

1 **Senescence in duckweed: age-related declines in survival, reproduction, and**  
2 **offspring quality**

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19 **Running title**

20 Senescence in duckweed

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24 **Summary**

25 **1.** As they grow old, most organisms experience progressive physiological deterioration  
26 resulting in declining rates of survival and reproduction – a seemingly maladaptive  
27 phenomenon known as senescence.

28 **2.** Although senescence is usually defined with respect only to survival and reproduction, a  
29 third component of fitness, offspring quality, may also decline with age. Few studies,  
30 however, have assessed age-related changes in offspring quality using measures that truly  
31 reflect fitness.

32 **3.** In a controlled environment, we tested for age-related declines in three demographic  
33 components of fitness (survival, reproduction, and offspring quality) in *Lemna minor*, a  
34 small aquatic plant in the subfamily Lemnoideae (the duckweeds) with a short lifespan and  
35 rapid rate of asexual reproduction. Our primary measure of offspring quality, the intrinsic  
36 rate of increase, more closely approximates fitness than measures used in previous studies  
37 such as size, lifespan, and total reproductive output.

38 **4.** We observed strong age-related declines in all three components of fitness: old plants  
39 had lower rates of survival and reproduction, and produced lower-quality offspring than  
40 younger plants.

41 **5.** Theoretical and empirical research on the evolutionary biology of senescence should  
42 devote more attention to offspring quality. This often unrecognized component of fitness  
43 may change with age – as we have shown in *L. minor* – and may be shaped by, and feed  
44 back into, the same evolutionary forces that give rise to senescence.

45

46 **Key-words:** ageing, Lansing effect, *Lemna minor*, life history, parental age effects

47 **Introduction**

48 Senescence is characterized by progressive physiological deterioration and age-related  
49 declines in survival and reproduction (reviewed in Kirkwood & Austad 2000; Hughes &  
50 Reynolds 2005; Williams *et al.* 2006; Sherratt & Wilkinson 2009). Such declines are  
51 seemingly deleterious from the perspective of an ageing individual, and yet senescence  
52 occurs in many taxa (Jones *et al.* 2014). Explaining the evolution and maintenance of  
53 senescence has therefore been an important challenge in evolutionary biology.

54 In the most general sense, the evolutionary paradox of senescence concerns age-  
55 related declines in the expectation of future genetic representation (i.e. fitness). All else  
56 being equal, a lineage that is not subject to age-related declines in fitness should have  
57 greater future representation than one that is. Although many authors define senescence  
58 with respect only to survival and reproduction, there is increasing evidence that another  
59 component of fitness, offspring quality, may also decline with age (Kern *et al.* 2001). For  
60 example, a decline in offspring lifespan with increasing parental age (known as the Lansing  
61 effect) has been observed in a variety of taxa including rotifers (Lansing 1947, 1948),  
62 ladybird beetles (Singh & Omkar 2009), duckweeds (Ashby & Wangermann 1949), and  
63 humans (Bell 1918; Gavrilov & Gavrilova 1997) (additional examples are cited in Priest,  
64 Mackowiak & Promislow 2002). Similarly, advanced parental age has been shown to  
65 negatively affect offspring fecundity schedules in great tits (Bouwhuis *et al.* 2010) and pre-  
66 industrial humans (Gillespie, Russell & Lummaa 2013a).

67 Age-related declines in offspring quality are paradoxical in much the same way as  
68 age-related declines in survival and reproduction. All else being equal, lineages not subject  
69 to age-related declines in offspring quality should have greater future representation than

70 those that are. Of course, this argument is only valid insofar as offspring 'quality' reflects  
71 biological fitness. Lifespan is generally a poor measure of fitness (e.g. Jenkins, McColl &  
72 Lithgow 2004), so despite the apparent prevalence of age-related declines in offspring  
73 lifespan, the extent to which offspring fitness declines with parental age remains unclear.  
74 Resolving this gap in our understanding is important because established theories of life  
75 history evolution and senescence implicitly assume that offspring fitness is constant with  
76 parental age (e.g. Williams 1957; Hamilton 1966; Kirkwood & Rose 1991). If this is not the  
77 case, then the force of selection cannot be understood simply in terms of age-specific  
78 survival and fecundity, but may also depend on age-specific patterns of change in offspring  
79 fitness (e.g. Pavard, Koons & Heyer 2007). As Caswell (2001, p. 280) points out: "The  
80 paradoxes of life history theory mean that selection must be studied in terms of the entire  
81 life cycle. The alternative – analysis in terms of a subset of vital rates, or what are called  
82 *components of fitness* – risks getting answers that are qualitatively wrong." Thus, if  
83 offspring fitness does indeed change with parental age, evolutionary analyses that ignore  
84 such changes may lead us astray.

85         Here we test for age-related declines in three major demographic components of  
86 fitness (survival, reproduction, and offspring fitness) in *Lemna minor* L., a small and short-  
87 lived aquatic plant (Landolt 1986). Our primary interest is to understand whether offspring  
88 fitness declines with increasing parental age. *Lemna minor* is an excellent species in which  
89 to address this question for two reasons. First, reproduction in *L. minor* is almost  
90 exclusively asexual, which simplifies the analysis of parental age effects (there is only one  
91 parent to account for and it is easy to identify). Second, previous research suggests *L. minor*  
92 may be subject to parental-age-related declines in various offspring traits potentially

93 relating to fitness, including offspring size, lifespan, and lifetime reproductive output  
94 (Wangermann & Ashby 1950, 1951; but see Claus 1972). Because there is a premium on  
95 early reproduction, lifespan and lifetime reproductive output may be poor measures of  
96 overall fitness (Stearns 1992; Partridge & Barton 1996). Thus, to understand whether *L.*  
97 *minor* is subject to age-related declines in offspring fitness (in addition to age-related  
98 declines in survival and reproduction), we employ a demographic measure that better  
99 approximates realized fitness – the intrinsic rate of increase ( $r$ ) measured at the level of  
100 individual offspring.

101

102

### 103 **Materials and methods**

#### 104 STUDY SPECIES

105 *Lemna minor* is a small aquatic plant belonging to Lemnoideae (the duckweeds), a  
106 subfamily comprising “the simplest and smallest of flowering plants” (Hillman 1961, p.  
107 222). It occurs in slow-moving freshwater bodies on every continent except Antarctica  
108 (Landolt 1986), and is tolerant to a wide range of environmental conditions (Wang 1990;  
109 Mkandawire & Dudel 2000). Individual plants are about 3–5 mm long and consist of a free-  
110 floating frond (also called a thallus; a combination of leaf and stem) and a single root that  
111 emanates from the frond’s lower surface (Lemon & Posluszny 2000). Proliferation of *L.*  
112 *minor* is dominated by vegetative reproduction – offspring (often referred to as daughter  
113 fronds) develop asexually in alternating succession from one of two meristematic pockets  
114 within the parent (Landolt 1986). Under optimal laboratory conditions, each plant will  
115 produce about 15 offspring within a lifespan of approximately 30 days (Lemon, Posluszny

116 & Husband 2001). We note that, unlike most vascular plants, duckweeds have a unitary  
117 growth form and determinate growth potential – maximum frond size is usually achieved  
118 prior to a frond detaching from its parent (Hillman 1961).

