

Dose-Response: An International Journal

Volume 10 | Issue 1

Article 11

3-2012

LANTHANUM ELEMENT INDUCED IMBALANCE OF MINERAL NUTRIENTS, HSP 70 PRODUCTION AND DNA-PROTEIN CROSSLINK, LEADING TO HORMETIC RESPONSE OF CELL CYCLE PROGRESSION IN ROOT TIPS OF VICIA FABA L. SEEDLINGS

Chengrun Wany

Huainan Normal University, Huainan, P.R. China

Cuie Shi

Huainan Normal University, Huainan, P.R. China

Ling Liu

Huainan Normal University, Huainan, P.R. China

Chen Wang

Huainan Normal University, Huainan, P.R. China

Recommended Citation

Wany, Chengrun; Shi, Cuie; Liu, Ling; Wang, Chen; Qiao, Wei; Gu, Zhimang; and Wang, Xiaorong (2012) "LANTHANUM ELEMENT INDUCED IMBALANCE OF MINERAL NUTRIENTS, HSP 70 PRODUCTION AND DNA-PROTEIN CROSSLINK, LEADING TO HORMETIC RESPONSE OF CELL CYCLE PROGRESSION IN ROOT TIPS OF VICIA FABA L. SEEDLINGS," *Dose-Response: An International Journal*: Vol. 10 : Iss. 1 , Article 11.

Available at: https://scholarworks.umass.edu/dose_response/vol10/iss1/11

This Article is brought to you for free and open access by ScholarWorks@UMass Amherst. It has been accepted for inclusion in Dose-Response: An International Journal by an authorized editor of ScholarWorks@UMass Amherst. For more information, please contact scholarworks@library.umass.edu.

Wei Qiao

Huainan Normal University, Huainan, P.R. China

See next page for additional authors

Follow this and additional works at: https://scholarworks.umass.edu/dose_response

LANTHANUM ELEMENT INDUCED IMBALANCE OF
MINERAL NUTRIENTS, HSP 70 PRODUCTION AND DNA-
PROTEIN CROSSLINK, LEADING TO HORMETIC RESPONSE
OF CELL CYCLE PROGRESSION IN ROOT TIPS OF VICIA FABA
L. SEEDLINGS

Authors

Chengrun Wany, Cuie Shi, Ling Liu, Chen Wang, Wei Qiao, Zhimang Gu, and Xiaorong Wang

LANTHANUM ELEMENT INDUCED IMBALANCE OF MINERAL NUTRIENTS, HSP 70 PRODUCTION AND DNA-PROTEIN CROSSLINK, LEADING TO HORMETIC RESPONSE OF CELL CYCLE PROGRESSION IN ROOT TIPS OF *VICIA FABA* L. SEEDLINGS

Chengrun Wang, Cuie Shi, Ling Liu, Chen Wang, and Wei Qiao □ Huainan Normal University, Huainan, P.R. China

Zhimang Gu □ University of Missouri, Columbia

Xiaorong Wang □ School of the Environment, Nanjing University, Nanjing, P.R. China

□ The effects and mechanisms of rare earth elements on plant growth have not been extensively characterized. In the current study, *Vicia faba* L. seedlings were cultivated in lanthanum (La)-containing solutions for 10 days to investigate the possible effects and mechanisms of La on cell proliferation and root lengthening in roots. The results showed that increasing La levels resulted in abnormal calcium (Ca), Ferrum (Fe) or Potassium (K) contents in the roots. Flow cytometry analysis revealed G1/S and S/G2 arrests in response to La treatments in the root tips. Heat shock protein 70 (HSP 70) production showed a U-shaped dose response to increasing La levels. Consistent with its role in cell cycle regulation, HSP 70 fluctuated in parallel with the S-phase ratios and proliferation index. Furthermore, DNA-protein crosslinks (DPCs) enhanced at higher La concentrations, perhaps involved in blocking cell progression. Taken together, these data provide important insights into the hormetic effects and mechanisms of REE(s) on plant cell proliferation and growth.

Keywords: Cell cycle phase; Hormetic Effect; DNA-Protein Crosslink (DPC); Heat Shock Protein 70 (HSP 70); Mineral Nutrients; Rare Earth Elements (REEs)

INTRODUCTION

Rare earth elements (REEs) include 15 lanthanide elements in group III A of the Periodic Table, exhibiting similar chemical and physical properties. In addition, yttrium and scandium have also been considered as REEs. Owing to their desirable properties and abundant sources in China, REE-based microfertilizers or additives have been utilized to enhance yield and quality of crops and vegetables (Hong *et al.* 2000; Wang *et al.* 2007).

Hormesis refers to concentration–response occurrence in various organisms, including numerous plant traits (Calabrese and Blain 2008).

Address correspondence to Chengrun Wang, Ph.D., Associated Professor, Xueyuan Road, Huainan, Anhui Province, China; 232001; Tel./fax: +86-554-6663083; E-mail: chengrunw66@yahoo.com.cn.

Hormetic effects of rare earth element on plant cell cycles

Hormesis is usually represented by two types of curves, including J-shaped curve showing a decrease below the control at low doses followed by an increase at higher doses, and inverted U-shaped curve showing an increase above the control at low doses followed by a decrease at higher doses (Calabrese and Baldwin 2003). The J-shaped curve is often followed immediately by an inverted U-shaped curve at a specific range of doses, together constituting so-called biphasic dose–response curves. Hormetic dose–response curves are widely used to determine the efficacy and hazard of pollutants (Qin *et al.* 2010). However, the underlying mechanism is not fully understood.

