



Communication

# A New Record for Microbial Perchlorate Tolerance: Fungal Growth in NaClO<sub>4</sub> Brines and its Implications for Putative Life on Mars

Jacob Heinz <sup>1</sup>, Tim Krahn <sup>1</sup> and Dirk Schulze-Makuch <sup>1,2,3,4,\*</sup>

- Astrobiology Research Group, Center for Astronomy and Astrophysics (ZAA), Technische Universität Berlin, Hardenbergstr. 36, 10623 Berlin, Germany; jacobheinz@web.de (J.H.); krahn.tim@web.de (T.K.)
- Section Geomicrobiology, German Research Centre for Geosciences (GFZ), 14473 Potsdam, Germany
- Department of Experimental Limnology, Leibniz-Institute of Freshwater Ecology and Inland Fisheries (IGB), 12587 Stechlin, Germany
- School of the Environment, Washington State University, Pullman, WA 99163, USA
- Correspondence: schulze-makuch@tu-berlin.de

Received: 14 April 2020; Accepted: 24 April 2020; Published: 28 April 2020



Abstract: The habitability of Mars is strongly dependent on the availability of liquid water, which is essential for life as we know it. One of the few places where liquid water might be found on Mars is in liquid perchlorate brines that could form via deliquescence. As these concentrated perchlorate salt solutions do not occur on Earth as natural environments, it is necessary to investigate in lab experiments the potential of these brines to serve as a microbial habitat. Here, we report on the sodium perchlorate (NaClO<sub>4</sub>) tolerances for the halotolerant yeast Debaryomyces hansenii and the filamentous fungus Purpureocillium lilacinum. Microbial growth was determined visually, microscopically and via counting colony forming units (CFU). With the observed growth of D. hansenii in liquid growth medium containing 2.4 M NaClO<sub>4</sub>, we found by far the highest microbial perchlorate tolerance reported to date, more than twice as high as the record reported prior (for the bacterium *Planococcus* halocryophilus). It is plausible to assume that putative Martian microbes could adapt to even higher perchlorate concentrations due to their long exposure to these environments occurring naturally on Mars, which also increases the likelihood of microbial life thriving in the Martian brines.

Keywords: Mars; habitability; brines; salts; perchlorate; microorganisms; halotolerant; fungi; yeast; microbial growth

## 1. Introduction

Several observations on Mars, such as characteristic surface morphologies like large fluid-eroded channels, dendritic networks, fluvial valleys, and glacial features [1], and the formation of water-depending minerals like hematite [2], indicate that the planet had a warmer (even though mostly freezing [3]), wetter and more habitable climate in its early history [4]. However, the loss of its magnetic field enabled the solar wind to sputter away large parts of the Martian atmosphere which caused a climate change leading to the dry, cold, and hostile planet that we know today [5,6]. Hence, putative Martian microorganisms would have had to adapt to a gradual decrease in the availability of liquid water. In this process of decreasing water activities, one of the last niches for the occurrence of liquid water would be deposits of hygroscopic salts that absorb water from the thin Martian atmosphere [7]. If these salts can absorb enough water, they will dissolve in the absorbed liquid and form a saturated salt solution (called "brine") that can be diluted by further water absorption. This process is called deliquescence. Indeed, there is strong evidence for the occurrence of deliquescence processes and of at least temporarily stable brines on Mars [8–10].

Several very hygroscopic salts have been detected on Mars that could undergo deliquescence processes [11]. Among these, the most hygroscopic class of salts are perchlorates which have been found in the Martian soil at the Phoenix landing site at concentrations of 0.4–0.6 wt.% [12]. Perchlorate brines have very low freezing points, down to -77.5 °C for a calcium perchlorate solution with an eutectic concentration [13], i.e., the salt concentration causing the most intense freezing point depression. This also enables the provision of liquid water at subzero temperatures and, hence, provides a promising cryo-environment that might serve as a habitat for putative Martian microbes.

