

**STATUS OF CITRUS CANKER IN ETHIOPIA AND MALAYSIA, AND
CHARACTERIZATION OF THE CAUSAL AGENT**

By

ESHETU DERSO KIDANU

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of the Requirements for the Degree of Doctor of Philosophy**

May 2006

DEDICATION

To the memory of my mother the late Shiwaye Derso who sacrificed everything to bring me up with love and affection.

Abstract of thesis presented to the Senate of Universiti Putra Malaysia
in fulfilment of the requirements for the degree of Doctor of Philosophy

**STATUS OF CITRUS CANKER IN ETHIOPIA AND MALAYSIA, AND
CHARACTERIZATION OF THE CAUSAL AGENT**

By

ESHETU DERSO KIDANU

May 2006

Chairman: Associate Professor Kamaruzaman Sijam, PhD

Faculty: Agriculture

Citrus canker disease surveys were conducted between August to November 2003 in Ethiopia and between January to April 2004 in West Malaysia. The pathogens were isolated and identified based on morphological characteristics and pathogenicity tests on seedlings of citrus cultivars using isolates collected from West Malaysia. Biochemical characterizations of the isolates were also carried out using standard determinative tests. In Ethiopia, citrus canker was observed only on sour orange (*Citrus aurantium*) and Mexican lime (*C. aurantifolia*). Disease incidence in the field on Mexican lime leaves in Ethiopia was 71.4% and severity was 26.8%; incidence on fruits was 30% and severity 21.25%. This is the first confirmed report of the disease in Ethiopia. In Malaysia, the disease was observed on Mexican lime (*C. aurantifolia*), pomelo (*C. grandis*) and kaffier lime (*C. hystrix*). Overall, the disease incidence in Malaysia was of 36.5% and severity of 12.5% on leaves, while incidence on fruits was 18.7% and severity 7.5%. Growth of *X. axonopodis* pv. *citri* in yeast dextrose chalk agar (YDCA) was not as fast as in peptone

sucrose agar (PSA) or nutrient glucose agar (NGA). However, the former medium was very selective to *Xanthomonas* species. There were highly significant differences in lesion size for cultivars ($P \leq 0.01$) but not for isolates. Significant positive regression ($P \leq 0.05$) was observed between lesion size and time after inoculation. Repeated measure analysis using general linear model (GLM) for correlation between times after inoculation was highly significant ($P \leq 0.01$). No variation in pathogenicity was observed among the isolates. Population sizes increased by over 2 Log cfu/lesion on Mexican lime and nearly by 1.5 Log units on sour orange and pomelo and remained around the initial inoculum level on calamondin. Significant positive correlations were observed between *X. axonopodis* pv. *citri* population and lesion size on sour orange ($r = 0.57$, $P = 0.024$), pomelo ($r = 0.73$, $P = 0.018$) and Mexican lime ($r = 0.76$, $P = 0.001$). The correlation was relatively the highest for Mexican lime ($r = 0.76$) and lowest for calamondin ($r = 0.25$). The interactions between isolates and cultivars were highly significant ($P \leq 0.01$). Tukey tests showed no significant differences in reaction to *X. axonopodis* pv. *citri* isolates between sour orange and pomelo, sour orange and sweet orange, Mexican lime and grapefruit, and also between calamondin and sweet orange. The six citrus test cultivars were all susceptible to the 15 *X. axonopodis* pv. *citri* isolates and citrus canker lesions were induced on the detached leaves. There was highly significant ($P \leq 0.01$) interaction between cultivars and all strains. Disease severity on detached leaves was significantly correlated ($P \leq 0.01$) with disease severity ratings in attached leaf test studies and was relatively the highest ($r = 0.97$) for Mexican lime. In the biochemical characterization study, all 15 isolates of *X. axonopodis* pv. *citri* showed similar results using standard determinative tests. Field host ranges for citrus canker in Ethiopia were Mexican lime and

sour orange, while in West Malaysia it appears to be wider than in Ethiopia. In conclusion, on the basis of their host range in seedling tests, morphological characteristics, pathogenicity tests, population growth in *planta* and biochemical characteristics, the 15 representative West Malaysian isolates of *X. axonopodis* pv. *citri* were characterized to be associated to the Asiatic type (“A” type) citrus canker. Sour orange, pomelo, Mexican lime, calamondin grapefruit and sweet orange were all susceptible to the 15 *X. axonopodis* pv. *citri* isolates.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**STATUS KANKER LIMAU DI ETHIOPIA DAN MALAYSIA, DAN
PENCIRIAN AGEN PENYEBAB**

