

Citation for published version:
Edwards, M, Saglani, S, Schwarze, J, Skevaki, C, Smith, J, Ainsworth, B, Almond, M, Andreakos, E, Belvisi, M, Chung, KF, Cookson, W, Cullinan, P, Hawrylowicz, C, Lommatzsch, M, Jackson, D, Lutter, R, Marsland, B, Moffat, M, Thomas, M, Virchow, JC, Xanthou, G, Edwards, J, Walker, S, Johnston, SL & Group, EARIPWPW 2017, Addressing unmer needs in understanding asthma mechanisms, European Respiratory Journal, vol. 49, pp. 1, 1602448, pp. 1, 142, https://doi.org/10.1182/12002003.00448.2016 no. 5, 1602448, pp. 1-43. https://doi.org/10.1183/13993003.02448-2016

10.1183/13993003.02448-2016

Publication date: 2017

Document Version Peer reviewed version

Link to publication

University of Bath

General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy
If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Download date: 05. Jun. 2020

© Copyright 2016, EARIP Consortium GA- 602077

Version: Final

Abstract

Asthma is a heterogeneous, complex disease with clinical phenotypes that incorporate persistent symptoms and acute exacerbations. It affects many millions of Europeans throughout their education and working lives and puts a heavy cost on European productivity. There is a wide spectrum of disease severity and control. Therapeutic advances have been slow despite greater understanding of basic mechanisms and the lack of satisfactory preventative and disease modifying management for asthma constitutes a significant unmet clinical need. Preventing, treating and ultimately curing asthma requires co-ordinated research and innovation across Europe. The European Asthma Research and Innovation Partnership (EARIP) is an FP7-funded programme which has taken a co-ordinated and integrated approach to analyse the future of asthma research and development. This report aims to identify the mechanistic areas in which investment is required to bring about significant improvements in asthma outcomes.





GA- 602077

Version: Final

1 Executive summary

This manuscript describes a state-of-the-art view of the mechanisms involved in asthma onset, asthma progression and asthma exacerbations and is an outcome of the European Asthma Research and Innovation Partnership (EARIP: www.earip.eu). Readers are also referred to companion EARIP reports including a report on phenotyping and improving precision and personalised medicine for asthma(1), a report on national and regional asthma programmes in Europe(2) and an editorial on building the case for more investment in asthma research in Europe(3).

The EARIP project was established in 2013 to harmonise efforts to reduce mortality and morbidity from asthma by agreeing the most important research priorities in Europe. The partnership has produced an evidence- and consensus-based list of the research priorities needed to reduce asthma deaths and hospitalisations(4). This coordinated agenda for asthma research and innovation will be used to address this major health and societal challenge and will be the foundation on which future EU, national and international research funding programmes can transform asthma outcomes throughout Europe.

International guidelines describe asthma as a chronic airway disease characterised by airway hyper-reactivity (AHR) caused by a range of processes including airway inflammation, bronchoconstriction of airway smooth muscle (ASM), airway infections, allergic responses, exposure to pollutants, exercise, stress, and nervous provocation(5). Asthma is one of the most common chronic long-term health conditions in Europe. It is estimated that the number of people aged 45 and under with asthma in Europe is over 30 million(6). The prevalence of asthma varies substantially across the European Union. A key feature of asthma is reversible decreases in lung function, measured either as changes in peak expiratory flow (PEF), or a decrease in forced expiratory volume in 1 second (FEV₁). Symptoms include dyspnoea, cough, wheeze and chest tightness. Asthma is a heterogeneous, complex disease with multiple clinical phenotypes that incorporate persistent symptoms and acute exacerbations with a wide spectrum of both disease severity and control (7, 8). The complex interplay between genetic and environmental influences that contribute to asthma pathogenesis makes management a significant ongoing challenge. The lack of preventative, curative and disease modifying management for asthma constitutes a significant unmet clinical need.

Asthma is most frequently associated with onset in early life, and is increasing in incidence in the developed world, but can also start in adulthood (that is late-onset asthma)(9). Current pharmacological management approaches are centered on the use of symptom relievers in the form of bronchodilators, which may be short acting or long acting and anti-inflammatory therapies, predominantly glucocorticosteroids (GCs)(10), their combinations(11, 12), leukotriene receptor antagonists (LTRAs)(13-15) and the non-selective anti-inflammatory and adenosine receptor antagonist, theophylline. Innovative novel monoclonal antibody therapies in phase II/III clinical trials or in recent





GA- 602077

Version: Final

clinical use include antagonists of T_H2 cytokines(16-20) and anti-IgE therapies (21-24). Sub-lingual and other immunotherapy approaches are also exciting, potentially disease modifying therapies(25), that require further study and further development. These have had varying levels of success at controlling asthma symptoms, improving lung function, and in particular reducing asthma exacerbation rates; however no therapy yet adequately modifies disease, is sufficiently accessible to asthma patients or could be considered a cure. Despite these advances, translating scientific progress to new treatment avenues has been slow for asthma, considering the innovations and development of therapies that have had substantial impact on disease in other fields. Significant disconnects are observed between objective disease severity parameters and patients' experience of their disease(26, 27), and outcomes are worse in those with psychosocial disadvantage(28, 29), mediated by mechanisms that remain unclear. Behavioural problems, such as non-adherence, are common even in patients with severe disease(30) and associated with worse outcomes(31).

As medical research enters the 'omics' era, a new challenge facing asthma research is how best to harness big data sets and apply them to models relevant to human disease, in order to benefit asthma treatment. This challenge is further complicated by a recent focus on asthma endotypes or phenotypes and personalised medicine applying novel mechanism-based therapies to those in which such mechanisms are active. Taking all the above into account it is timely to ask the question where next in asthma research? This report reviews current scientific knowledge (state of the art) on mechanisms of asthma onset, asthma progression from mild to severe disease and asthma exacerbation and identifies current unmet needs in asthma research to indicate where future research efforts should be focused.

The important mechanisms involved in asthma onset, asthma progression and asthma exacerbation and the strength of the supportive evidence for each mechanism are summarised in **Table 1**. The important methodologies or technologies that are required to beneficially impact on asthma research in future years are summarised in **Table 2**. Key unmet needs in understanding asthma mechanisms that we have identified are listed in **Table 3** and mechanisms of asthma onset, asthma progression from mild to severe disease and asthma exacerbation are pictorially represented in **Figures 1**, **2** and **3** respectively. These are the major areas in which future investment is needed to drive forward asthma research to hasten progress in preventing, controlling and curing asthma, and in preventing and treating asthma attacks and their associated morbidity and mortality.

In lay terms, these scientific goals can be better expressed as three broad aims that should translate into immediately understandable goals to boost:

- o Prevention (related to onset)
- o Cure or Control (related to progression)
- o Attack avoidance/treatment (related to exacerbations).





GA- 602077

Consortium Version: Final

2 Asthma onset

2.1 Introduction

Asthma onset frequently occurs early in life. Lung function is reduced by school-age(32) and this reduction is permanent and tracks to adulthood(33, 34). The key pathological abnormalities of asthma, including eosinophilic inflammation and airway remodelling also frequently develop early and may be established by school-age(35, 36). Therefore to achieve disease modification early intervention is essential. The ideal intervention would allow primary prevention. Disappointingly, even secondary prevention is currently not on the horizon.

In order to achieve primary prevention it is necessary to accurately predict which child will develop asthma. Although one third of preschool children wheeze, only a third will go on to develop asthma(37). Unfortunately, accurate prediction of asthma during the preschool years remains a significant challenge. Many clinical prediction scores, incorporating known risk factors such as atopy and family history, have been proposed(38), but they are either applicable too late; or have a poor predictive value(39). It is known that host and environmental influences interact to contribute to asthma risk.

In addition to studying disease mechanisms, following children who grow out of asthma and identifying differences between those who and those who do not at the molecular level may provide insight into mechanisms and potentially point towards a cure of asthma

A further significant problem is late onset asthma, which is frequently severe, often appears non-atopic (absent skin-prick test positivity or raised serum specific IgE) in pathogenesis and which may have other risk factors, can be quite a difficult differential diagnosis (sometimes overlapping with COPD) and may represent a different disease spectrum from early onset asthma. Also asthma can arise in adulthood as a result of occupational and irritant exposures frequently resulting in non-Th2 asthma.

Asthma is more likely to be diagnosed in people with anxiety and depression(40, 41). Research strategies therefore need to incorporate multiple risk factors and include complex interactions in order to reflect the disease and identify effective interventions(42). See **Figure 1** for a summary of important mechanisms involved in asthma onset.

2.2 Asthma onset - State of the art

2.2.1 Viruses

Viral infections represent critical events that likely contribute to asthma onset. Early life respiratory syncytial virus (RSV), human rhinovirus (RVs) and human metapneumovirus (HMPV) lower respiratory infections have all been strongly associated with progression to clinically defined asthma by age 6(43-47). However, little is known about the mechanisms underlying the progression of virus-induced early wheezing to later life well-defined asthma. Indeed, there is considerable debate over whether these virus infections are important causative agents of later asthma development, and/or are markers





GA- 602077

Version: Final

of risk for asthma development independent of causation. It is possible that different viruses fulfil different roles that are not obvious in the available cohort studies. Interactions between viral infections and underlying genetic susceptibility identified from Genome Wide Association Studies (GWAS) have been reported(45). Although this has shed light on gene-environment interactions, and identified several candidate genes with reproducible and strong associations, they remain associations and mechanistic advances are needed. For example polymorphisms spanning CDHR3 are associated with childhood asthma with severe exacerbations(48). CDHR3 encodes cadherinrelated family member 3 (CDHR3), which is involved in epithelial polarity and cell-cell interactions as well as being a receptor for rhinovirus C, the most common respiratory virus associated with asthma exacerbations and the key variant identified in GWAS modulates levels of CDHR3 providing a putative mechanism(49). Some mechanistic evidence has come from available small animal models. Infection of newborn mice with RSV, and the mouse equivalent pneumovirus, show striking long term changes to the airway and induction of pro-T_H2 responses to allergens later in life(50, 51). The interactions between virus infection with allergen exposure, and also the beneficial effects of environmental and airway microbes are important avenues to pursue for future research. (see Table 2 for an explanation of new methodologies in asthma research)

2.2.2 Allergen exposure & T_H2 pathways

T helper $(T_H)2$ immunity mediated by interleukin (IL)s -4, -5, -9 and -13 after sensitisation and exposure to allergens drives allergic asthma, one of the major subtypes of asthma, inducing IgE production, eosinophilia and tissue remodelling(52). Targeted therapies in subtypes of established T_H2 -mediated asthma have been successful in reducing exacerbations(53, 54). Interventions to prevent initial sensitisation may provide a long-lasting alternative to T_H2 -targeted therapies. As yet, initiating processes are incompletely understood, but likely involve dendritic cells (DCs), basophils, eosinophils, innate lymphoid cells (ILCs) and macrophages and airway epithelial cells secreting the newly identified pro- T_H2 cytokines IL-33, TSLP and IL-25, which thus may be potential alternative candidates for intervention.

ILCs are of particular interest as they have been recently described as tissue-resident and circulating innate immune cells(55), with three major subsets: ILC1s, ILC2s and ILC3s, which mirror T_H1 , T_H2 and T_H17 cells, respectively(56). There is little detailed information on the role of ILCs in asthma onset and progression, though roles for both IL-33 and IL-25, which act as growth factors and activators of ILC2s, have recently been proposed in acute exacerbations(57, 58). A particular shortcoming in our present understanding is information from experimental studies in humans.

Mouse models of asthma have provided evidence for ILC2s in development of allergic asthma(59-61) and ILC3s in a model of obesity-associated AHR(62). Whether these data can be translated to asthma depends on whether these models truly reflect human disease. In addition, asthma is a heterogeneous disease with respect to onset (early *versus* late), response to treatments and





Version: Final

GA-602077

concomitant pathology (allergic *versus* non-allergic, obesity), which likely reflects differences in underlying pathobiology. So, the contribution of ILCs may vary between the various subtypes of asthma. There is evidence that IL-33, IL-25 and TSLP, and their receptors, are involved in response to allergens and respiratory viruses(57, 58). This work provides at least preliminary evidence for a role of ILCs in asthma, as well as other innate and adaptive immune cells like eosinophils, basophils(63), mast cells and T cells, which may also be responsive to IL-33, IL-25 and TSLP.

2.2.3 Air pollution

The role of air pollution in asthma development is unclear (64). Three European birth cohort studies have reported positive relationships between traffic-related pollution and doctor-diagnosed asthma(65-67). The Southern Californian Children's Health Study has reported that traffic-related pollutants can cause asthma in children(68, 69), as has a Dutch study in which trafficrelated pollution levels at the birth address and incidence of asthma were considered during the first 8 years of life(70). Exposure to nitrogen dioxide (NO₂) and particulate matter (PM)_{2.5} at birth correlated with asthma development in childhood and adolescence(71) and a 5ppb increase in NO2 levels during the first year of life with physician-diagnosed asthma (OR: 1·17)(72). In women, a 3·6µg/m³ increase in PM_{2·5} correlated with greater odds (OR 1·20) of incident asthma, whereas a 3·6µg/m³ increase in PM_{2.5} (OR 1·14) and 5·8ppb increase in NO₂ (OR 1·08) correlated with incident wheeze(73). Not all studies, however, show such a relationship(74), and such inconsistencies may be due to incomplete exposure assessment or insufficient study power.

A recent report from the Royal College of Physicians London summarized the state of the art and also states that exposure of young children to second-hand tobacco smoke remains one of the most important sources of indoor air pollution and is associated with asthma prevalence, although the effects of exposure to maternal smoking in pregnancy may be stronger than for childhood exposure(75). Occupational exposures as well as second-hand smoking may also be related to late onset adult asthma.

Host characteristics that have been reported to influence the effects of air pollution on asthma development include nutritional status, atopy, and social stress(76-82). Polymorphisms in glutathione S-transferase (GSTP1) that facilitates the elimination of reactive oxygen species have also been associated with a greater risk of asthma(83) and sensitisation to allergen in association with traffic-related NOx during the first year of life (84). As the environment is a potentially modifiable factor in asthma development(64), funding is urgently needed to study the effects of the environment on asthma development, to reveal new approaches to reduce future development of asthma.

2.2.4 Bacteria & the microbiome

During and immediately following birth neonates are exposed to microbes that progressively colonize the gut, the skin and airways. Indeed, within a very





GA- 602077

Version: Final

short time frame post-partum, one moves from being 100% human, to a human super organism that lives with vast numbers of microbes(85, 86). Although much research to-date has focused on the gut, host-microbe interactions in the lung may also be powerful determinants of the presence or absence of asthma(87).

The human microbiome describes the totality of microorganisms (bacteria, viruses, fungi and parasites) associated with the human body, in both health and disease. Molecular techniques that include 16S rRNA gene sequencing, microbial whole genome sequencing and metagenomics are revolutionising our understanding of the diversity and function of the human microbiome(88, 89). The lower airways are now known to carry a characteristic community of microorganisms (90-92) with a similar density of bacteria to the upper intestines(91).

Presence of certain bacteria may be protective or harmful. A rich microbial environment in early life is associated with protection against development of asthma and allergies(93-95) and the protection appears to be mediated via signals transmitted via innate immune signalling(95). Molecular studies have since shown that particular genera, such as Bacteroidetes and Actinobacteria appear to be protective(91, 96). In support of the above, experimental animal models have shown that the early life formation of the lung microbiome provides signals to the immune system that shape its maturation and influences susceptibility to asthma(97-99). Several studies have reported that past exposure to tuberculosis or BCG vaccination may be associated with protection from development of asthma and allergies(100, 101), though others do not agree(102) and causal relationships in man are difficult to establish(103). Preclinical studies with BCG and bacterial products derived from *Mycobacterium tuberculosis* have shown protection from eosinophil recruitment and AHR in mouse models of allergic airway inflammation(104, 105).

It is clear that the microbiome influences the trajectory towards or away from asthma(106), so how could this be harnessed to reduce the onset of asthma? Spending the first year of life in a farming environment leads to a reduced risk of asthma and allergies (107). This protection is associated with increased microbial diversity(94), and could be related to bacterial components that are inhaled, as exposure of the lung to farm dust and endotoxin protect against house dust mite-induced asthma through A20 induction in lung epithelial cells(108) or to the constituents of the raw cow's milk(109). This latter observation may be particularly relevant from the standpoint of future interventions. In addition to a high microbial load, raw cow's milk contains a plethora of 'prebiotics', that is to say 'energy sources' for bacteria, which can support bacterial growth and function in the gut. Early life nutrition, and perhaps maternal nutrition, are key areas where more research is needed in order to identify strategies through which early life microbial exposure and colonization can be shaped. Answering which microbial signals, when and how we can provide them, holds the potential for reducing the onset of asthma.





© Copyright 2016,

Version: Final

EARIP Consortium

Molecular investigations also reveal that the bronchial microbiota in asthma is rich in pathogens(91, 110-114) (typically Proteobacteria such as Moraxella, Neisseria and Haemophilus spp.). Birth cohort studies have shown that the presence of the same pathogens in throat swabs predicts the later development of asthma(115), and that these bacteria have been associated with asthma exacerbations(116). A recent study applying 16S rRNA sequencing of the nasopharynx microbiome of 234 children in their first year of life showed that transient infections of Streptococcus, Moraxella, or Haemophilus were associated with virus-associated acute respiratory infections. Importantly, asymptomatic infection with Streptococccus was a predictor for asthma(117). The role these bacteria play is still unclear, and additional studies show that they may be independent risk factors for acute wheezing episodes in young children aged <3 years, as well as viruses(118). The interactions between viruses and bacteria to promote wheeze and asthma onset are largely unexplored.

Finally the role of antibiotics remains widely debated. While the maternal use of antibiotics has a dose-related association with the risk of asthma in offspring(119); there is evidence for efficacy of antibiotics in the treatment of asthma and related syndromes(120-123). The idea that the microbiome represents both harmful and protective genera, that help shape the early immune system; and that bacteria have a dual role in initiating and protecting against disease, may partly explain this controversy.

2.2.5 Diet & hormones

The prevalence of obesity has more than doubled since 1980, such that it is now the commonest nutritional disorder worldwide(124). The concomitant rise in the prevalence of asthma triggered speculation that obesity may be causally related to asthma incidence. Obesity is a major risk factor for asthma development, with the strength of association greater in females than in males(125). However, understanding of the pathophysiological mechanisms underlying this relationship remains incomplete.

Cluster analyses have identified two phenotypes of obesity-related asthma (ORA) distinguished primarily by age of onset(126). The early onset phenotype is characterised by classic, T_H2-driven inflammation, and is complicated by the subsequent development of obesity (126). Conversely, the late onset phenotype is typically non-atopic, exhibits lower markers of T_H2 inflammation, occurs more frequently in women and develops consequent to obesity(127). Little is understood about how obesity may affect asthma onset. The increased AHR observed in late onset ORA is likely multifactorial with contributions from mechanical, inflammatory, metabolic and dietary factors in addition to co-morbidities such as gastro-oesophageal reflux disease and sleep-disordered breathing. Studies to enhance comprehension of the mechanisms underpinning ORA onset should be prioritised to facilitate the development of phenotype-specific therapies.

Evidence suggests that obesity and asthma share common developmental origins(128). Common genetic, epigenetic, maternal and pre-natal factors may predispose to both conditions by influencing in utero foetal programming.





GA-602077

GA- 602077

Version: Final

Subsequent early-life events and exposures in the post-natal period may subsequently modify this risk. Further research efforts are warranted to help in the prevention of the development of both disorders.

We are cognisant of the fact that we have not adequately addressed the roles of a number of other dietary factors including vitamins, oxidants/anti-oxidants, fatty acids etc, in asthma onset. For example epidemiological evidence highlights strong associations with vitamin D status(129, 130), assessed as circulating levels of 25-hydroxyvitamin D, with an increased incidence, severity and poor control of asthma(131), and these data appear strongest in children(129, 130). A recent Cochrane systematic review concluded that vitamin D is likely to reduce the risk of severe asthma exacerbation and healthcare use(131). Two large trials of vitamin D supplementation in pregnancy show strong trends for improvement of respiratory outcomes in the infant(132, 133), through likely effects on lung maturation, appropriate immune system development and improved handling of respiratory infections. Vitamin D and oxidants are additionally addressed in part in the following section, but further research into these factors and their roles in asthma prevention are also needed, including further clinical studies of vitamin D throughout the life course.

Further studies on the roles of fatty acids are also clearly needed, as a recent study on the effects of n-3 long-chain polyunsaturated fatty acids (fish oil) vs placebo (olive oil) in the third trimester of pregnancy reduced the absolute risk of persistent wheeze or asthma at three to five years of age in the offspring by 31%, with a greater reduction of 54% seen in offspring of mothers whose blood levels of eicosapentaenoic and docosahexaenoic acid were in the lowest third of the trial population at randomization(134).

2.2.6 Immune reactivity vs immune tolerance

Immune tolerance is an active process involving specialised regulatory T-lymphocyte (Treg) subsets(135, 136). The airways are continuously exposed to harmful and non-harmful environmental antigens and in health these elicit immune responsiveness to achieve the effective handling of pathogens, but importantly also immune regulation to prevent potentially damaging immune responses to non-harmful antigens(135-137).

Allergic asthma is provoked by inhalation of otherwise innocuous aeroallergens such as house dust mite, mouse, cockroach, cat and dog proteins(138) accompanied by a lack of immune tolerance, leading to inappropriate immune responses to these harmless allergens. A major CD4⁺ Treg population expresses the transcription factor Foxp3; young boys with a rare, X-linked, genetic mutation causing loss of Foxp3⁺ Tregs, suffer from multisystem autoimmunity, and severe atopy including eczema, food allergy, and eosinophilic inflammation, highlighting the importance of Foxp3⁺ Tregs in preventing aberrant immune responses to allergens in early life(139). A second major Treg population synthesizes the anti-inflammatory cytokine IL-10. Blood cells from individuals with severe allergy/asthma show reduced IL-10 synthesis in response to allergen stimulation(136, 140).





Version: Final

GA-602077

Inappropriate immune responses/breaches of immune tolerance can result from failure in generation of Tregs, or inhibition of their actions, and both can be influenced by the environment. For example, vitamin D promotes both IL- 10^+ and Foxp3 $^+$ Tregs(141), whilst air pollution is associated with impaired Foxp3 $^+$ Treg function(142). Many other environmental factors, such as smoking and oxidative stress, can stimulate production of mediators that inhibit the action of Tregs(143). The manner in which antigen is presented by dendritic cells (DCs) also profoundly affects the immune response, including whether effector or regulatory lymphocyte responses are generated, and DCs are similarly affected by environmental factors(144, 145). The inappropriate immune response to innocuous allergens in asthma causes further immune dysregulation as the T_H2 -biased allergic response inhibits antiviral responses(146).

Understanding critical mechanisms of development of naturally occurring immune tolerance offers the potential for early life interventions to dramatically reduce the incidence of allergies(147). The same potential therefore exists for asthma. Funding is needed to study the effects of the environment on immune regulation, and to reveal new approaches to reduce future development of asthma.

2.2.7 Psycho-social and behavioural factors

Although frequently unrecognized, psychological dysfunction (particularly anxiety) is up to 6 times as common in people with asthma as in matched controls(28, 148) including in children, with evidence of a bi-directional relationship, in that although asthma may induce anxiety, anxiety may also precede asthma. Longitudinal cohort studies in adults have reported that the presence of anxiety and panic significantly increased the odds of subsequent asthma(40), and a birth cohort study reported that behavioural problems in the child at the age of 3 yrs. was a significant risk factor for both a subsequent asthma diagnosis and for late-onset wheezing(148). The mechanisms underlying this relationship are not understood, although it is known that stress can have significant but complex effects on immunological function(149). Psychological stress can affect the release of cortisol and the expression of inflammatory mediators in a complex and time-dependent way, with increased airway inflammation associated with stress(150). There is some evidence that psychological stress may predispose to the development and severity of atopic conditions, including asthma, through effects on the immune system(151). Although it is clear that stress can result in measurable neuroimmunological effects that can be associated with asthma expression and morbidity, the precise significance of these biological effects and the ways in which they interact with other stress-related factors such as health beliefs and behaviours is not fully understood(151).

2.3 Addressing the unmet need

The study of early life asthma onset presents some unique challenges that separate it from stable asthma progression and asthma exacerbation. A great unmet need in understanding mechanisms of asthma onset is the lack of





GA- 602077

Version: Final

available complex mouse models that accurately reflect human asthma development in early life. Allied to this are the difficulties in obtaining valuable early-life human samples from human birth cohort studies to validate findings in the models.

The discovery of age appropriate, efficacious interventions in preventing asthma onset is completely dependent on the development of age appropriate models that will allow discovery of critical mechanisms. Not only are some susceptibility genes different for childhood and adult onset asthma(152, 153), but the effects of immune maturation and lung development must also be considered(98). Effective prevention strategies that are clinically translatable will only become apparent if data is obtained from correct animal models and validated in early life/childhood studies of asthma onset. This is highlighted by the stark differences in allergic airways disease that result depending on age at first allergen exposure in mice(98). The significantly different responses that result from allergen exposure at different ages can also be explained by influences of the developing airway microbiome and the maturing immune system. Recently marked differences between human adult and neonatal immune systems have been highlighted by the demonstration of a unique group of CXCL8/IL-8 producing T cells in preterm babies that were particularly at risk of infections(154). Immune mechanisms that predict neonatal infection have also been investigated using a systems biology approach and investigating immune-metabolic networks in babies(155). Blood volumes as small as 0.5-1ml were sufficient to identify a 52 gene cluster; predictive of bacterial infection. However, an accurate prediction was only achieved when immune and metabolic pathways were integrated, emphasising the need to use complex models, and combine information accordingly.

Any distinction between the mechanisms of early- and adult-onset asthma may reflect not so much differences in the host response but in the timing of first exposure to an aetiological allergen. For example, certain types of occupational asthma – arising through specific IgE-associated responses to protein allergens encountered only in the workplace – appear clinically and mechanistically to be indistinguishable from atopic asthma acquired early in life. Moreover, there is good evidence that the incidence of occupational asthma can be related directly to the levels of allergen exposure in the workplace; and that the disease can be effectively controlled by reducing such exposures (156).

There is a similar dearth of complex mouse models that accurately reflect human asthma development in later life. Also allied to this are the difficulties in obtaining valuable human samples from human cohort studies in adulthood, or studies of incident late onset asthma, to validate findings in the models.

Another need is that of lower airway samples from accurately phenotyped patients (representing all the different asthma phenotypes/subtypes) and controls, from which meaningful information can be obtained from relatively small sample sizes(35, 157, 158). We also need to determine the degree to which information from more easily accessible peripheral blood or nasal airway samples, reflects lower airway changes. Mechanistic studies that allow manipulation of contributory factors and truly reflect human disease are





Version: Final

needed, but currently are not being undertaken. In children, this is likely because of the difficulties inherent in obtaining appropriate tissue samples that allow functional analyses to be performed. One example of circumventing these limitations is the identification of asthma susceptibility genes through large scale, multi-centre studies that have incorporated patients from different cohorts, with data analysis being performed by a single centre with the required expertise(159). This approach of collaboration and getting access to the targeted patients and specific sampling from multiple centres and ensuring the correct expertise in processing and analysis also needs to be adopted to allow mechanistic advances from more invasive or difficult samples. Although this approach to some extent does not differentiate the different phenotypes or endotypes and may lead to some genes being missed. Combined GWAS plus expression quantitative trait loci (eQTL) analysis for different phenotypes or endotypes may provide solutions.

The use of novel techniques that allow detailed information to be obtained from small and limited samples while challenging, is not impossible. These single cell transcriptome profiling(160) and multi-component immunohistochemistry. Models using human cells also need to be innovative and optimally reflect in vivo conditions. Utilisation of co-cultures of epithelial cells or their supernatants with leukocytes(57, 161) will improve functional information obtained. The limitations of animal models are recognised(162) but if efforts are made to optimally reflect human conditions by allowing for critical factors such as age, route of allergen exposure, appropriate species and strain findings can be translated (163).

With respiratory viral infections likely being important in asthma onset, an important unmet need in asthma onset is how best to target these viruses. Palivizumab is an anti-RSV antibody effective in reducing RSV induced bronchiolitis and it may reduce subsequent wheezing (164-166). Recently, two new oral RSV antivirals performed well in double-blind, placebo-controlled studies of experimental RSV infection in healthy subjects, showing impressive reductions in symptom score and virus load(167, 168). Development of new small molecule inhibitors or other anti-virals that can be used in early life or childhood is an area that must receive further support.

The role of non-pharmacological approaches to asthma prevention, including environmental manipulations and psycho-social approaches, remains uncertain.

3 Progression of stable asthma

3.1 Introduction

Mechanisms that promote disease activity and progression in established, but stable asthma overlap with, but may also be distinct from, those involved in asthma inception, and may thus require different scientific insights and treatments. This next section deals with stable asthma and asthma progression including the dynamics of the microbiome, and whether modifications may serve as future useful therapeutic targets, the process of





GA-602077

GA- 602077

Version: Final

airway remodelling and its impact on lung function over time, the role of airway nerves, of adverse effects of asthma treatments and of disease modifying therapies such as anti- T_H2 cytokine biologics, the identification and mechanisms of other non- T_H2 asthma endo-phenotypes, and immunoregulatory processes such as resolution of airway inflammation and the development of immunological tolerance. See **Figure 2** for a summary of important mechanisms involved in asthma progression and severity.

3.2 Progression of stable asthma - State of the art

3.2.1 Bacteria, fungi & the micro/mycobiome

Detection of pathogenic *Proteobacteria*, particularly *Haemophilus* spp., are more frequent in bronchi of asthmatics than controls(91). Only 3/11 adult asthmatic subjects had severe asthma, though all were on inhaled corticosteroids (ICS). Similar data have been reported in other studies and the bacterial burden correlated with AHR(110), longer asthma disease duration, lower FEV₁ and higher sputum neutrophils(112). In another study in sputum in severe and non-severe asthma *Bacteroidetes* and *Fusobacteria* were reduced in non-severe and severe asthmatic groups compared to healthy controls, *Proteobacteria* were more common in non-severe asthmatics compared to controls (OR=2.26) and *Firmicutes* were increased in severe asthmatics compared to controls (OR=2.15). Among *Firmicutes*, Streptococcal OTUs were associated with recent onset asthma, rhinosinusitis and sputum eosinophilia(169).

Potentially pathogenic bacteria have also been identified by culture in sputum of 15% of subjects with stable asthma and this was associated with greater airway inflammation(170), while in stable severe asthma, sputum was positive for bacteria in 52% of patients(171). IgE positivity to Staphylococcus aureus enterotoxin was significantly greater in patients with severe asthma (59.6%) than in healthy controls (13%) and enterotoxin IgE-positive subjects had increased risk of severe asthma (OR, 11.09; 95% CI, 4.1-29.6), greater oral steroid use and hospitalizations and lower FEV₁(172). Further case-control studies identified patients with severe asthma with non-tuberculous mycobacteria (NTM) infections, and infected subjects were older, had lower FEV₁, and had used ICS for longer than controls(173, 174). Additionally roles for the atypical bacteria Mycoplasma and Chlamydophila pneumoniae in asthma pathogenesis have long been postulated, as reviewed in(175, 176), however data to date are inconclusive and further work is needed to better understand the possible contribution of these organisms to asthma pathogenesis.