On Earth, most of the salt-rich habitats are based on sodium chloride (NaCl), e.g., in the Atacama Desert, Chile, where endolithic cyanobacteria thrive in NaCl crusts by gaining water absorbed by the salt [14,15]. Furthermore, it has also been shown very recently that methanogenic archaea can survive saturated NaCl concentrations and use water exclusively provided by the deliquescence of salt [16]. There are several environments on Earth that also provide high concentrations of salts other than NaCl, e.g., the Dead Sea with increased calcium (0.47 M Ca<sup>2+</sup>) and magnesium (1.98 M Mg<sup>2+</sup>) chloride concentrations (additional to saturated NaCl conditions) [17], the Spotted Lake (Canada) containing high sulfate concentrations (>3 M) [18], the Don Juan Pond (Antarctica) containing 3.7 M CaCl<sub>2</sub> [19,20], or the Discovery Basin (Mediterranean Sea) containing 5 M MgCl<sub>2</sub> [21]. While the Dead Sea and the Spotted Lake are inhabited by halophilic microorganisms [18,22], the Don Juan Pond and the Discovery Basin are thought to be sterile environments due to low water activities, high ionic strengths and chaotropic stress [21,23,24].

In contrast to Mars, there are only very few natural environments on Earth where low perchlorate concentrations have been detected, e.g., in the Atacama Desert, Chile [25–27], and in the Dry Valleys, Antarctica [28], and no environment is known that consists of concentrated perchlorate brines. Thus, it is appropriate to study these brines in lab experiments to better understand their potential in serving as a habitat for microbial life on Mars. To date, there is only a relatively small body of literature investigating the habitability of brines with perchlorate concentrations above  $\sim 0.1 \, \mathrm{M}$  [29–38]. More studies exist on perchlorate-reducing bacteria (for reviews see [39,40]) and archaea [41] at low perchlorate concentrations below 0.1 M. Prior to this study, the organism found tolerating the highest perchlorate concentration suitable for microbial growth was the halotolerant bacterial strain *Planococcus halocryophilus*, which tolerates 1.1 M sodium perchlorate (NaClO<sub>4</sub>) in its liquid growth medium at 25 °C [30]. Earlier studies reported lower perchlorate tolerances for halotolerant bacteria and halophilic archaea (see Table 1). However, there have been no investigations on the perchlorate tolerance of eukaryotes prior to this study, even though fungi are known to tolerate high concentrations of various other salts [42,43].

**Table 1.** Sodium perchlorate (NaClO<sub>4</sub>) tolerances (in mol/L, wt.% [w/w], and wt./vol.% [w/v]) for the two organisms of each domain of life tolerating the highest perchlorate concentrations reported to date.

Domain	Organism -	NaClO <sub>4</sub> Tolerance			T.11
		(mol/L)	(wt.%)	(wt./vol.%)	Literature
Archaea	Haloferax mediterranei	0.6	6.8	7.3	[31]
	Halorubrum lacusprofundi	0.8	8.9	9.8	[32]
Bacteria	Halomonas venusta	1.0	10.9	12.2	[33]
	Planococcus halocryophilus	1.1	12.0	13.6	[30]
Eukarya (Fungi)	Purpureocillium lilacinum Debaryomyces hansenii	1.9 2.4	19.0 23.0	23.5 29.9	This study This study

Here, we report on the NaClO<sub>4</sub> tolerance of the halotolerant yeast *Debaryomyces hansenii* and the filamentous fungus *Purpureocillium lilacinum*. *D. hansenii* can be found in hyper-saline environments like the Great Salt Lake of Utah or in salterns on the Atlantic coast of Namibia [44]. The yeast has been shown to grow in media containing up to 4 M NaCl [44]. Its high halotolerance results mainly from the accumulation of the compatible solutes glycerol and arabinitol in the exponential growth phase and

in the stationary phase, respectively [45]. Additional changes in the metabolism of the yeast during growth under saline conditions are reviewed elsewhere [44,46,47]. In contrast, there is no detailed research on the halotolerance of P. lilacinum. Lotlikar and Damare (2018) [48] showed that the fungus can grow in a medium with a salinity of S = 100 (corresponding to a NaCl concentration of 1.9 mol/L), but not at a salinity of S = 250 (5.7 mol/L NaCl). Arpini et al. (2019) [49] found the minimum inhibitory concentration (MIC) for NaCl to be 200 g/L (3.4 mol/L NaCl) for P. lilacinum.

