



Muscle-Type Nicotinic Receptor Blockade by Diethylamine, the Hydrophilic Moiety of Lidocaine

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OPEN ACCESS

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Received: 11 December 2015

Accepted: 29 January 2016

Published: 15 February 2016

Citation:

Alberola-Die A, Fernández-Ballester G, González-Ros JM, Ivorra I and Morales A (2016) Muscle-Type Nicotinic Receptor Blockade by Diethylamine, the Hydrophilic Moiety of Lidocaine.
Front. Mol. Neurosci. 9:12.
doi: 10.3389/fnmol.2016.00012

Lidocaine bears in its structure both an aromatic ring and a terminal amine, which can be protonated at physiological pH, linked by an amide group. Since lidocaine causes multiple inhibitory actions on nicotinic acetylcholine receptors (nAChRs), this work was aimed to determine the inhibitory effects of diethylamine (DEA), a small molecule resembling the hydrophilic moiety of lidocaine, on *Torpedo marmorata* nAChRs microtransplanted to *Xenopus* oocytes. Similarly to lidocaine, DEA reversibly blocked acetylcholine-elicited currents (I_{ACh}) in a dose-dependent manner (IC_{50} close to 70 μ M), but unlike lidocaine, DEA did not affect I_{ACh} desensitization. I_{ACh} inhibition by DEA was more pronounced at negative potentials, suggesting an open-channel blockade of nAChRs, although roughly 30% inhibition persisted at positive potentials, indicating additional binding sites outside the pore. DEA block of nAChRs in the resting state (closed channel) was confirmed by the enhanced I_{ACh} inhibition when pre-applying DEA before its co-application with ACh, as compared with solely DEA and ACh co-application. Virtual docking assays provide a plausible explanation to the experimental observations in terms of the involvement of different sets of drug binding sites. So, at the nAChR transmembrane (TM) domain, DEA and lidocaine shared binding sites within the channel pore, giving support to their open-channel blockade; besides, lidocaine, but not DEA, interacted with residues at cavities among the M1, M2, M3, and M4 segments of each subunit and also at intersubunit crevices. At the extracellular (EC) domain, DEA and lidocaine binding sites were broadly distributed, which aids to explain the closed channel blockade observed. Interestingly, some DEA clusters were located at the α - γ interphase of the EC domain, in a cavity near the orthosteric binding site pocket; by contrast, lidocaine contacted with all α -subunit loops conforming the ACh binding site, both in α - γ and α - δ and interphases, likely because of its larger size. Together, these results indicate that DEA mimics some, but not all, inhibitory actions of lidocaine on nAChRs and that even this small polar molecule acts by different mechanisms on this receptor. The presented results contribute to a better understanding of the structural determinants of nAChR modulation.

Keywords: diethylamine, lidocaine, nicotinic acetylcholine receptors, *Xenopus* oocytes, microtransplanted receptors, allosteric modulation

INTRODUCTION

Nicotinic acetylcholine receptors (nAChRs) belong to the “Cys-loop” superfamily of ligand-gated ion channels (LGICs). All members of this family of receptors are constituted by five subunits, each one contributing four transmembrane spanning-segments (M1–M4), which conform a channel pore lined by the M2 segment of each subunit (Albuquerque et al., 2009; Hurst et al., 2013). nAChRs are widely distributed in central and peripheral nervous systems, but they are also expressed in skeletal muscle fibers and other tissues where they play relevant functional roles (Gotti and Clementi, 2004). Although there is a large heterogeneity in the structure and function of nAChRs from different cells, all of them behave as allosteric proteins, undergoing conformational changes (from resting to active or desensitized states) when specific ligands bind to their orthosteric sites (Taly et al., 2009; Cecchini and Changeux, 2015).

nAChRs are involved in the etiopathology of several neurological disorders, including Parkinson’s disease, some myasthenic syndromes, addiction, depression, cognitive deficits (including Alzheimer’s disease), some types of epilepsy and also seem to contribute to modulate pain or inflammatory responses (Gotti and Clementi, 2004; Dani and Bertrand, 2007; Hurst et al., 2013). Hence, it is important to study the mechanisms of modulation of these receptors by allosteric ligands and to unravel their specific binding sites, in order to develop new therapeutic agents (Arias, 2010; Chatzidaki and Millar, 2015).

Lidocaine is commonly used in clinical practice as a local anesthetic and as an anti-arrhythmic agent because it reversibly blocks voltage-dependent Na⁺ channels (Hille, 1966). Besides, lidocaine modulates the function of other voltage-dependent channels, including Ca²⁺ (Xiong and Strichartz, 1998) and K⁺ channels (Trellakis et al., 2006) and different pentameric LGICs, such as 5-hydroxytryptamine-3 (Ueta et al., 2007), glycine (Gly) and GABA_A receptors (Hara and Sata, 2007), nAChRs (Steinbach, 1968; Gentry and Lukas, 2001; Alberola-Die et al., 2011, 2013) and the prokaryotic channels GLIC and ELIC (Hilf et al., 2010; Gonzalez-Gutierrez and Grosman, 2015). However, the specific mechanisms of action of lidocaine on neuroreceptors are not fully understood.

Recently, we have described that lidocaine blocks muscle-type (Alberola-Die et al., 2011) and neuronal heteromeric (Alberola-Die et al., 2013) nAChRs by multiple mechanisms that can be dissected, at least partially, by dose. At doses lower than the IC₅₀, lidocaine inhibits nAChRs mainly by open-channel blockade, whereas at higher concentrations (equal or higher than the IC₅₀) it also causes closed-channel blockade and enhances desensitization. Now, our aim is to unravel the structural determinants within the lidocaine molecule responsible for

specific effects on muscle-type nAChRs. So, in this work we have analyzed the effects of diethylamine (DEA), a small molecule resembling the hydrophilic moiety of the lidocaine molecule (see **Figure 1A**), on nAChRs.

When dealing with ligands bearing different chemical groups in their structure and exhibiting multiple functional effects, it would be ideal to be able to dissect the functional role of individual structural components by using smaller molecules mimicking specific domains of the larger ligand. This is the rationale behind this study on the hydrophilic DEA, which mimics the terminal amine group of lidocaine and indeed reproduces with high fidelity some, but not all, of its effects. Once this has been established, it opens the way to use small molecules resembling the hydrophobic moiety of lidocaine to determine which, if any, of the effects of the entire ligand on AChRs function can also be attributed to such hydrophobic domain.

Preliminary results have been published elsewhere (Alberola-Die et al., 2012).

MATERIALS AND METHODS

Purification and Reconstitution of nAChRs

nAChRs from *Torpedo marmorata* electroplax were purified by bromoacetylcholine-affinity chromatography in the presence of asolectin lipids using cholate as a detergent. After elution with carbamylcholine, purified receptors were dialyzed and reconstituted in asolectin lipids at a final protein concentration of 0.3–1.2 mg ml⁻¹. Samples were aliquoted and stored in liquid nitrogen (Ivorra et al., 2002).