There is clear evidence of increased susceptibility to bacterial infections in asthma. Increased risk (~3-fold) of invasive pneumococcal disease in non-severe asthma has been confirmed in two separate studies(177, 178), while that risk increases to 12-fold in severe asthma(177). Studies of mechanisms of increased susceptibility to bacterial infection in asthma report that toll-like receptor (TLR)5 and TLR7 expression is decreased in the lung in severe





an Ethani

GA-602077

Version: Final

asthma, thus severe asthmatics may suffer from insufficient TLR signalling during bacterial infections leading to impaired defense mechanisms(179-181). Effective antibacterial immunity requires type II IFN- γ and Th1 immune responses and mouse model data report that host defense against bacteria is also mediated by type I IFNs(182, 183). There is extensive data that type I, II & III IFN induction by viral and bacterial stimuli is deficient in asthma and that deficiency relates to underlying asthma severity(180, 184-188). IL-12 and IL-18 induction by LPS in macrophages are also deficient in asthma(186, 189, 190) and these are important for induction of IFN- γ and Th1 immune responses. Reduced phagocytosis of bacteria is reported in patients with severe asthma, thus persistence of bacteria in the lower airways in asthma may result from this defect(191). The mechanisms for increased susceptibility to bacterial infections in asthma are only partially understood, and further research in this field is clearly needed. The role ICS play in this increased susceptibility also needs further investigation.

The microbiota in established asthma is different to that in health(91), and recent preclinical evidence argues that the airway microbiota found in diseased lungs can influence chronicity and progression of airway inflammation(192), while treatment with certain microbes or their products, can reduce asthma severity(108). Microbial manipulation has been spectacularly successful in treating bowel inflammation(193, 194), and if asthma is driven by airway dysbiosis then manipulation of the airway microbiota may also ameliorate disease progression.

A pragmatic and potentially highly effective means of controlling asthma progression could be via the diet. A diet high in fermentable fiber fed to adult mice was effective at reducing the severity of allergic airway inflammation via fermentation of dietary fiber by gut microbiota, which directed the immune system away from allergic responses(195). The logical next steps are intervention studies in humans, where individuals would eat a diet high in fermentable fibers, or to receive supplements of certain downstream metabolites generated following microbial fermentation of fiber (e.g. short chain fatty acids). Alternative approaches whereby individuals are exposed to bacteria (probiotics) and/or their substrates (prebiotics) with the goal of directing an individual's immune system away from asthma, hold promise.

Fungal sensitisation and long term fungal infection/colonisation are associated with increased asthma severity and complications such as bronchiectasis and chronic pulmonary aspergillosis. Estimates suggest that many million people have severe asthma with fungal sensitizations and allergic bronchopulmonary aspergillosis and a substantial proportion of adult asthmatics attending secondary care have fungal sensitization. Little is known about which fungi and fungal allergens are relevant to asthma pathogenesis, and there is little data on the most effective management strategies. Further studies are needed on fungal exposure, sensitisation and infection/colonization and the role of host defense against fungi in asthma, as well as studies on the role of the mycobiome and of effective interventions(196).





Version: Final

© Copyright 2016, EARIP Consortium GA- 602077

3.2.2 Virus infections

Virus infections precipitate the great majority of asthma exacerbations and exacerbation frequency in asthma is strongly correlated with rates of decline in post-bronchodilator lung function (FEV₁)(197-199) and loss of bronchodilator reversibility(198). It therefore seems likely that virus induced exacerbations likely lead to structural alterations in the lung, and permanently worsened airflow and poor health status, and although the mechanisms are unclear, the effects are likely long lasting and may be irreversible(200). It is not known whether impaired host defense against virus (and bacterial) infections is associated with progression from mild to severe asthma.

3.2.3 Air pollution

There is substantial evidence linking traffic-related air pollution, exposure to second hand smoke and occupational exposures to progression of both allergic and non-allergic asthma to severe disease and perhaps to COPD. Diesel exhaust heightened lower airway eosinophilic inflammation in allergic subjects(201) and timing and duration of traffic-related air pollution exposure were found to be effect modifiers as early-life traffic related exposure correlated with persistent wheezing (OR 2.31) in children(202). The first 8 years of life represent a susceptible period, as exposure to traffic-derived particulate matter (PM) in that period has been associated with impaired lung function growth(203). Furthermore, in a study of asthmatic patients walking on a London polluted street for 2 hours, the reduction on FEV₁ and the degree of neutrophilic lung inflammation observed after the walk was associated most consistently with exposures to ultrafine particles and elemental carbon, and the reduction in FEV₁ was greater in the moderate compared to the mild asthmatics(204). Fine particulate matter was associated with increased respiratory resistance in children(205) and lifetime exposure to PM₁₀ and NO₂ was associated with retarded lung volume growth in elementary-school age children(206). However, since no specific pollutant or combination conferred more detrimental effects, it is important to use multi-pollutant models to analyse these associations (207). Many developing countries still employ open fires for cooking, which correlated with greater risks (OR 2·12) of asthma symptoms in children(208). The detrimental effects of pollution are readily demonstrated in certain subgroups. Young males are 3-times more likely to have asthma symptoms associated with exposure to truck traffic-related air pollution than those without (209), whereas being overweight (OR 4.36) or obese (OR 3.06) increases susceptibility to exercise-related asthma symptoms associated with exposure to $PM_{2.5}(210)$.

Mechanistic experimental exposure studies point to interactions between diesel exposure and allergen, in increasing airway inflammation(201) and suggested a possible mechanism contributing to epithelial wall damage following allergen exposure(211).

Epidemiological evidence supports a role for air pollutants in contributing to the spread of respiratory viral infections(212). This may be relevant to the mechanisms by which increased ambient ozone, nitrogen dioxide, PM2.5 and





Version: Final

GA-602077

sulphur dioxide levels are associated with increased admission for asthma exacerbations particularly in children(213).

3.2.4 Airway nerves & neural pathways

The role of airway innervation in asthma and the potential contribution of neuronal dysfunction to asthma pathophysiology are rarely studied. Human airways are richly innervated by parasympathetic efferent nerve fibres, as well as various sensory (afferent) nerve fibres(214), whereas contribution from the sympathetic nervous system is scant(215, 216). Afferent airway nerve activation mediates the noxious sensations patients associate with asthma e.g. chest tightness, air hunger, airway irritation, congestion and breathlessness(217) but also initiates reflexes producing cough. bronchospasm, AHR and mucus hypersecretion. Neuronal dysfunction therefore has the potential to play a major role in the pathogenesis of asthma symptoms.

Dysfunction of afferent and efferent nerves plays a major role in the pathogenesis of airway obstruction and AHR in pre-clinical asthma models(218-222) and in a major asthma symptom, cough(223). Cough in asthma is the archetypal airway reflex and is not only a common(224) and troublesome symptom(225), but is also associated with disease severity(226) and poor prognosis(227). Of the symptoms experienced by asthma patients cough is also the most readily objectively quantified and thus can provide insights into neuronal dysfunction. Recent work suggests that different airway diseases including asthma exhibit differing cough responses to a range of inhaled tussive agents, suggesting distinct neuro-phenotypes can be identified(223). Moreover, compared with healthy controls, different phenotypes of mild/moderate stable asthma exhibit heightened cough responses to the inhaled irritant capsaicin, which directly activates vagal afferent fibres. These responses were most exaggerated in females with nonatopic asthma, suggesting neuronal dysfunction may be particularly relevant in non-Th2 disease(228). Cough responses were unrelated to treatment with inhaled corticosteroids, exhaled nitric oxide, airflow obstruction or AHR, also supporting the hypothesis that neuronal hyper-responsiveness is a key feature contributing to treatment-resistant and non-allergic phenotypes.

Defensive respiratory reflexes, such as cough and bronchospasm, are regulated by vagal afferent nerves(229-232). Ion channels on the termini of airway sensory fibres located under the airway epithelium make them capable of directly responding to a diverse range of agents, many of which correspond to triggers of asthma symptoms identified by patients, including changes in temperature, humidity, pollution and irritant chemicals such as cigarette smoke, cleaning products, perfumes etc(229, 233), as well perhaps to changes in airway calibre during bronchospasm(234), Neuronal dysfunction in animal models of asthma can also be induced by eosinophils(235), viruses(236) and mediators such as neurotrophins(237, 238) and can mediate induction of AHR by β_2 agonists(239).





GA- 602077

Version: Final

Gastroesophageal reflux disease and a history of rhinitis or sinusitis are clearly identified risk factors for development of severe asthma(240), however the mechanisms involved are poorly understood, and the possible involvement of neural pathways requires further study.

The importance of nerves in asthma is highlighted by the effectiveness of muscarinic antagonists in the treatment of asthma(241, 242), blocking reflex bronchospasm mediated by acetylcholine release from parasympathetic efferent airway fibres. Animal models of asthma have demonstrated that in addition to anti-muscarinics, the late response to inhaled allergen can be prevented by blockade of afferent sensory nerves(220), implicating airway nerve function in allergic as well as irritant induced asthma symptoms. Although translation in clinical allergen challenge studies is still required. Of note AHR, airway obstruction and cough often persist despite treatment with highly effective anti-inflammatory drugs, perhaps as a consequence of persisting neuronal dysfunction(221). Therefore, there is a need for further research on the role of nerves in the pathogenesis of asthma and on new therapeutic approaches targeting neural dysfunction in this disease.

A substantial proportion of new or relapsing asthma in adulthood can be attributed to exposures encountered in the workplace (243). The paradigm of occupational asthma induced by classic allergic responses to workplace proteins such as flour or detergent enzymes is evidence that age itself is no protection against the development of IgE-associated respiratory disease. A substantial proportion of occupational asthma, however, arises in response to exposure to chemical agents that are of too low a molecular mass to act alone as antigens. In some cases these agents appear to conjugate with human proteins to form a hapten-protein allergenic complex; in many others this appears not to be the case and other mechanisms, such as neuronal dysfunction acquired through repeated exposure to respiratory irritants require exploration. The public health implications may be substantial since irritant exposures encountered at work - by, for example cleaners who consistently report high rates of asthma-like symptoms to cleaning products (244), or swimming pool attendants who regularly encounter chlorine(245) - can have very significant 'down-stream' effects in consumers.

Research on the role of nerves in asthma has been hampered by the different innervation of murine and human airways(246), by limited access to neuronal tissues (nerve fibres or ganglia) from patients with asthma, and by methodological challenges in analysis of these tissues. Therefore, more funding is needed for neuronal studies of human tissues, such as analysis of nerves in human whole-mount biopsies(216), to provide urgently needed data on the precise nature of airway innervation in asthma(162). In addition, experimental non-murine eg guinea pig asthma models are needed to provide more relevant functional information on airway nerves in asthma(247-249). Models and reagents in other species such as rats, rabbits and dogs would also aid both safety and efficacy studies. Finally, clinical studies of emerging therapies specifically targeting neuronal dysfunction in asthma(250, 251) are





Version: Final

needed to identify new therapeutic opportunities (223).

3.2.5 Airway remodeling

Airway remodeling represents one of the most challenging problems in asthma and is linked to disease progression. Airway remodeling includes epithelial cell shedding, goblet cell hypertrophy, basement membrane (BM) thickening and ASM cell hyperplasia, leading to progressive decline in lung function(252, 253). Another prominent feature of lower airway remodeling is increased vascularization that correlates with airflow limitation and AHR(252, 253). Growing evidence suggests that inherent changes in lung-resident cells are the principal drivers of fibrotic processes. Support for this concept comes from studies demonstrating that airway remodeling is observed in preschool children without signs of inflammation, and is comparable to that documented in adults(252, 253).

Other mechanisms involved in promoting airway remodeling processes include notch2(254), IL-13(255), the gamma-aminobutyric acid system(256), transcription factors regulating goblet cell differentiation(257) and many others. Further studies investigating mechanisms involved in promoting airway remodeling processes are required to lead to better therapies to prevent its development. Once developed, novel therapeutic approaches such as bronchial thermoplasty can ameliorate severity(258), however mechanisms involved are poorly understood, and better understanding of these mechanisms could lead to less invasive approaches to ameliorate severity.

Mainstay anti-inflammatory therapies, such as ICS, do not significantly affect airway remodeling processes, emphasizing the notion that uncontrolled structural changes may be a cause as well as a consequence of progressing asthmatic disease and future funding should be channeled to studies aiming to dissect the mechanisms involved(259, 260). Large-scale proteomic and genomic profiling of lung-resident cells may also guide the identification of novel mechanisms for airway remodeling. In addition, mathematical modeling approaches aimed at elucidating the effects of ASM contraction and the causes of altered airway mechanics in asthma may greatly facilitate the understanding of airway remodeling pathogenesis(259, 260).

Immune cells infiltrating the airways of asthmatics are potent sources of profibrotic factors, including transforming growth factor (TGF)- β , and therefore a role for immune cells in promoting structural changes is likely. In fact, eosinophils are a rich source of TGF- β and eosinophil depleted mice are protected from peri-bronchial collagen deposition and increased ASM mass in response to chronic allergen exposure(259). Recent studies have also uncovered key roles for other TGF- β superfamily members, including activin-A, and bone morphogenic proteins and the epithelial cell-derived cytokines, IL-33, TSLP and IL-25, in airway remodeling in asthma(261-263). Hence, the development of biologics targeting the effects of these cytokines on the control of airway remodeling represents a fruitful avenue for future research that may be translated to better treatment options for asthmatics(262, 263).

A major hindrance in our understanding of the mechanisms underlying airway remodeling is the lack of animal models that can effectively recapitulate





GA-602077

GA- 602077

Version: Final

structural changes(252, 253). It is, thus, essential that future funding should be targeted on the development of better animal models and novel *in vitro* assays that incorporate multiple cell types and mimic the intrinsic mechanical forces occurring in the airways. Finally, studies investigating the effects of environmental changes, such as diet and microbiota composition, on airway remodeling are warranted and future funding should be channeled in this area.

3.2.6 Immune reactivity vs immune tolerance

Because development of tolerance to allergens depends on the context (including dose, timing and route of exposure as well as presence of coexposures such as LPS) of exposure to the environmental antigens, this is
amenable to therapeutic manipulation. The best example of this is allergen
immunotherapy, which involves administering allergen to patients to induce
long-term, allergen-specific, immune tolerance. This can be achieved by
injection immunotherapy, which has been shown to induce benefits lasting
long after treatment has stopped(264), but which requires repeated
attendance at allergy clinics for 3 years. This can also be achieved by
sublingual immunotherapy(25), which is newer and therefore has unknown
duration of long-term benefit, but which has the advantage of being
administered by the patient at home.

Allergen immunotherapy increases Treg frequency and particularly of those Tregs that synthesize IL-10(265). Efforts are ongoing to standardize immunotherapy protocols, improve safety and efficacy, reduce costs and duration of treatment, and to better understand immune mechanisms involved to further improve its utility in treating asthma(265). Further current challenges in immunotherapy include understanding how to maintain immune tolerance and memory regulatory cell populations over extended periods of time.

If factors that underlie loss of such immune tolerance in asthma can be identified by research, then future therapies and environmental policies can be formulated to tackle the underlying immune dysregulation in asthma. Research is needed to study mechanisms underlying loss of immune tolerance and dysregulation in different asthma endotypes with the aim of designing better targeted treatments that restore normal immune responses, reducing asthmatic inflammation without the side-effects of suppressing healthy immune responses as seen with high dose corticosteroids. As the environment is a potentially modifiable factor in immune dysregulation, funding is also needed now to urgently study the effects of the environment on immune regulation, to reveal new approaches to treat asthma.

3.2.7 Resolution of airway inflammation

Chronic inflammation underlies the pathogenesis of asthma, the intensity of symptoms and progression of disease. Yet, efforts to tackle chronic inflammation in asthma have mostly explored the activation of anti-inflammatory or immunoregulatory responses on the assumption that the dampening of pro-inflammatory responses would suffice for inflammation to fade away.





© Copyright 2016, EARIP Consortium

-- 1

GA-602077

RIP Consortium Version: Final

Recently, it has become apparent that resolution of inflammation is by itself an active and highly orchestrated process, of similar complexity to the onset and progression of inflammation, responsible for the catabolism of the inflammatory response, the egress of immune cells and the restoration of tissue homeostasis(266). Among the multiple pathways involved, the production of specialized pro-resolving lipid mediators (SPMs) such as resolvins, protectins and maresins seems to be particularly important(267). These are generated through complex biosynthetic pathways from ω -3 and ω -6 polyunsaturated fatty acids and act in a stereospecific manner through G protein-coupled receptors to reverse vasodilation and suppress leukocyte infiltration, de-activate inflammatory cells, promote apoptotic cell and tissue debris clearance and repair damaged tissue(267).

In asthma, emerging evidence suggests that SPMs play a prominent role in the resolution of airway inflammation(268). SPM networks are altered in exhaled breath condensate, bronchoalveolar lavage, sputum or blood of patients with asthma(269-273), while administration of synthetic SPMs such as lipoxin A4, protectin D1, resolvin D1 (RvD1) or resolvin E1 (RvE1) promotes resolution of airway inflammation in experimental mouse models(271, 272, 274-278). SPMs pro-resolving actions in this context may additionally involve activation of macrophage efferocytic function and suppression of $T_{\rm H2}$ cytokines, IgE and ILC2 function, although their spectrum of activities remains largely uncharacterized(268). This raises the possibility of a key role of SPMs in the termination of airway inflammation with significant implications for the therapeutic potential of these molecules in halting asthma progression. This largely unexplored area is therefore in need of urgent funding.

3.2.8 Adverse effects of asthma therapies

 β_2 -agonists are almost universally used therapies in asthma as single agents, and as combination therapy with ICS. Inhaled short (SABA) and long acting (LABA) β_2 -agonists mediate their protective effects by inducing cyclic adenosine monophosphate (cAMP) in smooth muscle cells (SMCs) leading to smooth muscle relaxation and thereby, bronchodilatation. Although β_2 -agonists have undoubted beneficial effects, safety concerns have been repeatedly raised regarding the use of SABAs and LABAs in asthma.

Regular use of SABAs four times daily in stable asthma results in worse asthma control than use only when needed to relieve wheezing(279, 280) and overuse of SABAs (without ICS) in asthma exacerbations has been repeatedly associated with increased risk of hospitalisation or mortality(281, 282). LABA use without ICS has also been linked to increased asthma mortality(283-285) and the FDA has cautioned medical providers about risks associated with LABA use without ICS(286). The fact that LABA use, when combined in the same inhaler with an ICS, is safe was confirmed in two recent large studies in both adults(287) and children(288). Concerns about use of β_2 -agonists in the absence of ICS persist as excessive use of SABAs in the absence of ICS was recently identified in 40% of deaths and use of LABAs





GA- 602077

Version: Final

without ICS in 5(282). The mechanisms behind these safety concerns and the mechanisms through which ICS therapy is protective are poorly understood. Fenoterol is a SABA linked with an epidemic of asthma mortality in the 1980s(289) and reductions in hospitalisations due to asthma exacerbations following withdrawal of fenoterol suggested this was due to a beneficial effect of withdrawal on asthma severity(290).

 β_2 -agonists relax smooth muscle by raising levels of cAMP in SMCs. However, the most numerous cells in the airway and the cells most accessible to inhaled β_2 -agonists are not SMCs, but bronchial epithelial cells (BECs) and airway macrophages. It therefore seems likely that respiratory adverse effects of β_2 -agonists might be mediated via effects on these cells.

The pro-inflammatory cytokine IL-6, is induced in BECs by β_2 -agonists alone, and importantly in relation to overuse of β_2 -agonists in asthma exacerbations, IL-6 induction by RV infection was further augmented by β_2 -agonists. Promoter studies revealed that LABA augmentation of RV-induced IL-6 occurred via a cAMP response element (CRE) in the IL-6 promoter(291), indicating that an adverse effect of β_2 -agonists is mediated via cAMP elevation in BECs, just as their beneficial effects in SMCs are.

Two independent clinical trials have confirmed induction by β_2 -agonists of the asthma-related mediators brain-derived neurotrophic factor (BDNF)(292) and matrix metalloprotease (MMP)-9(293) in humans and both mediators are induced via cAMP/CREs(294, 295). Many other pro-inflammatory mediators with potential adverse effects in asthma have CREs in their promoters and are therefore likely to be induced by β_2 -agonists including IL-17, COX-2, amphiregulin, MMP-2, MUC5AC, MUC5B and MUC8(296). However, these have not to date been studied with β_2 -agonists. A genome-wide study suggests the number of human genes potentially inducible by β_2 -agonists via CREs in their promoters might extend into the hundreds(297). Thus- β_2 -agonists have the potential to induce many genes implicated in asthma pathogenesis.

Induction of and augmentation of RV-induced IL-6 by LABA were both abolished by ICS(291). ICS also blocked LABA induction of BDNF both in vitro and in vivo(292). IL-17, MMP-2, MUC5AC and MUC8 are all suppressed by steroids(296). These data suggest that use of β₂-agonists/ICS in combination inhalers would result in the ICS component blocking direct genomic adverse effects of β₂-agonists while maintaining the beneficial bronchodilator effects(296). This interpretation supported demonstration that taking both together, combined in a single inhaler, is clearly safe(287, 288). Mechanistic studies investigating adverse effects of β₂agonists and protective effects of ICS in BECs and airway macrophages in vitro, and in vivo in people with asthma, to identify genes induced by β₂agonists and suppressed by ICS, are urgently needed.

Use of leukotriene receptor antagonist (LTRA) therapy has been associated with an increased incidence of Churg-Strauss syndrome (CSS)(298). How LTRA therapy may interact with the pathogenesis of CSS is unknown. Potential mechanisms for an association between LTRAs and the CSS have been postulated including potential for allergic/hypersensitivity drug reactions





Version: Final

and leukotriene imbalance resulting from leukotriene receptor blockade. Further studies monitoring incidence of CSS in asthma patients receiving LTRAs are needed, including studies on the possible role of steroid/ICS withdrawal. Mechanistic studies investigating how LTRA therapy may interact with the pathogenesis of CSS are also needed.

Inhaled corticosteroids are widely used in chronic obstructive pulmonary disease (COPD) and their use is associated with an increased risk of pneumonia, but the mechanisms of this effect remain unclear(299). There is clear evidence of increased susceptibility to bacterial infections in asthma(177, 178), but this increased risk has not been directly linked to ICS use. The mechanisms for increased susceptibility to bacterial infections in asthma and the role ICS plays in this increased susceptibility are poorly understood. ICS also suppress antiviral immunity in the absence of asthma(300), and are associated with impaired innate IFN responses in asthma(301), but ICS also reduce exacerbation frequency(302), the majority of which are induced by viruses. Therefore, ICS presumably have mixed effects in asthma, and the mechanisms involved clearly need better understanding in order to guide future drug development.

Human genetics research will be critical to the development of genetic profiles for personalised medicine in asthma. Genetic profiles that predict individual disease susceptibility and risk for progression, may predict which pharmacologic therapies will result in a maximal therapeutic benefit, and may also predict whether a therapy will result in an adverse response in a given individual. Pharmacogenetic studies of the glucocorticoid, leukotriene, and β_2 -adrenergic receptor pathways have identified genetic loci associated with therapeutic responsiveness(303). Future studies, are needed to identify genetic profiles permitting personalised approaches to maximize therapeutic benefit for an individual, while minimizing the risk for adverse events.

3.2.9 Diet & hormones

Obesity has been associated with both over and under diagnosis of asthma(304), and with worse asthma control, impaired response to ICS therapy, increased exacerbation frequency, increased healthcare utilisation and diminished asthma-specific quality of life relative to normal-weight asthma(305). Patients with obesity-related asthma (ORA) are more likely to have obesity-related comorbidities such as obstructive sleep apnea and gastroesophageal disease that can influence responsiveness to asthma therapies(306). Obesity, although reportedly not associated with airflow obstruction(307), adversely affects pulmonary mechanics by reducing lung tidal volume, functional residual capacity and AHR(308), however, the mechanisms involved require clarification. Additional studies addressing the effect of obesity on airway remodelling may also help identify novel therapeutic targets.

Obesity is associated with chronic, low-grade, systemic inflammation and adipose tissue produces numerous pleiotropic adipokines in addition to serving as an energy-storage depot(309). The two most widely studied adipokines are leptin (pro-inflammatory) and adiponectin (anti-inflammatory)





GA-602077

GA- 602077

Version: Final

and both have been implicated in the pathogenesis of ORA(309). However, our understanding of the roles of adipokines within obese airways warrants further research. Adipokines are promising therapeutic targets as they are differentially expressed in obesity and several have been shown to exhibit immunomodulatory effects(309).

Although obesity is a state generated by positive calorie imbalance, dietary constituents, such as ω -3 and ω -6 polyunsaturated fatty acids and saturated fats, may also affect ORA pathogenesis(310). Furthermore, insulin resistance and oxidative stress may also play an important role(311). In addition to enhancing AHR obesity is thought to potentiate airway inflammation and reactivity to environmental stimuli such as ozone and particulate matter(127), with associated negative consequences on asthma control. Results from recent mouse studies have emerged to suggest potential mechanisms(312); however, further work is required.

The most rational strategy to improve the health status of individuals with ORA is weight reduction and several studies have demonstrated improvements in pulmonary function, asthma control, health status, AHR and systemic and airway inflammation following weight loss interventions(313, 314). However, whilst weight optimisation confers many health benefits and is recommended in the treatment of ORA, randomised controlled trial (RCT) data evaluating the efficacy of weight loss interventions (both surgical and non-surgical) in ORA are limited(315).

Evidence addressing the optimal pharmacological strategy for the treatment of ORA is lacking. Practical ORA management is complicated by the vicious cycle generated by obesity-induced poor asthma control and corticosteroid-associated weight gain. Greater understanding of the mechanisms underpinning obesity-associated corticosteroid resistance (in addition to promotion of weight-loss strategies) is key to breaking this cycle and improving clinical outcomes. Thus further work in this area should be prioritised.

Asthma is more common in males from birth until puberty(316, 317) but becomes more prevalent(318, 319) and more severe(320, 321) in women after puberty. Women are more likely to develop difficult-to-treat or steroid refractory asthma(317) and women >25 years of age account for >62% of hospitalisations and 64% of asthma deaths(317, 322). In addition to puberty, menstruation(323), pregnancy(324, 325), menopause(326, 327) and oral contraceptive use(328) have been associated with asthma outcomes in women(317). Peri-menstrual cyclic changes in lung function have been reported in women(329) and asthma symptoms and peak expiratory flow can deteriorate during high levels of oestrogens (330). These data suggest a role for sex hormones, most likely a pathogenic role for oestrogen and most immune cells involved in asthma express the oestrogen receptors (ER) ERa, ERB. The hypothesis that sex hormones play a role in the pathogenesis of asthma is further supported by the higher prevalence of asthma in women with early menarche(331) which is associated with higher oestrogen concentrations (332). Oestrogen levels have also been associated with the





GA- 602077

Version: Final

incidence of asthma(333) and women with a history of asthma who use oral contraceptives have reduced risk of current wheeze(334). But there are also developmental differences: female foetal lung development is more rapid than that of males(335) and male lungs are smaller with fewer respiratory bronchioles at birth(336) while at puberty boys have ~25% higher lung volumes than girls of identical height(337) and lung function development ends earlier in girls than in boys(338). Understanding gender differences in asthma which might in part be regulated by sex hormone levels appears important to optimize individualized treatment. Furthermore, a better understanding of gender differences in asthma pathogenesis and progression might lead to new treatment modalities.

Among the chronic diseases of adolescence, asthma has the highest prevalence and health care usage(339). During puberty children experience a shift in cognitive abilities from more concrete to more abstract thinking (340. 341) and children with asthma feel more lonely and depressed compared to healthy peers(342). Absenteeism or poor school performance can disrupt peer relationships and endanger development of independence(341). While self-care generally increases in children it may decrease again in adolescence(341, 343). Girls are more likely to incorporate asthma into their social and personal identities compared to boys who try to avoid this(344). Age and low emotional quality of life correlate with body mass index and level of asthma symptoms (341). Thus, in addition to hormonal changes discussed above which influence asthma severity, developmental changes in cognition and behaviour during puberty can severely affect asthma outcomes. Most management interventions do not account for the challenges faced by psychosocial and physiologic needs during puberty(341) and age specific programs are needed(341). Such programs could have vast influences on the future course of individual patients' asthma and could be instrumental in improving long term outcomes.

Around 10% of women of childbearing age suffer from asthma(325). Asthma prolongs time to pregnancy and has negative effects on fertility that increase with age and asthma severity(345). Pregnancy is associated with a large increase in circulating oestrogen levels that drop to baseline levels after delivery. The course of asthma during pregnancy is variable with approximately 1/3 reporting unchanged asthma control, 1/3 an improvement and 1/3 worsening asthma(325). Analysis of pulmonary function versus symptoms suggests that some reported deteriorations in symptoms might not be reflected in changes in airway function(325, 346). A small increase in the risk of congenital malformations in the offspring of patients with asthma has been reported with a higher incidence in more severe asthma(325). Preterm labour, low birth weight, small for gestational age and preeclampsia have associated with pregnancies in women with asthma(325). Understanding mechanisms that lead to deteriorations of asthma control during pregnancy and programs to monitor asthma control during pregnancy have a potential to avoid adverse outcomes during pregnancy.





Version: Final

© Copyright 2016, EARIP Consortium GA- 602077

3.2.10 Psycho-social and behavioral factors

Despite advances in pharmacotherapy that are theoretically capable of achieving high levels of asthma control for most patients, asthma outcomes have remained suboptimal and many patients remain symptomatic despite intensive pharmacological therapy. Psychological conditions such as anxiety and depression are common in people with asthma and associated with poor control(28). Consistent evidence from cross-sectional surveys using a variety of methodologies suggests that symptomatic asthma control and asthmarelated health status are impaired when anxiety or depression are also present(29, 347-349). This relationship is independent of confounding factors such as age, gender, socio-economic status, objective asthma severity and prescribed treatment level, with one study(349) estimating that the presence of psychiatric co-morbidity accounted for 29% of the variance in Asthma Control Questionnaire score. Anxiety and depression are associated with poor asthma outcomes across a range of different outcome measures. These include impaired asthma-related quality of life(350, 351), higher asthmarelated health resource utilisation(352), increased asthma-related health costs(352) and increased use of rescue medication(353). Patient reported outcomes correlate poorly with objective physiological measures of asthma control, while measures of psychological state are strongly predictive of most outcomes. The mechanisms underlying these associations are not well understood, and there is a paucity of interventional studies to show whether the recognition and treatment of co-morbid psychological dysfunction is effective in improving asthma control, with variable evidence supporting other behavioural interventions.

Breathing control exercises now have ever-increasing evidence to support their use as an adjuvant treatment for those uncontrolled on standard pharmacotherapy and are advocated in guidelines (354, 355), although the mechanism of action is incompletely understood. Behavioural interventions to support adherence(356) and internet-based behaviour change and selfmanagement support interventions for asthma show promise(357) but require further research to clarity which patients may benefit and in the optimal format of delivery. An accurate multidisciplinary assessment of adherence and psychological state are considered key parts of the multidimensional phenotyping of asthma required in a difficult asthma clinic before embarking on expensive treatment with new biological therapies. Other nonpharmacological interventions (including relaxation, mindfulness, biofeedback and cognitive-behavioural based) have some supportive evidence but an inadequate evidence-base and mechanisms of effectiveness are generally poorly understood(358). In view of how commonly the co-morbidity between psychological problems and asthma occurs, it is perhaps surprising that the evidence base for treatment is so meagre

To effectively complement pharmacological asthma management, there is a need to understand the neurocognitive, affective and behavioural mechanisms that impact asthma outcomes, particularly those relevant to anxiety and depression. Breathlessness is the fundamental symptom of asthma, with brain processing pathways possessing a strong affective





Version: Final

component that can cause anxiety and enhance symptom perception(359) as impacting cognition and negatively behavioural mechanisms (360). Neurocognitive studies have reported activation in the ventrolateral periaqueductal gray (vIPAG) associated with respiratory prefrontal activity stress-related and mav reflect inflammation(362). Investigation of neural pathways in asthma shows differential responses to a cognitive task in subgroups with different levels of inflammatory response to an allergen challenge, suggesting the possibility of neurophenotypes for asthma, with the potential of targeted interventions (363). Differential neural reactivity is related to disease severity in brain areas related to emotion-processing, indicating that neurophenotypes may exist within asthma populations. Such neurocognitive evidence supports cognitivebehavioural models (364) suggesting that some people with asthma may have dysfunctional cognitions (e.g. symptom perception) that interact with physiological mechanisms leading to the increased asthma onset risk noted in longitudinal studies. Cognitive-affective models have emphasized the importance of symptom perception in reduced asthma-specific quality of life(364). Anxiety is the strongest predictor of the unpleasantness of breathlessness during bronchoconstriction in people with asthma(365), and anxiety has a stronger relationship with asthma-related health status than lung function(366).

