



Johnson, J. L. (2021). The predictive potential of circulating microRNA for future cardiovascular events. *Cardiovascular Research*, 117(1), 1-3. [cvaa145]. <https://doi.org/10.1093/cvr/cvaa145>

Peer reviewed version

Link to published version (if available):
[10.1093/cvr/cvaa145](https://doi.org/10.1093/cvr/cvaa145)

[Link to publication record in Explore Bristol Research](#)
PDF-document

This is the author accepted manuscript (AAM). The final published version (version of record) is available online via Oxford University Press at <https://academic.oup.com/cardiovascres/advance-article/doi/10.1093/cvr/cvaa145/5839744?searchresult=1>. Please refer to any applicable terms of use of the publisher.

University of Bristol - Explore Bristol Research

General rights

This document is made available in accordance with publisher policies. Please cite only the published version using the reference above. Full terms of use are available: <http://www.bristol.ac.uk/red/research-policy/pure/user-guides/ebr-terms/>

The predictive potential of circulating microRNA for future cardiovascular events

Dr Jason L Johnson

Laboratory of Cardiovascular Pathology, Bristol Medical School, Faculty of Health Sciences, University of Bristol, England

Correspondence to Dr Jason L. Johnson, PhD, Laboratory of Cardiovascular Pathology, Bristol Medical School, Faculty of Health Sciences, University of Bristol, Level 7, Bristol Royal Infirmary, Bristol BS2 8HW, England.

E-mail Jason.L.Johnson@bristol.ac.uk.

Phone 0044 (0)117 3423583

Non-coding RNAs have received considerable attention within the cardiovascular research field over the last decade, especially the short non-coding RNA class commonly termed as microRNA (miRNA, miR). MicroRNAs are approximately 18–22 nucleotides in length and predicted to post-transcriptionally control 60–90% of protein-coding genes through inhibiting translation or promoting degradation of target messenger (m)RNA^{1 2}. Importantly, a single microRNA can potentially target a large number of genes¹, explaining why there are far less microRNAs than target genes, and there is evidence that multiple targets exist within the same functional networks. As such, mature microRNA possess the potential to play a central role in regulating cellular function, while their spatial and temporal expression patterns provide clues to their function^{1 2}. Accordingly, microRNAs are considered fine-tuners of gene and protein expression profiles during the progression of atherosclerosis, a proposition supported by human pathological studies alongside findings from diseased animal models where select microRNA function or expression have been modulated³. Such approaches identified multiple microRNAs which may serve as therapeutic targets for the prevention of clinical events associated with atherosclerotic plaque destabilisation, such as myocardial infarction and stroke³.

However, a marked proportion of patients who suffer a major adverse cardiovascular event (MACE) will have been previously asymptomatic, and even patients with confirmed clinically-relevant atherosclerosis and receiving optimal medical therapy, will experience additional future clinical events⁴. There is consequently a pertinent need for biomarkers which can readily inform a patient's risk of experiencing atherosclerotic plaque rupture and a subsequent clinical event. Relatedly, stable microRNAs have been detected within circulating blood, either conjugated with lipoproteins, packaged within extracellular vesicles (such as microvesicles and exosomes), or as a result of cell leakage during cell injury or apoptosis⁵. Moreover, circulating microRNA levels are associated with cardiovascular disease risk factors including hyperlipidaemia, and there are examples of specific miRNAs correlating with atherosclerosis and myocardial infarction³.

In this issue of Cardiovascular Research, Escate and colleagues⁶ examine if microRNA can be deployed as predictors of atherosclerotic plaque progression and related clinical events in patients with familial hypercholesterolaemia (FH), although potentially applicable to all individuals with atherosclerosis. The authors profiled microRNAs within exosomes isolated from circulating blood of patients with FH which had suffered a major adverse cardiovascular event (MACE) during an 8 year follow-up, alongside relatives without genetic diagnosis of FH and MACE, all participants within the SAFEHEART cohort⁷. This discovery approach identified 42 differentially expressed microRNAs which were subsequently subjected to pathway analysis and revealed 17 exosome-isolated microRNAs which were related to bioinformatically-defined atherosclerosis processes, such as 'inflammatory response'. Further RT-PCR validation of the 17 identified microRNAs was undertaken within exosomes, microvesicles, and plasma from a larger cohort of patients. This validation stage elucidated 9 microRNAs which were consistently increased within the plasma and exosome fraction of FH patients which had experienced MACE during

an 8-year follow-up compared to non-FH individuals with no evidence of MACE. Due to the correlation between plasma microRNA concentrations and those detected within exosomes, the authors propose plasma microRNA levels can serve as a surrogate for exosome microRNA content.