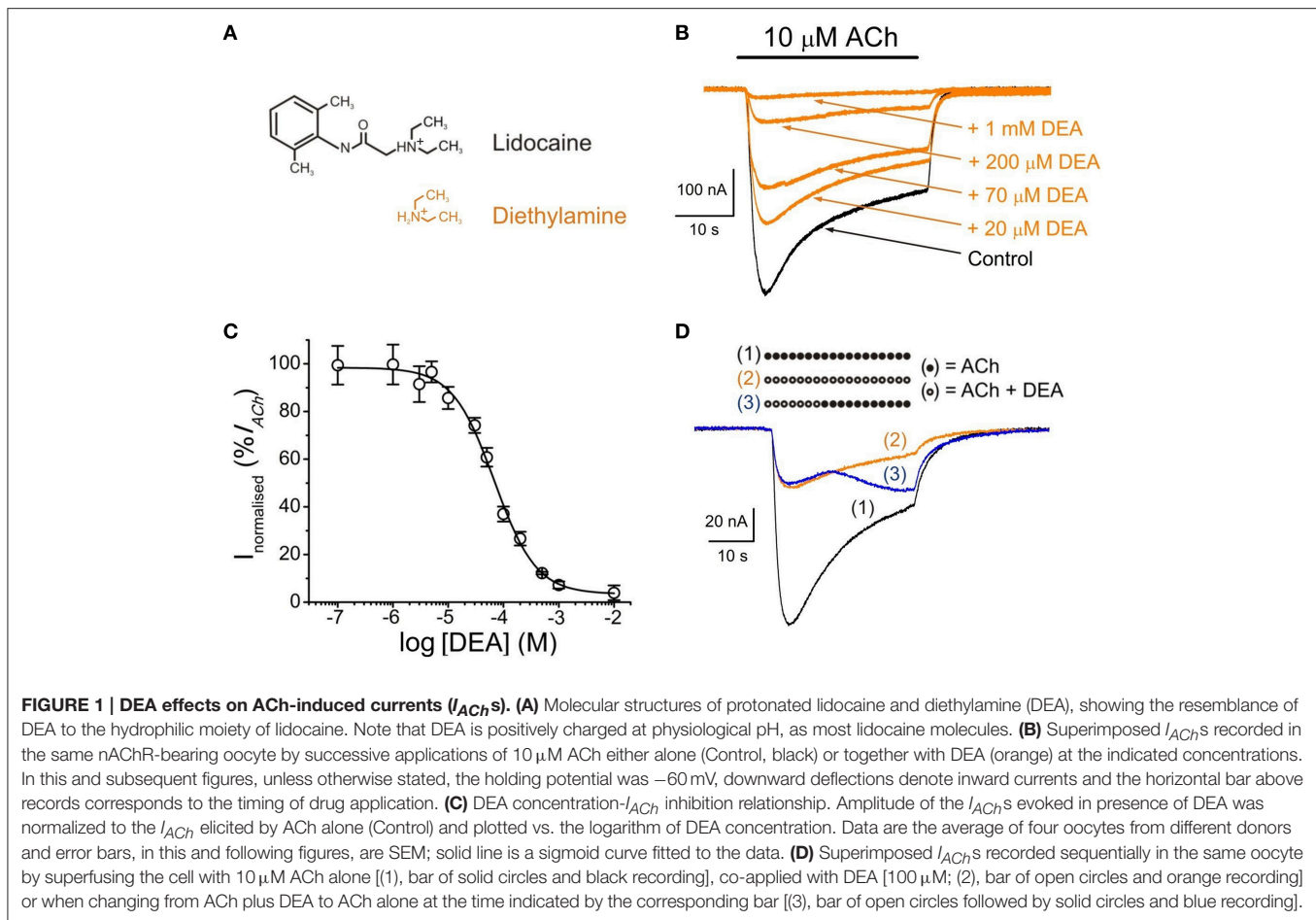
Oocyte Preparation and Microinjection

Adult female *Xenopus laevis* (purchased from Harlan Interfauna Ibérica S.L., Barcelona, Spain; and Centre National de la Recherche Scientifique, Montpellier, France) were immersed in cold 0.17% MS-222 for 20 min and a piece of ovary was drawn out aseptically. Animal handling was carried out in accordance with the guidelines for the care and use of experimental animals adopted by the E.U. and the animal protocol was approved by the ethic committee of Universidad de Alicante. Stage V and VI oocytes were isolated and their surrounding layers removed manually. Cells were kept at 15–16°C in a modified Barth’s solution [88 mM NaCl, 1 mM KCl, 2.40 mM NaHCO₃, 0.33 mM Ca(NO₃)₂, 0.41 mM CaCl₂, 0.82 mM MgSO₄, 10 mM HEPES (pH 7.4), 100 U ml⁻¹ penicillin, and 0.1 mg ml⁻¹ streptomycin] until used. Oocytes were microinjected with 100 nl of an aliquot of reconstituted nAChRs (Morales et al., 1995).

Two-Electrode Voltage-Clamp Recordings in Oocytes

Membrane current recordings were performed at 21–25°C, 16–72 h after proteoliposome injection, using a high compliance two-microelectrode voltage-clamp system (TurboTEC-10CD, npi Tamm, Germany). The recording methodology has been described previously (Morales et al., 1995; Olivera-Bravo et al., 2005). Briefly, intracellular microelectrodes (0.8–3 MΩ) were filled either with 3 M KCl or potassium acetate for voltage

Abbreviations: ACh, acetylcholine; ANR, normal Ringer solution with atropine; DEA, diethylamine; EC domain, extracellular domain; ELIC, *Erwinia chrysanthemi* ligand-gated ion channel; GABA_AR, gamma-aminobutyric acid type A receptors; GLIC, *Gloeobacter violaceus* ligand-gated ion channel; GlyR, glycine receptor; I_{ACh}, ACh-elicited current; LA, local anesthetic; LGIC, ligand gated ion channel; MS-222, ethyl 3-aminobenzoate methanesulfonate; *n*, number of oocytes; *N*, number of oocyte donors; nAChR, nicotinic acetylcholine receptor; NR, normal Ringer solution; TM, transmembrane segment.



recording and current injection, respectively. Oocytes were placed in a $150 \mu\text{l}$ recording chamber and continuously superfused with normal frog Ringer's solution (NR: 115 mM NaCl, 2 mM KCl, 1.8 mM CaCl_2 , 5 mM HEPES, pH 7.0) supplemented with $0.5 \mu\text{M}$ atropine sulfate (normal Ringer with atropine, ANR) to block any muscarinic response (Kusano et al., 1982). The membrane potential was held at -60 mV , unless otherwise stated. ACh and other tested drugs were diluted in ANR solution and oocytes were superfused with them at a flow rate of $13\text{--}17 \text{ ml min}^{-1}$. Membrane currents elicited by ACh (I_{ACh}) either alone or co-applied with DEA, were low-pass filtered at $30\text{--}1000 \text{ Hz}$ and, after sampling at fivefold the filter frequency (Digidata 1200 Series and Digidata 1440A; Axon Instruments, Foster City, CA, USA), recorded on two PC-computers, using the WCP v.3.2.8 package developed by J. Dempster (Strathclyde Electrophysiology Software, University of Strathclyde, Scotland, UK) and AxoScope v. 10.0.0.60 (Molecular Devices Corporation, Sunnyvale, U.S.A.).

Experimental Design

DEA concentration- I_{ACh} inhibition relationship was determined by measuring I_{ACh} s evoked by $10 \mu\text{M}$ ACh alone or together with different DEA concentrations. For competition assays, ACh concentration- I_{ACh} amplitude curves were obtained by exposing

injected oocytes to increasing ACh concentrations, either alone or together with $100 \mu\text{M}$ DEA. I_{ACh} s recorded in the presence or absence of DEA were normalized to the I_{ACh} evoked by 1 mM ACh alone and fitted to a sigmoid curve. To allow nAChRs to recover from desensitization, the interval between consecutive ACh applications was at least 5 min. To assess the blockade of resting nAChRs by DEA, we compared the I_{ACh} s elicited by ACh (from $1 \mu\text{M}$ to 1 mM) alone or co-applied with $100 \mu\text{M}$ DEA either directly or after pre-application of DEA (same concentration) for 12 s.

When studying the voltage dependence of the I_{ACh} blockade by DEA, series of 800 ms voltage pulses (in 10 mV steps from -120 to -20 mV , followed by 20 mV jumps from -20 to $+60 \text{ mV}$; occasionally, in 20 mV steps from -120 to $+60 \text{ mV}$) were given to the oocyte before ligand superfusion and during the I_{ACh} plateau elicited by $10 \mu\text{M}$ ACh, either alone or co-applied with DEA at different concentrations. In a few cells, the -120 mV pulse duration was extended up to 1500 ms to allow a more complete current relaxation.

Data Analysis and Statistical Procedures

Inhibition curves were determined by measuring I_{ACh} evoked by $10 \mu\text{M}$ ACh in the presence of different DEA concentrations. I_{ACh} s elicited in the presence of DEA were normalized to the

I_{ACh} evoked by ACh alone. Data were fitted to a single-site inhibition curve using the Origin 6.1 software (OriginLab Corp. Northampton, MA, U.S.A.).

Reversibility of I_{ACh} blockade by DEA was determined by giving 32 s pulses of ACh either alone or co-applied with DEA, for solely the first 12 s or during the whole pulse; I_{ACh} recovery was measured 20 s and 7 min after DEA washout. The percentage of recovery from blockade (% Recovery) was obtained using the equation:

$$\% \text{ Recovery} = \left[\frac{(I_{ACh \text{ after DEA}} - I_{ACh+DEA})}{(I_{ACh} - I_{ACh+DEA})} \right] \times 100 \quad (1)$$

where I_{ACh} is the current amplitude evoked by 10 μM ACh alone; $I_{ACh+DEA}$, is the current elicited by co-application of 10 μM ACh with 100 μM DEA; and $I_{ACh \text{ after DEA}}$ is the current obtained 20 s or 7 min after DEA removal.

The rate of desensitization was determined by measuring the I_{ACh} amplitude elicited by 100 μM ACh, either alone or co-applied with up to 200 μM DEA, at different times after I_{ACh} peak. Desensitization rates were obtained using the equation:

$$D_{ti} = [1 - (I_{ti}/I_{peak})] \times 100 \quad (2)$$

where D_{ti} is the desensitization value at the specified time; I_{peak} the I_{ACh} amplitude at the peak; and I_{ti} the current amplitudes remaining 2, 10, and 20 s after the peak (Olivera-Bravo et al., 2007). The apparent time-to-peak was determined as the time elapsed from I_{ACh} onset to the I_{ACh} peak. We have called this parameter as “apparent” time-to-peak, just to indicate that these values do not necessarily reflect “real” time-to-peak values of nAChR activation but those observed in our experimental conditions.

To characterize the pharmacological profile of DEA, nAChRs were activated by different ACh concentrations either alone or co-applied with DEA (at roughly its IC_{50}), just directly or after pre-application of DEA for 12 s. Dose-response data were fitted to the following form of the Hill equation:

$$I/I_{max} = [1 + (EC_{50}/[ACh])^{n_H}]^{-1} \quad (3)$$

where I is the I_{ACh} peak elicited at a given ACh concentration (applied either alone or together with DEA); I_{max} is the maximum I_{ACh} recorded; EC_{50} is the agonist concentration required to obtain one-half the maximum I_{ACh} ; and n_H is the Hill coefficient.