#### 2. Materials and Methods

## 2.1. Organisms and Culture Conditions

The halotolerant yeast *Debaryomyces hansenii* (DSM 3428) was obtained from the DSMZ (Leibniz Institute DSMZ—German Collection of Microorganisms and Cell Cultures). The yeast cells were grown aerobically at 25 °C (optimum growth temperature) in liquid DMSZ growth medium #90 (3% malt extract, 0.3% soya peptone) with various concentrations of NaClO<sub>4</sub>. The media were prepared by mixing the media components, NaClO<sub>4</sub> and water, followed by pH adjustment (pH  $\sim$ 5.6) and sterile filtration.

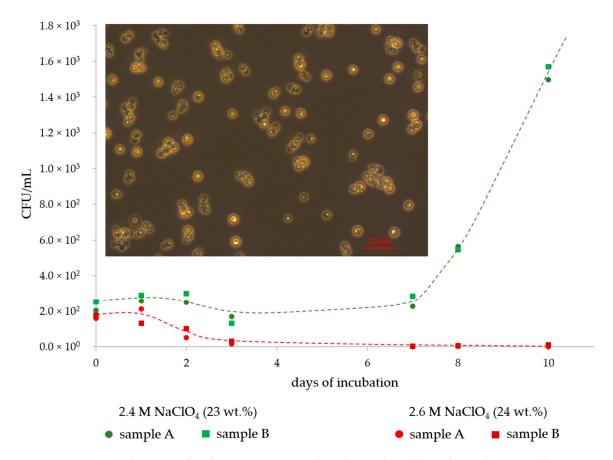
The filamentous fungus *Purpureocillium lilacinum* was found as a contamination at surprisingly high NaClO<sub>4</sub> concentrations (see Results) during growth experiments with *Planococcus halocryophilus* in liquid DMSZ growth medium #92 (3% tryptic soy broth, 0.3% yeast extract, pH 7.2–7.4, 25 °C) which have been described elsewhere [30]. *P. lilacinum* was isolated and characterized by 18S rDNA sequencing (data not shown). Due to safety restrictions (*P. lilacinum* is categorized as biosafety level S2) no further experiments were conducted with this fungus.

### 2.2. Determination of Perchlorate Tolerances

The survival and growth of the fungi were determined visibly in 15 mL centrifuge tubes containing the inoculated liquid growth medium, as well as by using light microscopy (Primo Star, Zeiss, equipped with Axio Cam 105 color) and counting colony forming units (CFU) on agar plates (1.5% agar) containing the respective growth medium. The maximum NaClO4 concentration suitable for growth was determined through progressive culture adaptation to higher perchlorate concentrations as described previously [30]. In short, 10  $\mu$ l of a stock culture was used to inoculate 5 ml of liquid growth medium containing 10 wt.% NaClO4 (corresponding to 0.9 mol/L). The culture growing from this medium was used to inoculate growth medium containing 15 wt.% NaClO4 (1.4 mol/L). This procedure was repeated with increasing NaClO4 concentrations in 5 wt.% steps. When no growth could be detected the increments of the NaClO4 concentration increase were lowered to 1 wt.%. The highest NaClO4 concentration that enabled growth was defined as the perchlorate tolerance of the organism with a technical error of  $\pm$  1 wt.% NaClO4. All growth experiments were conducted as biological duplicates, i.e., for each NaClO4 concentration, two separate samples were inoculated.

## 3. Results

The perchlorate tolerance of *D. hansenii* was found to be 2.4 M NaClO<sub>4</sub>. Growth curves (until the exponential growth phase) for the samples with this concentration are shown in Figure 1 together with the curves for samples with a concentration of 2.6 M NaClO<sub>4</sub>, in which the cells were dying within 10 days. Additionally, inserted into Figure 1 is a light microscopy image of *D. hansenii* cells after growth in DSMZ medium #90 showing single cells and some small and loosely-bound cell aggregates. Besides a decrease in cell density, no phenotypical changes in the yeast cell morphologies were found when the cells were grown in perchlorate-rich media. This contrasts with earlier studies on the bacterial strain *P. halocryophilus* where cells grown in perchlorate-rich media formed large cell clusters [30].



