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Direct trafficking pathways from the Golgi apparatus to the plasma membrane

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Abstract

In eukaryotic cells, protein sorting is a highly regulated mechanism important for many physiological events. After synthesis in the endoplasmic reticulum and trafficking to the Golgi apparatus, proteins sort to many different cellular destinations including the endolysosomal system and the extracellular space. Secreted proteins need to be delivered directly to the cell surface. Sorting of secreted proteins from the Golgi apparatus has been a topic of interest for over thirty years, yet there is still no clear understanding of the machinery that forms the post-Golgi carriers. Most evidence points to these post-Golgi carriers being tubular pleomorphic structures that bud from the *trans*-face of the Golgi. In this review, we present the background studies and highlight the key components of this pathway, we then discuss the machinery implicated in the formation of these carriers, their translocation across the cytosol, and their fusion at the plasma membrane.

Keywords: Secretion, Constitutive Secretion, post-Golgi carriers, Golgi to plasma membrane sorting, pleomorphic tubular carriers

Abbreviations: ATP, adenosine triphosphate; BFA, Brefeldin A; CARTS, CARriers of the TGN to the cell Surface; CI-MPR, cation-independent mannose-6 phosphate receptor; CtBP3/BARS, C-terminus binding protein 3/BFA adenosine diphosphate-ribosylated substrate; ER, endoplasmic reticulum; GlcCer, glucosylceramide; PAUF, pancreatic adenocarcinoma up-regulated factor; PM, plasma membrane; RUSH, retention using selective hooks; SBP, streptavidin-binding peptide; SM, sphingomyelin; SNARE, soluble N-ethylmaleimide sensitive fusion protein attachment protein receptor; SPCA1, secretory pathway calcium ATPase 1; TGN, trans-Golgi Network; TIRF, total internal reflection fluorescence; ts, temperature sensitive; VSV, vesicular stomatitis virus

1. Introduction

Cells are internally compartmentalised. This allows proteins and lipids that participate in the same biochemical pathways to have higher local concentrations. Additionally, compartmentalisation segregates proteins that could potentially damage other cellular components (eg. proteases). This system is maintained by the trafficking of lipids and proteins in vesicles within the cell. Vesicles bud from a 'donor' compartment and translocate across the cytosol to fuse with a 'receiver' compartment, a process orchestrated by the membrane trafficking machinery.

Membrane trafficking allows delivery of nascently biosynthesised proteins and lipids to the plasma membrane. Approximately 11-15% of human proteins are secreted as soluble proteins to the extracellular space [1–3] and an additional 10% localise to the plasma membrane [4]. This represents over 3500 proteins with a vast array of functions, including antibodies, signalling molecules, and integral endomembrane proteins which traffic to the plasma membrane before being internalised to the endolysosomal system. Secreted proteins are not only numerous, but abundant, with the secreted protein collagen family accounting for 30% of total protein mass in mammals [5]. This review will focus on the trafficking route and machinery required to deliver proteins from the Golgi apparatus to the plasma membrane.

2. Background studies on export from the Golgi apparatus

The fundamental principles of protein secretion were established during the early days of protein trafficking, by the pioneering work of George Palade. To track the intracellular biosynthetic routes, Palade used the highly secretory pancreatic exocrine cells, which accumulate proteins in zymogen granules before their regulated secretion [6]. Using analytical centrifugation and pulse-chase autoradiography, Palade demonstrated the fundamental principle that protein sorting is sequential and vectorial.

Later studies on the trafficking of immunoglobulin, a model secreted protein which traffics directly from the Golgi to the plasma membrane, identified the "smooth vesicles" or "secretory vacuoles" that contain this protein [7,8]. It was also realised, around the same time, that there were two different forms of secretion: constitutive (i.e. continuous) and regulated (i.e. upon stimulation) secretion. Both forms of secretion can occur in the same cell [9,10] and segregation between the two routes happens in the *trans*-Golgi network [11]. In this review we focus on the constitutive secretory route that is found in the vast majority of eukaryotic cells.

It soon became clear that specialised cells, such as endocrine cells and neurons, are able to secrete a subset of proteins from a preferred portion of the plasma membrane. This process was

later defined as polarised secretion [9,12]. This theory was formed from multiple lines of evidence including the polarised distribution of proteins in electron micrographs [13–15], immunoisolation of synaptic vesicles identifying specific proteins localised to polarised carriers [16], imaging studies tracking directional trafficking of organelles in the axons of squids [17–19], and secretion of viral proteins in polarised epithelial cells [20–22].

To identify the fundamental machinery of the secretory process, the laboratory of Randy Schekman performed a genetic screen in yeast [23]. Secretory mutants in yeast accumulate proteins and membrane intracellularly, making the cells denser. These mutant yeast would therefore differentially sediment from wild-type yeast on density gradients. This principle was employed to screen the yeast genome for mutants of the secretory system. To perform this screen they used ‘temperature sensitive’ (*ts*) mutants. *ts* mutants result in temperature dependent protein instability. Proteins with *ts* mutations fold properly in a ‘permissive’ temperature, but when the temperature is shifted they misfold, thus revealing the phenotype of protein loss. This allowed a set of 23 complementation groups (*Sec1-23*) to be discovered in what was called the *Sec* screen [23]. One of the major findings from this screen was the COPII protein complex that allows for endoplasmic reticulum (ER) to Golgi transport. Along with Rothman’s work [24,25], this landmark study identified the first genes encoding the machinery involved in protein trafficking and secretion. At this point, however, the route from the early secretory system to the plasma membrane and the machinery involved was still not understood.

In efforts to understand the principle of protein sorting and secretion, the ‘bulk flow’ hypothesis was formally proposed [9,26]. The ‘bulk flow’ model states that secretion from the ER through the Golgi apparatus to the plasma membrane does not require a signal and happens in ‘bulk’ by default. The first hints at this hypothesis came from the secretion of bacterial β -lactamase after microinjection of the RNA into *Xenopus* oocytes [27]. Subsequently, bulk peptide secretion was demonstrated by monitoring glycosylation and secretion of exogenously added peptides to mammalian cell lines and detecting their arrival at the cell surface [26]. A subset of the secreted peptides were glycosylated indicating they had been transported through the Golgi apparatus. The conclusion was that these peptides were secreted in bulk by default.

In the following years there were a number of studies that questioned the generality of this hypothesis. Most of these studies focussed on the ER export of membrane spanning proteins rather than the Golgi to plasma membrane route. Studies on soluble proteins reported increased concentration of secreted proteins as they are trafficked to the plasma membrane, suggesting that they are actively sorted. Concentrative transport of soluble proteins, however, was thought to be due to selective removal of other cargo rather than active sorting, supporting the idea that there is no selective transport on the soluble cargo itself [28,29]. There are a subset of soluble proteins that are actively sorted from the ER, such as GPI anchored proteins which are sorted by the p24 protein family [30] and proteins including cathepsin C [31], cathepsin Z [32], factor V, and factor VIII [33] which require the ERGIC-53/LMAN1 receptor. It is generally agreed, however, that the default fate of soluble proteins without any known signal is to be secreted.

Soluble protein secretion has been demonstrated to be highly efficient. In yeast, induction of secretion of acid phosphatase with a temperature shift indicates a six-fold increase in secreted protein with almost no detectable increase in intracellular levels [34]. Studies in plants demonstrated that 50-90% of synthesised soluble proteins are delivered to the extracellular space [35,36], consistent with studies in mammalian cells [37]. A quantitative study on the secretion of a soluble non-glycosylated protein found that after synthesis and folding the fastest molecules are secreted in about 15 mins with the half-life of secretion being 40 mins [38]. The resulting consensus for soluble secreted proteins is that transport is not signal-mediated, relatively fast and follows a default bulk flow pathway from the Golgi apparatus to the plasma membrane.

