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Levels of dioxin-like PCBs in low-volume serum samples of male patients attending fertility clinics

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Abstract An accurate and easy method for the extraction, cleanup, and HRGC-HRMS analysis of dioxin-like PCBs (DL-PCBs) in low-volume serum samples (1 mL) was developed. Serum samples were extracted several times using *n*-hexane and purified by acid washing. Recovery rates of labeled congeners ranged from 70 to 110 % and the limits of detection were below 1 pg/g on lipid basis. Although human studies are limited and contradictory, several studies have shown that DL-PCBs can have adverse effects on the male reproductive system. In this way, the present method was applied to 21 serum samples of male patients attending fertility clinics. The total levels obtained for the patients ranged from 6.90 to 84.1 pg WHO-TEQ/g lipid, with a mean value of 20.3 pg WHO-TEQ/g lipid. The predominant PCBs (the sum of PCB 118, 156, and 105) contributed 67 % to the mean concentration of total DL-PCBs in the samples analyzed.

Keywords Human serum · Male infertility · DL-PCBs · HRGC-HRMS · Pollutants · Analytical method

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Introduction

The possible relationship between environmental and occupational exposure to toxic substances and the decreasing quality of semen and the consequent increase in male infertility is a subject of great scientific interest (Rubes et al. 2005). Several laboratory animal studies have indicated that some endocrine disruptors as polychlorinated biphenyls (PCBs), polychlorinated dibenzo-*p*-dioxins (PCDDs), and polychlorinated dibenzofurans (PCDFs) may affect human reproduction and may be linked to reduced fertility in women and men (Cok et al. 2008).

The World Health Organization has estimated that nearly 8 % of couples around the world present some forms of infertility problems during their reproductive lives. This means that 50 to 80 million people suffer fertility problems (Woodruff et al. 2010). Specially, there are evidence for decreasing quality of semen during the last decades (Carlsen et al. 1992). Environmental factors (exposure to traffic exhaust fumes, dioxins, combustion products, etc.) and several lifestyles as obesity and smoking could negatively affect human fertility, underlining the importance of environmental and lifestyle impacts throughout the life course (Sharpe 2010).

Their lipophilic nature, low biodegradability, and chemical stability, together with the indiscriminate use of dioxin-contaminated PCBs, ensure their ubiquity and persistence in the environment (Esposito et al. 2014). Exposure of the general population to dioxin-like PCBs occurs mainly through food, and also dermal contact and inhalation. In order to quantify trends and patterns of human exposure to chemical agents, monitoring programs play a fundamental role (Angerer et al. 2007; Needham et al. 2007; Patterson et al. 2009; Viso et al. 2009). Moreover, monitoring programs act as a framework to evaluate health impacts and the effectiveness of public and private policies aimed at decreasing exposure to POPs (Department of Health and Human Services 2009; Viso et al. 2009).

63 There is little information on infertility and related condi- 113
64 tions on human exposure to DL-PCBs, most of the papers 114
65 published are relating to the presence of others PCBs conge- 115
66 ners in different fluids of the human reproductive tract such as 116
67 the seminal fluid, cervical mucus, and follicular fluid. 117

68 Nowadays, there is an increasing demand to reduce the size 118
69 of the specimen collected for human biomonitoring (Patterson 119
70 et al. 2011). In this way, there are several studies analyzing the 120
71 DL-PCBs content in serum samples from different group of 121
72 populations; however, there are scarce studies reporting the 122
73 analysis of serum from individuals using low-volume sam- 123
74 ples, due to the difficulty in reaching the required selectivity 124
75 and sensibility for the analysis of these compounds. 125

76 The goal of this work is to develop an analytical method for 126
77 the analysis of small-volume human serum samples (1 mL) in 127
78 order to provide an individualized patient study. The optimi- 128
79 zation of the analytical method has been carried out consider- 129
80 ing these important aspects: minimizing both the consumption 130
81 of chemical reagents and analysis time, in order to minimize 131
82 the cost of the analysis, but keeping the quality standards 132
83 required for the analysis of dioxin-like PCBs. 133

