1 Selfish genetic elements and male fertility

- 2 Rudi L. Verspoor¹, Tom A. R. Price¹, and Nina Wedell²
- 3 ¹ Institution for Integrative Biology, University of Liverpool, Liverpool L69 7ZB, UK
- 4 ² Biosciences, University of Exeter, Penryn Campus, Penryn TR10 9FE, UK

5

6 Selfish Genetic Elements (SGEs) are diverse and near ubiquitous in Eukaryotes and can be potent 7 drivers of evolution. Here we discuss SGEs that specifically act on sperm to gain a transmission 8 advantage to the next generation. The diverse SGEs that affect sperm often impose costs on carrier 9 males, including damaging ejaculates, skewing offspring sex-ratios and in particular reducing sperm 10 competitive success of SGE carrying males. How males and females tolerate and mitigate against these 11 costs is a dynamic and expanding area of research. The intense intra-genomic conflict that these selfish 12 elements generate could also have implications for male fertility and spermatogenesis more widely.

13

15 1. Introduction

16 For an allele securing a place in the next generation is critical. Achieving this success has traditionally 17 been explained by the forces of natural and sexual selection. However, a third route to evolutionary 18 success has been revealed, challenging the premise of 'fair' Mendelian inheritance. Here, Selfish 19 Genetic Elements (SGEs) can increase their own frequency across generations without increasing the 20 fitness of their carrier individuals, and often impose major costs on the rest of the genome (1). Several 21 SGEs specifically act through sperm, from paternal genome eliminators to endosymbionts to toxic 22 sperm killers (1-3). As a result, spermatogenesis through to fertilisation can be viewed as a series of 23 arenas that are vulnerable to the activities of SGEs. The actions of SGEs in sperm have far reaching 24 impacts, killing sperm and zygotes, changing the physiology and mating behaviour of males and 25 females, and perhaps influenced the evolution of some of the deep structures of spermatogenesis 26 (4,5,6; Figure 1).

27 2. SGEs and Sperm

28 SGEs affecting sperm were reported almost a century ago (7) and new SGEs are still being discovered

(8,9). Here we explore three types of SGEs that affect sperm; those that kill or damage sperm, those
that travel within sperm, and those that modify sperm to affect zygote formation.

31 Segregation distorters: sperm killers and disablers

32 Segregation distorters, sometimes referred to as killer meiotic drivers, are one of the best studied 33 SGEs that manipulate sperm (3,10). As sperm are haploid, carrying only one allele from their diploid 34 parent genome, one selfish haploid allele can gain a transmission advantage by sabotaging their 35 opposite haploid allele during spermatogenesis. This directly benefits the selfish haploid sperm allele, 36 as it then occurs in more than half of a male's sperm, despite its action generally being destructive to 37 the ejaculate (6). Segregation distorters can kill all non-carrier sperm, transmitting the driver to 100% 38 of offspring. However, this killing can reduce sperm number by up to 50%, and can even damage sperm 39 that carry the segregation distorter (3). This reduction can directly reduce male fertility (4) and these 40 costs can be exacerbated by contexts such as high polyandry, due to poor sperm-competitive ability 41 (Table 1), or high temperatures (11). When the driver occurs on a sex-chromosome they also strongly 42 bias offspring sex-ratios (10). This death of half a male's sperm, and potentially biased brood sex ratios, 43 can impose major costs on the rest of the genome, causing strong intra-genomic conflict and

44 promoting suppression of the SGE (12).

45 While our mechanistic understanding of how these SGEs kill sperm comes from only a few model 46 systems, we know they act at different stages of spermatogenesis (13,14). There are also differences 47 in the effect on non-carrier sperm, with some SGEs disabling sperm, for example the t-haplotype 48 system in *Mus musculus* (15), while many others kill non-carrier sperm, for example the Paris sex-ratio 49 drive system in *Drosophila simulans* (16). When and how non-carrier sperm are affected will likely 50 have differing impacts on sperm quality and male fertility costs (Figure 1; Table 1). In general, the 51 characterized drive systems share some commonalities in mechanism, frequently involving 52 heterochromatin binding and small RNA pathways (13).

53 Sperm hitchhikers

54 Other SGEs appear to use sperm as a vehicle to hitchhike to the next generation. For example, viruses

- 55 which have been found packaged within or on sperm can be paternally inherited (9,17). In *Diaphorina* 56 *citri* psyllid insects, a retrovirus makes use of a virus-encoded non-structural protein for efficient
- 57 vertical transmission (18) and remarkably, viruses can transmit through sperm without apparent costs,

like the rice gall dwarf arbovirus that interacts with proteoglycans on sperm heads (9). Supernumerary 58 59 chromosomes, commonly known as B-chromosomes, can also transmit themselves through sperm 60 (2,19), although paternal transmission is by no means universal with some B-chromosomes being 61 excluded from sperm during spermatogenesis and instead showing biased transmission through the 62 female germline (20). The Paternal Sex-Ratio (PSR) B-chromosome, which occurs in haplodiploid 63 parasitic wasps, is a remarkable example (2). PSR travels within sperm and upon fertilization eliminates the paternal genome component in the zygote. This turns the offspring male, which means PSR always 64 65 finds itself in the sex it uses for transmission. Given how recently many SGEs that travel within sperm 66 have been characterized (particularly viruses), understanding their diversity, mechanisms and impacts 67 on sperm is an emerging area of research.

