

SID 06978570

THE UNIVERSITY OF THE WEST OF ENGLAND, BRISTOL

FORM RD14

CANDIDATE'S DECLARATION FORM

---

1 \*\*NAME OF CANDIDATE: **AARON MARAMBA**2 DEGREE FOR WHICH THESIS IS SUBMITTED: **Professional Doctorate (DBMS)**3 TITLE OF THESIS: **ZAP-70 PHOSPHORYLATION AND OTHER PROGNOSTIC FACTORS IN ADULT B-CHRONIC LYMPHOCYTIC LEUKAEMIA**

4 MATERIAL SUBMITTED FOR ANOTHER AWARD

\* I declare that no material contained in the thesis has been used in any other submission for an academic award.

or

\* I declare that the following material contained in the thesis formed part of a submission for the award of

.....  
(State award and awarding body and list the material below)

5 CONFIRMATION OF CONTENTS

I confirm that the contents of the permanently bound thesis are identical with the version submitted for examination except where amendments have been made to meet the requirements of the examiners.

**\*\*THE NAME AT THE TOP OF THIS FORM IS THE ONE THAT APPEARS ON OUR RECORD AND WILL APPEAR ON YOUR CERTIFICATE** (signing below will confirm unless stated)

Signature of candidate: ..... Date: .....

\* Delete as appropriate

---

For official use

Date received:

## **Acknowledgements**

I would like to thank my supervisor and director of studies Dr Craig Donaldson for his invaluable guidance and support throughout my DBMS. I am very grateful to all the DBMS staff, especially Dr Lynne Lawrence, Dr Ruth Morse, and Dr Michael Ladomery. My appreciation goes to Dr Mark Lowdell for allowing me to use his laboratory and for his advice on aspects of my project, and Ms Janet North for the technical guidance throughout. I also would like to thank Dr Gitendra-Wickremasinghe for allowing me to use some of his frozen B-CLL samples for retrospective studies.

I would like to thank the British Society for Haematology for a grant they awarded me in support of this project. I am also very grateful to the haematology department at Royal Free for allowing me access to the IgVH gene mutation results, especially Mr Jim Griffiths, who supported me during the initial stages of this thesis and organised the financial support to kick start my study. I also would like to acknowledge Mrs Saida Solkar, Ms Faith Wright and Ms Juliette Gevao who supported me in undertaking this Professional Doctorate alongside my professional practice. Great appreciation goes to Dr Steve Hart for his guidance and support and to Mr Jeff Lucas for helping with data analysis.

I would like to acknowledge all my fellow colleagues at the Royal Free Hospital and all my friends who constantly encouraged me to complete this thesis. Special thanks go to my wife and children for their encouragements throughout this long journey.

## **Dedications**

I would like to dedicate this thesis to my late beloved brother Phillip Ganye Maramba who untimely passed away on his birthday in 2011 after a road traffic accident. May his soul rest with eternal peace!

## **Abstract**

B chronic lymphocytic leukaemia (B-CLL) is a disease with an enormously heterogeneous prognosis. Due to lack of effective treatments and the heterogeneity in its course, B-CLL has remained largely incurable. In the last two decades a number of biological markers have been recognised as prognostic parameters to replace clinical staging, chief among them is ZAP-70 (a T-cell cytoplasmic tyrosine protein) albeit without a full understanding of the mechanism involved.

The aim of this thesis was to identify reliable prognostic factors that could be used in the management of B-CLL patients. Seventy-six B-CLL patients were studied over 30 months using a random stratified method of sampling, whereby adult B-CLL patients were recruited at different stages of the disease. Retrospective studies enabled some patients to be studied for up to 192 months. ZAP-70 expression was demonstrated in some B-CLL cells and was examined for possible phosphorylation following B-CLL cells stimulation, implying its involvement in cell signalling. Through the use of a novel method developed during the present study, B-CLL cells were stimulated via the SDF1- $\alpha$ -CD184 signalling pathway resulting in ZAP-70 phosphorylation. Its expression and phosphorylation was investigated alongside other prognostic factors and also correlated with B-CLL progression. More importantly, this study demonstrated that ZAP-70 positive cells are more responsive to signals derived from their surrounding environment, such as SDF-1 $\alpha$ .