The 'default' route of integral membrane proteins is more complicated as there are multiple sorting signals and protein topologies. As discussed above there are two membrane trafficking steps to arrive at the plasma membrane: from the ER to the Golgi and from the Golgi to the plasma membrane. Following the proposal of the bulk-flow hypothesis, a number of groups demonstrated that ER export of transmembrane proteins is at least somewhat signal-mediated [36,39–41]. After arrival at the Golgi, membrane proteins need to sort to their final destination. Transport from the Golgi apparatus was hypothesised to be selectively mediated by clathrin after the discovery of the AP-1 clathrin adaptor complex [42–44]. Studies using a type-I membrane-spanning protein, the cation-independent mannose-6 phosphate receptor (CI-MPR), demonstrated that mutations in the cytoplasmic terminus cause retention in the Golgi apparatus [45]. It was later shown that monomeric GGA clathrin adaptors also play a role at the Golgi apparatus and are essential for export of CI-MPR [46,47]. Therefore, there are two clathrin adaptor complexes at the Golgi which selectively sort cargo in a signal mediated manner.

There are also trafficking routes for integral membrane proteins from the Golgi apparatus, independent of cytoplasmic signals. The mechanism of Golgi retention was investigated by making a series of chimeras made of two integral membrane proteins: a Golgi localised protein (β -1,4-galactosyltransferase (GT)) and an endosomal protein (human invariant chain (Ii)) [48]. A chimera with the transmembrane domain of GT and the cytoplasmic and luminal domain of Ii is retained in the Golgi, demonstrating that the transmembrane domain alone can mediate Golgi localisation [48]. Similarly, chimeras of the transmembrane domain of the Golgi-localised α -2,6-sialyltransferase (ST) with the cell surface protein dipeptidylpeptidase IV (DPPIV) are Golgi retained [49]. ST fusions with the transmembrane domain replaced with either 17 or 23 leucines localises to the Golgi apparatus and plasma membrane respectively. This effect was only observed, however, when the luminal domain of ST was also in the chimera [49,50]. Additionally, a type-I membrane protein (CD8) with a transmembrane domain of 23 leucine residues accumulates on the plasma membrane, whereas the same fusion with 17 leucines accumulates on the Golgi apparatus [50]. The relationship between transmembrane domain length and localisation was also analysed using temperature sensitive VSV-G (described in Section 4, below) [51]. A 7 amino acid truncation of the transmembrane domain of VSV-G causes less efficient Golgi export.

The E1 glycoprotein from an avian coronavirus localises to the Golgi apparatus in mammalian cells [52]. The E1 glycoprotein is a triple-pass transmembrane protein (with the C-terminus in the

cytoplasm). Replacing the transmembrane domain of VSV-G with the first transmembrane domain of E1 mislocalises VSV-G to the Golgi apparatus [53]. Another example, from plants, is the vacuolar transmembrane protein α -TIP which accumulates on the vacuolar membrane of tobacco leaf cells despite deletion of the cytosolic terminus [54]. Truncations of the transmembrane domain of α -TIP resulted in protein accumulation at the Golgi. This suggests that the length of the transmembrane domain is a determining factor for post-Golgi sorting, a finding supported by other studies in plants [36,55]. These observations were further corroborated in mammals by comparing the length of an extensive list of over 1000 single-pass membrane proteins with a known localisation. Proteins known to reside on the plasma membrane have, on average, a longer hydrophobic section when compared to Golgi localised proteins [56].

In summary, by the early 1990's the general rules of protein trafficking had been discovered. It was clear that proteins sort in a controlled way to various subcellular localisations. The studies above established a series of fundamental principles including: the binding of cytoplasmic adaptors to transmembrane cargos, the default secretion of soluble proteins and that membrane proteins do not have a 'default' route.

3. Navigating through the Early Secretory System

Proteins that enter the secretory system can be identified by a number of signals. Soluble proteins and type-I integral membrane proteins have a signal peptide on their N-terminus, type-II integral membrane proteins have a stop-transfer signal to allow C-terminal insertion into the ER, and multipass proteins have a combination of signal peptide and stop-transfers depending on the topology of the transmembrane domains. Once in the ER, the proteins fold due to an array of ATPase chaperones, the most characterised of which is BiP. The chaperones bind to unfolded hydrophobic portions of the protein and expend ATP to allow proper folding. The last step before export to the Golgi apparatus is the covalent addition of glycans [57].

There are two key processes happening in the Golgi stacks: proteins are progressively further glycosylated [58] and then sorted to their steady-state destination in the cell. Within the Golgi apparatus proteins are sorted into different domains [59–62] to be trafficked to at least four distinct locations (Fig.1). One subset of proteins return in COPI vesicles back to the ER. This includes the soluble resident ER chaperones that are 'rescued' from the *cis* Golgi stacks by the KDEL receptor, to allow steady-state localisation in the ER [63,64]. A second subset includes the Golgi resident enzymes that are retained in the Golgi apparatus [65]. These Golgi resident enzymes are thought to be retained by a combination of active sorting and oligomerisation (reviewed: [65]). The third subset traffic in clathrin-coated vesicles from the Golgi apparatus. These vesicles traffic directly to the endolysosomal system and contain lysosomal hydrolases and their receptors [59]. This process is mediated by the clathrin monomeric GGA adaptors and AP-1 complex which bind to both a motif in the cytosolic termini of the transmembrane cargos and accessory proteins present on the *trans*-Golgi [66]. The final subset of proteins leave the Golgi apparatus and traffic directly to the plasma membrane. This pathway and these carriers are the focus of this review.

4. Budding of constitutive cargos from the Golgi Apparatus

The *trans*-Golgi network was proposed to be the point at which cargos leave the Golgi apparatus [11] based on the observation of budding structures in electron micrographs [67]. To understand what machinery was involved in the formation of these membrane carriers budding from the Golgi, the well-characterised clathrin-mediated transport seemed an obvious candidate to investigate. Clathrin carriers range from the typical spherical 60-100 nm clathrin-coated vesicles to larger, up to 1200 nm in size, grape-like structures displaying several coated buds [68]. The clathrin coat and its accessory protein complex, AP-1, was also shown to drive the formation of Weibel-Palade bodies, large cigar-shaped secretory organelles produced by endothelial cells which can be up to 5 μm long [69,70]. It has been proposed that the diverse array of carrier morphologies could be an adaptive mechanism for a variety cargo size or abundance. In the same line, COPII coat, which mediates cargo transport from the ER to the Golgi apparatus, can adopt more flexible conformations than the conventional 80 nm COPII carriers and assemble into tubular structures [71] or even potentially compartment-to-compartment tunnels [72] to allow the transport of large cargos like procollagen. In addition, cargos could potentially also affect the architecture of the Golgi. For example, overexpression of uroplakins, proteins that follow the conventional trafficking route from Golgi to PM localising on the apical surface of urothelial cells, triggers the fragmentation of the Golgi apparatus [73]. This effect suggests an adaptation of the cell in response to an increased flux of secretory proteins.

Although some clathrin adaptors such as GGAs and AP1 associate to the cytosolic termini of transmembrane proteins at the *trans*-Golgi, it was soon identified that a clathrin-dependent mechanism was unlikely to be involved in the secretory carriers. In fact, the secreted protein invertase was unaffected by deletion of the clathrin heavy chain in *S. cerevisiae* [74]. It was therefore established relatively early that the secretory carriers are probably not clathrin coated vesicles and perhaps completely independent of clathrin.