68 Post-segregation distorters

69 Some SGEs modify sperm to cause serious downstream consequences during zygote formation (Figure 70 1), and so are often referred to as post-segregation distorters. These SGEs include maternally inherited 71 endosymbionts (e.g. Wolbachia, Rickettsia, Cardinium bacteria), that are transmitted in the cytoplasm 72 of eggs from mother to offspring. Many SGEs from these groups gain a transmission advantage by 73 killing males, or turning genetic males into females, which favours the transmitting sex and hence the 74 SGEs (21,22). However, many endosymbionts modify sperm into weapons that poison the eggs they 75 fertilise if the egg lacks the same endosymbiont. The resulting reproductive incompatibility 76 (cytoplasmic incompatibility, CI) can dramatically reduce offspring production of uninfected females 77 compared to infected females, allowing the endosymbiont to spread. These 'toxin' and 'rescue-factor' 78 systems favour the offspring production of SGE-carrying females that translates into a large 79 transmission advantage favouring the spread of the selfish endosymbiont through a population (23; 80 Figure 1). This sperm manipulation can also negatively impact male reproductive success by damaging 81 the weaponised sperm, for example *Wolbachia* may cause reduced fertility in infected males by 82 affecting expression of immune genes that result in oxidative damage and cell death in the males' 83 testes (24). However, the impact of Wolbachia on male fertility can vary in both magnitude and 84 direction (4,25–29) and can be context dependent, for example frequently reducing sperm 85 competitive ability (Table 1). Such sperm-modifying endosymbionts have been shown to occur in 86 numerous arthropods including spiders, mites and filarial nematodes, and have been particularly well 87 characterised in insects where they are predicted to be present in ~65% of all species (30).

88 Sperm competitive ability

There is strong evidence that sperm killing meiotic drive substantially reduces sperm competitive ability in insects and mice, and single studies find similar effects of endosymbionts and B chromosomes (Table 1). At present the vast array of other SGEs (1) have not yet been evaluated for their impact on sperm competitive success. Given how ubiquitous SGEs are in animals, it is likely that SGEs are affecting fertility, mating behaviour, and co-evolution in a far broader range of taxa. In particular, given how easy it is to PCR screen for common endosymbionts in insects, it is surprising how few studies have investigated their effect on sperm competitive success.

96 3. Mitigation strategies

97 An important impact of many SGEs in sperm is that they impose costs to their carrier. This results in a 98 fascinating intersection between sexual selection and SGEs where males and females may adapt to

- 99 mitigate against harm from SGEs.
- 100 Mitigation by males

101 Seeing that males of several species suffer reduced ejaculate quality due to harbouring SGEs, how can 102 they maximize their fitness? In flies infected with CI-inducing Wolbachia endosymbionts, repeated 103 male mating may lessen the severity of CI, possibly due to reduced exposure time to the Wolbachia 104 toxin during sperm development (31). In support of this suggestion, increased mating rate observed 105 by Wolbachia-infected D. simulans males is shown to restore their reproductive compatibility with 106 uninfected females resulting in increased male reproductive success (25,26). It is also suggested that 107 SGE-carrying males may benefit by dispersing to a low-density population with reduced risk of sperm 108 competition, which appears to be the case in house mice where t-carrying individuals show increased 109 dispersal, especially at higher densities (32). Sperm competition models predict that disfavoured 110 males (i.e. SGE-carrying males) consistently mating in a disfavoured role (e.g. after a non-carrying 111 male, 33) should increase their ejaculate expenditure, but that this will depend on the likelihood of 112 mating in a disfavoured role (34). To date, there is insufficient data to evaluate these predictions, and 113 what we know relates to the outcome of sperm competition rather than males' ejaculate allocation 114 strategies. The predictions will also depend on the severity of sperm limitation experienced by SGE 115 carrying males and females.

116 We also expect an evolutionary response in males to compensate the cost of reduced fertility. For 117 example, in *Teleopsis dalmanni* stalk-eyed flies, males carrying a sperm killing segregation distorter 118 transfer the same number of sperm as non-carrying males (35). They are able to maintain high fertility 119 by preferentially investing in testes size at the expense of accessory gland size (35, 36). However, this 120 trade-off could come at a cost of reduced mating rate, which is determined by accessory gland size 121 (37). Similar evolutionary compensation in sperm production may also be present in other taxa (e.g. 122 38; Figure 1), but it is currently unknown how widespread this is and likely to be shaped by the cost of 123 sperm production (e.g. sperm and ejaculate size). Nonetheless, it is clear there are male mitigation 124 strategies that reduce the cost imposed by SGEs.