**Figure 1.** Growth curves of *D. hansenii* at 25 °C in liquid growth media with a sodium perchlorate (NaClO<sub>4</sub>) concentration of 2.4 M (green line and symbols) and 2.6 M (red line and symbols). Experiments were run in biological duplicates (samples A and B). Microbial growth occurred only in the 2.4 M NaClO<sub>4</sub> samples while the cells in the 2.6 M NaClO<sub>4</sub> samples died within 10 days. Cells of *D. hansenii* after growth in liquid growth medium are shown in the implemented image.

Purpureocillium lilacinum was not investigated in detail due to safety restrictions (see Section 2.1). However, it was found as a contaminant growing in liquid growth medium having NaClO<sub>4</sub> concentrations of up to 1.9 M, which we interpret to be its uppermost tolerance to NaClO<sub>4</sub>.

#### 4. Discussion

Before the detection of significant amounts of perchlorate on Mars in 2008 [12], the scientific interest in determining the habitability of concentrated perchlorate brines was low due to their practical non-existence in natural habitats on Earth. Since 2008, the number of studies investigating this question has increased but is still insufficient for understanding the potential for life in Martian perchlorate-containing brines. Table 1 (see Introduction) lists the two record holders for each domain of life regarding their  $NaClO_4$  tolerance as described in the literature thus far.

The study presented here is the first one describing significant perchlorate tolerances for eukaryotes. Several other studies described growth of non-fungal organisms in perchlorate solutions with concentrations lower than the ones listed in Table 1 [31,33–38]. For example, Oren et al. (2014) [31] found that several halophilic archaea of the family Halobacteriaceae (*Halobacterium* strain NRC-1, *Hbt. salinarum* R1, *Haloferax volcanii*, *Hfx. mediterranei*, *Hfx. denitrificans*, *Hfx. gibbonsii*, *Haloarcula marismortui*, and *Har. vallismortis*) and the bacterium *Halomonas elongata* grew well in NaCl-based media supplemented with a perchlorate concentration of up to 0.4 M. However, among these, only *Hfx. mediterranei* was also able to grow in 0.6 M NaClO<sub>4</sub>. Al Soudi et al. (2017) [33] found that the halotolerant bacterial strains *Marinococcus halophilus*, *Halomonas venusta*, and *Bacillus licheniformis* grew

Life 2020, 10, 53 5 of 8

robustly at 0.5 M NaClO<sub>4</sub>, while only *H. venusta* also showed substantial growth at 1.0 M NaClO<sub>4</sub>. Furthermore, it has been shown that methanogenic archaea can metabolize and produce methane at perchlorate concentrations of up to 0.4 mol/L [38].

Therefore, the two fungi investigated in this study, *D. hansenii* and *P. lilacinum*, have by far the highest tolerances to NaClO<sub>4</sub> among all microorganisms investigated to date. The tolerance for *D. hansenii* (2.4 M NaClO<sub>4</sub>) is more than twice as high as for the bacterial strain *P. halocryophilus* that has been holding the NaClO<sub>4</sub> tolerance record (1.1 M NaClO<sub>4</sub>) prior to this study [30]. As the two fungi described in this study are the first ones ever investigated regarding their perchlorate tolerance, it is plausible to assume that other fungi (e.g., the extremely halotolerant black yeast *Hortaea werneckii*, or the obligately halophilic *Wallemia ichthyophaga* [43]) might tolerate even higher perchlorate concentrations, which we plan to investigate in upcoming experiments.

The perchlorate tolerance data available in the literature to date (Table 1) convey the impression that fungal species have a more efficient perchlorate defense machinery than bacteria and archaea. However, more research on microbial and fungal perchlorate tolerances and the adaptation mechanisms applied by the fungi grown in perchlorate-rich growth media is needed to confirm or discard this hypothesis. Furthermore, the effect of additional stress factors typical for Mars on the fungal survival and growth in perchlorate brines should prompt further investigations. The most relevant stressors in this context are low temperatures and pressures, high radiation levels, and stress induced by the higher chaotropicity and ionic strengths of ions from other perchlorate salts like magnesium or calcium perchlorate, which probably represent the majority of perchlorate salts on Mars [50,51].

