

## ARTHROPOD STRUCTURE AND DEVELOPMENT – SPECIAL VOLUME

**A defensive behavior and plant-insect interaction in Early Cretaceous amber – the case of the immature lacewing *Hallucinochrysa diogenesi***Ricardo Pérez-de la Fuente<sup>a,\*</sup>, Xavier Delclòs<sup>b</sup>, Enrique Peñalver<sup>c</sup>, Michael S. Engel<sup>d</sup><sup>a</sup> Department of Organismic and Evolutionary Biology, Museum of Comparative Zoology, Harvard University, 26 Oxford Street, Cambridge, MA 02138, United States<sup>b</sup> Departament d'Estratigrafia, Paleontologia i Geociències Marines, Facultat de Geologia, Universitat de Barcelona, E-08071 Barcelona, Spain<sup>c</sup> Museo Geominero, Instituto Geológico y Minero de España, E-28003 Madrid, Spain<sup>d</sup> Division of Entomology, Natural History Museum, and Department of Ecology and Evolutionary Biology, University of Kansas, Lawrence, KS 66045, United States**ABSTRACT**

Amber holds special paleobiological significance due to its ability to preserve direct evidence of biotic interactions and animal behaviors for millions of years. Here we review the finding of *Hallucinochrysa diogenesi* Pérez-de la Fuente, Delclòs, Peñalver and Engel, 2012, a morphologically atypical larva related to modern green lacewings (Insecta: Neuroptera) that was described in Early Cretaceous amber from the El Soplao outcrop (northern Spain). The fossil larva is preserved with a dense cloud of fern trichomes that corresponds to the trash packet the insect gathered and carried on its back for camouflaging and shielding, similar to that which is done by its extant relatives. This finding supports the prominent role of wildfires in the paleoecosystem and provides direct evidence of both an ancient plant-insect interaction and an early acquisition of a defensive behavior in an insect lineage. Overall, the fossil of *H. diogenesi* showcases the potential that the amber record offers to reconstruct not only the morphology of fossil arthropods but, more remarkably, their lifestyles and ecological relationships.

*Keywords:* fossil; reconstruction; insect; larva; Neuroptera

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**1. Introduction**

Amber – fossil resin – is a unique source of paleontological data on terrestrial arthropods and their environments, offering the most exquisite fossil preservation in deep time (Grimaldi and Engel, 2005; Labandeira, 2014). The resin often was able to protect small organisms (principally arthropods) from decay and disarticulation shortly after they became entombed, particularly as such resins have antiseptic and antimicrobial properties that hamper decomposition. Additionally, tissues of resin-included organisms dehydrate soon after they are trapped, leading to a process of mummification that normally ensures the fidelity of the fossilized structures down to the ultrastructural level (Martínez-Delclòs et al., 2004). Although most amber inclusions are preserved only as cuticle due to autolysis of internal tissues by endogenous bacteria (ibid.), soft tissues can also be preserved (e.g., Henwood, 1992; Pohl et al., 2008; Soriano et al., 2010; Perreau and Tafforeau, 2011; Labandeira, 2014). Moreover, resin entrapment and infilling, either partially or completely, protects inclusions from collapsing, preserving their three-dimensional structure. As a result of this exceptional morphological preservation, reliable inferences about the likely habits of the ancient organisms and the environments in which they lived can be extracted. Furthermore, the study of amber physicochemical properties provides information on its botanical source and the environment in which the resin was secreted, allowing for accurate assessment of the processes that permitted fossilization. Nevertheless, what makes the amber record most remarkable is

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4 its potential to preserve direct evidence of biotic interactions and complex behaviors for millions of years.  
5 This results from the ability of resin to encapsulate “small fragments” of the ecosystem in a relatively fast  
6 and unaltered way – in this sense, each amber piece can be considered a snapshot of the past. This ability  
7 is what distinguishes amber from other fossiliferous materials with a stability measured in millions of  
8 years, even others of exceptional preservation, such as some shales. A plethora of paleobiotic interactions,  
9 both intra- and interspecific, are known from the amber record (for a review, see Arillo, 2007). Among  
10 intraspecific interactions, there are cases of “frozen” reproductive behavior, such as mating and brood  
11 care, and social behaviors, such as cooperative feeding in social insects or gregarious habits (e.g.,  
12 Weitschat and Wichard, 1998; Grimaldi and Engel, 2005; Engel, 2009). Among the intraspecific  
13 interactions, direct evidence of predation, parasitism, mutualism, or defensive behaviors are known (e.g.,  
14 Janzen, 2002; Engel, 2005; Peñalver et al., 2012).

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16 Although the oldest amber with arthropod inclusions comes from the Triassic of Italy (Schmidt et al.,  
17 2012), fossil resin with abundant biological content is not known until the Early Cretaceous. From that  
18 epoch, the most significant ambers in terms of their paleobiotic diversity and abundance are found in  
19 –from west to east– Spain, France, Lebanon, and Myanmar (see review in Penney, 2010). In Spain, more  
20 than a hundred amber-bearing localities, mostly Cretaceous, have been discovered to date (Peñalver and  
21 Delclòs, 2010). From these localities, nine have provided amber with inclusions, although only three have  
22 done so significantly, i.e., the outcrops of Peñacerrada I (Burgos Province; Alonso et al., 2000), San Just  
23 (Teruel Province; Peñalver et al., 2007), and El Soplao (Cantabria Autonomous Community; Najjarro et  
24 al., 2009, 2010). More than 3,700 bioinclusions have been recovered from these three localities, which are  
25 dated to the Albian, ~105 million years ago (Peñalver and Delclòs, 2010; Barrón et al., 2015). In the El  
26 Soplao outcrop, amber is found embedded in sandstones and claystones that are associated with lignite,  
27 similar to the two deposits mentioned above, with accumulations of plant remains and marine to brackish  
28 invertebrates such as gastropods or bryozoans, indicating that the resin was deposited in delta-estuarine  
29 environments. Paleobotanical and chemotaxonomical data suggest that resin from which the El Soplao  
30 amber derives was secreted by conifers belonging to the extinct family Cheirolepidiaceae and, in some  
31 cases, another unidentified botanical source (Menor-Salván et al., 2010).