119

## 120 OVERVIEW

121 We tested for age-related declines in components of *L. minor* fitness in two phases.

122

### 123 *Phase One: Survival and Reproduction*

124 First, to measure the influence of age on rates of survival and reproduction, we  
125 isolated 216 fronds individually in Petri dishes containing a liquid growth medium, and  
126 observed the fronds daily for the duration of their lives. The first day of life was defined as  
127 the day that a frond detached from its parent, and death was defined as the day that a  
128 frond's final daughter detached (there are no obvious physiological definitions of death in *L.*  
129 *minor*, as the progression of cell death during frond senescence generally spans 10 or more  
130 days). Every day during a frond's lifetime, we observed whether or not the frond  
131 reproduced – i.e. whether any of its daughters detached since the previous day's  
132 observation. Detached daughters were aseptically removed from the Petri dish and  
133 discarded.

134

### 135 *Phase Two: Offspring Quality*

136 The second phase of our study examined changes in offspring quality (measures  
137 included the intrinsic rate of increase, total reproductive output, latency to reproduce,  
138 lifespan, and frond size) as a function of parental age. We isolated 41 'parental' fronds

139 individually in Petri dishes, and observed them daily for the duration of their lives as  
140 described above. This time, however, instead of being discarded, the daughters (the ‘focal’  
141 generation, N = 542) of the 41 parental fronds were transferred to their own Petri dish  
142 upon detaching from the parent, randomly assigned to one of three growth chambers, and  
143 observed for reproduction daily for the duration of their lives. Four of the 542 focal fronds  
144 (all of which were the final daughters produced by their respective parents) remained  
145 attached to their parent for a prolonged period of time – well into their reproductive  
146 lifespan. We defined the first day of life for these four individuals as the day that their first  
147 daughter detached.

148

#### 149 PLANTS AND GROWTH CONDITIONS

150 The plants used in this study were derived from a clonal lineage that we obtained from the  
151 Canadian Phycological Culture Centre (CPCC 492 *Lemna minor*; originally collected from  
152 Elk Lake, British Columbia, Canada; 48° 31' 30" N, 123° 23' 18" W). We studied a genetically  
153 homogeneous sample because heterogeneity (both genetic and environmental) can  
154 sometimes mask true patterns of senescence (Zens & Peart 2003). Due to the possibility of  
155 parental-age effects in *L. minor* (e.g. Wangermann & Ashby 1950, 1951), we also strove for  
156 ‘genealogical’ homogeneity among our focal plants. Specifically, the 216 focal fronds in  
157 *Phase one* and 41 parental fronds in *Phase two* were each first daughters of first daughters  
158 (etc.) going back at least five generations.

159         Plants were aseptically cultured in 60 × 10 mm Petri dishes containing 10 ml of  
160 Modified Hoagland’s E+ growth medium (Environment Canada 2007), and kept inside  
161 growth chambers set to 25°C with a 12:12 photoperiod and a photosynthetic photon flux

162 density at plant height of approximately  $500 \mu\text{mol m}^{-2} \text{sec}^{-1}$ . To ensure environmental  
163 constancy (e.g. to account for evaporation, nutrient depletion, etc.), we aseptically  
164 transferred each plant into a new Petri dish with 10 ml of fresh growth medium every four  
165 days. Two of the 216 fronds from *Phase one* developed bacterial contamination and so were  
166 discarded and not included in the analyses below. There was no bacterial contamination  
167 during *Phase two*. Low rates of fungal contamination occurred in both phases of our study,  
168 always taking the form of an isolated clump of stringy white fungus within the growth  
169 medium. When such contamination was detected, the corresponding frond was aseptically  
170 transferred to a new Petri dish with fresh growth medium. This intervention was  
171 successful given that no plant was ever subject to more than a single instance of fungal  
172 contamination.

173

## 174 FITNESS MEASURES

### 175 *Phase One: Survival and Reproduction*

176 Our measures of fitness in *Phase one* were daily rate of survival and daily rate of  
177 reproduction conditional on survival. Although fronds occasionally released two daughters  
178 on the same day (this occurred in 8.6% of the reproductive events that we observed), we  
179 chose to analyze reproduction as a binary event (0 = did not reproduce, 1 = released one or  
180 two daughters). Treating reproduction as binary instead of ordinal made it easier  
181 (statistically) to account for non-independence due to repeated observations on the same  
182 individuals.

183

### 184 *Phase two: Offspring Quality*



185 Our primary measure of offspring fitness was the intrinsic rate of increase ( $r$ )  
186 measured at the level of individual fronds, as described in McGraw and Caswell (1996).  
187 Intrinsic rate of increase is an appropriate measure of fitness for stable populations under  
188 constant environmental conditions (Metcalf & Pavard 2007), and can be calculated as the  
189 natural logarithm of the dominant eigenvalue of a Leslie matrix. To construct a Leslie  
190 matrix for single individuals, the age-specific survival rate was set to 1 for each age at  
191 which an individual survived, and 0 for every other age (McGraw & Caswell 1996).  
192 Measuring fitness in this way – at the level of the individual – is sometimes problematic due  
193 to a lack of replication (Link, Cooch & Cam 2002). However, our use of a single clone  
194 negates this problem. The realized fitness of replicate fronds of a given parental age should  
195 reflect the same underlying fitness propensity (or ‘latent fitness’), and thus, our approach  
196 entails appropriate replication.

197 In addition to our primary measure of offspring fitness (the intrinsic rate of  
198 increase), we examined four secondary measures of offspring quality (not necessarily  
199 directly related to fitness): total number of offspring produced, latency to first  
200 reproduction (days between detachment from parent and first daughter detaching;  
201 inversely related to fitness), lifespan (days between detachment from parent and last  
202 daughter detaching), and frond surface area. Frond surface area was measured in ImageJ v.  
203 1.43u (Rasband 2012) using images captured with a microscope-mounted digital camera.  
204 Images used for surface-area measurement were captured late in a frond’s life when it had  
205 no attached daughters. Occasionally, fronds produced late in their parent’s life were ‘curled’  
206 (see Figure S1 under Supporting Information), which complicated the measurement of  
207 surface area. For the 42 focal fronds in *Phase two* that were curled, we estimated surface

208 area based on the length of each frond's longitudinal axis (Fig. S1). These 'corrected'  
209 estimates were interpolated from a linear regression of surface area on length for the 500  
210 non-curved fronds (Fig. S2).

