

This is the peer reviewed version of the following article:

Tyrosine kinase inhibitors: potential use and safety considerations in HIV-1 infection

Coiras, M., Ambrosioni, J., Cervantes, F., Miró, J. M., & Alcamí, J. (2017). Tyrosine kinase inhibitors: potential use and safety considerations in HIV-1 infection. Expert opinion on drug safety, 16(5), 547–559.

which has been published in final form at:

https://doi.org/10.1080/14740338.2017.1313224

Tyrosine kinase inhibitors: potential use and safety considerations in HIV-1 infection

Abstract

<u>Introduction</u>: Infection caused by HIV-1 is nowadays a chronic disease due to a highly efficient antiretroviral treatment that is nevertheless, unable to eliminate the virus from the organism. New strategies are necessary in order to impede the formation of the viral reservoirs, responsible for the failure of the antiretroviral treatment to cure the infection. <u>Areas covered:</u> The purpose of this review is to discuss the possibility of using tyrosine kinase inhibitors (TKIs) for the treatment of HIV-1 infection. These inhibitors are successfully used in patients with distinct cancers such as chronic myeloid leukemia. The most relevant papers have been selected and commented.

<u>Expert opinion</u>: The family of TKIs are directed against the activation of tyrosine kinases from the Src family. Some of these kinases are essential for the activation of CD4+ T cells, the major target of HIV-1. During acute or primary infection the CD4+ T cells are massively activated, which is mostly responsible for the generation of the reservoirs, the spread of the infection and the destruction of activated CD4+ T cells, infected or not. Consequently, we discuss the possibility of using TKIs as adjuvant of the antiretroviral treatment against HIV-1 infection mostly, but not exclusively, during the acute/recent phase.

Keywords: HIV/AIDS; Tyrosine kinase inhibitors; chronic myeloid leukemia; immunomodulation; viral reservoirs; T-cell activation

Article highlights

- HIV-1 infection is currently incurable due to the formation of viral reservoirs.
- Antiretroviral treatment has transformed HIV-1 infection into a chronic disease that needs life-long treatment.
- CD4+ T cell massive activation during HIV-1 acute infection is mostly responsible for viral spread, the formation of the reservoirs and the destruction of CD4+ T cells.
- Control of CD4+ T cell activation could avoid viral replication and spread and, consequently, the formation of the viral reservoirs.
- Src tyrosine kinases are essential for the activation of CD4+ T cells and their inhibition could avoid the formation of HIV-1 reservoirs.
- Tyrosine kinase inhibitors currently used for treating chronic myeloid leukemia could be potential adjuvants of the antiretroviral treatment during HIV-1 infection.

Body of the review paper

1. Lymphocytic activation as an antiviral target

1.1. The role of immune activation in the HIV-1 cycle

HIV-1 may infect both quiescent and activated CD4+ T cells (1), but only activated cells support HIV-1 replication (2). Overall, resting/non-activated T cells are highly restrictive to HIV-1 infection and replication due to several factors (Figure 1). First, CCR5 chemokine receptor, the major HIV-1 co-receptor, is highly expressed only in activated T cells (3). Second, as resting T cells are in G0 phase, DNA synthesis is not required and dNTP levels are low. This prevents an efficient reverse transcription of the viral RNA. Among the checkpoint factors that regulate cell cycle, the phosphohydrolase SAMHD1 (SAM domain and HD domain-containing protein 1) decreases dNTP levels when is active, thereby constituting an essential antiviral factor against HIV-1 infection in resting cells (4). Third, resting, non-cycling T cells have low ATP levels, which impedes an active transport of the viral pre-integration complex to the nucleus (5). Finally, resting T cells do not require the active expression of genes involved in the generation of the immune response. Therefore, the activity of transcriptional factors such as NF- κ B, SP1 and NFAT that are necessary for HIV-1 transcription (6, 7) is very low in resting T cells. However, triggering an immune response leads to the activation of these transcription factors that are required not only for the expression of genes encoding receptors, cytokines and chemokines, but also for driving the expression of viral genes involved in the production of new virions.

The first three mechanisms are required for *de novo* HIV-1 infection, viral reverse transcription and integration, while the last mechanism is necessary to trigger an efficient transcription and elongation of the HIV-1 genome from a quiescent, integrated molecular double-stranded DNA form in the host genome called provirus. Therefore,

HIV-1 has successfully adapted to the cellular environment of CD4+ T cells by using a Trojan horse mechanism. The provirus remains in a state of latency in resting CD4+ T cells, allowing viral persistence and establishing viral reservoirs. Due to the lack of transcription and expression of viral mRNA and proteins, these infected resting cells are not recognized by the immune system and escape the immune surveillance. In contrast, when these latently infected lymphocytes become activated in the context of the normal generation of an immune response, the cell provides HIV-1 with the transcription factors required to induce the expression of the viral genome and a massive viral replication occurs in the activated CD4+ T cells (8).

This viral strategy represents a major barrier to HIV-1 cure due to the establishment of undetectable viral reservoirs in quiescent CD4+ T cells (9). Paradoxically, normal lymphocytic activation in response to HIV-1 infection or other pathogens, results in a robust viral replication, a permissive environment to new rounds of HIV-1 infection and the destruction of activated and thereafter infected lymphocytes. Lymphocytic activation is particularly detrimental in early phases of HIV-1 infection and in those lymphoid organs in which a high level of cellular activation occurs due to the encounter with antigens such as the lymph nodes and mostly, the gut-associated lymphoid tissue (GALT). Actually, it has been shown that in acute HIV-1 infection up to 80% of CD4+ T cells from the GALT are destroyed through different cytopathic mechanisms driven by HIV-1, being among them direct cytopathic effect, apoptosis and pyroptosis (10-14). However, a small subset of infected CD4+ T cells will persist and create the reservoir due to not completely known mechanisms such as the expression of the survival protein BCL-2 in long-lived CD4+ T cells (15) or the lingering presence of HIV-1 regulator protein Tat (16). On the other hand, T-cell activation is required to built-up effective immune responses targeting the virus but this sustained activation during the chronic

illness also contributes to the clonal exhaustion of virus-specific CD8+ T cells (17), as well as to the destruction of CD4+ T cells, particularly those committed to the recognition of HIV-1 antigens (18). These specific anti-HIV-1 T-cell cytotoxic and proliferative responses are not restored despite months of effective ART (19). Moreover, persistent replication occurs during the chronic phase of the infection even in the presence of an efficient antiretroviral treatment (ART) (20-23). In consequence, new strategies must be developed in order to avoid massive activation and infection of CD4+ T cells during the acute phase of HIV-1 infection and to interfere with the low-level, persistent viral replication that occurs during the chronic stage.

1.2. Immunomodulatory strategies in HIV-1 infection

Due to the close relationship between immune activation and viral replication, different authors proposed a controlled immune suppression together with ART. Actually, using immunomodulatory therapies in combination with ART could help control the spread of the infection, the formation of the viral reservoir, the development of escape mutant variants and the exhaustion of the immune system due to the sustained activation. In this regard, some clinical trials performed in HIV-infected patients who were treated with immunosuppressive drugs as adjuvant of ART showed contradictory results, such as those using cyclosporine A (CsA). Rizzardi et al. (24) did not report a significant effect in plasma viral load of HIV-infected patients treated with CsA and ART, but they observed a persistent increase in CD4 count that was sustained after withdrawal of CsA. However, Calabrese et al. (25) showed no positive effect on CD4 count and they even reported an increase in the viral load after administration of CsA. Markowitz et al. (26) also described that administering CsA and ART to patients with acute early HIV-1 infection did not provide any immunological or virological benefit. Therefore, while some studies showed a clinical benefit in using CsA with ART (24, 27, 28), others did not (25, 26, 29, 30).

It has also been described that both mycophenolic acid (MPA) and its ester derivate mycophenolate mofetil (MMF) suppress HIV-1 infection by interfering with the synthesis of guanosine nucleotides (31). Some clinical trials stated that MPA and MMF decreased both titers of infected CD4+ T cells (31) and viral load (32) in ART-treated HIV-infected patients. However, Sankatsing et al. (33) did not find significant changes in the viral RNA between MMF-treated and untreated groups. Besides, although it was determined that the combination of MMF and ART could delay viral rebound and improve control of viral replication during structured treatment interruption (STI), this strongly correlated with the simultaneous inhibition of lymphocyte proliferation (34, 35). This correlation was also observed in HIV-infected patients that were treated with hydroxyurea (HU) and ART before STI (36). Consequently, the interference with lymphocyte proliferation was essential to cause a delay in the viral rebound during STI likely due to a smaller size of the reservoir. However, this does not guarantee that the immunosuppressors could also have a detrimental activity against the viral reservoir that is already formed. Moreover, although no major toxicity was reported in these trials, clinicians were reluctant to use immunosuppressive drugs in patients that could already be immunosuppressed. Therefore, more specific immunosuppressants must be found in order to direct their inhibitory effect specifically to the HIV-1 target cells without causing a general immunosuppression.