84 Methods and materials

85 In this study, 21 blood serum samples were collected from 134
86 male patients recruited for assisted reproduction at the IVF 135
87 Spain Treatment Clinic from May 2012 to June 2014. The 136
88 age of the men ranged from 30 to 55 years and the body mass 137
89 index (BMI) ranged between 21.0 and 31.8 (mean 25.2). Gen- 138Q
90 eral information regarding age, place of residence, smoking 139
91 habits, dietary habits, and alcohol consumption, along with 140
92 occupational, residential, and clinical histories were collected. 141
93 Each patient provided an informed consent after receiving a 142
94 detailed explanation of the study and potential consequences 143
95 and signed an informed consent form. 144

96 Blood samples were obtained by venipuncture and collect- 145
97 ed in Vacutainer test tubes without anticoagulant and were 146
98 immediately transferred to a glass centrifuge tube. After cen- 147
99 trifugation and decantation, the serums were stored in the dark 148
100 at -20°C until analysis. 149

101 The procedure used for analytical determination was 150
102 adapted from a previously published method (Grimalt et al. 151
103 2010) for the analysis by gas chromatography with electron 152
104 capture detection of several halogenated organic pollutants, 153
105 including seven PCB congeners (PCBs 28, 52, 101, 118, 154
106 138, 153, and 180). 155

107 From an analytical point of view, gas chromatography 156
108 (GC) coupled with magnetic sector high-resolution mass spec- 157
109 trometry (HRMS) is the most sensitive and specific technique 158
110 in the field of ultra-trace measurement of dioxin-like PCB. 159
111 The use of isotope dilution methodology, based on ^{13}C -la- 160
112 beled standards at a minimum mass resolution of 10,000 161

113 resolving power in the selected ion monitoring (SIM) acqui- 114
115 sition mode, allows enhanced selectivity and the lowest sen- 116
117 sitivity threshold (Patterson et al. 2011). In this way, in this 117
118 study, the extraction and cleanup with *n*-hexane and concen- 118
119 trated sulfuric acid was followed by HRGC-HRMS determi- 119
120 nation. Figure 1 shows the analytical procedure optimized for 120
121 the determination of DL-PCBs in low-volume samples of hu- 121
122 man serum. To improve the separation between aqueous and 122

123 The samples were analyzed using an Agilent HP5890 gas 123
124 chromatograph equipped with programmable temperature va- 124
125 porization (PTV) inlet, coupled to a Micromass Autospec 125
126 Ultima-NT mass spectrometer. An Agilent DB5-MS chroma- 126
127 tographic column ($60\text{ m}\times 0.25\text{ mm i.d.}\times 0.25\text{ }\mu\text{m}$) was 127
128 used for the analysis. Isotopically labeled standards were ob- 128
129 tained from Wellington Laboratories Inc. (Canada). *n*-Hexane 129
130 and sulfuric acid were for organic trace analysis and were 130
131 purchased from Merck (Germany). 131

132 Concentration of lipophilic pollutants in serum are usually 132
133 expressed on a lipid weight basis instead of on a fresh weight 133
134 basis, with a view to obtain biomarker values that reflect body 134
135 burden of persistent organohalogen pollutants. In this work, 135
136 the total lipid concentration was determined by the sum of the 136
137 triglyceride and cholesterol concentrations, according to the 137

138Q following regression (Phillips et al. 1989): 138Q

139 $\text{Total lipid (mg/dL)} = 62.3 + \text{triglycerides (mg/dL)} + 2.27 * \text{total cholesterol (mg/dL)}$ 139

140 The levels of dioxin-like PCBs are expressed in picograms 140
141 of compound per gram of lipid (pg/g lipid) and the toxic 141
142 equivalents were calculated according to the toxic equivalen- 142
143 cy factors published in 2005 by the World Health Organiza- 143
144 tion (WHO₂₀₀₅-TEFs) (van den Berg et al. 2006). 144