211

## 212 DATA ANALYSIS

213 All analyses were conducted in R v. 3.0.2 (R Core Team 2013).

214

### 215 *Phase One: Survival and Reproduction*

216 To understand how daily rates of survival varied with age, we fit and compared four  
217 candidate survival models (described in Pletcher, Khazaeli & Curtsinger 2000; Sherratt *et*  
218 *al.* 2010): exponential, Weibull, Gompertz, and logistic. The exponential model serves as a  
219 null hypothesis of no senescence because it assumes a constant rate of survival with age,  
220 whereas survival may decline with age in the other models. All survival models were fit by  
221 maximizing log-likelihood functions using the *optim* function in R, and strength of evidence  
222 was assessed using the Akaike information criterion corrected for small sample sizes,  $AIC_c$   
223 (Burnham & Anderson 2002).

224 To test for age-related declines in the daily rate of reproduction, we used  
225 generalized estimating equations (GEE) with a binomial error structure and logit link, fit  
226 with the *geeglm* function in the R package *geepack* (Halekoh, Højsgaard & Yan 2006). The  
227 GEE approach was ideal for our analysis given the possibility of within-individual negative  
228 temporal autocorrelation in reproduction (i.e. an individual that reproduces on a given day  
229 is somewhat less likely to reproduce the very next day). Due to this possibility, we favoured  
230 (based on biological relevance) a first-order autoregressive (AR-1) correlation structure,

231 which assumes that the correlation between repeated observations on the same subject is  
232 inversely related to the distance (or time) between those observations. Other common  
233 correlation structures include ‘exchangeable’ (constant within-subject correlation; similar  
234 to a mixed-effects model with subject-level random intercepts) and ‘independence’ (no  
235 within-subject correlation; equivalent to a generalized linear model) (Zuur *et al.* 2009). We  
236 used the Rotnitzky–Jewell (RJ) criteria (Rotnitzky & Jewell 1990) and the rule-out criterion  
237 proposed by Shults *et al.* (2009) to compare the three correlation structures described  
238 above, and a Wald test to assess the effect of age on probability of reproduction. The RJ  
239 criteria include three metrics by which to compare robust (empirical) estimates of a  
240 covariance matrix to naïve (model-based) covariance estimates. The model in which the  
241 working correlation structure best approximates the ‘true’ correlation structure is the  
242 model for which empirical and model-based covariance estimates are most similar (Wang  
243 & Carey 2004; Shults *et al.* 2009). The rule-out criterion rejects correlation structures  
244 yielding estimated covariance matrices that are not positive definite – indicative of a  
245 misspecified correlation structure (Crowder 1995; Schults *et al.* 2009). Note that, in the  
246 analyses of reproduction described above, we excluded data for the first day of each frond’s  
247 life because none of the 216 focal fronds in *Phase one* reproduced on day one.

248

#### 249 *Phase Two: Offspring Quality*

250 To understand whether offspring quality declined with parental age, we modeled  
251 our primary measure of offspring fitness (intrinsic rate of increase) and secondary  
252 measures of offspring quality (total offspring, latency to first reproduction, lifespan, and  
253 surface area) as functions of the age of the parent when the focal frond (i.e. offspring)

254 detached, while controlling for the growth chamber that the focal frond was assigned to. All  
255 of the relationships between offspring quality and parental age were nonlinear and could  
256 not be transformed to linearity, so in all cases we examined polynomials of parental age up  
257 to a degree of three.

258         The modeling approach described hereafter follows Zuur *et al.* (2009). To account  
259 for potential non-independence of offspring derived from the same parent, we initially fit  
260 linear mixed models describing a given measure of offspring quality as a function (either  
261 linear, quadratic, or cubic) of parental age and linear function of growth chamber, with one  
262 of three random effect structures: (i) random intercept and slope terms for parent identity,  
263 (ii) random intercept term for parent identity, or (iii) no random effects. These models  
264 were fit via restricted maximum likelihood (REML) using the *lme* or *gls* functions (*gls* was  
265 used for models without random effects) in the package *nlme* (Pinheiro *et al.* 2010). To  
266 identify the best random effect structure (separately for each measure of offspring quality),  
267 we compared the nine models (3 random effect structures  $\times$  3 polynomials of parental age)  
268 using AIC<sub>c</sub>. We did not encounter any instances in which the ‘best’ random effect structure  
269 differed between the three polynomials of parental age for a given measure of offspring  
270 quality (i.e. selection of the best random effect structure was always unanimous).

271         Once the best random effect structure was established, we moved on to the fixed  
272 effects (parental age and growth chamber). In this portion of the analysis, models were fit  
273 via maximum likelihood (ML), again using either the *lme* or *gls* functions. Our approach  
274 here was to construct ‘full’ models describing each of the five measures of quality as a cubic  
275 function of parental age and linear function of growth chamber (with the appropriate  
276 random effect structure, as described above). We then compared all fixed-effect subsets of

277 each full model using the *dredge* function in the package *MuMIn* (Bartoń 2013) and AIC<sub>c</sub>  
278 values. Our all-subsets approach yielded eight models for each measure of offspring  
279 quality: three polynomials of parental age (either with or without a term for growth  
280 chamber), a growth chamber only model, and a null model with only an intercept.

281 We visually assessed model assumptions (independent, normally-distributed error  
282 with homogeneous variance) for each measure of offspring quality using standard  
283 diagnostic plots including quantile-quantile plots, histograms of model residuals,  
284 scatterplots of residuals versus fitted values, and scatterplots or histograms of residuals  
285 versus independent variables (including the random effect term for parent identity).

286 Diagnostic plots suggested that parametric assumptions were violated for the best model of  
287 intrinsic rate of increase (residuals were positively skewed). We therefore repeated the  
288 above-described protocol on log-transformed intrinsic rates of increase, which resulted in a  
289 best model that was more closely in line with parametric assumptions.

290

291

## 292 **Results**

### 293 PHASE ONE: SURVIVAL AND REPRODUCTION

294 We observed a significant decline in daily rates of survival with increasing frond age (Fig.  
295 1a). In particular, of the four candidate survival models that we examined, the three models  
296 in which survival rates declined with age received greater statistical support (i.e. had much  
297 lower AIC<sub>c</sub> values) than the exponential model which assumes a constant survival rate  
298 (Table 1). We also observed significant age-related declines in the daily probability of  
299 reproduction (Wald test,  $\chi^2 = 652.3$ ,  $df = 1$ ,  $P < 0.001$ ; Fig. 1b). Predicted daily probability of

300 reproduction from the fitted GEE declined from 0.65 at day one to 0.28 at day thirty. The  
301 Wald test and predicted probabilities of reproduction described above were based on a  
302 GEE with autoregressive (AR-1) correlation, which was selected as a more appropriate  
303 working correlation structure than ‘independence’ based on the RJ criteria (Table 2). The  
304 ‘exchangeable’ correlation structure was ruled out because it yielded an estimated  
305 covariance matrix that was not positive definite, potentially indicating a misspecified  
306 correlation structure (Crowder 1995; Schults *et al.* 2009). The estimate for the correlation  
307 parameter of the AR-1 model was  $-0.28 (\pm 0.02, SE)$ , indicating moderate within-subject  
308 negative temporal autocorrelation in reproduction.