Net i/v curves for I_{ACh} were obtained subtracting, for each voltage, the steady-state currents attained in ARN (measured during the last 100 ms of the pulse) from the corresponding ones recorded in presence of ACh. These net I_{ACh} values were normalized, for each oocyte, to the ACh response at -60 mV. The percentage of I_{ACh} inhibition at different membrane potentials (V_m) was computed using the following equation:

$$InV_m = [1 - (I_{(ACh+DEA) \text{ at } V_m} / I_{(ACh) \text{ at } V_m})] \times 100 \quad (4)$$

where InV_m is the percentage of I_{ACh} inhibition at the corresponding V_m ; $I_{(ACh+DEA) \text{ at } V_m}$ is the I_{ACh} amplitude in the presence of ACh and DEA at V_m ; and $I_{(ACh) \text{ at } V_m}$ is

the I_{ACh} elicited by ACh alone at V_m . Values from the i/v relationship obtained at different blocker concentrations were fitted to Equations (5) and (6) to estimate the fraction of voltage field (δ) sensed by DEA at its binding site inside the channel. The apparent K_i for each membrane potential, which is the concentration of DEA that reduces I_{ACh} amplitude to the half, was estimated from the following equation:

$$I_{ACh+DEA}/I_{ACh} = I_{min} + [(I_{max} - I_{min}) / (1 + ([DEA]/K_i)^n)] \quad (5)$$

where $I_{ACh+DEA}$ is the current evoked by co-application of 10 μM ACh with a given concentration of DEA; I_{ACh} is the current elicited by 10 μM ACh alone (control conditions); I_{min} and I_{max} are, respectively, the minimum and maximal fractional-current amplitudes evoked; $[DEA]$ is the DEA concentration; and n is the slope factor. To estimate the fraction of the voltage field experienced by the blocking particle (δ), we used the following form of the Woodhull equation (Woodhull, 1973):

$$\log K_i(V) = \log K_i(0mV) + (z\delta FV / 2.303RT) \quad (6)$$

where $K_i(V)$ is estimated from Equation (5) at each membrane potential; $K_i(0mV)$ is the K_i -value at 0 mV; z is the electric charge of DEA and R , T , and F have the usual thermodynamic meanings.

Unless otherwise specified, values given are the mean \pm SEM; “ n ” indicates the number of oocytes and “ N ” is the number of donors from which data were obtained. When comparing two-group means of normally distributed values, the Student’s t -test was used; otherwise, Mann-Whitney rank-sum test was applied. Among-group differences for non-normally distributed data were determined by the Kruskal–Wallis analysis of variance on ranks and multiple comparisons vs. a control group were carried out with the Dunn’s method. A significance level of $p < 0.05$ was considered for all cases.

Virtual Docking Assays

The structure of the full domains of *Torpedo* nAChR was taken from RCSB Protein Data Bank (code 2BG9), which was determined, in the closed channel state, by electron microscopy at 4 Å resolution (Unwin, 2005). The edition of the protein was made using DeepView v4.1 (Guex and Peitsch, 1997) and Yasara (Krieger et al., 2002, 2004) software without further optimization. Lidocaine and DEA structures (CID 8021 and 3676, respectively) were taken from NCBI Pubchem database (<http://www.ncbi.nlm.nih.gov/pccompound>). A global docking procedure was accomplished with AutoDock 4 (Morris et al., 2008) implemented in Yasara, where a total of 500–1000 flexible docking runs were set and clustered around the putative binding sites. The program then performed a simulated annealing minimization of the complexes, which moved the structure to a nearby stable energy minimum, by using the implemented AMBER 99 force field (Duan et al., 2003). The binding energy was obtained by calculating the energy at infinite distance between the ligand and the nAChR oligomer and subtracting the energy of the whole complex. The more positive the binding energy, the more favorable was the interaction in the context of the force

field. The best binding energy complex in each cluster was stored, analyzed and used to select the best orientation of the interacting partners. Figures were drawn with open source Pymol (The PyMOL Molecular Graphics System, Version 1.8 Schrödinger, LLC, at <http://www.pymol.org/>). Yasara pH command was set to 7, ensuring that molecules preserve their pH dependency of bond orders and protonation patterns. In this way, almost all DEA molecules remained charged during the docking procedure, but only 86% of the lidocaine molecules.

Drugs

ACh, atropine sulfate, DEA, MS-222, penicillin, and streptomycin were from Sigma (St. Louis, MO, USA). HEPES was obtained from Acros Organics (New Jersey, NJ, USA). Reagents of general use were purchased from Scharlau Chemie SA (Barcelona, Spain). All solutions were made in ANR just before each application, unless otherwise stated.

RESULTS

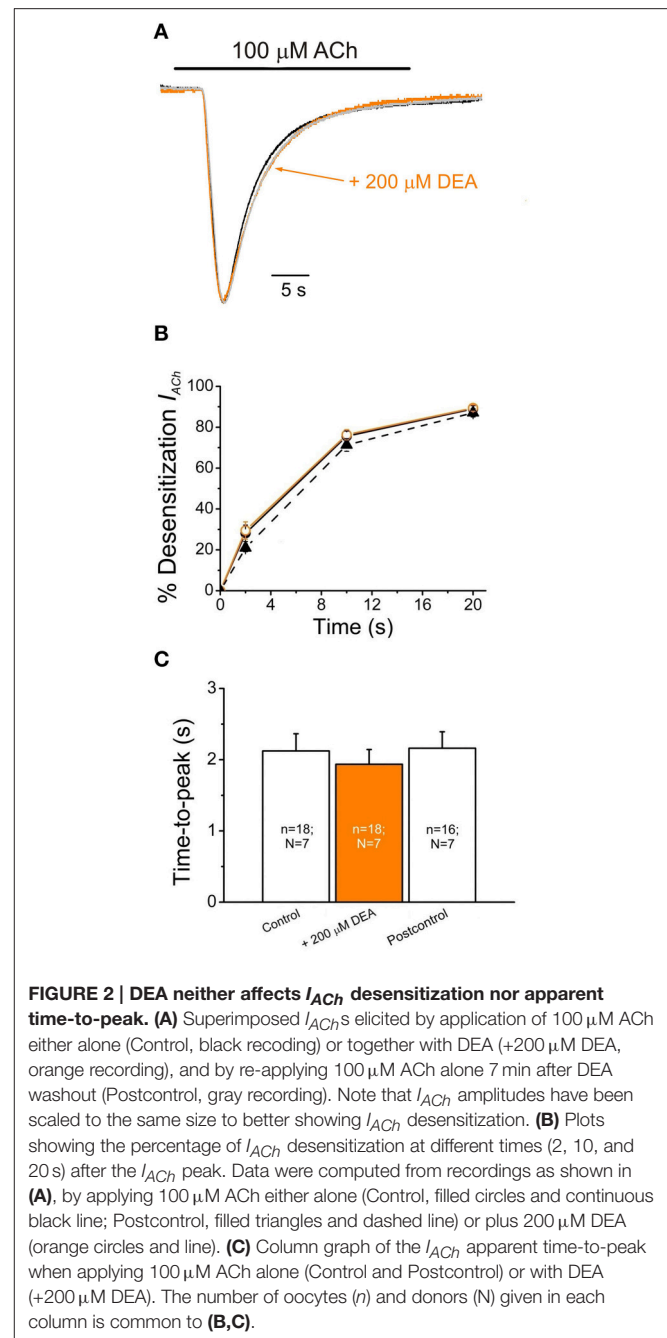
Inhibition of I_{ACh} by DEA

In either uninjected cells or oocytes bearing nAChRs, with the membrane potential held at -60 mV, DEA superfusion did not modify their membrane conductance, even at concentrations as high as 10 mM (not shown). However, in oocytes bearing nAChRs, co-application of $10 \mu\text{M}$ ACh with $0.1 \mu\text{M}$ – 10 mM DEA reversibly inhibited I_{ACh} s in a concentration-dependent manner (Figure 1B). The percentage of I_{ACh} , normalized to the control value obtained in presence of $10 \mu\text{M}$ ACh alone, was plotted vs. the logarithm of DEA concentration, and values were fitted to a sigmoid curve (Figure 1C). The half inhibitory DEA concentration (IC_{50}) was $68 \mu\text{M}$ (range 60 – $102 \mu\text{M}$), similar to that found for lidocaine ($73 \mu\text{M}$, range 62 – $83 \mu\text{M}$; Alberola-Die et al., 2011), and the Hill coefficient (n_H) was 1.15 ± 0.04 , suggesting that a single DEA molecule caused lidocaine-like nAChR blockade.

I_{ACh} recovery after DEA ($100 \mu\text{M}$) superfusion was slow and I_{ACh} did not reach control values even 7 min after DEA washout. So, the percentage of I_{ACh} recovery, estimated from Equation (1), was $73 \pm 8\%$ ($n = 7$, $N = 6$; Figure 1D) and $86 \pm 2\%$ ($n = 16$, $N = 11$; not shown) at 20 s and 7 min, respectively, indicating that a fraction of DEA molecules remains bound to the nAChR several minutes after DEA rinsing out.