Understanding of the mechanisms underpinning these relationships is needed to efficiently design and target appropriate interventions.

3.3 Addressing the unmet need

New therapies and emerging biologicals now target T_H2 asthma phenotypes, with success, showing that disease modifying agents have efficacy, even though asthma is clearly established. Limited information exists regarding other asthma endo-phenotypes particularly non-T_H2 asthma and treatmentresistant phenotypes. Therefore, novel approaches incorporating pre-clinical and clinical models are required in order to develop new therapeutic approaches for these neglected varieties of asthma.

Many of the agents known to precipitate asthma symptoms are also capable of activating airway nerves leading to the hypothesis that neuronal hyperresponsiveness is a key feature of some asthma endotypes, contributing to treatment-resistant phenotypes, particularly in non-T_H2 asthma. The role of airway innervation has received very little attention to date and has the potential to identify novel therapeutic targets. Major investment in consortia aimed at understanding and treating severe asthma has focused on inflammatory profiling in the various sub-phenotypes identified with little investment in the characterisation of neuronal function and associated targets. Anti-IgE and IL-5-targeted therapies are currently finding their place in the treatment of asthma. A word of caution may be required as long-term effects are unknown as yet. And further, both eosinophils (depleted by anti-IL-5 and anti-IL-5 receptor) and basophils (depleted by anti-IL-5 receptor) may exercise protective properties and thus their depletion may cause unwanted effects. A major challenge for these interventions will be to decide optimal timing of their





GA-602077

GA- 602077

Version: Final

use, particularly as sensitization and the start of asthma progression frequently occurs early in life. While much effort is put in obtaining antibodies that are highly effective, it should also be considered that dampening responses might be effective and at the same time may preserve useful functions of the targeted mediator. Thus, while anti- $T_{\rm H}2$ cytokine therapy will continue to impact on stable asthma and perhaps on asthma progression, there is still a lot of basic information missing on the best way to harness these therapies.

The successes with T_H2 -targeted therapy indicate that this approach is viable in the treatment of asthma in which these pathways are active. Intervention with the inhibition of T_H2 responses is still in its infancy and development would be greatly helped by relevant animal models. Efforts like U-BIOPRED and SARP, to obtain detailed molecular insights into various endotypes and phenotypes of severe asthma, should be pursued in parallel with the development of animal models to substantiate the functional relevance of specific molecular pathways for specific endotypes.

Airway epithelial cells and macrophages are an important source of IL-25, IL-33 and TSLP, which act on TH2 cells, ILC2s and DCs to enhance TH2 responses and we need to clarify whether epithelial cells and macrophages from individuals with asthma produce greater amounts of these T_H2-promoting cytokines compared to those without disease. We also need to develop inhibitors of IL-25, IL-33 and TSLP(367) to permit comparisons of their relative abilities to inhibit T_H2 cell, ILC2 and DC mediated enhancement of T_H2 responses. These studies will need to be carried out in pre-clinical models to permit greater understanding of their biology, as well as in RCTs with detailed phenotyping, to permit better understanding of which sub-phenotypes of asthma might respond to each therapy. Blocking these pathways holds great promise, as this should block T_H2 responses more effectively than targeting individual T_H2 cytokines, and they should also be more broadly applicable across different asthma phenotypes. Recent studies have also revealed that ILCs show plasticity. ILC1s can trans-differentiate into ILC3s, and vice versa(368) and ILC2s into ILC1s and vice versa(369). Therefore transdifferentiation is an option for restoring the balance between ILC subtypes and with that adjust effector responses.

There is a need for more intensive investment in mechanisms of resolution of inflammation. Key areas that will need to be investigated include the more global lipidomic analysis of ω -3 and ω -6 polyunsaturated fatty acid-derived lipid mediators, the better understanding of their regulation and biological functions at the 'system level', the effect of age, diet, environmental influences and infections in their generation, and their deregulation during disease. The identification of asthma subtypes that would most likely benefit from SPMs, and the development of appropriate therapeutic agents with desirable pharmaceutical properties will ultimately determine whether such rational new treatments aiming at pushing inflammation 'forward' towards its resolution can bring about significant improvements in asthma outcomes.

Studies analysing the effects of the disease itself, as well as treatments such as ICS on host immunity against viral and bacterial infections in asthma are





GA- 602077

Version: Final

urgently needed. These should include ex vivo studies in human primary cells as well as in vivo studies in models of asthma, investigating the effects of disease/ICS on host defense against virus and bacterial infections in BECs, macrophages and dendritic cells. Human experimental rhinovirus infection should be used to determine whether virus infection alters the microbiome and impairs host immunity to bacteria in patients with asthma. This model would then offer a model to study the effects of disease/ICS on the interactions between virus infection, host immunity, and the microbiome in small numbers of patients over a short period. By comparison of the effects of virus infection between asthmatic/healthy patients and patients using ICS and ICS-naive patients, experimental rhinovirus infection has the potential to identify mechanisms of increased susceptibility to viral/bacterial infections related to disease itself and to ICS use. Studies are also needed to investigate relationships between viral and bacterial infections and progression from mild to severe asthma, as well as studies on host immunity against these infections and asthma progression.

In relation to air pollution, the observation that exposure to traffic-derived PM during the first 8 years of life can impair lung function growth(203) must make us focus on the mechanisms by which traffic related air pollution, as well as perhaps work related exposures and second hand smoking may increase the severity of asthma. Epidemiological studies need to be pursued to find out whether there is any threshold safe levels of exposure or not in this respect. In addition, we need to find out what determines susceptibility to adverse consequences of exposure, the cellular and molecular mechanisms involved, and to devise and test preventive measures and treatments (for those particularly susceptible), as the problem of pollution will take time to be controlled at source. We need genome wide association studies (GWAS) and epigenetic studies in exposomics, and extension of these studies into asthma in particular, to better understand how asthma underlies increased susceptibility to stressors like cigarette smoking, occupational exposures and traffic related (eg. diesel exposure) and other sources of pollution.

We need to refine the understanding of mechanisms underlying the worsening of asthma progression with particular focus on oxidative stress pathways, T_H2 and non- T_H2 pathways (including the role of sensory nerves). We need to determine whether there is a particular endotype of asthma that results from high susceptibility to pollutant effects and we need to establish treatments (and life style measures eg dietary) to combat the effects of pollutants on asthmatic diathesis in the most susceptible individuals.

Studies are additionally needed to determine whether direct induction by β_2 -agonists via cAMP, as well as potentiation by β_2 -agonists of virus-induction, of pro-inflammatory mediators in airway epithelial cells and macrophages, may explain the adverse effects of β_2 -agonists in stable asthma and in asthma exacerbations. These studies should be carried out *in vitro* using human primary cells from people with asthma and should use methods such as whole genome transcriptomics or RNA-seq to identify all genes which are upregulated by β_2 -agonists. Further studies should test the hypothesis that these adverse effects will be resolved by the addition of ICS. Similar studies should





GA- 602077

Version: Final

be carried out *in vivo*, in people with asthma with both LABAs and SABAs delivered alone at the rapeutic doses and should sample airway lining fluid using the novel method of bronchosorption to measure soluble mediators, as well as airway BECs and macrophages for gene expression analyses. Again these studies should include β_2 -agonists alone, and β_2 -agonists in combination with ICS. Such studies are urgently needed to better inform safest use of these ubiquitous the rapies.

A major hindrance in our understating of the mechanisms underlying airway remodeling is the lack of animal models that can effectively recapitulate structural changes. This is due to the disparities in lung morphology between species and to the fact that most of the experimental protocols utilized are tested as preventive, rather than therapeutic, approaches(252, 253, 259). It is, thus, essential that future funding should be targeted to the development of better animal models, to mapping of different models to different asthma phenotypes and development of novel *in vitro* assays that incorporate multiple cell types and mimic the intrinsic mechanic forces occurring in the airways. In addition, more attention should be directed to the contribution of the remodeling of the small airways in asthma, the mechanisms of which remain incompletely defined.

For the microbiome, future studies are required to establish the range of commensals and Proteobacteria spp., fungi and other pathogens in the lower airway microbiome of normal and asthmatic subjects, to establish the degree of individuality, stability and anatomic variability of airway microbial communities, to relate the microbiota to severity and duration of asthma, current therapy, and measures of inflammation and prospectively study the effects of antibiotics on the asthmatic airway microbiome. Further work in preclinical models as well as in man is needed to understand the role of probiotics, prebiotics and the gut microbiome in regulating allergic airway inflammation and asthma. Finally, studies investigating the effects of environmental changes, such as diet and microbiota composition, on asthma progression are warranted and future funding should be channeled in this area. Pragmatic approaches, such as diet changes, have the potential to rapidly be tested and implemented, and consequently represent an opportunity for influencing asthma progression in the medium term. To achieve this, directed funding ensuring high quality and rigorous clinical intervention studies is needed.

Better understanding of the mechanisms underpinning the relationship between psychological and neurological factors and asthma outcomes is needed to help clinical care and to understand the complexity of asthma. The effectiveness of psychological and non-pharmacological interventions and the way such interventions fit in with pharmacological strategies need to be established, and the mechanisms of effectiveness need to be understood to allow efficient targeted delivery of appropriate interventions to the right patient. Patients consistently stress the need for a holistic approach and for better integration of non-pharmacological strategies, and a personalised medicine approach demands that a holistic strategy encompassing these factors be established.





GA- 602077

tium Version: Final

4 Asthma exacerbations

4.1 Introduction

Asthma exacerbations are universally accepted as the major cause of asthma morbidity, they contribute hugely to the healthcare costs associated with asthma and they are responsible for unacceptable asthma mortality(370). The pharmaceutical industry, academia and governing bodies all recognise the important impact of asthma exacerbations and the need for new asthma treatments that specifically address the burden of asthma exacerbations(5). Respiratory viral infections precipitate the vast majority of asthma exacerbations, however a number of other factors also contribute to exacerbation risk including allergen exposure(302, 371), air pollution(372), bacterial infection/reactivation(118, 373), eosinophils(374, 375), total IgE(376), asthma severity(377), genetic background(49. dampness(378), temperature variation(379), vitamin D deficiency(380, 381) medication adherence(382), absence of health insurance(377) and risk perception(383). Indeed, identification of other precipitants of asthma exacerbation is key to reduce asthma morbidity and health care utilization. Additional studies of risk scores for asthma exacerbation may help guide management of asthma patients(384, 385) while predictors of hospitalization and/or relapse in patients already experiencing an exacerbation are also being examined(386). See Figure 3 for a summary of important mechanisms involved in asthma exacerbations.

4.2 Asthma exacerbations - State of the art

4.2.1 Viruses

In adults and children, respiratory viruses cause approximately 80% of asthma exacerbations. The most common precipitants are RVs, but all respiratory viruses can do so. The roles of respiratory viruses, which may work both additively/synergistically with other stimuli including allergens and pollutants(302, 371, 372), and bacteria and atypical bacteria which are also important triggers of asthma exacerbation have been extensively covered in other recent reviews(387-389).

Interferons (IFNs) are anti-viral cytokines that induce >300 individual genes which aim to limit virus replication and dissemination(390). Many studies have reported deficient type I (β) and type III (λ) IFN production in BECs cultured from both children and adults with asthma, in response to RV infection(157, 180, 184, 185, 187, 391, 392). Studies have been extended to include IFN- α , other viruses/viral mimics and other cell types including airway macrophages(185, 393), peripheral blood mononuclear cells(301, 394), and blood derived DCs(395, 396). The degree to which this deficiency relates to asthma severity and/or control(301), or is found only in certain asthma phenotypes, but not others(397, 398), requires further study.





GA-602077

Version: Final

Impaired IFN responses in asthma are thought to be caused at least in part, by increased expression of the negative regulator suppressor of cytokine signaling (SOCS)-1(399). Various cytokines including IL-4, IL-13(146) and TGF- β (400, 401) can suppress virus-induced IFNs *in vitro*, however whether this is solely dependent upon SOCS1 remains to be established. Additionally TLR7 (a viral RNA sensor and potent IFN-inducer) expression(179, 180) and function(402) are reported to be deficient in asthma. Further studies on mechanisms of IFN deficiency in asthma are required.

Augmenting deficient IFN production in asthmatic patients could be a novel approach for therapeutic intervention in the treatment of virus induced asthma exacerbations. Recently, inhaled recombinant IFN- β therapy was studied in naturally occurring colds in mild-moderate asthmatics and it reduced moderate/severe exacerbations, symptoms and improved lung function in a sub-group of moderate asthmatics(403). Whether or not IFN- β was acting as an anti-viral cytokine, and/or was also effective due to antagonistic properties on T_{H2} cytokine pathways(404, 405) is currently unclear. Azithromycin has been shown to augment IFN responses during RV infection and reduce viral load *in vitro*(406). This property of azithromycin however has not been confirmed in clinical trials.

Neuronal dysfunction in asthma can be amplified by respiratory viral infections (407), but this is an area that requires further investigation to determine the degree to which this is important in man.

4.2.2 Bacteria, fungi & the microbiome

Bacteria are also associated with asthma exacerbations, specifically the atypical bacteria (*Mycoplasma* (*M.*) and *Chlamydophila* (*C.*) *pneumoniae*) with serological positivity rates as high as 40-60% in some studies(175, 373, 408-410), indicating that viral and atypical bacterial infections may act together to increase the risk of asthma exacerbations.

While asthmatics have increased susceptibility to respiratory bacterial infections(177, 178, 411), increased carriage of pathogenic respiratory bacteria identified by culture(412) and molecular techniques(91), in depth microbiome studies pre and post asthma exacerbation in longitudinal studies are lacking. Studies in mouse models have highlighted potential immune deviation properties of atypical bacteria that may help promote T_H2 immunity(413), and people with asthma have impaired IFN/T_H1 responses to bacterial polysaccharides(185, 186, 189, 190). In addition, viral infection impairs anti-bacterial innate immune responses(414) and increases bacterial adherence to BECs(415). There is therefore good evidence that respiratory bacterial infections are more common and more severe in asthma, and that viral infection can increase susceptibility to bacterial infection.

Acute wheezing episodes in children aged <3 years were associated with both bacterial and virus infection(118), however guidelines recommend that antibiotics should not be administered routinely in asthma exacerbations(416). Adults with asthma exacerbations treated with telithromycin showed statistically significantly greater reductions in asthma symptoms, improvement in lung function and faster recovery compared to placebo(410). However,





GA- 602077

Version: Final

acute liver toxicity limits telithromycin to severe life threatening infections. A recent study reported azithromycin treatment resulted in no statistically or clinically significant benefit in acute asthma exacerbations in adults, however, for each patient randomized, more than 10 were excluded because they had already received antibiotics, thus widespread antibiotic usage prevented firm conclusions being drawn from this study(417). Further studies of antibiotics in adults and children, in setting of low antibiotic use, are needed to determine which asthma patients may benefit from antibiotic therapy during asthma attacks(418).

Azithromycin treatment reduced the duration of acute episodes of asthma-like symptoms in 1 to 3 year old children(419). Furthermore, in 1 to 6 year old children with history of recurrent severe lower respiratory tract infections (LRTIs), azithromycin early during an apparent RTI reduced the likelihood of severe LRTI(420). In adults low-dose azithromycin prophylaxis for 6 months in subjects with exacerbation-prone severe asthma significantly reduced the rate of severe exacerbations and LRTIs requiring treatment with antibiotics in a predefined subgroup analysis of subjects with non-eosinophilic severe asthma(421). Further studies of antibiotics in both adults and children, in carefully phenotyped patient groups, are needed to determine in which asthma patients groups antibiotic therapy may prevent asthma attacks.

Emerging evidence indicates that the microbiota can influence the host's ability to fight respiratory infections(422), which could have profound implications for controlling exacerbations of asthma. Further studies of the effects of virus infections and of antibiotic therapy(423) on the airway microbiota and on risk and severity of asthma exacerbations are needed.

The role of fungi in precipitating asthma exacerbations is likely underappreciated. Exacerbations of allergic bronchopulmonary aspergillosis may be fungal in origin, yet clinical trials are few, and with disappointing results(424). Epidemics of severe exacerbations in the summer months are also thought likely related to alternaria sensitisation and exposure(425), while epidemics of exacerbations related to thunderstorms may be related to pollen and/or fungal exposures in sensitized individuals(426). Further research into the role of fungi in the aetiology of asthma exacerbations is needed.

4.2.3 Allergen exposure & T_H2 pathways

An important mechanistic process not completely understood is how virus infections exacerbate existing T_H2 immunity. Risk of hospital admissions for asthma exacerbations increases dramatically when allergen sensitisation and exposure occur together with a respiratory viral infection(302, 371) and mechanistic studies in human and mouse models of RV infection show induction of the pro- T_H2 cytokines IL-25 and IL-33 by RV, thus potentiating T_H2 immunity via increased IL-4, IL-5, IL-13 and eosinophilia within the airways(57, 58, 186). How important human respiratory viruses impact on the newly identified ILC2s is very understudied, but two studies suggest virus induced IL-25 and IL-33 potentiated T_H2 immunity via actions on these cells(57, 58).





GA- 602077

Version: Final

While corticosteroid based therapies impact only moderately on the severity of asthma exacerbations, anti- T_H2 biologics and anti-IgE therapies have shown a surprising efficacy(16, 18, 20, 23). Whether or not this added value is simply via suppressing allergen-induced T_H2 pathways, or is affecting other aspects of virus-allergen interactions (such as restoring deficient anti-viral immunity(24)) with a net beneficial effect on the outcome of virus-induced asthma exacerbations requires further exploration.

4.2.4 Pollution

A short-term increase in pollution exposure levels has been associated with asthma exacerbations: a $10\mu g/m^3$ increase in $PM_{2.5}$ correlated with emergency room visits and hospitalisation(427), while long-term exposure to NO_2 increased asthma hospitalisation in the elderly(428). Exposure to low dose air pollution acting directly on airway cells may cause chronic production of inflammatory mediators in asthmatics. These mediators may cause increased infiltration of airways tissue by immune cells and changes to the airways parenchyma leading to chronic remodelling(429). This could be responsible for a lowered threshold for other stimuli to cause asthma exacerbations. Acute exposure to high levels of air pollution alone may directly provoke asthma exacerbations in asthmatic individuals(430) either by action on innate & structural cells, and/or actions on the adaptive immune system(431).

There is emerging evidence providing a link between the increasing amounts of air pollutants/allergens in the environment and the worsening of symptoms in asthma(432). For example, air pollution has been linked to increases in hospital visits for respiratory disease(433). Synergy has also been suggested between air pollutants and allergens in the risk and exacerbation of asthma (especially in children), suggesting an advantage of avoiding co-exposure(372, 432).

Many epidemiological studies have identified ambient pollutant gases and airborne particles as risk factors for asthma exacerbations. However, the pollutants responsible remain unclear and causal relationships have often not been established. Among the most consistent observations are the direct effects of particle components on the generation of reactive oxygen species and induction of oxidative stress and inflammatory responses(434).

4.2.5 Diet and hormones

Individuals with ORA are at increased risk of asthma exacerbations(435). To the best of our knowledge, no studies have assessed the effect of obesity on the innate or adaptive immune response to respiratory viruses in asthmatics. Given that ~80% of asthma exacerbations are virus-induced(436), *ex vivo* and human challenge studies in this area may facilitate the development of novel preventative or therapeutic strategies.

Up to 20% of pregnant women with asthma experience asthma exacerbations during pregnancy and up to 6% require hospitalisations(325). During pregnancy the severity of asthma can change due to extrinsic as well as unknown factors. Uncontrolled asthma has a higher likelihood to deteriorate





Version: Final

during pregnancy and previous exacerbations predict emergency room visits during pregnancy(437), but its course is unpredictable and might even vary between pregnancies(325). Exacerbations have been reported to be more frequent during the early third trimester. Pregnancy has been reported to be associated with impaired anti-viral immunity in the presence(438) as well as absence(439) of asthma. Understanding the mechanisms that lead to deteriorations of asthma control during pregnancy has potential to improve

adverse outcome associated with poor asthma control during pregnancy.

4.2.6 Resolution of airway inflammation

A surprising recent observation has been that protectin D1 is induced by viral infection(440, 441) and viral RNA-sensing TLRs(278), and protectin D1 exhibits potent antiviral activity against influenza(440) while several resolvins and protectins can also enhance antibacterial defences(442). This raises the possibility of a dual role of SPMs in the regulation of inflammation and antiviral immunity in the respiratory tract with significant implications for the therapeutic potential of these molecules in the treatment or prevention of asthma exacerbations.

4.2.7 Psycho-social and behavioral factors

Stress, anxiety and depression are associated with an increased risk of exacerbation, less successful emergency treatment, and increased asthma hospitalisation rates(443, 444), and psychological co-morbidity may be associated with increased asthma mortality risks(445). The mechanisms underlying this increased risk are uncertain. Non-adherence with regular medication is common even in those with severe disease and exacerbation risk, and is associated with an increased risk of severe asthma attacks(31). Non-adherence occurs even in the period of deteriorating symptoms prior to and asthma attack(446). Risk stratification and targeted strategies to reduce exacerbation risk may be possible(447, 448).

4.3 Addressing the unmet need

While it is widely acknowledged that respiratory tract infections, particularly viral, account for the vast majority of asthma exacerbations (approximately 80%), a number of other factors are also likely to play a role. Identification of these other factors is vital if we are to reduce asthma morbidity and health care resources utilisation. Our incomplete understanding of asthma exacerbation pathophysiology and mechanisms is preventing development of effective treatments and preventive strategies. Many research programmes are seeing cross-European collaboration to integrate data, tools and techniques in order to explore related mechanisms and identify factors that play a decisive role in asthma. Similar cross-European collaboration is now needed to better understand aetiology and mechanism of asthma exacerbations.





GA-602077

GA-602077

© Copyright 2016, EARIP Consortium

. . .

Version: Final

At present there are few available therapies that target respiratory infections either non-specifically or specifically by virus type. A growing body of evidence suggests that augmenting deficient IFN production in asthmatic patients could be a novel target for therapeutic intervention in the context of virus induced asthma exacerbations. In addition, despite there being much interest and activity in developing viral inhibitors, no anti-virals have specifically been trialled in models of asthma exacerbations, despite the fact that such models exist and appear suitable(57). Again, the paucity of available anti-virals and a timely and effective model in which to test their efficacy must be considered an important unmet need. While IFN therapy is attractive in that it may target many viruses non-specifically, this argument can be extended to support the use of anti-viral therapies for asthma exacerbations. The recent developments in anti-viral therapy for important respiratory viruses have been covered elsewhere (449) and will not be systematically reviewed here. At the time of writing no anti-virals have specifically been trialled in models of asthma exacerbations, despite the multiple lines of evidence supporting the role of viruses as the most obvious asthma exacerbation trigger. Such studies are urgently needed.

Historically there has been much interest and activity in developing viral inhibitors. Small molecules and antibodies to viruses themselves or their receptors have been developed, or are currently undergoing clinical trials for RV(450, 451), RSV(452, 453) and influenza(454, 455). However, there has been slow progress in applying these to asthma. One question that is yet to be satisfactorily answered is in which model should they be tested? While the study of natural infections requires hundreds of subjects and is therefore expensive and time consuming, a useful alternative could be experimental infection models, which are performed with as few as 30 subjects. These have been successfully developed for RV(57, 186), and as RV is the most important viral trigger, there is an urgent need for these treatments to be tested in experimental RV challenge models to enable careful study of both efficacy and mechanisms. Another important consideration is which type and strain of virus should be used, and whether the experimental strains available behave in a manner comparable to wild type strains. This case is of particular importance for RV. The discovery of RV group C (RV-C)(456, 457), and its rise as probably the most clinically important group of RVs responsible for the majority of asthma exacerbation admissions(458-460) makes it a clear target for anti-viral therapy. There are several roadblocks concerning RV-C however; this is a difficult virus group to culture with a unique receptor(49), few laboratories have this expertise and there is as yet no RV-C stock available for experimental human challenge. Taken together, the case for the implementation of anti-viral therapy in asthma exacerbations, while clearly supported by the available scientific evidence, is hampered by several practical issues that requires further funding, careful planning and coordination between academia and industry to overcome.

Viral vaccines also represent an important unmet need, with either vaccines for some respiratory viruses completely unavailable or when available, of unclear benefit. New vaccine candidates against RSV including live





© Copyright 2016, EARIP Consortium GA- 602077

Version: Final

attenuated vaccines are currently being or have recently been tested in clinical trials(461-463). Immunization of mice with an RV capsid protein (VP0) containing highly conserved regions has been shown to induce cross-reactive cellular and humoral immune responses and therefore such capsid domains may prove useful for the development of a subunit RV vaccine(464). In the same model, RV VP1 vaccination has also been tested and provided some promising results in terms of cross-serotype antibody responses for future vaccine development(465). Further work to develop an effective and broadly cross-serotype protective vaccine against rhinovirus infections is urgently needed.

Regular vaccination against seasonal influenza is recommended for asthmatic children and adults(466, 467). A 2013 Cochrane review on the potential protective effect of the inactivated flu vaccine concluded that there was no significant reduction in the number, duration or severity of influenza-related asthma exacerbations in asthmatic children or adults(468). However, vaccinated children did display better symptom scores during influenza-positive weeks(468). Accumulating evidence now supports the concept of alternative influenza virus vaccine development strategies that will stimulate an immune response towards the conserved stalk rather than head domains of the virus hemagglutinin(469, 470). Such a universal vaccine would allow cross-immunisation against several influenza virus subtypes; however protective efficacy remains to be confirmed.

Our understanding of how asthma exacerbation frequency and severity relates to underlying asthma endotypes and phenotypes is also lacking. An important network running for over a decade is the National Institutes of Health-sponsored Severe Asthma Research Program (SARP), which is dedicated to the study of children and adults with severe asthma and aims at better understanding asthma endotypes with a focus on molecular and cellular processes. Subjects participating in the program undergo detailed characterization including clinical, physiological, radiological and genomic assessment and investigators seek to explore related mechanisms and identify factors that play a decisive role in disease history(471). In Europe, the BIOAIR and the U-BIOPRED severe asthma networks aim at improving our understanding of mechanisms and provide insight into endotypes or phenotypes such as those of frequent exacerbators (472), asthma exacerbation indicators (473), biomarkers (474) and pathophysiology of disease(475). These phenotypes need to be cross validated in different cohorts before they can be used to define patients. Asthma phenotyping has not been extensively studied among various neglected patient groups including smokers, drug users and the elderly. Identification of clusters may help recognize patients particularly prone to develop asthma exacerbations. Recent work towards this direction has defined phenotypic groups among elderly patients(476-478) and has attempted to profile asthma exacerbations in drug users(479). Finally, there is great potential for novel diagnostic technologies (in particular non-invasive techniques and biomarkers) to be developed to support prediction(480), assessment and management of





GA- 602077

Version: Final

asthma exacerbations(481, 482) and future studies in the field would help personalize intervention strategies.

The existing evidence points to the fact that asthma exacerbations are likely a result of synergistic and/or additive interactions between several factors. Perhaps most evidence is available for virus and T_H2 interactions, and how these precipitate asthma exacerbations in allergic asthma. No study has yet linked impaired IFN responses with a known asthma phenotype, however most studies observed this phenomenon in atopic asthma(157, 185, 187, 391, 393, 395). Sputum eosinophils in the past have been related to asthma exacerbation risk(483) and impaired IFN-λs in cultured cells negatively correlated with sputum eosinophilia(185) and both serum IgE and IL-4 staining levels in biopsies(187). Mechanistically, much evidence definitively shows that type I and III IFNs and T_H2 cytokines can exhibit potent negative regulation on the expression or actions of each other. IFN- α for example, can suppress T_H2 cell polarisation in pure T cell and mixed leukocyte culture systems(405, 484). IFN- λ (IL-28) when given to the airways can suppress the generation of allergic airways inflammation in the ovalbumin sensitisation and challenge model in mice, which is T_H2 driven(404). Conversely, crosslinking the Fc_ER1 on DCs prior to influenza virus or RV challenge down regulates IFN- α (395, 396); thus the overall evidence strongly underscores many potential mechanisms of counter-regulation of T_H2 cytokines and type I and III IFNs. Interestingly, SOCS1, linking with impaired IFN is a T_H2 inducible negative regulator and in the airway epithelium, correlates with serum IgE(399). There is still a lot to learn about how asthma exacerbations are precipitated, including how the underlying pathophysiology of asthma can be subtly affected by another insult or stimulus. This therefore represents an unmet need. The interactions between pathogen sensing, environmental microbiota and genetics are likely also important in asthma exacerbations and future research should explore the role of non-T_H2 pathways and other processes including inflammasome activation in airway remodelling and how these processes affect steroid resistance.

Further studies investigating the effectiveness of macrolide (including derivatives with and without antibiotic properties) and other antibiotics in treatment and prevention of acute exacerbations of asthma in adults and children in settings of low antibiotic usage, and studies including stratification on blood/sputum cell counts, are required. Further studies investigating the effects of viral infection on secondary disturbances of the microbiome in adult and children with asthma exacerbations are clearly needed, as well as studies investigating the mechanisms of impaired host defense against bacterial infections in asthma.

Asthma exacerbations constitute major outstanding challenges to treatment. Given the generation of danger signals during an exacerbation, the role of alarmins and of ILCs in exacerbations needs to be clarified. Understanding the role of these new ILC populations could eventually lead to new therapeutic targets. Similar work is needed on the role of neuronal mechanisms and respiratory reflexes.





© Copyright 2016, EARIP Consortium GA- 602077

Version: Final

The role of pollution in precipitating asthma exacerbations is also incompletely understood. Polymorphisms in glutathione S-transferases (GSTM1 and GSTP1) that facilitate the elimination of reactive oxygen species have been associated with breathing difficulties and respiratory symptoms in asthmatic children following increases in ambient O_3 concentrations(485) and an altered response to combined exposures to ragweed pollen and DEPs(486). How the innate immune response of asthmatics to infections is altered by air pollution should also be studied. Further research examining the influence of gene polymorphisms which could predict asthmatic patients that will benefit from antioxidant supplementation would be of interest.

Patients at higher risk of exacerbations may be identified through viable clinical strategies, with exacerbation prevention strategies targeted to improve exacerbation outcomes. Psychological and behavioural factors, for example substance abuse(487), may increase exacerbation risk, and may be amenable to intervention, although the mechanisms underlying this increased risk are not fully understood.





© Copyright 2016,

GA- 602077

EARIP Consortium Version: Final

5 Summary & conclusions

Asthma research now has loftier aims than in previous times, in that we now aim to prevent disease occurring, to cure it when it does occur, and if these are not possible, to treat and control asthma to prevent progression to severe disease, as well as to prevent and treat acute attacks of asthma. There is also a focus on studying mechanisms in different asthma phenotypes, including atopic and non-atopic asthma, so that new therapies can be best targeted towards the most appropriate phenotype.