In conclusion ZAP-70, CD38, and CD23 expressions together with IgVH gene mutational status were confirmed as indicators of poor prognosis. Response to external stimuli by some B-CLL cells, resulting in ZAP-70 phosphorylation represents the most novel aspect of this thesis and demonstrates ZAP-70 involvement in B-CLL signalling. Additionally, raised NK: B-CLL and T: B-CLL cell ratios were identified as good modifiable variable predictors of time to treatment that could be used for monitoring and therapeutic purposes. Since B-CLL is an incurable disease, the identification of prognostic factors could help in deciding the best time for intervention in B-CLL patients.

## CONTENTS

| <b>Page No.</b> |  |
|-----------------|--|
|                 | <b>Declaration</b> I   |
|                 | <b>Acknowledgements</b> II   |
|                 | <b>Dedications</b> III   |
|                 | <b>Abstract</b> IV   |
|                 | <b>List of Contents</b> V  |
|                 | <b>List of Figures</b> XI  |
|                 | <b>List of Tables</b> XVI  |
|                 | <b>List of Abbreviations</b> XVII                                      |
|                 | <b>Publications</b> XXI  |
|                 | <b>CHAPTER 1: General introduction</b> 1                               |
|                 | 1.1.0 Chronic lymphocytic leukaemia and B cell development 2           |
|                 | 1.1.1 Normal B-Cell development 6                                      |
|                 | 1.1.2 Normal B cell development in the bone marrow 8                   |
|                 | 1.1.3 B-cell surface IgM expression 11                                 |
|                 | 1.1.4 B cell activation 15   |
|                 | 1.2.0 Essential steps in B-CLL development and its biology 16          |
|                 | 1.2.1 Genesis of chronic lymphocytic leukaemia 18                      |
|                 | 1.2.2 Immunoglobulin V genes in B-CLL 21                               |
|                 | 1.2.3 B-cell receptors (BCR) in B-CLL 22                               |
|                 | 1.2.4 Stromal cell-derived factor-1 $\alpha$ and its receptor CD184 24 |
|                 | 1.2.5 CD184 expression in B-CLL 26                                     |
|                 | 1.3 Prognostic markers for B-CLL 28                                    |
|                 | 1.3.1 ZAP-70 in B-CLL 28   |

|   |  |           |
|---|--|-----------|
| 1.3.2                                   | Immunoglobulin heavy-chain gene mutation status in B-CLL | 32        |
| 1.3.3                                   | CD38 expression in B-CLL                                 | 33        |
| 1.3.4                                   | P53 expression in B-CLL                                  | 34        |
| 1.3.5                                   | Lymphocyte doubling time and overall survival in B-CLL   | 37        |
| 1.3.6                                   | Binet staging for B-CLL                                  | 38        |
| 1.4.0                                   | Treatment for B-CLL                                      | 39        |
| 1.4.1                                   | Radiotherapy for B-CLL                                   | 40        |
| 1.4.2                                   | Chemotherapy for B-CLL                                   | 42        |
| 1.4.2.1                                 | Chemotherapy drugs for B-CLL and mode of action          | 45        |
| 1.4.2.2                                 | Mechanism of action of steroids in B-CLL                 | 48        |
| 1.4.3                                   | Biologic or immunotherapy for B-CLL                      | 50        |
| 1.4.4                                   | Clinical trials for B-CLL                                | 52        |
| 1.4.5                                   | Bone marrow and stem cell transplant for B-CLL           | 55        |
| 1.4.6                                   | Novel therapies under clinical trials investigation      | 57        |
| 1.5                                     | Aims of the thesis                                       | 61        |
| <b>Chapter 2: Protocol optimisation</b> |  | <b>63</b> |
| 2.1                                     | Introduction   | 63        |
| 2.1.1                                   | Instrument settings                                      | 64        |
| 2.1.2                                   | Theory on antigen-antibody binding                       | 66        |
| 2.1.3                                   | Kinetics   | 66        |
| 2.1.4                                   | CD184 epitope density and its reactive potential         | 67        |
| 2.1.5                                   | IgM expression and its reactivity properties             | 68        |
| 2.1.6                                   | ZAP-70 expression  | 69        |
| 2.2                                     | Aims   | 69        |
| 2.3                                     | Methods  | 70        |