Oleh

ESHETU DERSO KIDANU

Mei 2006

Pengerusi: Associate Professor Kamaruzaman Sijam, PhD

Fakulti: Pertanian

Tinjauan penyakit kanker limau telah dijalankan di Ethiopia dan Malaysia. Pemencilan, pencirian morfologi dan ujian patogenisiti pada anak benih kultivar limau telah dijalankan. Pencirian biokimia strain turut dijalankan menggunakan ujian penentuan pawai. Di Ethiopia, kanker limau hanya didapati pada limau masam (*Citrus aurantium*) dan limau nipis (*C. aurantifolia*). Kejadian penyakit pada daun di Ethiopia ialah 71% dan keterukan adalah 26.8%; kejadian pada buah ialah 30% dan keterukan 21.25%. Ini adalah laporan pertama yang sah mengenai penyakit di Ethiopia. Di Malaysia, penyakit ini didapati pada *limau nipis* (*C. aurantifolia*), *limau bali* (*C. grandis*) dan *limau purut* (*C. hystrix*). Secara keseluruhannya kejadian penyakit kanker limau di Malaysia adalah 36.5% pada daun dengan keterukan 15.1%, manakala kejadian pada buah adalah 18.7%. Pertumbuhan di atas Yeast dextrose chalk agar (YDCA) tidak se pantas seperti di atas agar sukrosa peptone (PSA) atau agar glucosa nutrient (NGA). Terdapat perbezaan yang

sangat bererti di antara saiz lesi bagi kultivar ($P \leq 0.01$) tetapi bukan untuk strain. Regresi positif yang bererti ($P \leq 0.05$) telah didapati di antara saiz lesi dan masa selepas inokulasi. Analisis pengukuran berulang menggunakan model linear umum (GLM) untuk korelasi di antara masa selepas inokulasi adalah sangat bererti ($P \leq 0.01$). Tiada perbezaan dilihat dari segi patogenisiti antara strain. Saiz populasi meningkat lebih 2 log cfu/lesi pada limau nipis dan hampir 1.5 log unit pada limau masam dan limau bali manakala kekal di paras inokulum permulaan bagi limau kasturi. Korelasi positif yang bererti telah dilihat di antara populasi *X. axonopodis* dengan saiz lesi pada limau masam ($r = 0.58$, $P = 0.024$), limau bali ($r = 0.73$, $P = 0.018$) dan juga limau nipis ($r = 0.76$, $P = 0.001$). Korelasi ini adalah paling tinggi untuk limau nipis ($r = 0.76$) dan terendah bagi limau kasturi. Interaksi di antara strain dan kultivar-kultivar limau adalah sangat bererti ($P \leq 0.01$). Ujian-t menunjukkan tiada berbezaan bererti dari segi tindak balas terhadap pencilan *X. axonopodis* pv. *citri* kultivar limau masam dan limau bali, limau masam dan sweet orange; limau nipis dan grapefruit, serta limau kasturi dan sweet orange Navel. Keenam-enam kultivar limau yang diuji adalah peka kepada 15 pencilan *X. axonopodis* pv *citri* dan lesi kanker limau telah terangsang pada daun terpisah. Interaksi sangat bererti ($P \leq 0.01$) antara kultivar dan semua pencilan Terukan simptom pada ujian daun terpisah menunjukkan korelasi bererti ($P \leq 0.01$) dengan kadar keterukan simptom kajian ujian daun terlekat. Korelasi yang tertinggi ($r = 0.97$) adalah pada limau nipis. Dalam ujian pencirian secara biokimia kesemua 15 pencilan *X. axonopodis* pv. *citri* menunjukkan keputusan yang sama dengan menggunakan ujian penentuan piawai. Julat perumah kanker limau di Ethiopia adalah limau nipis dan limau masam, manakala di Malaysia Barat ia kelihatan lebih luas berbanding di Ethiopia. Selanjutnya, berdasarkan julat

perumah ujian anak benih, ciri morfologi, ujian patogenisiti, pertumbuhan populasi dalam tumbuhan dan ciri biokimia, 15 wakil pencilan *X. axonopodis* pv. *citri* Malaysia Barat telah dicirikan sebagai berkaitan dengan kanker limau jenis Asiatic (Jenis “A”). Limau manis kesemuanya adalah peka kepada 15 pencilan *X. axonopodis* pv. *citri*.

