

Programming of adiposity in childhood and adolescence: associations with birth weight and cord blood adipokines

Joy Simpson MRCOG, Andrew DAC Smith PhD, Abigail Fraser PhD, Naveed Sattar PhD, Robert S Lindsay PhD, Susan M Ring PhD, Kate Tilling PhD, George Davey Smith DSc, Debbie A Lawlor PhD, Scott M Nelson PhD

The Journal of Clinical Endocrinology & Metabolism Endocrine Society

Submitted: June 08, 2016 Accepted: November 08, 2016 First Online: November 14, 2016

Early Release articles are PDF versions of manuscripts that have been peer reviewed and accepted but not yet copyedited. The manuscripts are published online as soon as possible after acceptance and before the copyedited, typeset articles are published. They are posted "as is" (i.e., as submitted by the authors at the modification stage), and do not reflect editorial changes. No corrections/changes to the PDF manuscripts are accepted. Accordingly, there likely will be differences between the Early Release manuscripts and the final, typeset articles. The manuscripts remain listed on the Early Release page until the final, typeset articles are posted. At that point, the manuscripts are removed from the Early Release page.

DISCLAIMER: These manuscripts are provided "as is" without warranty of any kind, either express or particular purpose, or non-infringement. Changes will be made to these manuscripts before publication. Review and/or use or reliance on these materials is at the discretion and risk of the reader/user. In no event shall the Endocrine Society be liable for damages of any kind arising references to, products or publications do not imply endorsement of that product or publication.

Programming of adiposity in childhood and adolescence: associations with birth weight and cord blood adipokines

Cord blood measures and long-term adiposity

Joy Simpson MRCOG^{1*}, Andrew DAC Smith PhD^{2,3*}, Abigail Fraser PhD^{2,3}, Naveed Sattar PhD⁴, Robert S Lindsay PhD⁴, Susan M Ring PhD^{2,3}, Kate Tilling PhD^{2,3}, George Davey Smith DSc^{2,3}, Debbie A Lawlor PhD^{2,3}, Scott M Nelson PhD¹

^{*} These authors made equal contribution

¹School of Medicine, University of Glasgow, UK

² Medical Research Council Integrative Epidemiology Unit at the University of Bristol, UK

³ School of Social and Community Medicine, University of Bristol, UK

⁴ Institute of Cardiovascular and Metabolic Medicine, British Heart Foundation Glasgow Cardiovascular Research Centre, University of Glasgow, UK Received 8 June 2016. Accepted 8 November 2016.sss

Context: Exposure to maternal adiposity during pregnancy is associated with higher offspring birthweight and greater adiposity through childhood and adult life. As birthweight reflects the summation of lean and fat mass, the extent to which fat mass at birth tracks into later life is unknown.

Objective: Determine whether fat mass at birth is associated with child and adolescent adiposity.

Design, Setting and Participants: UK birth cohort with markers of neonatal fat mass; cord blood leptin, adiponectin, and birthweight and adiposity outcomes at age 9 (N=2775) and 17years (N=2138).

Main Outcomes: Offspring BMI, waist circumference, DXA-determined fat mass and obesity at age 9 and 17 years.

Results: Higher cord blood leptin was associated with higher z-scores of fat mass (difference in mean per 10pg/ml: 0.03SD,95%CI 0.00-0.06), waist circumference (0.04SD,95%CI 0.00-0.07), and BMI (0.04SD,95%CI 0.00-0.08), at age 9. However, by age 17 the adjusted results were attenuated to the null. Cord blood adiponectin was not associated with measures of adiposity at age 9. At age 17, cord blood adiponectin was positively associated with fat mass (0.02SD per 10µg/ml,95%CI 0.02-0.03) and waist circumference (0.04SD per 10µg/ml,95%CI 0.03-0.05). Birthweight was positively associated with waist circumference (0.03SD per 100g,95%CI 0.02-0.04) and BMI (0.02SD per 100g,95%CI 0.00-0.03), but not fat mass or odds of obesity. Cord blood leptin and adiponectin were not associated with obesity at either age.

Conclusions: Increased cord blood leptin and adiponectin, known surrogates of fetal fat mass, were weakly associated with increased fat mass in late childhood and adolescence respectively.

PRECIS: We found that cord blood leptin and adiponectin, known surrogates of fetal fat mass, were weakly positively associated with some measures of fat mass in late childhood and adolescence.

Introduction

Exposure to maternal adiposity during pregnancy is associated with higher offspring birth weight and greater adiposity through childhood and adult life (1). Developmental overnutrition has been proposed as a mechanism, by which excessive transplacental passage of nutrients facilitates the development of larger babies with greater fat mass. Evidence from

CRINE CRINE

ОСЛ Ц Ц Ц Ц Ц Ц Ц

within sibling studies, comparisons of maternal and paternal exposures and the use of genetic variants as proxies for the maternal exposures support maternal adiposity and developmental overnutrition causing greater adiposity in offspring at birth (2-4). However, whether this causal effect extends to long-term offspring adiposity is unclear. A longer-term effect may occur as a result of tracking of birth fatness across the life course. However, because birth weight is unable to distinguish relative contributions of lean versus fat mass (5,6), few studies to date have been able to determine the extent to which greater fat mass at birth tracks into later life.