Subsequently, the 9 identified microRNAs were retrospectively compared within the plasma of FH patients with MACE, without MACE, and non-FH/non-MACE individuals subjected to 8-year follow-up. While this refinement stage revealed elevated expression of all 9 microRNAs were discriminatory for FH patients, only significantly increased miR-133a plasma levels identified FH patients experiencing MACE. Further analysis supported the predictive power of miR-133a (alongside miR-200c and miR-339-3p) for incidence of future cardiovascular events, including sudden death, fatal and non-fatal myocardial infarction, unstable angina and cerebrovascular incidents, during 5, 8, and 10-year follow-up periods after entering the SAFEHEART cohort, and independent of other cardiovascular risk factors. The authors next deployed *in silico* analysis to identify 923 potential target genes of miR-133a, 42 of which were delineated in a biased fashion due to their association with cardiovascular disease related processes identified using bioinformatics software analysis. Through further *in silico* analysis, the transcription factors FOXL2 and DNAJB6 alongside the membrane receptor CD130 (a unit of the IL-6 receptor complex) were predicated, implying miR-133a regulates the Wnt/ β -catenin and IL-6 signalling pathways, respectively. Substantiating these propositions, the authors demonstrated increasing miR-133a exogenously in primary human macrophages or microvascular endothelial cells elevated mRNA expression of both CTNNB1 and IL6R.

The study by Escate and colleagues⁶ affords a further microRNA to add to the armamentarium of potential biomarkers for atherosclerosis progression and risk of an associated clinical event (as summarised within Figure and reviewed by³). Although the present study focussed on FH patients (the most common genetic disorder associated with premature atherosclerosis-related cardiovascular disease), the findings have likely broader utility to all individuals with atherosclerosis. Supporting evidence is provided by similar microRNA profiling clinical studies revealing that miR-133a plasma levels are increased in patients with angiographic-defined coronary artery disease (CAD) when compared to healthy subjects⁸ and those experiencing an acute myocardial infarction (AMI) relative to CAD patients without an AMI or healthy individuals⁹. In addition, intra-plaque concentrations of miR-133a are increased in carotid lesions deemed symptomatic relative to asymptomatic plaques¹⁰, indicating that elevated circulating levels of miR-133a are likely to derive from vulnerable and ruptured plaques.

However, miR-133a is considered a cardiac-specific microRNA primarily expressed within cardiomyocytes¹¹, and the elevated levels detected post myocardial infarction are speculated to originate from injured and dying cardiomyocytes^{9, 12, 13}. While in accordance with the study from Escate et al⁶, circulating levels of miR-133a display prognostic value for all-cause mortality in CAD patients which present with AMI, such predictive capacity correlated with high-sensitivity troponin T levels and therefore the extent of myocardial injury¹². Escate and colleagues do not clarify the cellular or tissue source of the elevated miR-133a detected within their study or define if the association between raised miR-133a levels and future MACE are causal. While they infer the increase is plaque-dependent, the above-mentioned studies argue otherwise and even suggest microRNAs released from the damaged myocardium after an AMI track to coronary plaques to exert subsequent biological effects¹³. An alternate hypothesis is that miR-133a is released from coronary plaques after rupture (not causal) and during their subsequent healing, a phenomenon known to be prevalent within culprit coronary atherosclerotic lesions¹⁴. This possibility would question the applicability of miR-133a as a therapeutic target, as retarding vascular smooth muscle cell growth and ensuing plaque healing, including the reformation of the protective fibrous cap, may accelerate another adverse clinical event. Indeed, 13% of individuals within the SAFEHEART cohort have evidence of previous clinical premature atherosclerosis⁷, while mouse studies have revealed a central role for miR-133a in vascular smooth muscle cell growth¹⁵.

The current study from Escate and colleagues alongside other research reveals miR-133a as a potential surrogate marker of previous plaque rupture and subsequent AMI, representing a vulnerable patient with a preponderance for future clinical events, particularly those with overt risk factors such as FH. Additional studies are warranted to identify the source of circulating miR-133a and its potential distal biological function, particularly if it is to be considered as a therapeutic target for cardiovascular disease prevention. Finally, larger profiling studies to substantiate miR-133a as a bona-fide biomarker for disease progression should be encouraged, alongside other circulating microRNAs proposed as prognostic biomarkers (see Figure).

ARTICLE INFORMATION

Affiliation

From the University of Bristol, United Kingdom.

Acknowledgments

None.

Sources of Funding

J.L. Johnson is supported by the British Heart Foundation (FS/18/1/33234).

Disclosures

None.