ACKNOWLEDGEMENTS

I would like to thank Associate professor Dr. Kamaruzaman Sijam, chairman of the supervisory committee, for his humble and diligent guidance and help in all my needs despite his multiple administrative responsibilities in the department of plant protection. My sincere appreciation also goes to members of the supervisory committee, Associate professor Dr. Zainal Abidin Mior Ahmad, Associate professor Dr. Suhaimi Napis and Dr. Ibrahim Omar for their assistance and constructive comments during the entire study period.

I am highly indebted to my wife Kassech Tsegaye and my son Tewodros Eshetu for their understanding and love while I was away. My heart felt thanks and appreciation goes to my sisters Abaynesh Derso, Etenat Eshete, Almaz Ayalew, Dadi Derso my brother Mebratu Derso and Mr. Alemu Temesgen who have been there for me in my needs and their never ending assistance and encouragement before and during my study period and in my entire life. I thank you very much for your love and assistance. I thank the Ethiopian Institute of Agricultural Research (EIAR) management for giving me the opportunity to pursue my study abroad and funding the research project.

I would also like to express my appreciation to the plant protection department office and laboratory staff Universiti Putra Malaysia (UPM) for the invaluable assistance rendered to me through out my entire study period. I am also thankful to all staff of the Malaysian Agricultural research and development institute (MARDI) ,Cameron

Highlands, especially to the director Dr. Ibrahim Omar for his unreserved help and for allowing me to use all facilities in the center, and to Mr. Tengku Aziz Muda for his assistance and subsequent follow up in the glass house.

I am equally indebted to crop science department staff Dr. Abdul Shukor Juraimi and Dr. Uma Rani Sinnia at UPM for allowing me to use the facilities in their laboratories. Thanks also go to Dr. Jalo for his assistance and guidance in computer programming. My sincere appreciation also goes to my lab mates, Antario Dikins, Mazmi, Ayman Faisal, Mehran and Reza. I am also equally indebted to my friends Wubshet Mamo, Birhanu Mintesinot, Senay Zena, Akil Hizam, Ahmed Kaid, Tadese Yohannes, Bayeta Belachew and Zebene Mikru for their encouragements.

I would like to express my sincere gratitude to fellow Ethiopian post-graduate students at Universiti Putra Malaysia (UPM), Mekasha Chiche, Mandefro Negussie, Ahmed Seid, Tesfaye Shimber, Abayneh Isayiyas, Gebeyehu G/ Selasie, Ahmed Sherif, Erenso Degu, Negash Demmisie, Deribe Gurumu and Mullugeta Negeri for their assistance in the glass house and in the field and for their brotherly encouragement and friendship. Thanks also go to Ms. Altayework Deneke for her help to my family while I was away. Last but not least, I am equally indebted to all Malaysian people for their unforgettable hospitality throughout my stay in this beautiful country.

I certify that an Examination Committee has met on 3rd May 2006 to conduct the final examination of Eshetu Derso Kidanu on his Doctor of Philosophy thesis entitled “Status of Citrus Canker in Ethiopia and Malaysia, and Characterization of the Causal Agent” in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher Degree) Regulations 1981. The Committee recommends that the candidate be awarded the relevant degree. Members of the Examination Committee are as follows:

Rita Muhamad, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Chairman)

Jugah Kedir, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Internal Examiner)

Wong Mui Yun, PhD

Faculty of Agriculture
Universiti Putra Malaysia
(Internal Examiner)

Baharudin Salleh, PhD

Professor
Faculty of Science
Universiti Malaya
(External Examiner)

HASANAH BT MOHD GHAZALI, PhD

Professor/Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

This thesis submitted to the Senate of Universiti Putra Malaysia, has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. Members of the Supervisory Committee are as follows:

Kamaruzaman Sijam, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Chairman)

Zainal Abidin Mior Ahmed, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

Ibrahim Omar, PhD

MARDI
Bukit Tinggi
(Member)

Suhaimi Napis, PhD

Associate Professor
Faculty of Biotechnology and Biomolecular Science
Universiti Putra Malaysia
(Member)

AINI IDERIS, PhD

Professor/Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

DECLARATION

I hereby declare that the thesis is based on my original work except for quotation and citations which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at UPM or any other institutions.