DEA has No Effect on I_{ACh} Desensitization and the Apparent Time-To-Peak

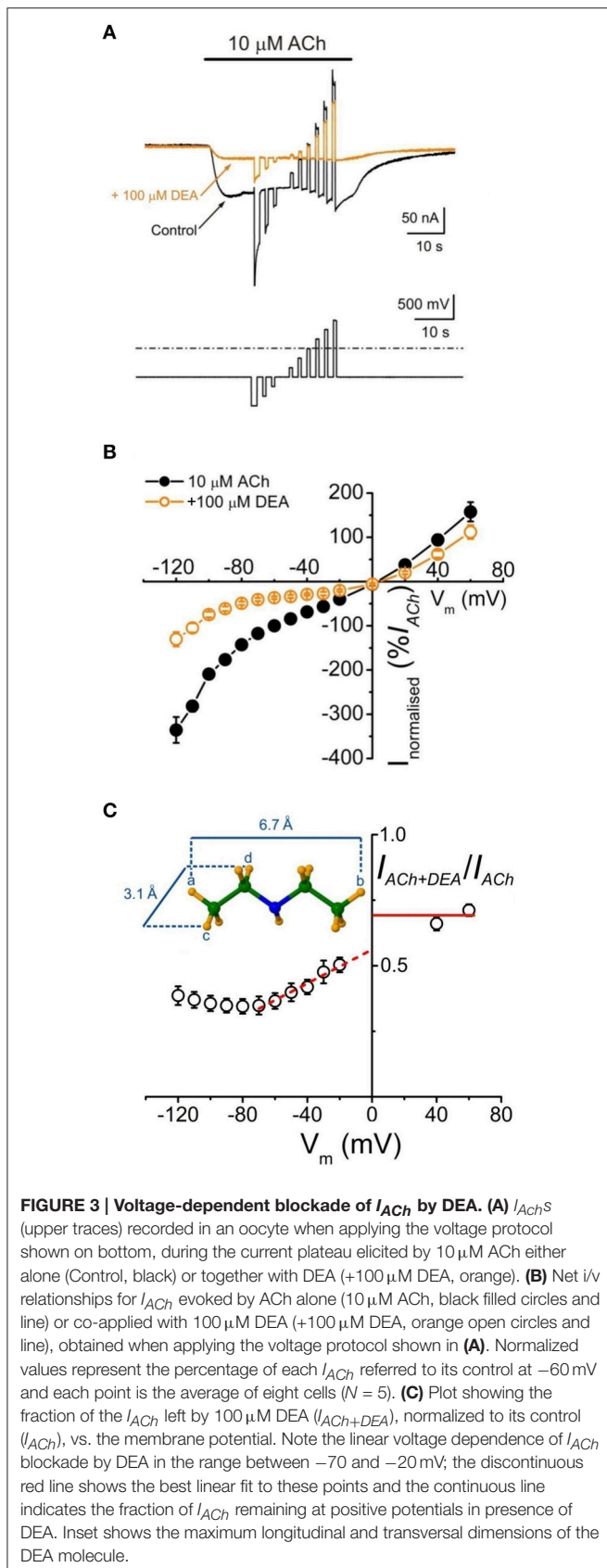
The I_{ACh} decay was not affected in oocytes superfused with $100 \mu\text{M}$ ACh together with DEA at the IC_{50} ($70 \mu\text{M}$). As shown in Figures 2A,B, DEA did not change I_{ACh} desensitization even when DEA concentration was increased to $200 \mu\text{M}$, roughly threefold its IC_{50} . The D_{fit} -values obtained at 2 and 20 s (see Equation 2 in Materials and Methods) were: $30 \pm 2\%$ and $89 \pm 1\%$ ($n = 18$, $N = 7$) for I_{ACh} s elicited by ACh alone vs. $29 \pm 4\%$ and $89 \pm 2\%$ (the same cells) when ACh was co-applied with $200 \mu\text{M}$ DEA (Figure 2B; $p > 0.05$, Mann-Whitney rank-sum test). Moreover, DEA at concentrations as high as $200 \mu\text{M}$ had no effect on the time elapsed from I_{ACh} onset to the I_{ACh}



peak (the apparent time-to-peak was 2.1 ± 0.2 s for $100 \mu\text{M}$ ACh alone against 1.9 ± 0.2 s for $100 \mu\text{M}$ ACh plus $200 \mu\text{M}$ DEA; data are from the same cells in which desensitization was measured; $p > 0.05$, Mann-Whitney rank-sum test; Figures 2A,C). DEA lack of effect on I_{ACh} desensitization and apparent time-to-peak is in sharp contrast with the action of the entire lidocaine molecule on nAChRs (Alberola-Die et al., 2011, 2013).

Voltage-Dependence of nAChR Blockade by DEA

To assess whether I_{ACh} inhibition by DEA is voltage-dependent, voltage pulses were applied to oocytes while superfusing the cell



with just ANR or during the I_{ACh} plateau elicited by $10\ \mu\text{M}$ ACh either alone or co-applied with $100\ \mu\text{M}$ DEA (Figure 3A; see Experimental Design in Materials and Methods). Figure 3B shows the i/v relationship obtained when plotting the net I_{ACh} s elicited by ACh, either alone or with DEA, normalized to its control I_{ACh} at $-60\ \text{mV}$, against the membrane potentials tested. The i/v curve for $10\ \mu\text{M}$ ACh alone showed a reversal potential close to $0\ \text{mV}$ and the characteristic inward rectification of I_{ACh} (Figure 3B, Morales et al., 1995). When DEA was co-applied with ACh the I_{ACh} reversal potential was unmodified, indicating that the channel selectivity was unaffected. However, in the presence of DEA, I_{ACh} amplitude decreased in a voltage-dependent way, the blockade being higher at negative potentials (Figure 3B). Nevertheless, the voltage dependency was only linear in the range from -70 to $-20\ \text{mV}$, since at potentials more negative than $-70\ \text{mV}$ the I_{ACh} blockade did not increase, but even decreased. So, as displayed in Figure 3C, the fraction of I_{ACh} remaining in the presence of DEA rose as the membrane was hyperpolarized beyond $-70\ \text{mV}$. This biphasic behavior of the voltage-dependent blockade of I_{ACh} by DEA at negative potentials was not observed for lidocaine (Alberola-Die et al., 2011), and could be due to the small size of DEA (see inset Figure 3C). Interestingly, at positive potentials DEA blocked roughly 30% of I_{ACh} (Figure 3C), indicating that DEA also causes a significant voltage-independent blockade of nAChRs.

Since DEA caused a voltage-dependent blockade of nAChRs, more evident between -70 and $-20\ \text{mV}$ (Figures 3B,C), we used in one cell the Woodhull's equation (see Equations 4–6 in Materials and Methods), restricted to this range of potentials, to estimate the δ -value, which indicates the fraction of the voltage field sensed by DEA at its binding site. The calculated δ -value was 0.30 (assuming that $z = 1$), indicating that DEA binds to the external third of the channel length.

Effects of DEA on nAChR Pharmacological Profile

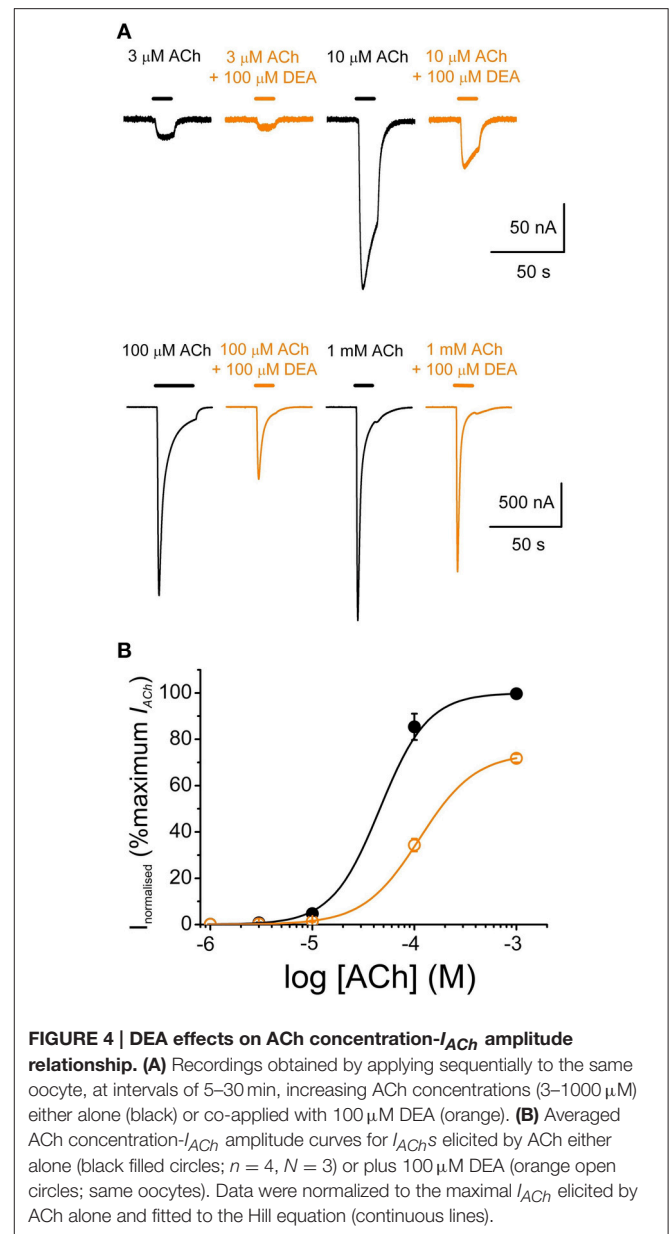
To study the effects of DEA on the ACh dose- I_{ACh} relationship, ACh was applied in the same oocyte at increasing concentrations ($1\ \mu\text{M}$ – $1\ \text{mM}$) either alone or with $100\ \mu\text{M}$ DEA (Figure 4A). Figure 4B shows the ACh dose- I_{ACh} curves obtained either in absence or in presence of $100\ \mu\text{M}$ DEA, normalizing I_{ACh} values to those evoked by $1\ \text{mM}$ ACh alone; data were fitted to sigmoid curves with the Hill equation (see Equation 3 in Materials and Methods). The EC_{50} for the control curve (ACh alone) was $46\ \mu\text{M}$ (range 16 – $59\ \mu\text{M}$) and the n_H 1.8 ± 0.1 ($n = 4$, $N = 3$), values which are in good agreement with previous data (Morales et al., 1995; Olivera-Bravo et al., 2005). In presence of $100\ \mu\text{M}$ DEA, the maximum I_{ACh} amplitude decreased, suggesting a non-competitive blockade (Figures 4A,B); besides, the dose-response curve shifted to the right, increasing the EC_{50} more than twice the original value ($112\ \mu\text{M}$, range 88 – $145\ \mu\text{M}$) and slightly decreasing the n_H to 1.6 ± 0.1 (Figure 4B). As shown in Figures 4A, 5C, co-application of $10\ \mu\text{M}$ ACh with $100\ \mu\text{M}$ DEA caused an I_{ACh} inhibition of $73 \pm 1\%$ ($n = 29$, $N = 10$), but when the same concentration of DEA was co-applied with $1\ \text{mM}$ ACh, the percentage of blockade was only $35 \pm 7\%$ ($n = 5$, $N = 4$;