Since there are no natural perchlorate-rich environments existing on Earth and thought not to have existed in the past, there is no obvious adaptation mechanism for the two fungal species investigated in this study to have adapted to these high NaClO<sub>4</sub> concentrations. If life on Mars exists in perchlorate brines, we may speculate that these microorganisms might have evolved tolerance to much higher perchlorate concentrations, as they—in contrast to Earth—would have been under natural selection pressures on Mars to achieve higher perchlorate tolerances. We recommend that this hypothesis should be tested in future Mars missions via life detection experiments in saline and perchlorate-rich environments, such as locations where the presumably deliquescence-driven "recurring slope lineae" (RSL) [10] occur.

**Author Contributions:** Conceptualization, J.H. and D.S.-M.; methodology, J.H.; validation, J.H., T.K. and D.S.-M.; formal analysis, J.H. and T.K.; investigation, J.H. and T.K.; resources, D.S.-M.; writing—original draft preparation, J.H.; writing—review and editing, J.H., T.K. and D.S.-M.; supervision, D.S.-M.; project administration, D.S.-M.; funding acquisition, D.S.-M. All authors have read and agreed to the published version of the manuscript.

**Funding:** Research was funded by the European Research Council Advanced Grant "Habitability of Martian Environments" (HOME, no. 339231).

**Acknowledgments:** We thank Hans-Peter Grossart and Solvig Pinnow for conducting the 18S rDNA sequencing of *Purpureocillium lilacinum* and two anonyms reviewers for helping us to improve this article.

**Conflicts of Interest:** The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to publish the results.

## References

- 1. Masson, P.; Carr, M.H.; Costard, F.; Greeley, R.; Hauber, E.; Jaumann, R. Geomorphologic Evidence for Liquid Water. *Space Sci. Rev.* **2001**, *96*, 333–364. [CrossRef]
- 2. Christensen, P.R.; Bandfield, J.L.; Clark, R.N.; Edgett, K.S.; Hamilton, V.E.; Hoefen, T.; Kieffer, H.H.; Kuzmin, R.O.; Lane, M.D.; Malin, M.C.; et al. Detection of crystalline hematite mineralization on Mars by the Thermal Emission Spectrometer: Evidence for near-surface water. *J. Geophys. Res.* **2000**, *105*, 9623–9642. [CrossRef]
- 3. Fairén, A.G. A cold and wet Mars. *Icarus* 2010, 208, 165–175. [CrossRef]
- 4. Wordsworth, R.D. The Climate of Early Mars. Annu. Rev. Earth Planet. Sci. 2016, 44, 381–408. [CrossRef]

5. Vaisberg, O. Mars atmospheric losses induced by the solar wind: Comparison of observations with models. *Planet. Space Sci.* **2015**, *119*, 69–91. [CrossRef]

