ompD (porin) genes are required for the efficient efflux of methyl viologen. *Mol Microbiol* 46:687–698. <https://doi.org/10.1046/j.1365-2958.2002.03204.x>.
- Nesme J, Cecillon S, Delmont TO, Monier JM, Vogel TM, Simonet P. 2014. Large-scale metagenomic-based study of antibiotic resistance in the environment. *Curr Biol* 24:1096–1100. <https://doi.org/10.1016/j.cub.2014.03.036>.
- Noronha MF, Lacerda Junior GV, Gilbert JA, de Oliveira VM. 2017. Taxonomic and functional patterns across soil microbial communities of global biomes. *Sci Total Environ* 609:1064–1074. <https://doi.org/10.1016/j.scitotenv.2017.07.159>.
- Kobayashi N, Nishino K, Yamaguchi A. 2001. Novel macrolide-specific ABC-type efflux transporter in Escherichia coli. *J Bacteriol* 183:5639–5644. <https://doi.org/10.1128/JB.183.19.5639-5644.2001>.
- Greene NP, Kaplan E, Crow A, Koronakis V. 2018. Antibiotic resistance mediated by the MacB ABC transporter family: a structural and functional perspective. *Front Microbiol* 9:950. <https://doi.org/10.3389/fmicb.2018.00950>.
- Fitzpatrick AWP, Llabres S, Neuberger A, Blaza JN, Bai XC, Okada U, Murakami S, van Veen HW, Zachariae U, Scheres SHW, Luisi BF, Du D. 2017. Structure of the MacAB-TolC ABC-type tripartite multidrug efflux pump. *Nat Microbiol* 2:17070. <https://doi.org/10.1038/nmicrobiol.2017.70>.
- Yamanaka H, Kobayashi H, Takahashi E, Okamoto K. 2008. MacAB is involved in the secretion of Escherichia coli heat-stable enterotoxin II. *J Bacteriol* 190:7693–7698. <https://doi.org/10.1128/JB.00853-08>.
- Turlin E, Heuck G, Simoes Brandao MI, Szili N, Mellin JR, Lange N, Wandersman C. 2014. Protoporphyrin (PPIX) efflux by the MacAB-TolC pump in Escherichia coli. *Microbiologyopen* 3:849–859. <https://doi.org/10.1002/mbo3.203>.
- Nishino K, Latifi T, Groisman EA. 2006. Virulence and drug resistance roles of multidrug efflux systems of Salmonella enterica serovar Typhimurium. *Mol Microbiol* 59:126–141. <https://doi.org/10.1111/j.1365-2958.2005.04940.x>.
- Bogomolnaya LM, Andrews KD, Talamantes M, Maple A, Ragoza Y, Vazquez-Torres A, Andrews-Polymenis H. 2013. The ABC-type efflux pump MacAB protects Salmonella enterica serovar Typhimurium from oxidative stress. *mBio* 4:e00630-13. <https://doi.org/10.1128/mBio.00630-13>.
- Winterbourn CC, Kettle AJ, Hampton MB. 2016. Reactive oxygen species and neutrophil function. *Annu Rev Biochem* 85:765–792. <https://doi.org/10.1146/annurev-biochem-060815-014442>.
- Fenlon LA, Slauch JM. 2014. Phagocyte roulette in Salmonella killing. *Cell Host Microbe* 15:7–8. <https://doi.org/10.1016/j.chom.2014.01.001>.
- Imlay JA. 2003. Pathways of oxidative damage. *Annu Rev Microbiol* 57:395–418. <https://doi.org/10.1146/annurev.micro.57.030502.090938>.
- Carlizo A, Touati D. 1986. Isolation of superoxide dismutase mutants in Escherichia coli: is superoxide dismutase necessary for aerobic life? *EMBO J* 5:623–630. <https://doi.org/10.1002/j.1460-2075.1986.tb04256.x>.
- Fang FC, DeGroot MA, Foster JW, Bäumlér AJ, Ochsner U, Testerman T, Bearson S, Giard JC, Xu Y, Campbell G, Laessig T. 1999. Virulent Salmonella typhimurium has two periplasmic Cu, Zn-superoxide dismutases.

- Proc Natl Acad Sci U S A 96:7502–7507. <https://doi.org/10.1073/pnas.96.13.7502>.
21. Robbe-Saule V, Coynault C, Ibanez-Ruiz M, Hermant D, Norel F. 2001. Identification of a non-haem catalase in *Salmonella* and its regulation by RpoS ( $\sigma$ S). *Mol Microbiol* 39:1533–1545. <https://doi.org/10.1046/j.1365-2958.2001.02340.x>.