1.2.1. Role of tyrosine kinases in lymphocyte activation

Lymphocyte activation by antigens occurs through the interaction of the T-cell receptor (TCR) with cytoplasmic non-receptor tyrosine protein kinases. One of the most important tyrosine kinases for lymphocyte activation is LCK that induces the activation

of downstream kinases such as the protein kinase C theta (PKC0) (37). PKC0 in turn activates transcription factors such as NF-KB, NFAT and AP-1 (38, 39) that are essential for both T-cell activation and HIV-1 replication. LCK belongs to the SRCfamily kinases (SFKs), a family of nine non-receptor tyrosine kinases that perform highly specialized functions within the various lineages (40). B cells primarily express LYN, FYN, and BLK, whereas T cells mostly express LCK and FYN-T, a spliced form of FYN that partially encodes the kinase domain (41). LCK (56 kDa) is expressed constantly through most of the lifespan of a T cell but the expression of FYN (59 kDa) mostly increases in mature cells (42). This implies that LCK has a unique role throughout T-cell development and function whereas FYN appears to be less critical for mature T-cell generation and function (43). Both molecules have several domains in common but LCK has a unique domain that mediates non-covalent interaction with CD4 and CD8 (44), which facilitates the initiation of T-cell receptor (TCR)/CD3 complex signal transduction during antigen recognition (43). Consequently, LCK is mostly found at the plasma membrane, whereas FYN colocalizes with centrosomal and mitotic structures (45, 46).

LCK activation is regulated through phosphorylation of the tyrosine residue Y394 located in the kinase domain. In resting T cells, this site is normally not phosphorylated, rendering LCK inactive. Upon TCR-major histocompatibility complex (MHC)/peptide interaction, LCK is recruited and phosphorylated at the activation loop Y394. Active LCK induces direct or indirect phosphorylation of several substrates such as SAMHD1 (47) and also specific tyrosine residues within the ITAMs (immunoreceptor tyrosine-based activation motifs) of CD3 chains, leading to the recruitment, phosphorylation and activation of SYK-family kinases (ZAP-70 in T cells) to the ITAMs. This triggers a signaling cascade that initiates Ca2+ influx, activates protein kinase C theta (PKC0) and

mitogen-activated protein kinase (MAPK) cascades, ultimately resulting in NFAT, NF- κ B, and AP-1 activation (39, 48), as well as full T-cell activation, characterized by entry into cell cycle, changes in gene expression and T-cell effector function (43). NF- κ B is a family of transcriptional regulators considered a central mediator of the immune response as it mostly participates in promoting the expression of cytokines and chemokines, receptors for immune recognition and migration, and proteins for antigen presentation (49). Activation of NF- κ B is very fast and strong and some viruses have evolved to modulate NF- κ B signaling pathway in order to enhance viral replication and host cell survival, as well as to evade immune response (50). HIV-1 replication succeeds mostly due to the presence of NF- κ B consensus binding sites in the enhancer proximal region of the viral long terminal repeat (LTR) promoter. In fact, the interference with NF- κ B in HIV-infected cells inhibits viral transcription and replication (51, 52).

Consequently, using immunomodulators able to suppress selectively the activity of SFKs such as LCK would be an advantage over the immunosuppressive strategies that have been tested so far for the treatment of HIV-1 infection as they would be directed specifically against the activation of the main target of the virus but would not cause a general immunosuppression (53, 54). In this regard, tyrosine kinase inhibitors (TKIs) are currently used in clinic for the treatment of chronic myeloid leukemia (CML) and other types of cancer such as lung cancer, thyroid cancer, or renal cell carcinoma.

2. Overview of TKI approved for CML: clinical practice, effects and toxicity

CML is a neoplastic disorder of the hematopoietic stem cell characterized by the uncontrolled growth of myeloid cells in the bone marrow and their accumulation in peripheral blood (55). CML occurs as a result of an acquired reciprocal translocation between chromosomes 9 and 22 in the hematopoietic stem cells, resulting in the

Philadelphia chromosome, whose molecular counterpart is the BCR-ABL oncogene. Thus, the BCR gene, encoding for a protein whose function is not completely understood, is located in the breakpoint of chromosome 22, whereas the ABL protooncogene, encoding for a tyrosine kinase protein, is located at the breakpoint of chromosome 9 (56, 57). Therefore, the translocation produces the chimeric fusion protein BCR-ABL, encoded by both *BCR* and *ABL1* genes and that is responsible for the development of CML due to its uncontrolled tyrosine kinase activity (58, 59).

The treatment of CML is currently based on the use of a group of oral small molecules that selectively inhibit the BCR-ABL oncogenic protein (60). These molecules are summarized in Table 1, adapted from Steegmann et al (61). Imatinib, the first TKI against BCR-ABL introduced in clinical practice, revolutionized the therapy of CML due to the achievement of unprecedentedly high rates of deep responses that translate into high progression-free and overall survival similar to that of the control population (62). The so-called second-generation TKIs, including nilotinib, dasatinib, and bosutinib, were introduced later in CML treatment (63) and were initially given to patients resistant or intolerant to imatinib. Nilotinib is at least 20-fold and dasatinib at least 300-fold more potent than imatinib (65). More recently, a third-generation TKI, ponatinib, has been approved for the treatment of patients with the T315I mutation of the *BCR-ABL* kinase domain, which is resistant to the other TKIs, and for patients with resistance to the second-generation TKIs (66).

In addition to inhibiting the BCR-ABL protein, the TKIs have other molecular targets, a fact that would explain their different toxicity profile (63). In this regard, imatinib is the TKI with a better toxicity profile since, although it causes frequent, low-grade chronic side effects, no safety issues have been observed with prolonged follow-up. The main

toxicity of nilotinib is the development of ischemic events (67), a complication registered in 12 to 18% of patients at 5 years of treatment. Pleural effusion is the most characteristic adverse effect of dasatinib (68), appearing in 30% of patients at one year, more frequently in patients with previous chronic obstructive pulmonary disease. This complication requires dose reduction or cessation of dasatinib administration. In CML patients, with the usual doses, other adverse reactions that have been reported as very frequent (> 1/10 patients) for dasatinib (69), as is shown in Table 2. Diarrhea and elevation in serum transaminases are the main toxicities of bosutinib (70). Finally, ponatinib is the most toxic TKI, due to its association with the appearance of ischemic events in a substantial proportion of the patients (71). All TKIs toxicity is detailed in Table 2.

A number of immunological effects of TKIs have also been described (72). Imatinib produces lymphopenia and decreased immunoglobulin plasma levels (73), whereas nilotinib inhibits CD8+ T cell function (74). Dasatinib inhibits the proliferation and activation of T cells and suppresses the cytotoxic activity of Natural Killer cells *in vitro* (75). In contrast, 30% of patients receiving dasatinib develop absolute lymphocytosis with clonal expansion of cytotoxic T cells and Natural Killer cells (76-78). However, patients usually show a good therapeutic outcome with deep responses to dasatinib (78) and no increased susceptibility to opportunistic infections (79). This supports the notion that the immunological response triggered by dasatinib and targeting Natural Killer cells and CD8+ T lymphocytes would be a mechanism that contributes to its therapeutic activity. In fact, control of lymphocyte activation has been associated to increased immune responses and disease control in cancer (80, 81).

3. Pre-clinical data on the use of TKIs in HIV-1 infection

Once the provirus is integrated, it remains hidden inside the resting CD4+ T cells in a latent state until the TCR/CD3-mediated activation promotes an active viral transcription and new viral particles are produced. Consequently, the control of T-cell activation might provide a way to control HIV-1 infection and replication. Our group described previously that the selective inhibition of PKC0 activity interfered with HIV-1 transcription because they inhibit the activity of the transcription factors NFAT, NF- κB and AP-1 (53, 82). We also demonstrated that PKC θ selective inhibitors were able to partially interfere with SAMHD1 phosphorylation at T592, preserving the antiviral activity and avoiding HIV-1 proviral integration. One of the main barriers that HIV-1 encounters when it enters the target cell is SAMHD1. As indicated above, SAMHD1 regulates cell cycle progression and HIV-1 reverse transcription by depleting the intracellular pool of dNTPs (83), thereby interfering with the integration of the provirus. SAMHD1 activity is regulated by phosphorylation of threonine residue T592 by Cdk1 and cyclin A2 during T-cell activation. Therefore, when SAMHD1 is phosphorylated at T592 it loses the antiviral activity because T-cell cycle progresses and HIV-1 vital cycle can be completed (84). Therefore, inhibition of PKC0 would inhibit through two different mechanisms: on one hand, a decrease in lymphocytic activation would result in a quiescent state highly restrictive to HIV infection; on the other hand, through inhibition of SAMHD1 phosphorylation, PKC0 inhibitors will directly tackle reverse transcription, integration and the formation of reservoirs. As LCK is upstream PKC0 signaling pathway, we determined that the inhibition of LCK activity with dasatinib was able to inhibit HIV-1 replication (54). Dasatinib inhibited HIV-1 replication of PBMCs isolated from healthy donors after activation with PHA/IL-2 in the presence of this TKI before infecting them in vitro with HIV-1 for 5 days (IC50 = 16 nM; $CC50 > 10 \mu M$) (Figure 2A). SAMHD1 phosphorylation at T592 was inhibited at 18.75 nM (Figure 2B),

proving that preservation of SAMHD1 antiviral activity and interference with HIV-1 reverse transcription and subsequent integration were the main mechanisms of action for dasatinib-mediated inhibition of HIV-1 infection in primary T cells (47, 54). It should also be considered that dasatinib has a wide range of activity, being able to inhibit BCR-ABL and all SFKs as Lck, whereas other ITKs such as imatinib are quite selective of BCR-ABL (85). This implies that not all the TKIs might have the same mechanism of action.