125 Mitigation by females

SGEs involving sperm manipulation confer direct fitness costs to males that carry them, therefore we expect females to mitigate against mating to such 'inferior' males (4–6). A simple strategy is precopulatory mate choice to avoid mating with SGE males entirely. However, evidence for direct mate choice is remarkably scarce; with only a few good examples of discrimination against SGE carrying males whereas most studies find no such evidence (for review see 5,39; Figure 1). However, SGEs can be costly to a male's fitness in a variety of ways and, as a result, any female-choice for high fitness males might generally select against SGE carrying low-fitness males (40).

133 Post-copulatory mechanisms offer another mitigation route for females. The importance of polyandry, 134 when females mate with multiple males, when at risk of mating with an SGE-carrying male has 135 received much attention (5,39). Polyandry is favoured because SGE-carrying males can be at a 136 disadvantage when competing against other males' undamaged ejaculates due to the production of 137 fewer sperm or sperm with lower vigour (41) and multiple studies across taxa have demonstrated SGE-138 carrying males to be inferior sperm-competitors (e.g. 33,42,43; Table 1). It is worth noting that studies 139 are heavily biased towards SGEs that kill sperm (Table 1). There is also evidence suggesting polyandry 140 could influence SGE frequency in the wild (44,45). The relationship between polyandry and SGEs is 141 dynamic (39). While polyandry can regulate the frequency of SGEs in populations (46) and maintain 142 population viability when at risk from SGEs (47), the presence of SGEs can also in turn directly affect 143 the level of polyandry in a population (48) in part due to female sperm limitation promoting increased 144 remating frequency (49). Apart from promoting sperm competition through polyandry, females at risk 145 of mating with SGE-carrying males may also bias against such males' ejaculates post-mating by 146 selective sperm dumping and/or sperm storage. However, these possibilities are yet to be examined

- 147 more widely (50). In summary, there is a growing body of evidence to suggest that due to the reduced
- sperm competitive ability of SGE-carrying males, polyandry is an effective female mitigation strategy
- 149 (Table 1).

150 4. Evolutionary consequences

The intra-genomic conflict caused by SGEs has implications for male fertility and spermatogenesis. First, the genome can evolve to counteract the costly effects of SGEs and this could disrupt male fertility between populations or between related species harbouring different SGEs. Secondly, intragenomic conflict stemming from SGEs could contribute to the complexity of spermatogenesis.

155 Suppressed SGEs and male fertility

156 The genome can respond to intra-genomic conflict by evolving to suppress the SGEs (12,51). This 157 means many segregation distorters may exist but are fully suppressed. Evidence from Drosophila 158 supports this prediction, with several drive systems only being revealed when closely related species 159 and subspecies hybridize, creating offspring that carry the driver but not its suppressors (52,53). In 160 mice, there is evidence that Sly, a multi-copy Y-linked gene is involved in a co-evolutionary arms-race with the X chromosome resulting in skewed offspring sex-ratios and disrupted gene expression (54). 161 162 These cryptic Drosophila and mouse drive systems result in abnormal spermatogenesis and damage 163 male fertility when expressed. Evidence from inter-population crosses involving a non-suppressed 164 sperm-killer in *D. subobscura* also show hybrid males suffer severe fertility costs (55). These studies 165 are consistent with sperm-killing SGEs and their suppressors playing a role in reducing male fertility in crosses between populations (or species) that harbour SGEs and those that do not. However, an open 166 167 question remains about how widespread a force this genetic conflict is in creating male fertility barriers between populations and potentially contributing to reproductive isolation (56,57). 168

169 SGEs and spermatogenesis

170 The dissection of spermatogenesis at the cellular and molecular level has revealed some intricacies 171 that could be attributed to SGE-fuelled intra-genomic conflict. SGEs could contribute to complexity in 172 spermatogenesis in several ways. First, spermatogenesis-genes may be particularly vulnerable to 173 harbouring SGEs themselves, because suppression of SGEs may come at a cost to male fertility. For 174 example there are three different sperm-killing SGEs in D. simulans, some that are unsuppressed (16,58). Second, if specific SGEs become suppressed or co-evolve with suppressors, over time, these 175 176 genes could become integral to achieving successful sperm production, with male fertility being 177 compromised if either the SGE or the suppression genes are lost (54,59,60). The co-evolution between 178 SGEs and their suppressors could thus lead to increasing number of genes being required for 179 successful spermatogenesis, as has been observed in the Winters meiotic drive system in D. simulans 180 (60). A third related explanation for increasing complexity is the evolution of a general defence against SGEs, whereby genes critical to guarding spermatogenesis against SGEs accumulate (61). The 181 182 proliferation of certain testes-specific gene families, for example the argonautes in Drosophila, is 183 suggested to have evolved to suppress the activity of transposable elements during spermatogenesis 184 (60,61). The impact of SGE-fuelled genomic conflict could therefore contribute new testes specific 185 genes and promote diversification of gene-families associated with generally suppressing a variety of 186 SGEs during spermatogenesis (62,63). Haploid silencing of many genes during spermatogenesis has 187 been implicated as management by the diploid genome to avoid such intra-genomic conflict, (see 188 Sutter et al 2020 in this issue). As our understanding of spermatogenesis deepens, some of its 189 intricacies may turn out to be the result of SGE-fuelled intra-genomic conflict.