  22. Hebrard M, Viala JP, Meresse S, Barras F, Aussel L. 2009. Redundant hydrogen peroxide scavengers contribute to *Salmonella* virulence and oxidative stress resistance. *J Bacteriol* 191:4605–4614. <https://doi.org/10.1128/JB.00144-09>.
  23. Achard ME, Chen KW, Sweet MJ, Watts RE, Schroder K, Schembri MA, McEwan AG. 2013. An antioxidant role for catecholate siderophores in *Salmonella*. *Biochem J* 454:543–549. <https://doi.org/10.1042/BJ20121771>.
  24. Peralta DR, Adler C, Corbalan NS, Paz Garcia EC, Pomares MF, Vincent PA. 2016. Enterobactin as part of the oxidative stress response repertoire. *PLoS One* 11:e0157799. <https://doi.org/10.1371/journal.pone.0157799>.
  25. Adler C, Corbalan NS, Peralta DR, Pomares MF, de Cristobal RE, Vincent PA. 2014. The alternative role of enterobactin as an oxidative stress protector allows *Escherichia coli* colony development. *PLoS One* 9:e84734. <https://doi.org/10.1371/journal.pone.0084734>.
  26. Ernst JF, Bennett RL, Rothfield LI. 1978. Constitutive expression of the iron-enterochelin and ferrichrome uptake systems in a mutant strain of *Salmonella typhimurium*. *J Bacteriol* 135:928–934. <https://doi.org/10.1128/JB.135.3.928-934.1978>.
  27. Troxell B, Hassan HM. 2013. Transcriptional regulation by ferric uptake regulator (Fur) in pathogenic bacteria. *Front Cell Infect Microbiol* 3:59. <https://doi.org/10.3389/fcimb.2013.00059>.
  28. Salvail H, Lanthier-Bourbonnais P, Sobota JM, Caza M, Benjamin J-AM, Mendieta MES, Lépine F, Dozois CM, Imlay J, Massé E. 2010. A small RNA promotes siderophore production through transcriptional and metabolic remodeling. *Proc Natl Acad Sci U S A* 107:15223–15228. <https://doi.org/10.1073/pnas.1007805107>.
  29. Barthel M, Hapfelmeier S, Quintanilla-Martínez L, Kremer M, Rohde M, Hogardt M, Pfeffer K, Rüssmann H, Hardt W-D. 2003. Pretreatment of mice with streptomycin provides a *Salmonella enterica* serovar Typhimurium colitis model that allows analysis of both pathogen and host. *Infect Immun* 71:2839–2858. <https://doi.org/10.1128/iai.71.5.2839-2858.2003>.
  30. Santos RL, Zhang S, Tsois RM, Baumler AJ, Adams LG. 2002. Morphologic and molecular characterization of *Salmonella typhimurium* infection in neonatal calves. *Vet Pathol* 39:200–215. <https://doi.org/10.1354/vp.39-2-200>.
  31. Imperi F, Tiburzi F, Visca P. 2009. Molecular basis of pyoverdine siderophore recycling in *Pseudomonas aeruginosa*. *Proc Natl Acad Sci U S A* 106:20440–20445. <https://doi.org/10.1073/pnas.0908760106>.
  32. Crosa JH, Walsh CT. 2002. Genetics and assembly line enzymology of siderophore biosynthesis in bacteria. *Microbiol Mol Biol Rev* 66:223–249. <https://doi.org/10.1128/mmb.66.2.223-249.2002>.
  33. Oves-Costales D, Kadi N, Challis GL. 14 September 2009, posting date. The long-overlooked enzymology of a nonribosomal peptide synthetase-independent pathway for virulence-conferring siderophore biosynthesis. *Chem Commun (Camb)* <https://doi.org/10.1039/b913092f>.
  34. Hantke K, Nicholson G, Rabsch W, Winkelmann G. 2003. Salmochelins, siderophores of *Salmonella enterica* and uropathogenic *Escherichia coli* strains, are recognized by the outer membrane receptor IroN. *Proc Natl Acad Sci U S A* 100:3677–3682. <https://doi.org/10.1073/pnas.0737682100>.
  35. Muller SI, Valdebenito M, Hantke K. 2009. Salmochelin, the long-overlooked catecholate siderophore of *Salmonella*. *Biomaterials* 22: 691–695. <https://doi.org/10.1007/s10534-009-9217-4>.