Umbilical cord blood leptin is widely recognized as an accurate biomarker for neonatal fat mass (7). Maternal exposures, including maternal adiposity which may cause developmental overnutrition, have been associated with increased cord leptin and neonatal adiposity at birth (8.9). In animal models, fetal leptin has also been proposed to contribute the long-term programming of hypothalamic feeding circuits, thereby providing a means by which leptin can influence long-term adiposity independent of tracking of adiposity from birth(10). Use of cord blood leptin in determining whether neonatal fat mass tracks across childhood has however been limited (11-14). This primarily reflects the scarcity of large prospective birth cohorts with cord blood samples and detailed measures of offspring adiposity as well as potential confounders. Studies that have made some assessment of this to date have had relatively small sample sizes (N=56-588) (11-14), and we are not aware of any study having followed children beyond age 7 years. These studies have reported nonconsistent results with higher cord leptin associated with both a lower (11) and higher (12) BMI at age 3 years, and a higher BMI at age 7 years (14).

Neonatal levels of adiponectin, which has insulin sensitizing effects in adults, are approximately 4-7 times higher than maternal levels. Furthermore, while maternal circulating concentrations of adiponectin are inversely associated with BMI, higher levels of cord blood adiponectin are associated with higher birth weight (11,15). That higher cord blood adiponectin concentrations might reflect increased fat mass in neonates is suggested by mouse studies where over-expression of fetal adiponectin was positively related to the size of fat depots in early life, while adiponectin knockout fetuses display lower body weight and lower fat content(16). Given this effect of adiponectin on body composition, specifically, its fat deposition enhancing effect in mice, and the known relationships of leptin in humans to fat mass, we hypothesized that both cord blood leptin and adiponectin would be positively associated with offspring adiposity in pre-pubertal children and adolescents.

The aim of this study was to determine whether cord blood leptin and adiponectin were positively associated with later obesity, BMI, waist circumference and fat mass and whether this is independent of maternal BMI. For comparison, we also examined associations of birthweight with these outcomes.

Research Design and Methods

Study Population

The Avon Longitudinal Study of Parents and Children (ALSPAC) is a prospective birth cohort study investigating the health and development of children (17,18). The study website contains details of all the data that is available through a fully searchable data dictionary; http://www.bris.ac.uk/alspac/researchers/data-access/data-dictionary/. Ethical approval was obtained from the ALSPAC Law and Ethics Committee and the National Health Service local ethics committees. A total of 14,541 women were initially enrolled, with 5011 motheroffspring pairs having a suitable cord blood sample. A detailed outline of the exclusion criteria for the analysis reported here and numbers with missing data is shown in Figure 1. We included participants if they had 1) attended and completed assessments at either the 9 or the 11-year clinic assessment, or 2) attended the 15 or 17-year clinic assessment. The eligible

cohort for the current analysis was 2775 mother-offspring pairs at age 9-11 years and 2138 mother-offspring pairs at age 15-17 years.

Cord blood Assays

Cord blood samples were collected at the time of delivery, initially stored at 4°C for 0 to 8 days before plasma was separated and then stored at -20°C before being transferred to longterm storage at -80°C. Cord blood leptin and adiponectin were measured using commercially available ELISA kits (Quantikine human leptin immunoassay (Cat No PDLP00), Quantikine Human Total Adiponectin/Acrp30 (Cat No PDRP300) both R&D Systems). Analysis of the cord blood was completed within a maximum of three freeze-thaw cycles and remained at -80°C in between thaws. The inter-assay coefficients of variability were 9.5% for leptin and 3.2% for adiponectin.

Obstetric/Perinatal Data

Six trained research midwives retrospectively extracted data from obstetric medical records and error rates were consistently <1%. These data included weight at every antenatal clinic visit (used to determine gestational weight gain), complications during pregnancy (hypertensive or diabetic disorders) and mode of delivery. Gestational age, offspring's sex and birthweight were obtained from hospital records at the time of birth. Maternal age, prepregnancy height, and weight, smoking status (defined as never smoked, smoked before but not during pregnancy and smoked during pregnancy), parity, occupational social class and highest educational attainment were obtained from questionnaires completed by the mothers in early and advanced stages of pregnancy. Occupation was used to allocate social class groups using the 1991 British Office of Population and Census Statistics classification.

Offspring Childhood and Adolescent Adiposity Measurements

Identical protocols were used at all follow-up clinics. At each clinic assessment participants' age in months was recorded and their weight and height measured in light clothing and without shoes. Weight was measured to the nearest 0.1kg using Tanita scales. Height was measured to the nearest 0.1cm using a Harpenden stadiometer. DXA scans were used to measure total fat mass. Waist circumference was measured to the nearest 1mm at the midpoint between the lower ribs and the pelvic bone with a flexible tape and with the child breathing normally. Offspring obesity was classified using BMI and criteria defined by the International Obesity Task Force (19).