190 **5. Summary and future perspectives**

We have discussed the widespread and diverse impact of SGEs on sperm and male fertility and their consequences for mating behaviour and spermatogenesis. There is no doubt that SGEs have profound impact on males' sperm production and reproductive success under polyandry, but that the impacts on male fertility are diverse, ranging from extreme to undetectable costs. Males carrying SGEs will suffer variable fitness consequences depending on the species' mating system biology. However, our current knowledge is limited to a few well studied taxa, and we anticipate that the impact of SGEs are far more widespread than discussed here.

198 While we have focused on vertically transmitted SGEs there are links to other SGEs that are 199 predominantly horizontally transmitted. The ejaculate contains not only sperm, but also a cocktail of 200 seminal proteins with diverse roles in reproduction (64). One speculation is that SGEs are associated 201 with accessory gland proteins (Acps) in the ejaculate. Is it possible that SGEs may be indirectly 202 associated with sperm if bound to Acps that in turn are bound to sperm (65) and/or are present in the 203 ejaculate at mating? The La Crosse virus and Zika virus in mosquitoes can be transmitted by male 204 accessory sex gland fluid rather than by sperm (66,67). However, such SGEs while adversely affecting 205 male fertility are less likely to be transmitted vertically and hence may have a different dynamic.

206 Another expanding area of research is the role of the microbiota in reproduction, and although we 207 know little about the male reproductive microbiome it can contain microorganisms from diverse taxa 208 (68–70) that can be transferred to the female at mating (71,72). There is evidence that the microbiota 209 within the male reproductive tract can adversely affect sperm performance. For example, in humans, 210 there is an association between the microbial community and sperm quality (73). However, it remains 211 unclear how these microbes influence sperm parameters or if this promotes their transmission (see 212 68 for review). Nonetheless, these impacts on male fertility have clear parallels with impacts of 213 vertically transmitted SGEs.

214 The next 50 years of sperm competition research promises to elucidate the prevalence and impact of 215 SGEs on the outcome of sperm competition, and their potency for shaping male mating and ejaculate 216 strategies. SGEs are likely to affect a multitude of areas where the conflict between SGEs and the rest 217 of the genome has not yet been identified. There are also aspects of male reproductive biology where 218 the presence of SGEs have not been extensively considered. No doubt SGEs, and other selfish agents 219 present in the male reproductive tract, have the potential to illuminate some unexplained aspects of 220 male fertility and spermatogenesis and may even be harnessed to suppress harmful vector and pest 221 populations (74).

222

224 Acknowledgements

- 225 We would like to thank Rebecca Jones and Tom Bishop for comments on the early manuscript. We
- would also like the thank Anna Lindholm and Jerry Wilkinson for constructive reviews. Finally, we
- 227 would like to acknowledge Geoff Parker for defining the field of sperm competition that continues to
- 228 inspire work in a diversity of research topics, including selfish genetic elements.

230 Table and Figure

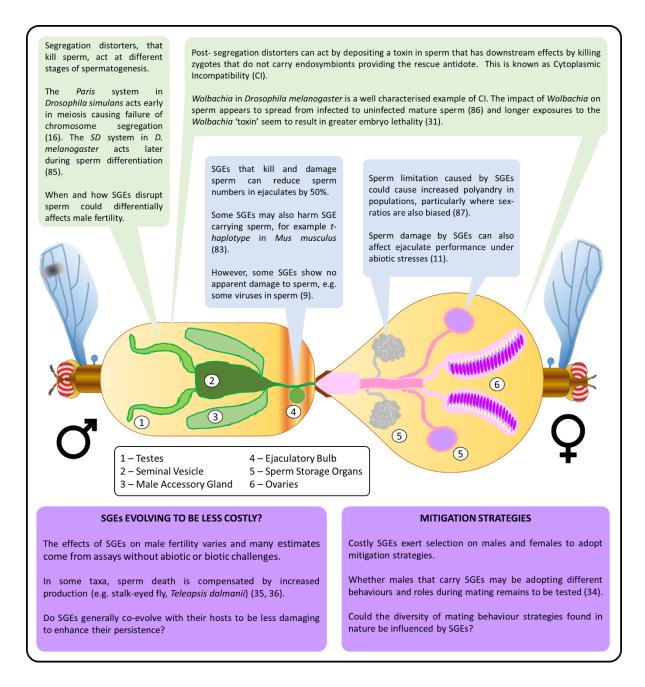
- 231 **Table 1**
- 232 Effects of selfish genetic elements (SGE) on sperm competitive ability.

