



This is a repository copy of *Alemtuzumab-induced halo nevus-like hypopigmentation – New insights into secondary skin autoimmunity in response to an immune cell-depleting antibody.*

White Rose Research Online URL for this paper:
<https://eprints.whiterose.ac.uk/161994/>

Version: Accepted Version

Article:

Bohm, M., Kemp, E. orcid.org/0000-0002-0313-8916, Metze, D. et al. (4 more authors) (2020) Alemtuzumab-induced halo nevus-like hypopigmentation – New insights into secondary skin autoimmunity in response to an immune cell-depleting antibody. *Journal of the European Academy of Dermatology and Venereology*. ISSN 0926-9959

<https://doi.org/10.1111/jdv.16781>

This is the peer reviewed version of the following article: Böhm, M., Kemp, E.H., Metze, D., Muresan, A.M., Neufeld, M., Luiten, R.M. and Ruck, T. (2020), Alemtuzumab-induced halo nevus-like hypopigmentation –. *J Eur Acad Dermatol Venereol*, which has been published in final form at <https://doi.org/10.1111/jdv.16781>. This article may be used for non-commercial purposes in accordance with Wiley Terms and Conditions for Use of Self-Archived Versions.

Reuse

Items deposited in White Rose Research Online are protected by copyright, with all rights reserved unless indicated otherwise. They may be downloaded and/or printed for private study, or other acts as permitted by national copyright laws. The publisher or other rights holders may allow further reproduction and re-use of the full text version. This is indicated by the licence information on the White Rose Research Online record for the item.

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.



eprints@whiterose.ac.uk
<https://eprints.whiterose.ac.uk/>

1
2
3 **Alemtuzumab-induced halo nevus-like hypopigmentation –**
4 **New insights into secondary skin autoimmunity in response**
5 **to an immune cell-depleting antibody**
6
7
8
9

10
11
12
13
14 **Markus Böhm^{1*}, E. Helen Kemp², Dieter Metze¹, Anna Maria Muresan¹,**
15 **Matthias Neufeld¹, Rosalie M. Luiten³, Tobias Ruck⁴**
16
17
18
19

20 ¹Department of Dermatology, University of Münster, Münster, Germany

21
22 ²Department of Oncology and Metabolism, University of Sheffield, Sheffield, UK

23
24 ³Department of Dermatology and Netherlands Institute for Pigment Disorders,
25 Amsterdam UMC, Amsterdam Institute for Infection and Immunity.
26
27

28 University of Amsterdam, Amsterdam, The Netherlands

29
30 ⁴Clinic of Neurology with Institute for Translational Neurology, University of
31 Münster, Münster, Germany
32
33

34
35
36 **Corresponding author:* Markus Böhm, MD
37 Associate Professor
38 Dept. of Dermatology
39 University of Münster
40 Von-Esmarch-Str. 58
41 48149 Münster
42 Germany
43 Tel.: 49-251-8356565
44 Fax: 49-251-8356522
45 email: bohmm@uni-muenster.de
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

The authors of this manuscript declare no conflicts of interest

1
2
3 Alemtuzumab is an antibody directed against the Cluster of Differentiation (CD)52. It
4 depletes B and T lymphocytes and is approved for treatment of active relapsing-
5 remitting multiple sclerosis (RRMS). Secondary autoimmunity is an important risk of
6 alemtuzumab-treated patients and can affect different organs. 41% of patients develop
7 autoimmune thyroid disease². Skin autoimmunity under alemtuzumab has but
8 reported only in the neurological literature^{3,4}.
9