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33 Several animal lineages possess a behavior by which they actively harvest and carry a wide variety of  
34 organic and/or inorganic remains on their bodies for protective purposes. Although some gastropods and  
35 sea urchins show this habit (Ponder, 1983, 1994; Allgaier, 2007; Dumont et al., 2007), arthropods  
36 exhibiting this behavior also possess associated morphological adaptations aimed at increasing the  
37 potential of retaining the exogenous elements, i.e., the presence of specialized setae, commonly hooked,  
38 and often also cuticular projections of their body dorsum. This is the case of decorating crabs (Hultgren  
39 and Stachowicz, 2011), sand-covering spiders (Duncan et al., 2007), some oribatid mites (Subías, 2012;  
40 Arillo and Subías, 2015 pers. comm.), some ants (Hölldobler and Wilson, 1986), and some immature  
41 forms from a few insect groups, i.e., assassin bugs (Weirauch, 2006), barklice (Betz, 1983; Lienhard,  
42 1988), and lacewings and their relatives (Henry, 1977; Eisner et al., 1978; New, 1982). Among the latter,  
43 in green lacewing larvae, the term “trash-carrying” has been commonly used to refer to the described  
44 behavior.  
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48 Green lacewings (Neuroptera: Chrysopidae) have a nearly cosmopolitan distribution and comprise  
49 around 1,200 described species (Brooks and Barnard, 1990). Their larvae are voracious predators,  
50 generally feeding on a wide variety of prey, which have been used widely as biological control agents  
51 (Canard et al., 1984). They use their sickle-shaped jaws to pierce their prey, inject salivary secretions, and  
52 then absorb the internal fluids and liquefied tissues. Not all green lacewing larvae are trash carriers –  
53 trash-carrying is present in different lacewing lineages, and their related morphological adaptations are  
54 helping elucidate the phylogenetic relationships within the family (Montserrat and Díaz-Aranda, 2012;  
55 Tauber et al., 2014). Trash-carrying larvae, unlike the “naked” morphotypes, tend to have humped bodies  
56 as an adaptation to carry great loads and pairs of setigerous tubercles on their backs for ensnaring the  
57 trash packet elements (McEwen et al., 2001). This trash packet can be composed of single or multiple  
58 types of materials, both animal (prey corpses, exuviae, shells, waxy secretions, etc.) and non-animal  
59 (bark, trichomes, terrestrial algae, lichens, etc.) in origin (Tauber et al., 2014). The trash-packet provides a  
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4 two-fold defense – a camouflage that prevents visual and tactile detection from both predators and prey,  
5 and a physical shield that can be actively moved towards the predator in instances where the larva is  
6 attacked (McEwen et al., 2001; Tauber et al., 2014).

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8 A four-millimeter-long immature green lacewing preserved together with its trash packet was  
9 discovered in the Spanish amber locality of El Soplao (Figs 1, 2, 3, Anim. 1; see above). This discovery is  
10 exceptional not only because fossil immature green lacewings are highly rare in the fossil record, but also  
11 because evidence of trash-carrying (in a broad sense) is also extremely scarce in the record of Arthropoda.  
12 The fossil was described as *Hallucinochrysa diogenesi* Pérez-de la Fuente, Delclòs, Peñalver and Engel,  
13 2012 (Pérez-de la Fuente et al., 2012a), after the mind-boggling aspect of the fossil and the Greek  
14 philosopher Diogenes of Sinope, whose name has been used for the human psychological disorder  
15 characterized by the compulsive accumulation of trash. The present paper reviews this discovery and its  
16 paleobiological implications, showcasing the amount and quality of the paleoecological and  
17 paleoethological information that can be extracted from amber inclusions, and which can be used to  
18 obtain highly reliable reconstructions.  
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## 20 21 **2. Material and methods**

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23 The holotype of *H. diogenesi* (CES 418.1) was isolated within a small piece of transparent amber and  
24 embedded in a regular prism of epoxy resin (EPO-TEK 301) for optimal viewing and curation  
25 (Nascimbene and Silverstein, 2000). The specimen is housed at the laboratory of the institutional El  
26 Soplao collection in El Soplao cave, Celis (Cantabria, northern Spain). The holotype is almost complete,  
27 although the distal parts of both antennae and some tubular tubercles are missing. A large, multi-planed  
28 fracture transversally crosses the larval abdomen, obscuring several tubular tubercles.  
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30 An Olympus BX51 transmitted-light microscope was used to study the insect in dorsal, ventral, and  
31 lateral views. Photography of the specimen used both a ColorView IIIu digital camera attached to an  
32 Olympus BX51 and a Nikon D1X digital camera attached to an Infinity K-2 long-distance microscope  
33 lens. Merged images were obtained with Helicon Focus v.5.3. The three-dimensional reconstruction was  
34 performed with the LightWave 3D computer graphics program (NewTek). Movement patterns, body  
35 coloration, antennal length, and number of abdominal tubular tubercle pairs (see below) were  
36 reconstructed as in modern trash-carrying green lacewing representatives, e.g., *Mallada desjardinsi*  
37 (Navás).  
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## 39 40 **3. Morphofunctional interpretation**