Statistical Analysis

The relation between exposures (birthweight and cord blood adipokines) and outcomes (BMI, waist circumference, and fat mass at ages 9-11 years and 15-17 years) was examined by Spearman correlation. Linear (offspring BMI, waist circumference, and fat mass) and logistic (offspring obesity) regression models were used to examine the associations between birthweight and cord blood measures and offspring BMI, waist circumference, fat mass and obesity at age 9 and 17 years. Offspring waist circumference and fat mass were log transformed to produce approximately normal distributions of regression model residuals. Within cohort logged fat mass and waist circumference z-scores (participant value minus mean for the sex and age category ÷ standard deviation for the sex and age category) were created using one year age categories. BMI z-scores were created using the UK 1990 British growth reference (20). Birthweight was adjusted for sex, gestational age and number of offspring (singletons or twins) using nonlinear regression fitting a Gompertz curve.

Three incremental analyses were performed to adjust for potential confounders (Supplemental Figure 1). The basic model (model 1) adjusted for offspring sex and age at outcome measurement alone (and offspring height when fat mass is the outcome). In model 2 we additionally adjusted for maternal confounders (age, smoking, parity, occupational social class, education, and pre-pregnancy BMI). In the fully adjusted model (model 3) we

additionally adjusted for pregnancy characteristics (gestational age at birth, mode of delivery, gestational weight gain, hypertensive and diabetic disorders of pregnancy). In these analyses since we have scaled the exposures (birthweight, cord blood leptin, and adiponectin) and outcomes (BMI, waist, and fat mass) on their standard deviations the resultant differences in means from the multivariable linear regression models are equivalent to partial (adjusted) correlation coefficients and can be interpreted in this way.

There were small amounts of missing data on some co-variables included in the multivariable models (Figure 1). Twenty imputation data sets were generated by chained equations (21), with all cord exposures, birthweight, the covariates specified for model 3 and the measurements from the 11-year clinic and 15-year clinic informing imputation of missing values in the 9-year clinic and 17-year clinic respectively. For convenience hereafter referred to as 9 and 17-year. The distributions of observed and imputed variables were similar (Supplemental Table 1). In the main paper, we present results from the imputed datasets and for present comparison results from those with complete confounders (N = 1041 to 1776) in Supplementary material (Supplemental Tables 5-8)

All statistical analyses were performed using Stata (version 13.0) software (Stata Inc., College Station, TX.).

Results

Table 1 summarizes the maternal and offspring characteristics for those participants with cord blood measures, who completed at least one clinic assessment, with Supplemental Table 1 demonstrating the similarity of the observed and imputed data. Supplemental Table 2 shows the Spearman's correlation between exposures (birthweight and cord blood adipokines) and outcomes (markers of anthropometry at age 9 and 17). Birthweight was positively correlated with cord blood leptin (n=4751, r=0.33) and, to a lesser degree, with cord blood adiponectin (n=4707, r=0.14). Cord leptin and adiponectin were positively correlated (n=4962, r=0.11). Birthweight and leptin also positively correlated with fat mass, BMI and waist circumference at age 9 and 17. There was a weak inverse association between cord adiponectin and waist circumference and BMI at age 9. Among those participants with assessments at both clinics (at age 9 and 17), measurements at each clinic were highly correlated (0.74 for BMI, 0.74 for fat mass and 0.66 for waist circumference).

Table 2 shows the multivariable associations between cord blood leptin, adiponectin and birthweight and z-scores of offspring fat mass, waist circumference, BMI and obesity at age 9 years. Cord blood leptin was positively associated with fat mass, waist circumference, and BMI at age 9 (model 1). The effect size was largely attenuated with adjustment for maternal and pregnancy characteristics (Table 2), with the individual univariate association of maternal and pregnancy characteristics on cord leptin, cord adiponectin and birthweight shown in Supplemental Table 3. A similar but weaker pattern was observed for measures at age 17 where cord leptin was associated with z-scores of fat mass, waist circumference, and BMI and with the risk of obesity (Table 3). These associations were similarly attenuated to the null after adjustment for potential confounders.

Cord blood adiponectin was not associated with any measures of adiposity at age 9 (Table 2). At age 17, cord blood adiponectin was positively associated with fat mass and waist circumference, with the effect size strengthened after adjustment for maternal and pregnancy characteristics (Table 3).

Birthweight was positively associated with fat mass, waist circumference and BMI at age 9 years and 17 years and showed a weak relationship with obesity in both age groups (Tables 2 and 3). After adjustment for maternal and pregnancy characteristics increasing birthweight remained associated with greater waist circumference and BMI, with the association with fat mass and obesity attenuated to the null.

Results did not differ substantially when absolute measures of adiposity at age 9 (Supplemental Table 4) or age 17 were considered (Supplemental Table 5). Results were similar for non-imputed analyses but with wider confidence intervals (Supplemental Tables 6-9).

Discussion

In this prospective birth cohort study, cord leptin, a marker of neonatal fat mass, exhibited relatively weak relationships with later measures of adiposity. These were largely attenuated by adjustment for maternal factors, particularly in later childhood. By contrast, adiponectin exhibited no relationship with measures of fat mass at age 9 and showed a weak relationship with fat mass and waist circumference at ages 15. Neither cord leptin nor adiponectin was associated with the risk of being classed as obese in late childhood or adolescence. Taken together this would suggest that neonatal fat mass per se has a limited contribution in determining fat mass in adolescence.

