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### Aqueous enantioselective aldol reaction of methyl- and phenylglyoxal organocatalyzed by N-Tosyl-(Sa)-binam-Lprolinamide

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## Aqueous enantioselective aldol reaction of methyl- and phenylglyoxal organocatalyzed by N-Tosyl- $(S_a)$ -binam-L-prolinamide.

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**Abstract:** The direct aldol reaction between methylglyoxal (40% aqueous solution) or phenylglyoxal monohydrate and ketones or aldehydes is catalyzed by *N*-tosyl-( $S_a$ )-binam-L-prolinamide to afford the corresponding chiral  $\gamma$ -oxo- $\beta$ -hydroxy carbonyl compounds, mainly as *anti* isomers with enantioselectivities up to 97%.

**Key words:** methylglyoxal, organocatalysis, aldol, prolinamide, aqueous conditions.

Methylglyoxal (1a), an endogenous  $\alpha$ -oxoaldehyde which is a potent protein modifier,<sup>1</sup> is a versatile reagent for the synthesis of heterocyclic compounds<sup>2</sup> using organocatalyzed methodologuies.<sup>3</sup> However, it use as electrophile in related organocatalyzed enantioselective aldol processes<sup>4</sup> have been scarcely described,<sup>5</sup> although it would afford to synthetically important chiral  $\gamma$ -oxo- $\beta$ -hydroxy carbonyl compounds. Probably, the reluctant use of this type of  $\alpha$ -alkyl- $\alpha$ -oxo aldehydes as electrophiles is due to their facile hydratation and polymerization tendency. On the other hand, methylglyoxal is only commercially available as an aqueous solution (40%)and the use of water as a reaction media to carry out organocatalytic processes remains a challenge, due to the fact that water can interfere with the formation of hydrogen bonds and polar interactions between the organocatalysts and substrates.<sup>6</sup> Only some privileged organocatalytic systems such as prolinamide and diaryl prolinol derivatives among others, have shown their efficiency as organocatalysts in water or aqueous media.<sup>4</sup> Most of these systems are highly hydrophobic molecules that diminished the contact with bulk water and the transition states, with actually the process taking place in a highly concentrated organic phase.<sup>8</sup> Recently, we have shown that prolinamides derived from 1,1'-binaphthyl-2,2'-diamine (binam)  $2^9$  and  $3^{10}$ and their supported related binam derivatives (4 and 5),<sup>11</sup> led to excellent results in the inter- and intramolecular aldol reactions under several reaction conditions, even using challenging aqueous electrophiles such as glyoxylic acid<sup>12</sup> dimethoxyacetaldehyde.<sup>13</sup> and 2,2-

Based on these previous results, we thought of interest the study of the efficiency of binam-prolinamide derivatives as organocatalysts in the reaction between methyl- and phenylglyoxal with ketones<sup>5a</sup> and with aldehydes.<sup>5b,c</sup>

First, the optimization of the reaction parameters in the reaction between acetone (**6a**) and methylglyoxal (**1a**, 40% aqueous solution) was studied. This reaction gave the Henze's ketol<sup>14</sup> (**7a**), which is involved in plants metabolism,<sup>15</sup> as a product.



Figure 1Binam-prolinamide derivatives as catalyst in the aldol reaction.

The efficiency of the two different binam-prolinamide derivatives 2 and 3(20 mol%) was evaluated using 10 equiv. of acetone as nucleophile (Table1). Better enantioselectivity was achieved with catalysts 3 than with catalysts 2 (Table 1, entries 1-4). While catalysts 2a and 3a afforded compound 7a, catalysts 2b and 3b gave its enantiomer (*ent*-7a) with lower enantioselectivity, showing that the configuration of the achieved aldol product was controlled by the chirality of the proline,<sup>16</sup> and that the match combination is  $(S_a)$ -binam and L-Pro. The results obtained with both catalysts were superior in terms of conversion, vields and enantioselectivities to the results achieved with L-proline (Table 1, compare entries 1-4 with entry 5), that gave product 7a as a racemic mixture. As the best enantioselectivity for this process was achieved with the  $(S_a)$ -binamsulfo-L-Pro derivative (3a) as catalyst, the rest of the reaction

parameters, were done using this catalyst (Table 1, entry 3).

The effect of the amount of nucleophile was evaluated. While decreasing the amount of acetone (6a) from 10 to 5 equiv. led to similar results, using only 2 equiv. of acetone provoked a decrease in the reaction rate and enantioselectivity (Table 1, compare entry 3 with 6 and 7). Changing the catalyst loading to 10 and 5 mol%, led to similar results in terms of conversion but with slight lower selectivity being found when only 5 mol% of 3a was used (Table 1, entries 8 and 9). Decreasing the temperature to 0 °C in the presence of 10 mol% of 3a and 5 equiv. of acetone, gave 88% ee but lower conversion and yield (Table 1, compare entries 3 and 10). Therefore, the effect of the addition of PhCO<sub>2</sub>H acid as co-catalyst was evaluated under these reaction conditions, but hardly any acceleration of the reaction was observed (Table 1, entry 12). Thus, under the best reaction conditions (10 mol% of 3a, 5 equiv. of 6a and 25°C), the time required for the reaction completion was reevaluated, finding that after 12h product 7a was achieved in almost quantitative yield and 90% ee (Table 1, entry 12). These results are better to those previously reported using simple L-prolinamide or the dipeptide L-Pro-L-Leu under neat conditions.<sup>5a</sup>Finally, the use of the supported binam-derivatives 4 and 5 as catalysts in the reaction between acetone and methylglyoxal was tested, but the reaction failed (Table 1, entries 13 and 14).

**Table 1** Optimization of reaction conditions between acetone (6a)and methylglyoxal  $(1)^a$ 

/	O − − − H − O 1a	+ ,	6a	T, a	Cat. dditives	7a	H V
Entry	Cat (mol%)	<b>6a</b> (equiv)	T (°C)	t (h)	Conv. <sup>b</sup>	Yield (%) <sup>c</sup>	ee (%) <sup>d</sup>
1	<b>2a (</b> 20)	10	25	24	100	-	80
2	<b>2b</b> (20)	10	25	24	100	-	-66
3	<b>3a</b> (20)	10	25	24	100	82	88
4	<b>3b</b> (20)	10	25	24	100	65	-68
5	L-Pro	10	25	72	80	66	0
	(20)						
6	<b>3a</b> (20)	5	25	24	100	-	87
7	<b>3a</b> (20)	2	25	24	95	-	80
8	<b>3a</b> (10)	5	25	24	100	-	87
9	<b>3a</b> (5)	5	25	24	100	-	80
10	<b>3a</b> (10)	5	0	24	90	75	88
11 <sup>e</sup>	<b>3a</b> (10)	5	0	24	93	84	88
12	<b>3a</b> (10)	5	25	12	100	94	90
13	4 (20)	5	25	72	-	-	-
14	5 (20)	5	25	168	-	-	-

<sup>*a*</sup> Reaction conditions: **1a** (0.25 mmol, 40% aq. solution), **6a** and 10 mol% of catalysts, otherwise stated.<sup>17b</sup>Conversion based on the unreacted aldehyde. <sup>*c*</sup> After purification by column chromatography. <sup>*d*</sup> Determined by chiral-phase HPLC.<sup>*e*</sup>5mol% of PhCO<sub>2</sub>H was added.

Once the best reaction conditions were established (Table 1, entry 12), the scope of the aldol reaction of

methylglyoxal (40% aqueous solution) with different ketones was studied (Scheme1 and Table 2).<sup>17</sup>



Scheme 1Aldol reaction of methylglyoxal with ketones.