In addition to the various clathrin carriers that bud from the Golgi apparatus, there is a second class of carriers often referred to as a 'pleomorphic tubular-vesicular carriers'. These are lipid tubules and vesicles that bud from the Golgi apparatus, and are heterogeneous in size and shape in contrast to a clathrin-coated vesicle, which are generally geometrically constrained by the clathrin lattice. In contrast, pleomorphic tubular-vesicular carriers that traffic from the Golgi apparatus directly to the plasma membrane seem devoid of coat [75]. Studies on temperature sensitive vesicular stomatitis virus (VSV) mutant strains revealed a subset which had a mutation in the envelope 'G' protein of the virus. This protein has incomplete glycosylation at the non-permissive temperature [76], and was shown by cellular fractionation to localise at a different site in the cell depending on the temperature [77]. Cells infected with the temperature sensitive mutant strain retain the G protein in the ER and, upon temperature shift, traffic the protein towards the PM [78]. This effect is recapitulated with overexpression of a cDNA clone of the mutant protein [79] allowing for an ectopically expressed *tsVSV-G* that misfolds and accumulates in the ER at the 'non-permissive' temperature of 40°C. Accumulated *tsVSV-G* is released *en masse* when the cells are shifted to the permissive temperature. This *en masse* export allows observation of *tsVSV-G* as it traffics to the plasma membrane, either biochemically or by imaging fluorophore

fusions. Visualisation of *tsVSV-G-GFP* as it buds from the Golgi revealed tubular carriers that bud from the Golgi apparatus and traffic to the plasma membrane [80,81].

These post-Golgi carriers were described to be large pleomorphic tubular structures which traffic directly to the plasma membrane with an average carrier containing ~10,000 *tsVSV-G-GFP* molecules [81,82]. These are likely not artifacts of an overloaded secretory system as they traffic along microtubules and fuse with the plasma membrane, demonstrating that cellular machinery exists to sort these carriers [81,83]. Later studies suggested that *tsVSV-G* traffics to the plasma membrane indirectly via recycling endosomes [84,85]. Whether *tsVSV-G* traffics via the indirect or direct route to the plasma membrane is likely to be dependent on cell type, analogous to the indirect and direct trafficking routes of the lysosomal protein LAMP1 [59,86,87].

Correlative light-electron microscopy imaging of the carriers revealed that they are 0.3–1.7 μm [80], a factor larger than typical clathrin coated vesicles, which are approximately between 0.06 and 0.1 μm [88]. These carriers were demonstrated to form *en bloc*, rather than by fusion of multiple smaller vesicular structures [75]. These initial observations set the stage for understanding the machinery that mediates this process.

5. Machinery associated with Golgi to Plasma Membrane Carriers

Unlike other cellular pathways, such as clathrin coated vesicles [88], or retromer recycling carriers [89], there is still no clear consensus for the machinery and molecular processes that drive the formation, budding, translocation and fusion of post-Golgi to plasma membrane carriers. There are, however, a number of candidate machineries that play a role in this process. We have detailed these below (see overview in figure 2).

5.1 Arf1 positive tubular carriers

Arf1, an extensively studied small GTPase of the Arf family, has a well-established role in COPI vesicle biogenesis at the Golgi [90]. Arf1 is also important in the budding of a large variety of other coated vesicles, recruiting adaptor protein complexes like AP1, AP3, AP4 as well as GGA, and exomer complexes [91,92]. Arf1 forms a dimer in its GTP-bound form which is crucial to induce positive membrane curvature *in vitro* [93]. A mutant of Arf1 that is unable to dimerise is not able to induce COPI vesicle formation. Arf1, therefore, seems to induce membrane bending by forming a dimer in order to generate vesicles. This property of Arf1 prompted Rothman's laboratory to investigate its role in the generation of Golgi-derived tubules [94]. Halo-tagged Arf1 was imaged at endogenous expression levels with super-resolution imaging and, only under these conditions (not by over-expression of Arf1) Arf1 labels transient tubular carriers emerging from the Golgi. Arf1 GTPase activity is required for the formation of these tubular carriers. There are at least two distinct populations of Arf1-containing tubules. A subset of these tubules, containing VSV-G, traffic along microtubules in an anterograde direction towards the cell periphery. Arf1 positive carriers, however, were not observed fusing directly to the plasma membrane implying that they either lose Arf1 before fusion or there is an additional trafficking step [94].

Arf1, cortactin and dynamin 2 can be detected on isolated Golgi membranes from a sucrose gradient. Incubating these membranes with the Arf1 inhibitor brefeldin A (BFA) causes loss of Arf1, cortactin and dynamin 2 from the membranes, which can be inhibited by supplementation with GTP-loaded Arf1 [95]. Imaging experiments show addition of brefeldin A causes loss of cortactin and dynamin 2 from the Golgi, suggesting that Arf1 is able to recruit actin, cortactin and dynamin 2 to the *trans*-Golgi. Finally, the expression of truncated cortactin proteins decreases the efficiency of *tsVSV-G* export from the TGN towards the cell surface. Cortactin promotes the formation of branched actin polymers through interactions with the Arp2/3 complex [96], and dynamin 2 is a small GTPase essential for membrane fission [97]. These findings show the importance of Arf1 in the recruitment of an actin/cortactin/dynamin 2 complex essential for post-Golgi transport.

Two binding partners of Arf1, PKD (Serine/threonine-protein kinase D, human gene isoforms: PRKD1-3) and FAPP2 (encoded by the gene PLEKHA8), are also associated with post-Golgi tubular carriers [98,99]. Addition of the metabolite ilimaquinone causes Golgi fragmentation by activating PKD through stimulation of a heterotrimeric G protein (G $\beta\gamma$) [100]. An inactive mutant of PKD, expressed at moderate levels in a stable HeLa cell line at 20°C, inhibits protein transport from the *trans*-Golgi to the cell surface by causing Golgi tubulation [101]. Cargo destined for the plasma membrane like *tsVSV-G* are trapped in large tubules that remain attached to the TGN. Similarly, depletion of PRKD2 and PRKD3 (human homologs of PKD) by siRNA also inhibits TGN-to-cell surface transport and shows cargo accumulation in large tubules [102]. These results strongly suggest that PKD is required for membrane fission at the *trans*-Golgi of carriers destined for the cell surface. PKD has two cysteine-rich domains at the N-terminus, one interacts with diacylglycerol and the second one with Arf1 [103,104]. The direct interaction of PKD2 with ARF1 is enhanced when the small GTPase is in its active conformation [104]. PKD is activated by G $\beta\gamma$ to stimulate specific lipid kinases like the enzyme PI(4)KIII β [98,100]. This promotes the production of PI(4)P, a crucial lipid that drives vesicle biogenesis at the TGN through a variety of mechanisms [105] - see also Section 7 below.

FAPP1 and FAPP2 were originally identified to bind specifically to PI(4)P through their PH domain [106,107]. Pull-down experiments with isolated Golgi membranes and with recombinant proteins show a direct interaction of FAPP proteins with Arf1-GTP [107]. Binding to both PI(4)P and Arf1 allows FAPP targeting to the *trans*-Golgi via coincidence detection. It was proposed that FAPP proteins are able to deform membranes [108–110]. Recombinant FAPP2, as well as the isolated PH domain of FAPP1, are able to induce tubulation of membrane sheets *in vitro* [108,110]. During imaging of synchronised transport of *tsVSV-G*, FAPP proteins localise on tubular carriers emerging from the TGN [107]. Knockdown of FAPPs by siRNA inhibits cargo delivery to the plasma membrane, indicating that FAPP proteins are essential for the constitutive post-Golgi transport. Furthermore, FAPP2 is specifically involved in apical transport in polarised MDCK cells [111]. FAPP2 has a similar domain structure to FAPP1 aside from an additional glycolipid-transfer-protein-homology domain at its C-terminus. FAPP2 specifically transfers glucosylceramide (GlcCer) *in vitro* and this transfer is stimulated in the presence of PI(4)P and Arf1 [112]. GlcCer is normally *trans*-Golgi localised, however in FAPP2 knock-down cells GlcCer is mislocalised to the *cis*-Golgi. As GlyCer is a precursor for glycosphingolipids which are key

components of the plasma membrane, this role of FAPP2 connects tubule formation, PI(4)P generation, and subsequent FAPP2 recruitment at the Golgi with plasma membrane homeostasis.