309

#### 310 PHASE TWO: OFFSPRING QUALITY

311 There was a strong decline in our primary measure of offspring fitness, the intrinsic rate of  
312 increase, with increasing parental age (Fig. 1c). We also observed parental-age-related  
313 declines in three of our four secondary measures of offspring quality: total offspring  
314 produced, latency to first reproduction (this inverse measure of quality technically  
315 increased with parental age), and frond surface area (Fig. 2a,b,d). Lifespan, conversely, did  
316 not decline with increasing parental age (Fig. 2c).

317         The models of offspring quality selected as best (lowest  $AIC_c$ ) were in all cases non-  
318 linear with respect to parental age. Specifically, best models always described offspring  
319 quality as either a quadratic or cubic function of parental age (Table 3). Except for frond  
320 surface area, best models (or a close second-best model in the case of latency to  
321 reproduction,  $\Delta AIC_c = 0.1$ ) always included a term for growth chamber, suggesting that  
322 measures of offspring quality consistently differed among the three growth chambers that

323 we used (Table 3). Excepting latency to reproduction and lifespan, best models also always  
324 included random intercept and slope terms for parent identity, suggesting non-  
325 independence of offspring derived from the same parent (Table 3).

326

327

## 328 **Discussion**

329 We observed strong age-related declines in three demographic components of fitness in *L.*  
330 *minor*. Old plants had lower rates of survival and reproduction, and produced offspring of  
331 lower fitness than younger plants. While many species are known to experience age-  
332 related declines in at least one component of fitness, our study is to our knowledge the first  
333 to demonstrate simultaneous age-related declines in these three major demographic  
334 components of fitness, and also one of few studies to demonstrate age-related declines in a  
335 measure of offspring quality that closely approximates fitness (see also Gillespie *et al.*  
336 2013a). Of course, these results were obtained in a controlled, laboratory setting, so we  
337 encourage further research examining how the demographic patterns we identified  
338 manifest in the wild.

339

340

## 341 OFFSPRING QUALITY AND THE EVOLUTION OF SENESCENCE

342 Classic theories for the evolution of senescence implicitly assume that all offspring are of  
343 equal fitness, so that the action of natural selection depends only on age-specific rates of  
344 survival and reproduction (e.g. Williams 1957; Hamilton 1966; Kirkwood & Rose 1991).

345 Our results suggest that this assumption does not always hold, in which case selection may

346 depend additionally on age-specific trajectories of offspring fitness. Why would this  
347 matter? There are few theoretical results to guide us here, but a recent analysis by Gillespie  
348 *et al.* (2013b) suggests that birth-order-related declines in offspring fitness (similar in  
349 principle to parental-age-related declines) lead to steeper declines in the force of selection  
350 compared to what would be expected under classical models of senescence. In other words,  
351 not accounting for declining offspring fitness, where it occurs, may lead us to  
352 underestimate age-related declines in the force of selection. As many authors have argued,  
353 senescence, or more generally the action of selection, cannot be understood in terms of a  
354 single 'vital rate' or component of fitness (Partridge & Barton 1996; Caswell 2001; Nussey  
355 *et al.* 2008). We suggest, following Kern *et al.* (2001), that research on the evolutionary  
356 biology of senescence should devote attention to one extra vital rate – offspring quality.  
357 This often unrecognized component of fitness can clearly change with age, as we have  
358 shown in *L. minor*, and may be just as important in shaping overall fitness as survival and  
359 fecundity.

360

## 361 SENESCENCE IN PLANTS

362 Evolutionary theories of senescence suggest that age-related declines in fitness evolve  
363 because, for populations subject to nonzero mortality, the force of natural selection  
364 declines with age (Medawar 1952; Williams 1957; Hamilton 1966). Simply put, natural  
365 selection discounts old age-classes because relatively few individuals survive into old age,  
366 even in the absence of senescence. However, a number of authors have suggested that  
367 senescence should be relatively rare among vascular plants (Vaupel *et al.* 2004; Peñueles &  
368 Munné-Bosch 2010) or even that plants are predisposed to immortality (Silvertown,



369 Franco & Perez-Ishiwara 2001). Such views are based on unique aspects of the plant form  
370 and life history. For example, unlike other organisms that exhibit determinate growth,  
371 many vascular plants exhibit continual growth and regeneration via totipotent apical  
372 meristems (Roach 2001). This indeterminate growth pattern potentially allows for a  
373 continual increase in reproductive potential with age, which may translate into an increase  
374 in the force of natural selection with age (Vaupel *et al.* 2004).

375         Although some iteroparous plants (e.g. Herrera & Jovani 2010; Shefferson & Roach  
376 2013) and all semelparous plant species exhibit senescence, comparative studies to date  
377 have largely confirmed the predicted rarity of senescence among iteroparous vascular  
378 plants (Silvertown *et al.* 2001; Baudisch *et al.* 2013). Furthermore, a recent analysis by  
379 Caswell and Salguero-Gómez (2013) found that the force of selection does in fact increase  
380 with age for many iteroparous plants, especially within later stages of the plant life cycle.  
381 Why then is the iteroparous *L. minor* subject to senescent decline when its relatives within  
382 Plantae seem mostly immune? Unlike most vascular plants, *L. minor* has a unitary growth  
383 form and exhibits determinate growth at the level of individual fronds, which usually reach  
384 their full growth potential prior to detaching from their parent (Hillman 1961). This  
385 determinate growth pattern, combined with potentially high rates of extrinsic mortality  
386 due to herbivory and disease (Landolt 1986), likely leads to a decline in the force of natural  
387 selection with age, making *L. minor*'s age-related declines in fitness consistent with  
388 evolutionary theory.