$p < 0.05$, t -test). Thus, the percentage of I_{ACh} inhibition by DEA markedly depended on the ACh concentration, suggesting that the blocking effect of DEA on nAChRs was not merely non-competitive. This apparently competitive inhibition of I_{ACh} by DEA could be explained, at least partially, by its binding to resting nAChRs. To test this hypothesis, we determined the percentages of I_{ACh} blockade induced by DEA (100 μ M) when it was pre-applied to the cell for 12 s before being co-applied with ACh at increasing concentrations (1 μ M–1 mM; **Figure 5A**). **Figure 5B** shows the ACh dose- I_{ACh} curves elicited by ACh either alone or when it was co-applied with 100 μ M DEA after its pre-application for 12 s. In these experiments, the EC_{50} for ACh alone was 37 μ M (range 14–115 μ M) and the n_H 1.9 ± 0.1 ($n = 7-11$, $N = 1-3$) whereas in presence of DEA the EC_{50} slightly increased, to 60 μ M (range 52–126 μ M), and the n_H was 1.6 ± 0.1 . Noticeably, when DEA was pre- and co-applied with ACh, a smaller maximum I_{ACh} amplitude was reached (**Figures 5A,B**) and the percentage of I_{ACh} inhibition caused was similar for any ACh concentration above 10 μ M ($p > 0.05$, Kruskal Wallis ANOVA on ranks; red plot of **Figure 5C**), indicating a non-competitive blockade of nAChRs. Thus, the effect of DEA pre- and co-application strikingly contrasted with that obtained by just DEA co-application with ACh, since the latter showed an apparent competitive mode of action while the former did not (**Figures 4B, 5C**). If DEA binds to, and blocks, closed (resting) nAChRs, it would be expected that DEA pre-application enhanced the I_{ACh} inhibition caused by just DEA and ACh co-application, preferentially at the highest ACh doses (1 mM), as we have indeed observed (**Figure 5C**). This is likely due to the fast activation of most nAChRs by 1 mM ACh, which prevented the effect of DEA on resting nAChRs when ACh and DEA were just co-applied. On the other hand, at low ACh doses only a small percentage of nAChRs were activated by the agonist and consequently most nAChRs remained resting and therefore susceptible to be blocked by DEA, acting as a closed-channel blocker.

Virtual Docking Assays of DEA- and Lidocaine-nAChR Interactions

Virtual docking assays were carried out to explore DEA- and lidocaine-nAChR interactions using as template the full structure of *Torpedo* nAChRs (see Materials and Methods). For these assays 1000 runs of interactions were performed for DEA and 500 for lidocaine molecules.

The runs for DEA-nAChR interactions disclosed 30 clusters of sites that differed in less than 5 Å of root-mean-square-deviation of which 23 (77%) corresponded to EC, 2 (7%) to TM, and the remaining 6 (16%) to intracellular domains; this latter solutions have been discarded for further analysis because DEA is impermeable through the cell membrane and so they lack of functional meaning. As shown in **Figure 6** (upper panels), the small and polar DEA barely binds to the TM domain, except inside the channel pore (**Figures 6A₁,A₂**), but it interacted quite well with different nAChR residues located at the EC domain (**Figures 6A₁,A₃**). Regarding the EC domain, most DEA-clusters were detected at α , β , or γ subunits (**Figures 6A₁,A₂**). Interestingly, some DEA clusters were located at the α - γ interphase, in a cavity near the orthosteric binding



site (**Figure 6A₃**). In fact, one of those clusters involved the Y93 (loop A) of the α subunit and the D176 (loop F) of the γ subunit, although due to the small size of DEA this cluster did not reach neither the B nor the C-loop (**Figure 6A₃**), which also contribute with key residues to the ACh binding pocket (Corringer et al., 2000). By contrast, the docking-assays did not show any equivalent hotspot for DEA binding at the interphase of α - δ subunits (see **Figure 6A₂**). The remaining DEA clusters at the EC domain were mostly at intersubunit crevices, although a few solutions involved a single chain of α , β , or γ subunits (**Figures 6A₁,A₂**). At the TM domain, we only detected 2 DEA clusters located near and into the channel pore. Both clusters involved interactions with α - β or α - δ subunits (**Figure 6A₂** and Supplementary Figure 2) and their binding sites were at a depth circa one third of the whole membrane thickness, from the extracellular side (see Supplementary Figure 2).

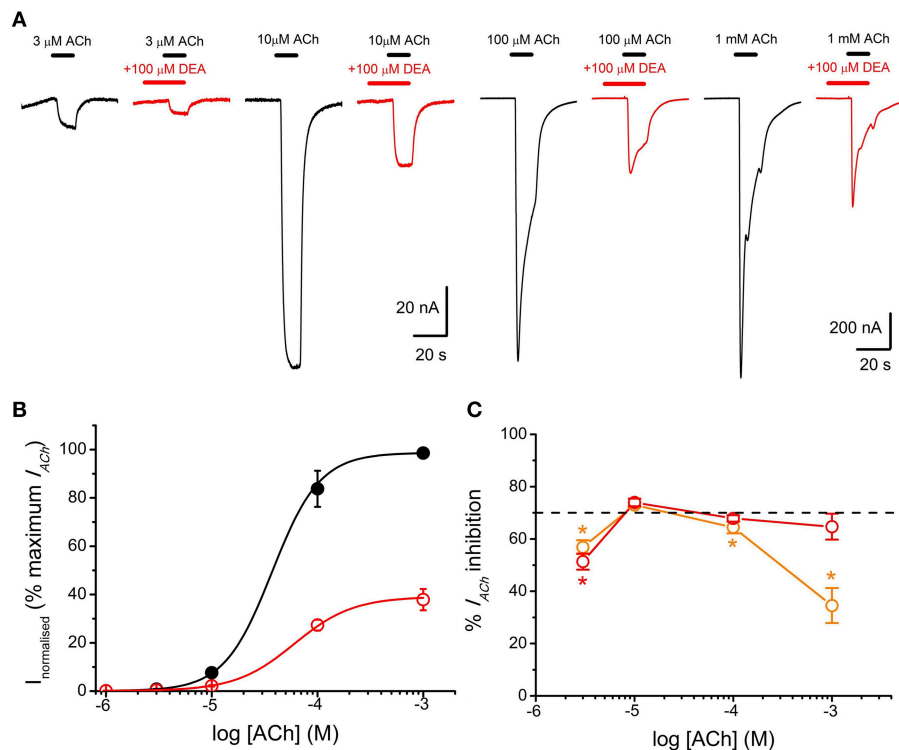


FIGURE 5 | DEA pre-application increases I_{ACh} inhibition and changes the pharmacological profile of nAChR inhibition. (A) I_{ACh} s elicited by pre-application of 100 μ M DEA for 12 s followed by its co-application with ACh at the indicated concentrations (red recordings) or by just ACh at the same concentrations (black recordings). **(B)** Averaged ACh concentration- I_{ACh} amplitude curves for I_{ACh} s evoked by ACh either alone (black filled circles; $n = 4-8$, $N = 1-3$) or co-applied with 100 μ M DEA, after 12 s pre-application of DEA at the same concentration (red open circles; same oocytes). Data were normalized to the maximal I_{ACh} elicited by ACh alone and fitted to the Hill equation (continuous lines). **(C)** Plot showing the percentage of I_{ACh} inhibition when ACh (at different concentrations) was directly co-applied with 100 μ M DEA (orange circles; $n = 5-29$, $N = 4-10$), or when ACh and DEA co-applications were preceded by 100 μ M DEA pre-application for 12 s (red circles; $n = 7-11$, $N = 1-3$). Asterisks indicate significant differences ($p < 0.05$, Kruskal Wallis ANOVA on ranks) respect to the I_{ACh} blockade caused by solely co-applying 10 μ M ACh and 100 μ M DEA; the dashed line indicates 70% inhibition. Note that the percentage of I_{ACh} inhibition decreased markedly when DEA was just co-applied with high ACh concentrations. By contrast, when DEA was pre-applied before its co-application with ACh, the percentage of I_{ACh} blockade was similar at the different ACh concentrations (see text). The slight decrease in I_{ACh} blockade by DEA observed at 3 μ M ACh might not be reliable because of the inaccuracies own to the small size of I_{ACh} at this agonist concentration.