Another binding partner of Arf1 at the *trans*-Golgi is the exomer complex [92,113,114]. To date, the exomer complex is the only known cargo adaptor involved in the direct transport of cargo from the TGN to the plasma membrane [92,115,116]. Discovered in yeast, there is no obvious exomer homolog in metazoans. The exomer complex is a heterotetramer composed of two Chs5 subunits and any two members of the four paralogous Chs5-Arf1 binding proteins (ChAPs), Chs6, Bud7, Bch1, and Bch2, which confer cargo specificity through direct binding to the cytoplasmic termini. Structural studies reveal that the exomer complex binds two Arf1 molecules [113]. By inserting a hydrophobic element into the membrane and coordinating the membrane insertion of two Arf1 molecules, exomer amplifies Arf1 membrane remodelling ability. Thus, exomer participates in cargo sorting and membrane fission.

In summary, Arf1 has a key role in the formation of *tsVSV-G* enriched post-Golgi tubular carriers by inducing membrane curvature and by recruiting a variety of binding partners such as lipid modifying enzymes, lipid transport proteins and fission promoting proteins.

5.2 Biosynthetic Lamp1 carriers

Post-Golgi carriers have also been observed using the “retention using selective hooks” (RUSH) system [59,117]. By fusing a protein of interest to streptavidin-binding peptide (SBP) and co-expressing an ER localised streptavidin, RUSH allows the protein of interest to accumulate in the ER due to the interaction between the streptavidin and the SBP [118]. Addition of exogenous biotin outcompetes the interaction between the SBP and streptavidin leaving the protein of interest free to traffic along its normal trafficking itinerary. By combining this approach with high-resolution microscopy to track the secretion of different endolysosomal proteins, Lamp1 was shown to sort into distinct Golgi domains when compared to the cation-dependent mannose-6-phosphate receptor or sortillin [59]. Lamp1 leaves the Golgi in tubular carriers depending on the transmembrane and luminal domains of Lamp1 but independently of sorting signals in the cytosolic tail. Finally, microscopy experiments show that Lamp1 tubular carriers that contain transferrin receptor and VSV-G, fuse directly with the plasma membrane. RUSH of VSV-G shows that VSV-G concentrates in Lamp1 carriers, suggesting that these carriers correspond to the *tsVSV-G* Golgi-to-PM carriers discussed above. The route of endogenous LAMP1 from the Golgi remains controversial as immunoelectron microscopy studies suggest that a fraction of these non-clathrin coated Lamp1 carriers travel instead directly to late endosomes and contain the HOPS component Vps41 and the SNARE VAMP7 which are both required for fusion [119].

In summary, the lysosomal protein Lamp1 traffics on VSV-G enriched post-Golgi tubular carriers to reach the cell surface before being recycled to the endolysosomal compartments. However, further investigations on this class of carriers is essential and will help determine if these carriers are the same as the Arf1 positive carriers described above.

5.3 Rab6 associated carriers

Rab6 was also found associated with about 50% of Golgi-derived vesicles enriched with *tsVSV-G* together with neuropeptide Y and BDNF which are secreted when overexpressed in HeLa cells [120]. TIRF experiments show the direct fusion of these Rab6 secretory vesicles with the plasma membrane. Furthermore, Rab6 knockdown delays the secretion of VSV-G, implying a role of Rab6 in the transport of the carriers to the cell periphery. The microtubule motors kinesin-1 (KIF5B) and dynein are also important for the delivery of the carriers [120]. A few years later, an unexpected role of Rab6 in the fission of these Rab6 secretory carriers was highlighted [121]. Long tubular carriers connected to the Golgi and seemingly unable to detach have been observed by Live cell imaging when cells are depleted of Rab6A, or its isoform Rab6A'. Pulldown experiments show that Rab6 interacts *in vivo* with myosin II and the direct interaction was observed with purified proteins. Inhibition or depletion of myosin II in different cell types prevents the fission of Rab6 secretory carriers which form long tubules connected to the Golgi apparatus. Therefore, myosin II has an important role at the Golgi in the fission step of secretory carriers. Further investigations show that myosin II works with F-actin filaments for vesicle fission since actin depolymerization leads to the formation of long tubules [121]. Altogether, this work shows that Rab6 is also associated with a subset *tsVSV-G* enriched post-Golgi tubular carriers and unravels a novel function of Rab GTPases in vesicle fission.

The kinesin KIF20A which interacts with Rab6 and myosin II, has a key role in the fission of Rab6-positive vesicles [122]. Long Golgi-attached Rab6 positive tubules can be observed by light microscopy upon either addition of a specific chemical inhibitor of KIF20A motor activity or depletion of KIF20A by siRNA. Rab6 participates in the recruitment of KIF20A to the Golgi complex confining it to growing microtubules at 'Golgi fission hotspots'. Rab6 and KIF20A recruit myosin II which, together with actin, drive fission of Rab6 secretory carriers. The carriers are then transported along microtubules towards the cell surface along microtubules due to the kinesin KIF5B [120,121].

Rab6 secretory carriers transport a large variety of cargos. Using the RUSH system, it was shown that Rab6 secretory carriers contain the anterograde cargos CD59, TNF α , and ColX and Rab6 depletion delays the delivery of these cargo to the plasma membrane [117]. Furthermore, Rab6 depletion in embryonic fibroblast cells reduces global protein secretion by 50%. This together implies that Rab6 is a major regulator of post-Golgi secretion by regulating fission as detailed above. However, Rab6 does not seem to be involved in cargo sorting.

5.4 CtBP1-S/BARS carriers

Brefeldin A (BFA) is a small molecule originally isolated from fungi that has been demonstrated to inhibit the Arf1 exchange factor, GBF1 [123]. As Arf1 is essential to Golgi structure and function, extended incubation with BFA leads to Golgi disassembly [124]. While investigating the molecular factors involved in the Golgi disassembly induced by BFA, the protein CtBP1-S/BARS, also known as CtBP3/BARS (C-terminus binding protein 3/BFA adenosine diphosphate-ribosylated substrate), was identified [125]. Isolated Golgi membranes are fragmented when incubated with either recombinant or rat brain purified CtBP1-S/BARS [126]. Co-incubation of CtBP1-S/BARS on

Golgi membranes with various lipids demonstrated that CtBP1-S/BARS acts as a lysophosphatidic acid acyltransferase to promote generation of phosphatidic acid on the membrane of the Golgi. Microinjection of CtBP1-S/BARS increases the number of post-Golgi carriers. *tsVSV-G* TGN-exit assays show that CtBP1-S/BARS is recruited to the Golgi and associates with *tsVSV-G* enriched post-Golgi carriers [127]. Microinjection of an anti-CtBP1-S/BARS antibody or siRNA experiments against CtBP1-S/BARS blocks the delivery of *tsVSV-G* carriers to the plasma membrane, whereas inhibition of dynamin 2 has no detectable effect [128]. Thus, CtBP1-S/BARS is hypothesised to be essential for membrane fission in a dynamin 2 independent mechanism. As the small GTPase dynamin 2 has been shown to be essential for the fission of Arf1 tubular carriers (section 5.1) and clathrin coated vesicles [129], this is a novel cellular fission mechanism.