10
11 We describe a 33-year-old male patient who developed hypopigmentation around his
12 melanocytic nevi with disappearance of the nevi in August 2018. In June 2016 he had
13 been diagnosed with highly active RRMS and treated with alemtuzumab in September
14 2016 for the first time. In September 2017 another cycle of alemtuzumab was
15 administered. The patient's family history for vitiligo was unremarkable and he had
16 no autoimmune thyroid disease. At time of the occurrence of the hypopigmented spots
17 the number of his circulating leukocytes in the blood was normal. Upon examination
18 he showed sharply demarcated hypopigmented spots around his melanocytic nevi
19 (Fig. 1A). Some of the nevi within the white spots had already disappeared. There
20 were no other signs of skin hypopigmentation. A biopsy specimen from one of the
21 halo-like nevi (insert of Fig. 1A) revealed absence of epidermal melanocytes (Fig. 1B)
22 which was confirmed by immunohistochemistry with an anti-Pan Mel antibody (Fig.
23 1C). In contrast, melanocytes were detected within the nevus (Fig. 1C).
24 Immunostaining with an anti-CD3 antibody disclosed a sparsely scattered infiltrate of
25 T lymphocytes within the upper dermis (Fig. 1D, E). To shed light into the
26 pathogenesis of the halo nevus-like lesions we wondered whether alemtuzumab might
27 directly attack epidermal melanocytes. CD52 immunohistochemistry of normal skin
28 and a melanocytic nevus from a healthy person did not show immunoreactivity in
29 melanocytes in contrast to lymphoid tissues used as positive control (data not shown).
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3 Since autoimmune thyroid disease is characterized by circulating anti-thyroid
4 antibodies, we hypothesized whether anti-melanocyte antibodies may be generated
5 following alemtuzumab treatment. In patients with non-segmental vitiligo (NSV)
6 antibodies against tyrosinase (TYR), the key enzyme of melanin synthesis, were also
7 detected⁵. Therefore, a TnT® T7-Coupled Reticulocyte Lysate System (Promega,
8 Southampton, UK) was used to produce various [³⁵S]-labelled melanocyte antigens *in*
9 *vitro* from the translation of cDNA in the appropriate plasmid. Serum samples of the
10 patient before and after alemtuzumab therapy were then analysed for circulating
11 antibodies against these antigens by radioligand binding assays⁵. An almost 6-fold
12 increase in anti-TYR antibodies and a ~3-fold increase in antibodies against TYR-
13 related protein 1 were detected following alemtuzumab (Table 1). In contrast, anti-
14 thyroid antibodies were not detected throughout the patient's history. In accordance
15 with this TSH levels were always normal.

16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34 Our findings are puzzling as alemtuzumab depletes both B and T cells but still evokes
35 an autoimmune attack against melanocytes. Recently, three patients with RRMS have
36 been described who developed NSV 14, 18 and 52 months after initiation of
37 alemtuzumab⁶. The immune-mediated destruction pathways of melanocytes in classic
38 halo nevus and NSV may be different⁷. It has been suggested that secondary
39 autoimmunity induced by alemtuzumab is related to proliferation of chronically
40 activated, oligoclonal, effector memory CD8⁺ T cells⁸. Interestingly, resident and
41 effector memory T cells have been identified in non-active skin of patients with
42 NSV⁹. Previous studies showed that alemtuzumab depletes all T cells from the blood
43 but does not deplete skin-resident memory T cells¹⁰. Thus, production of anti-
44 melanocyte-specific antibodies may be secondary to initial destruction of melanocytes
45 by melanocyte-specific CD8⁺ T cells.

1
2
3 In summary, alemtuzumab-induced skin autoimmunity is a condition a dermatologist
4 should be aware of. Our report underscores the complex pathogenesis of immune-
5 mediated destruction of epidermal melanocytes that may follow even upon depletion
6 of both T and B cells.
7
8
9
10
11
12
13
14
15

16 **Acknowledgement**

17
18
19 The patient in this manuscript has given written informed consent to the publication of
20 his case details.
21
22
23
24
25
26