<b>Table 2</b> Aldol reaction of methylglyoxal with ketones <sup>a</sup>								
Entry	Major product	Yield $\binom{9}{b}^{b}$	Dr <sup>c</sup>	$ee (\%)^d$				
1 <sup>e</sup>		92	-	90				
2		80	62:38	86				
3		79	22:78	86				
4		81	89:11	97				
5		72	86:14	82				
6 <sup>f</sup>		77	93:7	95				
7		70	40:60	34				
8		78	87:7:4:2	92				
9		67	68:29:2:1	87				
10		74	54:32:10:4	91				
11		42	63:37	90				

<sup>*a*</sup> Reaction conditions: Methylglyoxal (0.25 mmol, 40% aq. solution), ketone (5 equiv), catalyst **3a** (10 mol%) at 25 °C for 24 h, otherwise stated. <sup>*b*</sup> After purification by column chromatography. <sup>*c*</sup> Determined by the <sup>1</sup>H NMR of the crude product. <sup>*d*</sup> Determined by chiral-phase HPLC analysis for the major isomer. <sup>*e*</sup> Only 12h were required for reaction completion. <sup>*f*</sup> 30 h required for reaction completion.

In all cases, with the exception of cyclopentanone and 1,4-cyclohexadione (Table 2, entry 3 and 7, respectively), the major isomer achieved was the *anti* 

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isomer 7. The diastereoselectivities were rather moderate with the exception for cyclohexanone derivatives functionalized at the 4-position (products 7d-7f, entries 4-6). Only product 7g, achieved by reaction with 1,4-cyclohexadione, was obtained with low enantioselectivity (Table 2, entry 7), being the enantiomeric excesses for the rest of the examples higher that 82%. In the case of 4-substituted cyclohexanones, the major diastereoisomer formed was the expected *anti, anti-aldol*, being the diastereoselectivities highly dependent on the substituent at the 4-position (Table 1, entries 8-10). The relative configuration for compounds 7h-7j was done comparing the chemical shifts and coupling constants to those previously reported for related glyoxylic acid and ethyl glyoxylate derivatives,<sup>12</sup> and confirmed by NOESY experiments. The reaction with cyclobutanone led to the anti expected product 7k in good enantioselectivity but with low yield and diastereoselectivity (Table 2, entry 11). Attempts to extend the reaction to other ketones such as butanone ora-alkoxy ketones failed.

Once the scope of the reaction of aqueous methylglyoxal with ketones was accomplished, the cross aldol between methylglyoxal with enolizable aldehydes was studied under the same reaction conditions. This cross aldol reaction has previously reported,<sup>5b,c</sup> showing that been the corresponding  $\gamma$ -oxo- $\beta$ -hydroxy aldehydes isomerized easily during purification procedures. Therefore, these aldol products were in situ allowed to react with Ph<sub>3</sub>PCHCO<sub>2</sub>Et to give the corresponding Wittig adducts. Following this one-pot two step procedure, products 10 were obtained(Scheme 2, Table 3).<sup>18</sup> As before, in all cases the anti isomer was the major isomer, being the stereochemistry of the product assigned based in previously reported results.<sup>5b</sup> The reaction of methylglyoxal (1a) with propanal led to product 10a in moderate yield and diastereoselectivity but excellent enantioselectivity, comparable to the enantioselectivity achieved using diarylprolinol as catalyst (10 mol%) in THF<sup>5b</sup> (Table 3, entry 1). Better results were achieved in the reaction with octanal and heptanal and phenylpropanal, giving products 10b, 10c and 10d in excellent diastereo- and enantioselectivity, respectively (Table 3, entries 2-4). The reaction between  $phenylgly_{0}al^{19}$  (1b) and several aldehydes was also tested.<sup>20</sup> When propanal was used as nucleophile, moderate yield, diastereoand enantioselectivity was obtained (Table 3, entry 5). Meanwhile, phenylpropanal led to lower diastereoselectivity but better enantioselectivity (Table 3, entry 6).

**Table 3** Aldol reaction of glyoxals 1 with aldehydes followed by

 Wittig olefination<sup>a</sup>

Entry	Major product	Yield $(\%)^b$	Dr <sup>c</sup>	$ee (\%)^d$
1 <sup>e</sup>	QH ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓	40	76:24	92
2	10a OH UH UCH <sub>2</sub> γ,Me	57	98:2	95
3	10b OH (CH <sub>2</sub> ) <sub>6</sub> Me	65	99:1	96
4	10c OH Bn	48	99:1	97
5	10d OH Phy CO <sub>2</sub> Et	55	62:38	77
6 <sup><i>f</i></sup>	10e OH Ph O Bh CO <sub>2</sub> Et	48	55:45	86

<sup>*a*</sup> Reaction conditions: Methylglyoxal (0.25 mmol, 40% aq. solution)<sup>18</sup> or phenylglyoxal monohydrate (0.25 mmol),<sup>20</sup> aldehyde (2 equiv), catalyst **3a** (10 mol%) at 25 °C for 24 h, otherwise stated. <sup>*b*</sup> Overall yield after purification by column chromatography. <sup>*c*</sup> Determined by the <sup>1</sup>H NMR of the crude product. <sup>*d*</sup> Determined by chiral-phase HPLC analysis for the major isomer.

In conclusion, *N*-tosyl-( $S_a$ )-binam-L-prolinamide was an efficient catalysts to promote the aldol reaction between methylglyoxal under aqueous conditions or phenylglyoxal monohydrate with ketones or aldehydes, affording chiral  $\gamma$ -oxo- $\beta$ -hydroxy carbonyl compounds and  $\epsilon$ -oxo- $\delta$ -hydroxy  $\alpha$ , $\beta$ -unsaturated esters, respectively in good results in terms of yields, diastereo- and enantioselectivities.

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Scheme 2 Aldol reaction of glyoxals with aldehydes.

**Primary Data** for this article are available online at http://www.thieme-

connect.com/products/ejournals/journal/10.1055/s-

00000083 and can be cited using the following DOI: (number will be inserted prior to online publication).