145 Samples were spiked with ^{13}C -labeled internal standards 145
146 before extraction and with ^{13}C -labeled recovery standards be- 146
147 fore HRGC-HRMS analysis. Recovery rates of labeled conge- 147
148 ners ranged from 70 to 110 % and the limits of detection 148
149 were below 1 pg/g on lipid basis. Quality assurance criteria 149
150 were based on the minimum requirements described in US 150
151 EPA method 1668C for dioxin-like-PCBs (US EPA 2010). 151
152 A procedural blank was associated with each batch of four 152
153 samples and processed in the same manner. The congeners 153
154 below the limit of detection (LOD) were calculated consider- 154
155 ing a concentration equal to their respective LOD. 155

157 Results and discussion

158 In this study, DL-PCBs have been determined in 21 serum 158
159 samples from male patients recruited for assisted reproduc- 159
160 tion. Ten patients aged between 30 and 40 years and the rest 160
161 aged between 41 and 55 years. The BMI was categorized into 161
162 three groups: BMI <25, not overweight; BMI 25-29.9, 162

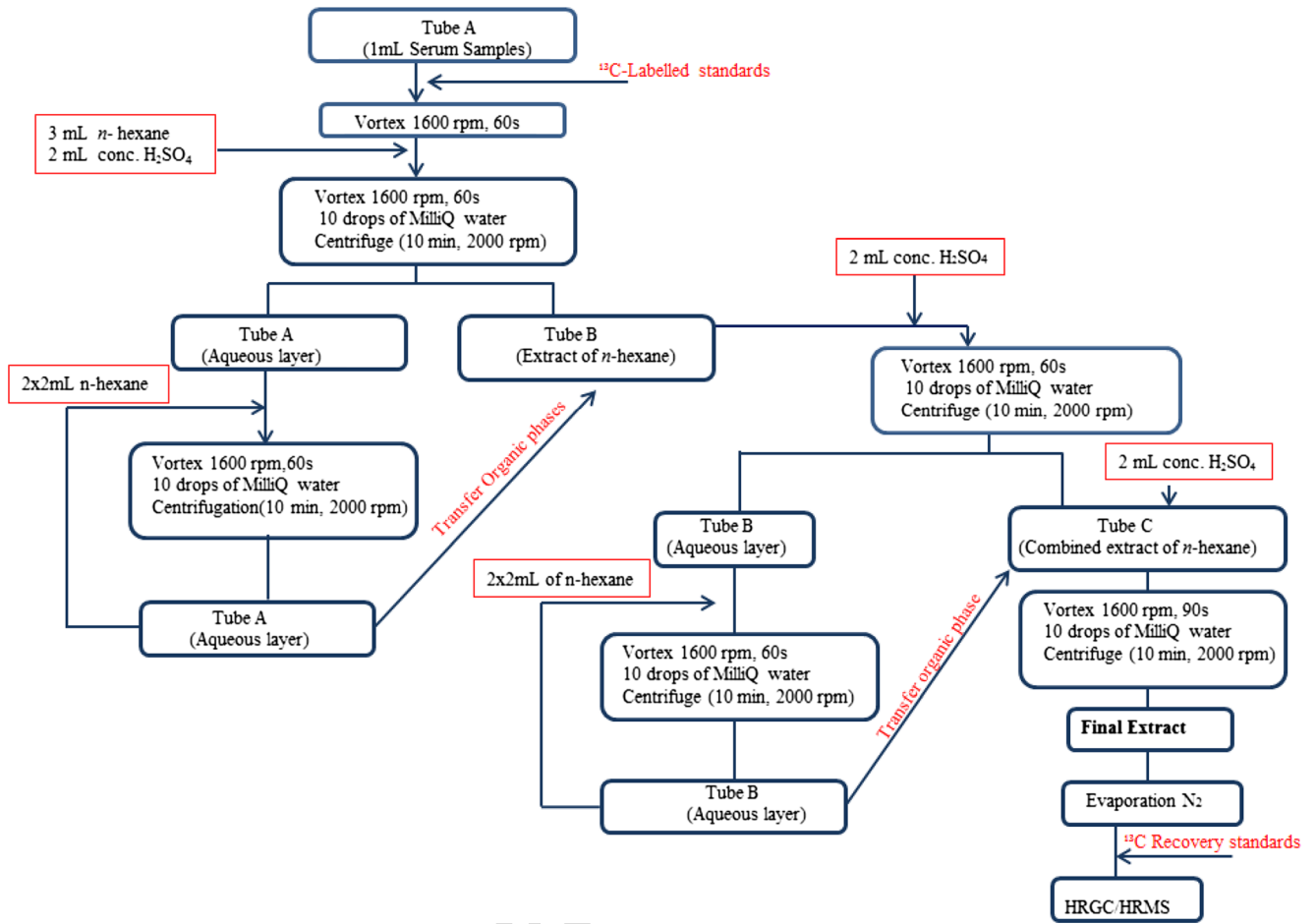
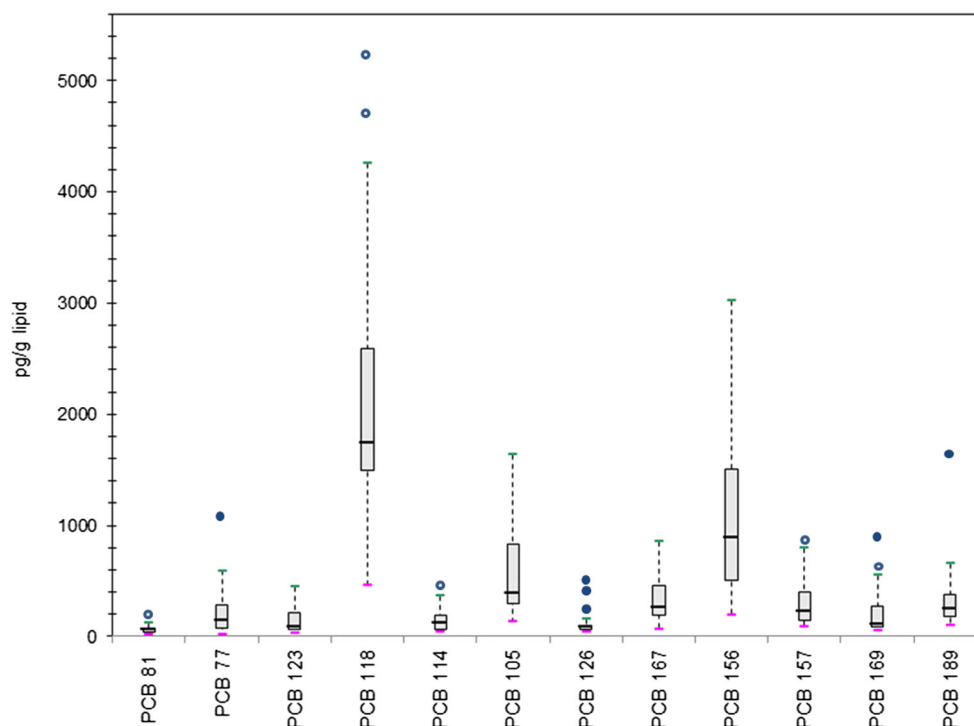