27 **References**

- 28
29
30 1. Tuohy O, Costelloe L, Hill-Cawthorne G, Bjornson I, Harding K, Robertson N,
31 May K, Button T, Azzopardi L, Kousin-Ezewu O, Fahey MT, Jones J, Compston
32 DA, Coles. Alemtuzumab treatment of multiple sclerosis: long-term safety and
33 efficacy. *J Neurol Neurosurg Psychiatry* 2015; **86**: 208–215.
34
35
36
37
38
39 2. Ruck T, Bittner S, Wiendl H, Meuth SG. Alemtuzumab in multiple sclerosis:
40 mechanism of action and beyond. *Int J Mol Sci* 2015; **16**: 16414–1639.
41
42
43
44
45 3. Chan JK, Traboulsee AL, Sayao AL. Case of alemtuzumab-related alopecia areata
46 management in MS. *Neurol Neuroimmunol Neuroinflamm* 2018; **6**: e516.
47
48
49
50 4. Alcalá C, Pzère-Miralles F, Gascón F, Evole M, Estutia M, Gil-Perotín S, Casanova
51 B. Recurrent and universal alopecia areata following alemtuzumab treatment in
52 multiple sclerosis: A secondary autoimmune disease. *Mult Scler Relat Disord*
53 2019; **27**: 406–408.
54
55
56
57
58
59
60

- 1
2
3 5. Kemp EH, Gawkrödger DJ, MacNeil S, Watson PF, Weetman AP. Detection of
4
5 tyrosinase autoantibodies in the sera of vitiligo patients using 35S-labelled
6
7 recombinant human tyrosinase in a radioimmunoassay. *J Invest Dermatol* 1997;
8
9 **109**: 69–73.
10
11
- 12
13 6. Ruck T, Pfeuffer S, Schulte-Mecklenbeck A, Gross CC, Lindner M, Metze D,
14
15 Ehrchen J, Sondermann W, Pul R, Kleinschnitz C, Wiendl H, Meuth SG, Klotz L
16
17 Vitiligo after alemtuzumab treatment: Secondary autoimmunity is not all about B
18
19 cells. *Neurology* 2018; **91**: e2233–e2237.
20
21
- 22
23 7. Jouary T, Taieb A. Halo nevi and vitiligo. In: Vitiligo (eds. Picardo M, Taieb A),
24
25 2010; Springer, Berlin, 61–64.
26
27
- 28
29 8. Jones JL, Phuah CL, Cox AL, Thompson SA, Ban M, Shawcross J, Walton A,
30
31 Sawcer SJ, Compston A, Coles AJ. IL-21 drives secondary autoimmunity in
32
33 patients with multiple sclerosis, following therapeutic lymphocyte depletion with
34
35 alemtuzumab (Campath-1H). *J Clin Invest* 2009; **119**: 2052–2061.
36
37
- 38
39 9. Boniface K, Jacquemin C, Darrigade AS, Dessarthe B, Martins C, Boukhedouni N,
40
41 Vernisse C, Grasseau A, Thiolat D, Rambert J, Lucchese F, Bertolotti A, Ezzedine
42
43 K, Taieb A, Seneschal J. Vitiligo skin is imprinted with resident memory CD8 T
44
45 cells expressing CXCR3. *J Invest Dermatol* 2018; **138**: 355–364.
46
47
- 48
49 10. Clark RA, Watanabe R, Teague JE, Schlapbach C, Tawa MC, Adams N,
50
51 Dorosario AA, Chaney KS, Cutler CS, Leboeuf NR, Carter JB, Fisher DC, Kupper
52
53 TS. Skin effector memory T cells do not recirculate and provide immune
54
55 protection in alemtuzumab-treated CTCL patients. *Sci Transl Med* 2012; **4**: 117ra7.
56
57
58
59
60

Figure 1

(A) Clinical appearance of the posterior trunk of the patient. The insert depicts the halo-like nevus from which the biopsy was taken; (B) Haematoxylin & eosin staining and (C) Pan Mel immunohistochemistry of the lesion (as shown in insert of Fig 1A). Note the absence of epidermal melanocytes; (D, E) CD3 immunostaining of the lesion. Note scattered T cells adjacent to the dermal melanocytes (D) and along the dermal-epidermal junction devoid of melanocytes (E).