14-3-3 γ and PI(4)KIII β co-immunoprecipitate with CtBP1-S/BARS from rat brain cytosol [127]. The resulting complex is stabilised by phosphorylation by the kinases PKD and PAK. CtBP1-S/BARS subsequently activates lysophosphatidic acid acyltransferase δ which produces phosphatidic acid, this process is essential to induce the fission of post-Golgi carriers [130]. In summary, CtBP1-S/BARS is a key component of a protein complex involving 14-3-3 γ , PI(4)KIII β , PKD and PAK, and has an essential role in the fission of *tsVSV-G* enriched post-Golgi carriers.

5.5 Sphingomyelin carriers

The importance of the lipid sphingomyelin (SM) for carrier formation was suggested while investigating lipid-raft formation at the Golgi [131], and confirmed by the identification of a new class of TGN-derived vesicles particularly enriched in that type of lipid [132]. SM, a principal component of the plasma membrane, is synthesised in the Golgi apparatus and transported to the cell surface. A non-toxic reporter, EQ-SM was used to monitor intracellular trafficking of SM along the biosynthetic pathway from the luminal side of the membrane. Proximity biotinylation identified new proteins associated with the SM secretory carriers [133]. EQ-SM (or a control protein that does not bind SM) was fused to the promiscuous biotin ligase APEX2 and expressed in HeLa cells. Cells were permeabilized and incubated with rat liver cytosol and an ATP regenerating system to generate Golgi-derived vesicles. APEX2-mediated biotinylation was then induced on vesicle fractions. Biotin-labelled proteins were isolated and identified by mass spectrometry. SM-rich secretory vesicles contain lysozyme C and GPI-anchored proteins and are highly enriched in Cab45 (calcium-binding protein 45), a luminal Golgi resident protein important for soluble cargo sorting in a calcium dependent manner [134,135].

The Golgi localised protein SPCA1 (Secretory Pathway Calcium ATPase 1), activated by its direct interaction with cofilin-1 and F-actin, pumps calcium into the TGN [134,136,137]. The luminal flux of calcium triggers the oligomerisation of the soluble Golgi localised protein, Cab45 [134,138]. As Cab45 binds to certain secreted cargoes (eg. lysozyme C), in a calcium dependent manner, the oligomerisation leads to sorting into secretory carriers. Interestingly, SiRNA experiments show that Cab45 affects the export of endogenous cargoes such as the cartilage oligomeric protein (COMP) [134]. Additionally, deletion of SPCA1 reduced the number of SM carriers [133] and RUSH assays show that secretion of lysozyme C containing vesicles are significantly reduced in SPCA1 null cells, which can be rescued by recovery of SPCA1. Furthermore, SPCA1 associates

with SM in Golgi membranes and Golgi calcium influx assays show that depletion of SM impairs the TGN calcium uptake by SPCA1. In summary, SPCA1 links sphingomyelin synthesis to calcium/Cab45-dependent cargo sorting in the *trans*-Golgi.

Lipoprotein lipase (LPL), identified in the proximity biotinylation experiment described above [133], is also secreted in SM carriers [139]. LPL binds to Syndecan-1 and gets co-secreted on SM-enriched secretory carriers. Therefore, SDC1 acts as a sorting receptor for LPL at the *trans*-Golgi and directs it into the secretory pathway.

In summary, the packaging of specific secretory proteins into SM rich carriers is mediated by SPCA1 and the subsequent Cab45 oligomerisation. Whether these carriers are the same as others described here requires further investigation and experimentation.

5.6 CARTS

In a series of experiments purifying post-Golgi carriers, a new class of secretory carriers named CARTS (CARriers of the TGN to the cell Surface) was discovered [140]. Partial permeabilisation of HeLa cells and the use of an ATP regenerating system allowed the formation and purification of post-Golgi vesicles. Isolation and analysis of these carriers indicate that CARTS are devoid of collagen and *tsVSV-G* but enriched in endogenous proteins such as pancreatic adenocarcinoma up-regulated factor (PAUF), lysozyme C, synaptotagmin II (SYT2), Rab6A, and Rab8A [140]. Earlier experiments had shown that protein kinase D (PKD) was essential for forming tubular carriers at the *trans*-Golgi [101], as discussed in Section 5.1. Knock-down of PKD resulted in extended CARTS tubules from the Golgi apparatus. Furthermore, vesicle-associated membrane protein-associated proteins (VAP-A/B) regulate CARTS formation upstream of PKD [141]. Interestingly, expression of an OSBP construct (PH-FFAT) which immobilises ER-Golgi contact sites, drastically reduced CARTS production and PAUF secretion [141,142], suggesting a key role of ER-Golgi contact sites in the generation of post-Golgi carriers. Similarly, FAPP1, localised at ER-Golgi contact sites, controls the level of PI(4)P at the *trans*-Golgi and regulates the secretion of the endogenous cargo ApoB100 [143]. Finally, the kinesin protein Eg5, well known for its role in bipolar spindle assembly during mitosis, has a role in the transport of CARTS toward the cell surface along microtubules [144].

CARTS are distinct from COPI, COPII and clathrin carriers and are pleomorphic structures of 100 to 250 nm in diameter [140]. In summary, CARTS constitute a totally distinct class of carriers destined to the cell surface as they exclude *tsVSV-G*. However, several cargo proteins sorted by Cab45 (like lysozyme C) are associated with CARTS [145] raising the question: how similar are these carriers to SM secretory carriers discussed above? Further investigation is essential to fully understand what distinguishes these carriers from the other ones.

6. Fusion Machinery

The final event of a protein trafficking pathway is the fusion of the carrier with the distal compartment. This involves membrane tethering ('recognition') and membrane fusion to allow soluble content mixing and membrane delivery. In the case of the post-Golgi carrier and the

plasma membrane there are a number of candidate tethering factors (see summary in figure 3), it is currently unclear if the machinery for carrier fusion is distinct for constitutive and regulated secretion.

6.1 ELKS

The large coiled-coil protein ELKS, also called CAST2, Rab6IP2, or ERC1 [146–149], is involved in synaptic vesicle exocytosis based on purification of highly exocytic synaptic junctions [150,151]. TIRF imaging in insulin-producing cells shows ELKS foci on the exocytic fusion pore, supporting a role in regulated secretion [152,153]. ELKS was identified as a Rab6 interactor [147]. An accumulation in Rab6 positive vesicular structures was observed after ELKS depletion [120], later shown to be positive for MICAL3 and Rab8 [154]. MICAL3 and Rab8 interact with each other and MICAL3 with ELKS, hinting at a functional complex for exocytosis. ARHGEF10, an exchange factor for RhoA has been shown to localise to Rab6/Rab8 positive carriers and is required for Rab8 recruitment suggesting that there is a small G protein cascade leading to carrier fusion [155,156]. The soluble biosynthetic cargo neuropeptide Y accumulates in pre-fusion ELKS dependent carriers, suggesting that this is the biosynthetic transport route [120]. ELKS and RAB6 are on plasma membrane ‘hotspots’ which are the fusion points of the secretory carriers [117].

6.2 Exocyst

Another obvious candidate for the fusion of the carriers at the plasma membrane is the exocyst complex. Exocyst is an octameric protein complex, with six of the subunits (Sec6, Sec8, Sec15, Sec3, Sec5 and Sec10) discovered in the original *Sec* yeast genetic screen and later characterised as a complex [23,157–159]. The remaining subunits (Exo70 and Exo84) were discovered subsequently as *bona fide* members of the complex [160–162]. Exocyst is important for exocytic transport (hence the name exocyst) based on the localisation in budding yeast and the *Sec* phenotype.