This analysis of available literature on asthma mechanisms has delineated the most important unmet needs in asthma research and proposes how these might be addressed. A stratification into asthma onset, progression of stable asthma, and asthma exacerbations allows identification of where in the continuity of asthma research future efforts need to be directed, indicating who would benefit and to what extent such efforts would impact on people with asthma. Our review of the literature and discussions with colleagues have clearly identified many important unmet needs. The most important mechanisms involved in asthma onset, asthma progression and asthma exacerbation and the strength of the supportive evidence for each mechanism are summarised in Table 1. The most important methodologies or technologies that are required to beneficially impact on asthma research in future years are summarised in **Table 2**. Key unmet needs in understanding asthma mechanisms that we have identified are listed in Table 3 and are pictorially represented in Figures 1-3. These are the major areas in which future investment is needed to drive forward asthma research to hasten progress in preventing asthma developing, controlling and curing established asthma, and in preventing and treating asthma attacks and their associated morbidity and mortality.

Firstly, we have shown that research into asthma onset suffers from several shortcomings in research effort. Importantly, this is an area that if addressed, could have the biggest impact, in both the short and long term, by preventing asthma occurring in the first place. We need to better understand the onset of both allergic and non-allergic asthma, including the role of air pollution. The development of new models, new sampling methods, and the structuring of research in productive, collaborative initiatives will deliver new information in the longer term and with time, new therapeutic targets for asthma inception. Funding this area of asthma research to amend these shortfalls should thus be considered a long term proposition. However, there are also more immediate goals to be achieved. The study of new vaccines and anti-virals, or application of existing ones to determine whether viruses are functionally associated with asthma onset could lead to exciting opportunities for asthma prevention. Even reducing respiratory viral infections by 20% in young infants could have vast financial effects on the healthcare burden associated with asthma. Prevention of acute respiratory viral infections could prevent wheeze and perhaps allergic sensitisation and asthma onset. Funding this research would deliver a short term gain, as anti-virals are already in phase II/III (for RSV) and would have to be applied to the right population at highest risk and





GA- 602077

Version: Final

at the optimal age window in clinical trials. For RV the prospects of pathway to impact are further away, as anti-viral programs are still in development. However vaccine candidates while still experimental, do hold promise and need to be prioritised.

Targeting mechanisms of progression of stable asthma to severe disease requires more basic research to hasten development of a more comprehensive array of disease modifying therapies that better address the needs of severe therapy resistant asthma. The studies should include the different endotypes and phenotypes of asthma and specific mechanisms should include neural pathways, airway remodelling and the understudied adverse effects of asthma therapies. The longer term prospects of such programs would be to achieve better asthma control with better targeted novel treatments, to reduce progression to severe asthma and to cure established asthma. Other areas that need more research include the role of comorbidities, better clinically defined groups and models, and also aspects of the microbiome and bacteria-virus interactions, together with their interactions with pollutants. These last two subjects would benefit specifically from basic research including new in vitro and in vivo models. The value in funding these areas should be considered a longer term proposition, with the initial aim of developing models and techniques, and applying them to gain new disease insights, which would eventually lead to new targets or agents for therapy.

Asthma exacerbations cause major morbidity, mortality and health care costs. Much more research funding needs to be channelled into anti-viral therapy and vaccine development programs to benefit asthma exacerbations, which have a clear viral aetiology. Funding also needs to be directed at development of better pre-clinical models of virus infections, and of interactions between virus infections, allergen exposure and airway microbes to study mechanisms of exacerbation. A major priority is the development of human challenge models in asthma with viruses, allergens and pollutants to enable identification of novel mechanisms of disease to lead to new therapeutic strategies. Development of human challenge models will also permit early proof of concept and proof of mechanism studies in development programs of novel therapeutic approaches for prevention/treatment of exacerbations. Funding also needs to prioritise better understanding of the mechanisms of increased susceptibility to viral and bacterial infection in asthma and better identification of the endotypes and phenotypes of asthma to which these specific mechanisms apply. More research is also needed to better understand of the interactions between T_H2 pathways and anti-viral immunity in exacerbation pathogenesis.

In summary, this paper describes a compelling picture of unmet need in asthma. Researchers, funders and the pharmaceutical industry need to work together to address these important priorities to prevent, treat and eventually cure asthma.





GA- 602077 Version: Final

Table 1: Mechanisms involved in asthma inception, progression and exacerbation

Phase	Mechanism & Strength of Supportive Evidence
Onset	Genetics ^{20,††}
	Allergen exposure & sensitization, T _H 2 pathways ^{§§§} Viruses ^{‡‡,§§§,∂∂,††}
	Viruses ^{+1-333,00,11} Bacteria ^{∂,††}
	Pollution ^{‡,††}
	Diet ^{§§,††}
	Hormones ^{††}
	Stress and psychological factors ^{††}
Progression	Allergen exposure & sensitization, T _H 2 pathways ^{‡‡‡‡,§§§,∂∂∂,†††}
	Bacteria ^{††}
	Pollution ^{‡,††}





© Copyright 2016,

EARIP Consortium	Version: Final
	Airway remodeling ^{‡‡‡,§§§,∂∂∂,†}
	Resolution of inflammation ^{T,SS,000,T}
	Immune responsiveness vs tolerance ^{‡,§§§,∂∂∂,†}
	Adverse effects of therapies ^{+,1}
	Diet ^{§§,∂∂,††}
	Hormones ^{††}
	Stress and psychological factors ^{††}
Exacerbation	Genetics ^{∂,†}
	Allergen exposure & sensitization, T _H 2 pathways ^{‡‡,§§§,∂∂,†††} Viruses ^{‡‡‡,§§§,∂∂∂,†††}
	Viruses ^{‡‡‡,§§§,∂∂∂,†††}
	Bacteria [†]
	Pollution ^{‡,††}
	Stress and psychological factors [†]

Evidence: [‡]*in vitro* model, [§]Animal model, [∂]*ex vivo* patient sample, [†]longitudinal cohort or cross-sectional clinical study. 3 symbols= very strong evidence, 2= validated or repeatable evidence, 1= weak evidence.





Version: Final

© Copyright 2016, EARIP Consortium GA- 602077

Table 2: Methodologies or technologies required to beneficially impact on asthma research

Method	Description			
NGS eQTL	RNA-seq, array techniques that capture all expressed genes in a sample. Can			
epigenetic, and				
array platforms	expression of genes in patient samples proving some translation of genetic effects			
GWAS and	GWAS identifies SNPs associated with diseases such as asthma. An association			
exome	of a SNP does not necessarily inform on a mechanism. Exome sequence refers to			
sequencing	sequencing the exons and other specified parts of that gene, such as adjacent			
	promoter sequences			
Metabolomics	Analysis of biomarkers either by mass spectrometry or other protein analysis eg.			
	bromotyrosine in urine			
Lipidomics	Analysis of pro-inflammatory and pro-resolving lipid networks by liquid chromatography-tandem mass spectrometry in sputum, BAL or plasma			
Volatile organic	Detects organic compounds identified by mass spectrometry in non-invasive			
compounds	samples such as exhaled breath. Can detect viruses and bacteria in theory.			
(VOCs)				
Nasosorption and	Detects proteins in very small volumes of biological fluid. Has been applied to the			
Bronchosorption	nose and lung in detecting rare cytokines.			
Mathematical	Can predict the efficacy of a new therapy, or how a new model may behave. This			
modelling	may replace executing the actual experimental work.			
Machine learning	Can interrogate big data sets in a free and non-hypothesis-driven manner. Can be			
	useful in the formulation of new hypotheses.			
Better pre-clinical	Needed to study mechanisms of interactions between virus infections, allergen			
models	exposure, defective innate immunity and airway microbes in initiation, progression			
	and exacerbation of asthma.			
Human challenge				
models (allergen, virus, pollution)	therapeutic approaches and enable identification of novel mechanisms of disease to lead to new therapeutic strategies.			
Anti-viral	Needed for use in early life in studies on asthma prevention and for prevention and			
therapies and				
vaccines				
Pre-clinical and	Needed to better understand mechanisms of neuronal dysfunction, airway			
clinical models of	remodelling etc in progression and severity of asthma. Pre-clinical models should			
disease	include development of models in and reagents for guinea pigs, rats, rabbits and			
Deputation based	dogs as well as mice.			
Population-based birth cohort	Needed to integrate genetic and epigenetic host factors with studies of virus infections, allergens and other environmental exposures and gut and airway			
studies	microbes, with studies of host innate and acquired immune reactivity vs tolerance			
Studies	with systems biologic analytic methodologies, to understand mechanisms			
	asthma onset.			
Clinic/community	Needed to investigate disease mechanisms and to validate mechanistic findings			
based studies of	from pre-clinical models in the human setting.			
paediatric and				
adult asthmatics.	Mandad to investigate officers and resolves of injection and the			
Clinical trials	Needed to investigate efficacy and mechanisms of injection and sublingual immunotherapy, and of novel asthma therapies.			
	ininiunounerapy, and or nover asumia unerapies.			

NGS = next generation sequencing loci GWAS = genome wide association studies polymorphism eQTL = expression quantitative trait SNP = single nucleotide





© Copyright 2016, EARIP Consortium

Version: Final





Version: Final

© Copyright 2016, EARIP Consortium

Table 3: Key unmet needs in understanding asthma mechanisms and the nature of the unmet need

Key Unmet Need	Where	Nature of the
	needed	unmet need*
Mechanisms of onset of both allergic and non-allergic asthma	Onset	Birth cohort studies, pre-clinical models, novel targets, new therapies, clinical studies
Role of early life viruses and bacteria as causative agents of asthma	Onset	Pre-clinical models, novel targets, new therapies, clinical studies
Understanding mechanisms underlying the progression of virus-induced early wheezing to later well-defined asthma	Onset Progression	Birth cohort studies, pre-clinical models, novel targets, new therapies, clinical studies
Identifying early life risk factors to enable accurate prediction of which early wheezers will later develop asthma to permit primary prevention	Onset Progression	Birth cohort studies, pre-clinical models, novel targets, new therapies, clinical studies
Mechanism of progression to treatment refractory severe asthma	Progression	Pre-clinical models, targets, therapies, clinical studies
Role of pro-T _H 2 cytokines IL-25, IL-33 and TSLP and of PGD ₂ /CRTH2	Onset Progression Exacerbation	Pre-clinical models, targets, therapies, clinical studies
Understanding mechanisms of development of immune reactivity vs immune tolerance in asthma	Onset Progression Exacerbation	Pre-clinical models, novel targets, new therapies, clinical studies
Mechanisms of non-allergic asthma	Onset Progression	Pre-clinical models, targets, therapies, clinical studies
Role of air pollution, occupational & other environmental exposures, second hand smoke	Onset Progression Exacerbation	Pre-clinical models, targets, therapies, clinical studies
Protective vs pathogenic role of gut and lung microbiomes	Onset,	Pre-clinical
* * * * *		



© Copyright 2016,

EARIP Consortium

Version: Final

Progression, models, targets, Exacerbation clinical studies Pre-clinical New anti-viral therapies and vaccines Onset. Exacerbation models, therapies, clinical studies Understanding mechanisms of increased susceptibility to viral Progression Pre-clinical and bacterial infection in asthma Exacerbation models, novel targets, new therapies Understanding asthma phenotypes expressing deficient anti-Progression Pre-clinical viral immunity, and the mechanisms involved Exacerbation models, novel targets, new therapies Understanding mechanisms of action of pro-resolving lipid Progression Pre-clinical Exacerbation models, novel mediators in asthma targets, new therapies Understanding neural pathways Onset, Pre-clinical Progression models, novel Exacerbation targets, new therapies, clinical studies Understanding role of obesity, diet, hormones & pregnancy Onset, Pre-clinical Progression models, novel Exacerbation targets, new therapies, clinical studies Mechanisms of airway remodelling Pre-clinical Progression models, novel targets, new therapies, clinical studies

Mechanisms underpinning relationship between psychological

Mechanisms of adverse effects of asthma therapies

Translating genetic findings to disease mechanisms



factors and asthma



Pre-clinical

Pre-clinical

Pre-clinical

studies

studies

studies

models, clinical

models, clinical

models, clinical

Onset,

Onset

Progression

Exacerbation

Progression,

Exacerbation

Exacerbation

^{*} Pre-clinical models may include *in vitro* studies in human cells as well as *in vivo* studies in animal models, clinical studies may include human challenge (virus, allergen, pollution) models as well as mechanistic and interventional clinical studies.

Version: Final

© Copyright 2016, EARIP Consortium GA- 602077

Figure legends

Figure 1. Major mechanisms of asthma onset. The major cell types, cytokines and mechanisms involved in asthma onset are depicted, with major mechanisms/research areas requiring substantial further investment highlighted in boxes.

Figure 2. Major mechanisms of asthma progression and severity. The major cell types, cytokines and mechanisms involved in asthma progression are depicted, with major mechanisms/research areas requiring substantial further investment highlighted in boxes.

Figure 3. Major mechanisms of asthma exacerbations. The major cell types, cytokines and mechanisms involved in asthma exacerbations are depicted, with major mechanisms/research areas requiring substantial further investment highlighted in boxes.





GA- 602077 Version: Final

6 References

- 1. Kupczyk M SS, James A, Walker S, Dahlén S-E; On behalf of the EARIP consortium (WP 3). The road-map to improve asthma phenotyping and thereby the implementation of precision and personalised medicine for asthma. (submitted) 2016.
- Selroos O, Kupczyk M, Kuna P, Lacwik P, Bousquet J, Brennan D, Palkonen S, Contreras J, FitzGerald M, Hedlin G, Johnston SL, Louis R, Metcalf L, Walker S, Moreno-Galdo A, Papadopoulos NG, Rosado-Pinto J, Powell P, Haahtela T. National and regional asthma programmes in Europe. *Eur Respir Rev* 2015; 24: 474-483.
- 3. Walker SM, Akdis C, Dahlen SE, Djukanovic R, Edwards J, Garcia-Marcos L, Johnston S, Kupczyk M, Martin T, Myles D, Palkonen S, Papadopoulos N, Powell P, Riley J. Building the investment case for asthma R&D: the European Asthma Research and Innovation Partnership argument. *Clin Exp Allergy* 2016; 46: 1136-1138.
- 4. Masefield SCea. Driving Investment in Asthma Research in Europe: Priorities to Prevent, Cure and Manage Asthma More Effectively. *Am J Respir Crit Care Med* 2016; 193: A2668.
- 5. Bateman ED, Hurd SS, Barnes PJ, Bousquet J, Drazen JM, FitzGerald M, Gibson P, Ohta K, O'Byrne P, Pedersen SE, Pizzichini E, Sullivan SD, Wenzel SE, Zar HJ. Global strategy for asthma management and prevention: GINA executive summary. *Eur Respir J* 2008; 31: 143-178.
- 6. Society ER. European Respiratory Society White Book. [cited 2015 February 27].
- 7. Anderson GP. Endotyping asthma: new insights into key pathogenic mechanisms in a complex, heterogeneous disease. *Lancet* 2008; 372: 1107-1119.
- 8. Wenzel SE. Asthma: defining of the persistent adult phenotypes. *Lancet* 2006; 368: 804-813.
- 9. Asher MI, Montefort S, Bjorksten B, Lai CK, Strachan DP, Weiland SK, Williams H. Worldwide time trends in the prevalence of symptoms of asthma, allergic rhinoconjunctivitis, and eczema in childhood: ISAAC Phases One and Three repeat multicountry cross-sectional surveys. *Lancet* 2006; 368: 733-743.
- 10. FitzGerald JM, Becker A, Sears MR, Mink S, Chung K, Lee J. Doubling the dose of budesonide versus maintenance treatment in asthma exacerbations. *Thorax* 2004; 59: 550-556.
- 11. Pauwels RA, Lofdahl CG, Postma DS, Tattersfield AE, O'Byrne P, Barnes PJ, Ullman A. Effect of inhaled formoterol and budesonide on exacerbations of asthma. Formoterol and Corticosteroids Establishing Therapy (FACET) International Study Group. *N Engl J Med* 1997; 337: 1405-1411.
- 12. O'Byrne PM, Bisgaard H, Godard PP, Pistolesi M, Palmqvist M, Zhu Y, Ekstrom T, Bateman ED. Budesonide/formoterol combination therapy as both maintenance and reliever medication in asthma. *Am J Respir Crit Care Med* 2005; 171: 129-136.
- 13. Bjermer L, Bisgaard H, Bousquet J, Fabbri LM, Greening A, Haahtela T, Holgate ST, Picado C, Leff JA. Montelukast or salmeterol combined with an inhaled steroid in adult asthma: design and rationale of a randomized, double-blind comparative study (the IMPACT Investigation of Montelukast as a Partner Agent for Complementary Therapy-trial). *Respir Med* 2000; 94: 612-621.





GA- 602077 Version: Final

14. Wilson AM, Dempsey OJ, Sims EJ, Lipworth BJ. Evaluation of salmeterol or

- 14. Wilson AM, Dempsey OJ, Sims EJ, Lipworth BJ. Evaluation of salmeterol or montelukast as second-line therapy for asthma not controlled with inhaled corticosteroids. *Chest* 2001; 119: 1021-1026.
- 15. Villaran C, O'Neill SJ, Helbling A, van Noord JA, Lee TH, Chuchalin AG, Langley SJ, Gunawardena KA, Suskovic S, Laurenzi M, Jasan J, Menten J, Leff JA. Montelukast versus salmeterol in patients with asthma and exercise-induced bronchoconstriction. Montelukast/Salmeterol Exercise Study Group. *J Allergy Clin Immunol* 1999; 104: 547-553.
- 16. Pavord ID, Korn S, Howarth P, Bleecker ER, Buhl R, Keene ON, Ortega H, Chanez P. Mepolizumab for severe eosinophilic asthma (DREAM): a multicentre, double-blind, placebo-controlled trial. *Lancet* 2012; 380: 651-659.
- 17. Wenzel S, Castro M, Corren J, Maspero J, Wang L, Zhang B, Pirozzi G, Sutherland ER, Evans RR, Joish VN, Eckert L, Graham NM, Stahl N, Yancopoulos GD, Louis-Tisserand M, Teper A. Dupilumab efficacy and safety in adults with uncontrolled persistent asthma despite use of medium-to-high-dose inhaled corticosteroids plus a long-acting beta2 agonist: a randomised double-blind placebo-controlled pivotal phase 2b dose-ranging trial. *Lancet* 2016; 388: 31-44.
- 18. Wenzel S, Ford L, Pearlman D, Spector S, Sher L, Skobieranda F, Wang L, Kirkesseli S, Rocklin R, Bock B, Hamilton J, Ming JE, Radin A, Stahl N, Yancopoulos GD, Graham N, Pirozzi G. Dupilumab in persistent asthma with elevated eosinophil levels. *N Engl J Med* 2013; 368: 2455-2466.
- 19. Wenzel S, Wilbraham D, Fuller R, Getz EB, Longphre M. Effect of an interleukin-4 variant on late phase asthmatic response to allergen challenge in asthmatic patients: results of two phase 2a studies. *Lancet* 2007; 370: 1422-1431.
- 20. Corren J, Lemanske RF, Hanania NA, Korenblat PE, Parsey MV, Arron JR, Harris JM, Scheerens H, Wu LC, Su Z, Mosesova S, Eisner MD, Bohen SP, Matthews JG. Lebrikizumab treatment in adults with asthma. *N Engl J Med* 2011; 365: 1088-1098.
- 21. Holgate ST, Chuchalin AG, Hebert J, Lotvall J, Persson GB, Chung KF, Bousquet J, Kerstjens HA, Fox H, Thirlwell J, Cioppa GD. Efficacy and safety of a recombinant anti-immunoglobulin E antibody (omalizumab) in severe allergic asthma. *Clin Exp Allergy* 2004; 34: 632-638.
- 22. Bousquet J, Wenzel S, Holgate S, Lumry W, Freeman P, Fox H. Predicting response to omalizumab, an anti-IgE antibody, in patients with allergic asthma. *Chest* 2004; 125: 1378-1386.
- 23. Busse WW, Morgan WJ, Gergen PJ, Mitchell HE, Gern JE, Liu AH, Gruchalla RS, Kattan M, Teach SJ, Pongracic JA, Chmiel JF, Steinbach SF, Calatroni A, Togias A, Thompson KM, Szefler SJ, Sorkness CA. Randomized trial of omalizumab (anti-IgE) for asthma in inner-city children. *N Engl J Med* 2011; 364: 1005-1015.
- 24. Teach SJ, Gill MA, Togias A, Sorkness CA, Arbes SJ, Jr., Calatroni A, Wildfire JJ, Gergen PJ, Cohen RT, Pongracic JA, Kercsmar CM, Khurana Hershey GK, Gruchalla RS, Liu AH, Zoratti EM, Kattan M, Grindle KA, Gern JE, Busse WW, Szefler SJ. Preseasonal treatment with either omalizumab or an inhaled corticosteroid boost to prevent fall asthma exacerbations. J Allergy Clin Immunol 2015; 136: 1476-1485.





Version: Final

GA-602077

25. Virchow JC, Backer V, Kuna P, Prieto L, Nolte H, Villesen HH, Ljorring C, Riis B, de Blay F. Efficacy of a House Dust Mite Sublingual Allergen Immunotherapy Tablet in Adults With Allergic Asthma: A Randomized Clinical Trial. *Jama* 2016; 315: 1715-1725.

- 26. Juniper EF, Wisniewski ME, Cox FM, Emmett AH, Nielsen KE, O'Byrne PM. Relationship between quality of life and clinical status in asthma: a factor analysis. *Eur Respir J* 2004; 23: 287-291.
- 27. Teeter JG, Bleecker ER. Relationship between airway obstruction and respiratory symptoms in adult asthmatics. *Chest* 1998; 113: 272-277.
- 28. Thomas M, Bruton A, Moffat M, Cleland J. Asthma and psychological dysfunction. *Primary care respiratory journal : journal of the General Practice Airways Group* 2011; 20: 250-256.
- 29. Rimington LD, Davies DH, Lowe D, Pearson MG. Relationship between anxiety, depression, and morbidity in adult asthma patients. *Thorax* 2001; 56: 266-271.
- 30. Gamble J, Stevenson M, McClean E, Heaney LG. The prevalence of nonadherence in difficult asthma. *Am J Respir Crit Care Med* 2009; 180: 817-822.
- 31. Suissa S, Ernst P, Kezouh A. Regular use of inhaled corticosteroids and the long term prevention of hospitalisation for asthma. *Thorax* 2002; 57: 880-884.
- 32. Bousquet J, Gern JE, Martinez FD, Anto JM, Johnson CC, Holt PG, Lemanske RF, Jr., Le Souef PN, Tepper RS, von Mutius ER, Arshad SH, Bacharier LB, Becker A, Belanger K, Bergstrom A, Bernstein DI, Cabana MD, Carroll KN, Castro M, Cooper PJ, Gillman MW, Gold DR, Henderson J, Heinrich J, Hong SJ, Jackson DJ, Keil T, Kozyrskyj AL, Lodrup Carlsen KC, Miller RL, Momas I, Morgan WJ, Noel P, Ownby DR, Pinart M, Ryan PH, Schwaninger JM, Sears MR, Simpson A, Smit HA, Stern DA, Subbarao P, Valenta R, Wang X, Weiss ST, Wood R, Wright AL, Wright RJ, Togias A, Gergen PJ. Birth cohorts in asthma and allergic diseases: report of a NIAID/NHLBI/MeDALL joint workshop. *J Allergy Clin Immunol* 2014; 133: 1535-1546.
- 33. Sears MR, Greene JM, Willan AR, Wiecek EM, Taylor DR, Flannery EM, Cowan JO, Herbison GP, Silva PA, Poulton R. A longitudinal, population-based, cohort study of childhood asthma followed to adulthood. *N Engl J Med* 2003; 349: 1414-1422.
- 34. Tai A, Tran H, Roberts M, Clarke N, Gibson AM, Vidmar S, Wilson J, Robertson CF. Outcomes of childhood asthma to the age of 50 years. *J Allergy Clin Immunol* 2014; 133: 1572-1578 e1573.
- 35. Bossley CJ, Fleming L, Gupta A, Regamey N, Frith J, Oates T, Tsartsali L, Lloyd CM, Bush A, Saglani S. Pediatric severe asthma is characterized by eosinophilia and remodeling without T(H)2 cytokines. *J Allergy Clin Immunol* 2012; 129: 974-982 e913.
- 36. Turato G, Barbato A, Baraldo S, Zanin ME, Bazzan E, Lokar-Oliani K, Calabrese F, Panizzolo C, Snijders D, Maestrelli P, Zuin R, Fabbri LM, Saetta M. Nonatopic children with multitrigger wheezing have airway pathology comparable to atopic asthma. *Am J Respir Crit Care Med* 2008; 178: 476-482.
- 37. Martinez FD, Vercelli D. Asthma. Lancet 2013; 382: 1360-1372.
- 38. Hafkamp-de Groen E, Lingsma HF, Caudri D, Levie D, Wijga A, Koppelman GH, Duijts L, Jaddoe VW, Smit HA, Kerkhof M, Moll HA, Hofman A, Steyerberg EW, de Jongste JC, Raat H. Predicting asthma in preschool children with





© Copyright 2016, EARIP Consortium GA- 602077

Version: Final

- asthma-like symptoms: validating and updating the PIAMA risk score. *J Allergy Clin Immunol* 2013; 132: 1303-1310.
- 39. Castro-Rodriguez JA. Another predictive score for childhood asthma: the search remains. *J Allergy Clin Immunol Pract* 2014; 2: 716-718.
- 40. Hasler G, Gergen PJ, Kleinbaum DG, Ajdacic V, Gamma A, Eich D, Rossler W, Angst J. Asthma and panic in young adults: a 20-year prospective community study. *Am J Respir Crit Care Med* 2005; 171: 1224-1230.
- 41. Brumpton BM, Leivseth L, Romundstad PR, Langhammer A, Chen Y, Camargo CA, Jr., Mai XM. The joint association of anxiety, depression and obesity with incident asthma in adults: the HUNT study. *International journal of epidemiology* 2013; 42: 1455-1463.
- 42. Walker ML, Holt KE, Anderson GP, Teo SM, Sly PD, Holt PG, Inouye M. Elucidation of pathways driving asthma pathogenesis: development of a systems-level analytic strategy. Front Immunol 2014; 5: 447.
- 43. Jackson DJ, Evans MD, Gangnon RE, Tisler CJ, Pappas TE, Lee WM, Gern JE, Lemanske RF, Jr. Evidence for a causal relationship between allergic sensitization and rhinovirus wheezing in early life. *Am J Respir Crit Care Med* 2011; 185: 281-285.
- 44. Ong BH, Gao Q, Phoon MC, Chow VT, Tan WC, Van Bever HP. Identification of human metapneumovirus and Chlamydophila pneumoniae in children with asthma and wheeze in Singapore. Singapore Med J 2007; 48: 291-293.
- 45. Caliskan M, Bochkov YA, Kreiner-Moller E, Bonnelykke K, Stein MM, Du G, Bisgaard H, Jackson DJ, Gern JE, Lemanske RF, Jr., Nicolae DL, Ober C. Rhinovirus wheezing illness and genetic risk of childhood-onset asthma. *N Engl J Med* 2013; 368: 1398-1407.
- 46. Jackson DJ, Gangnon RE, Evans MD, Roberg KA, Anderson EL, Pappas TE, Printz MC, Lee WM, Shult PA, Reisdorf E, Carlson-Dakes KT, Salazar LP, DaSilva DF, Tisler CJ, Gern JE, Lemanske RF, Jr. Wheezing rhinovirus illnesses in early life predict asthma development in high-risk children. *Am J Respir Crit Care Med* 2008; 178: 667-672.
- 47. Sigurs N, Aljassim F, Kjellman B, Robinson PD, Sigurbergsson F, Bjarnason R, Gustafsson PM. Asthma and allergy patterns over 18 years after severe RSV bronchiolitis in the first year of life. *Thorax* 2010; 65: 1045-1052.
- 48. Bonnelykke K, Sleiman P, Nielsen K, Kreiner-Moller E, Mercader JM, Belgrave D, den Dekker HT, Husby A, Sevelsted A, Faura-Tellez G, Mortensen LJ, Paternoster L, Flaaten R, Molgaard A, Smart DE, Thomsen PF, Rasmussen MA, Bonas-Guarch S, Holst C, Nohr EA, Yadav R, March ME, Blicher T, Lackie PM, Jaddoe VW, Simpson A, Holloway JW, Duijts L, Custovic A, Davies DE, Torrents D, Gupta R, Hollegaard MV, Hougaard DM, Hakonarson H, Bisgaard H. A genome-wide association study identifies CDHR3 as a susceptibility locus for early childhood asthma with severe exacerbations. *Nat Genet* 2014; 46: 51-55.
- 49. Bochkov YA, Watters K, Ashraf S, Griggs TF, Devries MK, Jackson DJ, Palmenberg AC, Gern JE. Cadherin-related family member 3, a childhood asthma susceptibility gene product, mediates rhinovirus C binding and replication. *Proc Natl Acad Sci U S A* 2015; 112: 5485-5490.
- 50. Culley FJ, Pollott J, Openshaw PJ. Age at first viral infection determines the pattern of T cell-mediated disease during reinfection in adulthood. *J Exp Med* 2002; 196: 1381-1386.





Version: Final

51. Kaiko GE, Loh Z, Spann K, Lynch JP, Lalwani A, Zheng Z, Davidson S, Uematsu S, Akira S, Hayball J, Diener KR, Baines KJ, Simpson JL, Foster PS, Phipps S. Toll-like receptor 7 gene deficiency and early-life Pneumovirus infection interact to predispose toward the development of asthma-like pathology in mice. *J Allergy Clin Immunol* 2013; 131: 1331-1339 e1310.