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#### References

- (a) Klöpfer, A.; Spanneberg, R.; Glomb, M. A. J. Agric. Food Chem. 2011, 59, 394. (b) Pfeifer, Y. V.; Haase, P. T.; Kroh, L. W. J. Agric. Food Chem. 2013, 61, 3090.
- (2) (a) Fernández, M; Vicario, J. L.; Reyes, E.; Carrillo, L.; Badia, D. *Chem. Commun.* 2012, *48*, 2092. (b) Ren, L.; Lian.X.-L.; Gong, L.-Z. *Chem. Eur. J.* 2013, *19*, 3315. (c) Xu, Z.; De Moliner, F.; Cappelli, A. P.; Hulme, C. *Org. Lett.* 2013, *15*, 2738.
- (3) (a) Enantioselective Organocatalysis, Dalko P. I. Ed WILEY-VCH: Weinheim, 2007. (b) Enantioselective Organocatalyzed Reactions, Mahrwald, R. Ed. Springer: Heidelberg, 2011; Vols 1 and 2; (c) Science of Synthesis, List, B. Maruoka,K. Ed. Georg Thieme Verlag, Stuttgart 2011; Vols 1 and 2; (d) Comprehensive Enantioselective Organocatalysis: Catalysis, Reactions and Applications, Dalko, P. I. Ed. WILEY-VCH: Weinheim, 2013.
- (4) (a) Guillena, G.; Nájera C.; Ramón, D. J. Tetrahedron: Asymmetry 2007, 18, 2249. (b) Geary L. M.; Hultin, P. G. Tetrahedron: Asymmetry 2009, 20, 131. (c) Zlotin, S. G.; Kucherenko A. S.; Beletskaya, I. P. Russ. Chem. Rev. 2009, 78, 737. (d) Trost B.; Brindle, C. S. Chem. Soc. Rev. 2010, 39, 1600. (e) Heravi M. M.; Asadi, S. Tetrahedron: Asymmetry 2012, 23, 1431. (f)Guillena, G. in Modern Methods in Stereoselective Aldol Reactions, Mahrwald, R. Ed. Wiley-VCH: Weinheim, 2013, 155.
- (5) (a) Alberg, D. G.; Poulsen, T. B.; Bertelsen, S.; Christensen, K. L.; Birkler, R. D.; Johannsen, M.; Jørgensen, K. A. *Bioorg. Med. Chem. Lett.* 2009, 19, 3888. (b) Hayashi, Y.; Yasui, Y.; Kojima, M. Kawamura, T., Ishikawa, H. *Chem. Commun.* 2012, 48, 4570. (c) Hayashi, Y.; Kojima, M. *ChemCatChem*, 2013, 5, 2883.
- (6) Lidström, U. M. Chem. Rev. 2002, 102, 2751.
- (7) (a) Mase, N.; Barbas, C. F. III, Org. Biomol. Chem.
  2010, 8, 4043. (b) Toma, S.; Sebesta, R.; Meciarova, M. Curr. Org. Chem. 2011, 15, 2257. (c) Bhowmick, S.; Bhowmick, K. C. Tetrahedron: Asymmetry 2011, 22, 1945. (d) Chen, F.; Gong, P.; Gao, Y.; Zhang, H.; Zhou, A. Mini-Rev. Org. Chem. 2013, 10, 207. (e) Mlynarski, J.; Baś, S. Chem. Soc. Rev. 2014, 43, 577.
- (8) Gruttadauria, M.; Giacalone, F.; Noto, R. *Adv. Synth. Catal.* **2009**, *351*, 33 and references quoted therein.
- (9) (a) Guillena, G.; Hita, M. C.; Nájera, C. Tetrahedron: Asymmetry 2006, 17, 729. (b) Gryko, D.; Kowalczyk, B. Zawadzki, L. Synlett 2006, 1059. (c) Guizzetti, S.; Benaglia, M.; Pignataro, L.; Puglisi, A. Tetrahedron: Asymmetry 2006, 17, 2754.(d) Ma, G-N.; Zhang, Y.-P.; Shi, M. Synthesis 2007, 197.(e) Guizzetti, S.; Benaglia, M.; Raimondi, L.; Celentano, G. Org. Lett. 2007, 9, 1247. (f) Kucherenko, A. S.; Syutkin, D. E.; Zlotin, S. G. Russ. Chem. Bull. 2008, 57, 591.(g) Guillena, G.; Hita, M. C. Nájera, C.; Viózquez, S. F. Tetrahedron: Asymmetry 2007, 18, 2300. (h) Guillena, G.; Hita, M. C.; Nájera, C.; Viózquez, S.

F.J. Org. Chem. **2008**, 73, 5933. (i) Viózquez, S. F.; Bañón-Caballero, A.; Guillena, G.; Nájera, C.; Gómez–Bengoa, E.Org. Biomol. Chem. **2012**, 10, 4029.

- (10) (a) Guillena, G; Nájera, C.; Viózquez, S. F. Synlett
  2008, 3031. (b) Viózquez, S. F.; Guillena, G; Nájera, C.; Bradshaw, B.; Etxebarria-Jardí, G; Bonjoch, J. Org. Synth. 2011, 88, 317.
- (11) (a) Bañón-Caballero, A.; Guillena, G;Nájera, C. Green Chem. 2010, 12, 1599. (b) Bañón-Caballero, A.; Guillena, G; Nájera, C. Helv. Chim. Acta. 2012, 95, 1831. (c) Bañón-Caballero, A.; Guillena, G; Nájera, C.; Faggi, E.;Sebastián, R. M.; Vallribera, A. Tetrahedron 2013, 69, 1307. (d) Bañon-Caballero, A.; Guillena, G; Nájera, C. J. Org. Chem.2013, 79, 5349.
- (12) (a)Moles, F. J. N.; Guillena, G.; Nájera, C. *RSC Adv.*,
  2014,4, 9963. (b) Moles, F. J. N.;Guillena, G.; Nájera,
  C.;Gómez-Bengoa, E. *Synthesis*, 2014, *in press*.
- (13) Moles, F. J. N.; Bañón-Caballero, A.; Guillena, G.; Nájera, C. *Tetrahedron: Asymmetry* **2014**, *25*, 1323.
- (14) (a) Henze, M.; Müller, R. Z. Physiol. Chem. 1933, 214,281. (b) Schechter, M. S.; Green, N.; LaForge, F. B. J. Am. Chem. Soc. 1949, 71, 3165.
- (15) (a) Holmes, F. L.; *Hans Krebs- The formation of a Scientific Life 1900-1933*; Oxford University Press: Oxford 1991, 245. (b) Fang, J-M.; Wang, K.-C.; Cheng, Y.-S. J. Chin. Chem. Soc. 1991, 38, 297. (c) Li, Y.; Shi, Y.-P. Pharmazie 2007, 62, 714.
- (16) Stereochemistry assigned by comparison of the optical rotation values of the literature, in reference 5a.
- (17) To a mixture of the methylglyoxal (40% aqueous solution, 0.25 mmol, 0.038 mL) and catalyst (10 mol%) at the indicated temperature was added the corresponding ketone (1.25 mmol). The reaction was stirred until the methylglyoxal was consumed (monitored by TLC). The resulting residue was purified by chromatography (hexanes/AcOEt) to yield the pure aldol product. During purification aldols 7d-7f undergo a slight epimerisation.
- (18) To a mixture of the methylglyoxal (40% aqueous solution, 0.25 mmol, 0.038 mL) and catalyst (10 mol%) at the indicated temperature was added the corresponding aldehyde (0.5 mmol). The reaction was stirred until the methylglyoxal was consumed (monitored by TLC). Ph<sub>3</sub>PCHCO<sub>2</sub>Et (0.178 g, 0.5 mmol) was added and reaction mixture was stirred for 2 h. Upon completion, the reaction was quenched by passing through silica gel pad, and concentrated in vacuo. The resulting residue was purified by chromatography (hexanes/AcOEt) to yield the  $\alpha,\beta$ -unsaturated ester.
- (19) Arylglyoxals are important reagents for the synthesis of heterocyclic compounds. See, for instance: Eftekhari-Sis, B.; Zirak, M.; Akbari, A. *Chem. Rev.* 2013, *113*, 2953.
- (20)To a mixture of the phenylglyoxal monohydrate (0.25 mmol, 0.028 g) and catalyst (10 mol%) at the indicated temperature was added the corresponding aldehyde (0.5 mmol). The reaction was stirred until the phenylglyoxal was consumed (monitored by TLC). Ph<sub>3</sub>PCHCO<sub>2</sub>Et (0.178 g, 0.5 mmol) was added and reaction mixture was stirred for 2 h. Upon completion, the reaction was quenched by passing through silica gel pad, and concentrated in vacuo. The resulting residue was purified bv chromatography (hexanes/AcOEt) to yield the  $\alpha$ , $\beta$ unsaturated ester.