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42 The morphological singularity of *H. diogenesi* compared to known diversity of green lacewing larvae,  
43 both fossil and extant, is remarkable. The most striking differences in the fossil are the highly specialized  
44 adaptations for carrying debris. First, *H. diogenesi*'s thorax and abdomen bear pairs of extremely elongate  
45 setigerous tubercles (= tubular tubercles; see Figs 1, 2, Anim. 1) dorsally, which exceed the body length.  
46 In extant green lacewing trash-carrying larvae, these tubercles are not developed in such an extreme form,  
47 never being more elongate than the body width. Also, the fossil bears two pairs of tubular tubercles on  
48 each thoracic segment (lateral and laterodorsal pairs), and these tubercles are similarly developed as to  
49 those on the abdomen. Conversely, only the laterodorsal thoracic tubercle pairs are well-developed in  
50 other known trash-carrying larvae, in which thoracic tubercles are often much more developed than those  
51 of the abdomen. The second remarkable adaptation of *H. diogenesi* to trash carrying is that the setae on  
52 the tubular tubercles show expanded, trumpet-shaped endings. Although this particular morphology of  
53 setae is unknown from other extinct and extant green lacewings, in which hooked ends and serrated  
54 margins are the most specialized setal adaptations for trash-carrying, it is known to be present in other  
55 neuropteran groups (New, 1982; McEwen et al., 2001). Additionally, and unlike any previously described  
56 green lacewing larvae, the fossil's head is broad, short, and with a strongly convex frons that resembles a  
57 banana (Fig. 3), its three labial palpomeres are not distinctly unequal in length, and the specimen lacks  
58 annulations on the second palpomere. The degree of tubercle development and setation indicates that the  
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4 specimen most likely corresponds to an advanced (third) instar. For a full description of the fossil larva  
5 see Pérez-de la Fuente et al. (2012a).

6 Based on the former characters, *H. diogenesi* was interpreted as a highly specialized, probably  
7 derived, trash-carrying morphotype, rather than representing the basal condition for the lineage that gave  
8 rise to modern green lacewings. For this reason, the fossil was not formally included in this family but left  
9 unclassified at familial level within the superfamily Chrysopoidea Schneider, 1851, which apart from  
10 modern green lacewings includes several extinct lineages known since the Early Jurassic (Nel et al.,  
11 2005). No other immature specimens assignable to any fossil chrysopoid lineage have been previously  
12 reported in the literature, as a few previous records of green lacewing larvae are only known from  
13 Cenozoic amber deposits and clearly fit within the diversity of modern green lacewings (Hagen, 1856;  
14 MacLeod, 1970; Engel and Grimaldi, 2007, 2008; Weitschat, 2009).

15 The dense and complete trash packet gathered by *H. diogenesi* (Figs 1 and 3) is exclusively composed  
16 of nonglandular, multicellular plant trichomes (Fahn, 1990; Evert, 2006), as indicated by their  
17 morphology, size range, ultrastructure, and autofluorescence signals (Pérez-de la Fuente et al., 2012a).  
18 These trichomes are tangled with the specialized setae of the larva's tubular tubercles. Trichomes show an  
19 oval cross-section (up to 30  $\mu\text{m}$  thick), a micropapillate surface, and a thick hyaline layer that corresponds  
20 to their cell wall. They have a branched, dendritic morphology, with a blunt insertion base and an axis  
21 from which several secondary branches alternatively emerge in a single plane. The trichomes show  
22 variation in part attributable to different stages of growth – the axis goes from straight in the less  
23 developed forms to zigzagged in the more mature forms, in which secondary branches are directed  
24 apically and arranged with more space between them (Fig. 4). After an exhaustive literature comparison  
25 (see Pérez-de la Fuente et al., 2012a), the most plausible attribution is that these trichomes belong to the  
26 fern family Gleicheniaceae, a group widespread during the Early Cretaceous (Skog, 2001). Very similar  
27 trichomes were reported in a gleicheniacean described from the Albian of England (Herendeen and Skog,  
28 1998). The family is present in the El Soplao outcrop sediments as two spore species attributed to these  
29 ferns (Najarro et al., 2010; Pérez-de la Fuente et al., 2012a).

30 Green lacewing larvae construct their trash packet using their jaws to gather debris in successive  
31 loads that are placed on the dorsum by bowing the head backwards as the rest of the body arches forwards  
32 (Jones, 1941; Eisner and Silberglied, 1988). Larvae are also capable of reallocating the elements of the  
33 trash-packet using their mouthparts and through peristaltic movements of their body (Eisner et al., 2002).  
34 In the trash packet of *H. diogenesi*, trichomes are often found in clusters with the same orientation,  
35 indicating that they were pulled out from the plant together by the larva. The extreme elongation of *H.*  
36 *diogenesi*'s tubercles suggests that it had its own stereotyped movements for constructing the trash  
37 packet. Trichomes were loaded into a dorsal basket shaped by the tube-like tubercles of the larva (Fig. 2,  
38 Anim. 1), which due to its extreme elongation would have allowed the construction of a thicker trash-  
39 packet, perhaps as a defense against a particular group of Cretaceous predators with an elongate piercing  
40 or sucking proboscis, such as true bugs, or elongate ovipositors, such as parasitoid wasps (Grimaldi and  
41 Engel, 2005). The trumpet-shaped endings of the tubercle setae acted as anchoring points among surfaces  
42 of trichomes, and the particular branched morphology of the trichomes added cohesion to the trash  
43 packet.  
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#### 49 50 **4. Paleobiological implications**

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52 This finding provides direct evidence that trash-carrying behavior, understood as actively harvesting  
53 and carrying exogenous materials for the purposes of camouflage and physical protection, has remained  
54 in stasis for more than 100 million years in the green lacewing lineage. Not only that, but it represents the  
55 earliest trash-carrying record known to date in the fossil record of arthropods (see Pérez-de la Fuente et  
56 al., 2012a). Previous fossil evidence of this behavior has only been reported from other immature insects  
57 in Cenozoic amber, i.e., in assassin bugs, barklice, and owlflies from Miocene Dominican and Mexican  
58 ambers (Wu, 1996; Engel and Grimaldi, 2007; Boucot and Poinar Jr., 2010), and in green lacewings from  
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4 Eocene Baltic amber (Weitschat, 2009). A similar behavior is present in case-building insect larvae like  
5 caddisflies (indirect camouflage), with records known since the Jurassic (Grimaldi and Engel, 2005).