- 52. Fajt ML, Wenzel SE. Asthma phenotypes and the use of biologic medications in asthma and allergic disease: the next steps toward personalized care. *J Allergy Clin Immunol* 2015; 135: 299-310; quiz 311.
- 53. Bel EH, Wenzel SE, Thompson PJ, Prazma CM, Keene ON, Yancey SW, Ortega HG, Pavord ID, Investigators S. Oral glucocorticoid-sparing effect of mepolizumab in eosinophilic asthma. *N Engl J Med* 2014; 371: 1189-1197.
- 54. Ortega HG, Liu MC, Pavord ID, Brusselle GG, FitzGerald JM, Chetta A, Humbert M, Katz LE, Keene ON, Yancey SW, Chanez P, Investigators M. Mepolizumab treatment in patients with severe eosinophilic asthma. *N Engl J Med* 2014; 371: 1198-1207.
- 55. McKenzie AN, Spits H, Eberl G. Innate lymphoid cells in inflammation and immunity. *Immunity* 2014; 41: 366-374.
- 56. Spits H, Artis D, Colonna M, Diefenbach A, Di Santo JP, Eberl G, Koyasu S, Locksley RM, McKenzie AN, Mebius RE, Powrie F, Vivier E. Innate lymphoid cells--a proposal for uniform nomenclature. *Nat Rev Immunol* 2013; 13: 145-149.
- 57. Jackson DJ, Makrinioti H, Rana BM, Shamji BW, Trujillo-Torralbo MB, Footitt J, Del-Rosario J, Telcian AG, Nikonova A, Zhu J, Aniscenko J, Gogsadze L, Bakhsoliani E, Traub S, Dhariwal J, Porter J, Hunt D, Hunt T, Stanciu LA, Khaitov M, Bartlett NW, Edwards MR, Kon OM, Mallia P, Papadopoulos NG, Akdis CA, Westwick J, Edwards MJ, Cousins DJ, Walton RP, Johnston SL. IL-33-dependent Type 2 Inflammation During Rhinovirus-induced Asthma Exacerbations In Vivo. Am J Respir Crit Care Med 2014; 190: 1373-1382.
- 58. Beale J, Jayaraman A, Jackson DJ, Macintyre JD, Edwards MR, Walton RP, Zhu J, Ching YM, Shamji B, Edwards M, Westwick J, Cousins DJ, Hwang YY, McKenzie A, Johnston SL, Bartlett NW. Rhinovirus-induced IL-25 in asthma exacerbation drives type 2 immunity and allergic pulmonary inflammation. *Sci Transl Med* 2014; 6: 256ra134.
- 59. Klein Wolterink RG, Kleinjan A, van Nimwegen M, Bergen I, de Bruijn M, Levani Y, Hendriks RW. Pulmonary innate lymphoid cells are major producers of IL-5 and IL-13 in murine models of allergic asthma. *Eur J Immunol* 2012; 42: 1106-1116.
- 60. Barlow JL, Bellosi A, Hardman CS, Drynan LF, Wong SH, Cruickshank JP, McKenzie AN. Innate IL-13-producing nuocytes arise during allergic lung inflammation and contribute to airways hyperreactivity. *J Allergy Clin Immunol* 2012; 129: 191-198.e191-194.
- 61. Halim TY, Krauss RH, Sun AC, Takei F. Lung natural helper cells are a critical source of Th2 cell-type cytokines in protease allergen-induced airway inflammation. *Immunity* 2012; 36: 451-463.
- 62. Kim HY, Lee HJ, Chang YJ, Pichavant M, Shore SA, Fitzgerald KA, Iwakura Y, Israel E, Bolger K, Faul J, DeKruyff RH, Umetsu DT. Interleukin-17-producing innate lymphoid cells and the NLRP3 inflammasome facilitate obesity-associated airway hyperreactivity. *Nat Med* 2014; 20: 54-61.
- 63. Gordon ED, Simpson LJ, Rios CL, Ringel L, Lachowicz-Scroggins ME, Peters MC, Wesolowska-Andersen A, Gonzalez JR, MacLeod HJ, Christian LS, Yuan S,





© Copyright 2016, EARIP Consortium GA- 602077

Version: Final

- Barry L, Woodruff PG, Ansel KM, Nocka K, Seibold MA, Fahy JV. Alternative splicing of interleukin-33 and type 2 inflammation in asthma. *Proc Natl Acad Sci U S A* 2016; 113: 8765-8770.
- 64. Dick S, Friend A, Dynes K, AlKandari F, Doust E, Cowie H, Ayres JG, Turner SW. A systematic review of associations between environmental exposures and development of asthma in children aged up to 9 years. *BMJ open* 2014; 4: e006554.
- 65. Brauer M, Hoek G, Smit HA, de Jongste JC, Gerritsen J, Postma DS, Kerkhof M, Brunekreef B. Air pollution and development of asthma, allergy and infections in a birth cohort. *Eur Respir J* 2007; 29: 879-888.
- 66. Morgenstern V, Zutavern A, Cyrys J, Brockow I, Koletzko S, Krämer U, Behrendt H, Herbarth O, von Berg A, Bauer CP, Wichmann HE, Heinrich J, Group GS, Group LS. Atopic diseases, allergic sensitization, and exposure to trafficrelated air pollution in children. *Am J Respir Crit Care Med* 2008; 177: 1331-1337.
- 67. Nordling E, Berglind N, Melén E, Emenius G, Hallberg J, Nyberg F, Pershagen G, Svartengren M, Wickman M, Bellander T. Traffic-related air pollution and childhood respiratory symptoms, function and allergies. *Epidemiology* 2008; 19: 401-408.
- 68. Gauderman WJ, Avol E, Gilliland F, Vora H, Thomas D, Berhane K, McConnell R, Kuenzli N, Lurmann F, Rappaport E, Margolis H, Bates D, Peters J. The effect of air pollution on lung development from 10 to 18 years of age. *N Engl J Med* 2004; 351: 1057-1067.
- 69. McConnell R, Berhane K, Gilliland F, London SJ, Islam T, Gauderman WJ, Avol E, Margolis HG, Peters JM. Asthma in exercising children exposed to ozone: a cohort study. *Lancet* 2002; 359: 386-391.
- 70. Gehring U, Wijga AH, Brauer M, Fischer P, de Jongste JC, Kerkhof M, Oldenwening M, Smit HA, Brunekreef B. Traffic-related air pollution and the development of asthma and allergies during the first 8 years of life. *Am J Respir Crit Care Med* 2010; 181: 596-603.
- 71. Gehring U, Wijga AH, Hoek G, Bellander T, Berdel D, Bruske I, Fuertes E, Gruzieva O, Heinrich J, Hoffmann B, de Jongste JC, Klumper C, Koppelman GH, Korek M, Kramer U, Maier D, Melen E, Pershagen G, Postma DS, Standl M, von Berg A, Anto JM, Bousquet J, Keil T, Smit HA, Brunekreef B. Exposure to air pollution and development of asthma and rhinoconjunctivitis throughout childhood and adolescence: a population-based birth cohort study. *Lancet Respir Med* 2015; 3: 933-942.
- 72. Nishimura KK, Galanter JM, Roth LA, Oh SS, Thakur N, Nguyen EA, Thyne S, Farber HJ, Serebrisky D, Kumar R, Brigino-Buenaventura E, Davis A, LeNoir MA, Meade K, Rodriguez-Cintron W, Avila PC, Borrell LN, Bibbins-Domingo K, Rodriguez-Santana JR, Sen S, Lurmann F, Balmes JR, Burchard EG. Early-life air pollution and asthma risk in minority children. The GALA II and SAGE II studies. Am J Respir Crit Care Med 2013; 188: 309-318.
- 73. Young MT, Sandler DP, DeRoo LA, Vedal S, Kaufman JD, London SJ. Ambient air pollution exposure and incident adult asthma in a nationwide cohort of U.S. women. *Am J Respir Crit Care Med* 2014; 190: 914-921.
- 74. Oftedal B, Nystad W, Brunekreef B, Nafstad P. Long-term traffic-related exposures and asthma onset in schoolchildren in oslo, norway. *Environ Health Perspect* 2009; 117: 839-844.





Version: Final

75. Physicians. RCo. Every breath we take: the lifelong impact of air pollution. Report of a working party. London: RCP, 2016.; 2016.

- 76. Gordian ME, Haneuse S, Wakefield J. An investigation of the association between traffic exposure and the diagnosis of asthma in children. *J Expo Sci Environ Epidemiol* 2006; 16: 49-55.
- 77. Sienra-Monge JJ, Ramirez-Aguilar M, Moreno-Macias H, Reyes-Ruiz NI, Del Río-Navarro BE, Ruiz-Navarro MX, Hatch G, Crissman K, Slade R, Devlin RB, Romieu I. Antioxidant supplementation and nasal inflammatory responses among young asthmatics exposed to high levels of ozone. *Clin Exp Immunol* 2004; 138: 317-322.
- 78. Behndig AF, Blomberg A, Helleday R, Kelly FJ, Mudway IS. Augmentation of respiratory tract lining fluid ascorbate concentrations through supplementation with vitamin C. *Inhal Toxicol* 2009; 21: 250-258.
- 79. Clougherty JE, Kubzansky LD. A framework for examining social stress and susceptibility to air pollution in respiratory health. *Environ Health Perspect* 2009; 117: 1351-1358.
- 80. Clougherty JE, Levy JI, Kubzansky LD, Ryan PB, Suglia SF, Canner MJ, Wright RJ. Synergistic effects of traffic-related air pollution and exposure to violence on urban asthma etiology. *Environ Health Perspect* 2007; 115: 1140-1146.
- 81. Meng YY, Wilhelm M, Rull RP, English P, Nathan S, Ritz B. Are frequent asthma symptoms among low-income individuals related to heavy traffic near homes, vulnerabilities, or both? *Ann Epidemiol* 2008; 18: 343-350.
- 82. Chen E, Schreier HM, Strunk RC, Brauer M. Chronic traffic-related air pollution and stress interact to predict biologic and clinical outcomes in asthma. *Environ Health Perspect* 2008; 116: 970-975.
- 83. Salam MT, Lin PC, Avol EL, Gauderman WJ, Gilliland FD. Microsomal epoxide hydrolase, glutathione S-transferase P1, traffic and childhood asthma. *Thorax* 2007; 62: 1050-1057.
- 84. Melén E, Nyberg F, Lindgren CM, Berglind N, Zucchelli M, Nordling E, Hallberg J, Svartengren M, Morgenstern R, Kere J, Bellander T, Wickman M, Pershagen G. Interactions between glutathione S-transferase P1, tumor necrosis factor, and traffic-related air pollution for development of childhood allergic disease. *Environ Health Perspect* 2008; 116: 1077-1084.
- 85. Macpherson AJ, Uhr T. Compartmentalization of the mucosal immune responses to commensal intestinal bacteria. *Ann N Y Acad Sci* 2004; 1029: 36-43.
- 86. Sender R, Fuchs S, Milo R. Are We Really Vastly Outnumbered? Revisiting the Ratio of Bacterial to Host Cells in Humans. *Cell* 2016; 164: 337-340.
- 87. Marsland BJ, Gollwitzer ES. Host-microorganism interactions in lung diseases. *Nat Rev Immunol* 2014; 14: 827-835.
- 88. Peterson J, Garges S, Giovanni M, McInnes P, Wang L, Schloss JA, Bonazzi V, McEwen JE, Wetterstrand KA, Deal C, Baker CC, Di Francesco V, Howcroft TK, Karp RW, Lunsford RD, Wellington CR, Belachew T, Wright M, Giblin C, David H, Mills M, Salomon R, Mullins C, Akolkar B, Begg L, Davis C, Grandison L, Humble M, Khalsa J, Little AR, Peavy H, Pontzer C, Portnoy M, Sayre MH, Starke-Reed P, Zakhari S, Read J, Watson B, Guyer M. The NIH Human Microbiome Project. *Genome Res* 2009; 19: 2317-2323.
- 89. Cox MJ, Cookson WO, Moffatt MF. Sequencing the human microbiome in health and disease. *Hum Mol Genet* 2013; 22: R88-94.





GA- 602077

Version: Final

- 90. Dickson RP, Erb-Downward JR, Huffnagle GB. The role of the bacterial microbiome in lung disease. *Expert Rev Respir Med* 2013; 7: 245-257.
- 91. Hilty M, Burke C, Pedro H, Cardenas P, Bush A, Bossley C, Davies J, Ervine A, Poulter L, Pachter L, Moffatt MF, Cookson WO. Disordered microbial communities in asthmatic airways. *PLoS One* 2010; 5: e8578.
- 92. Erb-Downward JR, Thompson DL, Han MK, Freeman CM, McCloskey L, Schmidt LA, Young VB, Toews GB, Curtis JL, Sundaram B, Martinez FJ, Huffnagle GB. Analysis of the lung microbiome in the "healthy" smoker and in COPD. *PLoS One* 2011; 6: e16384.
- 93. Eder W, Ege MJ, von Mutius E. The asthma epidemic. *N Engl J Med* 2006; 355: 2226-2235.
- 94. Ege MJ, Mayer M, Normand AC, Genuneit J, Cookson WO, Braun-Fahrlander C, Heederik D, Piarroux R, von Mutius E. Exposure to environmental microorganisms and childhood asthma. *N Engl J Med* 2011; 364: 701-709.
- 95. Stein MM, Hrusch CL, Gozdz J, Igartua C, Pivniouk V, Murray SE, Ledford JG, Marques dos Santos M, Anderson RL, Metwali N, Neilson JW, Maier RM, Gilbert JA, Holbreich M, Thorne PS, Martinez FD, von Mutius E, Vercelli D, Ober C, Sperling Al. Innate Immunity and Asthma Risk in Amish and Hutterite Farm Children. N Engl J Med 2016; 375: 411-421.
- 96. Marri PR, Stern DA, Wright AL, Billheimer D, Martinez FD. Asthma-associated differences in microbial composition of induced sputum. *J Allergy Clin Immunol* 2012; 131: 346-352 e341-343.
- 97. Horvat JC, Starkey MR, Kim RY, Phipps S, Gibson PG, Beagley KW, Foster PS, Hansbro PM. Early-life chlamydial lung infection enhances allergic airways disease through age-dependent differences in immunopathology. *J Allergy Clin Immunol* 2010; 125: 617-625.
- 98. Gollwitzer ES, Saglani S, Trompette A, Yadava K, Sherburn R, McCoy KD, Nicod LP, Lloyd CM, Marsland BJ. Lung microbiota promotes tolerance to allergens in neonates via PD-L1. *Nat Med* 2014; 20: 642-647.
- 99. Herbst T, Sichelstiel A, Schar C, Yadava K, Burki K, Cahenzli J, McCoy K, Marsland BJ, Harris NL. Dysregulation of allergic airway inflammation in the absence of microbial colonization. *Am J Respir Crit Care Med* 2011; 184: 198-205.
- 100. Linehan MF, Nurmatov U, Frank TL, Niven RM, Baxter DN, Sheikh A. Does BCG vaccination protect against childhood asthma? Final results from the Manchester Community Asthma Study retrospective cohort study and updated systematic review and meta-analysis. *J Allergy Clin Immunol* 2014; 133: 688-695 e614.
- 101. Shirakawa T, Enomoto T, Shimazu S, Hopkin JM. The inverse association between tuberculin responses and atopic disorder. *Science* 1997; 275: 77-79.
- 102. Flohr C, Nagel G, Weinmayr G, Kleiner A, Williams HC, Ait-Khaled N, Strachan DP, Group IPTS. Tuberculosis, bacillus Calmette-Guerin vaccination, and allergic disease: findings from the International Study of Asthma and Allergies in Childhood Phase Two. Pediatr Allergy Immunol 2012; 23: 324-331.
- 103. Shirtcliffe PM, Easthope SE, Weatherall M, Beasley R. Effect of repeated intradermal injections of heat-inactivated Mycobacterium bovis bacillus Calmette-Guerin in adult asthma. *Clin Exp Allergy* 2004; 34: 207-212.





Version: Final

104. Riffo-Vasquez Y, Spina D, Page C, Tormay P, Singh M, Henderson B, Coates A. Effect of Mycobacterium tuberculosis chaperonins on bronchial eosinophilia and hyper-responsiveness in a murine model of allergic inflammation. *Clin Exp Allergy* 2004; 34: 712-719.

- 105. Samary Cdos S, Antunes MA, Silva JD, Silva AL, Araujo CC, Bakker-Abreu I, Diaz BL, Fernezlian S, Parra ER, Capelozzi VL, Silva PL, Lapa e Silva JR, Rocco PR. Impact of Bacillus Calmette-Guerin Moreau vaccine on lung remodeling in experimental asthma. *Respiratory physiology & neurobiology* 2013; 189: 614-623.
- 106. Marsland BJ, Trompette A, Gollwitzer ES. The Gut-Lung Axis in Respiratory Disease. *Ann Am Thorac Soc* 2015; 12 Suppl 2: S150-156.
- 107. Braun-Fahrlander C, Riedler J, Herz U, Eder W, Waser M, Grize L, Maisch S, Carr D, Gerlach F, Bufe A, Lauener RP, Schierl R, Renz H, Nowak D, von Mutius E. Environmental exposure to endotoxin and its relation to asthma in school-age children. N Engl J Med 2002; 347: 869-877.
- 108. Schuijs MJ, Willart MA, Vergote K, Gras D, Deswarte K, Ege MJ, Madeira FB, Beyaert R, van Loo G, Bracher F, von Mutius E, Chanez P, Lambrecht BN, Hammad H. Farm dust and endotoxin protect against allergy through A20 induction in lung epithelial cells. *Science* 2015; 349: 1106-1110.
- 109. von Mutius E. Maternal farm exposure/ingestion of unpasteurized cow's milk and allergic disease. *Curr Opin Gastroenterol* 2012; 28: 570-576.
- 110. Huang YJ, Nelson CE, Brodie EL, Desantis TZ, Baek MS, Liu J, Woyke T, Allgaier M, Bristow J, Wiener-Kronish JP, Sutherland ER, King TS, Icitovic N, Martin RJ, Calhoun WJ, Castro M, Denlinger LC, Dimango E, Kraft M, Peters SP, Wasserman SI, Wechsler ME, Boushey HA, Lynch SV. Airway microbiota and bronchial hyperresponsiveness in patients with suboptimally controlled asthma. *J Allergy Clin Immunol* 2011; 127: 372-381 e371-373.
- 111. Cardenas PA, Cooper PJ, Cox MJ, Chico M, Arias C, Moffatt MF, Cookson WO. Upper airways microbiota in antibiotic-naive wheezing and healthy infants from the tropics of rural Ecuador. *PLoS One* 2012; 7: e46803.
- 112. Green BJ, Wiriyachaiporn S, Grainge C, Rogers GB, Kehagia V, Lau L, Carroll MP, Bruce KD, Howarth PH. Potentially pathogenic airway bacteria and neutrophilic inflammation in treatment resistant severe asthma. *PLoS One* 2014; 9: e100645.
- 113. Simpson JL, Daly J, Baines KJ, Yang IA, Upham JW, Reynolds PN, Hodge S, James AL, Hugenholtz P, Willner D, Gibson PG. Airway dysbiosis: Haemophilus influenzae and Tropheryma in poorly controlled asthma. *Eur Respir J* 2015; 47: 792-800.
- 114. Huang YJ, Nariya S, Harris JM, Lynch SV, Choy DF, Arron JR, Boushey H. The airway microbiome in patients with severe asthma: Associations with disease features and severity. *J Allergy Clin Immunol* 2015; 136: 874-884.
- 115. Bisgaard H, Hermansen MN, Buchvald F, Loland L, Halkjaer LB, Bonnelykke K, Brasholt M, Heltberg A, Vissing NH, Thorsen SV, Stage M, Pipper CB. Childhood asthma after bacterial colonization of the airway in neonates. *N Engl J Med* 2007; 357: 1487-1495.
- 116. Kraft M. The role of bacterial infections in asthma. *Clin Chest Med* 2000; 21: 301-313.
- 117. Teo SM, Mok D, Pham K, Kusel M, Serralha M, Troy N, Holt BJ, Hales BJ, Walker ML, Hollams E, Bochkov YA, Grindle K, Johnston SL, Gern JE, Sly PD, Holt PG, Holt KE, Inouye M. The infant nasopharyngeal microbiome impacts





GA-602077

Version: Final

- severity of lower respiratory infection and risk of asthma development. *Cell Host Microbe*; 17: 704-715.
- 118. Bisgaard H, Hermansen MN, Bonnelykke K, Stokholm J, Baty F, Skytt NL, Aniscenko J, Kebadze T, Johnston SL. Association of bacteria and viruses with wheezy episodes in young children: prospective birth cohort study. *BMJ* 2010; 341: c4978.
- 119. Stokholm J, Sevelsted A, Bonnelykke K, Bisgaard H. Maternal propensity for infections and risk of childhood asthma: a registry-based cohort study. *Lancet Respir Med* 2014; 2: 631-637.
- 120. Itkin IH, Menzel ML. The use of macrolide antibiotic substances in the treatment of asthma. *J Allergy* 1970; 45: 146-162.
- 121. Johnston SL. Macrolide antibiotics and asthma treatment. *J Allergy Clin Immunol* 2006; 117: 1233-1236.
- 122. Schwerk N, Brinkmann F, Soudah B, Kabesch M, Hansen G. Wheeze in preschool age is associated with pulmonary bacterial infection and resolves after antibiotic therapy. *PLoS One* 2011; 6: e27913.
- 123. Donnelly D, Critchlow A, Everard ML. Outcomes in children treated for persistent bacterial bronchitis. *Thorax* 2007; 62: 80-84.
- 124. Organisation WH. Obesity and overweight Fact sheet N°311. http://www.who.int/mediacentre/factsheets/fs311/en/; Updated January 2015. p. 8-11.
- 125. Ford ES. The epidemiology of obesity and asthma. *J Allergy Clin Immunol* 2005; 115: 897-909; quiz 910.
- 126. Wenzel SE. Severe asthma and its phenotype. *J Asthma* 2008; 45 Suppl 1: 32-36.
- 127. Sideleva O, Dixon AE. The many faces of asthma in obesity. *J Cell Biochem* 2014; 115: 421-426.
- 128. Litonjua AA, Gold DR. Asthma and obesity: common early-life influences in the inception of disease. *J Allergy Clin Immunol* 2008; 121: 1075-1084; quiz 1085-1076.
- 129. Brehm JM, Schuemann B, Fuhlbrigge AL, Hollis BW, Strunk RC, Zeiger RS, Weiss ST, Litonjua AA, Childhood Asthma Management Program Research G. Serum vitamin D levels and severe asthma exacerbations in the Childhood Asthma Management Program study. J Allergy Clin Immunol 2010; 126: 52-58 e55.
- 130. Goleva E, Searing DA, Jackson LP, Richers BN, Leung DY. Steroid requirements and immune associations with vitamin D are stronger in children than adults with asthma. *J Allergy Clin Immunol* 2012; 129: 1243-1251.
- 131. Martineau AR, Cates CJ, Urashima M, Jensen M, Griffiths AP, Nurmatov U, Sheikh A, Griffiths CJ. Vitamin D for the management of asthma. *Cochrane Database Syst Rev* 2016; 9: CD011511.
- 132. Litonjua AA, Carey VJ, Laranjo N, Harshfield BJ, McElrath TF, O'Connor GT, Sandel M, Iverson RE, Jr., Lee-Paritz A, Strunk RC, Bacharier LB, Macones GA, Zeiger RS, Schatz M, Hollis BW, Hornsby E, Hawrylowicz C, Wu AC, Weiss ST. Effect of Prenatal Supplementation With Vitamin D on Asthma or Recurrent Wheezing in Offspring by Age 3 Years: The VDAART Randomized Clinical Trial. *Jama* 2016; 315: 362-370.
- 133. Chawes BL, Bonnelykke K, Stokholm J, Vissing NH, Bjarnadottir E, Schoos AM, Wolsk HM, Pedersen TM, Vinding RK, Thorsteinsdottir S, Arianto L, Hallas HW, Heickendorff L, Brix S, Rasmussen MA, Bisgaard H. Effect of Vitamin D3





Version: Final

Supplementation During Pregnancy on Risk of Persistent Wheeze in the Offspring: A Randomized Clinical Trial. *Jama* 2016; 315: 353-361.

- 134. Bisgaard H, Stokholm J, Chawes BL, Vissing NH, Bjarnadottir E, Schoos AM, Wolsk HM, Pedersen TM, Vinding RK, Thorsteinsdottir S, Folsgaard NV, Fink NR, Thorsen J, Pedersen AG, Waage J, Rasmussen MA, Stark KD, Olsen SF, Bonnelykke K. Fish Oil-Derived Fatty Acids in Pregnancy and Wheeze and Asthma in Offspring. N Engl J Med 2016; 375: 2530-2539.
- 135. Vignali DA, Collison LW, Workman CJ. How regulatory T cells work. *Nat Rev Immunol* 2008; 8: 523-532.
- 136. Lloyd CM, Hawrylowicz CM. Regulatory T cells in asthma. *Immunity* 2009; 31: 438-449.
- 137. Holt PG, Strickland DH, Wikstrom ME, Jahnsen FL. Regulation of immunological homeostasis in the respiratory tract. *Nat Rev Immunol* 2008; 8: 142-152.
- 138. Nelson HS, Szefler SJ, Jacobs J, Huss K, Shapiro G, Sternberg AL. The relationships among environmental allergen sensitization, allergen exposure, pulmonary function, and bronchial hyperresponsiveness in the Childhood Asthma Management Program. *J Allergy Clin Immunol* 1999; 104: 775-785.
- 139. Chatila TA, Blaeser F, Ho N, Lederman HM, Voulgaropoulos C, Helms C, Bowcock AM. JM2, encoding a fork head-related protein, is mutated in X-linked autoimmunity-allergic disregulation syndrome. *J Clin Invest* 2000; 106: R75-81.
- 140. Akdis M, Verhagen J, Taylor A, Karamloo F, Karagiannidis C, Crameri R, Thunberg S, Deniz G, Valenta R, Fiebig H, Kegel C, Disch R, Schmidt-Weber CB, Blaser K, Akdis CA. Immune responses in healthy and allergic individuals are characterized by a fine balance between allergen-specific T regulatory 1 and T helper 2 cells. *J Exp Med* 2004; 199: 1567-1575.
- 141. Chambers ES, Hawrylowicz CM. The impact of vitamin D on regulatory T cells. Curr Allergy Asthma Rep 2011; 11: 29-36.
- 142. Nadeau K, McDonald-Hyman C, Noth EM, Pratt B, Hammond SK, Balmes J, Tager I. Ambient air pollution impairs regulatory T-cell function in asthma. *J Allergy Clin Immunol* 2010; 126: 845-852 e810.
- 143. Hawrylowicz CM. Regulatory T cells and IL-10 in allergic inflammation. *J Exp Med* 2005; 202: 1459-1463.
- 144. Vermaelen K, Pauwels R. Pulmonary dendritic cells. *Am J Respir Crit Care Med* 2005; 172: 530-551.
- 145. Lambrecht BN, Hammad H. Biology of lung dendritic cells at the origin of asthma. *Immunity* 2009; 31: 412-424.
- 146. Contoli M, Ito K, Padovani A, Poletti D, Marku B, Edwards MR, Stanciu LA, Gnesini G, Pastore A, Spanevello A, Morelli P, Johnston SL, Caramori G, Papi A. Th2 cytokines impair innate immune responses to rhinovirus in respiratory epithelial cells. *Allergy* 2015; 70: 910-920.
- 147. Du Toit G, Roberts G, Sayre PH, Bahnson HT, Radulovic S, Santos AF, Brough HA, Phippard D, Basting M, Feeney M, Turcanu V, Sever ML, Gomez Lorenzo M, Plaut M, Lack G, Team LS. Randomized trial of peanut consumption in infants at risk for peanut allergy. *N Engl J Med* 2015; 372: 803-813.
- 148. Calam R, Gregg L, Simpson A, Simpson B, Woodcock A, Custovic A. Behavior problems antecede the development of wheeze in childhood: a birth cohort study. *Am J Respir Crit Care Med* 2005; 171: 323-327.





© Copyright 2016, EARIP Consortium GA- 602077 Version: Final

- 149. Ader R, Cohen N, Felten D. Psychoneuroimmunology: interactions between the nervous system and the immune system. *Lancet* 1995; 345: 99-103.
- 150. Forsythe P, Ebeling C, Gordon JR, Befus AD, Vliagoftis H. Opposing effects of short- and long-term stress on airway inflammation. *Am J Respir Crit Care Med* 2004; 169: 220-226.
- 151. Wright RJ, Cohen RT, Cohen S. The impact of stress on the development and expression of atopy. *Current opinion in allergy and clinical immunology* 2005; 5: 23-29.
- 152. Kreiner-Moller E, Strachan DP, Linneberg A, Husemoen LL, Bisgaard H, Bonnelykke K. 17q21 gene variation is not associated with asthma in adulthood. *Allergy* 2014; 70: 107-114.
- 153. Sarnowski C, Sugier PE, Granell R, Jarvis D, Dizier MH, Ege M, Imboden M, Laprise C, Khusnutdinova EK, Freidin MB, Cookson WO, Moffatt M, Lathrop M, Siroux V, Ogorodova LM, Karunas AS, James A, Probst-Hensch NM, von Mutius E, Pin I, Kogevinas M, Henderson AJ, Demenais F, Bouzigon E. Identification of a new locus at 16q12 associated with time to asthma onset. *J Allergy Clin Immunol* 2016.
- 154. Gibbons D, Fleming P, Virasami A, Michel ML, Sebire NJ, Costeloe K, Carr R, Klein N, Hayday A. Interleukin-8 (CXCL8) production is a signatory T cell effector function of human newborn infants. *Nat Med* 2014; 20: 1206-1210.
- 155. Smith CL, Dickinson P, Forster T, Craigon M, Ross A, Khondoker MR, France R, Ivens A, Lynn DJ, Orme J, Jackson A, Lacaze P, Flanagan KL, Stenson BJ, Ghazal P. Identification of a human neonatal immune-metabolic network associated with bacterial infection. *Nat Commun* 2014; 5: 4649.
- 156. Cullinan P. Occupational asthma: risk factors, diagnosis and preventive measures. *Expert Rev Clin Immunol* 2005; 1: 123-132.
- 157. Edwards MR, Regamey N, Vareille M, Kieninger E, Gupta A, Shoemark A, Saglani S, Sykes A, Macintyre J, Davies J, Bossley C, Bush A, Johnston SL. Impaired innate interferon induction in severe therapy resistant atopic asthmatic children. *Mucosal Immunol* 2012; 6: 797-806.
- 158. Spann KM, Baturcam E, Schagen J, Jones C, Straub CP, Preston FM, Chen L, Phipps S, Sly PD, Fantino E. Viral and host factors determine innate immune responses in airway epithelial cells from children with wheeze and atopy. *Thorax* 2014; 69: 918-925.
- 159. Bonnelykke K, Sleiman P, Nielsen K, Kreiner-Moller E, Mercader JM, Belgrave D, den Dekker HT, Husby A, Sevelsted A, Faura-Tellez G, Mortensen LJ, Paternoster L, Flaaten R, Molgaard A, Smart DE, Thomsen PF, Rasmussen MA, Bonas-Guarch S, Holst C, Nohr EA, Yadav R, March ME, Blicher T, Lackie PM, Jaddoe VW, Simpson A, Holloway JW, Duijts L, Custovic A, Davies DE, Torrents D, Gupta R, Hollegaard MV, Hougaard DM, Hakonarson H, Bisgaard H. A genome-wide association study identifies CDHR3 as a susceptibility locus for early childhood asthma with severe exacerbations. *Nat Genet* 2013; 46: 51-55.
- 160. Stegle O, Teichmann SA, Marioni JC. Computational and analytical challenges in single-cell transcriptomics. *Nat Rev Genet* 2015; 16: 133-145.
- 161. Greer RM, Miller JD, Okoh VO, Halloran BA, Prince LS. Epithelial-mesenchymal co-culture model for studying alveolar morphogenesis. *Organogenesis* 2014: 1-10.
- 162. Edwards J, Belvisi M, Dahlen SE, Holgate S, Holmes A. Human tissue models for a human disease: what are the barriers? *Thorax* 2015; 70: 695-697.





Version: Final

163. Saglani S, Lui S, Ullmann N, Campbell GA, Sherburn RT, Mathie SA, Denney L, Bossley CJ, Oates T, Walker SA, Bush A, Lloyd CM. IL-33 promotes airway remodeling in pediatric patients with severe steroid-resistant asthma. *J Allergy Clin Immunol* 2013; 132: 676-685 e613.