SGE	Host	P1 ¹	P2 ²	Overall	Male mated	Reference
				paternity ³	status	
SR (X-linked driver)	Teleopsis* whitei	0.099	0.101	0.1	Virgin	(43)
SR (X-linked driver)	T. whitei	0.125 5	NA	NA	Virgin	(75)
SR (X linked driver)	T. dalmanni	NA	0.25	NA	Non-virgin	(76)
SR (X linked driver)	Drosophila pseudoobscura	0.02	0.83	0.42 ⁵	Virgin	(77)
SR (X linked driver)	D. pseudoobscura	0.38	0.32	0.35 ⁵	Non-virgin	(77)
SR (X linked driver)	D. pseudoobscura	0.35	0.14	0.25	Non-virgin	(33)
SR (X linked driver)	D. recens	NA	NA	0.30 ⁵	Non-virgin	(78)
SR (X linked driver)	D. simulans	0.10	0.50	0.30	Virgin	(79)
SR (X linked driver)	D. simulans	0.12	0.34	0.22	Virgin	(80)
<i>Wolbachia</i> (Riverside strain)	D. simulans	0.15	0.72	0.44	Non-virgin	(81)
PSR (B	Nasonia	0.08	0.58	0.25	Non-virgin	(82)
chromosome)	vitripennis					
t-haplotype	Mus musculus	0.22	0.05	0.13	Virgin	(83)
(autosomal driver)						

t-haplotype	M. musculus	NA	NA	0.24	Virgin	(84)
(autosomal driver)						

233

234 Sperm-competitive success of SGE males where a female is mated to 2 males unless otherwise stated. ¹P1 235 is the percentage of offspring fathered by the first of two males to mate with the same female. ²P2 is the 236 percentage of offspring fathered by the second of two males to mate with the same female. ³Overall 237 paternity is the mean of P1 and P2, and is the overall paternity expected under sperm competition when SGE status does not affect mating order. ⁴ 2nd male only transferred seminal fluids and not sperm, an 238 239 estimate of 0.125 is therefore extrapolated from SR males producing 25% as many offspring as ST males when exposed to the seminal fluid of a 2nd male. ⁵Paternity estimated from competition of Sex-Ratio and 240 241 standard males against an inferior tester mutant strain, potentially causing an underestimated P1 and 242 overestimated P2. Table modified from (4). *The Cyrtodiopsis genus was synonymized with Teleopsis in 2001. 243



²⁴⁵

Figure 1. The main arenas where SGEs are known to act from spermatogenesis to fertilisation (green), pictured in insects. In blue, highlights costs of carrying SGEs to sperm production and sperm competitive ability. In purple, examples of evolutionary impacts of SGEs on sperm and mitigation strategies.

251 References

Burt A, Trivers R. (2006) *Genes in conflict: The Biology of selfish genetic elements.* Cambridge, Massachusetts: Harvard University Press.

254 2. Werren JH, Stouthamer R. (2003) *PSR* (*paternal sex ratio*) chromosomes: The ultimate selfish 255 genetic elements. *Genetica* 117:85–101.

- 256 3. Lindholm AK, Dyer KA, Firman RC, Fishman L, Forstmeier W, Holman L, et al. (2016) The 257 ecology and evolutionary dynamics of meiotic drive. *Trends Ecol Evol*. 31:315–326.
- Price TAR, Wedell N. (2008) Selfish genetic elements and sexual selection: Their impact on
 male fertility. *Genetica* 134:99–111.
- Wedell N. (2020) Selfish genes and sexual selection: the impact of genomic parasites on host
 reproduction. *J Zool*. 311:1-12.
- 262 6. Zanders SE, Unckless RL. (2019) Fertility Costs of Meiotic Drivers. *Curr Biol*. 29:R512–520.
- 263 7. Gershenson S. (1928) A new sex-ratio abnormality in *Drosophila obscura*. *Genetics* 13:488–
 264 507.
- 265 8. Seidel HS, Rockman M V, Kruglyak L. (2008) Widespread genetic incompatibility in *C. elegans*266 maintained by balancing selection. *Science* 319:589–594.
- Mao Q, Wu W, Liao Z, Li J, Jia D, Zhang X, et al. (2019) Viral pathogens hitchhike with insect
 sperm for paternal transmission. *Nat Comm.* 10:1–10.
- 10. Jaenike J. (2001) Sex Chromosome Meiotic Drive. *Annu Rev Ecol Evol Syst.* 49:25–49.
- Price TAR, Hoskyns RC, Rapley H, Evans JC, Wedell N. (2012) No evidence that temperaturerelated fertility differences influence the distribution of a selfish genetic element. *Funct Ecol.*26:657–665.
- 273 12. Meiklejohn CD, Tao Y. (2009) Genetic conflict and sex chromosome evolution. *Trends Ecol*274 *Evol*. 25:215–223.
- 275 13. Courret C, Chang CH, Wei KHC, Montchamp-Moreau C, Larracuente AM. (2019) Meiotic
 276 drive mechanisms: Lessons from *Drosophila*. *Proc R Soc B Biol Sci*. 286: 20191430.
- 277 14. Bravo Núñez MA, Nuckolls NL, Zanders SE. (2018) Genetic villains: Killer meiotic drivers.
 278 *Trends Genet*. 34:424–33.
- 15. Bauer H, Véron N, Willert J, Herrmann BG. (2007) The t-complex-encoded guanine
 nucleotide exchange factor Fgd2 reveals that two opposing signaling pathways promote
 transmission ratio distortion in the mouse. *Genes Dev.* 21:143–147.
- Montchamp-Moreau C. (2006) Sex-ratio meiotic drive in *Drosophila simulans*: cellular
 mechanism, candidate genes and evolution. *Biochem Soc Trans*. 34:562–565.
- 17. Longdon B, Wilfert L, Obbard DJ, Jiggins FM. (2011) Rhabdoviruses in two species of
 Drosophila: Vertical transmission and a recent sweep. *Genetics* 188:141–150.
- 18. Chen Q, Godfrey K, Liu J, Mao Q, Kuo Y-W, Falk BW. (2019) A nonstructural protein
 responsible for viral spread of a novel insect reovirus provides a safe channel for biparental virus
 transmission to progeny. *J Virol.* 93:1–21.