In non-polarised mammalian cells exocyst components localise to the plasma membrane and Golgi apparatus [163]. *tsVSV-G* colocalises with exocyst in the Golgi stacks. It is not clear, however, if it colocalises to the post-Golgi *tsVSV-G* carriers or at the fusion point of the carriers. Exocyst inhibition with antibodies does not affect VSV-G delivery [163,164]. siRNA depletion of Exo70 decreased the efficiency but did not completely stop *tsVSV-G* delivery [165]. Colocalisation of *tsVSV-G* and Sec8 on the plasma membrane using TIRF microscopy showed only 20% overlap between the two markers [166]. To-date it is not clear if exocyst is directly involved in fusion of biosynthetic carriers to the plasma membrane.

6.3 SNAREs

The final membrane fusion step of the membrane of pathways is mediated by SNARE proteins. SNAREs interact to form four-helix bundles to promote membrane fusion, with each bundle requiring R-, Qa-, Qb-, and Qc-SNARE motifs [167]. *tsVSV-G* positive post-Golgi carriers accumulate upon SNARE inhibition by abrogation of the protein NSF [75]. As NSF is essential for SNARE function these experiments demonstrated that SNARE machinery is essential for fusion of the carriers. A targeted siRNA screen identified the specific SNAREs and SM (Sec1/Munc18-

like) proteins which help to organise the SNAREs [168]. 38 SNAREs, 4 SNARE- like proteins and 7 SM proteins were targeted, 9 of which were identified and validated as being strongly inhibitory to secretory exocytosis. These 9 hits included proteins for both the ER to Golgi transport step and Golgi to plasma membrane step. Candidate validation identified the SNAREs SNAP29 and syntaxin-19. SNAP29 has both a Qb- and Qc-SNARE domain [169], and syntaxin-19 is Qa-SNARE [170]. Depletion of both syntaxin-19 and SNAP29 decreased plasma membrane fusion events of secretory vesicles, additionally in SNAP29 depleted cells there was an accumulation of pre-fusion carriers in the cytosol. Surprisingly no R-SNAREs were identified in this screen [168].

In a subsequent targeted screen *Drosophila* cells were used, which have less genomic redundancy than mammalian cell lines. Results from the screen highlighted the R-SNAREs YKT6 and VAMP3 as important for fusion of post-Golgi secretory carriers, consistent with mammalian cells [171], although YKT6 is a promiscuous SNARE that also has a role in ER to Golgi transport [168]. In summary, one of the SNARE complexes for the secretory route to the plasma membrane requires SNAP29 and syntaxin-19 and perhaps VAMP3 redundantly with YKT6. There are likely multiple SNARE complexes for this route and multiple redundancies.

7. Importance of the lipid PI(4)P in post-Golgi trafficking

Lipids play a central role in the budding of carriers from the *trans*-Golgi (reviewed in [145]). One of the major lipids in this process is the phosphoinositide PI(4)P [105] see also section 5.1 above. PI(4)P is crucial for post-Golgi vesicle biogenesis including carriers destined directly for the cell surface. Enriched on the cytosolic outer membrane bilayer, PI(4)P can recruit an array of specific-binding proteins.

7.1 PI(4)KIII β

Mammals encode four different PI 4-kinases and PI(4)KIII β is involved in the formation of *tsVSV*-G carriers destined for the plasma membrane [107,127]. PI(4)KIII β is recruited to the Golgi by different mechanisms [105] including by the action of Arf1 [172] and the Golgi localised ACBD3 [173]. Once at the *trans*-Golgi, PI(4)KIII β generates PI(4)P that recruits further proteins such as GOLPH3 and lipid transport proteins as discussed below. PI(4)KIII β is also involved in the recruitment of the small G protein Rab11a [174,175] shown to be important for *tsVSV*-G trafficking from the *trans*-Golgi to the plasma membrane [176]. PI(4)KIII β was shown to have an important role in mitochondrial scission generating PI(4)P on *trans*-Golgi derived vesicles after recruitment by Arf1 [177]. These vesicles are targeted to mitochondria-ER contact sites and are thought to drive mitochondrial division downstream of Drp1 (Dynamin-related protein-1).

7.2 GOLPH3

One example of a PI(4)P mediated binding is GOLPH3, discovered via a proteomic lipid-binding screen, which is recruited to the *trans*-Golgi by interacting with PI(4)P [178]. GOLPH3 binds to the unconventional myosin MYO18A thus connecting the Golgi to F-actin. The model suggests that this complex applies tension to extract the carriers at the *trans*-Golgi. Knock-down of

GOLPH3 or MYO18A or actin depolymerisation impairs trafficking of *tsVSV-G* from the Golgi to the plasma membrane. Additionally, GOLPH3 induces membrane curvature, by inserting a loop into the proximal leaflet of the bilayer and this property is required for efficient *trans*-Golgi to plasma membrane trafficking [179]. However, GOLPH3 has also been shown to specifically bind and recycle a subset of Golgi enzymes involved in glycan assembly on sphingolipids [180]. This role of GOLPH3 matches the role of its yeast homolog Vps74 which binds to PI(4)P at the *trans*-Golgi and interacts with specific Golgi enzymes allowing their retrograde transport [181,182].

7.3 Lipid transport proteins

PI(4)P recruits a range of lipid transport proteins (reviewed in [183–185]), such as FAPP2, oxysterol-binding protein (OSBP), and ceramide transfer protein (CERT). These proteins share a similar domain organisation: an N-terminal PH domain that binds PI(4)P (and simultaneously Arf1-GTP for OSBP and FAPP2), a C-terminal FFAT motif that binds the ER membrane protein VAP and a C-terminal lipid transport domain. FAPP2 transfers GlcCer to the *trans*-Golgi [112] as described in section 5.1 above. At membrane contact sites, OSBP directs cholesterol transfer from the ER to the *trans*-Golgi through coupled counter-transport of PI(4)P [142,186]. CERT transports ceramide from the ER to the *trans*-Golgi [187], thereby promoting SM and diacylglycerol synthesis. Sterols and sphingolipids, which are suggested to assemble into lipid microdomains are important for the formation of secretory carriers [132,188]. In agreement, depletion of VAP inhibits *trans*-Golgi to plasma membrane transport of *tsVSV-G* [189]. Similarly, VAP knockdown, double knockdown of CERT/OSBP or the treatment of the cells with D-ceramide-C6, impairs the secretion of the cargo PAUF [141].

Another lipid transfer protein that functions at contact sites between the ER and the *trans*-Golgi is Nir2 [184]. Nir2 supplies the *trans*-Golgi with PI in exchange of phosphatidylcholine and thereby participates in PI(4)P synthesis and diacylglycerol homeostasis [190]. As a result, depletion of Nir2 affects the transport of *tsVSV-G* from the *trans*-Golgi to the cell surface. Similarly, two other PI transfer proteins, PITPNA and PITPNB, stimulate the production of PI(4)P which promotes the recruitment of GOLPH3 and CERT to the Golgi thus facilitating the apical targeting of membrane trafficking in neural stem cells [191].

Overall, lipid transfer proteins by executing non-vesicular lipid exchange are essential for the remodelling of the lipid landscape and, as a result, ER-Golgi contact sites could represent key sites for secretory cargo sorting and membrane fission.

7.4 Importance of PI(4)P in polarised secretion and cargo sorting in yeast

In yeast, PI(4)P has a key role in the generation of secretory vesicles during polarized growth. The PI(4)-kinase Pik1, responsible for PI(4)P synthesis at the *trans*-Golgi, regulates several transport events including the Golgi to surface route [192]. Together with the activated RabGTPase Ypt32, PI(4)P recruits the exchange factor Sec2 to the *trans*-Golgi which in turn activates another Rab, Sec4 [193,194]. The secretory vesicles subsequently pinch off and Sec4 recruits the exocyst component Sec15 to promote the tethering and fusion of the vesicles with the plasma membrane [195,196]. PI(4)P, Ypt32-GTP, Sec4-GTP and then Sec15 recruit Myo2, a type V myosin, to allow the transport of vesicles along actin cables toward the exocytic site [197–199].