REEs are mainly taken up by roots and transported to other parts of plants, potentially contributing to hormetic effects in various biological parameters (Ouyang *et al.* 2003). REE-dependent stimulatory effects on cell proliferation and division as well as plant growth have been previously documented (Hagenbeek *et al.* 2000; Wu *et al.* 2001; Hong *et al.* 2003; Dai *et al.* 2008). However, REEs have also been shown to displace biometals in membrane metallobiomolecules, metallic proteins and enzymes, leading to mineral nutrient imbalance, membrane permeability, altered conformation in biomolecules, and ultimately functional disorders (Qiu *et al.* 2005; Zeng *et al.* 2006). For example, REEs appeared to regulate plant growth by affecting distribution and contents of mineral elements such as Ca, Fe, Cu, K, P, and Mg in some plants (Hu *et al.* 2004; Wang *et al.* 2008). In addition, cell cycle progression was further proved to be altered with lanthanum (La) treatment in root tips of *Vicia faba* L. seedlings (Wang *et al.* 2011). However, the mechanisms for REE(s)-regulated cell cycle progression in plants remain unclear.

Generally, root lengthening correlates with apical meristem activity and cell extension. In this study, roots of *V. faba* L. seedlings were employed as the experimental system to further explore the potential effects and mechanisms of nutrient imbalance, heat shock protein 70 (HSP 70), and DNA-protein crosslinks (DPCs) on cell cycle progression in the root tips exposed to varying concentrations of La³⁺. Our results clearly demonstrated that lanthanum exposure led to abnormal accumulation of mineral elements, HSP 70 and DNA-protein crosslinks, which might be responsible for the alteration in cell cycle phases and root lengths of *V. faba* L. seedlings.

MATERIALS AND METHODS

Plant Materials and La³⁺ Treatment

Seeds of *V. faba* L. were surface sterilized with 0.1% (m/v) sodium hypochlorite solution for 10 min and rinsed thoroughly in distilled water. Subsequently, the seeds were sown in moist fine gravel in plastic cylinders with perforated bottom at 23°C. Eight uniformly germinated seeds with

C. Wang and others

nearly 2 cm of root length were selected and transplanted into 1.2-L containers filled with equal Hoagland solution prepared according to methods of Lucretti *et al.* (1999). After a day of pretreatment, lanthanum nitrate ($\text{La}(\text{NO}_3)_3$) was added to the containers and final concentrations of extraneous La were 0, 0.125, 0.25, 0.5, 1, 2, 4, 8 and 16 mg L^{-1} , respectively. The seedlings were hydroponically cultivated in these solutions with daily aeration. The solutions were renewed every two days and pH values were measured within 6.3-6.5 in the fresh solutions. The containers were placed in a growth chamber with day/night air temperatures of 23/20°C and relative humidity of 75%. Plants were provided light 15 h a day with photosynthetically active radiation (PAR) of 230 $\mu\text{mol m}^{-2} \text{sec}^{-1}$. Three containers were prepared in each treatment in two independent experiments. Roots were harvested for chemical analysis and biological measurements immediately after 10 days of the treatment.

Measurement of Shoot Heights and Root Lengths

Lengths between apical buds and basal stems were measured as heights of the seedlings, and lengths between basal stems and primary root tips as root lengths. Eight seedlings were measured in each container, and three containers were prepared in each treatment.

Contents of La and Other Metals

Fresh roots were consecutively rinsed with 1 M HCl and distilled water. Digestion of samples was performed according to previous protocol (Wang *et al.* 2010). La, calcium (Ca), Ferrum (Fe) or Potassium (K) content was detected by inductively coupled plasma optical emission spectrometer (ICP-OES) and expressed as $\mu\text{g g}^{-1}$ dry weight (DW). Certified standard samples (GBW07429) and triplicates of all samples were used to ensure accuracy and precision. The detection limit of 0.02 $\mu\text{g L}^{-1}$ for this method was applied for all results.

Cell Cycle Progression and Proliferation Index in Root Tips

Approximately 50 fresh root tips (5 mm long) were cut from each control and treatment, and immediately fixed in 4% (v/v) formaldehyde in Galbraith buffer (Galbraith *et al.* 1983), supplemented with 1% (m/v) polyvinylpyrrolidone and 10 mM sodium metabisulfite (pH 7.0) for 30 min at 4°C. Samples were then washed thoroughly in pre-chilled Galbraith buffer for 10 min. Nuclei of fixed root tips were isolated using 1mL of the buffer by crushing them with a glass rod in Petri dishes on ice. The dishes were kept tilted on ice to yield efficient collection of isolated nuclei in the buffer.

Nuclear suspensions were filtered through nylon mesh of 25 μm pore size to remove gross particles and centrifuged at 700×*g* for 10 min at 4°C. The supernatants were removed and the pellets resuspended in 0.75 M

Hormetic effects of rare earth element on plant cell cycles

hexanediol and stored at 4°C. The pelleted nuclei were incubated with propidium iodide (PI)/RNase Staining Buffer (BD Pharmingen™) in the dark for 2 h at room temperature. Cell cycle progression was characterized by flow cytometry (FACSCalibur, BD, USA) at 488 nm excitation and 525 nm emission. A total of 10,000 nuclei were measured for each sample; triplicate experiments were conducted for each treatment giving rise to similar results. Peaks at channel 200 and 400 denoted G₀/G₁ and G₂/M phase, respectively; and the area in between corresponded to the S-phase. The G₀/G₁, S and G₂/M phase ratios were calculated with the equipped software according to DNA distribution. Proliferation index was calculated by formula (S-phase ratio + G₂/M-phase ratio) / (G₀/G₁-phase ratio + S-phase ratio + G₂/M-phase ratio) × 100%.