To date birthweight, as a proxy for intrauterine growth, and its' relation to adult BMI has been extensively studied. Similar to our findings, studies principally demonstrate a positive association between birthweight and childhood and adult fat mass, BMI and waist circumference (22). To try to examine whether birthweight is simply acting as a surrogate for neonatal fat mass, we previously utilized ponderal index (birth weight/length³), a measure of fatness and demonstrated positive associations with lean body mass, total body fat and the fat-to-lean mass ratio at age 9-years (23). Although this suggests that neonatal fat mass is related to later adiposity, ponderal index is a relatively poor measure of neonatal total body fat (24).

To extend and improve on this work, the current study utilized cord blood leptin, a strong correlate of neonatal fat mass as assessed by skinfolds or total body electrical conductivity (25) and adiponectin, which in mouse studies is suggested to be a further positive correlate of fat mass (16). That cord blood leptin was positively associated with several adiposity measures and specifically fat mass z-score at age 9-years, suggests that there is either accretion of adipose tissue during intrauterine life that is maintained throughout childhood, the propensity to develop fat mass may be maintained, or there is a direct effect on the programming of hypothalamic feeding circuits. However, given our observed effect size, the contribution of neonatal fat to later fat mass is likely to be small. For example, a 10pg/ml increase in cord leptin would be associated with a BMI increase from 22 to 22.1kg/m² at age 9-years.

In accordance with some (26-28) but not all (29,30) previous studies we observed that adiponectin was weakly positively correlated with birthweight and cord leptin. We found some evidence for weak associations of cord blood adiponectin with adiposity at the older age (15-17) but none that this was mediated by increased (and persistent) fat mass through childhood. Why adiponectin is not related to adiposity outcomes in earlier childhood, as leptin is, is not clear. Perhaps these associations emerge after puberty which has a major impact on body composition and adipocyte number(31). It is also possible that given the multiple tests performed; some associations are due to chance, and we would caution against assuming these associations are real without further replication.

As previously shown in this cohort (32), we observed consistent positive associations of birth weight with later BMI and waist in both early childhood and adolescence, though null associations (coefficients equal to zero) were found for fat mass at both ages.

Our study has several strengths including its size, duration of follow-up, and the availability of data on a range of maternal, pregnancy and social factors pregnancy characteristics to facilitate a robust analysis. This is also one of the very few studies with DXA measurements of body composition at different time points, thereby overcoming the potential increase in overall mass attributed to the expected increase in bone density that results from increased adiposity. We do however acknowledge some limitations. The number of children who were overweight or obese was smaller than many contemporary populations. That birthweight and cord measures were not associated with the risk of being obese may reflect this. Another limitation of the study is the loss to follow-up. Our results may be biased if associations were substantially different among excluded participants due to conditioning on the variables in the model. We acknowledge that engaged participants may exhibit different characteristics at birth beyond gestational age and birthweight which are representative of the whole cohort, and also for the two outcomes. Replication of our analyses in additional birth cohorts with different metabolic risk profiles would strengthen our findings. Cord blood sample degradation may have contributed to variability, but leptin and adiponectin do appear to be stable with long-term storage (33-38). This is in stark contrast to c-peptide the preferred index of fetal glucose exposure, which we were unable to measure accurately due to degradation with long-term storage, a phenomenon previously reported by others (39).

In conclusion, we found that cord blood leptin and adiponectin, known surrogates of fetal fat mass, were weakly positively associated with some measures of fat mass in late childhood and adolescence. That these associations were robust to a wide range of confounders that may reflect intrauterine, maternal and shared environmental exposures suggests that neonatal fat mass may track into later life. However, we acknowledge replication of our findings in cohorts with a different risk profile is critical, and that the magnitude of the observed associations is small, potentially limiting the impact that neonatal life adiposity has on later outcomes.

Acknowledgments

We are grateful to all the families who took part in this study, the midwives for their help in recruiting them, and the whole Avon Longitudinal Study of Parents and Children team, which includes interviewers, computer and laboratory technicians, clerical workers, research scientists, volunteers, managers, receptionists, and nurses.

Corresponding author: Professor Scott M Nelson, School of Medicine, University of Glasgow, Room 2.52 Level 2, New Lister Building, Glasgow Royal Infirmary, Glasgow, G31 2ER, Tel: +44 141 201 8581, Email: scott.nelson@glasgow.ac.uk

Sources of funding

This work was funded by a grant from the Wellcome Trust (WT094311MA). J Simpson is funded by a Wellbeing of Women Research Training Fellowship. A. Fraser is funded by a UK Medical Research Council research fellowship (Grant ref: MR/M009351/1). The UK Medical Research Council (Grant ref: 102215/2/13/2), the Wellcome Trust (Grant ref: WT076467), and the University of Bristol provide core funding support for Avon Longitudinal Study of Parents and Children. The UK Medical Research Council (G0600705) and the University of Bristol provide core funding for the MRC Integrative Epidemiology Unit (grants MC UU 1201/1/5, MC UU 1201/1, and MC UU 1201/4).