|   |   |            |
|---|---|------------|
| 2.3.1   | Instrument setup  | 70         |
| 2.3.2   | Gating strategy   | 73         |
| 2.3.3   | Ficoll-paque versus lymphoprep: cell viability analysis   | 74         |
| 2.3.4   | ZAP-70 Analysis   | 75         |
| 2.3.5   | Optimizing the volume of the stimulant                    | 76         |
| 2.3.6   | Optimising the stimulation time                           | 77         |
| 2.3.7   | BD fixation and permeabilisation                          | 77         |
| 2.3.8   | Dako intrastain fixation and permeabilisation             | 78         |
| 2.3.9   | Comparison of the BD and Dako fix and perm protocols      | 78         |
| 2.4   | Results   | 79         |
| 2.4.1   | Gating strategy   | 79         |
| 2.4.2   | Isotype controls  | 81         |
| 2.4.3   | ZAP-70 labelling before surface antigen staining          | 84         |
| 2.4.4   | Jurkat cells as positive controls                         | 86         |
| 2.4.5   | Stimulant of choice                                       | 87         |
| 2.4.6   | Volume of the stimulant (SDF1 $\alpha$ and anti-IgM)      | 89         |
| 2.4.7   | Optimum time for stimulating fresh cells                  | 89         |
| 2.4.8   | Effect of freezing and thawing on phosphorylation         | 91         |
| 2.4.9   | Fixation and permeabilisation reagent choice (Dako Vs BD) | 93         |
| 3.5   | Discussion and concluding remarks                         | 96         |
| <b>Chapter 3: Materials and general methods</b> |   | <b>100</b> |
| 3.1   | Equipment   | 100        |
| 3.2   | Materials and reagents                                    | 100        |

|         |   |     |
|---------|---|-----|
| 3.3     | Patient selection and sample preparation                    | 103 |
| 3.3.1   | Preparation of stored samples                               | 108 |
| 3.3.2   | Thawing of previously stored samples                        | 108 |
| 3.3.3   | Viability testing   | 109 |
| 3.3.4   | Preparation of mononuclear cells from fresh samples         | 110 |
| 3.3.5   | Cell Counting   | 111 |
| 3.3.6   | Lyse-and-then-wash method for harvesting white cells        | 111 |
| 3.3.7   | Jurkat (human T-cell leukaemia) cell line                   | 112 |
| 3.4     | Flow cytometry  | 112 |
| 3.4.1   | Surface antigens labelling                                  | 112 |
| 3.4.2   | ZAP-70 analysis   | 112 |
| 3.4.3   | Phosphorylation of intracellular ZAP-70 (pY292 and 319)     | 113 |
| 2.4.3.1 | IgM stimulation for cell lines                              | 113 |
| 3.4.3.2 | SDF1- $\alpha$ stimulation of B-CLL cells                   | 113 |
| 3.4.4.3 | Stimulation of Jurkat cells                                 | 113 |
| 3.4.3.4 | Dako intra stain fixation and permeabilisation protocol     | 114 |
| 3.4.3.5 | Flow cytometric data analysis for pY-ZAP-70                 | 114 |
| 3.5     | IgVH gene mutation analyses                                 | 115 |
| 3.5.1   | Choice of materials and detection of clones                 | 116 |
| 3.5.2   | Screening for mutations in the clone hetero-duplex analysis | 117 |
| 3.5.3   | Sequencing of the IgVH gene                                 | 118 |
| 3.6     | Statistical analysis  | 119 |



|   |            |
|---|------------|
| <b>Chapter 4 ZAP-70 as a prognostic marker in B-CLL</b>       | <b>120</b> |
| 4.1 Introduction  | 120        |
| 4.2 Aims and objectives                                       | 124        |
| 4.2.1 Primary objectives                                      | 124        |
| 4.3 Methods   | 124        |
| 4.3.1 Laboratory determination of ZAP-70 expression           | 125        |
| 4.3.2 ZAP-70 phosphorylation                                  | 125        |
| 4.3.3 Statistical analysis                                    | 125        |
| 4.4 Results   | 126        |
| 4.4.1 Significance of ZAP-70 in B-CLL                         | 130        |
| 4.4.2 Correlation of ZAP-70 and its phosphorylation status    | 139        |
| 4.4.3 Multiple correlations for ZAP-70                        | 141        |
| 4.4.4 Changes in ZAP-70 expression during the course of B-CLL | 148        |
| 4.5 Discussion  | 152        |
| <b>Chapter 5: Other prognostic markers for B-CLL</b>          | <b>158</b> |
| 5.1 Current knowledge of prognostic markers in B-CLL          | 158        |
| 5.1.2 Established prognostic markers in B-CLL                 | 159        |
| 5.1.3 New prognostic markers                                  | 160        |
| 5.2 Aims  | 161        |
| 5.3 Methods   | 162        |
| 5.3.1 Full blood counts                                       | 162        |
| 5.3.2 Flow cytometry  | 162        |
| 5.3.3 IgVH gene mutation analyses                             | 162        |
| 5.3.4 Statistical analysis                                    | 162        |
| 5.4 Results   | 163        |