Determination of DNA-Protein Crosslink (DPC)

DPC formation was monitored based on fractionation of protein-bound and free DNA by potassium-SDS precipitation as described by Costa *et al.* (1996). Briefly, nuclei in root tips were isolated and purified as described by Gichner *et al.* (2004). The pelleted nuclei were lysed in 0.5% (m/v) SDS, and 0.5 ml of 100 mM KCl and 20 mM Tris (pH 7.5) were added to each tube. After 10 min incubation at 65°C, the tubes were placed on ice for 5 min, and then centrifuged at 6000×g for 5 min at 4°C. The supernatant was collected together by plastic tip pipette for quantification of free DNA contents in each treatment. The pellets were gently resuspended in 100 mM of KCl, 20 mM Tris-HCl (pH 7.5) and incubated at 65°C for 10 min. Crosslinked DNA was released from the final K⁺-SDS precipitates by treatment of 0.2 mg ml⁻¹ proteinase K (Amersham) at 50°C for 3 h. The tubes were placed on ice for 5 min and centrifuged at 12,000×g for 10 min at 4°C. The supernatant was collected to determine the content of crosslinked DNA using DNA standards (Sigma Chemical) prepared at concentrations of 100, 200, 500, 1000, 2000 and 5000 ng mL⁻¹. Two ml of standard DNA, free DNA or crosslinked DNA was mixed with 1 ml of 500 ng mL⁻¹ of freshly prepared Hoechst 33258 (Sigma Chemical), respectively. The mixtures were placed in the dark for 30 min. Fluorescence intensity was determined by excitation at 350 nm and emission at 450 nm using Shimadzu RF-5301PC Spectrofluorometer.

HSP 70 Western Blotting Analysis

Crude extracts in root tips were prepared according to a method described by Romero-Puertas *et al.* (2004). Soluble protein content was determined by method of Bradford (1976) with BSA as standard. All operations were performed at 4°C. Sodium dodecylsulfonate polyacrylate gel electrophoresis (SDS-PAGE) and Western blotting of HSP 70 were conducted according to Wang *et al.* (2010) with minor modification. Total pro-

C. Wang and others

tein extract was mixed with lysis buffer (0.5 M Tris, pH 6.8, 20% (v/v) glycerol, 3% (m/v) SDS, 0.01% (m/v) bromophenol blue, 10% (v/v) β -mercaptoethanol), boiled for 4 min, and cooled on ice. Twenty six μ g of total proteins per lane, together with PageRuler™ prestained protein ladder (Fermentas), were separated by 10% (m/v) SDS-PAGE. After electrophoresis, gels were either stained with Coomassie brilliant blue R-250 or transferred onto polyvinylidene fluoride membrane (Amersham). The membranes were blocked with 10% (m/v) non-fat milk/TBST buffer (50 mM Tris, 150 mM NaCl, pH 7.5, containing 0.05% (v/v) Tween-20) for 2 h. After washing, mouse anti-HSP 70/HSC 70 monoclonal antibody (SPA820, Stressgen Corp.) (diluted 1: 5000) was added and incubated overnight at 4°C. After washing, the membranes were incubated in goat anti-mouse IgG conjugated with horseradish peroxidase (Stressgen Corp.) (diluted 1: 25,000) at room temperature for 1.5 h. Bands were visualized by SuperSignal West Femto Maximum Sensitivity Substrate (Thermo Scientific) and exposed to X-ray film. Integrated densities in bands were quantificated by Image J software. The experiment was repeated 3 times.

Statistical Analysis

All statistical analyses were performed using SPSS 13.0 for Windows (SPSS, Chicago IL, USA). The data were presented as mean \pm standard deviations of three replicates. Difference was considered as significant at $p < 0.05$ (*) and highly significant at $p < 0.01$ (**) using one-way ANOVA by Dunnett's *t*-test. Representative photographs from each treatment were presented.

RESULTS

Contents of La, Ca, Fe and K Elements in Roots

ICP-OES was employed to monitor changes of La and mineral elements such as Ca, Fe or K in the roots. The results showed that contents of La in the roots increased with elevated extraneous La (Fig. 1A), which significantly enriched when the extraneous La was more than 1 mg L⁻¹. Contents of Ca, Fe or K were initially reduced before rebounding with increased levels of La, displaying a characteristic U-shaped curve. Particularly, Ca or Fe contents were significantly enhanced at 16 mg L⁻¹ of extraneous La. Generally, Low concentrations of La resulted in decreased contents of Ca, Fe or K, whereas higher concentrations led to opposite effects (Fig. 1, B-D).

Alteration in Cell Cycle Progression and Proliferation Index

Cell cycle phases and relative nuclear DNA contents were determined by flow cytometric analysis of nuclei released from the root tips (Fig. 2A).