  36. Crouch ML, Castor M, Karlinsey JE, Kalthorn T, Fang FC. 2008. Biosynthesis and IroC-dependent export of the siderophore salmochelin are essential for virulence of *Salmonella enterica* serovar Typhimurium. *Mol Microbiol* 67:971–983. <https://doi.org/10.1111/j.1365-2958.2007.06089.x>.
  37. Furrer JL, Sanders DN, Hook-Barnard IG, McIntosh MA. 2002. Export of the siderophore enterobactin in *Escherichia coli*: involvement of a 43 kDa membrane exporter. *Mol Microbiol* 44:1225–1234. <https://doi.org/10.1046/j.1365-2958.2002.02885.x>.
  38. Bleuel C, Grosse C, Taudte N, Scherer J, Wesenberg D, Krauss GJ, Nies DH, Grass G. 2005. TolC is involved in enterobactin efflux across the outer membrane of *Escherichia coli*. *J Bacteriol* 187:6701–6707. <https://doi.org/10.1128/JB.187.19.6701-6707.2005>.
  39. Lin H, Fischbach MA, Liu DR, Walsh CT. 2005. In vitro characterization of salmochelin and enterobactin trilactone hydrolases IroD, IroE, and Fes. *J Am Chem Soc* 127:11075–11084. <https://doi.org/10.1021/ja0522027>.
  40. Peng ED, Wyckoff EE, Mey AR, Fisher CR, Payne SM. 2016. Nonredundant roles of iron acquisition systems in *Vibrio cholerae*. *Infect Immun* 84: 511–523. <https://doi.org/10.1128/IAI.01301-15>.
  41. Neumann W, Sassone-Corsi M, Raffatellu M, Nolan EM. 2018. Esterase-catalyzed siderophore hydrolysis activates an enterobactin-ciprofloxacin conjugate and confers targeted antibacterial activity. *J Am Chem Soc* 140:5193–5201. <https://doi.org/10.1021/jacs.8b01042>.
  42. Nassif X, Sansonetti PJ. 1986. Correlation of the virulence of *Klebsiella pneumoniae* K1 and K2 with the presence of a plasmid encoding aerobactin. *Infect Immun* 54:603–608. <https://doi.org/10.1128/IAI.54.3.603-608.1986>.
  43. Arnow LE. 1937. Colorimetric determination of the components of 3,4-dihydroxyphenylalanine-tyrosine mixtures. *J Biol Chem* 118:531–537.
  44. Poole K. 2007. Efflux pumps as antimicrobial resistance mechanisms. *Ann Med* 39:162–176. <https://doi.org/10.1080/07853890701195262>.
  45. Alvarez-Ortega C, Olivares J, Martínez JL. 2013. RND multidrug efflux pumps: what are they good for? *Front Microbiol* 4:7. <https://doi.org/10.3389/fmicb.2013.00007>.
  46. Nguyen GT, Green ER, Mecsas J. 2017. Neutrophils to the ROScUE: mechanisms of NADPH oxidase activation and bacterial resistance. *Front Cell Infect Microbiol* 7:373. <https://doi.org/10.3389/fcimb.2017.00373>.
  47. Lim SP, Roongsawang N, Washio K, Morikawa M. 2009. Flexible exportation mechanisms of arthofactin in *Pseudomonas* sp. MIS38. *J Appl Microbiol* 107:157–166. <https://doi.org/10.1111/j.1365-2672.2009.04189.x>.
  48. Dubern JF, Coppoolse ER, Stiekema WJ, Bloemberg GV. 2008. Genetic and functional characterization of the gene cluster directing the biosynthesis of putisolvin I and II in *Pseudomonas putida* strain PCL1445. *Microbiology* 154:2070–2083. <https://doi.org/10.1099/mic.0.2008/016444-0>.
  49. Li W, Rokni-Zadeh H, De Vleeschouwer M, Ghequire MG, Sinnaeve D, Xie GL, Rozenski J, Madder A, Martins JC, De Mot R. 2013. The antimicrobial compound xantholysin defines a new group of *Pseudomonas* cyclic lipopeptides. *PLoS One* 8:e62946. <https://doi.org/10.1371/journal.pone.0062946>.
  50. Cho H, Kang H. 2012. The PseEF efflux system is a virulence factor of *Pseudomonas syringae* pv. *syringae*. *J Microbiol* 50:79–90. <https://doi.org/10.1007/s12275-012-1353-9>.
  51. Gehring AM, Mori I, Walsh CT. 1998. Reconstitution and characterization of the *Escherichia coli* enterobactin synthetase from EntB, EntE, and EntF. *Biochemistry* 37:2648–2659. <https://doi.org/10.1021/bi9726584>.