- 164. Simoes EA, Groothuis JR, Carbonell-Estrany X, Rieger CH, Mitchell I, Fredrick LM, Kimpen JL. Palivizumab prophylaxis, respiratory syncytial virus, and subsequent recurrent wheezing. *J Pediatr* 2007; 151: 34-42, 42 e31.
- 165. Simoes EA, Carbonell-Estrany X, Rieger CH, Mitchell I, Fredrick L, Groothuis JR. The effect of respiratory syncytial virus on subsequent recurrent wheezing in atopic and nonatopic children. *J Allergy Clin Immunol* 2010; 126: 256-262.
- 166. Blanken MO, Rovers MM, Molenaar JM, Winkler-Seinstra PL, Meijer A, Kimpen JL, Bont L. Respiratory syncytial virus and recurrent wheeze in healthy preterm infants. *N Engl J Med* 2013; 368: 1791-1799.
- 167. DeVincenzo JP, Whitley RJ, Mackman RL, Scaglioni-Weinlich C, Harrison L, Farrell E, McBride S, Lambkin-Williams R, Jordan R, Xin Y, Ramanathan S, O'Riordan T, Lewis SA, Li X, Toback SL, Lin SL, Chien JW. Oral GS-5806 activity in a respiratory syncytial virus challenge study. *N Engl J Med* 2014; 371: 711-722.
- 168. DeVincenzo JP, McClure MW, Symons JA, Fathi H, Westland C, Chanda S, Lambkin-Williams R, Smith P, Zhang Q, Beigelman L, Blatt LM, Fry J. Activity of Oral ALS-008176 in a Respiratory Syncytial Virus Challenge Study. N Engl J Med 2015; 373: 2048-2058.
- 169. Zhang Q, Cox M, Liang Z, Brinkmann F, Cardenas PA, Duff R, Bhavsar P, Cookson W, Moffatt M, Chung KF. Airway Microbiota in Severe Asthma and Relationship to Asthma Severity and Phenotypes. *PLoS One* 2016; 11: e0152724.
- 170. Wood LG, Simpson JL, Hansbro PM, Gibson PG. Potentially pathogenic bacteria cultured from the sputum of stable asthmatics are associated with increased 8-isoprostane and airway neutrophilia. *Free Radic Res* 2010; 44: 146-154.
- 171. Zhang Q, Illing R, Hui CK, Downey K, Carr D, Stearn M, Alshafi K, Menzies-Gow A, Zhong N, Fan Chung K. Bacteria in sputum of stable severe asthma and increased airway wall thickness. *Respiratory research* 2012; 13: 35.
- 172. Bachert C, van Steen K, Zhang N, Holtappels G, Cattaert T, Maus B, Buhl R, Taube C, Korn S, Kowalski M, Bousquet J, Howarth P. Specific IgE against Staphylococcus aureus enterotoxins: an independent risk factor for asthma. *J Allergy Clin Immunol* 2012; 130: 376-381 e378.
- 173. Fritscher LG, Marras TK, Bradi AC, Fritscher CC, Balter MS, Chapman KR. Nontuberculous mycobacterial infection as a cause of difficult-to-control asthma: a case-control study. *Chest* 2011; 139: 23-27.
- 174. Hojo M, Iikura M, Hirano S, Sugiyama H, Kobayashi N, Kudo K. Increased risk of nontuberculous mycobacterial infection in asthmatic patients using long-term inhaled corticosteroid therapy. *Respirology* 2012; 17: 185-190.
- 175. Johnston SL, Martin RJ. Chlamydophila pneumoniae and Mycoplasma pneumoniae: a role in asthma pathogenesis? *Am J Respir Crit Care Med* 2005; 172: 1078-1089.
- 176. Wong EH, Porter JD, Edwards MR, Johnston SL. The role of macrolides in asthma: current evidence and future directions. *Lancet Respir Med* 2014; 2: 657-670.





Version: Final

© Copyright 2016,

1

2 3 4

5

6

7

8

9

10

11

12

13

14

15

16

17

18

19

20

21

22 23

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45

46

47

48

49

50

51

52

53

54

60

- 177. Klemets P, Lyytikainen O, Ruutu P, Ollgren J, Kaijalainen T, Leinonen M, Nuorti JP. Risk of invasive pneumococcal infections among working age adults with asthma. Thorax 2010; 65: 698-702.
- 178. Talbot TR, Hartert TV, Mitchel E, Halasa NB, Arbogast PG, Poehling KA, Schaffner W, Craig AS, Griffin MR. Asthma as a risk factor for invasive pneumococcal disease. N Engl J Med 2005; 352: 2082-2090.
- 179. Johnston SL. IFN Deficiency in Asthma Attacks. Is Restoring Toll-like Receptor-7 Expression a New Treatment Approach in Severe Asthma? Am J Respir Crit Care Med 2016; 194: 1-3.
- 180. Rupani H, Martinez-Nunez RT, Dennison P, Lau LC, Jayasekera N, Havelock T, Francisco-Garcia AS, Grainge C, Howarth PH, Sanchez-Elsner T. Toll-like Receptor 7 Is Reduced in Severe Asthma and Linked to an Altered MicroRNA Profile. Am J Respir Crit Care Med 2016; 194: 26-37.
- 181. Shikhagaie MM, Andersson CK, Mori M, Kortekaas Krohn I, Bergqvist A, Dahl R, Ekblad E, Hoffmann HJ, Bjermer L, Erjefalt JS. Mapping of TLR5 and TLR7 in central and distal human airways and identification of reduced TLR expression in severe asthma. Clin Exp Allergy 2014; 44: 184-196.
- 182. Mancuso G, Midiri A, Biondo C, Beninati C, Zummo S, Galbo R, Tomasello F, Gambuzza M, Macri G, Ruggeri A, Leanderson T, Teti G. Type I IFN signaling is crucial for host resistance against different species of pathogenic bacteria. J Immunol 2007; 178: 3126-3133.
- 183. Rothfuchs AG, Trumstedt C, Mattei F, Schiavoni G, Hidmark A, Wigzell H, Rottenberg ME. STAT1 regulates IFN-alpha beta- and IFN-gamma-dependent control of infection with Chlamydia pneumoniae by nonhemopoietic cells. J Immunol 2006; 176: 6982-6990.
- 184. Wark PA, Johnston SL, Bucchieri F, Powell R, Puddicombe S, Laza-Stanca V, Holgate ST, Davies DE, Asthmatic bronchial epithelial cells have a deficient innate immune response to infection with rhinovirus. J Exp Med 2005; 201: 937-947.
- 185. Contoli M, Message SD, Laza-Stanca V, Edwards MR, Wark PA, Bartlett NW, Kebadze T, Mallia P, Stanciu LA, Parker HL, Slater L, Lewis-Antes A, Kon OM, Holgate ST, Davies DE, Kotenko SV, Papi A, Johnston SL. Role of deficient type III interferon-lambda production in asthma exacerbations. Nat Med 2006; 12: 1023-1026.
- 186. Message SD, Laza-Stanca V, Mallia P, Parker HL, Zhu J, Kebadze T, Contoli M, Sanderson G, Kon OM, Papi A, Jeffery PK, Stanciu LA, Johnston SL. Rhinovirus-induced lower respiratory illness is increased in asthma and related to virus load and Th1/2 cytokine and IL-10 production. Proc Natl Acad Sci U S A 2008; 105: 13562-13567.
- 187. Baraldo S, Contoli M, Bazzan E, Turato G, Padovani A, Marku B, Calabrese F, Caramori G, Ballarin A, Snijders D, Barbato A, Saetta M, Papi A. Deficient antiviral immune responses in childhood: distinct roles of atopy and asthma. J Allergy Clin Immunol 2012; 130: 1307-1314.
- 188. Edwards MR, Regamey N, Vareille M, Kieninger E, Gupta A, Shoemark A, Saglani S, Sykes A, Macintyre J, Davies J, Bossley C, Bush A, Johnston SL. Impaired innate interferon induction in severe therapy resistant atopic asthmatic children. Mucosal Immunol 2013; 6: 797-806.
- 189. Plummeridge MJ, Armstrong L, Birchall MA, Millar AB. Reduced production of interleukin 12 by interferon gamma primed alveolar macrophages from atopic asthmatic subjects. Thorax 2000; 55: 842-847.





Version: Final

190. Ho LP, Davis M, Denison A, Wood FT, Greening AP. Reduced interleukin-18 levels in BAL specimens from patients with asthma compared to patients with sarcoidosis and healthy control subjects. *Chest* 2002; 121: 1421-1426.

- 191. Liang Z, Zhang Q, Thomas CM, Chana KK, Gibeon D, Barnes PJ, Chung KF, Bhavsar PK, Donnelly LE. Impaired macrophage phagocytosis of bacteria in severe asthma. *Respiratory research* 2014; 15: 72.
- 192. Yadava K, Pattaroni C, Sichelstiel AK, Trompette A, Gollwitzer ES, Salami O, von Garnier C, Nicod LP, Marsland BJ. Microbiota Promotes Chronic Pulmonary Inflammation by Enhancing IL-17A and Autoantibodies. *Am J Respir Crit Care Med* 2016; 193: 975-987.
- 193. Lawley TD, Clare S, Walker AW, Stares MD, Connor TR, Raisen C, Goulding D, Rad R, Schreiber F, Brandt C, Deakin LJ, Pickard DJ, Duncan SH, Flint HJ, Clark TG, Parkhill J, Dougan G. Targeted restoration of the intestinal microbiota with a simple, defined bacteriotherapy resolves relapsing Clostridium difficile disease in mice. *PLoS Pathog* 2012; 8: e1002995.
- 194. Pham TA, Lawley TD. Emerging insights on intestinal dysbiosis during bacterial infections. *Current opinion in microbiology* 2014; 17: 67-74.
- 195. Trompette A, Gollwitzer ES, Yadava K, Sichelstiel AK, Sprenger N, Ngom-Bru C, Blanchard C, Junt T, Nicod LP, Harris NL, Marsland BJ. Gut microbiota metabolism of dietary fiber influences allergic airway disease and hematopoiesis. *Nat Med* 2014; 20: 159-166.
- 196. Denning DW, Pashley C, Hartl D, Wardlaw A, Godet C, Del Giacco S, Delhaes L, Sergejeva S. Fungal allergy in asthma-state of the art and research needs. *Clinical and translational allergy* 2014; 4: 14.
- 197. O'Byrne PM, Pedersen S, Lamm CJ, Tan WC, Busse WW, Group SI. Severe exacerbations and decline in lung function in asthma. *Am J Respir Crit Care Med* 2009; 179: 19-24.
- 198. Matsunaga K, Hirano T, Oka A, Tanaka A, Kanai K, Kikuchi T, Hayata A, Akamatsu H, Akamatsu K, Koh Y, Nakanishi M, Minakata Y, Yamamoto N. Progression of Irreversible Airflow Limitation in Asthma: Correlation with Severe Exacerbations. *J Allergy Clin Immunol Pract* 2015; 3: 759-764 e751.
- 199. Bai TR, Vonk JM, Postma DS, Boezen HM. Severe exacerbations predict excess lung function decline in asthma. *Eur Respir J* 2007; 30: 452-456.
- 200. Rennard SI, Farmer SG. Exacerbations and progression of disease in asthma and chronic obstructive pulmonary disease. *Proc Am Thorac Soc* 2004; 1: 88-92.
- 201. Carlsten C, Blomberg A, Pui M, Sandstrom T, Wong SW, Alexis N, Hirota J. Diesel exhaust augments allergen-induced lower airway inflammation in allergic individuals: a controlled human exposure study. *Thorax* 2016; 71: 35-44.
- 202. Brunst KJ, Ryan PH, Brokamp C, Bernstein D, Reponen T, Lockey J, Khurana Hershey GK, Levin L, Grinshpun SA, LeMasters G. Timing and Duration of Traffic-related Air Pollution Exposure and the Risk for Childhood Wheeze and Asthma. *Am J Respir Crit Care Med* 2015; 192: 421-427.
- 203. Schultz ES, Gruzieva O, Bellander T, Bottai M, Hallberg J, Kull I, Svartengren M, Melen E, Pershagen G. Traffic-related air pollution and lung function in children at 8 years of age: a birth cohort study. *Am J Respir Crit Care Med* 2012; 186: 1286-1291.
- 204. McCreanor J, Cullinan P, Nieuwenhuijsen MJ, Stewart-Evans J, Malliarou E, Jarup L, Harrington R, Svartengren M, Han IK, Ohman-Strickland P, Chung





GA- 602077 Version: Final

- KF, Zhang J. Respiratory effects of exposure to diesel traffic in persons with asthma. *N Engl J Med* 2007; 357: 2348-2358.
- 205. Eenhuizen E, Gehring U, Wijga AH, Smit HA, Fischer PH, Brauer M, Koppelman GH, Kerkhof M, de Jongste JC, Brunekreef B, Hoek G. Traffic-related air pollution is related to interrupter resistance in 4-year-old children. *Eur Respir J* 2013; 41: 1257-1263.
- 206. Molter A, Agius RM, de Vocht F, Lindley S, Gerrard W, Lowe L, Belgrave D, Custovic A, Simpson A. Long-term exposure to PM10 and NO2 in association with lung volume and airway resistance in the MAAS birth cohort. *Environ Health Perspect* 2013; 121: 1232-1238.
- 207. Pearce JL, Waller LA, Mulholland JA, Sarnat SE, Strickland MJ, Chang HH, Tolbert PE. Exploring associations between multipollutant day types and asthma morbidity: epidemiologic applications of self-organizing map ambient air quality classifications. *Environmental health*: a global access science source 2015; 14: 55.
- 208. Kraai S, Verhagen LM, Valladares E, Goecke J, Rasquin L, Colmenares P, Del Nogal B, Hermans PW, de Waard JH. High prevalence of asthma symptoms in Warao Amerindian children in Venezuela is significantly associated with open-fire cooking: a cross-sectional observational study. *Respiratory research* 2013; 14: 76.
- 209. Gonzalez-Barcala FJ, Pertega S, Garnelo L, Castro TP, Sampedro M, Lastres JS, San Jose Gonzalez MA, Bamonde L, Valdes L, Carreira JM, Silvarrey AL. Truck traffic related air pollution associated with asthma symptoms in young boys: a cross-sectional study. *Public health* 2013; 127: 275-281.
- 210. Lu KD, Breysse PN, Diette GB, Curtin-Brosnan J, Aloe C, Williams DL, Peng RD, McCormack MC, Matsui EC. Being overweight increases susceptibility to indoor pollutants among urban children with asthma. *J Allergy Clin Immunol* 2013; 131: 1017-1023, 1023 e1011-1013.
- 211. Rider CF, Yamamoto M, Gunther OP, Hirota JA, Singh A, Tebbutt SJ, Carlsten C. Controlled diesel exhaust and allergen coexposure modulates microRNA and gene expression in humans: Effects on inflammatory lung markers. *J Allergy Clin Immunol* 2016.
- 212. Ye Q, Fu JF, Mao JH, Shang SQ. Haze is a risk factor contributing to the rapid spread of respiratory syncytial virus in children. *Environmental science and pollution research international* 2016.
- 213. Guan W ZX, Chung KF, Zhong N. Impact of air pollution on the burden of chronic respiratory diseases in China: Time for urgent action. . Lancet 2016 (in press).
- 214. Proskocil BJ, Fryer AD. Beta2-agonist and anticholinergic drugs in the treatment of lung disease. *Proc Am Thorac Soc* 2005; 2: 305-310; discussion 311-302.
- 215. Mazzone SB, Undem BJ. Vagal Afferent Innervation of the Airways in Health and Disease. *Physiol Rev* 2016; 96: 975-1024.
- 216. West PW, Canning BJ, Merlo-Pich E, Woodcock AA, Smith JA. Morphologic Characterization of Nerves in Whole-Mount Airway Biopsies. *Am J Respir Crit Care Med* 2015; 192: 30-39.
- 217. Moy ML, Lantin ML, Harver A, Schwartzstein RM. Language of dyspnea in assessment of patients with acute asthma treated with nebulized albuterol. *Am J Respir Crit Care Med* 1998; 158: 749-753.





Version: Final

218. Keir S, Boswell-Smith V, Spina D, Page C. Mechanism of adenosine-induced airways obstruction in allergic guinea pigs. *Br J Pharmacol* 2006; 147: 720-728.

- 219. Reynolds SM, Docherty R, Robbins J, Spina D, Page CP. Adenosine induces a cholinergic tracheal reflex contraction in guinea pigs in vivo via an adenosine A1 receptor-dependent mechanism. *Journal of applied physiology (Bethesda, Md*: 1985) 2008; 105: 187-196.
- 220. Raemdonck K, de Alba J, Birrell MA, Grace M, Maher SA, Irvin CG, Fozard JR, O'Byrne PM, Belvisi MG. A role for sensory nerves in the late asthmatic response. *Thorax* 2012; 67: 19-25.
- 221. Lommatzsch M. Airway hyperresponsiveness: new insights into the pathogenesis. Seminars in respiratory and critical care medicine 2012; 33: 579-587.
- 222. Lommatzsch M, Virchow JC. The neural underpinnings of asthma. *J Allergy Clin Immunol* 2007; 119: 254-255; author reply 255.
- 223. Belvisi MG, Birrell MA, Khalid S, Wortley MA, Dockry R, Coote J, Holt K, Dubuis E, Kelsall A, Maher SA, Bonvini S, Woodcock A, Smith JA. Neuro-phenotypes in Airway Diseases: Insights From Translational Cough Studies. *Am J Respir Crit Care Med* 2016.
- 224. Manfreda J, Becklake MR, Sears MR, Chan-Yeung M, Dimich-Ward H, Siersted HC, Ernst P, Sweet L, Van Til L, Bowie DM, Anthonisen NR, Tate RB. Prevalence of asthma symptoms among adults aged 20-44 years in Canada. *Cmaj* 2001; 164: 995-1001.
- 225. Osman LM, McKenzie L, Cairns J, Friend JA, Godden DJ, Legge JS, Douglas JG. Patient weighting of importance of asthma symptoms. *Thorax* 2001; 56: 138-142.
- 226. Mincheva R, Ekerljung L, Bjerg A, Axelsson M, Popov TA, Lundback B, Lotvall J. Frequent cough in unsatisfactory controlled asthma--results from the population-based West Sweden Asthma study. *Respiratory research* 2014; 15: 79.
- 227. Thomson NC, Chaudhuri R, Messow CM, Spears M, MacNee W, Connell M, Murchison JT, Sproule M, McSharry C. Chronic cough and sputum production are associated with worse clinical outcomes in stable asthma. *Respir Med* 2013; 107: 1501-1508.
- 228. Satia I, Tsamandouras N, Holt K, Badri H, Woodhead M, Ogungbenro K, Felton TW, O'Byrne PM, Fowler SJ, Smith JA. Capsaicin-evoked cough responses in asthmatic patients: Evidence for airway neuronal dysfunction. *J Allergy Clin Immunol* 2016.
- 229. Grace MS, Baxter M, Dubuis E, Birrell MA, Belvisi MG. Transient receptor potential (TRP) channels in the airway: role in airway disease. *Br J Pharmacol* 2014; 171: 2593-2607.
- 230. Grace M, Birrell MA, Dubuis E, Maher SA, Belvisi MG. Transient receptor potential channels mediate the tussive response to prostaglandin E2 and bradykinin. *Thorax* 2012; 67: 891-900.
- 231. Carr MJ, Kollarik M, Meeker SN, Undem BJ. A role for TRPV1 in bradykinin-induced excitation of vagal airway afferent nerve terminals. *J Pharmacol Exp Ther* 2003; 304: 1275-1279.
- 232. Kollarik M, Undem BJ. Activation of bronchopulmonary vagal afferent nerves with bradykinin, acid and vanilloid receptor agonists in wild-type and TRPV1-/- mice. *J Physiol* 2004; 555: 115-123.





GA-602077

Version: Final

- 233. Price D, Dale P, Elder E, Chapman KR. Types, frequency and impact of asthma triggers on patients' lives: a quantitative study in five European countries. *J Asthma* 2014; 51: 127-135.
- 234. Grainge CL, Lau LC, Ward JA, Dulay V, Lahiff G, Wilson S, Holgate S, Davies DE, Howarth PH. Effect of bronchoconstriction on airway remodeling in asthma. *N Engl J Med* 2011; 364: 2006-2015.
- 235. Fryer AD, Stein LH, Nie Z, Curtis DE, Evans CM, Hodgson ST, Jose PJ, Belmonte KE, Fitch E, Jacoby DB. Neuronal eotaxin and the effects of CCR3 antagonist on airway hyperreactivity and M2 receptor dysfunction. *J Clin Invest* 2006; 116: 228-236.
- 236. Zaccone EJ, Lieu T, Muroi Y, Potenzieri C, Undem BE, Gao P, Han L, Canning BJ, Undem BJ. Parainfluenza 3-Induced Cough Hypersensitivity in the Guinea Pig Airways. *PLoS One* 2016; 11: e0155526.
- 237. Lommatzsch M, Schloetcke K, Klotz J, Schuhbaeck K, Zingler D, Zingler C, Schulte-Herbruggen O, Gill H, Schuff-Werner P, Virchow JC. Brain-derived neurotrophic factor in platelets and airflow limitation in asthma. *Am J Respir Crit Care Med* 2005; 171: 115-120.
- 238. Lieu TM, Myers AC, Meeker S, Undem BJ. TRPV1 induction in airway vagal low-threshold mechanosensory neurons by allergen challenge and neurotrophic factors. *American journal of physiology Lung cellular and molecular physiology* 2012; 302: L941-948.
- 239. Keir S, Page C, Spina D. Bronchial hyperresponsiveness induced by chronic treatment with albuterol: Role of sensory nerves. *J Allergy Clin Immunol* 2002; 110: 388-394.
- 240. Kupczyk M, Wenzel S. U.S. and European severe asthma cohorts: what can they teach us about severe asthma? *Journal of internal medicine* 2012; 272: 121-132.
- 241. Lipworth BJ. Emerging role of long acting muscarinic antagonists for asthma. British journal of clinical pharmacology 2014; 77: 55-62.
- 242. Kerstjens HA, Casale TB, Bleecker ER, Meltzer EO, Pizzichini E, Schmidt O, Engel M, Bour L, Verkleij CB, Moroni-Zentgraf P, Bateman ED. Tiotropium or salmeterol as add-on therapy to inhaled corticosteroids for patients with moderate symptomatic asthma: two replicate, double-blind, placebo-controlled, parallel-group, active-comparator, randomised trials. *Lancet Respir Med* 2015; 3: 367-376.
- 243. Toren K, Blanc PD. Asthma caused by occupational exposures is common a systematic analysis of estimates of the population-attributable fraction. *BMC pulmonary medicine* 2009; 9: 7.
- 244. Siracusa A, De Blay F, Folletti I, Moscato G, Olivieri M, Quirce S, Raulf-Heimsoth M, Sastre J, Tarlo SM, Walusiak-Skorupa J, Zock JP. Asthma and exposure to cleaning products a European Academy of Allergy and Clinical Immunology task force consensus statement. *Allergy* 2013; 68: 1532-1545.
- 245. Thickett KM, McCoach JS, Gerber JM, Sadhra S, Burge PS. Occupational asthma caused by chloramines in indoor swimming-pool air. *Eur Respir J* 2002; 19: 827-832.
- 246. Wenzel S, Holgate ST. The mouse trap: It still yields few answers in asthma. Am J Respir Crit Care Med 2006; 174: 1173-1176; discussion 1176-1178.
- 247. Seehase S, Schleputz M, Switalla S, Matz-Rensing K, Kaup FJ, Zoller M, Schlumbohm C, Fuchs E, Lauenstein HD, Winkler C, Kuehl AR, Uhlig S, Braun A, Sewald K, Martin C. Bronchoconstriction in nonhuman primates: a species





Version: Final

comparison. *Journal of applied physiology (Bethesda, Md : 1985)* 2011; 111: 791-798.

- 248. Birrell MA, Bonvini SJ, Dubuis E, Maher SA, Wortley MA, Grace MS, Raemdonck K, Adcock JJ, Belvisi MG. Tiotropium modulates transient receptor potential V1 (TRPV1) in airway sensory nerves: A beneficial off-target effect? *J Allergy Clin Immunol* 2014; 133: 679-687 e679.
- 249. Braun A. Animal models of asthma. Current drug targets 2008; 9: 436-437.
- 250. Abdulqawi R, Dockry R, Holt K, Layton G, McCarthy BG, Ford AP, Smith JA. P2X3 receptor antagonist (AF-219) in refractory chronic cough: a randomised, double-blind, placebo-controlled phase 2 study. *Lancet* 2015; 385: 1198-1205.
- 251. Khalid S, Murdoch R, Newlands A, Smart K, Kelsall A, Holt K, Dockry R, Woodcock A, Smith JA. Transient receptor potential vanilloid 1 (TRPV1) antagonism in patients with refractory chronic cough: a double-blind randomized controlled trial. *J Allergy Clin Immunol* 2014; 134: 56-62.
- 252. Hall S, Agrawal DK. Key mediators in the immunopathogenesis of allergic asthma. *Int Immunopharmacol* 2014; 23: 316-329.
- 253. Szefler SJ. Advances in pediatric asthma in 2014: Moving toward a population health perspective. *J Allergy Clin Immunol* 2015; 135: 644-652.
- 254. Danahay H, Pessotti AD, Coote J, Montgomery BE, Xia D, Wilson A, Yang H, Wang Z, Bevan L, Thomas C, Petit S, London A, LeMotte P, Doelemeyer A, Velez-Reyes GL, Bernasconi P, Fryer CJ, Edwards M, Capodieci P, Chen A, Hild M, Jaffe AB. Notch2 is required for inflammatory cytokine-driven goblet cell metaplasia in the lung. *Cell reports* 2015; 10: 239-252.
- 255. Bellini A, Marini MA, Bianchetti L, Barczyk M, Schmidt M, Mattoli S. Interleukin (IL)-4, IL-13, and IL-17A differentially affect the profibrotic and proinflammatory functions of fibrocytes from asthmatic patients. *Mucosal Immunol* 2012; 5: 140-149.
- 256. Xiang YY, Wang S, Liu M, Hirota JA, Li J, Ju W, Fan Y, Kelly MM, Ye B, Orser B, O'Byrne PM, Inman MD, Yang X, Lu WY. A GABAergic system in airway epithelium is essential for mucus overproduction in asthma. *Nat Med* 2007; 13: 862-867.
- 257. Shi N, Zhang J, Chen SY. Runx2, a novel regulator for goblet cell differentiation and asthma development. FASEB journal: official publication of the Federation of American Societies for Experimental Biology 2017; 31: 412-420.
- 258. Wechsler ME, Laviolette M, Rubin AS, Fiterman J, Lapa e Silva JR, Shah PL, Fiss E, Olivenstein R, Thomson NC, Niven RM, Pavord ID, Simoff M, Hales JB, McEvoy C, Slebos DJ, Holmes M, Phillips MJ, Erzurum SC, Hanania NA, Sumino K, Kraft M, Cox G, Sterman DH, Hogarth K, Kline JN, Mansur AH, Louie BE, Leeds WM, Barbers RG, Austin JH, Shargill NS, Quiring J, Armstrong B, Castro M, Asthma Intervention Research 2 Trial Study G. Bronchial thermoplasty: Long-term safety and effectiveness in patients with severe persistent asthma. *J Allergy Clin Immunol* 2013; 132: 1295-1302.
- 259. Pain M, Bermudez O, Lacoste P, Royer PJ, Botturi K, Tissot A, Brouard S, Eickelberg O, Magnan A. Tissue remodelling in chronic bronchial diseases: from the epithelial to mesenchymal phenotype. *Eur Respir Rev* 2014; 23: 118-130.





Version: Final

© Copyright 2016, EARIP Consortium

- 260. Chachi L, Gavrila A, Tliba O, Amrani Y. Abnormal corticosteroid signalling in airway smooth muscle: mechanisms and perspectives for the treatment of severe asthma. *Clin Exp Allergy* 2015; 45: 1637-1646.
- 261. Hardy CL, Rolland JM, O'Hehir RE. The immunoregulatory and fibrotic roles of activin A in allergic asthma. *Clin Exp Allergy* 2015; 45: 1510-1522.
- 262. Boorsma CE, Dekkers BG, van Dijk EM, Kumawat K, Richardson J, Burgess JK, John AE. Beyond TGFB--novel ways to target airway and parenchymal fibrosis. *Pulm Pharmacol Ther* 2014; 29: 166-180.
- 263. Lloyd CM, Saglani S. Epithelial cytokines and pulmonary allergic inflammation. *Curr Opin Immunol* 2015; 34: 52-58.
- 264. Durham SR, Walker SM, Varga EM, Jacobson MR, O'Brien F, Noble W, Till SJ, Hamid QA, Nouri-Aria KT. Long-term clinical efficacy of grass-pollen immunotherapy. *N Engl J Med* 1999; 341: 468-475.
- 265. Akdis CA, Akdis M. Advances in allergen immunotherapy: aiming for complete tolerance to allergens. *Sci Transl Med* 2015; 7: 280ps286.
- 266. Serhan CN. Pro-resolving lipid mediators are leads for resolution physiology. *Nature* 2014; 510: 92-101.
- 267. Buckley CD, Gilroy DW, Serhan CN. Proresolving lipid mediators and mechanisms in the resolution of acute inflammation. *Immunity* 2014; 40: 315-327.
- 268. Levy BD, Serhan CN. Resolution of acute inflammation in the lung. *Annu Rev Physiol* 2014; 76: 467-492.
- 269. Levy BD, Bonnans C, Silverman ES, Palmer LJ, Marigowda G, Israel E, Severe Asthma Research Program NHL, Blood I. Diminished lipoxin biosynthesis in severe asthma. *Am J Respir Crit Care Med* 2005; 172: 824-830.
- 270. Vachier I, Bonnans C, Chavis C, Farce M, Godard P, Bousquet J, Chanez P. Severe asthma is associated with a loss of LX4, an endogenous anti-inflammatory compound. *J Allergy Clin Immunol* 2005; 115: 55-60.
- 271. Levy BD, Kohli P, Gotlinger K, Haworth O, Hong S, Kazani S, Israel E, Haley KJ, Serhan CN. Protectin D1 is generated in asthma and dampens airway inflammation and hyperresponsiveness. *J Immunol* 2007; 178: 496-502.
- 272. Planaguma A, Kazani S, Marigowda G, Haworth O, Mariani TJ, Israel E, Bleecker ER, Curran-Everett D, Erzurum SC, Calhoun WJ, Castro M, Chung KF, Gaston B, Jarjour NN, Busse WW, Wenzel SE, Levy BD. Airway lipoxin A4 generation and lipoxin A4 receptor expression are decreased in severe asthma. Am J Respir Crit Care Med 2008; 178: 574-582.
- 273. Kazani S, Planaguma A, Ono E, Bonini M, Zahid M, Marigowda G, Wechsler ME, Levy BD, Israel E. Exhaled breath condensate eicosanoid levels associate with asthma and its severity. *J Allergy Clin Immunol* 2013; 132: 547-553.
- 274. Levy BD, De Sanctis GT, Devchand PR, Kim E, Ackerman K, Schmidt BA, Szczeklik W, Drazen JM, Serhan CN. Multi-pronged inhibition of airway hyper-responsiveness and inflammation by lipoxin A(4). *Nat Med* 2002; 8: 1018-1023.
- 275. Aoki H, Hisada T, Ishizuka T, Utsugi M, Kawata T, Shimizu Y, Okajima F, Dobashi K, Mori M. Resolvin E1 dampens airway inflammation and hyperresponsiveness in a murine model of asthma. *Biochem Biophys Res Commun* 2008; 367: 509-515.
- 276. Haworth O, Cernadas M, Yang R, Serhan CN, Levy BD. Resolvin E1 regulates interleukin 23, interferon-gamma and lipoxin A4 to promote the resolution of allergic airway inflammation. *Nat Immunol* 2008; 9: 873-879.





Version: Final

277. Rogerio AP, Haworth O, Croze R, Oh SF, Uddin M, Carlo T, Pfeffer MA, Priluck R, Serhan CN, Levy BD. Resolvin D1 and aspirin-triggered resolvin D1 promote resolution of allergic airways responses. *J Immunol* 2012; 189: 1983-1991.