Hormetic effects of rare earth element on plant cell cycles

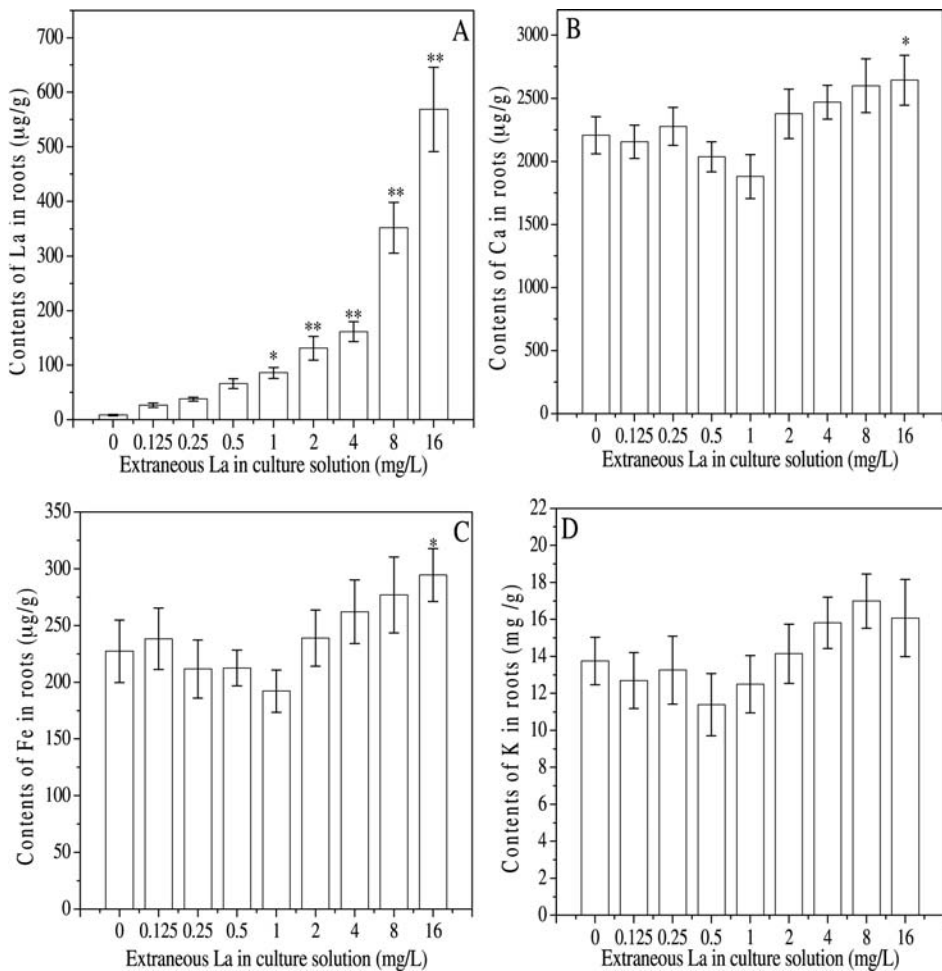


FIG. 1. La (A), Ca (B), Fe (C), and K (D) contents in roots of *V. faba* seedlings cultivated in 0-16 mg L⁻¹ of extraneous La in culture solution for 10 days. Values are denoted as mean ± SD, n=3. Significant differences from the controls are indicated as follows: * $p < 0.05$, ** $p < 0.01$.

The ratios of G0/G1, S or G2/M phase displayed a biphasic curve in response to increasing levels of La. The G0/G1 phase ratios mirrored that of the G2/M phase, showing initial enhancement at 0-0.5 mg L⁻¹, followed by a steady decline from 0.5 to 4 mg L⁻¹ before the rebound. The S phase ratios, on the other hand, exhibited an essentially opposite pattern (Fig. 2B). Therefore, the cell cycles were likely arrested at G1/S and/or S/G2 interphases by La³⁺ in the root tip cells. Furthermore, the proliferation index displayed a pattern closely in parallel with the S-phase ratio, with initial reduction at 0-0.5 mg L⁻¹, followed by enhancement from 0.5 to 4 mg L⁻¹ and decline thereafter (Fig. 2 C). However, no significant elevation or reduction was observed in cell cycle phase ratios and proliferation index (Fig. 2 B, C).