- 6. Jakosky, B.M.; Slipski, M.; Benna, M.; Mahaffy, P.; Elrod, M.; Yelle, R.; Stone, S.; Alsaeed, N. Mars' atmospheric history derived from upper-atmosphere measurements of 38Ar/36Ar. *Science* 2017, 355, 1408–1410. [CrossRef]
- 7. Davila, A.F.; Schulze-Makuch, D. The Last Possible Outposts for Life on Mars. *Astrobiology* **2016**, *16*, 159–168. [CrossRef]
- 8. Martínez, G.M.; Renno, N.O. Water and Brines on Mars: Current Evidence and Implications for MSL. *Space Sci. Rev.* **2013**, 175, 29–51. [CrossRef]
- 9. Martín-Torres, F.J.; Zorzano, M.-P.; Valentín-Serrano, P.; Harri, A.-M.; Genzer, M.; Kemppinen, O.; Rivera-Valentin, E.G.; Jun, I.; Wray, J.; Bo Madsen, M.; et al. Transient liquid water and water activity at Gale crater on Mars. *Nat. Geosci.* **2015**, *8*, 357–361. [CrossRef]
- 10. Ojha, L.; Wilhelm, M.B.; Murchie, S.L.; McEwen, A.S.; Wray, J.J.; Hanley, J.; Massé, M.; Chojnacki, M. Spectral evidence for hydrated salts in recurring slope lineae on Mars. *Nat. Geosci.* **2015**, *8*, 829–832. [CrossRef]
- 11. Kounaves, S.P.; Hecht, M.H.; Kapit, J.; Gospodinova, K.; DeFlores, L.; Quinn, R.C.; Boynton, W.V.; Clark, B.C.; Catling, D.C.; Hredzak, P.; et al. Wet Chemistry experiments on the 2007 Phoenix Mars Scout Lander mission: Data analysis and results. *J. Geophys. Res.* **2010**, *115*, E00E10. [CrossRef]
- 12. Hecht, M.H.; Kounaves, S.P.; Quinn, R.C.; West, S.J.; Young, S.M.M.; Ming, D.W.; Catling, D.C.; Clark, B.C.; Boynton, W.V.; Hoffman, J.; et al. Detection of perchlorate and the soluble chemistry of martian soil at the Phoenix lander site. *Science* **2009**, *325*, 64–67. [CrossRef] [PubMed]
- 13. Pestova, O.N.; Myund, L.A.; Khripun, M.K.; Prigaro, A.V. Polythermal Study of the Systems M(ClO4)2-H2O (M2+ = Mg2+, Ca2+, Sr2+, Ba2+). *Russ. J. Appl. Chem.* **2005**, *78*, 409–413. [CrossRef]
- 14. Davila, A.F.; Gómez-Silva, B.; de los Rios, A.; Ascaso, C.; Olivares, H.; McKay, C.P.; Wierzchos, J. Facilitation of endolithic microbial survival in the hyperarid core of the Atacama Desert by mineral deliquescence. *J. Geophys. Res.* **2008**, *113*. [CrossRef]
- 15. Wierzchos, J.; Ascaso, C.; McKay, C.P. Endolithic cyanobacteria in halite rocks from the hyperarid core of the Atacama Desert. *Astrobiology* **2006**, *6*, 415–422. [CrossRef] [PubMed]
- 16. Maus, D.; Heinz, J.; Schirmack, J.; Airo, A.; Kounaves, S.P.; Wagner, D.; Schulze-Makuch, D. Methanogenic Archaea Can Produce Methane in Deliquescence-Driven Mars Analog Environments. *Sci. Rep.* **2020**, *10*. [CrossRef] [PubMed]
- 17. Oren, A. The dying Dead Sea: The microbiology of an increasingly extreme environment. *Lakes Reserv. Res. Manag.* **2010**, *15*, 215–222. [CrossRef]
- 18. Pontefract, A.; Zhu, T.F.; Walker, V.K.; Hepburn, H.; Lui, C.; Zuber, M.T.; Ruvkun, G.; Carr, C.E. Microbial Diversity in a Hypersaline Sulfate Lake: A Terrestrial Analog of Ancient Mars. *Front. Microbiol.* **2017**, *8*, 1819. [CrossRef]
- 19. Marion, G.M. A theoretical evaluation of mineral stability in Don Juan Pond, Wright Valley, Victoria Land. *Antarct. Sci.* **1997**, *9*, 92–99. [CrossRef]
- 20. Dickson, J.L.; Head, J.W.; Levy, J.S.; Marchant, D.R. Don Juan Pond, Antarctica: Near-surface CaCl(2)-brine feeding Earth's most saline lake and implications for Mars. *Sci. Rep.* **2013**, *3*, 1166. [CrossRef]
- 21. Hallsworth, J.E.; Yakimov, M.M.; Golyshin, P.N.; Gillion, J.L.M.; D'Auria, G.; de Lima Alves, F.; La Cono, V.; Genovese, M.; McKew, B.A.; Hayes, S.L.; et al. Limits of life in MgCl2-containing environments: Chaotropicity defines the window. *Environ. Microbiol.* **2007**, *9*, 801–813. [CrossRef] [PubMed]
- 22. Oren, A. Life in Magnesium- and Calcium-Rich Hypersaline Environments: Salt Stress by Chaotropic Ions. In *Polyextremophiles: Life under Multiple Forms of Stress;* Seckbach, J., Oren, A., Stan-Lotter, H., Eds.; Springer: Dordrecht, The Netherlands, 2013; pp. 215–232. ISBN 978-94-007-6487-3.
- 23. Samarkin, V.A.; Madigan, M.T.; Bowles, M.W.; Casciotti, K.L.; Priscu, J.C.; McKay, C.P.; Joye, S.B. Abiotic nitrous oxide emission from the hypersaline Don Juan Pond in Antarctica. *Nat. Geosci.* **2010**, *3*, 341–344. [CrossRef]
- 24. Fox-Powell, M.G.; Hallsworth, J.E.; Cousins, C.R.; Cockell, C.S. Ionic Strength Is a Barrier to the Habitability of Mars. *Astrobiology* **2016**, *16*, 427–442. [CrossRef] [PubMed]
- 25. Ericksen, G.E. *Geology and Origin of the Chilean Nitrate Deposits*; U.S. Department of the Interior, U.S. Government Publishing Office: Washington, DC, USA, 1981.
- 26. Catling, D.C.; Claire, M.W.; Zahnle, K.J.; Quinn, R.C.; Clark, B.C.; Hecht, M.H.; Kounaves, S. Atmospheric origins of perchlorate on Mars and in the Atacama. *J. Geophys. Res.* **2010**, *115*, E00E11. [CrossRef]