  52. Carpenter C, Payne SM. 2014. Regulation of iron transport systems in Enterobacteriaceae in response to oxygen and iron availability. *J Inorg Biochem* 133:110–117. <https://doi.org/10.1016/j.jinorgbio.2014.01.007>.
  53. Fischbach MA, Lin H, Liu DR, Walsh CT. 2006. How pathogenic bacteria evade mammalian sabotage in the battle for iron. *Nat Chem Biol* 2:132–138. <https://doi.org/10.1038/nchembio771>.
  54. Bister B, Bischoff D, Nicholson GJ, Valdebenito M, Schneider K, Winkelmann G, Hantke K, Sussmuth RD. 2004. The structure of salmochelins: C-glycosylated enterobactins of *Salmonella enterica*. *Biomaterials* 17: 471–481. <https://doi.org/10.1023/b:biom.0000029432.69418.6a>.
  55. Ellermann M, Arthur JC. 2017. Siderophore-mediated iron acquisition and modulation of host-bacterial interactions. *Free Radic Biol Med* 105: 68–78. <https://doi.org/10.1016/j.freeradbiomed.2016.10.489>.
  56. Pi H, Jones SA, Mercer LE, Meador JP, Caughron JE, Jordan L, Newton SM, Conway T, Klebba PE. 2012. Role of catecholate siderophores in gram-negative bacterial colonization of the mouse gut. *PLoS One* 7:e50020. <https://doi.org/10.1371/journal.pone.0050020>.
  57. Fischbach MA, Lin H, Zhou L, Yu Y, Abergel RJ, Liu DR, Raymond KN, Wanner BL, Strong RK, Walsh CT, Aderem A, Smith KD. 2006. The pathogen-associated iroA gene cluster mediates bacterial evasion of lipocalin 2. *Proc Natl Acad Sci U S A* 103:16502–16507. <https://doi.org/10.1073/pnas.0604636103>.
  58. Nagy TA, Moreland SM, Andrews-Polymenis H, Detweiler CS. 2013. The ferric enterobactin transporter Fep is required for persistent *Salmonella enterica* serovar Typhimurium infection. *Infect Immun* 81:4063–4070. <https://doi.org/10.1128/IAI.00412-13>.
  59. Rabsch W, Voigt W, Reissbrodt R, Tsois RM, Baumler AJ. 1999. *Salmonella typhimurium* IroN and FepA proteins mediate uptake of enterobactin but differ in their specificity for other siderophores. *J Bacteriol* 181: 3610–3612. <https://doi.org/10.1128/JB.181.11.3610-3612.1999>.
  60. Chaturvedi KS, Hung CS, Giblin DE, Urushidani S, Austin AM, Dinauer MC, Henderson JP. 2014. Cupric yersiniabactin is a virulence-associated superoxide dismutase mimic. *ACS Chem Biol* 9:551–561. <https://doi.org/10.1021/cb400658k>.
  61. Batinic-Haberle I, Rajic Z, Tovmasyan A, Reboucas JS, Ye X, Leong KW,

- Dewhirst MW, Vujaskovic Z, Benov L, Spasojevic I. 2011. Diverse functions of cationic Mn(III) N-substituted pyridylporphyrins, recognized as SOD mimics. *Free Radic Biol Med* 51:1035–1053. <https://doi.org/10.1016/j.freeradbiomed.2011.04.046>.
62. Day BJ. 2009. Catalase and glutathione peroxidase mimics. *Biochem Pharmacol* 77:285–296. <https://doi.org/10.1016/j.bcp.2008.09.029>.
63. Doctrow SR, Huffman K, Marcus CB, Tocco G, Malfroy E, Adinolfi CA, Kruk H, Baker K, Lazarowych N, Mascarenhas J, Malfroy B. 2002. Salen-manganese complexes as catalytic scavengers of hydrogen peroxide and cytoprotective agents: structure-activity relationship studies. *J Med Chem* 45:4549–4558. <https://doi.org/10.1021/jm020207y>.
64. Povie G, Villa G, Ford L, Pozzi D, Schiesser CH, Renaud P. 2010. Role of catechol in the radical reduction of B-alkylcatecholboranes in presence of methanol. *Chem Commun (Camb)* 46:803–805. <https://doi.org/10.1039/b917004a>.