Author Contributions

JS performed the laboratory cord blood analysis, contributed to statistical analysis, participated in data interpretation and drafted the manuscript. AS contributed to the statistical analysis and data interpretation. AF, NS, RL, SR, GDS and DAL contributed to obtaining funding and data interpretation. SMN conceived the study, obtained funding, contributed to the statistical analysis, data interpretation and drafted the manuscript. All authors contributed to the preparation of the manuscript and approved the final version.



Conflict of Interest: none

Guarantor: Professor Scott Nelson

References

Nelson SM, Matthews P, Poston L. Maternal metabolism and obesity: modifiable 1. determinants of pregnancy outcome. Hum Reprod Update 2009; 16:255-275

Lawlor DA, Relton C, Sattar N, Nelson SM. Maternal adiposity -- a determinant of 2. perinatal and offspring outcomes? Nature reviews Endocrinology 2012; 8:679-688

Lawlor DA. The Society for Social Medicine John Pemberton Lecture 2011. 3. Developmental overnutrition—an old hypothesis with new importance? International Journal of Epidemiology 2013; 42:7-29

4. Tyrrell J, Richmond RC, Palmer TM, Feenstra B, Rangarajan J, Metrustry S, Cavadino A, Paternoster L, Armstrong LL, De Silva NM, Wood AR, Horikoshi M, Geller F, Myhre R, Bradfield JP, Kreiner-Moller E, Huikari V, Painter JN, Hottenga JJ, Allard C, Berry DJ, Bouchard L, Das S, Evans DM, Hakonarson H, Haves MG, Heikkinen J, Hofman A, Knight B, Lind PA, McCarthy MI, McMahon G, Medland SE, Melbye M, Morris AP, Nodzenski M, Reichetzeder C, Ring SM, Sebert S, Sengpiel V, Sorensen TI, Willemsen G, de Geus EJ, Martin NG, Spector TD, Power C, Jarvelin MR, Bisgaard H, Grant SF, Nohr EA, Jaddoe VW, Jacobsson B, Murray JC, Hocher B, Hattersley AT, Scholtens DM, Davey Smith G, Hivert MF, Felix JF, Hypponen E, Lowe WL, Jr., Frayling TM, Lawlor DA, Freathy RM. Genetic Evidence for Causal Relationships Between Maternal Obesity-Related Traits and Birth Weight. Jama 2016; 315:1129-1140

Jain V, Kurpad AV, Kumar B, Devi S, Sreenivas V, Paul VK, Body composition of 5. term healthy Indian newborns. European journal of clinical nutrition 2015;

Sewell MF, Huston-Presley L, Super DM, Catalano P. Increased neonatal fat mass, 6. not lean body mass, is associated with maternal obesity. Am J Obstet Gynecol 2006; 195:1100-1103

Hauguel-de Mouzon S, Lepercq J, Catalano P. The known and unknown of leptin in 7. pregnancy. Am J Obstet Gynecol 2006; 194:1537-1545

Catalano PM, Presley L, Minium J, Hauguel-de Mouzon S. Fetuses of obese mothers 8. develop insulin resistance in utero. Diabetes Care 2009; 32:1076-1080

Lawlor DA, West J, Fairley L, Nelson SM, Bhopal RS, Tuffnell D, Freeman DJ, 9. Wright J, Whitelaw DC, Sattar N. Pregnancy glycaemia and cord-blood levels of insulin and leptin in Pakistani and white British mother-offspring pairs: findings from a prospective pregnancy cohort. Diabetologia 2014; 57:2492-2500

10. Bouret SG. Nutritional programming of hypothalamic development: critical periods and windows of opportunity. International journal of obesity supplements 2012; 2:S19-24

Mantzoros CS, Rifas-Shiman SL, Williams CJ, Fargnoli JL, Kelesidis T, Gillman 11. MW. Cord Blood Leptin and Adiponectin as Predictors of Adiposity in Children at 3 Years of Age: A Prospective Cohort Study. Pediatrics 2009; 123:682-689

Boeke CE, Mantzoros CS, Hughes MD, S LR-S, Villamor E, Zera CA, Gillman MW. 12. Differential associations of leptin with adiposity across early childhood. Obesity (Silver Spring) 2013; 21:1430-1437

13. Nakano Y, Itabashi K, Maruyama T. Association between serum adipocytokine and cholesterol levels in cord blood. Pediatrics International 2009; 51:790-794

Lindsay RS, Nelson SM, Walker JD, Greene SA, Milne G, Sattar N, Pearson DW. 14. Programming of Adiposity In Offspring Of Mothers With Type 1 Diabetes At Age 7. Diabetes Care 2010; 33:1080 - 1085

15. Aye ILMH, Powell TL, Jansson T. Review: Adiponectin – The missing link between maternal adiposity, placental transport and fetal growth? Placenta 2013; 34, Supplement:S40-S45

16. Oiao L, Yoo Hs, Madon A, Kinney B, Hay WW, Shao J. Adiponectin Enhances Mouse Fetal Fat Deposition. Diabetes 2012; 61:3199-3207