|   |  |            |
|---|--|------------|
| 5.4.1   | Contextualising stable and progressive disease             | 163        |
| 5.4.2   | B-CLL course in relation to traditional prognostic markers | 167        |
| 5.4.3   | CD23 expression patterns in B-CLL patients                 | 170        |
| 5.4.4   | Common IgVH genes in B-CLL                                 | 174        |
| 5.4.5   | Multiple correlations                                      | 177        |
| 5.4.6   | BCR expression in B-CLL                                    | 181        |
| 5.4.7   | Response to therapy  | 184        |
| 5.5   | Discussion   | 187        |
| <b>Chapter 6: General discussion and conclusion</b> |  | <b>193</b> |
| 6.1   | Method development   | 194        |
| 6.2   | ZAP-70 expression in B-CLL and its prognostic significance | 194        |
| 6.3   | ZAP-70 correlations with other prognostic indicators       | 196        |
| 6.4   | B-Cell receptors in B-CLL                                  | 199        |
| 6.5   | Modifiable prognostic characteristics in B-CLL             | 200        |
| 6.6   | Interpretation and application of prognostic markers       | 201        |
| 6.7   | Conclusion   | 205        |
| 6.8   | Future work  | 206        |
| <b>Chapter 7: Reflective</b>                        |  | <b>208</b> |
| 7.1   | Introduction   | 208        |
| 7.2   | Commencing the DBMS  | 210        |
| 7.3   | The research project                                       | 212        |
| <b>Chapter 8: References</b>                        |  | <b>219</b> |
| <b>APPENDICES</b>                                   |  | <b>252</b> |

## List of Figures

|             |  |    |
|-------------|--|----|
| Figure 1.1  | Incidence of chronic lymphocytic leukaemia in the UK   | 3  |
| Figure 1.2  | Appearance of Romanowsky stained peripheral blood film from a patient with B-CLL   | 5  |
| Figure 1.3  | Sequence of events and characteristics of the stages in B cell maturation in the bone marrow and periphery   | 7  |
| Figure 1.4  | B cell development in the bone marrow  | 8  |
| Figure 1.5  | Schematic diagram of sequential expression of membrane immunoglobulin and surrogate light chain at different stages of B-cell differentiation in the bone marrow | 9  |
| Figure 1.6  | Comparison between B-1 and B-2 B cells' attributes   | 10 |
| Figure 1.7  | Pentameric IgM and monomeric IgG1 structures   | 11 |
| Figure 1.8  | Complementarity determining regions (CDR) on the monomeric structure of the IgM  | 12 |
| Figure 1.9  | BCR and the B-cell co-receptor activation and signalling   | 14 |
| Figure 1.10 | BCR and CXCR4 activation and signalling demonstrating the interactions between B-CLL cell and its microenvironment   | 24 |
| Figure 1.11 | Schematic structure of ZAP-70  | 28 |
| Figure 1.12 | The balance of negative and positive signals   | 30 |
| Figure 1.13 | The <i>p53</i> signalling pathway  | 36 |
| Figure 1.14 | Mechanism of action of DNA-damaging drugs  | 44 |
| Figure 1.15 | Mechanism of action and resistance of traditional DNA-damaging anti-cancer drugs   | 47 |
| Figure 2.1  | CST beads dot plot and histograms  | 72 |
| Figure 2.2  | Sequential gating strategy for surface antigens  | 79 |