6 *Hallucinochrysa diogenesi* and its trichome-based trash-packet showcase an ancient plant-insect  
7 interaction, in which the structures used by a fern to protect itself from herbivores (trichomes) are used by  
8 a predator to protect itself and provide camouflage from both its predators and prey. As the lacewing larva  
9 fed on the fern's herbivores, the fern could have invested a part of its defensive structures in the lacewing.  
10 For this reason, it seems reasonable to consider that both the fern and the insect obtained a mutual benefit  
11 from their interaction. Further evidence, however, is required to regard the described plant-insect  
12 relationship as a result of a symbiotic interaction, even if the relationship seems to have been specific to  
13 this fern: *H. diogenesi*'s trash packet is solely composed of the described trichomes, suggesting that the  
14 larvae might have exclusively fed on herbivores living in these ferns. In this regard, the predatory habits  
15 of these immature insects can be delimited. First, although the prey range for extant green lacewing larvae  
16 covers a high diversity of arachnid and insect groups (McEwen et al., 2001), their most common prey are  
17 sternorrhynchan hemipterans such as aphids, coccoids, and psyllids. Aphids, however, were scarce in the  
18 paleoenvironment in comparison with coccoids, and their low relative abundance has been attributed to a  
19 low climatic seasonality during the Cretaceous (Peñalver and Wegierek, 2008). Second, insects associated  
20 with ferns today are mainly hemipterans, beetles, and lepidopterans, with a high proportion of specialist  
21 feeders restricted to ferns (Cooper-Driver, 1978). Third, the broadened cephalic capsule of *H. diogenesi*,  
22 together with its long jaws, suggest specialization or, at least, ability to feed on large prey, as both  
23 cephalic width and jaw length are related to prey size (Tauber et al., 1995).

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27 Lastly, evidence exists that indicates fires were important agents controlling the dynamics of the  
28 Iberian Cretaceous amber forest. Charcoal has been copiously found in the sediments associated with El  
29 Soplao amber (and other coetaneous Spanish deposits), and charcoalified plant fibers were discovered as  
30 inclusions within the amber (Najarro et al., 2010; Pérez-de la Fuente et al., 2012b). Fires induced resin  
31 production by damaging the resin-producing trees (in the same manner by which systemic and pandemic  
32 insect outbreaks similarly induce resin production: McKellar et al., 2011), and promoted resin  
33 accumulation after burning the plant mass due to increased erosion of the soil in which resin was  
34 primarily buried. In that sense, *H. diogenesi* and its trash packet composed by fern trichomes with  
35 gleicheniaceae affinity supports that scenario. Gleicheniacean ferns are today primary succession  
36 pioneers following wildfires or lava flows (Gillison, 1969; Walker and Boneta, 1995; Russell et al., 1998)  
37 and this was most likely the same in the past due to their abundance as charcoalified remains since the  
38 Late Jurassic (Herendeen and Skog, 1998; Collinson, 2002; Van Konijnenburg-Van Cittert, 2002). There  
39 also exists among the paleoentomological record further evidence supporting the presence of paleofires in  
40 the Iberian Cretaceous amber forests (Ortega-Blanco et al., 2008; Pérez-de la Fuente et al., 2012b).

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42 Overall, the evolutionary, behavioral, paleoecological, and paleoenvironmental information that *H.*  
43 *diogenesi* provides represents a nice example of how the extraordinary paleobiological potential that  
44 amber holds can be reliably aimed at reconstructing the past and "give life" to fossils.