REFERENCES

1. Kozomara A, Birgaoanu M, Griffiths-Jones S. miRBase: from microRNA sequences to function. *Nucleic Acids Res* 2018;**47**:D155-D162.
2. Ha M, Kim VN. Regulation of microRNA biogenesis. *Nat Rev Mol Cell Biol* 2014;**15**:509-524.
3. Fasolo F, Di Gregoli K, Maegdefessel L, Johnson JL. Non-coding RNAs in cardiovascular cell biology and atherosclerosis. *Cardiovasc Res* 2019;**115**:1732-1756.
4. Cohn JN. The Message is Clear: Prevent as Well as Treat Acute Myocardial Infarction. *Circulation* 2013.
5. Théry C, Ostrowski M, Segura E. Membrane vesicles as conveyors of immune responses. *Nat Rev Immunol* 2009;**9**:581-593.
6. Escate R, Padró T, Suades R, Camino S, Muñoz O, Diaz-Diaz JL, Sionis A, Mata P, Badimon L. High miR-133a levels in the circulation anticipates presentation of clinical events in familial hypercholesterolaemia patients. *Cardiovasc Res* 2020;**XX**:XXX-XXX.
7. Perez de Isla L, Alonso R, Watts GF, Mata N, Saltijeral Cerezo A, Muñoz O, Fuentes F, Diaz-Diaz JL, de Andrés R, Zambón D, Rubio-Marin P, Barba-Romero MA, Saenz P, Sanchez Muñoz-Torrero JF, Martinez-Faedo C, Miramontes-Gonzalez JP, Badimón L, Mata P, Aguado R, Almagro F, Arrieta F, Barba MÁ, Brea Á, Cepeda JM, De Andrés R, Díaz G, Díaz JL, Fuentes F, Galiana J, Garrido JA, Irigoyen L, Manjón L, Martin A, Piedecausa M, Martínez-Faedo C, Mauri M, Miramontes P, Muñoz O, Pereyra F, Pérez L, Pintó X, Pujante P, Ruiz E, Sáenz P, Sánchez JF, Vidal JJ, Argüeso R, Zambón D. Attainment of LDL-Cholesterol Treatment Goals in Patients With Familial Hypercholesterolemia: 5-Year SAFEHEART Registry Follow-Up. *J Am Coll Cardiol* 2016;**67**:1278-1285.
8. Fichtlscherer S, De Rosa S, Fox H, Schwietz T, Fischer A, Liebetrau C, Weber M, Hamm CW, Roxel T, Muller-Ardogan M, Bonauer A, Zeiher AM, Dimmeler S. Circulating MicroRNAs in Patients With Coronary Artery Disease. *Circ Res* 2010;**107**:677-684.
9. Wang G-K, Zhu J-Q, Zhang J-T, Li Q, Li Y, He J, Qin Y-W, Jing Q. Circulating microRNA: a novel potential biomarker for early diagnosis of acute myocardial infarction in humans. *Eur Heart J* 2010;**31**:659-666.
10. Cipollone F, Felicioni L, Sarzani R, Uchino S, Spigonardo F, Mandolini C, Malatesta S, Bucci M, Mammarella C, Santovito D, de Lutiis F, Marchetti A, Mezzetti A, Buttitta F. A Unique MicroRNA Signature Associated With Plaque Instability in Humans. *Stroke* 2011;**42**:2556-2563.
11. Carè A, Catalucci D, Felicetti F, Bonci D, Addario A, Gallo P, Bang M-L, Segnalini P, Gu Y, Dalton ND, Elia L, Latronico MVG, Høydal M, Autore C, Russo MA, Dorn GW, Ellingsen Ø, Ruiz-Lozano P, Peterson KL, Croce CM, Peschle C, Condorelli G. MicroRNA-133 controls cardiac hypertrophy. *Nat Med* 2007;**13**:613-618.
12. Widera C, Gupta SK, Lorenzen JM, Bang C, Bauersachs J, Bethmann K, Kempf T, Wollert KC, Thum T. Diagnostic and prognostic impact of six circulating microRNAs in acute coronary syndrome. *J Mol Cell Cardiol* 2011;**51**:872-875.
13. Rosa SD, Fichtlscherer S, Lehmann R, Assmus B, Dimmeler S, Zeiher AM. Transcoronary Concentration Gradients of Circulating MicroRNAs. *Circulation* 2011;**124**:1936-1944.
14. Burke AP, Kolodgie FD, Farb A, Weber DK, Malcom GT, Smialek J, Virmani R. Healed plaque ruptures and sudden coronary death - Evidence that subclinical rupture has a role in plaque progression. *Circulation* 2001;**103**:934-940.
15. Torella D, Iaconetti C, Catalucci D, Ellison GM, Leone A, Waring CD, Boicchio A, Vicinanza C, Aquila I, Curcio A, Condorelli G, Indolfi C. MicroRNA-133 Controls Vascular Smooth Muscle Cell Phenotypic Switch In Vitro and Vascular Remodeling In Vivo. *Circ Res* 2011;**109**:880-893.

FIGURE LEGEND

Figure title: MicroRNA expression within circulating blood of patients with atherosclerosis.

This diagram illustrates the elevated microRNAs identified through profiling approaches within circulating plasma samples (including exosomes) and peripheral blood mononuclear cells (PBMCs) of patients with clinical atherosclerosis. Blue-filled box indicates intra-plaque cellular sources of circulating microRNA. Green-filled box signifies the forms in which circulating microRNA are packaged and potential other tissue sources. Line-coloured boxes indicate the patient cohorts from within which the associated upregulated microRNAs were identified. Yellow highlighting shows elevated miR-133a has been verified in an independent study, while microRNA with grey highlighting have been independently reported to be up- and down-regulated.