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#### Binam-prolinamides catalyzed the aqueous aldol reaction of glyoxals.



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**Supporting Information for:** 

Aqueous enantioselective aldol reaction of methyl- and phenylglyoxal organocatalyzed by N-Tosyl- $(S_a)$ -binam-L-prolinamide.

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**1.** General information: Catalysts **2** and **3** were prepared according to literature.<sup>1</sup> All the reagents were commercially available and used without further purification. <sup>1</sup>H NMR (300 MHz, 400 MHz) and <sup>13</sup>C NMR (75 MHz) spectra were obtained at 25 °C using CDCl<sub>3</sub> as solvent and chemical shifts are reported as  $\delta$  values relative to TMS as internal standard. IR spectra were obtained with Jasco 4100 LE (Pike Piracle ATR). High resolution mass spectra (HRMS-ESI) were obtained on a Waters LCT Premier XE apparatus equipped with a time of flight (TOF) analyzer and the samples were ionized by ESI techniques and introduced through an ultra-high pressure liquid chromatography (UPLC) model Waters ACQUITY H CLASS. Optical rotations were measured on a Jasco P-1030 Polarimeter with a 5 cm cell (c given in g/100 mL). HPLC analyses were performed on equipped with a chiral column and automatic injector, using mixtures of n-hexane/isopropyl alcohol (IPA) as mobile phase, at 25 °C. Analytical TLC was performed on silica gel plates and the spots were visualized using KMnO<sub>4</sub> solution as revelator. For flash chromatography we employed silica gel 60 (0.040-0.063 mm).

### 2. General procedures for the aldol reaction (

# 2.1 General procedure for the aldehyde-ketone aldol reaction using methylglyoxal 40% aqueous solution:

To a mixture of the methyl lyoxal 40% aqueous solution (0.25 mmol, 0.038 mL) and catalyst (10 mol%) at the indicated temperature was added the corresponding ketone (1.25 mmol). The reaction was stirred until the methylglyoxal was consumed (monitored by TLC). The resulting residue was purified by chromatography (hexanes/AcOEt) to yield the pure aldol product. During purification the aldols **7d**, **7e**and **7f** undergo an epimerisation and therefore the diastereoselectivities of the crude <sup>1</sup>H-RMN is different than the one showed in the <sup>1</sup>H NMR spectra.

# 2.2 General procedure for the aldehyde-aldehyde aldol reaction using methylglyoxal 40% aqueous solution:

To a mixture of the methylglyoxal 40% aqueous solution (0.25 mmol, 0.038 mL) and catalyst (10 mol%) at the indicated temperature was added the corresponding aldehyde (0.5 mmol). The reaction was stirred until the methylglyoxal was consumed (monitored by TLC). Wittig Reagent (0.178 g, 0.5 mmol) was added and reaction mixture was stirred for 2 h. Upon completion, the Witting reaction was quenched by passing through silica gel pad, and concentrated in vacuo. The resulting residue was purified by chromatography (hexanes/AcOEt) to yield the  $\Box,\Box$ -unsaturated ester.

# 2.3 General procedure for the aldehyde-aldehyde aldol reaction using phenylglyoxal monohidrate:

To a mixture of the phenylglyoxal monohidrate (0.25 mmol, 0.028 g) and catalyst (10 mol%) at the indicated temperature was added the corresponding aldehyde (0.5 mmol). The reaction was stirred until the phenylglyoxal was consumed (monitored by TLC). Wittig Reagent (0.178 g, 0.5 mmol) was added and reaction mixture was stirred for 2 h. Upon completion, the Witting reaction was quenched by passing through silica gel pad, and concentrated in vacuo. The resulting residue was purified by chromatography (hexanes/AcOEt) to yield the  $\Box$ ,  $\Box$  -unsaturated ester.

### 3. Spectra data of aldol products



(*R*)-3-hydroxyhexane-2,5-dione (7a).<sup>2</sup> Yellow oil. (0.029 g, 90%);  $[\alpha]^{26}_{D} = -17$  (*c*= 0.9; CHCl<sub>3</sub>);  $R_{\rm f}$ = 0.38 (Hex/EtOAc 1:1, revealed with KMnO<sub>4</sub>). IR: v 3423.0 (OH), 1721.6 (C=O), 1707.7 (C=O), 1357.6 (CH<sub>2</sub>-C=O). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  4.36 (dd, *J* = 6.4, 3.8 Hz, 1 H,

<u>CH</u>OH), 3.00 (dd, J = 17.3, 3.8 Hz, 1 H, <u>CH<sub>a</sub>H<sub>b</sub>-CHOH</u>), 2.87 (dd, J = 17.3, 6.4 Hz, 1 H, CH<sub>a</sub><u>H<sub>b</sub>-CHOH</u>), 2.28 (s, 3 H, CH<sub>3</sub>), 2.24 (s, 3 H, CH<sub>3</sub>). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  209.2 (C), 207.1 (C), 73.8 (CH), 46.1 (CH<sub>2</sub>), 30.8 (CH<sub>3</sub>), 25.4 (CH<sub>3</sub>). MS (IE) m/z (%) for C<sub>6</sub>H<sub>10</sub>O<sub>3</sub>: M<sup>+</sup> = 130 (2), 112 (3), 97 (6), 87 (100), 70 (10), 55 (11).



(*S*)-2-((*R*)-1-hydroxy-2-oxopropyl)cyclohexanone (7b). Data for the isomer (2*S*,1'*R*). Yellow oil. (0.034 g, 80%);  $[\alpha]^{26}_{D} = -29$  (*c*= 0.5; CHCl<sub>3</sub>); *R*<sub>f</sub>= 0.43 (Hex/EtOAc 7:3, revealed with KMnO<sub>4</sub>). IR: v 3460.6 (OH), 1733.69 (C=O), 1703.8 (C=O), 1421.3 (CH<sub>3</sub>-C=O). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  3.87 (dd, *J* = 7.9, 3.0 Hz, 1 H, <u>CH</u>OH), 3.56 (d, *J* = 7.9 Hz, 1 H, OH), 3.15 - 3.03 (m, 1 H, H<sub>cyclo</sub>), 2.51 - 2.32 (m, 2 H, H<sub>cyclo</sub>), 2.30 (s, 3 H, CH<sub>3</sub>), 2.20 - 2.06 (m, 2 H, H<sub>cyclo</sub>), 2.06 - 1.65 (m, 4 H, H<sub>cyclo</sub>). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  212.3 (C), 210.0 (C), 77.9 (CH), 53.7 (CH), 42.0 (CH<sub>2</sub>), 30.3 (CH<sub>2</sub>), 26.9 (CH<sub>2</sub>), 25.7 (CH<sub>3</sub>), 24.8 (CH<sub>2</sub>). HRMS calculated for C<sub>9</sub>H<sub>14</sub>O<sub>3</sub>: 170.0943 found: 171.1020 (M<sup>+</sup> + H, calculated 171.1021).