Dasatinib is administered chronically to CML patients at a dose of 100 mg dq. Our results proved that concentrations of dasatinib displaying antiviral activity in vitro may be lower than those used in clinical practice for the treatment of CML. Full inhibition of SAMHD1 phosphorylation at 18.75 nM demonstrated that dasatinib exhibits linear pharmacokinetics, thereby suggesting a proportional increase in AUC (area under the concentration-time curve) and linear elimination characteristics over the dose range of 25 mg to 120 mg BID (twice a day). The antiviral activity of dasatinib in vitro was also confirmed ex vivo by isolating PBMCs from five CML patients on chronic treatment with dasatinib for at least 2 years. PBMCs were activated with PHA/IL-2 48 hours before ex vivo infection with HIV-1 and then incubated for 5 days. Similar results to PBMCs treated in vitro with dasatinib were obtained, observing a decrease in HIV-1 proviral integration and transcription, as well as lower phosphorylation of SAMHD1 in CD4+ T cells from CML patients upon activation (54). These results proved not only that the dose of dasatinib used for the treatment of CML was able to interfere with HIV-1 infection, but also that long-lasting intracellular dasatinib levels able to inhibit HIV-1 replication can be achieved with doses currently used in CML treatment.

4-Potential use of TKIs in HIV-1 infection: timing of infection and risk of toxicity

ART is the cornerstone for HIV-1 treatment. Although ART has tremendously evolved since 1995 (start of modern triple ART) and has transformed the lethal HIV-1 infection into a chronic disease, no other adjuvant or alternative strategy has proved to be effective so far (86). We hypothesized that TKIs, particularly dasatinib that inhibits a large spectrum of tyrosine kinases, could reduce the size of the viral reservoirs during acute/recent infection (54), and even reduce replenishment of the reservoirs during chronic infection. Consequently, although its impact may be higher in acute/recent infection, it could also be useful in chronically infected patients. Dasatinib, when administered during short-time periods, might also decrease immune activation, interfering with the characteristic chronic inflammation that leads to non-AIDS comorbidities related to HIV-1 infection. This sustained, low-grade inflammation and increased immune activation are strongly associated with a heightened risk for cardiovascular disease, osteoporosis, cancer, physical function impairments and frailty, among other non-AIDS-defining events and mortality (87, 88).

As described above, a number on immunological approaches have been evaluated trying to improve the clinical outcome of HIV-1 infection. In this regard, TKIs, and particularly dasatinib, may provide an interesting additional antiretroviral tool due to several different mechanisms: first, a direct antiretroviral effect through the inhibition of SAMHD1 phosphorylation and consequently, of proviral integration, which would lead to a reduced viral reservoir; second, an indirect anti-reservoir effect through the inhibition of the clonal expansion of the infected cells due to its cytostatic effect (54); and finally, a putative role of increased Natural Killer or $\gamma\delta$ T lymphocytes activity after TKI discontinuation has been proposed in CML patients with treatment-free remission (89) (90, 91). If such mechanisms produced by TKI could also contribute to the immune control of HIV replication remains speculative. All three effects might be useful to improve viral control at any stage of HIV-1 infection, both in acute/recent and chronically infected patients. Although the third mechanism is completely speculative in the HIV field, a growing body of evidence suggests that the immune system has a major role in determining the therapeutic efficacy of TKIs (92). Figure 3 summarizes the potential mechanisms of TKIs to interfere with HIV-1 replication and spread, as well as their possible effect on the size of the viral reservoir. Besides, TKIs could also be beneficial against other aspects of HIV infection such as the Kaposi sarcoma (93).

However, the addition of one TKI to ART during early HIV-1 infection might have the highest impact, reducing the final size of the reservoir, which has been associated to the time of virological control when ART is stopped (the lowest the reservoir, the longest the period without viral replication) (94, 95). Since the reservoir is formed very early during initial Fiebig stages (Figure 4) (96), only a short period (4 to 12 weeks) of TKI administration might be enough to obtain a clinical benefit, reducing the risk of increased toxicity by TKI. In addition, as previously described, doses required may be lower than those used in clinical practice for CML, decreasing even further the potential for side effects. Finally, if an immunomodulatory effect and increased long-acting immune control of HIV-1 replication effect induced by dasatinib is confirmed, this may be the basis for intermittent administration during brief periods, reducing further the potential for serious toxicity. Indeed, some CML patients exhibit what is called a treatment-free remission of the disease after treatment with TKI (89). In the particular case of HIV-1 infection, treatment with TKIs in acute infection could block the development of specific HIV-1 immune responses, mostly cytotoxic responses that are raised in the first weeks of infection. This delay in building up immune responses against HIV-1 could paradoxically be beneficial at mid-term by two different reasons: first, the immune responses will be generated in the absence of very high viremia levels

that result in abnormal immune activation and apoptosis (97). Actually, it has been demonstrated that very early treatment of HIV-1 infection blocks high peaks of viremia and results in better immune responses against viral replication (98). Second, it has been described that HIV-1 variants resistant to cytotoxic T lymphocyte (CTL)-specific responses are easily generated and stored in the reservoirs in the acute phase of infection due to the accelerated replication and the generation of escape variants (99). This viral escape that leads to failure of immune control compromises the future potential use of CTL-based therapeutic vaccines raised against immunodominant epitopes. The down-modulation of immune activation during the acute phase of infection could contribute to overcome the emergence of resistant variants by delaying specific HIV-1 responses within a context of lower viremia and normal cellular activation. Moreover, Shytaj et al (100) described that the restriction of CD4+ T cell activation in macaques infected with the simian immunodeficiency virus (SIV) was not only well tolerated, but even the SIV-specific cell-mediated immunity was increased. This suggests that likely TKIs would not negatively influence HIV-specific immune responses.

Available information regarding the antiretroviral activity of TKIs is very limited for all the drugs in this family (101). However, the anti-HIV-1 effects of dasatinib have been evaluated and it seems to be the most attractive drug of the class for this use (54, 102). There is not clinical experience with dasatinib in HIV-infected population. Therefore, the potential toxicity should be considered from the available information in CML patients and then extrapolated to the eventual clinical use in HIV-infected patients. Currently, dasatinib is indicated at usual doses ranging from 100 to 140 mg/day for the treatment of adult patients with Philadelphia chromosome positive (Ph+) CML in chronic, accelerated or blastic phase, and Ph+ acute lymphoblastic leukemia (ALL) (69). In patients who tolerate this medication and show haematological response, the drug is administered indefinitely. Safety profile of dasatinib has been recorded from almost 3,000 patients included in clinical trials, most of them patients with CML who failed to or were intolerant to imatinib (69). The tolerability and rate of adverse reactions in patients with HIV-1 infection, using lower doses (ranging 20 to 70 mg) and for a shorter periods (up to 16 weeks) is unknown, but is expected to be lower.

However, it is essential to consider that in patients with both acute/recent or chronic HIV-1 infection, the addition of one TKI to ART regimen may represent a risk for increased toxicity and frequency of the side effects listed in Table 2. Potential raising issues include, first, bone marrow suppression, as HIV-1 infection itself has been associated with decreased numbers of red blood cells, leukocytes and platelets (103). Lymphopenia related to uncontrolled HIV-1 replication is especially prominent during acute infection (104) and, as previously explained, some TKIs such as imatinib induces lymphopenia. In chronic and advanced HIV-1 disease, there is a global dysfunction of bone marrow function and all three blood series may be reduced (103). The addition of a potential bone marrow suppressor such as the TKIs may increase this effect. Second, the high risk of infections should be considered as the most remarkable clinical expression of fully developed AIDS is the emergence of opportunistic diseases and tumors related to advanced immunodeficiency (105). Although infrequent, opportunistic infections may even arise as a complication of acute HIV-1 infection, when CD4 T cell count reaches the lowest level (nadir). In this context, the addition of one TKI to ART regimen may potentiate this risk. Hepatitis B reactivation has been described in patients receiving TKIs who were previously infected with hepatitis B virus (HBV), even those with no active replication (isolated anti-HBVc positive patients) (106). Since approximately 10% of HIV-1 infected patients are co-infected with HBV (107), particular attention should be paid to the potential reactivation of HBV in these patients,

although several drugs used in current antiretroviral regimens, such as lamivudine, emtricitabine or tenofovir have also anti-HBV activity.

Finally, potential serious drug-drug-interactions (DDI) may occur as several antiretroviral drugs are metabolized by Cytochrome P450 3A4 (CYP3A4). In particular, non-nucleoside reverse transcriptase inhibitors (NNRTI) are inducers of this enzyme, whereas cobicistat and ritonavir are powerful inhibitors (108), which is the basis of pharmacokinetics enhancement of protease inhibitors (PI) and elvitegravir (an integrasestrand transfer inhibitor -InSTI-). Dasatinib, as well as other TKIs such as nilotinib, bosutinib and ponatinib, are also metabolized though CYP3A4 pathway (109-111). In consequence, standard doses of dasatinib or other TKIs might become toxic when coadministered with ritonavir, cobicistat or any other potent CYP3A4 inhibitor. In that context, ART regimens containing drugs such as dolutegravir or raltegravir should be considered if dasatinib is co-administered, since these molecules have no expected drugdrug interactions with TKI and are among the preferred regimens in the most recent guidelines (112, 113). On the other hand, non-specific, general, side effects, such as fatigue or gastrointestinal intolerance may also be relevant, since they could reduce the optimal adherence to ART, which is essential for treatment efficacy, particularly during ART initiation.