Boyd A, Golding J, Macleod J, Lawlor DA, Fraser A, Henderson J, Molloy L, Ness 17. A, Ring S, Davey Smith G. Cohort Profile: The 'Children of the 90s'-the index offspring of the Avon Longitudinal Study of Parents and Children. International Journal of Epidemiology 2013; 42:111-127

18. Fraser A, Macdonald-Wallis C, Tilling K, Boyd A, Golding J, Davey Smith G, Henderson J, Macleod J, Molloy L, Ness A, Ring S, Nelson SM, Lawlor DA. Cohort Profile: the Avon Longitudinal Study of Parents and Children: ALSPAC mothers cohort. Int J Epidemiol 2013; 42:97-110

19. Cole TJ, Bellizzi MC, Flegal KM, Dietz WH. Establishing a standard definition for child overweight and obesity worldwide: international survey. BMJ (Clinical research ed) 2000; 320:1240-1243

20. Cole TJ, Freeman JV, Preece MA. British 1990 growth reference centiles for weight, height, body mass index and head circumference fitted by maximum penalized likelihood. Statistics in medicine 1998; 17:407-429

21. White IR, Royston P, Wood AM. Multiple imputation using chained equations: Issues and guidance for practice. Statistics in medicine 2011; 30:377-399

Brisbois TD, Farmer AP, McCargar LJ. Early markers of adult obesity: a review. 22. Obes Rev 2012; 13:347-367

Rogers IS, Ness AR, Steer CD, Wells JCK, Emmett PM, Reilly JR, Tobias J, Smith 23. GD. Associations of size at birth and dual-energy X-ray absorptiometry measures of lean and fat mass at 9 to 10 y of age. Am J Clin Nutr 2006; 84:739-747

de Bruin NC, van Velthoven KA, Stijnen T, Juttmann RE, Degenhart HJ, Visser HK. 24. Body fat and fat-free mass in infants: new and classic anthropometric indexes and prediction equations compared with total-body electrical conductivity. Am J Clin Nutr 1995; 61:1195-1205

Okereke NC, Uvena-Celebrezze J, Hutson-Presley L, Amini SB, Catalano PM. The 25. effect of gender and gestational diabetes mellitus on cord leptin concentration. Am J Obstet Gynecol 2002; 187:798-803

Sivan E, Mazaki-Tovi S, Pariente C, Efraty Y, Schiff E, Hemi R, Kanety H. 26. Adiponectin in Human Cord Blood: Relation to Fetal Birth Weight and Gender. J Clin Endocrinol Metab 2003; 88:5656-5660

Tsai PJ, Yu CH, Hsu SP, Lee YH, Chiou CH, Hsu YW, Ho SC, Chu CH. Cord 27. plasma concentrations of adiponectin and leptin in healthy term neonates: positive correlation with birthweight and neonatal adiposity. Clinical Endocrinology 2004; 61:88-93

Kotani Y, Yokota I, Kitamura S, Matsuda J, Naito E, Kuroda Y. Plasma adiponectin 28. levels in newborns are higher than those in adults and positively correlated with birth weight. Clinical Endocrinology 2004; 61:418-423

Lindsay RS, Walker JD, Havel PJ, Hamilton BA, Calder AA, Johnstone FD. 29. Adiponectin is present in cord blood but is unrelated to birth weight. Diabetes Care 2003; 26:2244-2249

Nelson SM, Freeman DJ, Sattar N, Lindsay RS. Role of adiponectin in matching of 30. fetal and placental weight in mothers with type 1 diabetes. Diabetes Care 2008; 31:1123-1125

31. Knittle JL, Timmers K, Ginsberg-Fellner F, Brown RE, Katz DP. The growth of adipose tissue in children and adolescents. Cross-sectional and longitudinal studies of adipose cell number and size. J Clin Invest 1979; 63:239-246



32. Anderson EL, Howe LD, Fraser A, Callaway MP, Sattar N, Day C, Tilling K, Lawlor DA. Weight trajectories through infancy and childhood and risk of non-alcoholic fatty liver disease in adolescence: the ALSPAC study. J Hepatol 2014; 61:626-632

33. Flower L, Ahuja RH, Humphries SE, Mohamed-Ali V. EFFECTS OF SAMPLE HANDLING ON THE STABILITY OF INTERLEUKIN 6, TUMOUR NECROSIS FACTOR-α AND LEPTIN. Cytokine 2000; 12:1712-1716

34. Gislefoss RE, Grimsrud TK, Morkrid L. Long-term stability of serum components in the Janus Serum Bank. Scandinavian journal of clinical and laboratory investigation 2008; 68:402-409

35. Shih WJ, Bachorik PS, Haga JA, Myers GL, Stein EA. Estimating the long-term effects of storage at -70 degrees C on cholesterol, triglyceride, and HDL-cholesterol measurements in stored sera. Clinical chemistry 2000; 46:351-364

36. Boyanton BL, Jr., Blick KE. Stability studies of twenty-four analytes in human plasma and serum. Clinical chemistry 2002; 48:2242-2247