Additionally, Sec2 is able to bind directly to the Sec4 effector Sec15 [200]. Sec15 and Ypt32-GTP compete for binding to Sec2. Also, the association of Sec2 with its alternate binding partners is regulated by PI(4)P [193]. Thus, when secretory vesicles are initially formed and enriched in PI(4)P, Sec2 interacts with its initial binding partner Ypt32-GTP. It was shown that the concentration of PI(4)P on secretory vesicles is reduced by the time of their arrival at the cell surface [193,201] by the action of the protein Osh4 [201]. This suggests that during the maturation of secretory vesicles, as the PI(4)P concentration drops, Ypt32-GTP is replaced on Sec2 by Sec15 leading to a positive feedback loop and thus preparing the vesicles for docking and fusion with the plasma membrane.

Furthermore, an additional regulatory level is accomplished by the phosphorylation of Sec2 by casein kinases Yck1/Yck2 [202,203]. Sec2 phosphorylation inhibits its interaction with its initial binding partners PI(4)P and Ypt32-GTP and promotes its interaction with its downstream effector Sec15. Additionally, Yck2 binds to Sec2 near the PI(4)P binding region and the presence of PI(4)P inhibits phosphorylation of Sec2 by Yck2 *in vitro* [202,203]. This suggests that phosphorylation of Sec2 is initially prevented by the high level of PI(4)P in nascent secretory vesicles. As the secretory vesicle matures and the PI(4)P level drops, Sec2 gets phosphorylated promoting the switch in Sec2 binding partners and thus pushing the reaction forwards. Altogether, this complex regulatory circuit helps to impose a directionality in vesicular transport and shows the key role of PI(4)P in the secretory route. This Rab cascade is somewhat conserved in mammalian cells and involves the Ypt32-homologue Rab11 recruiting the exchange factor Rabin8 to activate the Sec4-homologue Rab8 [204]. There are some differences, however, for example the lipid phosphatidylserine participating in the Golgi recruitment of Rabin8 in mammalian cells instead of PI(4)P in yeast [205].

Glucose induces a change in intracellular pH that governs the protonation state of PI(4)P at the *trans*-Golgi [206]. This regulates recruitment of Osh1, a member of the OSBP family of lipid transfer proteins, which promotes the trafficking of the Tat2 amino acid permease to the plasma membrane regulating tryptophan uptake. This study shows that pH biosensing by PI(4)P in response to nutrient availability regulates cargo sorting at the *trans*-Golgi.

8. Open Questions

At this stage, we still do not have a clear picture of the exact molecular machinery of the pleomorphic sorting carriers that bud from the Golgi apparatus and new types of carriers are emerging [207]. This is complicated by the fact that these carriers are transient and contain low cargo abundance. Accordingly, they have mostly been observed using temporal kinetic assays, such as *tsVSV-G* and RUSH. Many of the cargos used are exogenous proteins, including non-mammalian proteins such as GFP and HRP fused to a signal peptide and human proteins that are not expressed normally in that cell type. These techniques are essential and have facilitated fundamental discoveries over the years. It is important, although technically challenging, to investigate these carriers using native proteins under more physiological conditions.

Mutations in genes encoding for proteins involved in the *trans*-Golgi export machinery have been shown to cause severe genetic diseases [66,208]. For example, mutations in different subunits of the AP-1-complex, component of clathrin-coated vesicles and associated with the *trans*-Golgi and early/recycling endosomes, are linked with X-linked mental retardation and with MEDNIK syndrome (Mental retardation, Enteropathy, Deafness, peripheral Neuropathy, Ichthyosis and Keratoderma). Other genetic diseases have been linked to mutations in AP subunits [66]. Therefore a better understanding of the mechanistic basis of these pathologies is essential.

There are also a number of diseases associated with secreted proteins. One example is transthyretin (TTR) amyloidosis: a rare genetic disease caused by over 80 known mutations in the secreted protein TTR [209], resulting in protein aggregation into amyloid fibrils [210]. Monoclonal gammopathies, caused by an abundance of a particular immunoglobulin into the blood (IgM, IgG, IgE or a combination), are another example of a disease associated with protein secretion [211,212]. Our current understanding of the machinery is still incomplete and it is possible that diseases associated with mutations in this pathway are yet to be genetically characterised.

How many routes are there from the Golgi apparatus to the plasma membrane? It is possible that *tsVSV-G* takes multiple routes from the Golgi apparatus (fig, 2), which makes the number of routes that leave the Golgi hard to interpret. Similarly, glycosylphosphatidylinositol-anchored proteins (GPI-APs) traffic to the apical surface of most epithelial cells, however, there are some exceptions such as Fischer rat thyroid cells where GPI-APs are sorted to the basolateral surface [213]. In figure 2, the schematic shows the various carriers that have been shown experimentally. It is of course possible, for example, that the Rab6 carriers and Lamp1-RUSH carriers shown in the image are the same. One reason for multiple pathways from the Golgi apparatus to the plasma membrane is due to polarised sorting in some cell types. Examples of polarised cells are epithelial tissues such as the choroid plexus that makes up a portion of the blood-brain barrier and is responsible for the secretion of cerebrospinal fluid (CSF). Neurons, another polarised cell type, are characterised by a long axonal process, reaching a meter in length in humans.

Polarised cells need to maintain differential protein trafficking in the cellular subdomains, for example, the choroid plexus polarised epithelial layer faces fenestrated capillaries on the basolateral side and the ventricular space on the apical side where the CSF is secreted. The generation, maintenance and trafficking of cell polarity has been comprehensively reviewed elsewhere [214–216]. The machinery that mediates the sorting into the two different routes is still being characterised. It seems likely that some of the machinery discussed in this review is essential, for example loss of FAPP2 in MDCK cells causes loss of complete polarisation and defective cilium formation and is important for delivery of apical cargos [111,217]. It is possible that the routes that exist in polarised cells also exist in ‘non-polarised’ cells and act as redundant or alternative routes to the plasma membrane, as demonstrated in comparisons of trafficking non-polarised Fao cells with polarised WIF-B cells [218].

A final outstanding question is the presence of a ‘coat’. All other known carriers characterised to-date have a coat of some sort. In the case of clathrin coated vesicles it is a geometric lattice that

directly forms and scaffolds the forming vesicles, in the retromer coat it is a core complex that allows for cargo selectivity and binding to other machinery. Although, as discussed above, there are some candidate proteins, there is still no *bona fide* coat for the secretory carriers. Perhaps there is no need for a coat in these carriers or perhaps it has yet to be discovered. With the recent advances in genetics and proteomics there is surely more to be discovered on this elusive trafficking pathway.

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Legends

Figure 1: Destinations of proteins from the Golgi apparatus. Proteins arrive into the Golgi apparatus from either the ER (bottom of the schematic) or endolysosomal system (top of the schematic). After arrival proteins can either return to the ER in COPI vesicles (light blue), can be retained in the Golgi apparatus, can be delivered directly to the plasma membrane (yellow) or delivered to the endolysosomal system by either clathrin (red) or other clathrin independent mechanism (eg, AP4, green).

Figure 2. Proteins associated with tubular carriers in mammalian cells. Models from experimental data on different types of secretory tubular carriers that bud towards the plasma membrane in mammalian cells. The carriers have been drawn as separate entities unless there is strong experimental evidence to combine the carriers. Some of the machinery described here might be overlapping on the same carriers, for example the SM carriers and CARTS, and the Lamp1-RUSH carriers, Rab6 carriers and the Arf1 carriers. Protein cartoons are not to scale and do not meaningfully reflect protein structure. LYZ = Lysozyme C.