For lidocaine-nAChR interactions, the docking assays revealed multiple hotspots both at EC and TM domains (Figure 6, lower panels), being remarkable the large number of solutions found at the TM domain as compared with DEA. At the EC region, it is noteworthy that one of the lidocaine clusters fitted quite well at the nAChR ligand binding sites in the α - γ (Figures 6B₂, B₃) and the α - δ interphases (Figure 6B₂), involving interactions with key residues of A, B, and C loops of the α subunit. In contrast to DEA, the adjustment of this cluster to the ACh-binding site is possible because the longitudinal and transversal molecular dimensions of lidocaine (10.7 and 4.8–6.7 Å, respectively) are relatively close to those of the ACh molecule (9.1 and 3.7 Å, respectively). Besides, some lidocaine clusters at the EC domain were located at intersubunit crevices and, less frequently, at intrasubunit sites, with a roughly similar pattern to that found for DEA (compare Figure 6A₁ and Figure 6B₁). At the TM domain, lidocaine clusters could be grouped in three main types of interactions: (i) inside the pore of the channel, where lidocaine contacted residues of M2 segments from four or five subunits; (ii) intrasubunit interactions, with low binding energy, at the cavities among the M1, M2, M3, and M4 segments

of each subunit; and (iii) intersubunit interactions, where α - γ , α - δ , and β - δ hotspots presented the strongest interactions with lidocaine.

DISCUSSION

Lidocaine is an aminoethylamide local anesthetic that in physiological solutions exists as a mixture of charged and uncharged species, due to its pK_a of 7.8 (Liu et al., 2003). Though it is believed that the charged form of lidocaine mediates most of its therapeutic action (Narahashi et al., 1969), the neutral form is also important because of its higher ability to penetrate inside membranes. Interestingly, both charged and uncharged forms of lidocaine seem to contribute to the complex inhibitory action of this molecule on muscle- (Alberola-Die et al., 2011) and neuronal-type (Alberola-Die et al., 2013) nAChRs. Now, we report the effects of DEA, a small amine mimicking the hydrophilic moiety of lidocaine, on muscle-type nAChR, which is mostly protonated at physiological pH, since it has a pK_a of 10.49 (Sergeeva et al., 2000).

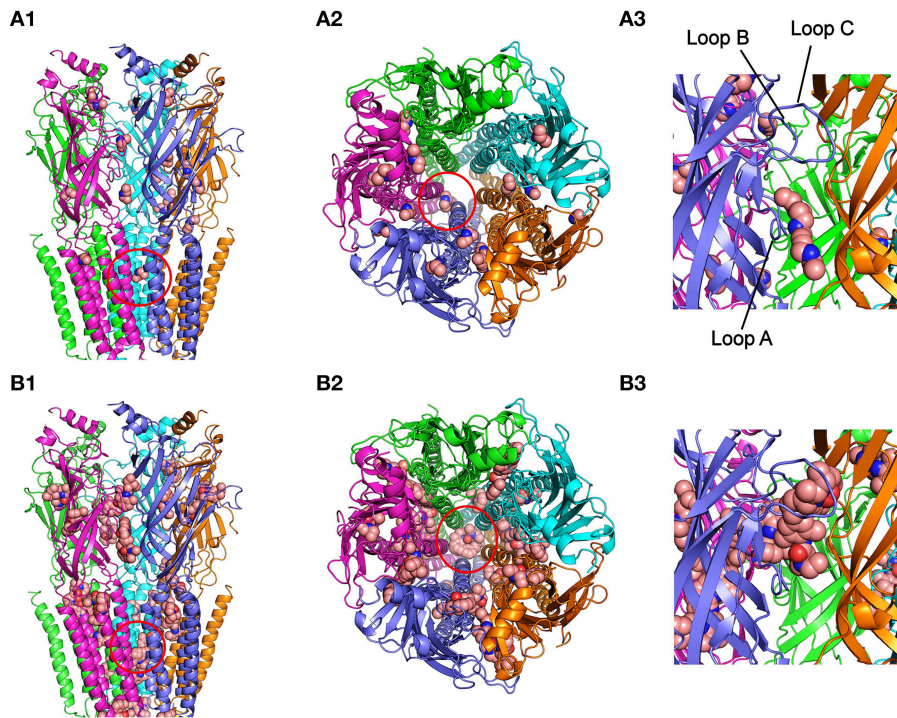


FIGURE 6 | Modeling of DEA and lidocaine binding to the EC and TM domains of the nAChR. (A₁) Lateral view, in the membrane plane (top corresponding to the synaptic cleft), of nAChR with bound DEA molecules. For this and following panels, the color code for nAChR subunit is: α (blue and cyan), β (magenta), γ (orange), and δ (green). Ligand molecules are colored brown and represented in van der Waals spheres. Note that very few DEA-nAChR binding solutions (clusters) were located at the TM domain, except two inside the channel pore (red circle), but there were several clusters distributed at the EC domain. **(A₂)** Upper view, from the synaptic cleft, of the nAChR with bound DEA. Note that at the EC domain DEA mainly interacts at intersubunit interphases, although some DEA clusters involved single subunits. The red circle indicates the cluster sited within the channel pore. **(A₃)** An expanded view of the EC domain at the α - γ interphase, corresponding to the ACh-binding site (loops A, B, and C of the α subunit are indicated as reference). Note that one DEA cluster is near, but not inside, the pocket of the ACh-binding site (see text for details). **(B₁)** nAChR with lidocaine binding solutions is shown in a similar view as in **(A₁)**. Note that lidocaine clusters were more numerous at the TM domain than those observed for DEA. Several TM clusters were located in intra- and intersubunit cavities and others inside the channel pore (red circle). At the EC domain there were also several lidocaine clusters, with a distribution roughly similar to that found for DEA. **(B₂)** Upper view, from the synaptic cleft, of the nAChR with bound lidocaine. Note that both at the EC and TM domains lidocaine interacted at intra and intersubunit interphases and that several clusters were grouped inside the channel pore (red circle). **(B₃)** The same nAChR area as in **(A₃)**, showing a lidocaine cluster into the ACh-binding site.

DEA effects on nAChRs reported here showed important analogies with those described for lidocaine. So, the IC_{50} for DEA ($68 \mu\text{M}$) was quite similar to that found for lidocaine ($73 \mu\text{M}$; Alberola-Die et al., 2011); besides, DEA and lidocaine blocked nAChRs in a voltage-dependent manner, but also bound to resting (closed) nAChRs. However, there were important differences between lidocaine and DEA actions on nAChRs, mainly in the desensitization rate, which was markedly increased by lidocaine (Alberola-Die et al., 2011), but unaffected by DEA, even at concentrations well over its IC_{50} . This differential effect is of great functional relevance because it points out that the faster I_{ACh} decay induced by lidocaine is indeed caused by an enhancement of nAChR desensitization, rather than by a slow nAChR blockade.

It has been previously shown that DEA, and also triethylamine, can directly activate muscle-type nAChRs, and so act as partial agonists (Sánchez et al., 1986). However, this effect was observed at concentrations far higher (mM range) than those used here to characterize DEA inhibitory actions on nAChRs. In our hands, such high doses would almost completely

block the I_{ACh} (Figure 1). Besides, in oocytes bearing nAChRs, we could not detect any current activated by DEA alone, even when applied up to 10 mM. Thus, although it cannot be fully ruled out that DEA causes some competitive inhibition on nAChRs, the marked shift to the right of the ACh dose- I_{ACh} curve (Figure 4) rather suggests the involvement of other mechanisms (i.e., blockade of closed nAChRs, as indicated below). We have also rule out that DEA caused an unspecific blockade of LGICs, since $100 \mu\text{M}$ DEA showed no effect on GABA-elicited currents from rat neuronal GABA_ARs microtransplanted to *Xenopus* oocytes (see Supplementary Figure 1), which belong to the same family of LGICs. Nevertheless, it remains to be determined whether or not DEA has specific effects or different potencies on the neuronal nAChR subtypes.