- 278. Koltsida O, Karamnov S, Pyrillou K, Vickery T, Chairakaki AD, Tamvakopoulos C, Sideras P, Serhan CN, Andreakos E. Toll-like receptor 7 stimulates production of specialized pro-resolving lipid mediators and promotes resolution of airway inflammation. *EMBO Mol Med* 2013; 5: 762-775.
- 279. Salpeter SR, Ormiston TM, Salpeter EE. Meta-analysis: respiratory tolerance to regular beta2-agonist use in patients with asthma. *Annals of internal medicine* 2004; 140: 802-813.
- 280. Drazen JM, Israel E, Boushey HA, Chinchilli VM, Fahy JV, Fish JE, Lazarus SC, Lemanske RF, Martin RJ, Peters SP, Sorkness C, Szefler SJ. Comparison of regularly scheduled with as-needed use of albuterol in mild asthma. Asthma Clinical Research Network. *N Engl J Med* 1996; 335: 841-847.
- 281. Anderson HR, Ayres JG, Sturdy PM, Bland JM, Butland BK, Peckitt C, Taylor JC, Victor CR. Bronchodilator treatment and deaths from asthma: case-control study. *Bmj* 2005; 330: 117.
- 282. Levy ML. The national review of asthma deaths: what did we learn and what needs to change? *Breathe (Sheff)* 2015; 11: 14-24.
- 283. Hasford J, Virchow JC. Excess mortality in patients with asthma on long-acting beta2-agonists. *Eur Respir J* 2006; 28: 900-902.
- 284. Nelson HS, Weiss ST, Bleecker ER, Yancey SW, Dorinsky PM. The Salmeterol Multicenter Asthma Research Trial: a comparison of usual pharmacotherapy for asthma or usual pharmacotherapy plus salmeterol. *Chest* 2006; 129: 15-26.
- 285. Castle W, Fuller R, Hall J, Palmer J. Serevent nationwide surveillance study: comparison of salmeterol with salbutamol in asthmatic patients who require regular bronchodilator treatment. *Bmj* 1993; 306: 1034-1037.
- 286. Drazen JM, O'Byrne PM. Risks of long-acting beta-agonists in achieving asthma control. *N Engl J Med* 2009; 360: 1671-1672.
- 287. Stempel DA, Raphiou IH, Kral KM, Yeakey AM, Emmett AH, Prazma CM, Buaron KS, Pascoe SJ, Investigators A. Serious Asthma Events with Fluticasone plus Salmeterol versus Fluticasone Alone. *N Engl J Med* 2016; 374: 1822-1830.
- 288. Stempel DA, Szefler SJ, Pedersen S, Zeiger RS, Yeakey AM, Lee LA, Liu AH, Mitchell H, Kral KM, Raphiou IH, Prillaman BA, Buaron KS, Yun Kirby S, Pascoe SJ, Investigators V. Safety of Adding Salmeterol to Fluticasone Propionate in Children with Asthma. *N Engl J Med* 2016; 375: 840-849.
- 289. Crane J, Pearce N, Flatt A, Burgess C, Jackson R, Kwong T, Ball M, Beasley R. Prescribed fenoterol and death from asthma in New Zealand, 1981-83: case-control study. *Lancet* 1989; 1: 917-922.
- 290. Sears MR. Adverse effects of beta-agonists. *J Allergy Clin Immunol* 2002; 110: S322-328.
- 291. Edwards MR, Haas J, Panettieri RA, Jr., Johnson M, Johnston SL. Corticosteroids and beta2 agonists differentially regulate rhinovirus-induced interleukin-6 via distinct Cis-acting elements. *J Biol Chem* 2007; 282: 15366-15375.
- 292. Lommatzsch M, Lindner Y, Edner A, Bratke K, Kuepper M, Virchow JC. Adverse effects of salmeterol in asthma: a neuronal perspective. *Thorax* 2009; 64: 763-769.





RIP Consortium Version: Final

- 293. O'Kane CM, McKeown SW, Perkins GD, Bassford CR, Gao F, Thickett DR, McAuley DF. Salbutamol up-regulates matrix metalloproteinase-9 in the alveolar space in the acute respiratory distress syndrome. *Crit Care Med* 2009; 37: 2242-2249.
- 294. Tabuchi A, Sakaya H, Kisukeda T, Fushiki H, Tsuda M. Involvement of an upstream stimulatory factor as well as cAMP-responsive element-binding protein in the activation of brain-derived neurotrophic factor gene promoter I. *J Biol Chem* 2002; 277: 35920-35931.
- 295. Park JK, Park SH, So K, Bae IH, Yoo YD, Um HD. ICAM-3 enhances the migratory and invasive potential of human non-small cell lung cancer cells by inducing MMP-2 and MMP-9 via Akt and CREB. *International journal of oncology* 2010; 36: 181-192.
- 296. Johnston SL, Edwards MR. Mechanisms of adverse effects of {beta}-agonists in asthma. *Thorax* 2009; 64: 739-741.
- 297. Zhang X, Odom DT, Koo SH, Conkright MD, Canettieri G, Best J, Chen H, Jenner R, Herbolsheimer E, Jacobsen E, Kadam S, Ecker JR, Emerson B, Hogenesch JB, Unterman T, Young RA, Montminy M. Genome-wide analysis of cAMP-response element binding protein occupancy, phosphorylation, and target gene activation in human tissues. *Proc Natl Acad Sci U S A* 2005; 102: 4459-4464.
- 298. Bibby S, Healy B, Steele R, Kumareswaran K, Nelson H, Beasley R. Association between leukotriene receptor antagonist therapy and Churg-Strauss syndrome: an analysis of the FDA AERS database. *Thorax* 2010; 65: 132-138.
- 299. Finney L, Berry M, Singanayagam A, Elkin SL, Johnston SL, Mallia P. Inhaled corticosteroids and pneumonia in chronic obstructive pulmonary disease. *Lancet Respir Med* 2014; 2: 919-932.
- 300. Singanayagam A, Glanville N, Bartlett N, Johnston S. Effect of fluticasone propionate on virus-induced airways inflammation and anti-viral immune responses in mice. *Lancet* 2015; 385 Suppl 1: S88.
- 301. Simpson JL, Carroll M, Yang IA, Reynolds PN, Hodge S, James AL, Gibson PG, Upham JW. Reduced Antiviral Interferon Production in Poorly Controlled Asthma Is Associated With Neutrophilic Inflammation and High-Dose Inhaled Corticosteroids. *Chest* 2016; 149: 704-713.
- 302. Murray CS, Poletti G, Kebadze T, Morris J, Woodcock A, Johnston SL, Custovic A. Study of modifiable risk factors for asthma exacerbations: virus infection and allergen exposure increase the risk of asthma hospital admissions in children. *Thorax* 2006; 61: 376-382.
- 303. Ortega VE, Meyers DA, Bleecker ER. Asthma pharmacogenetics and the development of genetic profiles for personalized medicine. *Pharmacogenomics and personalized medicine* 2015; 8: 9-22.
- 304. van Huisstede A, Castro Cabezas M, van de Geijn GJ, Mannaerts GH, Njo TL, Taube C, Hiemstra PS, Braunstahl GJ. Underdiagnosis and overdiagnosis of asthma in the morbidly obese. *Respir Med* 2013; 107: 1356-1364.
- 305. Pradeepan S, Garrison G, Dixon AE. Obesity in asthma: approaches to treatment. *Curr Allergy Asthma Rep* 2013; 13: 434-442.
- 306. Bhatt NA, Lazarus A. Obesity-related asthma in adults. *Postgraduate medicine* 2016; 128: 563-566.
- 307. Sin DD, Jones RL, Man SF. Obesity is a risk factor for dyspnea but not for airflow obstruction. *Arch Intern Med* 2002; 162: 1477-1481.





Version: Final

308. Shore SA. Obesity and asthma: possible mechanisms. *J Allergy Clin Immunol* 2008; 121: 1087-1093; quiz 1094-1085.

- 309. Ali Assad N, Sood A. Leptin, adiponectin and pulmonary diseases. *Biochimie* 2012; 94: 2180-2189.
- 310. Wood LG, Garg ML, Gibson PG. A high-fat challenge increases airway inflammation and impairs bronchodilator recovery in asthma. *J Allergy Clin Immunol* 2011; 127: 1133-1140.
- 311. Lugogo NL, Bappanad D, Kraft M. Obesity, metabolic dysregulation and oxidative stress in asthma. *Biochim Biophys Acta* 2011; 1810: 1120-1126.
- 312. Kasahara DI, Kim HY, Williams AS, Verbout NG, Tran J, Si H, Wurmbrand AP, Jastrab J, Hug C, Umetsu DT, Shore SA. Pulmonary inflammation induced by subacute ozone is augmented in adiponectin-deficient mice: role of IL-17A. *J Immunol* 2012; 188: 4558-4567.
- 313. Sivapalan P, Diamant Z, Ulrik CS. Obesity and asthma: current knowledge and future needs. *Curr Opin Pulm Med* 2015; 21: 80-85.
- 314. van Huisstede A, Rudolphus A, Castro Cabezas M, Biter LU, van de Geijn GJ, Taube C, Hiemstra PS, Braunstahl GJ. Effect of bariatric surgery on asthma control, lung function and bronchial and systemic inflammation in morbidly obese subjects with asthma. *Thorax* 2015; 70: 659-667.
- 315. Adeniyi FB, Young T. Weight loss interventions for chronic asthma. *Cochrane Database Syst Rev* 2012; 7: CD009339.
- 316. Peroni DG, Piacentini GL, Bodini A, Boner AL. Preschool asthma in Italy: prevalence, risk factors and health resource utilization. *Respir Med* 2009; 103: 104-108.
- 317. Keselman A, Heller N. Estrogen Signaling Modulates Allergic Inflammation and Contributes to Sex Differences in Asthma. *Front Immunol* 2015; 6: 568.
- 318. Zein JG, Erzurum SC. Asthma is Different in Women. *Curr Allergy Asthma Rep* 2015; 15: 28.
- 319. Govaere E, Van Gysel D, Verhamme KM, Doli E, De Baets F. The association of allergic symptoms with sensitization to inhalant allergens in childhood. *Pediatr Allergy Immunol* 2009; 20: 448-457.
- 320. Almqvist C, Worm M, Leynaert B. Impact of gender on asthma in childhood and adolescence: a GA2LEN review. *Allergy* 2008; 63: 47-57.
- 321. Tantisira KG, Colvin R, Tonascia J, Strunk RC, Weiss ST, Fuhlbrigge AL. Airway responsiveness in mild to moderate childhood asthma: sex influences on the natural history. *Am J Respir Crit Care Med* 2008; 178: 325-331.
- 322. Baibergenova A, Thabane L, Akhtar-Danesh N, Levine M, Gafni A, Leeb K. Sex differences in hospital admissions from emergency departments in asthmatic adults: a population-based study. *Ann Allergy Asthma Immunol* 2006; 96: 666-672.
- 323. Rao CK, Moore CG, Bleecker E, Busse WW, Calhoun W, Castro M, Chung KF, Erzurum SC, Israel E, Curran-Everett D, Wenzel SE. Characteristics of perimenstrual asthma and its relation to asthma severity and control: data from the Severe Asthma Research Program. *Chest* 2013; 143: 984-992.
- 324. Schatz M, Harden K, Forsythe A, Chilingar L, Hoffman C, Sperling W, Zeiger RS. The course of asthma during pregnancy, post partum, and with successive pregnancies: a prospective analysis. *J Allergy Clin Immunol* 1988; 81: 509-517.
- 325. Virchow JC. Asthma and pregnancy. Seminars in respiratory and critical care medicine 2012; 33: 630-644.





GA-602077

© Copyright 2016, EARIP Consortium

Version: Final

- 326. Troisi RJ, Speizer FE, Willett WC, Trichopoulos D, Rosner B. Menopause, postmenopausal estrogen preparations, and the risk of adult-onset asthma. A prospective cohort study. *Am J Respir Crit Care Med* 1995; 152: 1183-1188.
- 327. Foschino Barbaro MP, Costa VR, Resta O, Prato R, Spanevello A, Palladino GP, Martinelli D, Carpagnano GE. Menopausal asthma: a new biological phenotype? *Allergy* 2010; 65: 1306-1312.
- 328. Tan KS, McFarlane LC, Lipworth BJ. Modulation of airway reactivity and peak flow variability in asthmatics receiving the oral contraceptive pill. *Am J Respir Crit Care Med* 1997; 155: 1273-1277.
- 329. Farha S, Asosingh K, Laskowski D, Hammel J, Dweik RA, Wiedemann HP, Erzurum SC. Effects of the menstrual cycle on lung function variables in women with asthma. *Am J Respir Crit Care Med* 2009; 180: 304-310.
- 330. Pauli BD, Reid RL, Munt PW, Wigle RD, Forkert L. Influence of the menstrual cycle on airway function in asthmatic and normal subjects. *Am Rev Respir Dis* 1989; 140: 358-362.
- 331. Lieberoth S, Gade EJ, Brok J, Backer V, Thomsen SF. Age at menarche and risk of asthma: systematic review and meta-analysis. *J Asthma* 2014; 51: 559-565.
- 332. Apter D, Reinila M, Vihko R. Some endocrine characteristics of early menarche, a risk factor for breast cancer, are preserved into adulthood. *Int J Cancer* 1989; 44: 783-787.
- 333. Lange P, Parner J, Prescott E, Ulrik CS, Vestbo J. Exogenous female sex steroid hormones and risk of asthma and asthma-like symptoms: a cross sectional study of the general population. *Thorax* 2001; 56: 613-616.
- 334. Salam MT, Wenten M, Gilliland FD. Endogenous and exogenous sex steroid hormones and asthma and wheeze in young women. *J Allergy Clin Immunol* 2006; 117: 1001-1007.
- 335. Hepper PG, Shannon EA, Dornan JC. Sex differences in fetal mouth movements. *Lancet* 1997; 350: 1820.
- 336. Thurlbeck WM. Postnatal human lung growth. Thorax 1982; 37: 564-571.
- 337. DeGroodt EG, van Pelt W, Borsboom GJ, Quanjer PH, van Zomeren BC. Growth of lung and thorax dimensions during the pubertal growth spurt. *Eur Respir J* 1988; 1: 102-108.
- 338. Hibbert M, Lannigan A, Raven J, Landau L, Phelan P. Gender differences in lung growth. *Pediatr Pulmonol* 1995; 19: 129-134.
- 339. Clark NM, Shah S, Dodge JA, Thomas LJ, Andridge RR, Little RJ. An evaluation of asthma interventions for preteen students. *J Sch Health* 2010; 80: 80-87.
- 340. Piaget J. The stages of the intellectual development of the child. *Bull Menninger Clin* 1962; 26: 120-128.
- 341. Clark NM, Dodge JA, Thomas LJ, Andridge RR, Awad D, Paton JY. Asthma in 10- to 13-year-olds: challenges at a time of transition. *Clin Pediatr (Phila)* 2010; 49: 931-937.
- 342. Forero R, Bauman A, Young L, Booth M, Nutbeam D. Asthma, health behaviors, social adjustment, and psychosomatic symptoms in adolescence. *J Asthma* 1996; 33: 157-164.
- 343. Rew L. The relationship between self-care behaviors and selected psychosocial variables in children with asthma. *J Pediatr Nurs* 1987; 2: 333-341.





Version: Final

344. Williams C. Doing health, doing gender: teenagers, diabetes and asthma. Soc Sci Med 2000; 50: 387-396.

- 345. Gade EJ, Thomsen SF, Lindenberg S, Kyvik KO, Lieberoth S, Backer V. Asthma affects time to pregnancy and fertility: a register-based twin study. *Eur Respir J* 2014; 43: 1077-1085.
- 346. Murphy VE, Gibson PG. Asthma in pregnancy. *Clin Chest Med* 2011; 32: 93-110, ix.
- 347. Scott KM, Von Korff M, Ormel J, Zhang MY, Bruffaerts R, Alonso J, Kessler RC, Tachimori H, Karam E, Levinson D, Bromet EJ, Posada-Villa J, Gasquet I, Angermeyer MC, Borges G, de Girolamo G, Herman A, Haro JM. Mental disorders among adults with asthma: results from the World Mental Health Survey. *General hospital psychiatry* 2007; 29: 123-133.
- 348. Vuillermin PJ, Brennan SL, Robertson CF, Carlin JB, Prior M, Jenner BM, South M. Anxiety is more common in children with asthma. *Archives of disease in childhood* 2010; 95: 624-629.
- 349. Lavoie KL, Bacon SL, Barone S, Cartier A, Ditto B, Labrecque M. What is worse for asthma control and quality of life: depressive disorders, anxiety disorders, or both? *Chest* 2006; 130: 1039-1047.
- 350. Cluley S, Cochrane GM. Psychological disorder in asthma is associated with poor control and poor adherence to inhaled steroids. *Respir Med* 2001; 95: 37-39.
- 351. Goldney RD, Ruffin R, Fisher LJ, Wilson DH. Asthma symptoms associated with depression and lower quality of life: a population survey. *The Medical journal of Australia* 2003; 178: 437-441.
- 352. ten Brinke A, Ouwerkerk ME, Zwinderman AH, Spinhoven P, Bel EH. Psychopathology in patients with severe asthma is associated with increased health care utilization. *Am J Respir Crit Care Med* 2001; 163: 1093-1096.
- 353. Dahlem NW, Kinsman RA, Horton DJ. Panic-fear in asthma: requests for asneeded medications in relation to pulmonary function measurements. *J Allergy Clin Immunol* 1977; 60: 295-300.
- 354. Thomas M, McKinley RK, Mellor S, Watkin G, Holloway E, Scullion J, Shaw DE, Wardlaw A, Price D, Pavord I. Breathing exercises for asthma: a randomised controlled trial. *Thorax* 2009; 64: 55-61.
- 355. Freitas DA, Holloway EA, Bruno SS, Chaves GS, Fregonezi GA, Mendonca KP. Breathing exercises for adults with asthma. *Cochrane Database Syst Rev* 2013: CD001277.
- 356. Gamble J, Stevenson M, Heaney LG. A study of a multi-level intervention to improve non-adherence in difficult to control asthma. *Respir Med* 2011; 105: 1308-1315.
- 357. McLean G, Murray E, Band R, Moffat KR, Hanlon P, Bruton A, Thomas M, Yardley L, Mair FS. Interactive digital interventions to promote self-management in adults with asthma: systematic review and meta-analysis. *BMC pulmonary medicine* 2016; 16: 83.
- 358. Yorke J, Fleming S, Shuldham C, Rao H, Smith HE. Nonpharmacological interventions aimed at modifying health and behavioural outcomes for adults with asthma: a critical review. *Clin Exp Allergy* 2015; 45: 1750-1764.
- 359. von Leupoldt A, Sommer T, Kegat S, Baumann HJ, Klose H, Dahme B, Buchel C. Dyspnea and pain share emotion-related brain network. *NeuroImage* 2009; 48: 200-206.





GA-602077

- 360. Lavoie KL, Bouthillier D, Bacon SL, Lemiere C, Martin J, Hamid Q, Ludwig M, Olivenstein R, Ernst P. Psychologic distress and maladaptive coping styles in patients with severe vs moderate asthma. *Chest* 2010; 137: 1324-1331.
- 361. Faull OK, Jenkinson M, Ezra M, Pattinson K. Conditioned respiratory threat in the subdivisions of the human periaqueductal gray. *eLife* 2016; 5.
- 362. Rosenkranz MA, Esnault S, Christian BT, Crisafi G, Gresham LK, Higgins AT, Moore MN, Moore SM, Weng HY, Salk RH, Busse WW, Davidson RJ. Mind-body interactions in the regulation of airway inflammation in asthma: A PET study of acute and chronic stress. *Brain, behavior, and immunity* 2016.
- 363. Rosenkranz MA, Busse WW, Sheridan JF, Crisafi GM, Davidson RJ. Are there neurophenotypes for asthma? Functional brain imaging of the interaction between emotion and inflammation in asthma. *PLoS One* 2012; 7: e40921.
- 364. Janssens T, Verleden G, De Peuter S, Van Diest I, Van den Bergh O. Inaccurate perception of asthma symptoms: a cognitive-affective framework and implications for asthma treatment. *Clinical psychology review* 2009; 29: 317-327.
- 365. Spinhoven P, van Peski-Oosterbaan AS, Van der Does AJ, Willems LN, Sterk PJ. Association of anxiety with perception of histamine induced bronchoconstriction in patients with asthma. *Thorax* 1997; 52: 149-152.
- 366. Laveneziana P, Lotti P, Coli C, Binazzi B, Chiti L, Stendardi L, Duranti R, Scano G. Mechanisms of dyspnoea and its language in patients with asthma. *Eur Respir J* 2006; 27: 742-747.
- 367. Gauvreau GM, O'Byrne PM, Boulet LP, Wang Y, Cockcroft D, Bigler J, FitzGerald JM, Boedigheimer M, Davis BE, Dias C, Gorski KS, Smith L, Bautista E, Comeau MR, Leigh R, Parnes JR. Effects of an anti-TSLP antibody on allergen-induced asthmatic responses. *N Engl J Med* 2014; 370: 2102-2110.
- 368. Bernink JH, Krabbendam L, Germar K, de Jong E, Gronke K, Kofoed-Nielsen M, Munneke JM, Hazenberg MD, Villaudy J, Buskens CJ, Bemelman WA, Diefenbach A, Blom B, Spits H. Interleukin-12 and -23 Control Plasticity of CD127(+) Group 1 and Group 3 Innate Lymphoid Cells in the Intestinal Lamina Propria. *Immunity* 2015; 43: 146-160.
- 369. Bal SM, Bernink JH, Nagasawa M, Groot J, Shikhagaie MM, Golebski K, van Drunen CM, Lutter R, Jonkers RE, Hombrink P, Bruchard M, Villaudy J, Munneke JM, Fokkens W, Erjefalt JS, Spits H, Ros XR. IL-1beta, IL-4 and IL-12 control the fate of group 2 innate lymphoid cells in human airway inflammation in the lungs. *Nat Immunol* 2016; 17: 636-645.
- 370. Lane S, Molina J, Plusa T. An international observational prospective study to determine the cost of asthma exacerbations (COAX). *Respir Med* 2006; 100: 434-450.
- 371. Green RM, Custovic A, Sanderson G, Hunter J, Johnston SL, Woodcock A. Synergism between allergens and viruses and risk of hospital admission with asthma: case-control study. *Bmj* 2002; 324: 763.
- 372. Chauhan AJ, Inskip HM, Linaker CH, Smith S, Schreiber J, Johnston SL, Holgate ST. Personal exposure to nitrogen dioxide (NO2) and the severity of virus-induced asthma in children. *Lancet* 2003; 361: 1939-1944.
- 373. Wark PA, Johnston SL, Simpson JL, Hensley MJ, Gibson PG. Chlamydia pneumoniae immunoglobulin A reactivation and airway inflammation in acute asthma. *Eur Respir J* 2002; 20: 834-840.





Version: Final

374. Tran TN, Khatry DB, Ke X, Ward CK, Gossage D. High blood eosinophil count is associated with more frequent asthma attacks in asthma patients. *Ann Allergy Asthma Immunol* 2014; 113: 19-24.

- 375. Zeiger RS, Schatz M, Li Q, Chen W, Khatry DB, Gossage D, Tran TN. High blood eosinophil count is a risk factor for future asthma exacerbations in adult persistent asthma. *J Allergy Clin Immunol Pract* 2014; 2: 741-750.
- 376. Tanaka A, Jinno M, Hirai K, Miyata Y, Mizuma H, Yamaguchi M, Ohta S, Watanabe Y, Yamamoto M, Suzuki S, Yokoe T, Adachi M, Sagara H. Longitudinal increase in total IgE levels in patients with adult asthma: an association with poor asthma control. *Respiratory research* 2014; 15: 144.
- 377. Hasegawa K, Sullivan AF, Tovar Hirashima E, Gaeta TJ, Fee C, Turner SJ, Massaro S, Camargo CA, Jr., Multicenter Airway Research Collaboration I. A multicenter observational study of US adults with acute asthma: who are the frequent users of the emergency department? *J Allergy Clin Immunol Pract* 2014; 2: 733-740.
- 378. Kanchongkittiphon W, Mendell MJ, Gaffin JM, Wang G, Phipatanakul W. Indoor environmental exposures and exacerbation of asthma: an update to the 2000 review by the institute of medicine. *Environ Health Perspect* 2015; 123: 6-20.
- 379. Qiu H, Yu IT, Tse LA, Chan EY, Wong TW, Tian L. Greater temperature variation within a day associated with increased emergency hospital admissions for asthma. *The Science of the total environment* 2015; 505: 508-513.
- 380. Confino-Cohen R, Brufman I, Goldberg A, Feldman BS. Vitamin D, asthma prevalence and asthma exacerbations: a large adult population-based study. *Allergy* 2014; 69: 1673-1680.
- 381. Salas NM, Luo L, Harkins MS. Vitamin D deficiency and adult asthma exacerbations. *J Asthma* 2014; 51: 950-955.
- 382. Engelkes M, Janssens HM, de Jongste JC, Sturkenboom MC, Verhamme KM. Medication adherence and the risk of severe asthma exacerbations: a systematic review. *Eur Respir J* 2015; 45: 396-407.
- 383. Jaen C, Dalton P. Asthma and odors: the role of risk perception in asthma exacerbation. *Journal of psychosomatic research* 2014; 77: 302-308.
- 384. Bateman ED, Buhl R, O'Byrne PM, Humbert M, Reddel HK, Sears MR, Jenkins C, Harrison TW, Quirce S, Peterson S, Eriksson G. Development and validation of a novel risk score for asthma exacerbations: The risk score for exacerbations. *J Allergy Clin Immunol* 2015; 135: 1457-1464 e1454.
- 385. Maekawa T, Oba MS, Katsunuma T, Ishiguro A, Ohya Y, Nakamura H. Modified pulmonary index score was sufficiently reliable to assess the severity of acute asthma exacerbations in children. *Allergology international: official journal of the Japanese Society of Allergology* 2014; 63: 603-607.
- 386. Schneider JE, Lewis LM, Ferguson I, House SL, Liu J, Matsuda K, Johnson K. Repeated dyspnea score and percent FEV1 are modest predictors of hospitalization/relapse in patients with acute asthma exacerbation. *Respir Med* 2014; 108: 1284-1291.
- 387. Papadopoulos NG, Christodoulou I, Rohde G, Agache I, Almqvist C, Bruno A, Bonini S, Bont L, Bossios A, Bousquet J, Braido F, Brusselle G, Canonica GW, Carlsen KH, Chanez P, Fokkens WJ, Garcia-Garcia M, Gjomarkaj M, Haahtela T, Holgate ST, Johnston SL, Konstantinou G, Kowalski M, Lewandowska-Polak A, Lodrup-Carlsen K, Makela M, Malkusova I, Mullol J, Nieto A, Eller E,





GA- 602077

- Ozdemir C, Panzner P, Popov T, Psarras S, Roumpedaki E, Rukhadze M, Stipic-Markovic A, Todo Bom A, Toskala E, van Cauwenberge P, van Drunen C, Watelet JB, Xatzipsalti M, Xepapadaki P, Zuberbier T. Viruses and bacteria in acute asthma exacerbations--a GA(2) LEN-DARE systematic review. *Allergy* 2010; 66: 458-468.
- 388. Gern JE. The ABCs of rhinoviruses, wheezing, and asthma. *J Virol* 2010; 84: 7418-7426.
- 389. Edwards MR, Bartlett NW, Hussell T, Openshaw P, Johnston SL. The microbiology of asthma. *Nat Rev Microbiol* 2012; 10: 459-471.
- 390. Schoggins JW, Wilson SJ, Panis M, Murphy MY, Jones CT, Bieniasz P, Rice CM. A diverse range of gene products are effectors of the type I interferon antiviral response. *Nature* 2011; 472: 481-485.
- 391. Uller L, Leino M, Bedke N, Sammut D, Green B, Lau L, Howarth PH, Holgate ST, Davies DE. Double-stranded RNA induces disproportionate expression of thymic stromal lymphopoietin versus interferon-beta in bronchial epithelial cells from donors with asthma. *Thorax* 2010; 65: 626-632.
- 392. Collison A, Hatchwell L, Verrills N, Wark PA, de Siqueira AP, Tooze M, Carpenter H, Don AS, Morris JC, Zimmermann N, Bartlett NW, Rothenberg ME, Johnston SL, Foster PS, Mattes J. The E3 ubiquitin ligase midline 1 promotes allergen and rhinovirus-induced asthma by inhibiting protein phosphatase 2A activity. *Nat Med* 2013; 19: 232-237.
- 393. Sykes A, Edwards MR, Macintyre J, Del Rosario A, Bakhsoliani E, Trujillo-Torralbo MB, Kon OM, Mallia P, McHale M, Johnston SL. Rhinovirus 16induced IFN-alpha and IFN-beta are deficient in bronchoalveolar lavage cells in asthmatic patients. *J Allergy Clin Immunol* 2012; 129: 1506-1514 e1506.
- 394. Bufe A, Gehlhar K, Grage-Griebenow E, Ernst M. Atopic phenotype in children is associated with decreased virus-induced interferon-alpha release. *Int Arch Allergy Immunol* 2002; 127: 82-88.
- 395. Gill MA, Bajwa G, George TA, Dong CC, Dougherty, II, Jiang N, Gan VN, Gruchalla RS. Counterregulation between the FcepsilonRI pathway and antiviral responses in human plasmacytoid dendritic cells. *J Immunol* 2010; 184: 5999-6006.
- 396. Durrani SR, Montville DJ, Pratt AS, Sahu S, Devries MK, Rajamanickam V, Gangnon RE, Gill MA, Gern JE, Lemanske RF, Jr., Jackson DJ. Innate immune responses to rhinovirus are reduced by the high-affinity IgE receptor in allergic asthmatic children. *J Allergy Clin Immunol* 2012; 130: 489-495.
- 397. Sykes A, Macintyre J, Edwards MR, Del Rosario A, Haas J, Gielen V, Kon OM, McHale M, Johnston SL. Rhinovirus-induced interferon production is not deficient in well controlled asthma. *Thorax* 2014; 69: 240-246.
- 398. Sykes A, Edwards MR, Macintyre J, Del Rosario A, Gielen V, Haas J, Kon OM, McHale M, Johnston SL. TLR3, TLR4 and TLRs7-9 Induced Interferons Are Not Impaired in Airway and Blood Cells in Well Controlled Asthma. *PLoS One* 2013; 8: e65921.
- 399. Gielen V, Sykes A, Zhu J, Chan B, Macintyre J, Regamey N, Kieninger E, Gupta A, Shoemark A, Bossley C, Davies J, Saglani S, Walker P, Nicholson SE, Dalpke AH, Kon OM, Bush A, Johnston SL, Edwards MR. Increased nuclear suppressor of cytokine signaling 1 in asthmatic bronchial epithelium suppresses rhinovirus induction of innate interferons. *J Allergy Clin Immunol* 2015; 136: 177-188 e111.





Version: Final

400. Bedke N, Sammut D, Green B, Kehagia V, Dennison P, Jenkins G, Tatler A, Howarth PH, Holgate ST, Davies DE. Transforming growth factor-Beta promotes rhinovirus replication in bronchial epithelial cells by suppressing the innate immune response. *PLoS One* 2012; 7: e44580.