19. Beukeboom LW, Seif M, Mettenmeyer T, Plowman AB, Michiels NK. (1996) Paternal
inheritance of B chromosomes in a parthenogenetic hermaphrodite. *Heredity* 77:646–654.

20. Cabrero, J., Martín-Peciña, M., Ruiz-Ruano, F.J., Gómez, R. and Camacho, J.P.M., 2017. Postmeiotic B chromosome expulsion, during spermiogenesis, in two grasshopper species. *Chromosoma*126:633-644.

Hurst GD, Werren JH. (2001) The role of selfish genetic elements in eukaryotic evolution. *Nat Rev Genet.* 2:597–606

296 22. Werren JH, Baldo L, Clark ME. (2008) *Wolbachia*: master manipulators of invertebrate
297 biology. *Nat Rev Microbiol.* 6:741–751.

298 23. Werren JH. (1997) Biology of Wolbachia. Annu Rev Entomol. 42:587–609.

24. Biwot JC, Zhang HB, Liu C, Qiao JX, Yu XQ, Wang YF. (2020) *Wolbachia*-induced expression of
kenny gene in testes affects male fertility in *Drosophila melanogaster*. *Insect Sci.* in press
(doi.org/10.1111/1744-7917.12730)

302 25. Awrahman ZA, Champion de Crespigny F, Wedell N. (2014) The impact of *Wolbachia*, male
303 age and mating history on cytoplasmic incompatibility and sperm transfer in *Drosophila simulans*. J
304 *Evol Biol*. 27:1–10.

26. De Crespigny FEC, Wedell N. (2006) *Wolbachia* infection reduces sperm competitive ability in an insect. *Proc R Soc B Biol Sci.* 273:1455–1458.

307 27. Hariri AR, Werren JH, Wilkinson GS. (1998) Distribution and reproductive effects of
308 Wolbachia in stalk-eyed flies (Diptera: Diopsidae). *Heredity* 81:254–60.

Snook RR, Cleland SY, Wolfner MF, Karr TL. (2000) Offsetting effects of *Wolbachia* infection
and heat shock on sperm production in *Drosophila simulans*: Analyses of fecundity, fertility and
accessory gland proteins. *Genetics* 155:168–178.

Wade MJ, Chang NW. (1995) Increased male fertility in *Tribolium confusum* beetles after
infection with the intracellular parasite *Wolbachia*. *Nature* 373:72–74.

30. Hilgenboecker K, Hammerstein P, Schlattmann P, Telschow A, Werren JH. (2008) How many
species are infected with *Wolbachia*? - A statistical analysis of current data. *FEMS Microbiol Lett.*281:215–220.

31. Karr TL, Yang W, Feder ME, Karr TL, Yang W, Feder ME. (1998) Overcoming cytoplasmic
318 incompatibility in *Drosophila. Proc R Soc B Biol Sci.* 265:391–395.

319 32. Runge JN, Lindholm AK. (2018) Carrying a selfish genetic element predicts increased
320 migration propensity in free-living wild house mice. *Proc R Soc B Biol Sci.* 285(1888).

32. Price TAR, Bretman AJ, Avent TD, Snook RR, Hurst GDD, Wedell N. (2008) Sex ratio distorter
 reduces sperm competitive ability in an insect. *Evolution* 62:1644–52.

323 34. Ball MA, Parker GA. (2000) Sperm competition games: A Comparison of loaded raffle models
324 and their biological implications. *J Theor Biol*. 206:487–506.

325 35. Meade LC, Dinneen D, Kad R, Lynch DM, Fowler K, Pomiankowski A. (2019) Ejaculate sperm
326 number compensation in stalk-eyed flies carrying a selfish meiotic drive element. *Heredity* 122:916–
327 926.