389

390 PROXIMATE EXPLANATIONS FOR DECLINING OFFSPRING QUALITY

391 Age-related declines in fitness generally coincide with various forms of physiological  
392 deterioration or damage (Munné-Bosch 2007; Lindner *et al.* 2008; Monaghan 2010).  
393 Although our study did not specifically examine proximate explanations for senescence, we  
394 briefly touch on a potential explanation for age-related declines in offspring quality in *L.*  
395 *minor*. In general, age-related declines in offspring quality (including the Lansing effect) are  
396 thought to relate either to declines in parental care or provisioning of offspring (Fox 1993),  
397 the accumulation of mutations in parental reproductive tissue (Crow 1997), or the  
398 accumulation and transfer of deleterious compounds from parent to offspring (Ashby &  
399 Wangermann 1951). Of the three explanations above, mutation accumulation seems the  
400 least likely in this case given *L. minor*'s almost-exclusive asexual reproduction, which would  
401 render it subject to Muller's ratchet and mutation meltdown (Lynch *et al.* 1993). While we  
402 are not able to rule out the other two explanations in the case of *L. minor*, we suggest a  
403 another, non-exclusive possibility – that declining offspring quality in *L. minor* is caused by  
404 age-related structural changes in the environment in which fronds develop. In particular,  
405 Lemon and Posluszny (2000) found that when a daughter frond detaches from its parent, a  
406 small amount of connective tissue (deriving from a structure called the stipe) is left behind  
407 in the parent's meristematic pocket. They report, "after several daughter fronds have been  
408 produced, a large amount of stipe tissue will have accumulated in the pockets" (p. 743). We  
409 hypothesize that the accumulation of stipe tissue in the meristematic pockets of *L. minor*  
410 fronds progressively constricts or otherwise modifies the growth environment experienced  
411 by successive daughters, which may play a role in the age-related declines in offspring size  
412 and fitness observed in our study. This hypothesis yields a potentially testable prediction:

413 the artificial removal of accumulated stipe tissue should delay age-related declines in  
414 offspring size and/or fitness.

415

#### 416 SENESENCE IN LEMNA

417 Wangermann and Ashby (1950, 1951) documented parental-age-related declines in  
418 offspring size, lifespan, and lifetime reproductive output in *L. minor*, whereas Claus (1972)  
419 observed a slight increase in offspring lifespan and no change in lifetime reproductive  
420 output with increasing birth order (similar in principle to parental age). In Claus's study,  
421 birth order was confounded with other aspects of genealogy and there were very few  
422 plants representing the highest birth orders (i.e. greatest parental ages), so his results are  
423 difficult to interpret and we do not consider them further. Similar to Wangermann and  
424 Ashby, our results demonstrate age-related declines in offspring size and lifetime  
425 reproductive output, and we extend the results of Wangermann and Ashby in a manner  
426 relevant to evolutionary theories of senescence by specifically demonstrating age-related  
427 declines in offspring fitness (i.e. intrinsic rate of increase). We did not, however, observe  
428 declines in offspring lifespan with increasing parental age. One possible explanation for the  
429 conflicting results relates to how we defined death (i.e. the day that a frond's final daughter  
430 detached). It is not clear to us exactly how Wangermann and Ashby defined death, but they  
431 seem to have assessed death visually based on a loss of pigment. The difference between  
432 these two definitions of death might be considered the post-reproductive lifespan (i.e. the  
433 time between a final reproduction and the complete loss of pigment). If post-reproductive  
434 lifespans (but not reproductive lifespans) tend to decline with increasing parental age in *L.*

435 *minor*, we would expected to see age-related declines in offspring lifespan under  
436 Wangermann and Ashby's (presumed) definition of death, but not under our own.

437

## 438 CONCLUSIONS

439 We found that, in a controlled laboratory environment, *L. minor* fronds exhibited age-  
440 related declines in three major demographic components of fitness – survival, reproduction,  
441 and offspring fitness. Following Kern *et al.* (2001), we suggest that both theoretical and  
442 empirical research on the evolutionary biology of senescence should devote more attention  
443 to age-related changes in offspring quality. This often unrecognized component of fitness  
444 can clearly change with age, as we have shown in *L. minor*, and may be just as important in  
445 shaping overall fitness as survival and fecundity. Incorporating offspring quality into  
446 demographic and evolutionary analyses will no doubt be challenging. Indeed, determining  
447 the appropriate measure of fitness is difficult even when only the traditional fitness  
448 components – survival and fecundity – are considered (Link *et al.* 2002; Metcalf & Pavard  
449 2007). Nonetheless, we suggest that treating offspring quality as a component of fitness  
450 that may covary or trade-off with other fitness components, and be shaped by age-specific  
451 changes in the force of natural selection alongside other fitness components, may provide  
452 important insight into the evolutionary biology of senescence.

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454

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#### 464 **Data accessibility**

465 Data deposited in the Dryad repository: <http://doi.org/10.5061/dryad.t938n> (Barks &  
466 Laird 2014)

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630 **Supporting Information**

631 Additional supporting information may be found in the online version of this article:

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633 **Figure S1.** Images of non-curled and curled fronds.

634 **Figure S2.** Estimating the surface area of curled fronds.

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653 **Table 1.** Comparison of models describing age-specific rates of frond survival. The best

654 model is in bold.

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Model	Parameters	Deviance	AIC <sub>c</sub>	ΔAIC <sub>c</sub>	AIC <sub>c</sub> weight
<b>Logistic</b>	<b>3</b>	<b>1195.9</b>	<b>1202.0</b>	<b>0.0</b>	<b>0.99</b>
Weibull	2	1222.1	1226.2	24.2	<0.001
Gompertz	2	1258.5	1262.5	60.5	<0.001
Exponential (no senescence)	1	1808.1	1810.1	608.1	<0.001

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670 **Table 2.** Comparison of working correlation structures for GEE models describing age-  
 671 specific rates of reproduction. The ‘best’ working correlation structure (in bold) is the one  
 672 that yields values of RJ1 and RJ2 closest to 1, and a value of RJ3 closest to 0. Working  
 673 correlation structures that fail to yield a positive definite covariation matrix are ruled out.  
 674

Positive definite				
Working correlation structure	covariation matrix?	RJ1 ( $\bar{c}_1$ )	RJ2 ( $\bar{c}_2$ )	RJ3 ( $\bar{d}$ )
Independence	yes	0.25	0.07	0.58
<b>Autoregressive (AR-1)</b>	<b>yes</b>	<b>0.42</b>	<b>0.19</b>	<b>0.36</b>
Exchangeable	no	–	–	–