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## 57 58 59 **References**

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4 Allgaier, C., 2007. Active camouflage with lichens in a terrestrial snail, *Napaeus (N.) barquini*  
5 (Gastropoda, Pulmonata, Enidae). *Zoolog Sci* 24(9), 869–876.
- 6 Alonso, J., Arillo, A., Barrón, E., Corral, J.C., Grimalt, J., López, J.F., López, R., Martínez-Delclòs, X.,  
7 Ortuño, V., Peñalver, E., Trincão, P.R., 2000. A new fossil resin with biological inclusions in Lower  
8 Cretaceous deposits from Álava (northern Spain, Basque-Cantabrian Basin). *J Paleontol* 74(1), 158–  
9 178.
- 10 Arillo, A., 2007. Paleoethology: fossilized behaviors in amber. *Geol Acta* 5(2), 159–166.
- 11 Barrón, E., Peyrot, D., Rodríguez-López, J.P., Meléndez, N., López del Valle, R., Najarro, M., Rosales, I.,  
12 Comas-Rengifo, M. J., 2015. Palynology of Aptian and upper Albian (Lower Cretaceous) amber-  
13 bearing outcrops of the southern margin of the Basque-Cantabrian basin (northern Spain). *Cretaceous*  
14 *Res* 52, 292–312.
- 15 Betz, B.W., 1983. The biology of *Trichadenotecnum alexandrae* Sommerman (Psocoptera, Psocidae). 1.  
16 Habitat, life stages and events. *Entomol News* 94, 152–158.
- 17 Boucot, A.J., Poinar Jr., G.O., 2010. *Fossil Behaviour Compendium* (CRC, Boca Raton, FL).
- 18 Brooks, S.J., Barnard, P.C., 1990. The green lacewings of the world: A generic review (Neuroptera:  
19 Chrysopidae). *Bull Br Mus Nat Hist Ent* 59(2), 117–286.
- 20 Canard, M., Séméria, Y., New, T.R., 1984. *Biology of Chrysopidae* (Dr. W. Junk Publishers, The Hague).
- 21 Collinson, M.E., 2002. The ecology of Cainozoic ferns. *Rev Palaeobot Palynol* 119, 51–68.
- 22 Cooper-Driver, G.A., 1978. Insect-fern associations. *Entomol Exp Appl* 24(3), 310–316.
- 23 Dumont, C.P., Drolet, D., Deschênes, I., Himmelman, J.H., 2007. Multiple factors explain the covering  
24 behaviour in the green sea urchin, *Strongylocentrotus droebachiensis*. *Anim Behav* 73(6), 979–986.
- 25 Duncan, R.P., Autumn, K., Binford, G.J., 2007. Convergent setal morphology in sandcovering spiders  
26 suggests a design principle for particle capture. *P R Soc B* 274(1629), 3049–3056.
- 27 Eisner, T., Silberglied, R.E., 1988. A chrysopid larva that cloaks itself in mealybug wax. *Psyche* 95, 15–  
28 19.
- 29 Eisner, T., Hicks, K., Eisner, M., Robson, D.S., 1978. “Wolf-in-sheep’s-clothing” strategy of a  
30 predaceous insect larva. *Science* 199(4330), 790–794.
- 31 Eisner, T., Carrel, J.E., Van Tassel, E., Hoebeke, E.R., Eisner, M., 2002. Construction of a defensive trash  
32 packet from sycamore leaf trichomes by a chrysopid larva (Neuroptera: Chrysopidae). *P Entomol Soc*  
33 *Wash* 104(2), 437–446.
- 34 Engel, M.S., 2005. An Eocene ectoparasite of bees: The oldest definitive record of phoretic meloid  
35 triungulins (Coleoptera: Meloidae; Hymenoptera: Megachilidae). *Acta Zool Cracov* 48B(1–2), 43–48.
- 36 Engel, M.S., 2009. Gregarious behaviour in Cretaceous earwig nymphs (Insecta, Dermaptera) from  
37 southwestern France. *Geodiversitas* 31(1), 129–135.
- 38 Engel, M.S., Grimaldi, D.A., 2007. The neuropterid fauna of Dominican and Mexican amber  
39 (Neuropterida: Megaloptera, Neuroptera). *Am Mus Novit* 3587, 1–58.
- 40 Engel, M.S., Grimaldi, D.A., 2008. Diverse Neuropterida in Cretaceous amber, with particular reference  
41 to the paleofauna of Myanmar (Insecta). *Nova Suppl Entomol Keltern* 20, 1–86.
- 42 Evert, R.F., 2006. *Esau’s Plant Anatomy. Meristemes, Cells and Tissues of Plant Body: Their Structure,*  
43 *Function and Development* (Wiley, Hoboken, NJ), 3rd Ed.
- 44 Fahh, A., 1990. *Plant Anatomy* (Pergamon, Oxford), 4th Ed.
- 45 Gillison, A.N., 1969. Plant succession in an irregularly fired grassland area – Doma Peaks Region, Papua.  
46 *J Ecol* 57(2), 415–428.
- 47 Grimaldi, D., Engel, M.S., 2005. *Evolution of the Insects* (Cambridge Univ. Press, Cambridge, UK).