|             |  |     |
|-------------|--|-----|
| Figure 2.3  | Sequential and reverse gating for ZAP-70   | 80  |
| Figure 2.4  | Isotype control for a ZAP-70 negative sample   | 82  |
| Figure 2.5  | Isotype control for a ZAP-70 positive sample   | 83  |
| Figure 2.6  | Effect of Fixation and permeabilisation on surface antigens  | 85  |
| Figure 2.7  | Effect of H <sub>2</sub> O <sub>2</sub> treatment of Jurkat cells  | 86  |
| Figure 2.8  | Comparison of IgM and CD184 expression on B-CLL cells from two patients                                      | 87  |
| Figure 2.9  | Determination of optimum time for ZAP-70 phosphorylation   | 90  |
| Figure 2.10 | Freeze and thaw effect on ZAP-70 phosphorylation   | 92  |
| Figure 2.11 | Comparison between BD Biosciences and Dako Cytomation fix and perm reagents for tyrosine 292 phosphorylation | 94  |
| Figure 2.12 | Comparison between BD Biosciences and Dako Cytomation fix and perm reagents for tyrosine 319 phosphorylation | 95  |
| Figure 3.1  | Lymphocyte harvesting  | 111 |
| Figure 3.2  | Representative photograph of IgVH somatic hypermutation hetero-duplex gel                                    | 118 |
| Figure 4.1  | Structural organisation of Inactive ZAP-70 and comparison with auto-inhibited Scr family kinases             | 122 |
| Figure 4.2  | Comparison of the lymphocyte counts at diagnosis   | 127 |
| Figure 4.3  | Comparison of patients' survival times based on ZAP-70 expression in B-CLL cells                             | 128 |
| Figure 4.4  | ZAP-70 in female and male patients   | 130 |
| Figure 4.5  | ZAP-70 expressions for kappa and lambda chain restriction patients   | 131 |

|             |  |     |
|-------------|--|-----|
| Figure 4.6  | Comparison of ZAP-70 between treated and untreated patients  | 132 |
| Figure 4.7  | Survival times for patients who did not receive treatment  | 133 |
| Figure 4.8  | Comparison of time to treatment (TT) between ZAP-70 negative and ZAP-70 positive patients                                      | 134 |
| Figure 4.9  | Comparison of survival times and time to first treatment for B-CLL patients with <20% ZAP-70 expression who received treatment | 135 |
| Figure 4.10 | Comparison of survival times and time to first treatment for B-CLL patients with >20% ZAP-70 expression who received treatment | 136 |
| Figure 4.11 | Comparison of the survival times for patients with ZAP-70 in >20% B-CLL cells  | 137 |
| Figure 4.12 | Survival times for both treated and untreated B-CLL patients   | 138 |
| Figure 4.13 | One-way ANOVA test with Dunn's multiple comparison tests   | 143 |
| Figure 4.14 | Tyrosine 292 phosphorylation and other prognostic markers in adult B-CLL patients  | 144 |
| Figure 4.15 | Tyrosine 319 phosphorylation and other prognostic markers in adult B-CLL patients  | 145 |
| Figure 4.16 | ZAP-70 phosphorylation in the presence other markers   | 146 |
| Figure 4.17 | Comparison between tyrosines 292 and 319 phosphorylation   | 147 |
| Figure 4.18 | ZAP-70 expression changes in 14 B-CLL patients   | 148 |

|             |   |     |
|-------------|---|-----|
| Figure 4.19 | Comparison between peripheral blood and bone marrow ZAP-70 expression                 | 151 |
| Figure 5.1  | Lymphocyte counts in stable and progressive B-CLL                                     | 164 |
| Figure 5.2  | Comparison of mean lymphocyte counts between treated and untreated B-CLL cases        | 165 |
| Figure 5.3  | CD5 expressions as a function of peripheral blood lymphocyte counts in B-CLL patients | 166 |
| Figure 5.4  | Comparison of CD38 expression in stable and progressive B-CLL patients                | 169 |
| Figure 5.5  | ZAP-70 levels between stable and progressive B-CLL                                    | 169 |
| Figure 5.6  | Comparison between CD5 and CD23 on B-CLL cells  | 170 |
| Figure 5.7  | Comparison of CD23 expression between low level B-CLL and advanced disease            | 171 |
| Figure 5.8  | One-way ANOVA Comparisons of CD23 expression and B-CLL course                         | 173 |
| Figure 5.9  | IgVH gene mutations detected  | 175 |
| Figure 5.10 | Comparisons for ZAP-70 expression in relation to VH3-21                               | 176 |
| Figure 5.11 | Comparisons for CD38 expression in relation to VH3-21                                 | 177 |
| Figure 5.12 | CD38 expressions between IgVH mutated and unmutated B-CLL patients                    | 179 |
| Figure 5.13 | One-way ANOVA comparison of CD23 expression and other prognostic markers              | 180 |
| Figure 5.14 | Multiple comparisons for BCR and ZAP-70   | 181 |
| Figure 5.15 | Linear regression curves for IgM versus CD184 expressions                             | 182 |