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688 **Table 3.** Comparison of models describing measures of offspring quality as functions of  
689 parental age (p.age) and growth chamber (chamb). For each measure of quality, the best  
690 model is in bold. Only the five best models are displayed for each measure of offspring  
691 quality.  
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Measure of offspring quality	Model <sup>a</sup>	df	Deviance	AIC <sub>C</sub>	ΔAIC <sub>C</sub>	AIC <sub>C</sub> weight
log (Intrinsic rate of increase) <sup>c</sup>	<b>p.age<sup>2</sup> + chamb</b>	<b>9</b>	<b>-192.8</b>	<b>-174.4</b>	<b>0</b>	<b>0.44</b>
	p.age <sup>2</sup>	7	-187.8	-173.6	0.9	0.29
	p.age <sup>3</sup> + chamb	10	-192.9	-172.5	2.0	0.17
	p.age <sup>3</sup>	8	-187.9	-171.7	2.8	0.11
	p.age <sup>1</sup>	6	-127.4	-115.2	59.2	<0.001
Total offspring <sup>c</sup>	<b>p.age<sup>3</sup> + chamb</b>	<b>10</b>	<b>1854.1</b>	<b>1874.5</b>	<b>0</b>	<b>0.83</b>
	p.age <sup>3</sup>	8	1861.4	1877.6	3.1	0.17
	p.age <sup>2</sup> + chamb	9	1871.0	1889.3	14.9	<0.001
	p.age <sup>2</sup>	7	1877.0	1891.2	16.7	<0.001
	p.age <sup>1</sup> + chamb	8	1963.0	1979.3	104.8	<0.001
Latency to reproduce <sup>b</sup>	<b>p.age<sup>2</sup></b>	<b>4</b>	<b>1502.4</b>	<b>1510.5</b>	<b>0</b>	<b>0.30</b>
	p.age <sup>2</sup> + chamb	6	1498.4	1510.6	0.1	0.28
	p.age <sup>3</sup>	5	1501.0	1511.1	0.6	0.22
	p.age <sup>3</sup> + chamb	7	1497.1	1511.3	0.8	0.20
	p.age <sup>1</sup>	3	1571.7	1577.8	67.3	<0.001
Lifespan <sup>b</sup>	<b>p.age<sup>3</sup> + chamb</b>	<b>7</b>	<b>3005.0</b>	<b>3019.2</b>	<b>0</b>	<b>0.95</b>
	p.age <sup>2</sup> + chamb	6	3013.0	3025.2	6.0	0.05
	chamb	4	3025.2	3033.3	14.1	0.001
	p.age <sup>1</sup> + chamb	5	3025.2	3035.3	16.1	<0.001
	p.age <sup>3</sup>	5	3033.2	3043.3	24.1	<0.001
Fronde surface area <sup>c</sup>	<b>p.age<sup>3</sup></b>	<b>8</b>	<b>970.2</b>	<b>986.5</b>	<b>0</b>	<b>0.67</b>
	p.age <sup>3</sup> + chamb	10	967.5	987.9	1.4	0.34
	p.age <sup>2</sup>	7	1002.0	1016.2	30.0	<0.001
	p.age <sup>2</sup> + chamb	9	998.4	1016.7	30.2	<0.001
	p.age <sup>1</sup>	6	1236.4	1248.5	262.0	<0.001

693 a. Numeric superscripts beside the parental age term (p.age) indicate polynomial degree. For example, p.age<sup>3</sup>  
694 indicates that the measure of offspring quality was modeled as a cubic function of parental age.