27. Schulze-Makuch, D.; Wagner, D.; Kounaves, S.P.; Mangelsdorf, K.; Devine, K.G.; de Vera, J.-P.; Schmitt-Kopplin, P.; Grossart, H.-P.; Parro, V.; Kaupenjohann, M.; et al. Transitory microbial habitat in the hyperarid Atacama Desert. *Proc. Natl. Acad. Sci. USA* **2018**, *115*, 2670–2675. [CrossRef]

- 28. Kounaves, S.P.; Stroble, S.T.; Anderson, R.M.; Moore, Q.; Catling, D.C.; Douglas, S.; McKay, C.P.; Ming, D.W.; Smith, P.H.; Tamppari, L.K.; et al. Discovery of natural perchlorate in the Antarctic Dry Valleys and its global implications. *Environ. Sci. Technol.* **2010**, *44*, 2360–2364. [CrossRef]
- 29. Heinz, J.; Schirmack, J.; Airo, A.; Kounaves, S.P.; Schulze-Makuch, D. Enhanced Microbial Survivability in Subzero Brines. *Astrobiology* **2018**, *18*, 1171–1180. [CrossRef]
- 30. Heinz, J.; Waajen, A.C.; Airo, A.; Alibrandi, A.; Schirmack, J.; Schulze-Makuch, D. Bacterial Growth in Chloride and Perchlorate Brines: Halotolerances and Salt Stress Responses of Planococcus halocryophilus. *Astrobiology* **2019**, *19*, 1377–1387. [CrossRef]
- 31. Oren, A.; Elevi Bardavid, R.; Mana, L. Perchlorate and halophilic prokaryotes: Implications for possible halophilic life on Mars. *Extremophiles* **2014**, *18*, 75–80. [CrossRef]
- 32. Laye, V.J.; DasSarma, S. An Antarctic Extreme Halophile and Its Polyextremophilic Enzyme: Effects of Perchlorate Salts. *Astrobiology* **2018**, *18*, 412–418. [CrossRef]
- 33. Al Soudi, A.F.; Farhat, O.; Chen, F.; Clark, B.C.; Schneegurt, M.A. Bacterial growth tolerance to concentrations of chlorate and perchlorate salts relevant to Mars. *Int. J. Astrobiol.* **2017**, *16*, 229–235. [CrossRef]
- 34. Matsubara, T.; Fujishima, K.; Saltikov, C.W.; Nakamura, S.; Rothschild, L.J. Earth analogues for past and future life on Mars: Isolation of perchlorate resistant halophiles from Big Soda Lake. *Int. J. Astrobiol.* **2017**, 16, 218–228. [CrossRef]
- 35. Beblo-Vranesevic, K.; Huber, H.; Rettberg, P. High Tolerance of Hydrogenothermus marinus to Sodium Perchlorate. *Front. Microbiol.* **2017**, *8*, 1369. [CrossRef] [PubMed]
- 36. Shcherbakova, V.; Oshurkova, V.; Yoshimura, Y. The Effects of Perchlorates on the Permafrost Methanogens: Implication for Autotrophic Life on Mars. *Microorganisms* **2015**, *3*, 518–534. [CrossRef] [PubMed]
- 37. Flores, N.; Hoyos, S.; Venegas, M.; Galetović, A.; Zúñiga, L.M.; Fábrega, F.; Paredes, B.; Salazar-Ardiles, C.; Vilo, C.; Ascaso, C.; et al. Haloterrigena sp. Strain SGH1, a Bacterioruberin-Rich, Perchlorate-Tolerant Halophilic Archaeon Isolated From Halite Microbial Communities, Atacama Desert, Chile. *Front. Microbiol.* 2020, 11, 324. [CrossRef]
- 38. Kral, T.A.; Goodhart, T.H.; Harpool, J.D.; Hearnsberger, C.E.; McCracken, G.L.; McSpadden, S.W. Sensitivity and adaptability of methanogens to perchlorates: Implications for life on Mars. *Planet. Space Sci.* **2016**, 120, 87–95. [CrossRef]