C. Wang and others

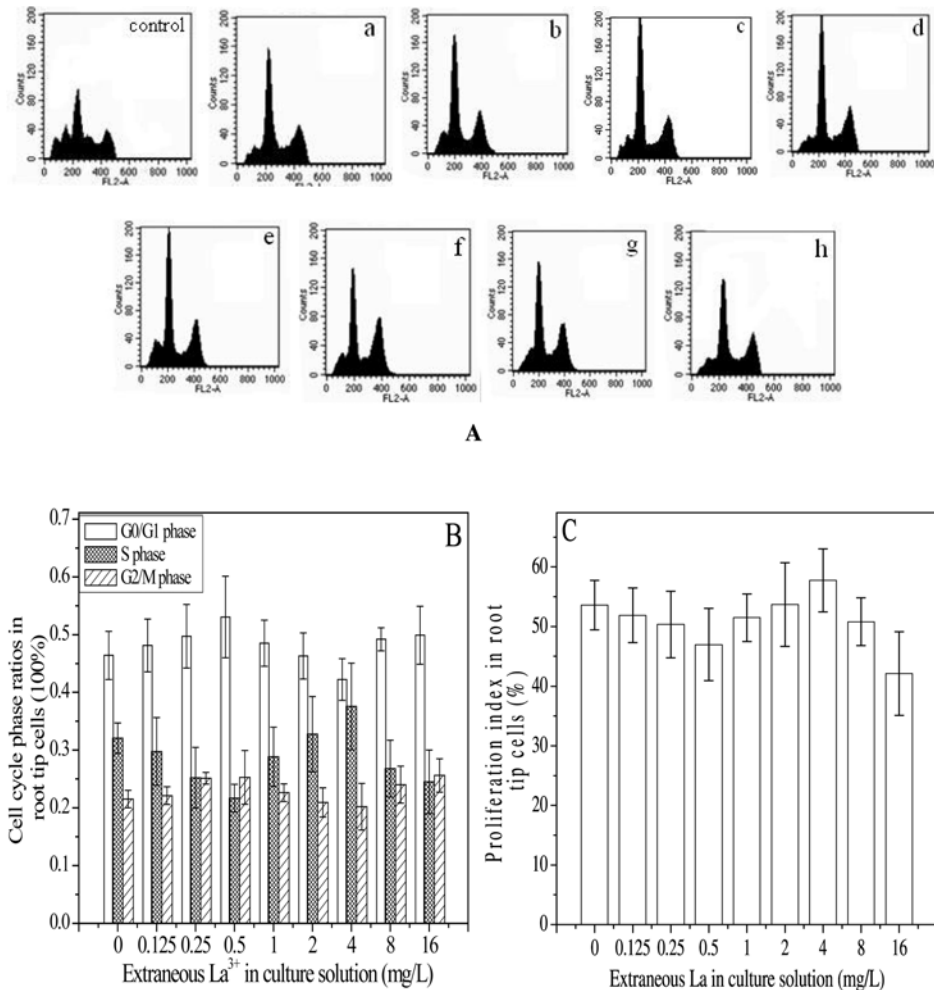


FIG. 2. Histograms of relative nuclear DNA contents and cell cycles (A) obtained by flow cytometric analysis of nuclei released from root tips of *V. faba* seedlings cultivated in 0-16 mg L⁻¹ of extraneous La for 10 days. B: ratios of cell cycle phases; C: proliferation indexes in the root tips. Control and a-h represents 0, 0.125, 0.25, 0.5, 1, 2, 4, 8 and 16 mg L⁻¹ of extraneous La, respectively. Peaks at channels 200 and 400 denote G0/G1 and G2/M phase, respectively, and the area between the two channels denotes S phase. Ratios of G0/G1, S and G2/M phases are shown in B. Proliferation indexes are shown in C. Values are denoted as mean \pm SD, $n=3$.

In addition, root lengths were found to be positively correlated with the S-phase ratios ($r = 0.829$, $p < 0.05$) and proliferation index ($r = 0.929$, $p < 0.01$), but negatively with the G0/G1-phase ($r = -0.741$, $p < 0.05$) or G2/M-phase ratios ($r = -0.899$, $p < 0.01$) in the root tips, respectively.

Changes of DPC and Seedling Growth

DPCs were involved in DNA replication and repair, which may interfere with cell proliferation. In the roots, contents of DPCs appeared to

Homeotic effects of rare earth element on plant cell cycles

decrease at 0-0.5 mg L⁻¹ of extraneous La, and then increased with the increasing La, which significantly elevated when the extraneous La was more than 4 mg L⁻¹ (Fig. 3A). In addition, the contents of DPCs were well correlated with the S-phase ratios ($r = 0.808$, $p < 0.05$) and proliferation index ($r = 0.823$, $p < 0.01$) at 0-4 mg L⁻¹ of extraneous La, respectively.

Moreover, root length fluctuated in parallel with the proliferation index, showing decreases at 0.125-0.5 mg L⁻¹ followed by enhancement from 0.5 to 4 mg L⁻¹ and subsequent decline (Fig. 3B).

Induction of HSP 70 Production

HSP 70 participates in many physiological activities, including cell proliferation. HSP 70 production in the roots was investigated by Western blotting method. The results showed that the relative HSP 70 levels decreased at 0-0.5 mg L⁻¹ of extraneous La, and enhanced with the increasing La, showing a biphasic dose response. In addition, the HSP 70 levels were significantly elevated at 4-8 mg L⁻¹ (Fig. 4).

DISCUSSION

REEs mainly accumulate in plant roots as heavy metals. The content of La in the roots increased with elevated La concentrations, contributing to imbalance of Ca, Fe or K contents in the roots. The varying doses of extraneous La were responsible for the U-shaped dose response of mineral metal contents in the roots. Similar findings have also been reported for Cd on tomato plants. Low dose of Cd²⁺ caused a moderate Fe deficiency, whereas higher Cd²⁺ abolished any nutrient interaction due to excess Cd

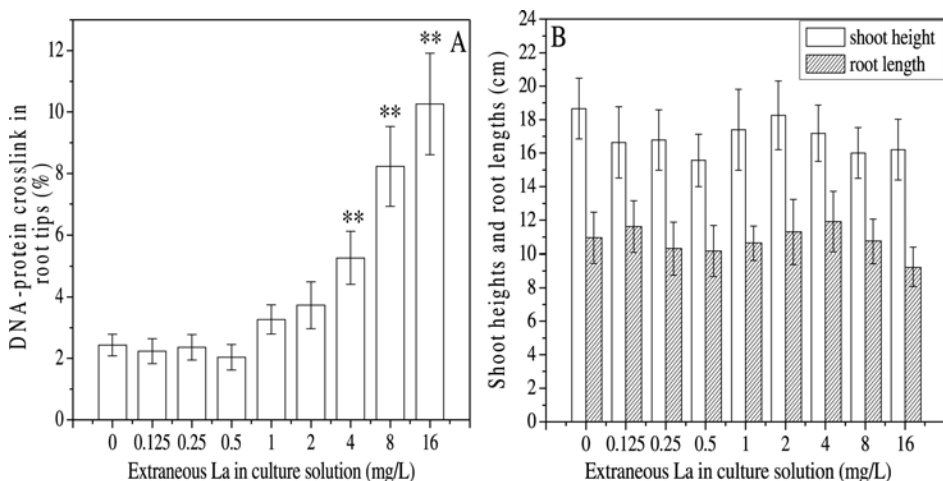


FIG. 3. Contents of DNA-protein crosslinks in nuclei separated from the root tips (A), and shoot heights and root lengths (B) of *V. faba* seedlings cultivated in 0-16 mg L⁻¹ of extraneous La for 10 days. Values are denoted as mean \pm SD, $n=3$. Significant differences from the controls are indicated as follows: ** $p < 0.01$.