695 b. Models do not include random effects

696 c. Models include random intercept and slope terms for parent identity

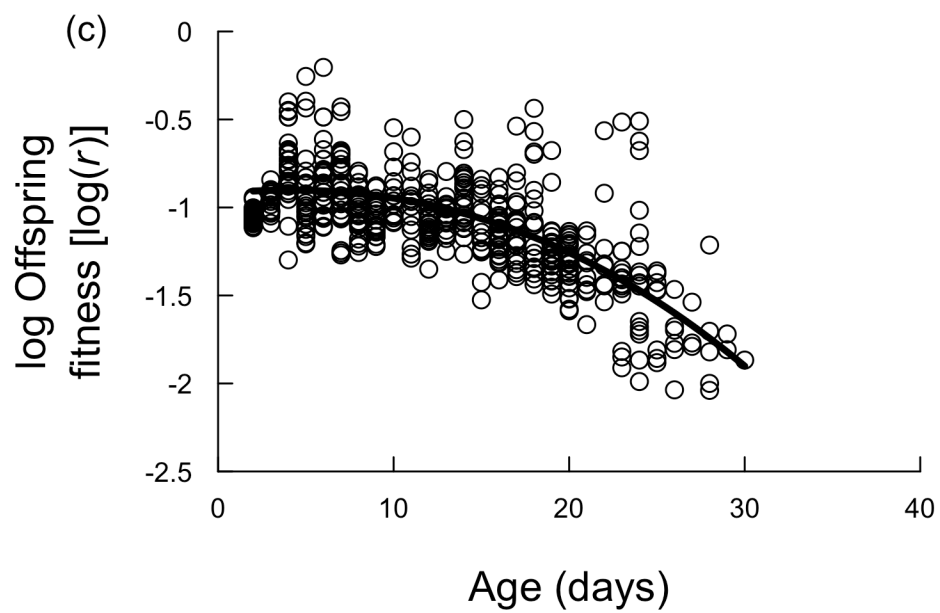
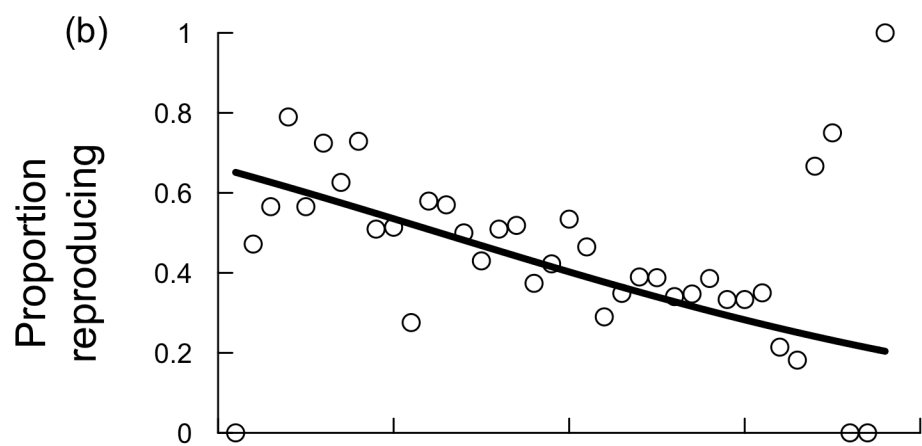
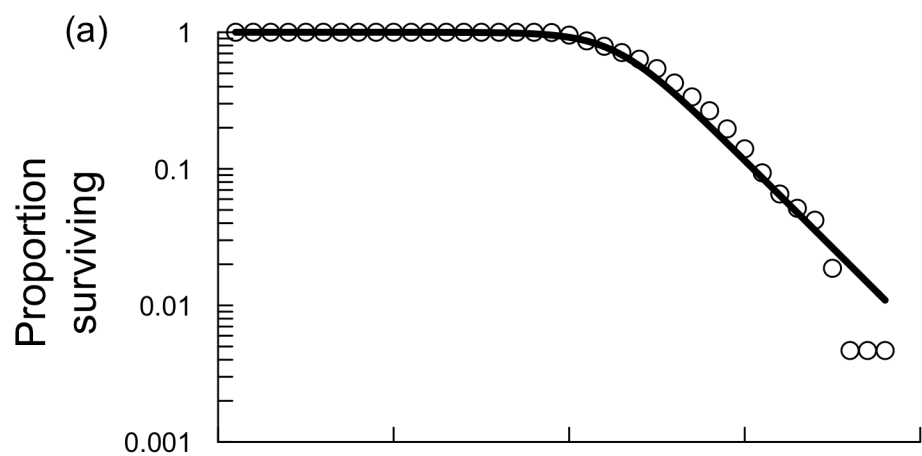
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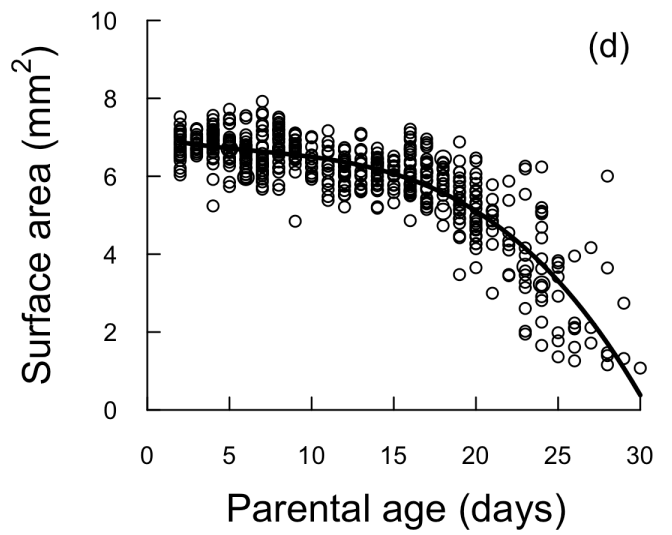
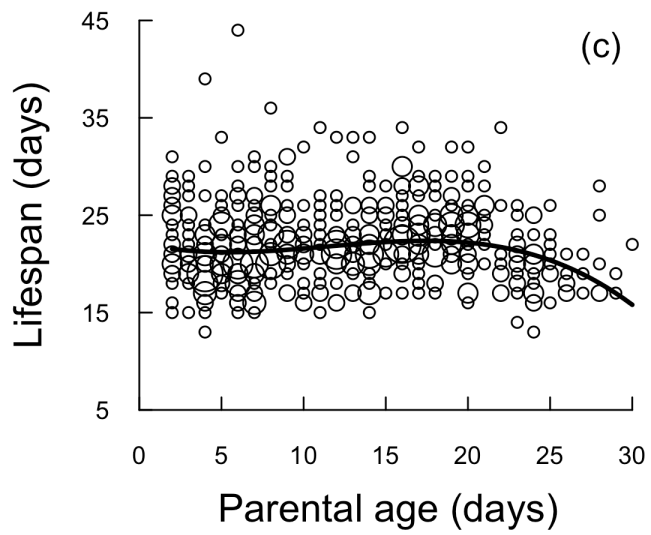
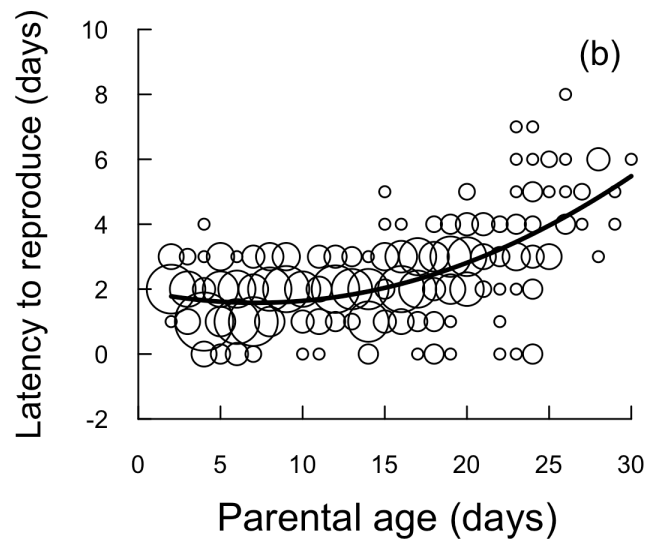
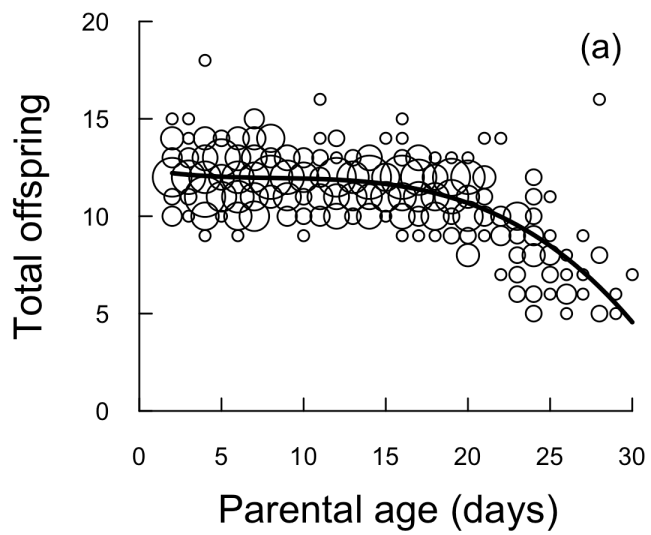
698 **Figure 1.** Age-related changes in rates of survival (a), rates of reproduction (b), and  
699 offspring fitness (c) in *L. minor*. Offspring fitness is measured as the log-transformed  
700 intrinsic rate of increase ( $r$ ), which has units of day<sup>-1</sup>. Best-fit models are described in the  
701 text and Tables 1-3. In semi-log survival plots such as in panel a, a population with constant  
702 survival rates (i.e. with no senescence) would appear as a straight line.