Conclusion:

At first, TKIs, and particularly dasatinib, may appear very attractive as adjunctive therapy for treating HIV-1 infection, especially in specific clinical settings such as acute/recent infection. However, the risk for increased toxicity and adverse events has to be carefully balanced. Initial evaluation should target patients who most likely could be helped by this intervention with the lowest risk for toxicity, such as patients with high CD4 T cell count, no active co-infections, and no previous conditions that could risk for

increased toxicity from TKIs (e.g., leucopenia). In addition, DDI should be carefully considered and drugs interfering with CYP3A4 inhibiting activity should be avoided; raltegravir or dolutegravir based-regimens can be used in combination with TKI. However, the potential virological benefit of TKIs and particularly dasatinib seems to be enough to consider performing controlled pilot trials, closely monitored and with carefully selected patients. Reduction of doses and/or shortening the duration of treatment, or even intermittent use to decrease the potential side effects rate, as well as avoiding DDI, might prove that using TKIs in HIV-1 infected individuals could be highly beneficial to reduce the reservoir size from the beginning, making possible longer structured treatment interruption of ART.

Expert opinion

A cure for HIV-1 infection is not available yet due to viral persistence in the reservoirs. To address this issue different strategies have been proposed (for a review see Coiras et al. (8) and Table 3). Shock and kill strategies with latency reversing agents (LRAs), aiming at the reactivation and destruction of the viral reservoirs, have been assayed in clinical trials with few success due to low drug potency and toxic effects. Bone-marrow transplantation approaches using HIV-resistant CCR5 deficient lymphocytes are only indicated in very special cases. Gene therapy tools are under development but in case an efficient system could be selected in the near future, this strategy is not affordable for all candidate patients due to economic and technical constrains. Finally, enhancement of immune responses aiming at the elimination of the HIV-1 reservoirs has shown interesting results in the SIV model but raises safety concerns for its use in humans (114). Therapeutic vaccination to increase specific cell responses against HIV-1 faces the challenge of hidden reservoirs carrying escape variants to immunodominant responses (99). Overall, there is not a clear strategy to tackle the HIV-1 reservoirs and achieve a functional cure of infected patients. Therefore, new approaches should be considered. In this context, the use of immunosuppressive/immunomodulatory drugs in a disease leading to immune suppression appears to be a paradox. However, although HIV-1 can infect both resting and activated CD4+ T cells, it only replicates in activated cells because they have adequate amounts of dNTPs, ATP, and active transcription factors to permit a complete replication cycle. Moreover, in activated CD4+ T cells the antiviral factor SAMHD1 is phosphorylated, which permits not only the progression of the cell cycle but also an effective viral reverse transcription. Therefore, HIV-1 replication is mostly successful due to the massive activation of CD4+ T cells that

occurs at very high levels during the first stages of the infection, allowing the early formation of viral reservoirs that renders the infection, from this moment, incurable.

Based on these observations, some attempts have been made to try to control immune activation during HIV-1 infection. However, although the rationale for using immunosuppressants as adjuvants of conventional ART was solid, the results were controversial as the experimental design and the stages of infection of the patients that entered in the different clinical trials were very diverse and hardly comparable. Moreover, there was reluctance to use immunosuppressants in a disease that already causes immunosuppression, even in the presence of the antiretroviral treatment. However, one important conclusion was obtained from these clinical trials: the control of viral replication during STI was improved in those patients previously treated with a combination of immunosuppressants and ART, in whom a delay in viral rebound was observed in comparison with patients only treated with ART. But this improvement was achieved only when lymphocyte proliferation inhibited by the was immunomodulatory/cytostatic agent, suggesting that administering immunosuppressants during acute infection could be potentially useful to avoid massive CD4+ T cell activation and consequently, to restrain the formation of viral reservoirs. In fact, the smaller the size of the reservoir, the later the viral rebound during STI, and the better the prognosis of the disease. Besides, decreasing immune activation during acute infection can result in lower lymphocyte destruction by HIV-1 through different mechanisms and contributes to preserve specific HIV-1 immune responses and avoid viral escape mutants. Although the rationale for using immunosuppressants with ART in patients with early acute infection can be considered, their use in chronically infected patients is more controversial as they would not directly beneficiate from this regimen by having reduced the size of the viral reservoir. However, it could be recommendable to

administer the cytostatic drugs in these patients in order to avoid low-level viral replication and the chronic activation of the immune system that eventually, leads to the failure of the immune response. TKIs could also be used to limit the viral reservoir replenishment during STI following the administration of LRAs.

The family of tyrosine kinase inhibitors (TKIs) that are currently used in clinic for the treatment of several types of cancer such as chronic myeloid leukemia, emerges as a potential adjuvant of ART in acutely HIV-infected patients to reduce or impede the formation of the viral reservoir, as well as in chronically infected patients to avoid viral replication and to help the immune system to preserve its function. Some of these TKIs are directed against SRC kinases that are specifically expressed in the main viral targets: the CD4+ T cells. Therefore, their use would not cause a general immunosuppression like the immunosuppressants that were used in previous clinical trials. In fact, they would achieve two specific goals, depending on the targeted tyrosine kinase: 1) to avoid the phosphorylation – and subsequent inactivation – of SAMHD1 through the interference with LCK activity; and 2) to restrict CD4+ T cell activation due to their cytostatic effect, impeding HIV-1 replication.

From a clinical point of view, several aspects should be considered. The most adequate TKI should display a broad range of inhibition of tyrosine kinases to make possible the interference with HIV-1 replication in vitro by preserving the antiviral effect of SAMHD1 and by inducing a cytostatic effect on CD4+ T cells. This would ensure a resistance to infection in the most important targets of HIV-1, as well as to avoid the formation of the viral reservoir. Dasatinib, a TKI shares these characteristics but although is generally well tolerated, several serious adverse effects have been described for this drug and the possible interaction with the antiretroviral treatment should be carefully considered. Moreover, dasatinib is a potent cytostatic of T cells, and although

it does not appear to make CML patients on chronic treatment more susceptible to infection, is unknown whether they could enhance opportunistic infections in immunodeficient patients. A pilot clinical trial with a small number of patients carefully selected would be advisable to determine the feasibility of this approach, evaluating both immunological and virological benefits for the patients with this chronic infection.

Figure legends

Figure 1. Factors that induce high restriction to HIV-1 infection and replication in resting/non-activated CD4+ T cells.

Figure 2. (A) PBMCs isolated from healthy donors were activated with PHA/IL-2 in the presence of dasatinib at different concentrations before infecting them in vitro with NL4-3_Renilla strain (1ng p24 per million of cells) for 5 days. The presence of Renilla in this strain allowed the monitorization of the infection with luminescence (relative light units, RLUs). IC50 and CC50 were calculated by using GraphPad software. The selectivity index (SI = CC50/IC50) was calculated to determine the therapeutic index, giving maximum antiviral activity with minimal cytotoxicity. R² is a measure of goodness-of-fit of linear regression. (B) PBMCs isolated from healthy donors were activated with IL-7 for 5 days in the presence of dasatinib at different concentrations and then SAMHD1 phosphorylation was analyzed by immunoblotting using specific antibodies. β-actin was used as loading control.

Figure 3. Mechanisms of inhibition of the HIV-1 cycle by TKIs. TKIs specifically block SAMHD1 phosphorylation and inhibits viral reverse transcription (1). Through inhibition of T-cell activation, TKIs inhibit full viral replication (2) and the reservoir expansion by interfering with the homeostatic proliferation of latently infected T cells (3). Finally, enhancement of the antiviral activity of Natural Killer (NK) cells can contribute to the destruction of HIV-infected CD4+ T lymphocytes (4).

Figure 4. Fiebig stages of HIV-1 infection (modified from Fiebig et al. (96)).

References

1. Zhang Z, Schuler T, Zupancic M, Wietgrefe S, Staskus KA, Reimann KA, et al. Sexual transmission and propagation of SIV and HIV in resting and activated CD4+ T cells. Science. 1999;286(5443):1353-7. PubMed PMID: 10558989.

2. Zack JA, Arrigo SJ, Weitsman SR, Go AS, Haislip A, Chen IS. HIV-1 entry into quiescent primary lymphocytes: molecular analysis reveals a labile, latent viral structure. Cell. 1990;61(2):213-22. PubMed PMID: 2331748.

3. Bermejo M, Martin-Serrano J, Oberlin E, Pedraza MA, Serrano A, Santiago B, et al. Activation of blood T lymphocytes down-regulates CXCR4 expression and interferes with propagation of X4 HIV strains. Eur J Immunol. 1998;28(10):3192-204. doi: 10.1002/(SICI)1521-4141(199810)28:10<3192::AID-IMMU3192>3.0.CO;2-E. PubMed PMID: 9808188.

Baldauf HM, Pan X, Erikson E, Schmidt S, Daddacha W, Burggraf M, et al. SAMHD1 restricts HIV-1 infection in resting CD4(+) T cells. Nat Med. 2012;18(11):1682-7. doi: 10.1038/nm.2964. PubMed PMID: 22972397; PubMed Central PMCID: PMCPMC3828732.
** This article highlights for the first time one of the major mechanisms of resistance to infection in resting CD4 lymphocytes, the presence of the restriction factot SAMHD11

5. Bukrinsky MI, Sharova N, Dempsey MP, Stanwick TL, Bukrinskaya AG, Haggerty S, et al. Active nuclear import of human immunodeficiency virus type 1 preintegration complexes. Proc Natl Acad Sci U S A. 1992;89(14):6580-4. PubMed PMID: 1631159; PubMed Central PMCID: PMCPMC49545.