37. Brinc D, Chan MK, Venner AA, Pasic MD, Colantonio D, Kyriakopolou L, Adeli K. Long-term stability of biochemical markers in pediatric serum specimens stored at -80 degrees C: a CALIPER Substudy. Clinical biochemistry 2012; 45:816-826

38. Paltiel L, Ronningen KS, Meltzer HM, Baker SV, Hoppin JA. Evaluation of Freeze Thaw Cycles on stored plasma in the Biobank of the Norwegian Mother and Child Cohort Study. Cell preservation technology 2008; 6:223-230

39. Gislefoss RE, Grimsrud TK, Morkrid L. Stability of selected serum proteins after long-term storage in the Janus Serum Bank. Clinical chemistry and laboratory medicine : CCLM / FESCC 2009; 47:596-603

Figure 1: ALSPAC participant flow chart.

Maternal Characteristics	Attended at least one clinic assessment (n=2955)						
	N obs (%)	Median (IQR)					
Age	2914	29 (26, 32)					
Smoking							
Never	2103 (73.8)						
Before, not during pregnancy	212 (7.4)						
During pregnancy	533 (18.7)						
BMI	2587	22.2 (20.5, 24.4)					
Parity							
0	1274 (45.5)						
1	1011 (36.1)						
2	383 (13.7)						
3	101 (3.6)						
4+	30 (1.1)						
Education							
Left school at 16	1713 (61.3)						
A level	689 (24.8)						
Degree	391 (14.0)						
Social Class							
I (least disadvantaged)	140 (5.9)						
II	807 (33.7)						
IIIa	1038 (43.5)						
IIIb	162 (6.8)						
IV	203 (8.5)						
V (most disadvantaged)	40 (1.7)						
Pregnancy Characteristics							
Gestational age at birth (weeks)	2914	40 (39, 41)					
Model of delivery							
Spontaneous	2253 (77.9)						

Table 1: Maternal and Offspring Characteristics

Breech	36 (1.3)	
Caesarean	249 (8.6)	
Forceps	167 (5.8)	
Vacuum	154 (5.3)	
Other	32 (1.1)	
Gestational weight gain (kg)	2668	12.5 (9.5, 15.2)
Hypertension and pre-eclampsia		
No hypertensive disorders	2449 (84.5)	
Hypertension, no pre-eclampsia	420 (13.9)	
Hypertension and pre-eclampsia	49 (1.7)	
Diabetes		
No glycosuria or diabetes	2651 (95.8)	
Existing diabetes	10 (0.4)	
Gestational diabetes	16 (0.6)	
Glycosuria	91 (3.3)	
Offspring Characteristics		
Sex		
Male	1414 (47.9)	
Female	1541 (52.2)	
Birthweight (kg)	2891	3.5 (3.1, 3.8)
Cord leptin (pg/ml)	2952	6.4 (3.6, 12.1)
Cord adiponectin (µg/ml)	2927	75.7 (53.6, 98.4)
Height (cm)	Age 9: 2561	140 (136, 144)
-	Age 11: 2363	151 (146, 156)
	Age 15: 1816	169 (163, 175)
	Age 17: 1648	170(164, 178)
Fat mass (kg)	Age 9: 2460	7.3 (4.9, 11.2)
	Age 11: 2327	10.0 (6.8, 15.7)
	Age 15: 1716	13.7 (8.6, 20.6)
	Age 17: 1594	16.7 (11.0, 23.5)
Waist circumference (cm)	Age 9: 2574	61.1 (57.4, 66.6)
	Age 11: 2362	66.0 (61.8, 73.5)
	Age 15: 1475	75.4 (71.0, 81.5)
BMI (kg/m ²)	Age 9: 2560	17.0 (15.7, 19.1)
	Age 11: 2359	18.4(16.6, 21.0)
	Age 15: 1811	20.7 (19.0, 23.1)
	Age 17: 1647	22.0 (20.2, 24.7)
Obese	Age 9: 102 (4.0)	
	Age 11:116 (4.9)	
	Age 15: 78 (4.3)	
	Age 17:105 (6.4)	
Age at clinic attendance (years)	Age 9: 2583	9.8 (9.6, 10.0)
- /	Age 11: 2378	11.8 (11.6, 11.8)
	Age 15: 1838	15.4 (15.3, 15.6)
	Age 17: 1695	17.8 (17.6, 17.9)

Median (Interquartile range)

Figures are numbers (%) unless stated otherwise

THE JOURNAL OF CLINICAL ENDOCRINOLOGY & METABOLISM	
\geq	

Table 2: Associations of birthweight and cord blood analyte with fat mass	, waist circumference and BMI z-scores, and obesity outcome at age 9
years. N= 2775	