- 401. Thomas BJ, Lindsay M, Dagher H, Freezer NJ, Li D, Ghildyal R, Bardin PG. Transforming growth factor-beta enhances rhinovirus infection by diminishing early innate responses. *American journal of respiratory cell and molecular biology* 2009; 41: 339-347.
- 402. Roponen M, Yerkovich ST, Hollams E, Sly PD, Holt PG, Upham JW. Toll-like receptor 7 function is reduced in adolescents with asthma. *Eur Respir J* 2010; 35: 64-71.
- 403. Djukanovic R, Harrison T, Johnston SL, Gabbay F, Wark P, Thomson NC, Niven R, Singh D, Reddel HK, Davies DE, Marsden R, Boxall C, Dudley S, Plagnol V, Holgate ST, Monk P. The effect of inhaled IFN-beta on worsening of asthma symptoms caused by viral infections. A randomized trial. *Am J Respir Crit Care Med* 2014; 190: 145-154.
- 404. Koltsida O, Hausding M, Stavropoulos A, Koch S, Tzelepis G, Ubel C, Kotenko SV, Sideras P, Lehr HA, Tepe M, Klucher KM, Doyle SE, Neurath MF, Finotto S, Andreakos E. IL-28A (IFN-lambda2) modulates lung DC function to promote Th1 immune skewing and suppress allergic airway disease. *EMBO Mol Med* 2011; 3: 348-361.
- 405. Pritchard AL, Carroll ML, Burel JG, White OJ, Phipps S, Upham JW. Innate IFNs and Plasmacytoid Dendritic Cells Constrain Th2 Cytokine Responses to Rhinovirus: A Regulatory Mechanism with Relevance to Asthma. *J Immunol* 2012; 188: 5898-5905.
- 406. Gielen V, Johnston SL, Edwards MR. Azithromycin induces anti-viral responses in bronchial epithelial cells. *Eur Respir J* 2010; 36: 646-654.
- 407. Zaccone EJ, Undem BJ. Airway Vagal Neuroplasticity Associated with Respiratory Viral Infections. *Lung* 2016; 194: 25-29.
- 408. Esposito S, Blasi F, Arosio C, Fioravanti L, Fagetti L, Droghetti R, Tarsia P, Allegra L, Principi N. Importance of acute Mycoplasma pneumoniae and Chlamydia pneumoniae infections in children with wheezing. *Eur Respir J* 2000; 16: 1142-1146.
- 409. Cunningham AF, Johnston SL, Julious SA, Lampe FC, Ward ME. Chronic Chlamydia pneumoniae infection and asthma exacerbations in children. *Eur Respir J* 1998; 11: 345-349.
- 410. Johnston SL, Blasi F, Black PN, Martin RJ, Farrell DJ, Nieman RB. The effect of telithromycin in acute exacerbations of asthma. *N Engl J Med* 2006; 354: 1589-1600.
- 411. Pilishvili T, Zell ER, Farley MM, Schaffner W, Lynfield R, Nyquist AC, Vazquez M, Bennett NM, Reingold A, Thomas A, Jackson D, Schuchat A, Whitney CG. Risk factors for invasive pneumococcal disease in children in the era of conjugate vaccine use. *Pediatrics* 2010; 126: e9-17.
- 412. Jounio U, Juvonen R, Bloigu A, Silvennoinen-Kassinen S, Kaijalainen T, Kauma H, Peitso A, Saukkoriipi A, Vainio O, Harju T, Leinonen M. Pneumococcal carriage is more common in asthmatic than in non-asthmatic young men. *Clin Respir J* 2010; 4: 222-229.
- 413. Kaiko GE, Phipps S, Hickey DK, Lam CE, Hansbro PM, Foster PS, Beagley KW. Chlamydia muridarum infection subverts dendritic cell function to promote





© Copyright 2016, EARIP Consortium GA- 602077

- Th2 immunity and airways hyperreactivity. *J Immunol* 2008; 180: 2225-2232.
- 414. Oliver BG, Lim S, Wark P, Laza-Stanca V, King N, Black JL, Burgess JK, Roth M, Johnston SL. Rhinovirus exposure impairs immune responses to bacterial products in human alveolar macrophages. *Thorax* 2008; 63: 519-525.
- 415. Avadhanula V, Rodriguez CA, Devincenzo JP, Wang Y, Webby RJ, Ulett GC, Adderson EE. Respiratory viruses augment the adhesion of bacterial pathogens to respiratory epithelium in a viral species- and cell type-dependent manner. *J Virol* 2006; 80: 1629-1636.
- 416. British Guideline on the Management of Asthma. *Thorax* 2008; 63 Suppl 4: iv1-121.
- 417. Johnston SL, Szigeti M, Cross M, Brightling C, Chaudhuri R, Harrison T, Mansur A, Robison L, Sattar Z, Jackson D, Mallia P, Wong E, Corrigan C, Higgins B, Ind P, Singh D, Thomson NC, Ashby D, Chauhan A, Team AT. Azithromycin for Acute Exacerbations of Asthma: The AZALEA Randomized Clinical Trial. *JAMA internal medicine* 2016.
- 418. Brusselle GG, Van Braeckel E. AZALEA Trial Highlights Antibiotic Overuse in Acute Asthma Attacks. *JAMA internal medicine* 2016.
- 419. Stokholm J, Chawes BL, Vissing NH, Bjarnadottir E, Pedersen TM, Vinding RK, Schoos AM, Wolsk HM, Thorsteinsdottir S, Hallas HW, Arianto L, Schjorring S, Krogfelt KA, Fischer TK, Pipper CB, Bonnelykke K, Bisgaard H. Azithromycin for episodes with asthma-like symptoms in young children aged 1-3 years: a randomised, double-blind, placebo-controlled trial. *Lancet Respir Med* 2016; 4: 19-26.
- 420. Bacharier LB, Guilbert TW, Mauger DT, Boehmer S, Beigelman A, Fitzpatrick AM, Jackson DJ, Baxi SN, Benson M, Burnham CA, Cabana M, Castro M, Chmiel JF, Covar R, Daines M, Gaffin JM, Gentile DA, Holguin F, Israel E, Kelly HW, Lazarus SC, Lemanske RF, Jr., Ly N, Meade K, Morgan W, Moy J, Olin T, Peters SP, Phipatanakul W, Pongracic JA, Raissy HH, Ross K, Sheehan WJ, Sorkness C, Szefler SJ, Teague WG, Thyne S, Martinez FD. Early Administration of Azithromycin and Prevention of Severe Lower Respiratory Tract Illnesses in Preschool Children With a History of Such Illnesses: A Randomized Clinical Trial. Jama 2015; 314: 2034-2044.
- 421. Brusselle GG, Vanderstichele C, Jordens P, Deman R, Slabbynck H, Ringoet V, Verleden G, Demedts IK, Verhamme K, Delporte A, Demeyere B, Claeys G, Boelens J, Padalko E, Verschakelen J, Van Maele G, Deschepper E, Joos GF. Azithromycin for prevention of exacerbations in severe asthma (AZISAST): a multicentre randomised double-blind placebo-controlled trial. *Thorax* 2013; 68: 322-329.
- 422. Ichinohe T, Pang IK, Kumamoto Y, Peaper DR, Ho JH, Murray TS, Iwasaki A. Microbiota regulates immune defense against respiratory tract influenza A virus infection. *Proc Natl Acad Sci U S A* 2011; 108: 5354-5359.
- 423. Slater M, Rivett DW, Williams L, Martin M, Harrison T, Sayers I, Bruce KD, Shaw D. The impact of azithromycin therapy on the airway microbiota in asthma. *Thorax* 2014; 69: 673-674.
- 424. Chishimba L, Langridge P, Powell G, Niven RM, Denning DW. Efficacy and safety of nebulised amphotericin B (NAB) in severe asthma with fungal sensitisation (SAFS) and allergic bronchopulmonary aspergillosis (ABPA). *J Asthma* 2015; 52: 289-295.





359-363.

Version: Final

425. O'Hollaren MT, Yunginger JW, Offord KP, Somers MJ, O'Connell EJ, Ballard DJ, Sachs MI. Exposure to an aeroallergen as a possible precipitating factor in respiratory arrest in young patients with asthma. *N Engl J Med* 1991; 324:

- 426. Pulimood TB, Corden JM, Bryden C, Sharples L, Nasser SM. Epidemic asthma and the role of the fungal mold Alternaria alternata. *J Allergy Clin Immunol* 2007; 120: 610-617.
- 427. Zheng XY, Ding H, Jiang LN, Chen SW, Zheng JP, Qiu M, Zhou YX, Chen Q, Guan WJ. Association between Air Pollutants and Asthma Emergency Room Visits and Hospital Admissions in Time Series Studies: A Systematic Review and Meta-Analysis. *PLoS One* 2015; 10: e0138146.
- 428. Andersen ZJ, Bonnelykke K, Hvidberg M, Jensen SS, Ketzel M, Loft S, Sorensen M, Tjonneland A, Overvad K, Raaschou-Nielsen O. Long-term exposure to air pollution and asthma hospitalisations in older adults: a cohort study. *Thorax* 2012; 67: 6-11.
- 429. Esposito S, Tenconi R, Lelii M, Preti V, Nazzari E, Consolo S, Patria MF. Possible molecular mechanisms linking air pollution and asthma in children. *BMC pulmonary medicine* 2014; 14: 31.
- 430. Kim J, Kim H, Kweon J. Hourly differences in air pollution on the risk of asthma exacerbation. *Environmental pollution* 2015; 203: 15-21.
- 431. Cai J, Zhao A, Zhao J, Chen R, Wang W, Ha S, Xu X, Kan H. Acute effects of air pollution on asthma hospitalization in Shanghai, China. *Environmental pollution* 2014; 191: 139-144.
- 432. Wang IJ, Tung TH, Tang CS, Zhao ZH. Allergens, air pollutants, and childhood allergic diseases. *Int J Hyg Environ Health* 2016; 219: 66-71.
- 433. Peel JL, Tolbert PE, Klein M, Metzger KB, Flanders WD, Todd K, Mulholland JA, Ryan PB, Frumkin H. Ambient air pollution and respiratory emergency department visits. *Epidemiology* 2005; 16: 164-174.
- 434. Huang SK, Zhang Q, Qiu Z, Chung KF. Mechanistic impact of outdoor air pollution on asthma and allergic diseases. *Journal of thoracic disease* 2015; 7: 23-33.
- 435. Schatz M, Zeiger RS, Zhang F, Chen W, Yang SJ, Camargo CA. Overweight/obesity and risk of seasonal asthma exacerbations. *J Allergy Clin Immunol Pract* 2013; 1: 618-622.
- 436. Jackson DJ, Sykes A, Mallia P, Johnston SL. Asthma exacerbations: origin, effect, and prevention. *J Allergy Clin Immunol* 2011; 128: 1165-1174.
- 437. Kwon HL, Belanger K, Bracken MB. Asthma prevalence among pregnant and childbearing-aged women in the United States: estimates from national health surveys. *Ann Epidemiol* 2003; 13: 317-324.
- 438. Forbes RL, Gibson PG, Murphy VE, Wark PA. Impaired type I and III interferon response to rhinovirus infection during pregnancy and asthma. *Thorax* 2012; 67: 209-214.
- 439. Forbes RL, Wark PA, Murphy VE, Gibson PG. Pregnant women have attenuated innate interferon responses to 2009 pandemic influenza A virus subtype H1N1. *The Journal of infectious diseases* 2012; 206: 646-653.
- 440. Morita M, Kuba K, Ichikawa A, Nakayama M, Katahira J, Iwamoto R, Watanebe T, Sakabe S, Daidoji T, Nakamura S, Kadowaki A, Ohto T, Nakanishi H, Taguchi R, Nakaya T, Murakami M, Yoneda Y, Arai H, Kawaoka Y, Penninger JM, Arita M, Imai Y. The lipid mediator protectin D1 inhibits influenza virus replication and improves severe influenza. *Cell* 2013; 153: 112-125.





GA- 602077

- 441. Tam VC, Quehenberger O, Oshansky CM, Suen R, Armando AM, Treuting PM, Thomas PG, Dennis EA, Aderem A. Lipidomic profiling of influenza infection identifies mediators that induce and resolve inflammation. *Cell* 2013; 154: 213-227.
- 442. Chiang N, Fredman G, Backhed F, Oh SF, Vickery T, Schmidt BA, Serhan CN. Infection regulates pro-resolving mediators that lower antibiotic requirements. *Nature* 2012; 484: 524-528.
- 443. Wainwright NW, Surtees PG, Wareham NJ, Harrison BD. Psychosocial factors and incident asthma hospital admissions in the EPIC-Norfolk cohort study. *Allergy* 2007; 62: 554-560.
- 444. Sandberg S, Paton JY, Ahola S, McCann DC, McGuinness D, Hillary CR, Oja H. The role of acute and chronic stress in asthma attacks in children. *Lancet* 2000; 356: 982-987.
- 445. Barton CA, McKenzie DP, Walters EH, Abramson MJ, Victorian Asthma Mortality Study G. Interactions between psychosocial problems and management of asthma: who is at risk of dying? *J Asthma* 2005; 42: 249-256.
- 446. Patel M, Pilcher J, Hancox RJ, Sheahan D, Pritchard A, Braithwaite I, Shaw D, Black P, Weatherall M, Beasley R, Group SS. The use of beta2-agonist therapy before hospital attendance for severe asthma exacerbations: a post-hoc analysis. NPJ primary care respiratory medicine 2015; 25: 14099.
- 447. Smith JR, Noble MJ, Musgrave S, Murdoch J, Price GM, Barton GR, Windley J, Holland R, Harrison BD, Howe A, Price DB, Harvey I, Wilson AM. The at-risk registers in severe asthma (ARRISA) study: a cluster-randomised controlled trial examining effectiveness and costs in primary care. *Thorax* 2012; 67: 1052-1060.
- 448. Price D, Wilson AM, Chisholm A, Rigazio A, Burden A, Thomas M, King C. Predicting frequent asthma exacerbations using blood eosinophil count and other patient data routinely available in clinical practice. *Journal of asthma and allergy* 2016; 9: 1-12.
- 449. Dhariwal J, Edwards MR, Johnston SL. Anti-viral agents: potential utility in exacerbations of asthma. *Curr Opin Pharmacol* 2013; 13: 331-336.
- 450. Hayden FG, Coats T, Kim K, Hassman HA, Blatter MM, Zhang B, Liu S. Oral pleconaril treatment of picornavirus-associated viral respiratory illness in adults: efficacy and tolerability in phase II clinical trials. *Antivir Ther* 2002; 7: 53-65.
- 451. Traub S, Nikonova A, Carruthers A, Dunmore R, Vousden KA, Gogsadze L, Hao W, Zhu Q, Bernard K, Zhu J, Dymond M, McLean GR, Walton RP, Glanville N, Humbles A, Khaitov M, Wells T, Kolbeck R, Leishman AJ, Sleeman MA, Bartlett NW, Johnston SL. An anti-human ICAM-1 antibody inhibits rhinovirus-induced exacerbations of lung inflammation. *PLoS Pathog* 2013; 9: e1003520.
- 452. Bitko V, Musiyenko A, Shulyayeva O, Barik S. Inhibition of respiratory viruses by nasally administered siRNA. *Nat Med* 2005; 11: 50-55.
- 453. DeVincenzo J, Lambkin-Williams R, Wilkinson T, Cehelsky J, Nochur S, Walsh E, Meyers R, Gollob J, Vaishnaw A. A randomized, double-blind, placebo-controlled study of an RNAi-based therapy directed against respiratory syncytial virus. *Proc Natl Acad Sci U S A* 2010; 107: 8800-8805.
- 454. Moore TW, Sana K, Yan D, Krumm SA, Thepchatri P, Snyder JP, Marengo J, Arrendale RF, Prussia AJ, Natchus MG, Liotta DC, Plemper RK, Sun A.





Version: Final

Synthesis and Metabolic Studies of Host Directed Inhibitors for Anti Viral Therapy. ACS Med Chem Lett 2013; 4: 762-767.

- 455. Krumm SA, Ndungu JM, Yoon JJ, Dochow M, Sun A, Natchus M, Snyder JP, Plemper RK. Potent host-directed small-molecule inhibitors of myxovirus RNA-dependent RNA-polymerases. *PLoS One* 2011; 6: e20069.
- 456. Palmenberg AC, Spiro D, Kuzmickas R, Wang S, Djikeng A, Rathe JA, Fraser-Liggett CM, Liggett SB. Sequencing and analyses of all known human rhinovirus genomes reveal structure and evolution. *Science* 2009; 324: 55-59.
- 457. Bochkov YA, Palmenberg AC, Lee WM, Rathe JA, Amineva SP, Sun X, Pasic TR, Jarjour NN, Liggett SB, Gern JE. Molecular modeling, organ culture and reverse genetics for a newly identified human rhinovirus C. *Nat Med* 2011; 17: 627-632.
- 458. Bizzintino J, Lee WM, Laing IA, Vang F, Pappas T, Zhang G, Martin AC, Khoo SK, Cox DW, Geelhoed GC, McMinn PC, Goldblatt J, Gern JE, Le Souef PN. Association between human rhinovirus C and severity of acute asthma in children. *Eur Respir J* 2011; 37: 1037-1042.
- 459. Calvo C, Casas I, Garcia-Garcia ML, Pozo F, Reyes N, Cruz N, Garcia-Cuenllas L, Perez-Brena P. Role of rhinovirus C respiratory infections in sick and healthy children in Spain. *Pediatr Infect Dis J* 2010; 29: 717-720.
- 460. Miller EK, Edwards KM, Weinberg GA, Iwane MK, Griffin MR, Hall CB, Zhu Y, Szilagyi PG, Morin LL, Heil LH, Lu X, Williams JV. A novel group of rhinoviruses is associated with asthma hospitalizations. *J Allergy Clin Immunol* 2009; 123: 98-104 e101.
- 461. Bernstein DI, Malkin E, Abughali N, Falloon J, Yi T, Dubovsky F. Phase 1 study of the safety and immunogenicity of a live, attenuated respiratory syncytial virus and parainfluenza virus type 3 vaccine in seronegative children. *Pediatr Infect Dis J* 2011; 31: 109-114.
- 462. Glenn GM, Smith G, Fries L, Raghunandan R, Lu H, Zhou B, Thomas DN, Hickman SP, Kpamegan E, Boddapati S, Piedra PA. Safety and immunogenicity of a Sf9 insect cell-derived respiratory syncytial virus fusion protein nanoparticle vaccine. *Vaccine* 2012; 31: 524-532.
- 463. Malkin E, Yogev R, Abughali N, Sliman J, Wang CK, Zuo F, Yang CF, Eickhoff M, Esser MT, Tang RS, Dubovsky F. Safety and immunogenicity of a live attenuated RSV vaccine in healthy RSV-seronegative children 5 to 24 months of age. *PLoS One* 2013; 8: e77104.
- 464. Glanville N, McLean GR, Guy B, Lecouturier V, Berry C, Girerd Y, Gregoire C, Walton RP, Pearson RM, Kebadze T, Burdin N, Bartlett NW, Almond JW, Johnston SL. Cross-serotype immunity induced by immunization with a conserved rhinovirus capsid protein. *PLoS Pathog* 2013; 9: e1003669.
- 465. McLean GR, Walton RP, Shetty S, Peel TJ, Paktiawal N, Kebadze T, Gogsadze L, Niespodziana K, Valenta R, Bartlett NW, Johnston SL. Rhinovirus infections and immunisation induce cross-serotype reactive antibodies to VP1. Antiviral research 2012; 95: 193-201.
- 466. WHO. Vaccines against influenza WHO position paper November 2012. Releve epidemiologique hebdomadaire / Section d'hygiene du Secretariat de la Societe des Nations = Weekly epidemiological record / Health Section of the Secretariat of the League of Nations 2012; 87: 461-476.
- 467. ACIP. Prevention and control of seasonal influenza with vaccines. Recommendations of the Advisory Committee on Immunization Practices-





GA- 602077

- United States, 2013-2014. MMWR Recommendations and reports: Morbidity and mortality weekly report Recommendations and reports / Centers for Disease Control 2013; 62: 1-43.
- 468. Cates CJ, Rowe BH. Vaccines for preventing influenza in people with asthma. *Cochrane Database Syst Rev* 2013; 2: CD000364.
- 469. Fiers W, De Filette M, El Bakkouri K, Schepens B, Roose K, Schotsaert M, Birkett A, Saelens X. M2e-based universal influenza A vaccine. *Vaccine* 2009; 27: 6280-6283.
- 470. Nachbagauer R, Wohlbold TJ, Hirsh A, Hai R, Sjursen H, Palese P, Cox RJ, Krammer F. Induction of broadly reactive anti-hemagglutinin stalk antibodies by an H5N1 vaccine in humans. *J Virol* 2014; 88: 13260-13268.
- 471. Jarjour NN, Erzurum SC, Bleecker ER, Calhoun WJ, Castro M, Comhair SA, Chung KF, Curran-Everett D, Dweik RA, Fain SB, Fitzpatrick AM, Gaston BM, Israel E, Hastie A, Hoffman EA, Holguin F, Levy BD, Meyers DA, Moore WC, Peters SP, Sorkness RL, Teague WG, Wenzel SE, Busse WW, Program NSAR. Severe asthma: lessons learned from the National Heart, Lung, and Blood Institute Severe Asthma Research Program. Am J Respir Crit Care Med 2012; 185: 356-362.
- 472. Kupczyk M, ten Brinke A, Sterk PJ, Bel EH, Papi A, Chanez P, Nizankowska-Mogilnicka E, Gjomarkaj M, Gaga M, Brusselle G, Dahlen B, Dahlen SE, investigators B. Frequent exacerbators--a distinct phenotype of severe asthma. Clin Exp Allergy 2014; 44: 212-221.
- 473. Kupczyk M, Haque S, Sterk PJ, Nizankowska-Mogilnicka E, Papi A, Bel EH, Chanez P, Dahlen B, Gaga M, Gjomarkaj M, Howarth PH, Johnston SL, Joos GF, Kanniess F, Tzortzaki E, James A, Middelveld RJ, Dahlen SE, investigators B. Detection of exacerbations in asthma based on electronic diary data: results from the 1-year prospective BIOAIR study. *Thorax* 2013; 68: 611-618.
- 474. Wheelock CE, Goss VM, Balgoma D, Nicholas B, Brandsma J, Skipp PJ, Snowden S, Burg D, D'Amico A, Horvath I, Chaiboonchoe A, Ahmed H, Ballereau S, Rossios C, Chung KF, Montuschi P, Fowler SJ, Adcock IM, Postle AD, Dahlen SE, Rowe A, Sterk PJ, Auffray C, Djukanovic R, Group UBS. Application of 'omics technologies to biomarker discovery in inflammatory lung diseases. *Eur Respir J* 2013; 42: 802-825.
- 475. Rochlitzer S, Hoymann HG, Muller M, Braun A, consortium UB. No exacerbation but impaired anti-viral mechanisms in a rhinovirus-chronic allergic asthma mouse model. *Clinical science* 2014; 126: 55-65.
- 476. Wardzynska A, Kubsik B, Kowalski ML. Comorbidities in elderly patients with asthma: Association with control of the disease and concomitant treatment. *Geriatrics & gerontology international* 2015; 15: 902-909.
- 477. Milanese M, Di Marco F, Corsico AG, Rolla G, Sposato B, Chieco-Bianchi F, Costantino MT, Crivellaro MA, Guarnieri G, Scichilone N, Group ES. Asthma control in elderly asthmatics. An Italian observational study. *Respir Med* 2014; 108: 1091-1099.
- 478. Park HW, Song WJ, Kim SH, Park HK, Kim SH, Kwon YE, Kwon HS, Kim TB, Chang YS, Cho YS, Lee BJ, Jee YK, Jang AS, Nahm DH, Park JW, Yoon HJ, Cho YJ, Choi BW, Moon HB, Cho SH. Classification and implementation of asthma phenotypes in elderly patients. *Ann Allergy Asthma Immunol* 2015; 114: 18-22.



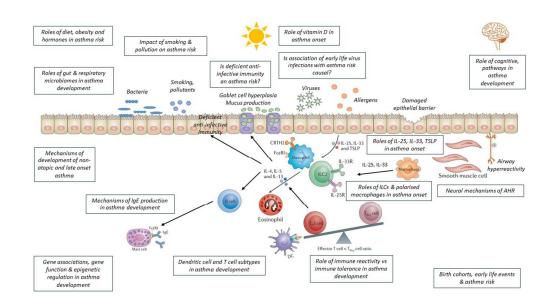


GA- 602077

- 479. Doshi V, Shenoy S, Ganesh A, Rishi MA, Molnar J, Henkle J. Profile of Acute Asthma Exacerbation in Drug Users. *American journal of therapeutics* 2014.
- 480. Manthei DM, Schwantes EA, Mathur SK, Guadarrama AG, Kelly EA, Gern JE, Jarjour NN, Denlinger LC. Nasal lavage VEGF and TNF-alpha levels during a natural cold predict asthma exacerbations. *Clin Exp Allergy* 2014; 44: 1484-1493.
- 481. Petsky HL, Li AM, Au CT, Kynaston JA, Turner C, Chang AB. Management based on exhaled nitric oxide levels adjusted for atopy reduces asthma exacerbations in children: A dual centre randomized controlled trial. *Pediatr Pulmonol* 2015; 50: 535-543.
- 482. Long W, Li LJ, Huang GZ, Zhang XM, Zhang YC, Tang JG, Zhang Y, Lu G. Procalcitonin guidance for reduction of antibiotic use in patients hospitalized with severe acute exacerbations of asthma: a randomized controlled study with 12-month follow-up. *Critical care* 2014; 18: 471.
- 483. Green RH, Brightling CE, McKenna S, Hargadon B, Parker D, Bradding P, Wardlaw AJ, Pavord ID. Asthma exacerbations and sputum eosinophil counts: a randomised controlled trial. *Lancet* 2002; 360: 1715-1721.
- 484. Huber JP, Ramos HJ, Gill MA, Farrar JD. Cutting edge: Type I IFN reverses human Th2 commitment and stability by suppressing GATA3. *J Immunol* 2010; 185: 813-817.
- 485. Romieu I, Sienra-Monge JJ, Ramírez-Aguilar M, Moreno-Macías H, Reyes-Ruiz NI, Estela del Río-Navarro B, Hernández-Avila M, London SJ. Genetic polymorphism of GSTM1 and antioxidant supplementation influence lung function in relation to ozone exposure in asthmatic children in Mexico City. *Thorax* 2004; 59: 8-10.
- 486. Gilliland FD, Li YF, Saxon A, Diaz-Sanchez D. Effect of glutathione-Stransferase M1 and P1 genotypes on xenobiotic enhancement of allergic responses: randomised, placebo-controlled crossover study. *Lancet* 2004; 363: 119-125.
- 487. Caponnetto P, Auditore R, Russo C, Alamo A, Campagna D, Demma S, Polosa R. "Dangerous relationships": asthma and substance abuse. *Journal of addictive diseases* 2013; 32: 158-167.

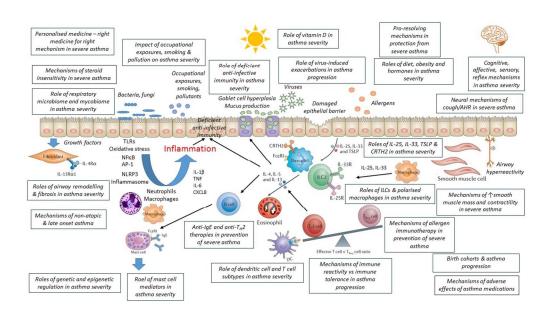






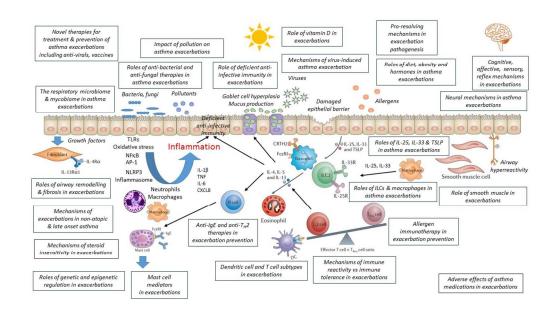
Major mechanisms of asthma onset. The major cell types, cytokines and mechanisms involved in asthma onset are depicted, with major mechanisms/research areas requiring substantial further investment highlighted in boxes

108x60mm (300 x 300 DPI)



Major mechanisms of asthma progression and severity. The major cell types, cytokines and mechanisms involved in asthma progression are depicted, with major mechanisms/research areas requiring substantial further investment highlighted in boxes.

108x60mm (300 x 300 DPI)



Major mechanisms of asthma exacerbations. The major cell types, cytokines and mechanisms involved in asthma exacerbations are depicted, with major mechanisms/research areas requiring substantial further investment highlighted in boxes.

108x60mm (300 x 300 DPI)

Appendix

The European Asthma Research and Innovation Partnership (EARIP) was funded under the European Union in the 7th Framework Programme, grant agreement 602077.

In addition to those listed as authors on the title page, the following people were members of the EARIP WP2 working group and contributed to the writing and revision of this manuscript:

lan Adcock	Imperial College London, London, UK
Cezmi Akdis	University of Zurich Medical Faculty, Zurich,
	Switzerland
Elisabeth Bel Jean Bousquet	Department of Respiratory Medicine,
	Academic Medical Centre, University of
	Amsterdam, Amsterdam, Netherlands
	University Hospital, Montpellier, France;
	MACVIA-LR, Centre les Maladies Chroniques
	pour un Vieillissement Actif en Languedoc-
	Roussillon, EIP-AHA Reference Site, France
Anneke ten Brinke	Department of Pulmonary Diseases, Medical
	Center Leeuwarden, Leeuwarden, The
	Netherlands
Guy Brusselle	Department of Respiratory Medicine,
	Laboratory for Translational Research in
	Obstructive Pulmonary Diseases, Ghent
	University Hospital, Ghent, Belgium
Sven-Erik Dáhlen	Experimental Asthma and Allergy Research,
	Institute of Environmental Medicine,
	Karolinska Institutet, Stockholm, Sweden
Ratko Djukanovic	Southampton University Hospitals Trust,
	United Kingdom
Mina Gaga	7th Respiratory Medicine Dept. and Asthma
	Center, Athens Chest Hospital "Sotiria",
	Athens, Greece
Luis Garcia-Marcos	Arrixaca University Children's Hospital.
	University of Murcia, Spain
Pieter Hiemstra	Department of Pulmonology, Leiden
	University Medical Center, Leiden,
	Netherlands
Enrico Heffler	Unit of Respiratory Medicine and
	Allergology, Department of Clinical and
	Experimental Medicine, Azienda
	Ospedaliero Universitaria Policlinico-
	Vittorio Emanuele Hospital, University of
	Catania, Catania, Italy
Stephen Holgate	Southampton General Hospital,
	Southampton, UK
Maciek Kupczyk	Medical University of Łódź, Poland
Clare Lloyd	Imperial College London, London, UK
Jane Lucas	Clinical and Experimental Sciences, Faculty
	of Medicine, University of Southampton,
	Southampton, United Kingdom

Antoine Magnan	Service de pneumologie, L'institut du
	thorax, CHU Nantes, Nantes, France
Thomas Martin	Novartis Pharmaceuticals, USA
David Myles	GlaxoSmithKline, United Kingdom
Clive Page	Sackler Institute of Pulmonary Pharmacology, King's College London UK
Susanna Palkonen	EFA, Belgium
Nikos Papadopoulos	University of Manchester, United Kingdom
Alberto Papi	University of Ferrara
Pippa Powell	European Lung Foundation, United Kingdom
John Riley	GlaxoSmithKline, United Kingdom
lan Sayers	Division of Respiratory Medicine, University of Nottingham, Nottingham, United Kingdom
Antonio Spanevello	Pneumology Unit, Fondazione Maugeri, IRCCS, Tradale, Italy; Department of Clinical and Experimental Medicine, University of Insubria, Varese, Italy
Peter Sterk	University of Groningen, Department of Respiratory Medicine, Amsterdam, Academic Medical Centre, University of Amsterdam, Netherlands
Paraskevi Xepapadaki	Allergy Department, 2nd Pediatric Clinic, National and Kapodistrian University of Athens, Greece
Craig Wheelock	Department of Medical Biochemistry and Biophysics, Division of Physiological Chemistry 2, Karolinska Institutet, SE- 17177, Stockholm, Sweden