DEA, as the entire lidocaine molecule, inhibited nAChRs in a voltage-dependent manner, the blockade being higher at negative potentials, which strongly suggests an open-channel blockade caused by this positively-charged molecule. However, at potentials more negative than -70 mV , the I_{ACh} remaining upon DEA did not decrease but rather increased (Figure 3C),

in contrast to the voltage-dependent effect of lidocaine, which kept increasing at very negative potentials (Alberola-Die et al., 2011). This difference might arise because of the smaller size of DEA, which could allow its permeation through the channel pore, by a “punch-through” mechanism, as it has been proposed for other small molecules in this receptor (Sine and Steinbach, 1984). In fact, from data on permeation of various sized cations, the estimated size of the pore was 8.4 Å (Cohen et al., 1992), which agrees well with the 9–10 Å pore diameter estimated from images of the open state of *Torpedo* nAChRs (Unwin, 1995). Given the small size of DEA (smaller than 7 Å in the long axis; see **Figure 3C**) and its positive charge in physiological solutions, this molecule could be electrostatically forced to move from its binding site to the cytoplasm, releasing the nAChR from its blockade at highly negative potentials. Virtual docking assays also showed that both DEA and lidocaine might bind inside the channel pore (**Figure 6**), though lidocaine protruding more into it (**Figure 6** and Supplementary Figure 2). The predicted DEA binding sites within the pore were slightly shallower than the lidocaine loci, in spite that for both molecules the estimated δ -values were similar (about 30% of the electrical field). Nevertheless, interpretation of the electrical distances (δ) is not always straightforward because the IC_{50} s are dependent not only on the equilibrium dissociation constants of the blockers, but also on the kinetics of the transitions between functional states (Pascual and Karlin, 1998). Interestingly, whereas lidocaine could interact within the pore at all five subunits, DEA interacted with residues belonging to just two subunits (α - β or α - δ), which might cause fainter interactions and thus facilitate the punch-through occurrence.

I_{ACh} blockade by DEA could not be fully reversed by applying pulses to positive potentials, which should unplug the positively charged DEA from the channel pore. Thus, at +60 mV, the I_{ACh} inhibition caused by 100 μ M DEA (roughly 30%) was voltage-independent (**Figure 3C**) and, so, it is not mediated by open-channel blockade of nAChRs. Consequently, DEA should also bind outside the pore and cause the blockade of resting nAChRs, as it has been proposed for other non-competitive blockers of nAChRs, including some cholinesterase inhibitors, such as tacrine (Prince et al., 2002) or edrophonium (Olivera-Bravo et al., 2007) and local anesthetics, such as tetracaine (Papke and Oswald, 1989; Gallagher and Cohen, 1999; Middleton et al., 1999) or lidocaine (Alberola-Die et al., 2011). DEA binding to closed nAChRs is also supported by its pronounced inhibitory action on nAChRs when it is pre-applied to the cells before being co-applied with ACh (**Figure 5**). This was particularly evident at high ACh doses (**Figure 5C**), since the fast opening of nAChR channels by ACh prevented the DEA block of resting receptors when DEA and ACh were solely co-applied. As DEA caused nAChR blockade at positive potentials and it is permanently charged (preventing their permeation through the membrane), it is suggested that DEA blocked resting nAChRs by its binding to extracellular residues outside the pore, which differs from the binding of tetracaine within the channel on resting nAChRs (Gallagher and Cohen, 1999; Middleton et al., 1999). Accordingly, our virtual docking results showed that both DEA and lidocaine bind at multiple loci on the EC domain, including

several sites at intersubunit interphases. Of particular functional relevance seems the DEA binding to a cavity near the ligand-binding pocket at the α - γ interphase (**Figure 6A₃**). Interestingly, this DEA binding site is very similar to that found for ketamine binding to the homologous GLIC channel, as resolved by X-ray crystallography at 2.99 Å (Pan et al., 2012; compare panels A₁ and A₂ of Supplementary Figure 2). It is also remarkable that ketamine, a small water soluble amine, blocked GLIC channels with an IC_{50} of 58 μ M, which is very close to that of DEA for nAChRs blockade (**Figure 1**). Even more, ketamine also blocked, at this range of concentrations, mammalian muscle- and neuronal-type nAChRs by acting on the open and closed states (Scheller et al., 1996), although it neither blocked GABA_AR nor GlyRs (Yamakura et al., 2000). Therefore, since DEA, and most likely ketamine, binding site at the α - γ interphase of nAChRs overlaps partially with the ACh-binding pocket, it might additionally inhibit nAChR by competitive antagonism. Finally, our docking results do not support a role of DEA as a partial nAChR agonist because of, at least partially, its small size. Thus, although DEA interacted with A-loop residues from the α -subunit, this binding site was 15–18 Å below the C loop, which is known to contribute with key residues to the ligand-binding site (Corringer et al., 2000). Moreover, the docking-assays did not show any equivalent hotspot for DEA binding at the interphase of α - δ subunits. By contrast, lidocaine interacted with residues from all α loops conforming the ACh-binding pocket, both at the α - γ and α - δ interphases, which suggests a possible additional action as a competitive antagonist.

In conclusion, we found that DEA, a small charged molecule closely resembling the hydrophilic moiety of the lidocaine molecule, mimics some, but not all the inhibitory actions of the complete lidocaine molecule on nAChRs. So, DEA accounts for the voltage-dependent blockade of nAChRs by lidocaine and might also contribute to inhibit resting nAChRs, likely acting on extracellular residues lying outside the channel pore.

AUTHOR CONTRIBUTIONS

All authors listed, have made substantial, direct and intellectual contribution to the work, and approved it for publication.

FUNDING

This work was supported by grants BFU2012-31359, BFU2012-39092-C02-01, BFU2011-25920, and CSD2008-00005 from the MINECO and PROMETEO/2014/11 from GVA (Spain).

ACKNOWLEDGMENTS

We thank Mr. Simón Moya for expert technical assistance.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <http://journal.frontiersin.org/article/10.3389/fnmol.2016.00012>