65. Puertas-Bartolomé M, Benito-Garzón L, Fung S, Kohn J, Vázquez-Lasa B, San Román J. 2019. Bioadhesive functional hydrogels: controlled release of catechol species with antioxidant and antiinflammatory behavior. *Mater Sci Eng C Mater Biol Appl* 105:110040. <https://doi.org/10.1016/j.msec.2019.110040>.
66. Puertas-Bartolomé M, Vázquez-Lasa B, San Roman J. 13 July 2018, posting date. Bioactive and bioadhesive catechol conjugated polymers for tissue regeneration. *Polymers (Basel)* <https://doi.org/10.3390/polym10070768>.
67. Barclay LRC, Edwards CE, Vinqvist MR. 1999. Media effects on antioxidant activities of phenols and catechols. *J Am Chem Soc* 121:6226–6231. <https://doi.org/10.1021/ja990878u>.
68. Heijnen CG, Haenen GR, van Acker FA, van der Vijgh WJ, Bast A. 2001. Flavonoids as peroxynitrite scavengers: the role of the hydroxyl groups. *Toxicol In Vitro* 15:3–6. [https://doi.org/10.1016/s0887-2333\(00\)00053-9](https://doi.org/10.1016/s0887-2333(00)00053-9).
69. Datsenko KA, Wanner BL. 2000. One-step inactivation of chromosomal genes in *Escherichia coli* K-12 using PCR products. *Proc Natl Acad Sci U S A* 97:6640–6645. <https://doi.org/10.1073/pnas.120163297>.
70. Santiviago CA, Reynolds MM, Porwollik S, Choi SH, Long F, Andrews-Polymeris HL, McClelland M. 2009. Analysis of pools of targeted *Salmonella* deletion mutants identifies novel genes affecting fitness during competitive infection in mice. *PLoS Pathog* 5:e1000477. <https://doi.org/10.1371/journal.ppat.1000477>.
71. Bogomolnaya LM, Santiviago CA, Yang HJ, Baumler AJ, Andrews-Polymeris HL. 2008. 'Form variation' of the O12 antigen is critical for persistence of *Salmonella* Typhimurium in the murine intestine. *Mol Microbiol* 70:1105–1119. <https://doi.org/10.1111/j.1365-2958.2008.06461.x>.
72. Sternberg NL, Maurer R. 1991. Bacteriophage-mediated generalized transduction in *Escherichia coli* and *Salmonella typhimurium*. *Methods Enzymol* 204:18–43. [https://doi.org/10.1016/0076-6879\(91\)04004-8](https://doi.org/10.1016/0076-6879(91)04004-8).
73. Wang RF, Kushner SR. 1991. Construction of versatile low-copy-number vectors for cloning, sequencing and gene expression in *Escherichia coli*. *Gene* 100:195–199. [https://doi.org/10.1016/0378-1119\(91\)90366-J](https://doi.org/10.1016/0378-1119(91)90366-J).
74. Effenbein JR, Endicott-Yazdani T, Porwollik S, Bogomolnaya LM, Cheng P, Guo J, Zheng Y, Yang HJ, Talamantes M, Shields C, Maple A, Ragoza Y, DeAtley K, Tatsch T, Cui P, Andrews KD, McClelland M, Lawhon SD, Andrews-Polymeris H. 2013. Novel determinants of intestinal colonization of *Salmonella enterica* serotype Typhimurium identified in bovine enteric infection. *Infect Immun* 81:4311–4320. <https://doi.org/10.1128/IAI.00874-13>.
75. Zhang S, Adams LG, Nunes J, Khare S, Tsois RM, Baumler AJ. 2003. Secreted effector proteins of *Salmonella enterica* serotype Typhimurium elicit host-specific chemokine profiles in animal models of typhoid fever and enterocolitis. *Infect Immun* 71:4795–4803. <https://doi.org/10.1128/iai.71.8.4795-4803.2003>.
76. Zhang S, Santos RL, Tsois RM, Stender S, Hardt WD, Baumler AJ, Adams LG. 2002. The *Salmonella enterica* serotype Typhimurium effector proteins SipA, SopA, SopB, SopD, and SopE2 act in concert to induce diarrhea in calves. *Infect Immun* 70:3843–3855. <https://doi.org/10.1128/iai.70.7.3843-3855.2002>.
77. Winkelmann G, Cansier A, Beck W, Jung G. 1994. HPLC separation of enterobactin and linear 2,3-dihydroxybenzoylserine derivatives: a study on mutants of *Escherichia coli* defective in regulation (*fur*), esterase (*fes*) and transport (*fepA*). *Biometals* 7:149–154. <https://doi.org/10.1007/bf00140485>.