(*S*)-2-((*S*)-1-hydroxy-2-oxopropyl)cyclopentanone (7c).<sup>3</sup> As a diastereoisomer mixture (22:78, *anti:syn*). Yellow oil. (0.031 g, 79%);  $[\alpha]^{26}_{D} = -35$  (*c*= 2 ; CHCl<sub>3</sub>); *R*<sub>f</sub>= 0.54 (Hex/EtOAc 1:1, revealed with KMnO<sub>4</sub>). IR: v 3455.8 (OH), 1733.7 (C=O), 1710.5 (C=O), 1402.9 (CH<sub>3</sub>-C=O). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  4.71 (dd, *J* = 4.8, 2.3 Hz, 1 H, *syn*), 4.12 (dd, *J* = 3.6, 3.4 Hz, 1 H, *anti*), 3.87 (d, *J* = 3.6 Hz, 1 H, *anti*), 3.54 (d, *J* = 4.8 Hz, 1 H, *syn*), 2.81 - 2.71 (m, 1 H, H<sub>cyclo</sub>, *anti*), 2.58 - 2.48 (m, 1 H, H<sub>cyclo</sub>, *syn*), 2.27 (s, 3 H, CH<sub>3</sub>, *anti*), 2.22 (s, 3 H, CH<sub>3</sub>, *syn*), 2.20 - 1.98 (m, 6 H, H<sub>cyclo</sub>), 1.90 - 1.67 (m, 6 H, H<sub>cyclo</sub>). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  218.5 (C), 217.8 (C), 208.7 (C), 207.9 (C), 76.3 (CH), 75.5 (CH), 51.4 (CH), 50.6 (CH), 38.7 (CH<sub>2</sub>), 38.3 (CH<sub>2</sub>), 26.3 (CH<sub>2</sub>), 25.3 (CH<sub>3</sub>), 25.1 (CH<sub>3</sub>), 21.7 (CH<sub>2</sub>), 21.0 (CH<sub>2</sub>), 20.6 (CH<sub>2</sub>). MS (IE) m/z (%) for C<sub>8</sub>H<sub>12</sub>O<sub>4</sub>: M<sup>+</sup> = 156 (2), 138 (3), 113 (100), 96 (30), 85 (40), 67 (84), 57 (28).

(*S*)-3-((*R*)-1-hydroxy-2-oxopropyl)dihydro-2*H*-pyran-4(3*H*)-one (7d). As a diastereoisomer mixture (89:11, *anti:syn*). Yellow oil. (0.035 g, 81%);  $[\alpha]^{26}_{D} = -38$  (*c*= 1.5 ; CHCl<sub>3</sub>); *R*<sub>f</sub>= 0.26 (Hex/EtOAc; 1:1, revealed with KMnO<sub>4</sub>). IR: v 3449.1 (OH), 1769.6 (C=O), 1712.5 (C=O), 1373.1 (CH<sub>3</sub>-C=O). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, diastereomer mixture (1:1)):  $\delta$  4.70 (dd, *J* = 5.4, 3.1 Hz, 1 H, CHOH, *syn*), 4.35 (dd, *J* = 11.3, 6.8 Hz, 1 H, H<sub>cyclo</sub>), 4.29 - 4.18 (m, 2 H, H<sub>cyclo</sub>), 4.03 (dd, *J* = 11.3, 6.7 Hz, 1 H, H<sub>cyclo</sub>), 3.90 - 3.65 (m, 6 H, H<sub>cyclo</sub>), 3.43 (d, *J* = 5.5 Hz, 1 H, OH), 3.26 (ddd, *J* = 10.9, 6.8, 2.9 Hz, 1 H, H<sub>cyclo</sub>), 3.02 (ddd, *J* = 10.7, 6.7, 3.1 Hz, 1 H, H<sub>cyclo</sub>), 2.75 - 2.56 (m, 2 H, H<sub>cyclo</sub>), 2.52 - 2.34 (m, 2 H, H<sub>cyclo</sub>), 2.26 (s, 3 H, CH<sub>3</sub>), 2.25 (s, 3 H, CH<sub>3</sub>). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  209.3 (C), 207.8 (C), 206.4 (C), 206.1 (C), 74.6 (CH), 73.7 (CH), 70.0 (CH<sub>2</sub>), 67.7 (CH<sub>2</sub>), 67.6 (CH<sub>2</sub>), 67.5 (CH<sub>2</sub>), 53.6 (CH), 53.2 (CH), 42.3 (CH<sub>2</sub>), 42.2 (CH<sub>2</sub>), 26.1 (CH<sub>3</sub>), 25.2 (CH<sub>3</sub>). HRMS calculated for C<sub>8</sub>H<sub>12</sub>O<sub>4</sub>: 172.0736 found: 173.0814 (M<sup>+</sup> + H, calculated 173.0814).



(*S*)-3-((*R*)-1-hydroxy-2-oxopropyl)dihydro-2*H*-thiopyran-4(3*H*)-one (7e). Colorless oil. Data for the isomer (2*S*,1'*R*). (0.034 g, 72%);  $[\alpha]^{26}{}_{D} = -14$  (*c*= 1 ; CHCl<sub>3</sub>); *R*<sub>f</sub>= 0.33 (Hex/EtOAc 1:1, revealed with KMnO<sub>4</sub>); IR: v 3413.4 (OH), 1715.5 (C=O), 1704.8 (C=O), 1419.3 (CH<sub>3</sub>-C=O). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 3.93 (dd, *J* = 7.3, 2.8 Hz, 1 H, <u>CH</u>OH), 3.60 (d, *J* = 7.4 Hz, 1 H, OH), 3.46 - 3.36 (m, 1 H, H<sub>cyclo</sub>), 3.29 - 2.75 (m, 6 H, H<sub>cyclo</sub>), 2.29 (s, 3 H, CH<sub>3</sub>). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 208.9 (C), 208.2 (C), 77.8 (CH), 55.8 (CH), 44.2 (CH<sub>2</sub>), 32.5 (CH<sub>2</sub>), 29.5 (CH<sub>2</sub>), 25.4 (CH<sub>3</sub>). HRMS calculated for C<sub>8</sub>H<sub>12</sub>O<sub>3</sub>S: 188.0507 found: 189.0583 (M<sup>+</sup> + H, calculated 189.0585).



(S)- *tert*-butyl 3-((*R*)-1-hydroxy-2-oxopropyl)-4-oxopiperidine-1-carboxylate (7f). As a diastereomer mixture (93:7, *anti:syn*). Colorless oil. (0.064 g, 95%);  $[\alpha]_{D}^{26} = -34$  (*c*= 0.7 ; CHCl<sub>3</sub>);

*R*<sub>f</sub>= 0.55 (Hex/EtOAc; 1:1, revealed with KMnO<sub>4</sub>). IR: v 3390.2 (OH), 1791.6 (C=O), 1691.3 (C=O), 1419.3 (CH<sub>3</sub>-C=O). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, diastereoisomer mixture 1:1):  $\delta$  4.70 (dd, *J* = 4.9, 2.9 Hz, 1 H, <u>CH</u>OH, *syn*), 3.93 (dd, *J* = 6.2, 2.7 Hz, 1 H, <u>CH</u>OH, *anti*), 3.75 - 3.62 (m, 1 H, H<sub>cyclo</sub>), 3.49 - 3.09 (m, 6 H, H<sub>cyclo</sub>), 2.87 (ddd, *J* = 10.6, 6.2, 2.9 Hz, 1 H, H<sub>cyclo</sub>), 2.63 - 2.38 (m, 6 H, H<sub>cyclo</sub>), 2.30 (s, 3 H, CH<sub>3</sub>, *anti*), 2.29 (s, 3 H, CH<sub>3</sub>, *syn*), 1.52 (s, 9 H, C(CH<sub>3</sub>)<sub>3</sub>, *anti*), 1.50 (s, 9 H, C(CH<sub>3</sub>)<sub>3</sub>, *syn*). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  209.0 (C), 207.7 (C), 207.5 (C), 207.0 (C), 154.6 (C), 154.5 (C), 80.8 (2xC), 75.6 (CH), 74.6 (C), 52.4 (CH), 52.0 (CH), 45.3 (2xCH<sub>2</sub>), 42.9 (CH<sub>2</sub>), 42.7 (CH<sub>2</sub>), 40.8 (2xCH<sub>2</sub>), 28.3 (6xCH<sub>3</sub>), 25.9 (CH<sub>3</sub>), 25.2 (CH<sub>3</sub>). HRMS calculated for C<sub>13</sub>H<sub>21</sub>NO<sub>5</sub>: 271.1421 found: 294.1322 (M<sup>+</sup> + H, calculated 294.1317).