- 48 Hagen, H., 1856. Die im Bernstein befindlichen organischen Reste der Vorwelt. Zweiter Band, II  
49 [Organic remains found in amber from the Ancient World, second volume, II], ed. Berendt, G.C.  
50 (Nicholaischen Buchhandlung, Berlin), pp 41–125. In German.
- 51 Henry, C.S., 1977. The behavior and life histories of two North American ascalaphids. *Ann Entomol Soc*  
52 *Am* 70(2), 179–195.
- 53 Henwood, A. 1992. Excepcional preservation of dipteran flight muscle and taphonomy of insects in  
54 amber. *Palaios* 7, 203–212.
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- 1  
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4 Herendeen, P.S., Skog, J.E., 1998. *Gleichenia chaloneri* – a new fossil fern from the Lower Cretaceous  
5 (Albian) of England. *Int J Plant Sci* 159(5), 870–879.
- 6 Hölldobler, B., Wilson, E.O., 1986. Soil-binding pilosity and camouflage in ants of the tribes Basicerotini  
7 and Stegomyrmecini (Hymenoptera, Formicidae). *Zoomorphology* 106, 12–20.
- 8 Hultgren, K., Stachowicz, J., 2011. *Animal Camouflage*, eds Stevens, M., Merilaita, S. (Cambridge Univ.  
9 Press, Cambridge, UK), pp 214–238.
- 10 Janzen, J.W., 2002. *Arthropods in Baltic amber*. Ampyx-Verlag, Halle (Saale), 167pp.
- 11 Jones, D.T., 1941. Further notes on the snail-collecting aphid-lion larva (Neuroptera: Chrysopidae).  
12 *Entomol News* 52, 39–44.
- 13 Labandeira, C.C., 2014. Amber. In: *Reading and Writing of the Fossil Record: Preservational Pathways*  
14 *to Exceptional Fossilization*. The Paleontological Society Papers, vol. 20. Laflamme, M., Schiffbauer,  
15 J.D., Darroch, S.A.F. (eds), The Paleontological Society Short Course, 163–217.
- 16 Lienhard, C., 1988. Three new extra-neotropical species of Troctopsocidae (Insecta: Psocoptera). *J Nat*  
17 *Hist* 22(3), 575–587.
- 18 MacLeod, E.G., 1970. The Neuroptera of the Baltic Amber. I. Ascalaphidae, Nymphidae, and  
19 Psychopsidae. *Psyche* 77, 147–180.
- 20 Martínez-Delclòs, X., Briggs, D.E.G., Peñalver, E., 2004. Taphonomy of insects in carbonates and amber.  
21 *Palaeogeogr Palaeoclimatol Palaeoecol* 203, 19–64.
- 22 McEwen, P.K., New, T.R., Whittington, A.E., 2001. *Lacewings in the Crop Environment* (Cambridge  
23 Univ. Press, Cambridge, UK).
- 24 McKellar, R.C., Wolfe, A.P., Muehlenbachs, K., Tappert, R., Engel, M.S., Cheng, T., Sánchez-Azofeifa,  
25 G.A., 2011. Insect outbreaks produce distinctive carbon isotope signatures in defensive resins and  
26 fossiliferous ambers. *P R Soc B* 278(1722), 3219–3224.
- 27 Menor-Salván, C., Najarro, M., Velasco, F., Rosales, I., Tornos, F., Simoneit, B.R.T., 2010. Terpenoids in  
28 extracts of Lower Cretaceous ambers from the Basque Cantabrian Basin (El Soplao, Cantabria,  
29 Spain): Paleochemotaxonomic aspects. *Org Geochem* 41(10), 1089–1103.
- 30 Monserrat, V.J., Díaz-Aranda, L.M., 2012. Los estadios larvarios de los crisópidos ibéricos (Insecta,  
31 Neuroptera, Chrysopidae), nuevos elementos sobre la morfología larvaria aplicables a la sistemática  
32 de la familia. *Graellsia* 68: 31–158.
- 33 Najarro, M., Peñalver, E., Rosales, I., Pérez-de la Fuente, R., Daviero-Gomez, V., Gomez, B., Delclòs,  
34 X., 2009. Unusual concentration of Early Albian arthropod-bearing amber in the Basque-Cantabrian  
35 Basin (El Soplao, Cantabria, northern Spain): Palaeoenvironmental and palaeobiological implications.  
36 *Geol Acta* 7(3), 363–387.
- 37 Najarro, M., Peñalver, E., Pérez-de la Fuente, R., Ortega-Blanco, J., Menor-Salván, C., Barrón, E.,  
38 Soriano, C., Rosales, I., López del Valle, R., Velasco, F., Tornos, F., Daviero-Gomez, V., Gomez, B.,  
39 Delclòs, X., 2010. A review of the El Soplao amber outcrop, Early Cretaceous of Cantabria (Spain).  
40 *Acta Paleontol Sin* 84(4), 959–976.
- 41 Nascimbene, P., Silverstein, H., 2000. The preparation of fragile Cretaceous ambers for conservation and  
42 study of organismal inclusions. In: *Studies on Fossils in Amber, with Particular Reference to the*  