6. Alcami J, Lain de LT, Folgueira L, Pedraza MA, Jacque JM, Bachelerie F, et al. Absolute dependence on kappa B responsive elements for initiation and Tat-mediated amplification of HIV transcription in blood CD4 T lymphocytes. EMBO J. 1995;14(7):1552-60.

7. Bosque A, Planelles V. Induction of HIV-1 latency and reactivation in primary memory CD4+ T cells. Blood. 2009;113(1):58-65. doi: 10.1182/blood-2008-07-168393. PubMed PMID: 18849485; PubMed Central PMCID: PMCPMC2614643.

8. Coiras M, Lopez-Huertas MR, Perez-Olmeda M, Alcami J. Understanding HIV-1 latency provides clues for the eradication of long-term reservoirs. Nat Rev Microbiol. 2009;7(11):798-812. doi: 10.1038/nrmicro2223. PubMed PMID: 19834480.

** A classical review covering both the molecular mechanisms of HIV latency and reactivation and the different strategies to tackle the viral reservoirs

9. Finzi D, Hermankova M, Pierson T, Carruth LM, Buck C, Chaisson RE, et al. Identification of a reservoir for HIV-1 in patients on highly active antiretroviral therapy. Science. 1997;278(5341):1295-300. PubMed PMID: 9360927.

* A classical paper first describing and quantifying the viral reservoir in CD4 lymphocytes from HIV-infected patients

10. Brenchley JM, Schacker TW, Ruff LE, Price DA, Taylor JH, Beilman GJ, et al. CD4+ T cell depletion during all stages of HIV disease occurs predominantly in the gastrointestinal tract. J Exp Med. 2004;200(6):749-59. doi: 10.1084/jem.20040874. PubMed PMID: 15365096; PubMed Central PMCID: PMCPMC2211962.

11. Doitsh G, Cavrois M, Lassen KG, Zepeda O, Yang Z, Santiago ML, et al. Abortive HIV infection mediates CD4 T cell depletion and inflammation in human lymphoid tissue. Cell. 2010;143(5):789-801. doi: 10.1016/j.cell.2010.11.001. PubMed PMID: 21111238; PubMed Central PMCID: PMCPMC3026834.

12. Doitsh G, Galloway NL, Geng X, Yang Z, Monroe KM, Zepeda O, et al. Cell death by pyroptosis drives CD4 T-cell depletion in HIV-1 infection. Nature. 2014;505(7484):509-14. doi: 10.1038/nature12940. PubMed PMID: 24356306; PubMed Central PMCID: PMCPMC4047036.

13. Mehandru S, Poles MA, Tenner-Racz K, Horowitz A, Hurley A, Hogan C, et al. Primary HIV-1 infection is associated with preferential depletion of CD4+ T lymphocytes from effector sites in the gastrointestinal tract. J Exp Med. 2004;200(6):761-70. doi: 10.1084/jem.20041196. PubMed PMID: 15365095; PubMed Central PMCID: PMCPMC2211967.

14. Monroe KM, Yang Z, Johnson JR, Geng X, Doitsh G, Krogan NJ, et al. IFI16 DNA sensor is required for death of lymphoid CD4 T cells abortively infected with HIV. Science. 2014;343(6169):428-32. doi: 10.1126/science.1243640. PubMed PMID: 24356113; PubMed Central PMCID: PMCPMC3976200.

** In this article a new mechanisms of cell death linked to the increase of reverse transcripts in the cytosol of infected lymphocytes due to HIV-1 restriction by SAMHD1 is proposed

15. Cummins NW, Sainski AM, Dai H, Natesampillai S, Pang YP, Bren GD, et al. Prime, Shock, and Kill: Priming CD4 T Cells from HIV Patients with a BCL-2 Antagonist before HIV Reactivation Reduces HIV Reservoir Size. J Virol. 2016;90(8):4032-48. doi: 10.1128/JVI.03179-15. PubMed PMID: 26842479; PubMed Central PMCID: PMCPMC4810548.

16. Lopez-Huertas MR, Mateos E, Sanchez Del Cojo M, Gomez-Esquer F, Diaz-Gil G, Rodriguez-Mora S, et al. The presence of HIV-1 Tat protein second exon delays fas proteinmediated apoptosis in CD4+ T lymphocytes: a potential mechanism for persistent viral production. J Biol Chem. 2013;288(11):7626-44. doi: 10.1074/jbc.M112.408294. PubMed PMID: 23364796; PubMed Central PMCID: PMCPMC3597804.

17. Pantaleo G, Demarest JF, Schacker T, Vaccarezza M, Cohen OJ, Daucher M, et al. The qualitative nature of the primary immune response to HIV infection is a prognosticator of disease progression independent of the initial level of plasma viremia. Proc Natl Acad Sci U S A. 1997;94(1):254-8. PubMed PMID: 8990195; PubMed Central PMCID: PMCPMC19306.

18. Rosenberg ES, Billingsley JM, Caliendo AM, Boswell SL, Sax PE, Kalams SA, et al. Vigorous HIV-1-specific CD4+ T cell responses associated with control of viremia. Science. 1997;278(5342):1447-50. PubMed PMID: 9367954.

19. Plana M, Garcia F, Gallart T, Miro JM, Gatell JM. Lack of T-cell proliferative response to HIV-1 antigens after 1 year of highly active antiretroviral treatment in early HIV-1 disease. Immunology Study Group of Spanish EARTH-1 Study. Lancet. 1998;352(9135):1194-5. PubMed PMID: 9777842.

20. Boritz EA, Darko S, Swaszek L, Wolf G, Wells D, Wu X, et al. Multiple Origins of Virus Persistence during Natural Control of HIV Infection. Cell. 2016;166(4):1004-15. doi: 10.1016/j.cell.2016.06.039. PubMed PMID: 27453467; PubMed Central PMCID: PMCPMC4983216.

* A new actively replicating HIV-1 reservoir -CD4 T Helper follicular cells- that are placed in a CTL-resistant sanctuary is described

21. Douek DC, Brenchley JM, Betts MR, Ambrozak DR, Hill BJ, Okamoto Y, et al. HIV preferentially infects HIV-specific CD4+ T cells. Nature. 2002;417(6884):95-8. doi: 10.1038/417095a. PubMed PMID: 11986671.

* In this article the paradoxical destruction of CD4 specific lymphocytes targeting the HIV-1 virus due to enhanced susceptibility to infection is described as a qualitative mechanism of immune suppression

22. Lore K, Smed-Sorensen A, Vasudevan J, Mascola JR, Koup RA. Myeloid and plasmacytoid dendritic cells transfer HIV-1 preferentially to antigen-specific CD4+ T cells. J Exp Med. 2005;201(12):2023-33. doi: 10.1084/jem.20042413. PubMed PMID: 15967828; PubMed Central PMCID: PMCPMC2212038.

23. Lorenzo-Redondo R, Fryer HR, Bedford T, Kim EY, Archer J, Kosakovsky Pond SL, et al. Persistent HIV-1 replication maintains the tissue reservoir during therapy. Nature. 2016;530(7588):51-6. doi: 10.1038/nature16933. PubMed PMID: 26814962; PubMed Central PMCID: PMCPMC4865637.

24. Rizzardi GP, Harari A, Capiluppi B, Tambussi G, Ellefsen K, Ciuffreda D, et al. Treatment of primary HIV-1 infection with cyclosporin A coupled with highly active antiretroviral therapy. J Clin Invest. 2002;109(5):681-8. doi: 10.1172/JCI14522. PubMed PMID: 11877476; PubMed Central PMCID: PMCPMC150896.

* First paper describing the role of immune suppression to preserve CD4 lymphocyte counts in acutely infected HIV patients. Unfortunately this article was not confirmed by other authors

25. Calabrese LH, Lederman MM, Spritzler J, Coombs RW, Fox L, Schock B, et al. Placebocontrolled trial of cyclosporin-A in HIV-1 disease: implications for solid organ transplantation. J Acquir Immune Defic Syndr. 2002;29(4):356-62. PubMed PMID: 11917239.

26. Markowitz M, Vaida F, Hare CB, Boden D, Mohri H, Hecht FM, et al. The virologic and immunologic effects of cyclosporine as an adjunct to antiretroviral therapy in patients treated during acute and early HIV-1 infection. J Infect Dis. 2010;201(9):1298-302. doi:

10.1086/651664. PubMed PMID: 20235838; PubMed Central PMCID: PMCPMC2851487.
27. Andrieu JM, Even P, Venet A, Tourani JM, Stern M, Lowenstein W, et al. Effects of cyclosporin on T-cell subsets in human immunodeficiency virus disease. Clin Immunol Immunopathol. 1988;47(2):181-98. PubMed PMID: 3258211.

28. Rizzardi GP, Vaccarezza M, Capiluppi B, Tambussi G, Lazzarin A, Pantaleo G. Cyclosporin A in combination with HAART in primary HIV-1 infection. J Biol Regul Homeost Agents. 2000;14(1):79-81. PubMed PMID: 10763900.

29. Nicolas D, Ambrosioni J, Sued O, Brunet M, Lopez-Dieguez M, Manzardo C, et al. Cyclosporine A in addition to standard ART during primary HIV-1 infection: pilot randomized clinical trial. J Antimicrob Chemother. 2016. doi: 10.1093/jac/dkw462. PubMed PMID: 27999018.

30. Phillips A, Wainberg MA, Coates R, Klein M, Rachlis A, Read S, et al. Cyclosporineinduced deterioration in patients with AIDS. CMAJ. 1989;140(12):1456-60. PubMed PMID: 2720530; PubMed Central PMCID: PMCPMC1269983.