Fig. 1 Scheme of the analytical procedure developed

Q3 t1.1 Table 1 Levels of DL-PCB congeners in the blood serum, expressed as pg/g lipid, and total DL-PCB concentrations, expressed as pg WHO-TEQ/g lipid

t1.2	Mean	Median	Standard deviation	Minimum	Maximum	% above LOD	% WHO-TEQ
t1.3	Non-ortho dioxin-like PCBs						
t1.4	230	147	254	18.7	1110	81	0.14
t1.5	72.5	61.5	53.8	14.3	250	19	0.15
t1.6	138	88.9	156	41.2	638	57	65.6
t1.7	211	109	232	56.1	1010	81	30.1
t1.8	Mono-ortho dioxin-like PCBs						
t1.9	527	394	315	137	1040	100	0.11
t1.10	147	117	100	34.2	387	76	0.03
t1.11	2170	1740	1270	462	5220	100	0.47
t1.12	138	87.8	94.3	23.0	368	62	0.03
t1.13	1080	886	716	196	2630	100	0.25
t1.14	269	227	180	85.5	823	95	0.06
t1.15	322	262	212	66.6	788	95	0.07
t1.16	345	244	337	103	1690	95	2.95
t1.17	Sum						
t1.18	5650	5210	2780	2020	13,200		
t1.19	20.3	11.6	22.4	6.90	84.1		

Fig. 2 Boxplots of median values of DL-PCBs



163 overweight; and BMI ≥ 30 , obese. Only one patient had a BMI
 164 >30 , while for the rest of the patients, the number of not
 165 overweight and overweight was the same. To describe the
 166 levels of DL-PCBs, we calculated the medians and arithmetic
 167 means, their standard deviations, the maximum and minimum
 168 values, and the percentage of samples that present a concen-
 169 tration above the limit of detection. An overview of the values
 170 obtained is shown in Table 1.

171 The total levels of DL-PCBs obtained for the patients
 172 ranged from 6.90 to 84.1 pg WHO-TEQ/g lipid, with a mean
 173 value of 20.3 pg WHO-TEQ/g lipid. The predominant conge-
 174 ners, in order, were PCB 118, PCB 156, and PCB 105, with
 175 mean values of 2170, 1080, and 527 pg/g lipid, respectively.
 176 The predominant PCBs (the sum of the three previously men-
 177 tioned) contributed 67 % to the mean concentration of total
 178 DL-PCBs in the samples analyzed. Two congeners, PCB 126
 179 and PCB 169 contributed 96 % to the total WHO-TEQ value,
 180 due to their high toxic equivalency factors. The mean value for
 181 non-ortho PCBs was 20.2 pg WHO-TEQ/g lipid and for
 182 mono-ortho PCBs 0.53 pg WHO-TEQ/g lipid. All the conge-
 183 ners were detected in the samples, although PCB 81 was de-
 184 tected only in 4 of the 21 samples analyzed. PCBs 118, 156,
 185 and 105 were detected in all the serum samples.

186 The total TEQ at the 90th percentile of the US population
 187 in NHANES 2003–2004 was 30.0 pg/g of lipid (Patterson
 188 et al. 2009) for total dioxin-like compounds (PCDD/Fs and
 189 DL-PCBs). Wittsiepe et al. (2007) found a total dioxin-like
 190 compounds value of 28.36 pg TEQ/g lipid in the blood from
 191 pregnant women. In a review of worldwide literature (works
 192 published from 1989 until June 2010) (Consonni et al. 2012)

193 in general populations not directly exposed to recognized
 194 emission sources, a mean value of 3.7 pg WHO-TEQ/g lipid
 195 for the mono-ortho and non-ortho PCBs is given. Several
 196 authors have studied the DL-PCBs in blood serum samples
 197 from populations living in industrial areas or near urban waste
 198 treatment or incineration plants. Zubero et al. (2009) com-
 199 pared the DL-PCBs in serum samples of different populations
 200 and found that the mean values ranged between 4.1 and
 201 15.6 pg WHO-TEQ/g lipid; the values obtained for the male
 202 patients facing fertility problems are higher than those present-
 203 ed by these studies.