(*S*)-2-((*S*)-1-hydroxy-2-oxopropyl)cyclohexane-1,4-dione (7g). As a diastereoisomer mixture (40:60, *anti:syn*). Yellow oil. (0.032 g, 70%);  $[α]^{26}{}_{D} = -20$  (*c*= 0.6 ; CHCl<sub>3</sub>);  $R_{f}$ = 0.18 (Hex/EtOAc; 1:1, revealed with KMnO<sub>4</sub>). IR: v 3427.8 (OH), 1720.9 (C=O), 1703.8 (C=O), 1310.4 (CH<sub>3</sub>-C=O). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 4.84 (dd, *J* = 4.8, 2.0 Hz, 1 H, <u>CH</u>OH, *syn*), 3.98 (dd, *J* = 4.3, 2.5 Hz, 1 H, <u>CH</u>OH, *anti*), 3.82 (d, *J* = 4.3 Hz, 1 H, OH, *anti*), 3.60 (d, *J* = 4.8 Hz, 1 H, OH, *syn*), 3.36 (ddd, *J* = 10.2, 6.4, 2.5 Hz, 1 H, H<sub>cyclo</sub>), 3.16 - 3.03 (m, 2 H, H<sub>cyclo</sub>), 2.91 - 2.61 (m, 11 H, H<sub>cyclo</sub>), 2.35 (s, 3 H, CH<sub>3</sub>, *anti*), 2.27 (s, 3 H, *syn*). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 208.1 (C), 207.9 (2xC), 207.3 (C), 207.0 (C), 206.5 (C), 77.7 (CH), 77.2 (CH), 48.9 (CH), 48.1 (CH), 41.3 (CH<sub>2</sub>), 37.2 (CH<sub>2</sub>), 37.1 (CH<sub>2</sub>), 36.7 (CH<sub>2</sub>), 35.9 (CH<sub>2</sub>), 35.8 (CH<sub>2</sub>), 25.4 (CH<sub>3</sub>), 25.1 (CH<sub>3</sub>). HRMS calculated for C<sub>9</sub>H<sub>12</sub>O<sub>4</sub>: 184.0736 found: 185.0816 (M<sup>+</sup> + H, calculated 185.0814).



(2*S*,4*S*)-2-((*R*)-1-hydroxy-2-oxopropyl)-4-methylcyclohexanone (7h). As a diastereoisomer mixture (87:7:4:2). Colorless oil. (0029 g, 78%);  $[\alpha]^{26}_{D} = -70$  (*c*= 3.2 ; CHCl<sub>3</sub>); *R*<sub>f</sub>= 0.64 (Hex/EtOAc; 1:1, revealed with KMnO<sub>4</sub>). IR: v 3450.0 (OH), 1736.5 (C=O), 1708.6 (C=O), 1239.0 (CH<sub>3</sub>-C=O). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  4.66 (dd, *J* = 4.8, 2.3 Hz, 1 H, <u>CH</u>OH, *syn*), 3.87 (dd, *J* 

= 7.2, 3.1 Hz, 1 H, <u>CH</u>OH, *anti*), 3.58 (d, J = 7.2 Hz, 1 H, OH, *anti*), 3.27 (d, J = 4.8 Hz, 1 H, *syn*), 3.22 - 3.12 (m, 1 H, H<sub>cyclo</sub>), 2.92 - 2.82 (m, 1 H, H<sub>cyclo</sub>), 2.54 - 2.36 (m, 2 H, H<sub>cyclo</sub>), 2.33 - 2.08 (m, 10 H, H<sub>cyclo</sub>), 2.05 - 1.63 (m, 8 H, H<sub>cyclo</sub>), 1.18 (d, J = 7.0 Hz, 3 H, CH<u>CH<sub>3</sub></u>, *anti*), 1.00 (d, J = 6.4 Hz, 3 H, CH<u>CH<sub>3</sub></u>, *syn*). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  212.6 (C), 211.6 (C), 210.4 (C), 209.6 (C), 78.5 (CH), 75.3 (CH), 52.1 (CH), 49.1 (CH), 41.2 (CH<sub>2</sub>), 38.1 (CH<sub>2</sub>), 36.1 (CH<sub>2</sub>), 34.7 (CH<sub>2</sub>), 34.3 (CH<sub>2</sub>), 32.2 (CH<sub>2</sub>), 31.3 (CH), 26.8 (CH), 26.4 (CH<sub>3</sub>), 25.7 (CH<sub>3</sub>), 21.3 (CH<sub>3</sub>), 18.5 (CH<sub>3</sub>). HRMS calculated for C<sub>10</sub>H<sub>16</sub>O<sub>3</sub>: 184.1099 found: 185.1184 (M<sup>+</sup> + H, calculated 185.1178).



(2*S*,4*S*)-2-((*R*)-1-hydroxy-2-oxopropyl)-4-phenylcyclohexanone (7i). As a diastereoisomer mixture (68:29:2:1). Colorless oil. (0041 g, 67%);  $[α]^{26}_{D} = -19$  (*c*= 2.3 ; CHCl<sub>3</sub>); *R*<sub>f</sub>= 0.61 (Hex/EtOAc; 1:1, revealed with KMnO<sub>4</sub>). IR: v 3439.4 (OH), 1710.1 (C=O), 1704.7 (C=O), 1102.1 (CH<sub>3</sub>-C=O). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 7.41 - 7.21 (m, 5 H, ArH), 4.75 (dd, *J* = 4.8, 2.3 Hz, 1 H, <u>CH</u>OH, *syn*), 4.08 (dd, *J* = 5.8, 3.1 Hz, 1 H, <u>CH</u>OH, *anti*), 3.75 (d, *J* = 5.8 Hz, 1 H, OH, *anti*), 3.49 - 3.40 (m, 1 H, H<sub>cyclo</sub>), 3.33 (d, *J* = 4.9 Hz, 1H, OH, *syn*), 3.21 - 3.03 (m, 2 H, H<sub>cyclo</sub>), 2.67 - 2.38 (m, 5 H, H<sub>cyclo</sub>), 2.36 - 1.79 (m, 14 H, H<sub>cyclo</sub>). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 211.8 (C), 210.3 (C), 210.2 (C), 208.5 (C), 144.2 (C), 143.6 (C), 128.8 (CH), 128.7 (2xCH), 126.8 (CH), 126.7 (3xCH), 126.5 (C), 79.2 (CH), 75.4 (CH), 52.5 (CH), 49.9 (CH), 42.6 (CH), 41.5 (CH<sub>2</sub>), 39.9 (CH<sub>2</sub>), 37.2 (CH), 35.4 (CH<sub>2</sub>), 33.9 (CH<sub>2</sub>), 33.5 (CH<sub>2</sub>), 30.6 (CH<sub>2</sub>), 26.5 (CH<sub>3</sub>), 25.6 (CH<sub>3</sub>). HRMS calculated for C<sub>15</sub>H<sub>18</sub>O<sub>3</sub>: 246.1256 found: 247.1342 (M<sup>+</sup> + H, calculated 247.1334).