C. Wang and others

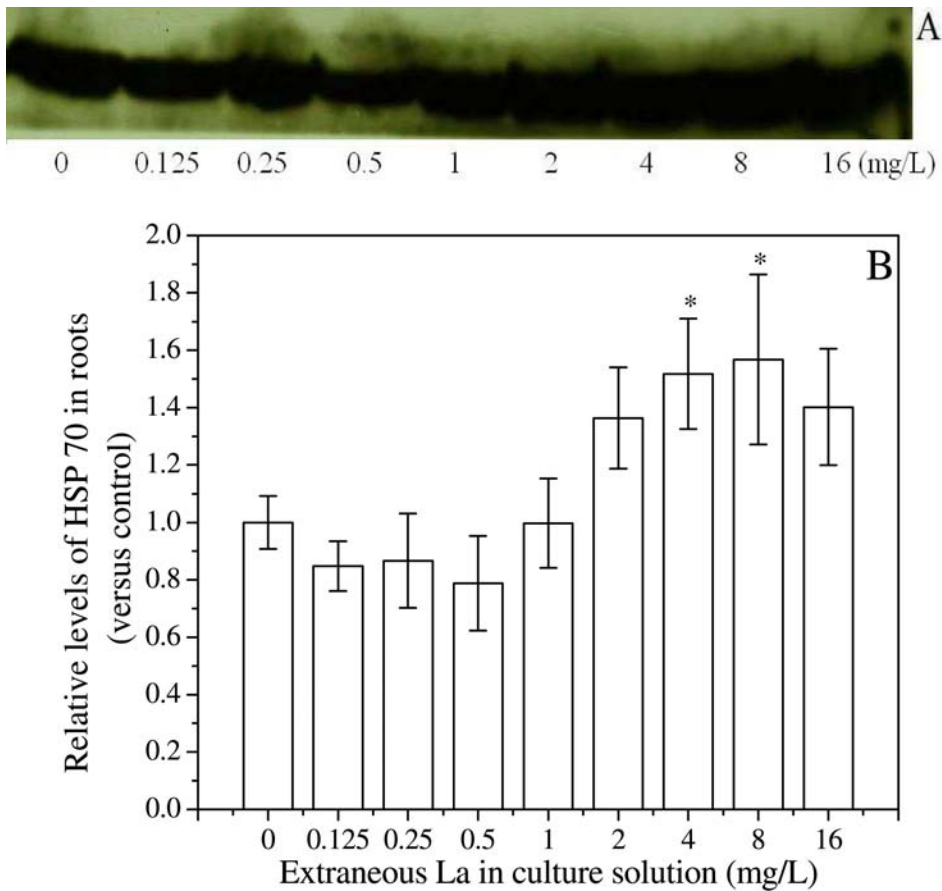


FIG. 4. Western blotting and relative levels of HSP 70 in roots of *V. faba* seedlings exposed to 0-16 mg L⁻¹ of extraneous La for 10 days. Twenty six μ g of total protein loaded in each lane was separated by SDS-PAGE and transferred to PVDF membrane. A, Western blotting of HSP 70; B, relative levels of HSP 70 normalized to controls. Values are denoted as mean \pm SD, $n=3$. Significant differences from the controls are indicated as follows: * $p < 0.05$.

(López-Millán *et al.* 2009). Thus, REEs likely have similar properties in altering the balance of metal contents in plant tissues as heavy metals.

Ca uptake has been shown to be completely inhibited by a voltage-dependent Ca channel antagonist such as La in plants (Tammeba *et al.* 2004). In fact, La³⁺ can bind to surface-localized Ca²⁺ absorption sites with lower degrees of reversibility than Ca²⁺, thus blocking Ca²⁺ channels and disrupting the uptake of nutrient ions through Ca²⁺ channels (Hu *et al.* 2006). In the current study, increasing La induced a U-shaped dose response of Ca, Fe or K in the roots (Fig. 1), suggesting that low concentrations of La contributed to the reduction of these metals. Thus, lower concentrations of La appeared to function as Ca antagonist in the roots. Meanwhile, the U-shaped dose response curve of Ca content also indicated that higher La increased Ca contents in the roots. La at higher doses

Homeotic effects of rare earth element on plant cell cycles

might inactivate the Ca channels in the membrane and promote the entry of metals such as Ca, Fe and K into the roots. However, La at lower concentrations also led to decreased contents of Fe and K. Whether La at low doses may act as an antagonist of Fe or K needs to be further investigated.

Mitotic activity, cell cycle progression and cell expansion are major factors involved in cell growth (Kononowig *et al.* 1992). Mineral elements such as Ca, Fe, K, Cu, Mg, etc are known to be essential nutrients for plant development. Absence or deficiency of these elements may have adverse effects on plant growth. In the present study, the concentrations of La resulted in the imbalance of mineral nutrients in the roots. Previous studies also showed that REE (s) regulated plant growth by affecting the distribution of mineral elements in crop seedlings (Wang *et al.* 2008, Wang *et al.* 2011). Likewise, the U-shaped fluctuation of mineral element contents is likely to affect cell proliferation and extension in the roots and shoots. Therefore, cell cycle progression and root lengthening may be altered due to nutrient imbalance in the roots subjected to La treatment.