31. Chapuis AG, Paolo Rizzardi G, D'Agostino C, Attinger A, Knabenhans C, Fleury S, et al. Effects of mycophenolic acid on human immunodeficiency virus infection in vitro and in vivo. Nat Med. 2000;6(7). Epub 768. PubMed Central PMCID: PMC10888924.

32. Coull JJ, Turner D, Melby T, Betts MR, Lanier R, Margolis DM. A pilot study of the use of mycophenolate mofetil as a component of therapy for multidrug-resistant HIV-1 infection. J Acquir Immune Defic Syndr. 2001;26(5):423-34. PubMed PMID: 11391161.

33. Sankatsing SU, Jurriaans S, van Swieten P, van Leth F, Cornelissen M, Miedema F, et al. Highly active antiretroviral therapy with or without mycophenolate mofetil in treatment-naive HIV-1 patients. AIDS. 2004;18(14):1925-31. PubMed PMID: 15353978.

34. Garcia F, Plana M, Arnedo M, Brunet M, Castro P, Gil C, et al. Effect of mycophenolate mofetil on immune response and plasma and lymphatic tissue viral load during and after interruption of highly active antiretroviral therapy for patients with chronic HIV infection: a randomized pilot study. J Acquir Immune Defic Syndr. 2004;36(3):823-30. PubMed PMID: 15213566.

35. Fumero E, Garcia F, Gatell JM. Immunosuppressive drugs as an adjuvant to HIV treatment. J Antimicrob Chemother. 2004;53(3):415-7. doi: 10.1093/jac/dkh123. PubMed PMID: 14963070.

* A review on the potential benefit of immunomodulatory drugs in HIV infection

36. Garcia F, Plana M, Arnedo M, Ortiz GM, Miro JM, Lopalco L, et al. A cytostatic drug improves control of HIV-1 replication during structured treatment interruptions: a randomized study. AIDS. 2003;17(1):43-51. doi: 10.1097/01.aids.0000042953.95433.79. PubMed PMID: 12478068.

37. Kong KF, Yokosuka T, Canonigo-Balancio AJ, Isakov N, Saito T, Altman A. A motif in the V3 domain of the kinase PKC-theta determines its localization in the immunological synapse and functions in T cells via association with CD28. Nat Immunol. 2011;12(11):1105-12. doi: 10.1038/ni.2120. PubMed PMID: 21964608; PubMed Central PMCID: PMCPMC3197934.

38. Gruber T, Pfeifhofer-Obermair C, Baier G. PKCtheta is necessary for efficient activation of NFkappaB, NFAT, and AP-1 during positive selection of thymocytes. Immunol Lett. 2010;132(1-2):6-11. doi: 10.1016/j.imlet.2010.04.008. PubMed PMID: 20433868; PubMed Central PMCID: PMCPMC2937209.

39. Wang X, Chuang HC, Li JP, Tan TH. Regulation of PKC-theta function by phosphorylation in T cell receptor signaling. Front Immunol. 2012;3:197. doi: 10.3389/fimmu.2012.00197. PubMed PMID: 22798961; PubMed Central PMCID: PMCPMC3393885.

40. Thomas SM, Brugge JS. Cellular functions regulated by Src family kinases. Annu Rev Cell Dev Biol. 1997;13:513-609. doi: 10.1146/annurev.cellbio.13.1.513. PubMed PMID: 9442882.

41. Cooke MP, Perlmutter RM. Expression of a novel form of the fyn proto-oncogene in hematopoietic cells. New Biol. 1989;1(1):66-74. PubMed PMID: 2488273.

42. Olszowy MW, Leuchtmann PL, Veillette A, Shaw AS. Comparison of p56lck and p59fyn protein expression in thymocyte subsets, peripheral T cells, NK cells, and lymphoid cell lines. J Immunol. 1995;155(9):4236-40. PubMed PMID: 7594580.

43. Palacios EH, Weiss A. Function of the Src-family kinases, Lck and Fyn, in T-cell development and activation. Oncogene. 2004;23(48):7990-8000. doi: 10.1038/sj.onc.1208074. PubMed PMID: 15489916.

* An interesting revision on the role of Tyrosin kinases in lymphocyte activation

44. Turner JM, Brodsky MH, Irving BA, Levin SD, Perlmutter RM, Littman DR. Interaction of the unique N-terminal region of tyrosine kinase p56lck with cytoplasmic domains of CD4 and CD8 is mediated by cysteine motifs. Cell. 1990;60(5):755-65. PubMed PMID: 2107025.

45. Ley SC, Marsh M, Bebbington CR, Proudfoot K, Jordan P. Distinct intracellular localization of Lck and Fyn protein tyrosine kinases in human T lymphocytes. J Cell Biol. 1994;125(3):639-49. PubMed PMID: 7513706; PubMed Central PMCID: PMCPMC2119993.

46. Lin K, Longo NS, Wang X, Hewitt JA, Abraham KM. Lck domains differentially contribute to pre-T cell receptor (TCR)- and TCR-alpha/beta-regulated developmental transitions. J Exp Med. 2000;191(4):703-16. PubMed PMID: 10684862; PubMed Central PMCID: PMCPMC2195836.

47. Coiras M, Bermejo M, Descours B, Mateos E, Garcia-Perez J, Lopez-Huertas MR, et al. IL-7 Induces SAMHD1 Phosphorylation in CD4+ T Lymphocytes, Improving Early Steps of HIV-1 Life Cycle. Cell Rep. 2016;14(9):2100-7. doi: 10.1016/j.celrep.2016.02.022. PubMed PMID: 26923586; PubMed Central PMCID: PMCPMC5063304.

** First description of the ability of dasatinib to interfere with SAMHD1

48. Simeoni L, Smida M, Posevitz V, Schraven B, Lindquist JA. Right time, right place: the organization of membrane proximal signaling. Semin Immunol. 2005;17(1):35-49. doi: 10.1016/j.smim.2004.09.002. PubMed PMID: 15582487.

49. Ghosh S, May MJ, Kopp EB. NF-kappa B and Rel proteins: evolutionarily conserved mediators of immune responses. Annu Rev Immunol. 1998;16:225-60. doi: 10.1146/annurev.immunol.16.1.225. PubMed PMID: 9597130.

50. Hiscott J, Kwon H, Genin P. Hostile takeovers: viral appropriation of the NF-kappaB pathway. J Clin Invest. 2001;107(2):143-51. doi: 10.1172/JCI11918. PubMed PMID: 11160127; PubMed Central PMCID: PMCPMC199181.

51. Kwon H, Pelletier N, DeLuca C, Genin P, Cisternas S, Lin R, et al. Inducible expression of IkappaBalpha repressor mutants interferes with NF-kappaB activity and HIV-1 replication in Jurkat T cells. J Biol Chem. 1998;273(13):7431-40. PubMed PMID: 9516441.

52. Quinto I, Mallardo M, Baldassarre F, Scala G, Englund G, Jeang KT. Potent and stable attenuation of live-HIV-1 by gain of a proteolysis-resistant inhibitor of NF-kappaB (IkappaB-alphaS32/36A) and the implications for vaccine development. J Biol Chem.

1999;274(25):17567-72. PubMed PMID: 10364191.

53. Bermejo M, Lopez-Huertas MR, Hedgpeth J, Mateos E, Rodriguez-Mora S, Maleno MJ, et al. Analysis of protein kinase C theta inhibitors for the control of HIV-1 replication in human CD4+ T cells reveals an effect on retrotranscription in addition to viral transcription. Biochem Pharmacol. 2015;94(4):241-56. doi: 10.1016/j.bcp.2015.02.009. PubMed PMID: 25732195.

54. Bermejo M, Lopez-Huertas MR, Garcia-Perez J, Climent N, Descours B, Ambrosioni J, et al. Dasatinib inhibits HIV-1 replication through the interference of SAMHD1 phosphorylation in

CD4+ T cells. Biochem Pharmacol. 2016;106:30-45. doi: 10.1016/j.bcp.2016.02.002. PubMed PMID: 26851491.

* First description of the role of tyrosine kinase inhibitors in SAMHD1 regulation
55. D'Antonio J. Chronic myelogenous leukemia. Clin J Oncol Nurs. 2005;9(5):535-8. doi:
10.1188/05.CJON.535-538. PubMed PMID: 16235580.

56. Sabir N, Iqbal Z, Aleem A, Awan T, Naeem T, Asad S, et al. Prognostically significant fusion oncogenes in Pakistani patients with adult acute lymphoblastic leukemia and their association with disease biology and outcome. Asian Pac J Cancer Prev. 2012;13(7):3349-55. PubMed PMID: 22994759.

57. Andreieva SV, Korets KV, Kyselova OA, Ruzhinska OE, Serbin IM. Chronic myeloid leukemia in patient with the Klinefelter syndrome. Exp Oncol. 2016;38(3):195-7. PubMed PMID: 27685529.

58. Sawyers CL. Chronic myeloid leukemia. N Engl J Med. 1999;340(17):1330-40. doi: 10.1056/NEJM199904293401706. PubMed PMID: 10219069.

59. Quintas-Cardama A, Kantarjian H, Cortes J. Imatinib and beyond--exploring the full potential of targeted therapy for CML. Nat Rev Clin Oncol. 2009;6(9):535-43. doi: 10.1038/nrclinonc.2009.112. PubMed PMID: 19652654.

60. Woessner DW, Lim CS, Deininger MW. Development of an effective therapy for chronic myelogenous leukemia. Cancer J. 2011;17(6):477-86. doi: 10.1097/PPO.0b013e318237e5b7. PubMed PMID: 22157291; PubMed Central PMCID: PMCPMC3251313.