(2*S*,4*S*)-4-(*tert*-butyl)-2-((**R**)-1-hydroxy-2-oxopropyl)cyclohexanone (7j). As a diastereoisomer mixture (54:32:10:4). Colorless oil. (0041 g, 74%);  $[\alpha]^{26}{}_{D} = -87$  (*c*= 3.6 ; CHCl<sub>3</sub>); *R*<sub>f</sub>= 0.69 (Hex/EtOAc; 1:1, revealed with KMnO<sub>4</sub>). IR: v 3453.9 (OH), 1745.2 (C=O), 1708.6 (C=O), 1240.0 (CH<sub>3</sub>-C=O). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  4.68 (dd, *J* = 4.9, 2.4 Hz, 1 H, <u>CH</u>OH, *syn*), 4.03 (dd, *J* 

= 6.3, 3.4 Hz, 1 H, <u>CH</u>OH, *anti*), 3.67 (d, *J* = 6.3 Hz, 1 H, OH, *anti*), 3.31 (d, *J* = 4.9 Hz, 1 H, OH, *syn*), 3.06 (td, *J* = 7.9, 3.4 Hz, 1 H, H<sub>cyclo</sub>), 2.51 - 2.38 (m, 2 H, H<sub>cyclo</sub>), 2.30 (s, 3 H, CH<sub>3</sub>, *anti*), 2.24 (s, 3 H, CH<sub>3</sub>, *syn*), 2.13 - 1.47 (m, 11 H, H<sub>cyclo</sub>), 0.93 (s, 9 H, C(CH<sub>3</sub>)<sub>3</sub>, *anti*), 0.90 (s, 9 H, C(CH<sub>3</sub>)<sub>3</sub>, *syn*). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): □ 214.1 (C), 211.4 (C), 210.3 (C), 209.0 (C), 78.7 (CH), 75.6 (CH), 52.4 (CH), 49.7 (CH), 46.4 (CH), 42.6 (CH), 41.3 (CH<sub>2</sub>), 40.1 (CH<sub>2</sub>), 33.0 (C), 32.6 (C), 27.5 (3xCH<sub>3</sub>), 27.4 (CH<sub>2</sub>), 27.3 (CH<sub>3</sub>), 27.0 (3xCH<sub>3</sub>), 26.4 (CH<sub>3</sub>), 25.7 (CH<sub>3</sub>), 23.5 (2xCH<sub>2</sub>). HRMS calculated for C<sub>13</sub>H<sub>22</sub>O<sub>3</sub>: 226.1569 found: 227.1638 (M<sup>+</sup> + H, calculated 227.1647).

(*S*)-2-((*R*)-1-hydroxy-2-oxopropyl)cyclobutanone (7k).<sup>4</sup> As a diastereoisomer mixture (63:37, *anti:syn*). Colorless oil. (0.015 g, 42%);  $[\alpha]^{26}_{D} = -20$  (*c*= 1.2 ; CHCl<sub>3</sub>); *R*<sub>f</sub>= 0.46 (Hex/EtOAc 1:1, revealed with KMnO<sub>4</sub>); IR: v 3445.2 (OH), 1776.1 (C=O), 1711.5 (C=O), 1417.4 (CH<sub>3</sub>-C=O). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, diastereoisomer mixture 1:1): δ 4.62 (dd, *J* = 3.6, 3.2 Hz, 1 H, <u>CH</u>OH, *syn*), 4.21 (dd, *J* = 3.8, 3.4 Hz, 1 H, <u>CH</u>OH, *anti*), 3.88 - 3.62 (m, 4 H, H<sub>cyclo</sub>), 3.15 2.94 (m, 4 H, H<sub>cyclo</sub>), 2.29 (s, 3 H, CH<sub>3</sub>, *anti*), 2.22 (s, 3 H, CH<sub>3</sub>, *syn*), 1.97 (dd, *J* = 16.9, 8.5 Hz, 2 H, H<sub>cyclo</sub>). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 208.1 (C), 207.5 (C), 207.1 (C), 206.5 (C), 75.2 (CH), 74.1 (CH), 61.3 (2xCH), 46.7 (CH<sub>2</sub>), 46.1 (CH<sub>2</sub>), 25.4 (CH<sub>3</sub>), 25.0 (CH<sub>3</sub>), 13.6 (CH<sub>2</sub>), 10.6 (CH<sub>2</sub>). MS (IE) m/z (%) for C<sub>7</sub>H<sub>10</sub>O<sub>3</sub>: M<sup>+</sup> = 142 (2), 124 (10), 99 (34), 86 (24), 71 (80), 57 (100).



(4*R*,5*R*,*E*)-ethyl 5-hydroxy-4-methyl-6-oxohept-2enoato (10a).<sup>5</sup> As a diastereoisomer mixture (76:24, *anti:syn*). Colorless oil. (0.020 g, 40%);  $[α]^{26}_{D} = -15$  (*c*= 0.6 ; CHCl<sub>3</sub>); *R*<sub>f</sub>= 0.20 (Hex/EtOAc; 85:15, revealed with KMnO<sub>4</sub>). IR: v 3326.6 (OH), 1715.4 (C=O), 1665.2 (C=O), 1226.5 (CH<sub>3</sub>-C=O). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 7.06 (dd, *J* = 15.8, 7.1 Hz, 1 H, <u>CH</u>=CH, *syn*), 6.80 (dd, *J* = 15.8, 8.2 Hz, 1 H, <u>CH</u>=CH, *anti*), 5.93 (dd, *J* = 15.8, 1.4 Hz, 1 H, CH=<u>CH</u>, *syn*), 5.84 (dd, *J* = 15.8, 1.2 Hz, 1 H, CH=<u>CH</u>, *syn*), 4.27 (dd, *J* = 4.6, 2.8 Hz, 1 H, <u>CH</u>OH, *syn*), 4.24 - 4.12 (m, 5 H), 3.58 (d, *J* = 4.7 Hz, 1 H, OH, *anti*), 3.52 (d, *J* = 4.9 Hz, 1 H, OH, *syn*), 2.98 - 2.79 (m, 2 H), 2.25 (s, 3 H, CH<sub>3</sub>, *syn*), 2.20 (s, 3 H, CH<sub>3</sub>, *anti*), 1.32 - 1.24 (m, 9 H), 0.95 (d, *J* = 6.9 Hz, 3 H, CH<u>CH<sub>3</sub>, *syn*).<sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 208.2 (C), 208.0 (C), 166.3 (C), 165.9 (C), 149.4 (CH),</u>

146.6 (CH), 122.8 (CH), 121.9 (CH), 80.1 (CH), 79.0 (CH), 60.5 (CH<sub>2</sub>), 60.4 (CH<sub>2</sub>), 39.8 (CH), 39.2 (CH), 25.8 (CH<sub>3</sub>), 25.6 (CH<sub>3</sub>), 16.7 (CH<sub>3</sub>), 14.2 (2xCH<sub>3</sub>), 12.0 (CH<sub>3</sub>). MS (IE) m/z (%) for  $C_{10}H_{16}O_4$ : M<sup>+</sup> = 200 (2), 182 (3), 157 (40), 128 (38), 111 (100), 100 (35), 83 (28), 55 (42).