Figure 5.16 Comparison of CD184 expression between IgM weak and IgM  
medium expressing B-CLL cells

182

**List of Tables**

|           |   |     |
|-----------|---|-----|
| Table 1.1 | B-cell chronic lymphocytic leukaemia immunophenotype in comparison to mantle cell and follicular lymphoma | 20  |
| Table 1.2 | A summary of some published studies of consolidation B-CLL trials   | 54  |
| Table 3.1 | Patient demographics  | 104 |
| Table 4.1 | Observed trends in the three groups based on B-CLL cells' ZAP-70 levels                                   | 126 |
| Table 4.2 | Characterisation of B-CLL patients who died during the study  | 129 |
| Table 4.3 | ZAP-70 correlation tests  | 140 |
| Table 4.4 | ZAP-70 associations with other prognostic markers   | 141 |
| Table 4.5 | Nonparametric correlations  | 142 |
| Table 4.6 | ZAP-70 fluctuation in peripheral blood over time  | 149 |
| Table 4.7 | ZAP-70 expression differences between PB and BM at the same time  | 150 |
| Table 5.1 | B-CLL course and the associated prognostic markers  | 168 |
| Table 5.2 | Characteristic of patients with low level disease   | 172 |
| Table 5.3 | Pearson correlation analyses  | 178 |
| Table 5.4 | Spearman's correlations for BCR expression in B-CLL   | 183 |
| Table 5.5 | B-CLL response to therapy and the associated markers  | 185 |
| Table 6.1 | Studies to evaluate prognostic power of different markers   | 202 |



## List of Abbreviations

|       |  |
|-------|--|
| ADCC  | Antibody-dependent cellular cytotoxicity   |
| ALL   | Acute lymphoblastic leukaemia  |
| AML   | Acute myeloid leukaemia  |
| APC   | Allophycocyanin  |
| ATM   | Ataxia telangiectasia mutated  |
| BCR   | B cell receptor  |
| BLNK  | B-cell linker  |
| BM    | Bone marrow  |
| BMT   | Bone marrow transplant   |
| BSAP  | B-cell lineage specific activator protein  |
| CADPR | Cyclin adenosine diphosphate-ribose  |
| CXC   | Chemokines that are classified depending on the spacing and number of their conserved cysteine residue |
| CCR   | Chemokine receptor   |
| CD    | Cluster of differentiation   |
| CDR   | Complimentarity determining region   |
| CHOP  | Cyclophosphamide + Hydroxydaunorubicin + Oncovin + Prednisone  |
| CLL   | Chronic lymphocytic leukaemia  |
| CML   | Chronic myeloid leukaemia  |
| CPD   | Continuous Professional Development  |
| CST   | Cytometer setup and tracking   |
| DBMS  | Professional doctorate in biomedical science   |
| DMSO  | Dimethyl sulphoxide  |

|                               |  |
|-------------------------------|--|
| DNA                           | Deoxyribonucleic acid                      |
| EBF                           | Early B-cell factor                        |
| EBRT                          | External beam radiotherapy                 |
| EBV                           | Epstein barr virus                         |
| EDTA                          | Ethylene-diamine-tetra-acetic acid         |
| ELC                           | EBV-induced receptor ligand chemokine      |
| ERK                           | Extracellular signal-regulated kinases     |
| FACS                          | Fluorescence-activated cell sorting        |
| FBC                           | Full blood counts                          |
| FCR                           | Fludarabine + Cyclophosphamide + Rituximab |
| FCS                           | Foetal calf serum                          |
| FDA                           | Food and drug administration               |
| FITC                          | Fluorescein isothiocyanate                 |
| FR                            | Fragment region                            |
| FMC7                          | Flinders medical centre, Australia         |
| FSC                           | Forward scatter channel                    |
| GPCR                          | Glycoprotein-coupled receptor              |
| GR                            | Glucocorticoid receptors                   |
| GPI                           | Glycosylphosphatidylinositol               |
| GVHD                          | Graft-versus-host disease                  |
| GVL                           | Graft-versus leukaemia                     |
| H <sub>2</sub> O <sub>2</sub> | Hydrogen peroxide                          |
| HBSS                          | Hank's balanced salt solution              |
| HDMP                          | High dose methylprednisolone               |
| HIV                           | Human immunodeficiency virus               |