The U-shaped dose response of HSP 70 parallels the patterns of mineral nutrients, S-phase ratios and proliferation index, but was opposite to those of the G0/G1-phase and G2/M-phase ratios in the root tips. Under normal conditions, Ca binds calmodulin and initiates a cascade of signal transduction events including HSP 70 induction. HSP 70 mRNA levels are rapidly elevated at early S phase, but subsequently decreased at late S and G2 phases (Milarski and Morimoto, 1986). In nuclei, the spatial and temporal association of HSP 70 and CaM has been shown to regulate cell cycle progression (Huang *et al.* 2009). HSP 70 proteins were found to be associated with the spindle, thus functioning in cell cycle progression (Vargas-Roig *et al.* 1997). Furthermore, HSP 70 appeared to positively correlate with cell cycle progression in human cells (Karlseder *et al.* 1996; Kim *et al.* 2006) and plant cells (Giménez-Abián *et al.* 2004). In addition, HSP 70 overexpression was particularly pronounced during S phase, leading to enlarged S-phase populations (Helmbrecht and Rensing, 1999). However, whether HSP 70 function in cell cycle progression of plants is not fully understood.

Our results suggest that the decreased HSP 70 levels may be involved in the retardation of root tip cell proliferation, since increased G0/G1- and G2/M-phase ratios and decreased S-phase ratios were observed at the 0-0.5 mg L⁻¹ dose range. On the other hand, the enhanced HSP 70 at 1-4 mg L⁻¹ may stimulate proliferation by decreasing G0/G1 and G2/M populations accompanied by increased S-phase ratios in the root tips. Therefore, the La-induced HSP 70 may play a role in the control of cell cycle progression in the root tips at 0-4 mg L⁻¹ of extraneous La. Similar findings have also been reported in human lung cells (Bromberg *et al.* 2008)

DPCs are formed when proteins become covalently bound to DNA, and serve as bulky helix-distorting adducts to physically block DNA repli-

C. Wang and others

cation, repair and transcription (Barker *et al.* 2005). Thus, DPCs may interfere with cell proliferation and extension. In the present study, the DPC content was found to increase with the increase of extraneous La from 0.5 to 16 mg L⁻¹. DPCs enrichment may be involved in the decline of the S-phase ratio at more than 4 mg L⁻¹ La, leading to reduced proliferation index and root length.

Finally, our results suggest that the imbalance of mineral nutrients was involved in the induction of HSP 70, which in turn were associated with the cell cycle control. Together, the results in the current study provide useful insights into the functional mechanisms of hormetic effects of La³⁺ on plant cell proliferation and growth.

ACKNOWLEDGMENTS

We acknowledge supports from the National Natural Science Foundations of China (grant No. 20877032) and the Foundation of State Key Laboratory of Pollution Control and Resources Reuse of China (grant No. PCRRF08011). We also thank Ling Wang and Fan Hu in Center of Medical Analysis of Nanjing Medical University for cell cycle analysis in *V. faba* seedlings by flow cytometry.

REFERENCES

- Barker S, Weinfeld M, and Murray D. 2005. DNA–protein crosslinks: their induction, repair, and biological consequences. *Mutat. Res.* 589: 111–135
- Bradford MM. 1976. A rapid and sensitive method for the quantification of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal. Biochem.* 72: 248–254
- Bromberg Z, Raj N, Goloubinoff P, Deutschman CS, and Weiss YG. 2008. Enhanced expression of 70-kilodalton heat shock protein limits cell division in a sepsis-induced model of acute respiratory distress syndrome. *Crit. Care Med.* 36: 246–255
- Calabrese EJ and Baldwin LA. 2003. Toxicology rethinks its central belief: Hormesis demands a reappraisal of the way risks are assessed. *Nature* 421: 691–69
- Calabrese EJ and Blain RB. 2008. Hormesis and plant biology. *Environ. Pollut.* 157: 42–48
- Costa M, Zhitkovich A, Gargas M, Paustenbach D, Finley B, Kuykendall J, Billings R, Carlson TJ, Wetterhahn K, Xu J, Patierno S, and Bogdanffy M. 1996. Interlaboratory validation of a new assay for DNA-protein crosslinks. *Mutat. Res.* 369: 13–21
- Dai J, Zhang YZ, and Liu Y. 2008. Microcalorimetric investigation on metabolic activity and effects of La (III) in mitochondria isolated from Indica rice 9311. *Biol. Trace Elem. Res.* 121: 60–68
- Galbraith DW, Harkins KR, Maddox JR, Ayres NM, Sharma DP, and Firoozabady E. 1983. Rapid flow cytometric analysis of the cell cycle in intact plant tissues. *Science* 220: 1049–1051
- Gichner T, Patková Z, Száková J, and Demnerová K. 2004. Cadmium induces DNA damage in tobacco roots, but no DNA damage, somatic mutations or homologous recombination in tobacco leaves. *Mutat. Res.* 559: 49–57
- Jiménez-Abián MI, Rozalén AE, Carballo JA, Botella LM, Pincheira J, López-Sáez JF, and de la Torre C. 2004. HSP 90 and checkpoint-dependent lengthening of the G2 phase observed in plant cells under hypoxia and cold. *Protoplasma* 223: 191–196
- Hagenbeek D, Quatrano RS, and Rock CD. 2000. Trivalent ions activate abscisic acid-inducible promoters through an ABI1-dependent pathway in rice protoplasts. *Plant Physiol.* 123: 1553–1560
- Helmbrecht K and Rensing L. 1999. Different constitutive heat shock protein 70 expression during proliferation and differentiation of rat C6 glioma cells. *Neurochem. Res.* 24: 1293–1299
- Hong FH, Wei ZG, and Zhao GW. 2000. Effect of lanthanum on aged seed germination of rice. *Biol. Trace Elem. Res.* 75: 205–213