331 accessory reproductive organ size in the stalk-eyed fly Cyrtodiopsis dalmanni BMC Evol Biol. 6:1-6. 332 Hauschteck-Jungen E, Burkard W, Jungen H, Burch-Schwaller R. (1987) The loss of Y-sperm in 38. 333 "sex-ratio" (SR) males of Drosophila subobscura is compensated. Genetica 74:27–30. 334 39. Wedell N. (2013) The dynamic relationship between polyandry and selfish genetic elements: 335 The dynamic relationship between polyandry and selfish genetic elements. *Proc R Soc B Biol Sci.* 368: 336 20120049. 337 40. Verspoor RL, Hurst GDD, Price TAR. 2016 The ability to gain matings, not sperm competition, 338 reduces the success of males carrying a selfish genetic element in a fly. Anim Behav. 115:207–215. 339 41. Haig D, Bergstrom CT. (1995) Multiple mating, sperm competition and meiotic drive. J Evol 340 Biol. 8:265-282. 42. 341 Sutter A, Lindholm AK. (2016) Meiotic drive changes sperm precedence patterns in house 342 mice: Potential for male alternative mating tactics? BMC Evol Biol. 16:1–15. 343 43. Wilkinson GS, Fry CL. (2001) Meiotic drive alters sperm competitive ability in stalk-eyed flies. 344 Proc R Soc B Biol Sci. 268:2559-2564. 345 44. Pinzone CA, Dyer KA. (2013) Association of polyandry and sex-ratio drive prevalence in 346 natural populations of Drosophila neotestacea. Proc R Soc B Biol Sci. 2013:280. 347 45. Price TAR, Bretman A, Gradilla AC, Reger J, Taylor ML, Giraldo-perez P, et al. (2014) Does 348 polyandry control population sex ratio via regulation of a selfish gene? Proc R Soc B Biol Sci. 281: 20133259. 349 350 46. Manser A, Lindholm AK, Konig B, Bagheri HC. (2011) Polyandry and the decrease of a selfish 351 genetic element in a wild house. Evolution 65:2435–2447. 352 47. Price TAR, Hurst GDD, Wedell N. (2010) Polyandry prevents extinction. Curr Biol. 20:471-475. 353 354 48. Price TAR, Hodgson DJ, Lewis Z, Hurst GDD, Wedell N. (2008) Selfish genetic elements 355 promote polyandry in a fly. *Science* 322:1241–1243. 356 49. Charlat S, Reuter M, Dyson EA, Hornett EA, Duplouy A, Davies N, et al. (2007) Male-killing 357 bacteria trigger a cycle of increasing male fatigue and female promiscuity. Curr Biol. 20:471–475. 358 50. Price T, Lewis Z, Wedell N. (2009) Sperm dumping as a defense against meiotic drive. J Biol. 359 6:8-11. 360 51. Bastide H, Gérard PR, Ogereau D, Cazemajor M, Montchamp-Moreau C. (2013) Local 361 dynamics of a fast-evolving sex-ratio system in Drosophila simulans. Mol Ecol. 22:5352–5367. 362 52. Tao Y, Hartl DL, Laurie CC. (2001) Sex-ratio segregation distortion associated with 363 reproductive isolation in Drosophila. Proc Natl Acad Sci U S A 98:13183–13188. 364 Phadnis N, Orr HA. (2009) Sterility and Segregation Distortion in Drosophila Hybrids. Science 53. 323:376-379. 365

36. Meade, L., Finnegan, S. R., Kad, R., Fowler, K., & Pomiankowski, A. (2020). Maintenance of

Rogers DW, Chapman T, Fowler K, Pomiankowski A. (2005) Mating-induced reduction in

fertility in the face of meiotic drive. The American Naturalist, 195:743-751.

328

329

330

37.

- Second Sec
- 368 55. Verspoor RL, Price TAR, Smith JML, Mannion NLM, Hurst GDD. (2018) Strong hybrid male
 369 incompatibilities impede the spread of a selfish chromosome between populations of a fly. *Evol Lett.*370 169–179.
- 371 56. Presgraves DC. (2010) The molecular evolutionary basis of species formation. *Nat Rev Genet*.
 372 11:175–180.
- 373 57. Crespi B, Nosil P. (2013) Conflictual speciation: species formation via genomic conflict.
 374 *Trends Ecol Evol.* 28:48–57.
- Tao Y, Araripe L, Kingan SB, Ke Y, Xiao H, Hartl DL, 2007 A sex-ratio meiotic drive system in *Drosophila simulans*. II: An X-linked Distorter. PLoS Biology 5:e293
- 377 598. Price TAR, Verspoor RL, Wedell N. (2019) Ancient gene drives: an evolutionary paradox. *Proc*378 *R Soc B Biol Sci.* 2267:10.1098.
- Lin, C.J., Hu, F., Dubruille, R., Vedanayagam, J., Wen, J., Smibert, P., Loppin, B. and Lai, E.C.,
 2018. The hpRNA/RNAi pathway is essential to resolve intragenomic conflict in the *Drosophila* male
 germline. Developmental cell, 46(3), pp.316-326.
- 382 61. Dechaud C, Volff JN, Schartl M, Naville M. (2019) Sex and the TEs: Transposable elements in
 383 sexual development and function in animals. *Mobile DNA* 10:1–15.
- Lewis SH, Webster CL, Salmela H, Obbard DJ. (2016) Repeated duplication of Argonaute2 is
 associated with strong selection and testis specialization in *Drosophila*. *Genetics* 204:757–769.
- 386 63. Baker, R.H., Narechania, A., DeSalle, R., Johns, P.M., Reinhardt, J.A. and Wilkinson,
- G.S. 2016 Spermatogenesis drives rapid gene creation and masculinization of the X chromosome in
 stalk-eyed flies (Diopsidae). *Genome Biology and Evolution* 8: 896-914
- Wigby S, Sirot LK, Linklater JR, Buehner N, Calboli FCF, Bretman A, et al. (2009) Seminal fluid
 protein allocation and male reproductive success. *Curr Biol.* 19:751–757.
- Bound Sex-Peptide Controls Female Postmating Behavior in *Drosophila*. *Curr Biol*. 15:207–213.
- 393 66. Thompson WH, Beaty BJ. (1977) Venereal transmission of La Crosse (California encephalitis)
 394 arbovirus in *Aedes triseriatus* mosquitoes. *Science* 196:530–531.
- 395 67. Campos SS, Fernandes RS, Araujo A, Miranda RM De, Telleria EL, Ferreira-de-brito A, et al.
 396 (2019) Zika virus can be venereally transmitted between *Aedes aegypti* mosquitoes. *Parasites and*397 *Vectors* 2017:1–4.
- Rowe M, Veerus L, Trosvik P, Buckling A, Pizzari T. (2020) The reproductive microbiome: An
 emerging driver of sexual selection, sexual conflict, mating systems, and reproductive isolation. *Trends Ecol Evol.* 35:220–234.
- 401 69. Skau PA, Folstad I. (2003) Do bacterial infections cause reduced ejaculate quality? A meta-402 analysis of antibiotic treatment of male infertility. *Behav Ecol.* 14:40–47.
- 403 70. Baud D, Pattaroni C, Vulliemoz N, Castella V. (2019) Sperm microbiota and its impact on
 404 semen parameters. *Front Microbiol.* 10:1–9.