43 *Cretaceous of New Jersey*, ed. Grimaldi, D. (Backhuys Publishers, Leiden, The Netherlands), pp 93–  
44 102.
- 45 Nel, A., Delclòs, X., Hutin, A., 2005. Mesozoic chrysopid-like Planipennia: A phylogenetic approach  
46 (Insecta: Neuroptera). *Ann Soc Entomol Fr* 41(1), 29–69.
- 47 New, T.R., 1982. The larva of *Nymphes* Leach (Neuroptera: Nymphidae). *Neurop Int* 2(2), 79–84.
- 48 Ortega-Blanco, J., Rasnitsyn, A.P., Delclòs, X., 2008. First record of anaxyelid woodwasps  
49 (Hymenoptera: Anaxyelidae) in Lower Cretaceous Spanish amber. *Zootaxa* 1937, 39–50.
- 50 Penney, D. (Ed.), 2010. *Biodiversity of fossils in amber from the major world deposits*. Siri Scientific  
51 Press.
- 52 Peñalver, E., Delclòs, X., 2010. Spanish amber. In: *Biodiversity of fossils in amber from the major world*  
53 *deposits*, ed. Penney, D. (Siri Scientific Press, Manchester, UK), pp 236–270.
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3  
4 Peñalver, E., Wegierek, P., 2008. A new genus and species of the family Tajmyraphididae (Hemiptera:  
5 Sternorrhyncha) in Early Cretaceous amber from Peñacerrada I (Spain). *Alavesia* 2, 187–192.
- 6 Peñalver, E., Delclòs, X., Soriano, C., 2007. A new rich amber outcrop with palaeobiological inclusions  
7 in the Lower Cretaceous of Spain. *Cretaceous Res* 28, 791–802.
- 8 Peñalver, E., Labandeira, C.C., Barrón, E., Delclòs, X., Nel, P., Nel, A., Tafforeau, P., Soriano, C., 2012.  
9 Thrips pollination of Mesozoic gymnosperms. *P Natl Acad Sci USA* 109(22), 8623–8628.
- 10 Pérez-de la Fuente, R., Delclòs, X., Peñalver, E., Speranza, M., Wierzchos, J., Ascaso, C., Engel, M.S.,  
11 2012a. Early evolution and ecology of camouflage in insects. *P Natl Acad Sci USA* 109(52), 21414–  
12 21419.
- 13 Pérez-de la Fuente, R., Peñalver, E., Delclòs, X., Engel, M.S., 2012b. Snakefly diversity in Early  
14 Cretaceous amber from Spain (Neuropterida, Raphidioptera). *Zookeys* 204, 1–40.
- 15 Perreau, M., Tafforeau, P., 2011. Virtual dissection using phase-contrast X-ray synchrotron  
16 microtomography: reducing the gap between fossils and extant species. *Syst Entomol* 36(3), 573–580.
- 17 Pohl, H., Friedrich, F., Beckmann, F., Herzen, J., Beutel, R.G., 2008. External and internal structures of  
18 fossil and extant Strepsiptera (Hexapoda) using SR $\mu$ CT. *Annual Rep., Photon Sci. – HASYLAB,*  
19 2008. Online at: [http://photon-science.desy.de/annual\\_report/files/2008/2008275.pdf](http://photon-science.desy.de/annual_report/files/2008/2008275.pdf)
- 20 Ponder, W.F., 1983. A revision of the Recent Xenophoridae of the world and of the Australian fossil  
21 species (Mollusca, Gastropoda). *Aust Mus Mem* 17, 1–126.
- 22 Ponder, W.F., 1994. The Malacofauna of Hong Kong and Southern China III, ed. Morton, B. (Honk Kong  
23 Univ. Press, Hong Kong), pp 215–239.
- 24 Russell, A.E., Raich, J.W., Vitousek, P.M., 1998. The ecology of the climbing fern *Dicranopteris linearis*  
25 on windward Mauna Loa, Hawaii. *J Ecol* 86, 765–779.
- 26 Schmidt, A.R., Jancke, S., Lindquist, E.E., Ragazzi, E., Roghi, G., Nascimbene, P.C., Schmidt, K.,  
27 Wappler, T., Grimaldi, D.A., 2012. Arthropods in amber from the Triassic Period. *P Natl Acad Sci*  
28 USA 109, 14796–14801.
- 29 Skog, J.E., 2001. Biogeography of Mesozoic leptosporangiate ferns related to extant ferns. *Brittonia*  
30 53(2), 236–269.
- 31 Soriano, C., Archer, M., Azar, D., Creaser, P., Delclòs, X., Godhelp, H., Hand, S., Jones, A., Néraudeau,  
32 D., Ortega-Blanco, J., Pérez-de la Fuente, R., Perrichot, V., Saupe, E., Solórzano-Kraemer, M.,  
33 Tafforeau, P., 2010. Synchrotron x-ray imaging of inclusions in amber. *C R Palevol* 9(6–7), 361–368.
- 34 Subías, L.S., 2012. Un nuevo oribátido cavernícola, *Damaeus gevi* n. sp., de España (Acari: Oribatida:  
35 Damaeidae) con un camuflaje de cadáveres de oribátidos adheridos a sus exuvias. *Ib J Arac* 20, 31–  
36 34.