Hormetic effects of rare earth element on plant cell cycles

- Hong FS, Wang L, and Liu C. 2003. Study of lanthanum on seed germination and growth of rice. *Biol. Trace Elem. Res.* 94: 273–286
- Hu X, Wang XR, and Wang C. 2006. Bioaccumulation of lanthanum and its effect on growth of maize seedlings in a red loamy soil. *Pedosphere* 16: 799–805
- Hu ZY, Richter H, Sparovek G, and Schnug E. 2004. Physiological and biochemical effects of rare earth elements on plants and their agricultural significance: A Review. *J. Plant Nutr.* 12: 183–220
- Huang M, Wei J N, Peng W X, Liang J, Zhao C, Qian Y, Dai G, Yuan J, Pan F Y, Xue B, Sha J H, and Li C J. 2009. The association of CaM and HSP 70 regulates S-phase arrest and apoptosis in a spatially and temporally dependent manner in human cells. *Cell Stress Chaperon.* 14: 343–353
- Karlseder J, Wissing D, Holzer G, Orel L, Sliutz G, Auer H, Jäättelä M, and Simon M M. 1996. HSP 70 overexpression mediates the escape of a doxorubicin-induced G2 cell cycle arrest. *Biochem. Bioph. Res. Co.* 220: 153–159
- Kim EK, Park JD, Shim SY, Kim HS, Kim BI, Choi JH, and Kim JE. 2006. Effect of chronic hypoxia on proliferation, apoptosis, and HSP 70 expression in mouse bronchiolar epithelial cells. *Physiol. Res.* 55: 405–411
- Kononowig AK, Hasegawa PM, and Bressan RA. 1992. Cell cycle duration in tobacco cells adapted to NaCl. *Environ. Exp. Bot.* 32: 1–9
- López-Millán AF, Sagardoy R, Solanas M, Abadía A, and Abadía J. 2009. Cadmium toxicity in tomato (*Lycopersicon esculentum*) plants grown in hydroponics. *Environ. Exp. Bot.* 65: 376–385
- Lucretti S, Nardi L, Nisini PT, Moretti F, Gualberti G, and Doležel J. 1999. Bivariate flow cytometry DNA/BrdUrd analysis of plant cell cycle. *Meth. Cell Sci.* 21: 155–166
- Milarski KL and Morimoto RI. 1986. Expression of human HSP 70 during the synthetic phase of the cell cycle. *Proc. Natl. Acad. Sci. USA* 83: 9517–9521
- Ouyang J, Wang XD, Zhao B, Yuan XF, and Wang YC. 2003. Effects of rare earth elements on the growth of *Cistanche deserticola* cell and the production of phenylethanoid glycosides. *J. Biotechnol.* 102: 129–134
- Qin LT, Liu SS, Liu HL, and Zhang YH. 2010. Support vector regression and least squares support vector regression for hormetic dose–response curves fitting. *Chemosphere* 78: 327–334
- Qiu G, Li W, Li X, Zhou W, and Yang C. 2005. Biological intelligence of rare earth elements in animal cells. *J. Rare Earth.* 23: 554–573
- Romero-Puertas MC, McCarthy I, Gómez M, Sandalio LM, Corpas FJ, Del Río LA, and Palma JM. 2004. Reactive oxygen species-mediated enzymatic systems involved in the oxidative action of 2, 4-dichlorophenoxyacetic acid. *Plant Cell Environ.* 27: 1135–1148
- Tammeba P, Wennberg T, Vuorela H, and Vuorela P. 2004. HPLC microfraction coupled to a cell-based assay for automated online primary screening of calcium antagonistic components in plant extracts. *Anal. Bioanal. Chem.* 380: 614–618
- Vargas-Roig LM, Fanelli MA, López LA, Gago FE, Tello O, Aznar JC, and Ciocca D R. 1997. Heat shock proteins and cell proliferation in human breast cancer biopsy samples. *Cancer Detect. Prev.* 21: 441–451
- Wang CR, Lu XW, Tian Y, Cheng T, Hu LL, Chen FF, Jiang CJ, and Wang XR. 2011. Lanthanum resulted in unbalance of nutrient elements and disturbance of cell proliferation cycles in *V. faba* L. seedlings. *Biol. Trace Elem. Res.* DOI: 10.1007/s12011-010-8939-z
- Wang CR, Tian Y, Wang XR, Yu HX, Lu XW, Wang C, and Wang H. 2010. Hormesis effects and implicative application in assessment of lead-contaminated soils in roots of *Vicia faba* seedlings. *Chemosphere* 80: 965–971
- Wang DF, Sun JP, Du DH, Sun LP, Chen ZD, and Xue CH. 2007. Degradation of extraction from seaweed and its complex with rare earths for organophosphorous pesticides. *J. Rare Earth.* 25: 93–99
- Wang LH, Huang XH, and Zhou Q. 2008. Effects of rare earth elements on the distribution of mineral elements and heavy metals in horseradish. *Chemosphere* 73: 314–319
- Wu JY, Wang CG, and Mei XG. 2001. Stimulation of taxol production and excretion in *Taxus* cell cultures by rare earth chemical lanthanum. *J. Biotechnol.* 85: 67–73
- Zeng Q, Zhu JG, Cheng HL, Xie ZB, and Chu HY. 2006. Phytotoxicity of lanthanum in rice in *haplic acrisols* and *cambisols*. *Ecotoxicol. Environ. Safe.* 64: 226–233