	Outcome	utcome Fat mass z-score *				Waist circumference z-score			BMI z-score			Obesity		
Exposure		Coefficient	95% CI	Р	Coefficient	95% CI	Р	Coefficient	95% CI	Р	OR	95% CI	Р	
Leptin (per	Model 1	0.07	0.04, 0.10	< 0.001	0.08	0.05, 0.12	< 0.001	0.11	0.07, 0.15	< 0.001	1.15	1.00, 1.31	0.046	
10p5/111)	Model 2	0.04	0.00, 0.07	0.023	0.05	0.01, 0.08	0.008	0.06	0.02, 0.10	0.003	1.00	0.85, 1.17	0.993	
	Model 3	0.03	0.00, 0.06	0.086	0.04	0.00, 0.07	0.045	0.04	0.00, 0.08	0.029	0.95	0.81, 1.12	0.548	
Adiponectin (per 10µg/ml)	Model 1	0.00	-0.01, 0.01	0.828	-0.01	-0.02, 0.00	0.072	0.00	-0.02, 0.01	0.602	0.99	0.94, 1.05	0.845	
	Model 2	0.00	-0.01, 0.01	0.916	-0.01	-0.02, 0.00	0.118	0.00	-0.01, 0.01	0.858	1.00	0.94, 1.05	0.874	
	Model 3	0.00	-0.01, 0.01	0.875	-0.01	-0.02, 0.00	0.100	0.00	-0.01, 0.01	0.767	0.99	0.94, 1.05	0.834	
Birthweight‡ (per 100g)	Model 1	0.01	0.00, 0.02	0.006	0.03	0.03, 0.04	< 0.001	0.04	0.03, 0.05	< 0.001	1.06	1.02, 1.10	0.006	
	Model 2	0.00	0.00, 0.01	0.192	0.03	0.02, 0.04	< 0.001	0.04	0.03, 0.04	< 0.001	1.03	0.99, 1.07	0.193	
	Model 3	0.00	-0.01, 0.01	0.741	0.02	0.02, 0.03	< 0.001	0.03	0.02, 0.04	< 0.001	1.01	0.96, 1.05	0.852	

Model 1: Adjusted for offspring sex and age at measurement.

Model 2: Adjusted for offspring sex, age at measurement and maternal confounders (age, smoking, parity, occupational social class, education and pre-pregnancy BMI). Model 3: Adjusted for offspring sex, age at measurement and maternal confounders plus pregnancy confounders (gestational age at birth, mode of delivery, gestational weight gain, hypertensive disorders and diabetic disorders of pregnancy).

* Fat mass adjusted for height

‡ Birthweight adjusted for sex, gestational age and singleton/twin pregnancy

	Outcome	Fa	t mass z-score *		Waist circumference z-score			BMI z-score			Obesity		
Exposure		Coefficient	95% CI	Р	Coefficient	95% CI	Р	Coefficient	95% CI	Р	OR	95% CI	Р
Leptin (per	Model 1	0.07	0.03, 0.11	<0.001	0.06	0.02, 0.10	0.003	0.09	0.04, 0.14	< 0.001	1.13	0.99, 1.28	0.060
10pg/111)	Model 2	0.02	-0.02, 0.06	0.263	0.01	-0.03, 0.05	0.545	0.03	-0.02, 0.07	0.272	0.96	0.83. 1.12	0.629
	Model 3	0.02	-0.02, 0.05	0.444	0.01	-0.03, 0.05	0.598	0.02	-0.03, 0.06	0.481	0.95	0.81, 1.11	0.497
Adiponectin (per 10µg/ml)	Model 1	0.01	0.00, 0.03	0.034	0.01	0.00, 0.03	0.033	0.01	-0.01, 0.02	0.245	1.03	0.98, 1.08	0.238
	Model 2	0.02	0.00, 0.03	0.006	0.02	0.00, 0.03	0.008	0.01	0.00, 0.03	0.076	1.04	0.99, 1.10	0.660
	Model 3	0.02	0.00, 0.03	0.007	0.02	0.00, 0.03	0.008	0.01	0.00, 0.03	0.080	1.05	0.99, 1.10	0.613

Table 3: Associations of birthweight and cord blood analyte with fat mass, waist circumference (at age 15 years), BMI z-scores and obesity outcomes at age 17 years. N= 2138

EARLY RELEASE:

Birthweight‡ (per 100g)	Model 1	0.02	0.02, 0.03	< 0.001	0.04	0.03, 0.05	< 0.001	0.04	0.03, 0.05	< 0.001	1.05	1.02, 1.09	0.004
	Model 2	0.01	0.00, 0.02	0.010	0.03	0.02, 0.04	< 0.001	0.02	0.01, 0.03	< 0.001	1.02	0.98, 1.06	0.241
	Model 3	0.01	0.00, 0.02	0.098	0.03	0.02, 0.04	< 0.001	0.02	0.01, 0.03	< 0.001	1.01	0.97, 1.05	0.516

Model 1: Adjusted for offspring sex and age at measurement.

Model 2: Adjusted for offspring sex, age at measurement and maternal confounders (age, smoking, parity, occupational social class, education and pre-pregnancy BMI). Model 3: Adjusted for offspring sex, age at measurement and maternal confounders plus pregnancy confounders (gestational age at birth, mode of delivery, gestational weight gain, hypertensive disorders and diabetic disorders of pregnancy).

* Fat mass adjusted for height

‡ Birthweight adjusted for sex, gestational age and singleton/twin pregnancy

EARLY



{individualUser.displayName}] on 22 November 2016. at 03:50 For perso