- 37 Tauber, C.A., Ruberson, J.R., Tauber, M.J., 1995. Size and morphological differences among the larvae  
38 of two predacious species and their hybrids (Neuroptera: Chrysopidae). *Ann Entomol Soc Am* 88(4),  
39 502–511.
- 40 Tauber, C.A., Tauber, M. J., Albuquerque, G. S., 2014. Debris-carrying in larval Chrysopidae: Unraveling  
41 its evolutionary history. *Ann Entomol Soc Am*, 107(2), 295–314.
- 42 Van Konijnenburg-Van Cittert, J.H.A., 2002. Ecology of some Late Triassic to Early Cretaceous ferns in  
43 Eurasia. *Rev Palaeobot Palynol* 119, 113–124.
- 44 Walker, L.R., Boneta, W., 1995. Plant and soil responses to fire on a fern-covered landslide in Puerto  
45 Rico. *J Trop Ecol* 11, 473–479.
- 46 Weirauch, C., 2006. Anatomy of disguise: Camouflaging structures in nymphs of some Reduviidae  
47 (Heteroptera). *Am Mus Novit* 3542, 1–18.
- 48 Weitschat, W., 2009. Jäger, Gejagte, Parasiten und Blindepassagiere – Momentaufnahmen aus dem  
49 Bernsteinwald [Predator, prey, parasites and stowaways – snapshots of the amber forest]. *Denisia* 26,  
50 243–256. In German.
- 51 Weitschat, W., Wichard, W., 1998. Atlas der Pflanzen und Tiere im Baltischen Bernstein. München.  
52 Verlag Dr. Friedrich Pfeil, 256pp.
- 53 Wu, R.J.C., 1996. *Secrets of a Lost World: Dominican Amber and Its Inclusions* (R.J.C. Wu, Santo  
54 Domingo, Dominican Republic).
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5 **Figure captions**  
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8 **Figure 1.** Lateroventral habitus of *Hallucinochrysa diogenesi* Pérez-de la Fuente, Delclòs, Peñalver and  
9 Engel, 2012 (Neuroptera: Chrysopoidea), holotype, preserved together with its trash packet.  
10 Abbreviations: ab – abdomen; h – head; l1 – prothoracic leg; l2 – mesothoracic leg; l3 – metathoracic leg;  
11 tb – tubular tubercle; tx – thorax. Only the legs from the right side of the body and well-visible tubercles  
12 in the image have been tagged. Legs tagged at the femoro-tibial joint. A multiple fracture (f) is  
13 transversally crossing the abdomen. The image was obtained by combining photographs taken at  
14 successive focal planes.  
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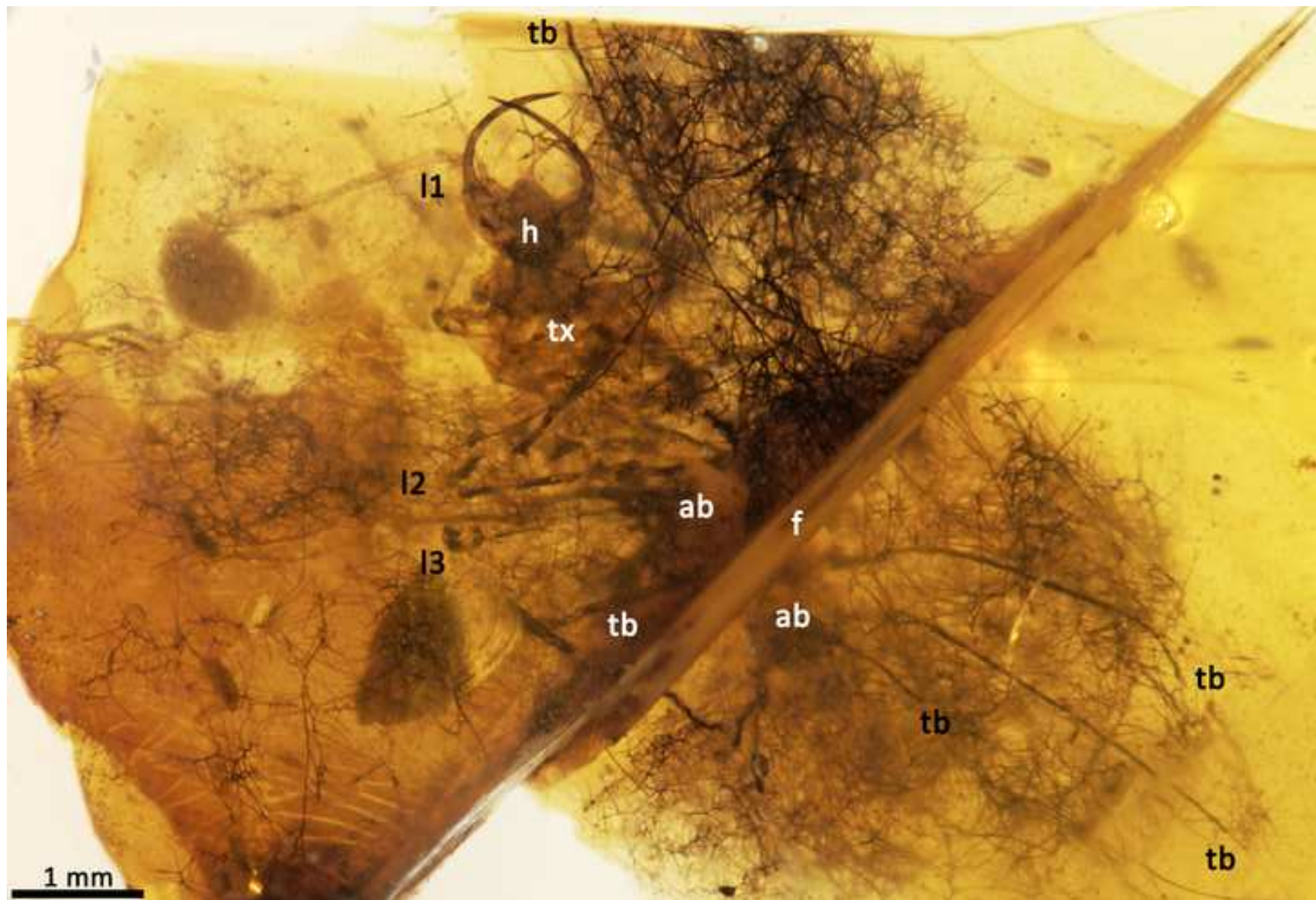
16 **Figure 2.** Reconstruction of *H. diogenesi* depicting its life aspect. A) Dorsolateral view. B) Frontal view.  
17 C) Sagittal view. Head and leg setation have been omitted for clarity. Tubular tubercle setae have been  
18 modified for improved visibility (these setae are variable in length in the fossil, usually longer than  
19 modeled, and their trumpet-shaped endings are somewhat smaller than as shown here). Length of  
20 antennae, body coloration, and number of abdominal tubular tubercle pairs are based on extant larvae.  
21 (Author: J.A. Peñas.)  
22  
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24 **Figure 3.** Detail of the head and the right prothoracic leg of *H. diogenesi* in lateroventral view; note also  
25 the abundant trichomes from its trash packet. Abbreviations: a – antenna; fe – femur; lp – labial palpi; o –  
26 ommatidium; ta – tarsus; tb – tubular tubercle; ti – tibia. A circular fracture is crossing the head (f). The  
27 image was obtained combining pictures taken at successive focal planes.  
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30 **Figure 4.** Detail of two trichomes from the trash-packet of *H. diogenesi*. These two trichomes appear to  
31 be tangled with one another and show different stages of development, the left one (A) being a more  
32 mature form than the one at right (B).  
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34 **Animation 1.** Morphological and locomotive reconstruction of *Hallucinochrysa diogenesi*. See figure 2  
35 caption for additional information.  
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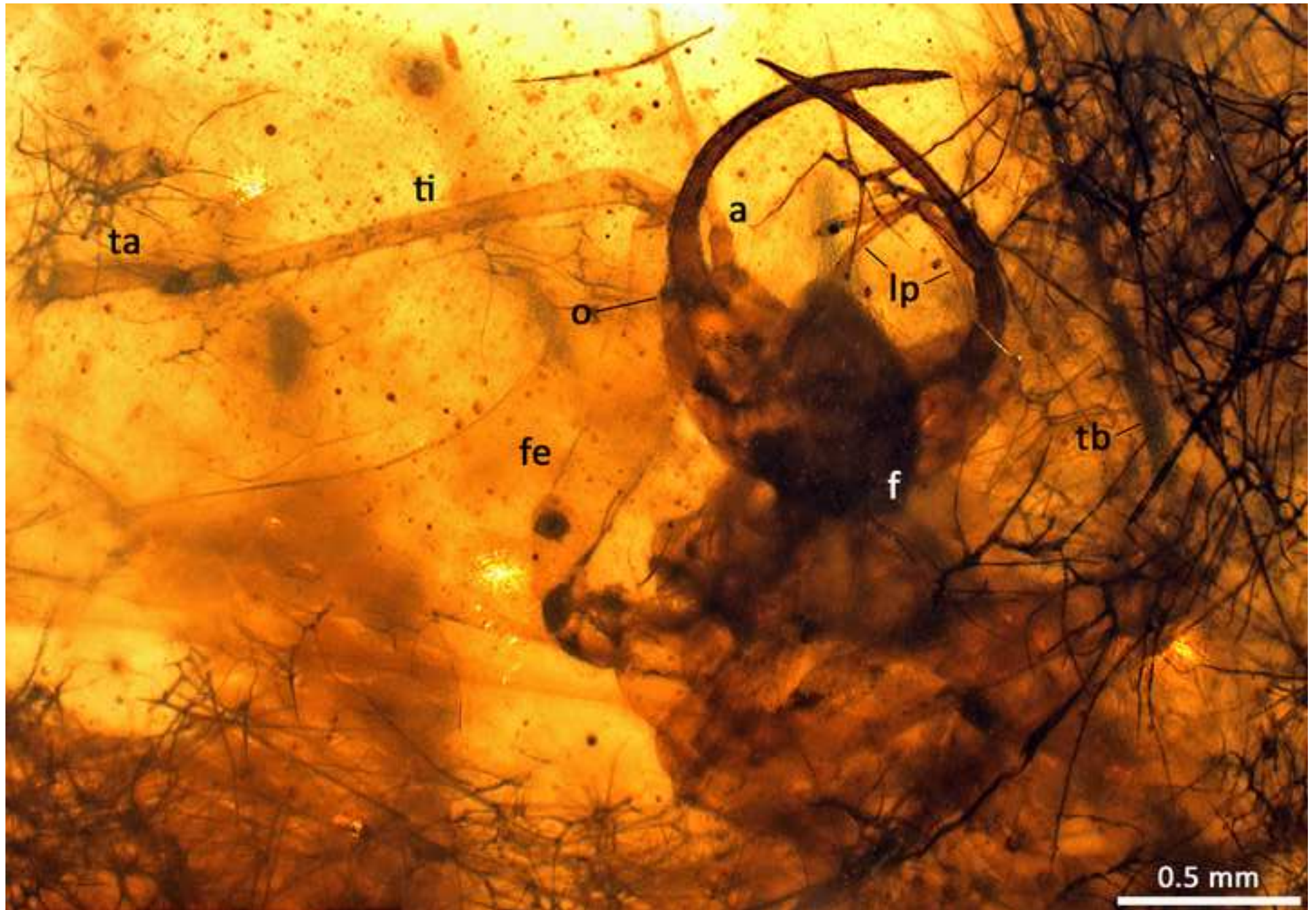


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