405 71. Knell RJ, Webberley KM. (2004) Sexually transmitted diseases of insects: distribution,
406 evolution, ecology and host behaviour. *Biol Rev.* 79:557–581.

407 72. Otti O. (2015) Genitalia-associated microbes in insects. *Insect Sci.* 22:325–39.

Weng S, Chiu C, Lin F, Huang W, Liang C, Yang T, et al. (2014) Bacterial communities in semen
from men of infertile couples: metagenomic sequencing reveals relationships of seminal microbiota
to semen quality. *PLoS One*. 9(10).

411 74. Wedell N, Price TAR, Lindholm AK. (2019) Gene drive: progress and prospects. *Proc R Soc B*412 *Biol Sci.* 286: 20192709.

413 75. Fry CL, Wilkinson GS. (2014) Sperm survival in female stalk-eyed flies depends on seminal
414 fluid and meiotic drive. *Evolution* 58:1622–1626.

415 76. Wilkinson GS, Johns PM, Kelleher ES, Muscedere ML, Lorsong A. (2006) Fitness effects of X
416 chromosome drive in the stalk-eyed fly, *Cyrtodiopsis dalmanni*. *J Evol Biol*. 9:1851–1860.

417 77. Wu Cl. (1983) Virility deficiency and the sex-ratio trait in *Drosophila pseudoobscura*. II.
418 Multiple mating and overall virility selection. *Genetics* 105:663–679.

419 78. Dyer KA, Hall DW. (2019) Fitness consequences of a nonrecombining sex-ratio drive
420 chromosome can explain its prevalence in the wild. *Proc R Soc B Biol Sci.* 286: 20192529.

421 79. Atlan, A., Joly D, Capillon C, Montchamp-Moreau C. (2004) Sex-ratio distorter of *Drosophila*422 *simulans* reduces male productivity and sperm competition ability. *J Evol Biol.* 17:744–751.

423 80. Angelard C, Montchamp-Moreau C, Joly D. (2008) Female-driven mechanisms, ejaculate size
424 and quality contribute to the lower fertility of sex-ratio distorter males in *Drosophila simulans*. *BMC*425 *Evol Biol.* 8:326.

426 81. De Crespigny FEC, Pitt TD, Wedell N. (2006) Increased male mating rate in *Drosophila* is 427 associated with *Wolbachia* infection. *J Evol Biol.* 19:1964–1972.

Beukeboom LW. (1994) Phenotypic fitness effects of the selfish B chromosome, paternal
sex-ratio (PSR) in the parasitic wasp *Nasonia vitripennis*. *Evol Ecol.* 8:1–24.

430 83. Sutter A, Lindholm AK. (2015) Detrimental effects of an autosomal selfish genetic element
431 on sperm competitiveness in house mice. *Proc R Soc B Biol Sci.* 282.

432 84. Manser, A., Lindholm, A. K., Simmons, L. W., & Firman, R. C. (2017) Sperm competition
433 suppresses gene drive among experimentally evolving populations of house mice. *Molecular ecology*434 26:5784-5792.

435 85. Larracuente AM, Presgraves DC. (2012) The selfish Segregation Distorter gene complex of 436 *Drosophila melanogaster. Genetics* 192: 33–53.

437 86. Riparbelli MG, Giordano R, Callaini G. (2007) Effects of *Wolbachia* on sperm maturation and 438 architecture in *Drosophila simulans* Riverside. *Mech Dev.* 124: 699–714.

439 87. Holman L, Snook RR. (2008) A sterile sperm caste protects brother fertile sperm from
440 female-mediated death in *Drosophila pseudoobscura*. *Curr. Biol.* 18:292-296.

441