REFERENCES

- Alberola-Die, A., Martínez-Pinna, J., González-Ros, J. M., Ivorra, I., and Morales, A. (2011). Multiple inhibitory actions of lidocaine on *Torpedo* nicotinic acetylcholine receptors transplanted to *Xenopus* oocytes. *J. Neurochem.* 117, 1009–1019. doi: 10.1111/j.1471-4159.2011.07271.x
- Alberola-Die, A., Reboreda, A., Lamas, J. A., and Morales, A. (2013). Lidocaine effects on acetylcholine-elicited currents from mouse superior cervical ganglion neurons. *Neurosci. Res.* 75, 198–203. doi: 10.1016/j.neures.2013.01.005
- Alberola-Die, A., Soriano, S., Fernández-Ballester, G., Martínez-Pinna, J., Gonzalez-Ros, J. M., Ivorra, I., et al. (2012). Diethylamine blocks muscle-type nicotinic acetylcholine receptors by mimicking the action of charged lidocaine. *Acta Physiol.* 206(Suppl. 693), P120.
- Albuquerque, E. X., Pereira, E. F. R., Alkondon, M., and Rogers, S. W. (2009). Mammalian acetylcholine receptors: from structure to function. *Physiol. Rev.* 89, 73–120. doi: 10.1152/physrev.00015.2008
- Arias, H. R. (2010). Positive and negative modulation of nicotinic receptors. *Adv. Protein. Chem. Struct. Biol.* 80, 153–203. doi: 10.1016/B978-0-12-381264-3.00005-9
- Cecchini, M., and Changeux, J. P. (2015). The nicotinic acetylcholine receptor and its prokaryotic homologues: structure, conformational transitions & allosteric modulation. *Neuropharmacology* 96, 137–149. doi: 10.1016/j.neuropharm.2014.12.006
- Chatzidakis, A., and Millar, N. S. (2015). Allosteric modulation of nicotinic acetylcholine receptors. *Biochem. Pharmacol.* 97, 408–417. doi: 10.1016/j.bcp.2015.07.028
- Cohen, B. N., Labarca, C., Davidson, N., and Lester, H. A. (1992). Mutations in M2 alter the selectivity of the mouse nicotinic acetylcholine receptor for organic and alkali metal cations. *J. Gen. Physiol.* 100, 373–400. doi: 10.1085/jgp.100.3.373
- Corringer, P. J., Le Novère, N., and Changeux, J. P. (2000). Nicotinic receptors at the amino acid level. *Annu. Rev. Pharmacol. Toxicol.* 40, 431–458. doi: 10.1146/annurev.pharmtox.40.1.431
- Dani, J. A., and Bertrand, D. (2007). Nicotinic acetylcholine receptors and nicotinic cholinergic mechanisms of the central nervous system. *Annu. Rev. Pharmacol. Toxicol.* 47, 699–729. doi: 10.1146/annurev.pharmtox.47.120505.105214
- Duan, Y., Wu, C., Chowdhury, S., Lee, M. C., Xiong, G., Zhang, W., et al. (2003). A point-charge force field for molecular mechanics simulations of proteins based on condensed-phase quantum mechanical calculations. *J. Comput. Chem.* 24, 1999–2012. doi: 10.1002/jcc.10349
- Gallagher, M. J., and Cohen, J. B. (1999). Identification of amino acids of the *Torpedo* nicotinic acetylcholine receptor contributing to the binding site for the noncompetitive antagonist [³H]tetracaine. *Mol. Pharmacol.* 56, 300–307.
- Gentry, C. L., and Lukas, R. J. (2001). Local anesthetics noncompetitively inhibit function of four distinct nicotinic acetylcholine receptor subtypes. *J. Pharmacol. Exp. Ther.* 299, 1038–1048.
- Gonzalez-Gutierrez, G., and Grosman, C. (2015). The atypical cation-conduction and gating properties of ELIC underscore the marked functional versatility of the pentameric ligand-gated-ion-channel fold. *J. Gen. Physiol.* 146, 15–36. doi: 10.1085/jgp.201411333
- Gotti, C., and Clementi, F. (2004). Neuronal nicotinic receptors: from structure to pathology. *Prog. Neurobiol.* 74, 363–396. doi: 10.1016/j.pneurobio.2004.09.006
- Guex, N., and Peitsch, M. C. (1997). SWISS-MODEL and the Swiss-PdbViewer: an environment for comparative protein modeling. *Electrophoresis* 18, 2714–2723. doi: 10.1002/elps.1150181505
- Hara, K., and Sata, T. (2007). The effects of the local anesthetics lidocaine and procaine on glycine and gamma-aminobutyric acid receptors expressed in *Xenopus* oocytes. *Anesth. Analg.* 104, 1434–1439. doi: 10.1213/01.ane.0000261509.72234.a6
- Hilf, R. J., Bertozzi, C., Zimmermann, I., Reiter, A., Trauner, D., and Dutzler, R. (2010). Structural basis of open channel block in a prokaryotic pentameric ligand-gated ion channel. *Nat. Struct. Mol. Biol.* 17, 1330–1336. doi: 10.1038/nsmb.1933
- Hille, B. (1966). Common mode of action of three agents that decrease the transient change in sodium permeability in nerves. *Nature* 5042, 1220–1222. doi: 10.1038/2101220a0
- Hurst, R., Rollema, H., and Bertrand, D. (2013). Nicotinic acetylcholine receptors: from basic science to therapeutics. *Pharmacol. Therapeut.* 137, 22–54. doi: 10.1016/j.pharmthera.2012.08.012
- Ivorra, I., Fernández, A., Gal, B., Aleu, J., González-Ros, J. M., Ferragut, J. A., et al. (2002). Protein orientation affects the efficiency of functional protein transplantation into the *Xenopus* oocyte membrane. *J. Membr. Biol.* 185, 117–127. doi: 10.1007/s00232-001-0118-x
- Krieger, E., Darden, T., Nabuurs, S. B., Finkelstein, A., and Vriend, G. (2004). Making optimal use of empirical energy functions: force-field parameterization in crystal space. *Proteins* 57, 678–683. doi: 10.1002/prot.20251
- Krieger, E., Koraimann, G., and Vriend, G. (2002). Increasing the precision of comparative models with YASARA NOVA; a self-parameterizing force field. *Proteins* 47, 393–402. doi: 10.1002/prot.10104
- Kusano, K., Miledi, R., and Stinnakre, J. (1982). Cholinergic and catecholaminergic receptors in the *Xenopus* oocyte membrane. *J. Physiol.* 328, 143–170. doi: 10.1113/jphysiol.1982.sp014257
- Liu, H., Atkins, J., and Kass, R. S. (2003). Common molecular determinants of flecainide and lidocaine block of heart Na⁺ channels: evidence from experiments with neutral and quaternary flecainide analogues. *J. Gen. Physiol.* 121, 199–214. doi: 10.1085/jgp.20028723
- Middleton, R. E., Strnad, N. P., and Cohen, J. B. (1999). Photoaffinity labeling the *Torpedo* nicotinic acetylcholine receptor with [³H]tetracaine, a nondesensitizing noncompetitive antagonist. *Mol. Pharmacol.* 56, 290–299.
- Morales, A., Aleu, J., Ivorra, I., Ferragut, J. A., González-Ros, J. M., and Miledi, R. (1995). Incorporation of reconstituted acetylcholine receptors from *Torpedo* into the *Xenopus* oocyte membrane. *Proc. Natl. Acad. Sci. U.S.A.* 92, 8468–8472. doi: 10.1073/pnas.92.18.8468
- Morris, G. M., Huey, R., and Olson, A. J. (2008). Using AutoDock for ligand-receptor docking. *Curr. Protoc. Bioinformatics* 24, 8.14.1–8.14.40. doi: 10.1002/0471250953.bi0814s24
- Narahashi, T., Frazier, D. T., and Yamada, M. (1969). Cationic forms of local anesthetics block action potentials from inside nerve membrane. *Nature* 223, 748–749. doi: 10.1038/223748a0
- Olivera-Bravo, S., Ivorra, I., and Morales, A. (2005). The acetylcholinesterase inhibitor BW284c51 is a potent blocker of *Torpedo* nicotinic AChRs incorporated into the *Xenopus* oocyte membrane. *Br. J. Pharmacol.* 144, 88–97. doi: 10.1038/sj.bjpp.0705965
- Olivera-Bravo, S., Ivorra, I., and Morales, A. (2007). Diverse inhibitory actions of quaternary ammonium cholinesterase inhibitors on *Torpedo* nicotinic ACh receptors transplanted to *Xenopus* oocytes. *Br. J. Pharmacol.* 151, 1280–1292. doi: 10.1038/sj.bjpp.0707329
- Pan, J., Chen, Q., Willenbring, D., Mowrey, D., Kong, X. P., Cohen, A., et al. (2012). Structure of the pentameric ligand-gated ion channel GLIC bound with anesthetic ketamine. *Structure* 20, 1463–1469. doi: 10.1016/j.str.2012.08.009
- Papke, R. L., and Oswald, R. E. (1989). Mechanisms of noncompetitive inhibition of acetylcholine-induced single-channel currents. *J. Gen. Physiol.* 93, 785–811. doi: 10.1085/jgp.93.5.785
- Pascual, J. M., and Karlin, A. (1998). Delimiting the binding site for quaternary ammonium lidocaine derivatives in the acetylcholine receptor channel. *J. Gen. Physiol.* 112, 611–621. doi: 10.1085/jgp.112.5.611
- Prince, R. J., Pennington, R. A., and Sine, S. M. (2002). Mechanism of tacrine block at adult human muscle nicotinic acetylcholine receptors. *J. Gen. Physiol.* 120, 369–393. doi: 10.1085/jgp.20028583
- Sánchez, J. A., Dani, J. A., Siemen, D., and Hille, B. (1986). Slow permeation of organic cations in acetylcholine receptor channels. *J. Gen. Physiol.* 87, 985–1001. doi: 10.1085/jgp.87.6.985
- Scheller, M., Bufler, J., Hertle, I., Schneck, H. J., Franke, C., and Kochs, E. (1996). Ketamine blocks currents through mammalian nicotinic acetylcholine receptor channels by interaction with both the open and the closed state. *Anesth. Analg.* 83, 830–836.
- Sergeeva, V. S., Efrementko, E. N., Kazankov, G. M., and Varfolomeyev, S. D. (2000). Double effect of organic amines (activation and inhibition) on the phosphotriesterase. *J. Mol. Cat. B Enzymatic* 10, 571–576. doi: 10.1016/S1381-1177(00)00102-8
- Sine, S. M., and Steinbach, J. H. (1984). Agonists block currents through acetylcholine receptor channels. *Biophys. J.* 46, 277–283. doi: 10.1016/S0006-3495(84)80422-9

- Steinbach, A. B. (1968). Alteration by Xilocaine (lidocaine) and its derivatives of the time course of the end plate potential. *J. Gen. Physiol.* 52, 144–161. doi: 10.1085/jgp.52.1.144
- Taly, A., Corringier, P. J., Guedin, D., Lestage, P., and Changeux, J. P. (2009). Nicotinic receptors: allosteric transitions and therapeutic targets in the nervous system. *Nat. Rev. Drug Discov.* 8, 733–750. doi: 10.1038/nrd2927
- Trellakis, S., Benzenberg, D., Urban, B. W., and Friederich, P. (2006). Differential lidocaine sensitivity of human voltage-gated potassium channels relevant to the auditory system. *Otol. Neurotol.* 27, 117–123. doi: 10.1097/01.mao.0000186443.11832.8a
- Ueta, K., Suzuki, T., Sugimoto, M., Uchida, I., and Mashimo, T. (2007). Local anesthetics have different mechanisms and sites of action at recombinant 5-HT₃ receptors. *Reg. Anesth. Pain Med.* 32, 462–470. doi: 10.1097/00115550-200711000-00002
- Unwin, N. (1995). Acetylcholine receptor channel imaged in the open state. *Nature* 373, 37–43. doi: 10.1038/373037a0
- Unwin, N. (2005). Refined structure of the nicotinic acetylcholine receptor at 4 Å resolution. *J. Mol. Biol.* 346, 967–989. doi: 10.1016/j.jmb.2004.12.031
- Woodhull, A. M. (1973). Ionic blockage of sodium channels in nerve. *J. Gen. Physiol.* 61, 687–708. doi: 10.1085/jgp.61.6.687
- Xiong, Z., and Strichartz, G. R. (1998). Inhibition by local anesthetics of Ca²⁺ channels in rat anterior pituitary cells. *Eur. J. Pharmacol.* 363, 81–90. doi: 10.1016/S0014-2999(98)00769-9
- Yamakura, T., Chavez-Noriega, L. E., and Harris, R. A. (2000). Subunit-dependent inhibition of human neuronal nicotinic acetylcholine receptors and other ligand-gated ion channels by dissociative anesthetics ketamine and dizocilpine. *Anesthesiology* 92, 1144–1153. doi: 10.1097/00000542-200004000-00033

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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