For Peer Review

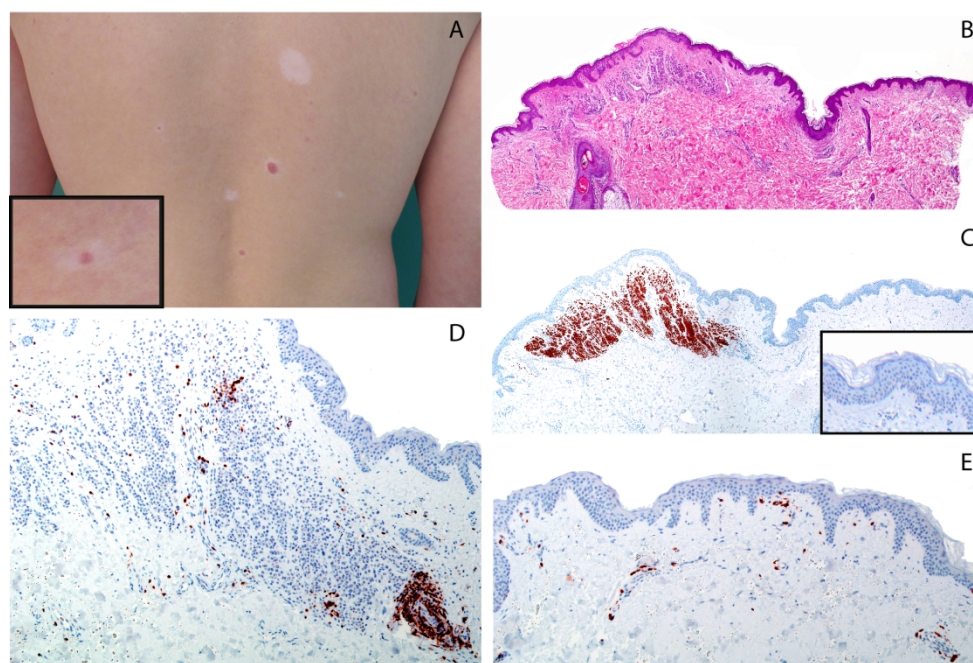
Table 1: Presence of circulating anti-melanocyte antibodies in the patient's serum before (Sept/2016) and after initiation of alemtuzumab (Aug/2018 and Feb/2019).

Antigen used in radioligand binding assay ¹	Before treatment antibody index ²	After ² treatment antibody index ²	After ² treatment antibody index ²	Upper limit of normal for radioligand binding assay ³
LMNA	1.23	1.09	1.15	1.59
TYR	2.23	12.89	13.12	1.72
TYRP1	1.25	3.04	3.89	1.57
DCT	0.95	0.99	1.03	1.36
PMEL	1.14	1.26	1.19	1.45
TH	1.09	1.12	1.15	1.84
MCHR1	0.95	0.94	0.98	1.48

¹Radioligand binding assays (RLBA) were used to test for antibodies against LMNA, laminA, TYR, tyrosinase; TYRP1, TYR-related protein 1; DCT, dopachrome tautomerase; PMEL, premelanosome protein; TH, tyrosine hydroxylase; and MCHR1, melanin-concentrating hormone receptor 1.

²An antibody index for the patient's serum was calculated for each RLBA as: counts per min (cpm) immunoprecipitated by serum/mean cpm immunoprecipitated by 20 healthy control sera. Positive antibody indices are in bold.

³The upper limit of normal for the RLBA was calculated using the mean antibody index + 3 SD of the population of 20 healthy control sera.



(A) Clinical appearance of the posterior trunk of the patient. The insert depicts the halo-like nevus from which the biopsy was taken; (B) Haematoxylin & eosin staining and (C) Pan Mel immunohistochemistry of the lesion (as shown in insert of Fig 1A). Note the absence of epidermal melanocytes; (D, E) CD3 immunostaining of the lesion. Note scattered T cells adjacent to the dermal melanocytes (D) and along the dermal-epidermal junction devoid of melanocytes (E).

196x136mm (300 x 300 DPI)