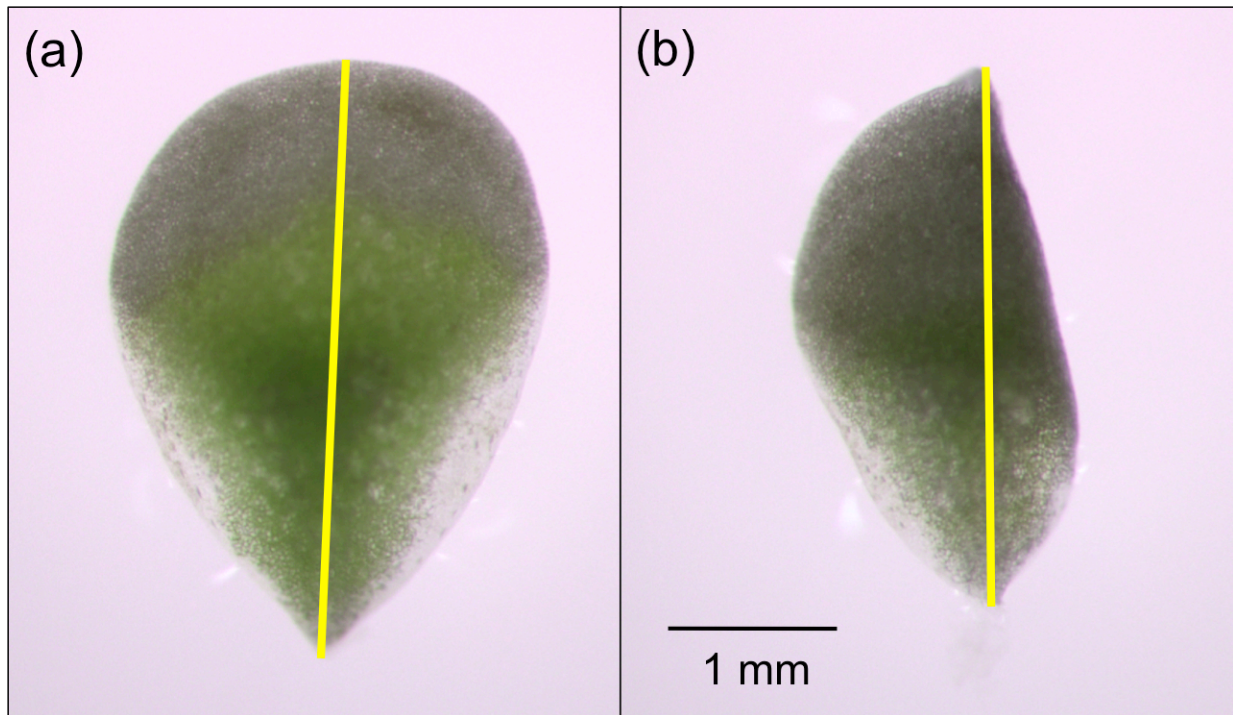
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704 **Figure 2.** Parental-age-related changes in secondary measures of offspring quality  
705 including total offspring produced (a), latency to first reproduction (inversely related to  
706 fitness; b), lifespan (c), and frond surface area (d). Point area is proportional to the number  
707 of observations at a given set of coordinates. Best-fit models are described in the text and  
708 Table 3.

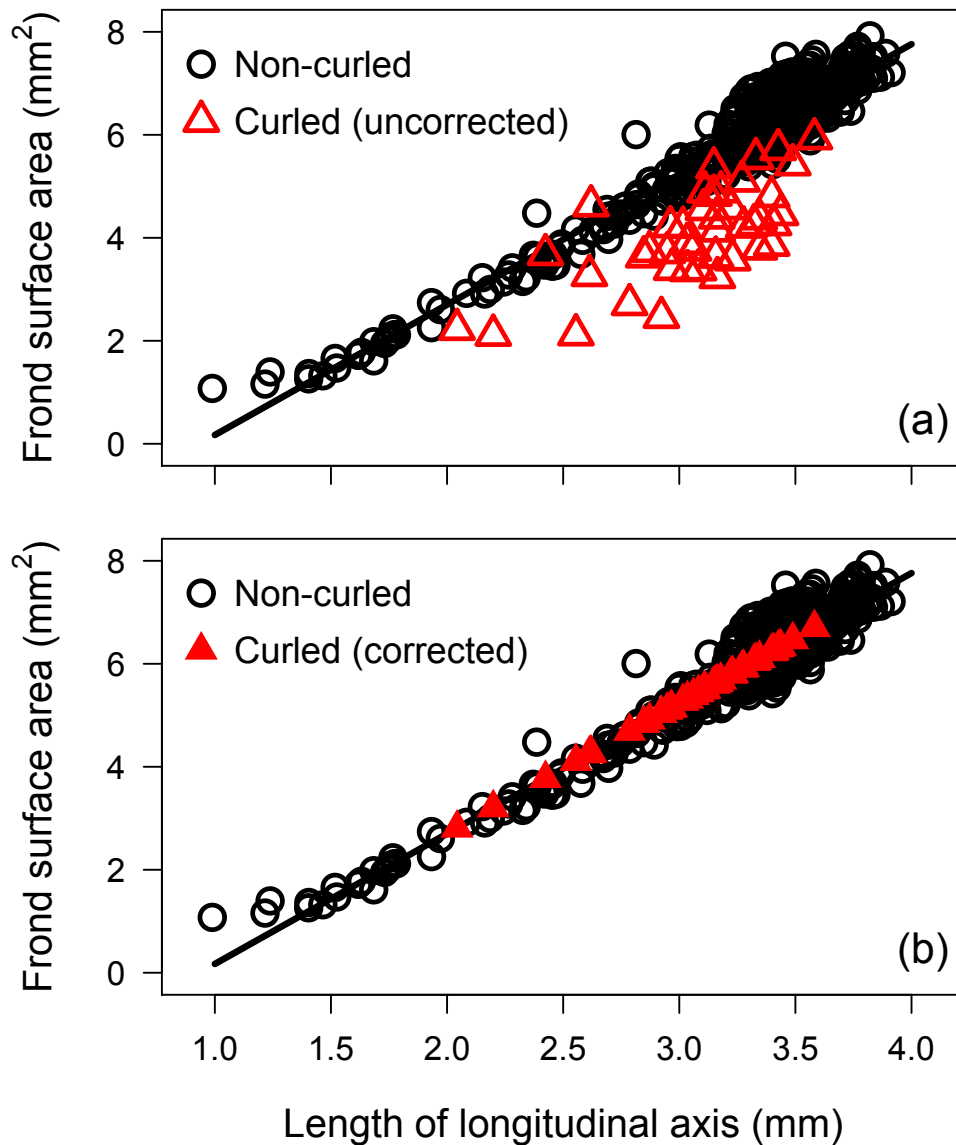








**Figure S1.** Comparison of non-curved (a) and curved (b) fronds of *Lemna minor*. Yellow lines correspond to each frond's longitudinal axis. We used the strong correlation between surface area and length of the longitudinal axis to estimate the surface area of the 42 (out of 542) fronds in *Phase two* of our study that were curved (see also Fig. S2). Note that the 1 mm scale bar applies to both panels.



**Figure S2.** Linear regression of frond surface area (mm<sup>2</sup>) versus length of the longitudinal axis (mm) for the 500 non-curved fronds (open black circles) in *Phase two* of our study. Uncorrected (open red triangles, panel a) and corrected (filled red triangles, panel b) surface areas of curled fronds are depicted for comparison. We ‘corrected’ estimates of surface area for curled fronds by interpolating from the regression line (which was fit using data from the 500 non-curved fronds).