61. Steegmann JL, Cervantes F, le Coutre P, Porkka K, Saglio G. Off-target effects of BCR-ABL1 inhibitors and their potential long-term implications in patients with chronic myeloid leukemia. Leuk Lymphoma. 2012;53(12):2351-61. doi: 10.3109/10428194.2012.695779. PubMed PMID: 22616642.

62. Thompson PA, Kantarjian HM, Cortes JE. Diagnosis and Treatment of Chronic Myeloid Leukemia in 2015. Mayo Clin Proc. 2015;90(10):1440-54. doi: 10.1016/j.mayocp.2015.08.010. PubMed PMID: 26434969.

* An interesting revision on the treatment of chronic myeloid leukemia, describing the different tyrosine kinase inhibitors

63. Steegmann JL, Baccarani M, Breccia M, Casado LF, Garcia-Gutierrez V, Hochhaus A, et al. European LeukemiaNet recommendations for the management and avoidance of adverse events of treatment in chronic myeloid leukaemia. Leukemia. 2016;30(8):1648-71. doi:

10.1038/leu.2016.104. PubMed PMID: 27121688; PubMed Central PMCID: PMCPMC4991363. 64. O'Hare T, Walters DK, Stoffregen EP, Jia T, Manley PW, Mestan J, et al. In vitro activity of Bcr-Abl inhibitors AMN107 and BMS-354825 against clinically relevant imatinib-resistant Abl kinase domain mutants. Cancer Res. 2005;65(11):4500-5. doi: 10.1158/0008-5472.CAN-05-0259. PubMed PMID: 15930265.

65. Puttini M, Coluccia AM, Boschelli F, Cleris L, Marchesi E, Donella-Deana A, et al. In vitro and in vivo activity of SKI-606, a novel Src-Abl inhibitor, against imatinib-resistant Bcr-Abl+ neoplastic cells. Cancer Res. 2006;66(23):11314-22. doi: 10.1158/0008-5472.CAN-06-1199. PubMed PMID: 17114238.

66. Simoneau CA. Treating chronic myeloid leukemia: improving management through understanding of the patient experience. Clin J Oncol Nurs. 2013;17(1):E13-20. doi: 10.1188/13.CJON.E13-E20. PubMed PMID: 23372106.

67. Moslehi JJ, Deininger M. Tyrosine Kinase Inhibitor-Associated Cardiovascular Toxicity in Chronic Myeloid Leukemia. J Clin Oncol. 2015;33(35):4210-8. doi: 10.1200/JCO.2015.62.4718. PubMed PMID: 26371140; PubMed Central PMCID: PMCPMC4658454.

68. Brixey AG, Light RW. Pleural effusions due to dasatinib. Curr Opin Pulm Med. 2010;16(4):351-6. doi: 10.1097/MCP.0b013e328338c486. PubMed PMID: 20375898.

69. (EMA) EMA. Sprycel[®] product information.

70. Cortes JE, Kantarjian HM, Brummendorf TH, Kim DW, Turkina AG, Shen ZX, et al. Safety and efficacy of bosutinib (SKI-606) in chronic phase Philadelphia chromosome-positive chronic

myeloid leukemia patients with resistance or intolerance to imatinib. Blood. 2011;118(17):4567-76. doi: 10.1182/blood-2011-05-355594. PubMed PMID: 21865346; PubMed Central PMCID: PMCPMC4916618.

71. Miller GD, Bruno BJ, Lim CS. Resistant mutations in CML and Ph(+)ALL - role of ponatinib. Biologics. 2014;8:243-54. doi: 10.2147/BTT.S50734. PubMed PMID: 25349473; PubMed Central PMCID: PMCPMC4208348.

72. Rohon P, Porkka K, Mustjoki S. Immunoprofiling of patients with chronic myeloid leukemia at diagnosis and during tyrosine kinase inhibitor therapy. Eur J Haematol. 2010;85(5):387-98. doi: 10.1111/j.1600-0609.2010.01501.x. PubMed PMID: 20662899.

73. Cwynarski K, Laylor R, Macchiarulo E, Goldman J, Lombardi G, Melo JV, et al. Imatinib inhibits the activation and proliferation of normal T lymphocytes in vitro. Leukemia. 2004;18(8):1332-9. doi: 10.1038/sj.leu.2403401. PubMed PMID: 15190258.

74. Chen J, Schmitt A, Chen B, Rojewski M, Rubeler V, Fei F, et al. Nilotinib hampers the proliferation and function of CD8+ T lymphocytes through inhibition of T cell receptor signalling. J Cell Mol Med. 2008;12(5B):2107-18. doi: 10.1111/j.1582-4934.2008.00234.x. PubMed PMID: 18194453; PubMed Central PMCID: PMCPMC4506175.

75. Schade AE, Schieven GL, Townsend R, Jankowska AM, Susulic V, Zhang R, et al. Dasatinib, a small-molecule protein tyrosine kinase inhibitor, inhibits T-cell activation and proliferation. Blood. 2008;111(3):1366-77. doi: 10.1182/blood-2007-04-084814. PubMed PMID: 17962511; PubMed Central PMCID: PMCPMC2214733.

76. Kreutzman A, Juvonen V, Kairisto V, Ekblom M, Stenke L, Seggewiss R, et al. Mono/oligoclonal T and NK cells are common in chronic myeloid leukemia patients at diagnosis and expand during dasatinib therapy. Blood. 2010;116(5):772-82. doi: 10.1182/blood-2009-12-256800. PubMed PMID: 20413659.

* A description of immunological changes induced by tyrosine kinase inhibitors related with toxic effects

77. Mustjoki S, Auvinen K, Kreutzman A, Rousselot P, Hernesniemi S, Melo T, et al. Rapid mobilization of cytotoxic lymphocytes induced by dasatinib therapy. Leukemia.

2013;27(4):914-24. doi: 10.1038/leu.2012.348. PubMed PMID: 23192016.

78. Schiffer CA, Cortes JE, Hochhaus A, Saglio G, le Coutre P, Porkka K, et al. Lymphocytosis after treatment with dasatinib in chronic myeloid leukemia: Effects on response and toxicity. Cancer. 2016;122(9):1398-407. doi: 10.1002/cncr.29933. PubMed PMID: 26998677.

* A description of the potential role of recovered lymphocytes in the response and control of chronic myeloid leukemia

79. Rodriguez GH, Ahmed SI, Al-akhrass F, Rallapalli V, Safdar A. Characteristics of, and risk factors for, infections in patients with cancer treated with dasatinib and a brief review of other complications. Leuk Lymphoma. 2012;53(8):1530-5. doi: 10.3109/10428194.2012.656626. PubMed PMID: 22263567.

80. Benhar M, Shytaj IL, Stamler JS, Savarino A. Dual targeting of the thioredoxin and glutathione systems in cancer and HIV. J Clin Invest. 2016;126(5):1630-9. doi:

10.1172/JCI85339. PubMed PMID: 27135880; PubMed Central PMCID: PMCPMC4855928.
81. Zitvogel L, Apetoh L, Ghiringhelli F, Andre F, Tesniere A, Kroemer G. The anticancer immune response: indispensable for therapeutic success? J Clin Invest. 2008;118(6):1991-2001. doi: 10.1172/JCI35180. PubMed PMID: 18523649; PubMed Central PMCID: PMCPMC2396905.

82. Lopez-Huertas MR, Mateos E, Diaz-Gil G, Gomez-Esquer F, Sanchez del Cojo M, Alcami J, et al. Protein kinase Ctheta is a specific target for inhibition of the HIV type 1 replication in CD4+ T lymphocytes. J Biol Chem. 2011;286(31):27363-77. doi: 10.1074/jbc.M110.210443. PubMed PMID: 21669868; PubMed Central PMCID: PMCPMC3149330.

83. Lahouassa H, Daddacha W, Hofmann H, Ayinde D, Logue EC, Dragin L, et al. SAMHD1 restricts the replication of human immunodeficiency virus type 1 by depleting the intracellular

pool of deoxynucleoside triphosphates. Nat Immunol. 2012;13(3):223-8. doi: 10.1038/ni.2236. PubMed PMID: 22327569; PubMed Central PMCID: PMCPMC3771401.

84. Cribier A, Descours B, Valadao AL, Laguette N, Benkirane M. Phosphorylation of SAMHD1 by cyclin A2/CDK1 regulates its restriction activity toward HIV-1. Cell Rep. 2013;3(4):1036-43. doi: 10.1016/j.celrep.2013.03.017. PubMed PMID: 23602554.

85. Giansanti P, Preisinger C, Huber KV, Gridling M, Superti-Furga G, Bennett KL, et al. Evaluating the promiscuous nature of tyrosine kinase inhibitors assessed in A431 epidermoid carcinoma cells by both chemical- and phosphoproteomics. ACS Chem Biol. 2014;9(7):1490-8. doi: 10.1021/cb500116c. PubMed PMID: 24804581.

86. Bullen CK, Laird GM, Durand CM, Siliciano JD, Siliciano RF. New ex vivo approaches distinguish effective and ineffective single agents for reversing HIV-1 latency in vivo. Nat Med. 2014;20(4):425-9. doi: 10.1038/nm.3489. PubMed PMID: 24658076; PubMed Central PMCID: PMCPMC3981911.