ESHETU DERSO KIDANU

Date:

TABLE OF CONTENTS

	Page
DEDICATION	ii
ABSTRACT	iii
ABSTRAK	vi
ACKNOWLEDGMENTS	ix
APPROVAL	xi
DECLARATION	xiii
LIST OF TABLES	xvii
LIST OF FIGURES	xxi
LIST OF PLATES	xxiii
LIST OF ABBREVIATIONS	xxv

CHAPTER

I	GENERAL INTRODUCTION	1
II	LITERATURE REVIEW	6
	Introduction	6
	Importance of Citrus	6
	Global Production	8
	Citrus Canker	10
	Etiology of Citrus Canker Bacterium	13
	Symptoms	17
	Disease Cycle	18
	Host Range	21
	Geographic Distribution	21
	Means of Dispersal	22
	Canker Resistance	23
	Pathogenicity and inoculation techniques	26
	Detection and Identification	29
	Use of Selective Media	29
	Pathogenicity Tests	29
	Enzyme –Linked Immunosorbent Assay (ELISA)	30
	Molecular Technique	31
	Disease Management	32
	Regulatory Control	32
	Cultural Control	32
	Biological Control	33
	Chemical Control	33
	Phytosanitary Measures	33

III	CITRUS CANKER DISTRIBUTION AND INTENSITY IN ETHIOPIA AND MALAYSIA	
	Introduction	36
	Materials and Methods	36
	Survey Areas	36
	Sampling and Data Collection	37
	Data Analysis	37
	Results	40
	Surveyed Areas	43
	Disease Intensity in Ethiopia	43
	Disease Intensity in Malaysia	43
	Discussion	44
	Conclusion	50
		57
IV	CHARACTERIZATION OF <i>XANTHOMONAS</i> <i>AXONOPODIS</i> PV.<i>CITRI</i> ISOLATES	63
	Introduction	
	Materials and Methods	64
	Morphological Characterization	
	Sample Collection	
	Bacterial Isolation and Identity Confirmation	64
	Cellular Morphology	65
	Cultural Morphology	65
	Pathogenicity and Population Dynamics of <i>X. axonopodis</i> <i>pv. citri</i> Isolates on Citrus Cultivars	65
	Plant Materials	66
	Inoculum Preparation	67
	Leaf Inoculation	
	Data Collection	67
	Enumeration of Bacterial Population in Inoculated Cultivars	67
	Measurement of Bacterial Colony Number	69
	Disease Assessment	71
	Statistical Analysis	
	Biochemical and Physiological Characterization	71
	Reaction of Citrus Cultivars to <i>X. axonopodis</i> <i>pv. citri</i> Isolates	73
	Detached Leaf Test	74
	Bacterial Inoculation	75
	Disease Assessment	75
	Statistical Analysis	75
	Results	76
	Morphological Characterization	77

Bacterial Isolation and Identity Confirmation	78
Cellular Morphology	78
Cultural Morphology	78
Pathogenicity and Population Dynamics of <i>X. axonopodis</i>	78
<i>pv. citri</i> Isolates on Citrus Cultivars	80
Leaf Inoculation	80
Enumeration of Bacterial Population in Inoculated	84
Cultivars	
Biochemical and Physiological Characterization	84
Reaction of Citrus Cultivars to <i>X. axonopodis pv. citri</i>	93
Isolates	
Detached Leaf Test	102
Discussion	106
Conclusion	
	106
V GENERAL DISCUSSION AND CONCLUSION	118
	136
REFERENCES	138
APPENDICES	
BIODATA OF THE AUTHOR	147
	165
	200