|               |   |
|---------------|---|
| HPC           | Health professions council                      |
| HSCT          | Haematopoietic stem cell transplant             |
| IBMS          | Institute of biomedical science                 |
| Ig            | Immunoglobulin                                  |
| IgVH          | Immunoglobulin heavy-chain variable region      |
| IL            | Interleukin                                     |
| IMGT          | International <b>ImMunoGeneTics</b>             |
| ITAM          | Immunoreceptor tyrosine-based activation motifs |
| ITIM          | Immunoreceptor tyrosine inhibitory motifs       |
| ITP           | Immune thrombocytopenia                         |
| JNK           | Jun N-terminal kinase                           |
| LDT           | Lymphocyte doubling time                        |
| LN            | Lymph node                                      |
| MAPK          | Mitogen-activated protein kinase                |
| MBC           | Monoclonal B Cells                              |
| MDM2          | Mouse/ murine double minute 2                   |
| MDS           | Myelodysplastic syndromes                       |
| MFI           | Median fluorescence intensity                   |
| MRD           | Minimal residual disease                        |
| MRNA          | Messenger RNA                                   |
| mIg           | Membrane immunoglobulin                         |
| MIP-3 $\beta$ | Macrophage inflammatory protein 3 $\beta$       |
| MNC           | Mononuclear cell                                |
| MWM           | Molecular weight marker                         |
| NFAT          | Nuclear factor of activated T-cell              |

|                |   |
|----------------|---|
| NHS            | Nation health service   |
| NICE           | National institute of clinical excellence                         |
| NK             | Natural killer  |
| NRTI           | Nucleoside analogue that work as reverse transcriptase inhibitors |
| OS             | Overall survival  |
| P53            | Tumour protein 53   |
| PCR            | Polymerase chain reaction   |
| PE             | Phycoerythrin   |
| Per CP         | Peridicin chlorophyll-protein                                     |
| PFS            | Progression free survival   |
| PLC $\gamma$ 2 | Phospholipase- C $\gamma$ 2                                       |
| PMT            | Photomultiplier tube  |
| PRBCA          | Pure red blood cell aplasia                                       |
| PTK            | Protein tyrosine kinase   |
| PTP            | Protein tyrosine phosphatase                                      |
| RA             | Refractory anaemia  |
| RAG            | Recombination activating genes                                    |
| RIC            | Reduced intensity conditioning                                    |
| RNA            | Ribonucleic acid  |
| RPMI           | Roswell park memorial institute                                   |
| SCF            | Stem cell factor  |
| SDF-1 $\alpha$ | Stromal cell-derived factor 1 $\alpha$                            |
| SH2            | Src homology 2  |
| SHM            | Somatic hyper-mutations   |

|        |   |
|--------|---|
| SHIP   | Src homology inositol phosphate                             |
| SHP1   | Src homology containing protein 1                           |
| SLC    | Secondary lymphoid tissue chemokine                         |
| SLP-76 | Src homology 2 domain containing leukocyte protein of 76kDa |
| SSC    | Side scatter channel  |
| ST     | Survival times  |
| Syk    | Spleen tyrosine kinase                                      |
| TAPA-1 | Target of anti-proliferative antibody-1 (CD81)              |
| TCR    | T-Cell receptor   |
| TD     | Thymus dependent  |
| TdT    | Terminal deoxyribonucleotidyl transferase                   |
| TEM    | Transendothelial migration                                  |
| TFS    | Treatment free survival                                     |
| TK     | Tyrosine kinase   |
| TP53   | p53 gene  |
| TT     | Time to treatment   |
| VCAM-1 | Vascular cell adhesion molecule -1                          |
| VLA-4  | Very late antigen -4  |
| XIAP   | X-linked inactivator of apoptosis                           |
| WBC    | White blood cells   |
| ZAP-70 | Zeta associated protein of 70 kDa                           |

## **PUBLICATIONS**

Maramba A., North J., Lowdell MW and Donaldson C. IgM ligation triggers Zeta associated protein of 70 kDa (ZAP-70) tyrosine 292 and 319 phosphorylation in B-Chronic Lymphocytic Leukaemia (B-CLL). *Poster presentation at the British Society for Haematology British Journal of Haematology* 2011; **153 (Suppl. 1): 72-73.**