204 Boxplots with whiskers for DL-PCBs congeners distribu-
 205 tion are shown in Fig. 2. Boxes represent interquartile ranges
 206 (IQR), medians are marked in each box with a line, the whis-
 207 ker on the appropriate side is taken to 1.5·IQR from the quar-
 208 tile (the “inner fence”) and individual outlying data points are
 209 displayed as unfilled circles (for suspected outliers) or filled
 210 circles (for outliers). Suspected outliers are above 1.5·IQR and
 211 individual outlying data points are above 3·IQR. There are
 212 two serum samples analyzed that present several outliers
 213 values for different isomers, one serum for PCB 169, PCB

Table 2 Concentrations of DL-PCBs (p WHO-TEQ/g lipid) in male serum samples by age and BMI

	Mean	Median	Minimum	Maximum	
30–40 years	19.2	11.5	7.40	84.2	t2.2
41–55 years	21.3	11.4	6.90	83.9	t2.3
BMI <25	29.9	17.8	6.90	84.2	t2.4
BMI 25–29	11.9	11.1	7.40	19.2	t2.5

214 189, and PCB 126 and another sample for PCB 81, PCB 77,
215 and PCB 126.

216 Table 2 summarizes the concentrations of DL-PCBs
217 (pg WHO-TEQ/g lipid) in male serum samples by age and
218 BMI. Generally, the concentration of PCBs in human samples
219 tended to increase with age (Chen et al. 2013), which is con-
220 sistent with the fact that these compounds are liposoluble sub-
221 stances that accumulate in the adipose tissue. The results ob-
222 tained in this study are in accordance with this general trend,
223 although the values are quite similar in the two groups of age.
224 The role played by BMI has not yet been established and there
225 is controversy with respect to the direction of its association.
226 One of the main findings is that some of the PCBs are posi-
227 tively related to fat mass, while others show an opposite asso-
228 ciation. Typically the low-chlorinated PCBs were related to fat
229 mass in a positive way, while the opposite trend was generally
230 seen for the highly chlorinated compounds (Ronn et al. 2011).
231 In this study, overweight subjects had much lower mean
232 values than the normal weight patients; this could be ex-
233 plained by the distribution of a given body burden in a higher
234 amount of fat, so the lipid concentration is lower in the over-
235 weight patients.

236 PCBs have been associated with a range of adverse health
237 effects, including adverse effects on reproduction. However,
238 the human data available on this topic remain vague and large-
239 ly inconclusive (Meeker et al. 2011). Although there are sev-
240 eral studies that try to relate the serum concentrations of PCBs
241 (mainly analyzed by GC-ECD) with male and female fertility
242 problems, we have not found any study about the levels of
243 DL-PCBs in male serum samples of patients facing fertility
244 problems, and that makes the comparison of our results a quite
245 complicated task. Dallinga et al. (2002) analyzed PCB 118,
246 153, 138, and 180 in blood samples from a group of men with
247 very poor semen quality and another group with normal se-
248 men quality in order to determine whether a difference in these
249 levels could be established and found a significant relationship
250 between the combined levels of these PCBs and the corre-
251 sponding level in seminal plasma.

252 In this work, an analytical method for the analysis of
253 dioxin-like PCBs in low volume (1 mL) serum samples has
254 been carried out, obtaining an easy and cost-effective method
255 with appropriate detection limits. This method has allowed to
256 obtain the amount of DL-PCBs present in the serum of 21 men
257 attending fertility clinics who suffer different fertility prob-
258 lems. Despite the limitations of our study, to the best of our
259 knowledge, this is the first study to investigate the levels of
260 DL-PCBs in the serum of male patients facing fertility prob-
261 lems. Further number of samples, having a better understand-
262 ing of the lifestyle of patients, and knowing other factors that
263 could affect the relationship between these pollutants and fer-
264 tility problems could be very useful. It would also be interest-
265 ing to compare the results of this study with serum samples
266 from fertile patients.

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272

Compliance with ethical standards 273

Consent to participate Each patient provided an informed consent 274Q6
after receiving a detailed explanation of the study and potential conse- 275
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