Figure 3. Fusion of post-Golgi carriers in mammalian cells. There are 2 tethering complexes associated with post-Golgi carriers, ELKS (left) and exocyst (center). The SNAREs associated with the final fusion event are shown on the right. YKT6 and VAMP3 appear to work either redundantly or as two parallel pathways [171]. Soluble cargo represents bulk flow as assayed by exogenously expressed cargos (eg. [168]).

Figure 1

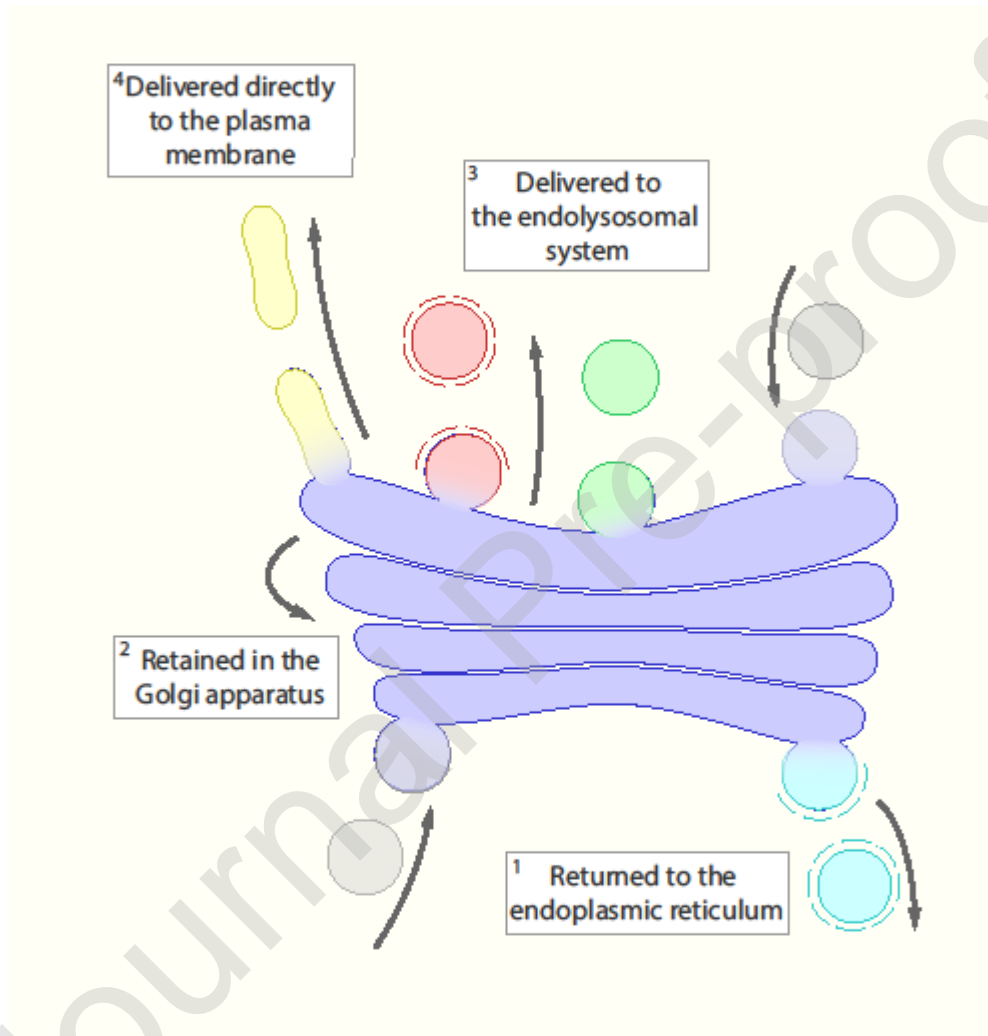


Figure 2

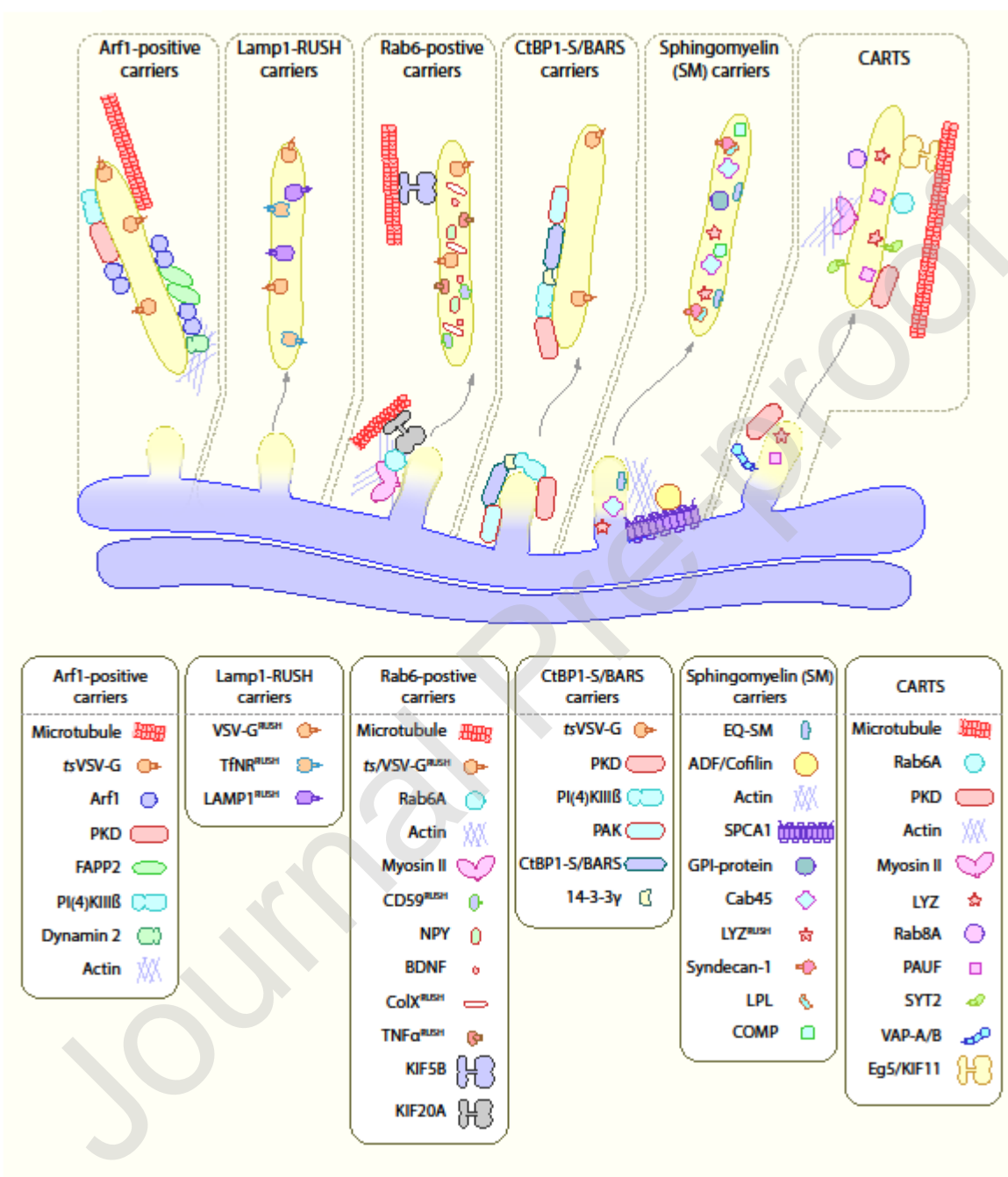


Figure 3