- 39. Coates, J.D.; Achenbach, L.A. Microbial perchlorate reduction: Rocket-fueled metabolism. *Nat. Rev. Microbiol.* **2004**, *2*, 569–580. [CrossRef]
- 40. Bardiya, N.; Bae, J.-H. Dissimilatory perchlorate reduction: A review. *Microbiol. Res.* **2011**, *166*, 237–254. [CrossRef]
- 41. Liebensteiner, M.G.; Pinkse, M.W.H.; Schaap, P.J.; Stams, A.J.M.; Lomans, B.P. Archaeal (per)chlorate reduction at high temperature: An interplay of biotic and abiotic reactions. *Science* **2013**, *340*, 85–87. [CrossRef]
- 42. Zajc, J.; Džeroski, S.; Kocev, D.; Oren, A.; Sonjak, S.; Tkavc, R.; Gunde-Cimerman, N. Chaophilic or chaotolerant fungi: A new category of extremophiles? *Front. Microbiol.* **2014**, *5*, 708. [CrossRef]
- 43. Gunde-Cimerman, N.; Zalar, P. Extremely halotolerant and halophilic fungi inhabit brine in solar salterns around the globe. *Food Technol. Biotech.* **2014**, *52*, 170–179.
- 44. Breuer, U.; Harms, H. Debaryomyces hansenii—an extremophilic yeast with biotechnological potential. *Yeast* **2006**, 23, 415–437. [CrossRef] [PubMed]
- 45. Nobre, M.F.; Costa, M.S.d. The accumulation of polyols by the yeast Debaryomyces hansenii in response to water stress. *Can. J. Microbiol.* **1985**, *31*, 1061–1064. [CrossRef]
- 46. Ramos, J. Introducing Debaryomyces Hansenii, a Salt Loving Yeast. In *Adaptation to Life at High Salt Concentrations in Archaea, Bacteria, and Eukarya*; Gunde-Cimerman, N., Oren, A., Plemenitaš, A., Eds.; Springer: Berlin/Heidelberg, Germany, 2005; pp. 441–451. ISBN 1-4020-3632-9.
- 47. Prista, C.; Michán, C.; Miranda, I.M.; Ramos, J. The halotolerant Debaryomyces hansenii, the Cinderella of non-conventional yeasts. *Yeast* **2016**, *33*, 523–533. [CrossRef] [PubMed]
- 48. Lotlikar, N.P.; Damare, S.R. Variability in Protein Expression in Marine-Derived Purpureocillium lilacinum Subjected to Salt and Chromium Stresses. *Indian J. Microbiol.* **2018**, *58*, 360–371. [CrossRef] [PubMed]

49. Arpini, C.M.; Nóbrega, Y.C.; Castheloge, V.D.; Neves, D.S.; Tadokoro, C.E.; Costa, G.L.d.; Oliveira, M.M.E.; Santos, M.R.d.D. Purpuriocillium lilacinum infection in captive loggerhead sea turtle hatchlings. *Med. Mycol. Case Rep.* **2019**, *23*, 8–11. [CrossRef] [PubMed]

- 50. Kounaves, S.P.; Chaniotakis, N.A.; Chevrier, V.F.; Carrier, B.L.; Folds, K.E.; Hansen, V.M.; McElhoney, K.M.; O'Neil, G.D.; Weber, A.W. Identification of the perchlorate parent salts at the Phoenix Mars landing site and possible implications. *Icarus* 2014, 232, 226–231. [CrossRef]
- 51. Toner, J.D.; Catling, D.C.; Light, B. Modeling salt precipitation from brines on Mars: Evaporation versus freezing origin for soil salts. *Icarus* **2015**, 250, 451–461. [CrossRef]



© 2020 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).