87. Deeks SG. HIV infection, inflammation, immunosenescence, and aging. Annu Rev Med. 2011;62:141-55. doi: 10.1146/annurev-med-042909-093756. PubMed PMID: 21090961; PubMed Central PMCID: PMCPMC3759035.

**A review on the pathogenic mechanisms leading to immunological damage due to chronic activation and exhaustion of the immune system by persistent HIV replication

88. Aberg JA. Aging, inflammation, and HIV infection. Top Antivir Med. 2012;20(3):101-5. PubMed PMID: 22954610.

89. Saussele S, Richter J, Hochhaus A, Mahon FX. The concept of treatment-free remission in chronic myeloid leukemia. Leukemia. 2016;30(8):1638-47. doi: 10.1038/leu.2016.115. PubMed PMID: 27133824; PubMed Central PMCID: PMCPMC4980559.

* A concept paper discussing the successful interruption of treatment with tyrosine kinase inhibitors in patients with chronic myeloid leukemia that show deep molecular response
 90. Imagawa J, Tanaka H, Okada M, Nakamae H, Hino M, Murai K, et al. Discontinuation of

dasatinib in patients with chronic myeloid leukaemia who have maintained deep molecular response for longer than 1 year (DADI trial): a multicentre phase 2 trial. Lancet Haematol. 2015;2(12):e528-35. doi: 10.1016/S2352-3026(15)00196-9. PubMed PMID: 26686407.

91. Mizoguchi I, Yoshimoto T, Katagiri S, Mizuguchi J, Tauchi T, Kimura Y, et al. Sustained upregulation of effector natural killer cells in chronic myeloid leukemia after discontinuation of imatinib. Cancer Sci. 2013;104(9):1146-53. doi: 10.1111/cas.12216. PubMed PMID: 23758044.

92. Zitvogel L, Rusakiewicz S, Routy B, Ayyoub M, Kroemer G. Immunological off-target effects of imatinib. Nat Rev Clin Oncol. 2016;13(7):431-46. doi: 10.1038/nrclinonc.2016.41. PubMed PMID: 27030078.

93. Cao W, Vyboh K, Routy B, Chababi-Atallah M, Lemire B, Routy JP. Imatinib for highly chemoresistant Kaposi sarcoma in a patient with long-term HIV control: a case report and literature review. Curr Oncol. 2015;22(5):e395-9. doi: 10.3747/co.22.2635. PubMed PMID: 26628884; PubMed Central PMCID: PMCPMC4608417.

94. Ananworanich J, Schuetz A, Vandergeeten C, Sereti I, de Souza M, Rerknimitr R, et al. Impact of multi-targeted antiretroviral treatment on gut T cell depletion and HIV reservoir seeding during acute HIV infection. PLoS One. 2012;7(3):e33948. doi:

10.1371/journal.pone.0033948. PubMed PMID: 22479485; PubMed Central PMCID: PMCPMC3316511.

95. Buzon MJ, Martin-Gayo E, Pereyra F, Ouyang Z, Sun H, Li JZ, et al. Long-term antiretroviral treatment initiated at primary HIV-1 infection affects the size, composition, and decay kinetics of the reservoir of HIV-1-infected CD4 T cells. J Virol. 2014;88(17):10056-65. doi: 10.1128/JVI.01046-14. PubMed PMID: 24965451; PubMed Central PMCID: PMCPMC4136362. * This article describes the impact of early treatment on the establishment and size of the HIV reservoir

96. Fiebig EW, Wright DJ, Rawal BD, Garrett PE, Schumacher RT, Peddada L, et al. Dynamics of HIV viremia and antibody seroconversion in plasma donors: implications for diagnosis and staging of primary HIV infection. AIDS. 2003;17(13):1871-9. doi: 10.1097/01.aids.0000076308.76477.b8. PubMed PMID: 12960819.

** Classical article describing the criteria to define the different stages in acute HIV-1 infection 97. Ndhlovu ZM, Kamya P, Mewalal N, Kloverpris HN, Nkosi T, Pretorius K, et al. Magnitude and Kinetics of CD8+ T Cell Activation during Hyperacute HIV Infection Impact Viral Set Point. Immunity. 2015;43(3):591-604. doi: 10.1016/j.immuni.2015.08.012. PubMed PMID: 26362266; PubMed Central PMCID: PMCPMC4575777.

98. Kloverpris HN, Kazer SW, Mjosberg J, Mabuka JM, Wellmann A, Ndhlovu Z, et al. Innate Lymphoid Cells Are Depleted Irreversibly during Acute HIV-1 Infection in the Absence of Viral Suppression. Immunity. 2016;44(2):391-405. doi: 10.1016/j.immuni.2016.01.006. PubMed PMID: 26850658.

99. Deng K, Pertea M, Rongvaux A, Wang L, Durand CM, Ghiaur G, et al. Broad CTL response is required to clear latent HIV-1 due to dominance of escape mutations. Nature. 2015;517(7534):381-5. doi: 10.1038/nature14053. PubMed PMID: 25561180; PubMed Central PMCID: PMCPMC4406054.

100. Shytaj IL, Nickel G, Arts E, Farrell N, Biffoni M, Pal R, et al. Two-Year Follow-Up of Macaques Developing Intermittent Control of the Human Immunodeficiency Virus Homolog Simian Immunodeficiency Virus SIVmac251 in the Chronic Phase of Infection. J Virol. 2015;89(15):7521-35. doi: 10.1128/JVI.00396-15. PubMed PMID: 25972547; PubMed Central PMCID: PMCPMC4505651.

101. Harmon B, Campbell N, Ratner L. Role of Abl kinase and the Wave2 signaling complex in HIV-1 entry at a post-hemifusion step. PLoS Pathog. 2010;6(6):e1000956. doi: 10.1371/journal.ppat.1000956. PubMed PMID: 20585556; PubMed Central PMCID: PMCPMC2887473.

102. Pogliaghi M, Papagno L, Lambert S, Calin R, Calvez V, Katlama C, et al. The tyrosine kinase inhibitor Dasatinib blocks in-vitro HIV-1 production by primary CD4+ T cells from HIV-1 infected patients. AIDS. 2014;28(2):278-81. doi: 10.1097/QAD.0000000000000073. PubMed PMID: 24361684.

103. Dhurve SA, Dhurve AS. Bone Marrow Abnormalities in HIV Disease. Mediterr J Hematol Infect Dis. 2013;5(1):e2013033. doi: 10.4084/MJHID.2013.033. PubMed PMID: 23795271; PubMed Central PMCID: PMCPMC3684351.

104. Ambrosioni J, Nicolas D, Sued O, Aguero F, Manzardo C, Miro JM. Update on antiretroviral treatment during primary HIV infection. Expert Rev Anti Infect Ther. 2014;12(7):793-807. doi: 10.1586/14787210.2014.913981. PubMed PMID: 24803105.

105. Shiels MS, Engels EA. Evolving epidemiology of HIV-associated malignancies. Curr Opin HIV AIDS. 2016. doi: 10.1097/COH.000000000000027. PubMed PMID: 27749369.

106. Ando T, Kojima K, Isoda H, Eguchi Y, Honda T, Ishigami M, et al. Reactivation of resolved infection with the hepatitis B virus immune escape mutant G145R during dasatinib treatment for chronic myeloid leukemia. Int J Hematol. 2015;102(3):379-82. doi: 10.1007/s12185-015-1788-y. PubMed PMID: 25842192.

107. Soriano V, Labarga P, de Mendoza C, Pena JM, Fernandez-Montero JV, Benitez L, et al. Emerging challenges in managing hepatitis B in HIV patients. Curr HIV/AIDS Rep. 2015;12(3):344-52. doi: 10.1007/s11904-015-0275-7. PubMed PMID: 26156570.

108. Nguyen T, McNicholl I, Custodio JM, Szwarcberg J, Piontkowsky D. Drug Interactions with Cobicistat- or Ritonavir-Boosted Elvitegravir. AIDS Rev. 2016;18(2):101-11. PubMed PMID: 27196356.

Johnson FM, Agrawal S, Burris H, Rosen L, Dhillon N, Hong D, et al. Phase 1 pharmacokinetic and drug-interaction study of dasatinib in patients with advanced solid tumors. Cancer. 2010;116(6):1582-91. doi: 10.1002/cncr.24927. PubMed PMID: 20108303.
Abbas R, Hsyu PH. Clinical Pharmacokinetics and Pharmacodynamics of Bosutinib. Clin Pharmacokinet. 2016;55(10):1191-204. doi: 10.1007/s40262-016-0391-6. PubMed PMID: 27113346.

111. (EMA) EMA. Iclusig[®] Prescribing Information. Available from:

http://www.ema.europa.eu/docs/es_ES/document_library/EPAR_Product_Information/hum an/002695/WC500145646.pdf

112. (EACS) EACS. EACS 2015 Guidelines.

113. Gunthard HF, Saag MS, Benson CA, del Rio C, Eron JJ, Gallant JE, et al. Antiretroviral Drugs for Treatment and Prevention of HIV Infection in Adults: 2016 Recommendations of the International Antiviral Society-USA Panel. JAMA. 2016;316(2):191-210. doi:

10.1001/jama.2016.8900. PubMed PMID: 27404187; PubMed Central PMCID: PMCPMC5012643.

114. Velu V, Titanji K, Zhu B, Husain S, Pladevega A, Lai L, et al. Enhancing SIV-specific immunity in vivo by PD-1 blockade. Nature. 2009;458(7235):206-10. doi:

10.1038/nature07662. PubMed PMID: 19078956; PubMed Central PMCID: PMCPMC2753387.