(*R*,*E*)-ethyl 4-((*R*)-1-hydroxy-2-oxopropyl)undec-2-enoate (10b). As a diastereoisomer mixture (99:1, *anti:syn*). Colorless oil. (0.046 g, 65%);  $[\alpha]^{26}{}_{D} = -56$  (*c*= 1.2 ; CHCl<sub>3</sub>); *R*<sub>f</sub>= 0.23 (Hex/EtOAc; 85:15, revealed with KMnO<sub>4</sub>). IR: v 3463.5 (OH), 1715.4 (C=O), 1653.7 (C=O), 1231.3 (CH<sub>3</sub>-C=O). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  6.72 (dd, *J* = 15.8, 9.7 Hz, 1 H, <u>CH</u>CH), 5.81 (dd, *J* = 15.8, 0.7 Hz, 1 H, CH<u>CH</u>), 4.27 (dd, *J* = 4.5, 2.4 Hz, 1 H, <u>CH</u>OH), 4.18 (q, *J* = 7.1 Hz, 2 H, O<u>CH<sub>2</sub></u>CH<sub>3</sub>), 3.55 (d, *J* = 4.5 Hz, 1 H, OH), 2.71 - 2.57 (m, 1 H), 2.20 (s, 3 H, CH<sub>3</sub>), 1.79 - 1.62 (m, 2 H), 1.42 - 1.18 (m, 13 H), 0.90 (t, *J* = 6.7 Hz, 3 H, CH<sub>3</sub>). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  208.0 (C), 165.7 (C), 145.7 (CH), 123.5 (C), 79.0 (CH), 60.5 (CH<sub>2</sub>), 45.6 (CH), 31.8 (CH<sub>2</sub>), 31.5 (CH<sub>2</sub>), 29.4 (CH<sub>2</sub>), 29.2 (CH<sub>2</sub>), 27.3 (CH<sub>2</sub>), 25.4 (CH<sub>3</sub>), 22.6 (CH<sub>2</sub>), 14.2 (CH<sub>3</sub>), 14.1 (CH<sub>3</sub>). HRMS calculated for C<sub>16</sub>H<sub>28</sub>O<sub>4</sub>: 284.1988 found: 285.2072 (M<sup>+</sup> + H, calculated 285.2066).



(*R*,*E*)-ethyl 4-((*R*)-1-hydroxy-2-oxopropyl)dodec-2-enoate (10c). As a diastereoisomer mixture (98:2, *anti:syn*). Colorless oil. (0.042 g, 57%);  $[\alpha]^{26}_{D} = -50$  (*c*= 1.4 ; CHCl<sub>3</sub>); *R*<sub>f</sub>= 0.22 (Hex/EtOAc; 85:15, revealed with KMnO<sub>4</sub>). IR: v 3463.5 (OH), 1713.4 (C=O), 1642.0 (C=O), 1367.3 (CH<sub>3</sub>-C=O). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 6.72 (dd, *J* = 15.8, 9.7 Hz, 1 H, <u>CH</u>CH), 5.81 (dd, *J* = 15.8, 0.6 Hz, 1 H, CH<u>CH</u>), 4.27 (d, *J* = 2.4 Hz, 1 H, <u>CH</u>OH), 4.18 (q, *J* = 7.1 Hz, 2 H, O<u>CH<sub>2</sub>CH<sub>3</sub>), 2.71 - 2.57 (m, 1 H), 2.20 (s, 3 H, CH<sub>3</sub>), 1.78 - 1.56 (m, 2 H), 1.41 - 1.19 (m, 15 H), 0.90 (t, *J* = 6.7 Hz, 3 H, CH<sub>3</sub>). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>, diastereoisomer mixture 1:1): δ 208.5 (C), 208.1 (C), 166.2 (C), 165.8 (C), 148.5 (CH), 145.8 (CH), 79.6 (CH), 79.0 (CH), 60.5 (2xCH<sub>2</sub>), 45.6 (CH<sub>3</sub>), 45.5 (CH<sub>3</sub>), 33.9 (CH<sub>2</sub>), 31.8 (2xCH<sub>2</sub>), 31.4 (CH<sub>2</sub>), 29.5 (CH<sub>2</sub>), 29.4 (2xCH<sub>2</sub>), 29.2 (2xCH<sub>2</sub>), 29.1 (CH<sub>2</sub>), 27.4 (CH<sub>2</sub>), 27.3 (CH<sub>2</sub>), 25.9 (CH<sub>3</sub>), 25.4 (CH<sub>3</sub>), 24.7 (CH<sub>2</sub>), 22.6 (CH<sub>2</sub>), 14.2 (CH<sub>3</sub>),</u>

14.1 (CH<sub>3</sub>). HRMS calculated for  $C_{17}H_{30}O_4$ : 298.2144 found: 299.2243 (M<sup>+</sup> + H, calculated 299.2222).



(4*R*,5*R*,*E*)-ethyl 4-benzyl-5-hidroxy-6-oxohept-2-enoate (10d).<sup>5</sup> As a diastereoisomer mixture (99:1, *anti:syn*). Colorless oil. (0.033 g, 48%);  $[\alpha]^{26}_{D} = -42$  (*c*= 1.5 ; CHCl<sub>3</sub>); *R*<sub>f</sub>= 0.15 (Hex/EtOAc; 80:20, revealed with KMnO<sub>4</sub>). IR: v 3414.3 (OH), 1712.9 (C=O), 1709.6 (C=O), 1132.1 (CH<sub>3</sub>-C=O). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 7.38 -7.22 (m, 5 H, ArH), 6.81 (dd, *J* = 15.8, 9.0 Hz, 1 H, <u>CH</u>=CH), 5.81 (d, *J* = 15.8 Hz, 1 H, CH=<u>CH</u>), 4.29 - 4.14 (m, 2 H), 4.11 (dd, *J* = 4.5, 2.0 Hz, 1 H, <u>CH</u>OH), 3.60 (d, *J* = 4.4 Hz, 1 H, OH), 3.14 - 2.68 (m, 3 H), 2.11 (s, 3 H, CH<sub>3</sub>), 1.29 (t, *J* = 7.1 Hz, 3 H, CH<sub>3</sub>). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>, diastereoisomer mixture (1:1)): δ 208.2 (2xC), 166.0 (C), 165.6 (C), 147.6 (CH), 144.8 (CH), 138.4 (2xC), 138.2 (2xC), 129.3 (CH), 128.7 (CH), 128.5 (2xCH), 128.3 (2xCH), 126.7 (2xCH), 126.4 (2xCH), 123.8 (CH), 123.1 (CH), 79.0 (CH), 77.2 (CH), 60.5 (2xCH<sub>2</sub>), 47.3 (2xCH), 34.3 (CH<sub>2</sub>), 30.6 (CH<sub>2</sub>), 25.7 (CH<sub>3</sub>), 25.2 (CH<sub>3</sub>), 14.2 (CH<sub>3</sub>), 14.1 (CH<sub>3</sub>). MS (IE) m/z (%) for C<sub>16</sub>H<sub>20</sub>O<sub>4</sub>: M<sup>+</sup> = 276 (2), 258 (8), 233 (16), 203 (28), 187 (27), 129 (37), 91 (100).



(4*R*,5*R*,*E*)-ethyl 5-hydroxy-4-methyl-6-oxo-6-phenylhex-2-enoate (10e). As a diastereoisomer mixture (61:39, *anti:syn*). Colorless oil. (0.035 g, 53%);  $[\alpha]^{26}_{D} = -12$  (*c*= 1.2 ; CHCl<sub>3</sub>);  $R_{\rm f}$ = 0.30 (Hex/EtOAc; 70:30, revealed with KMnO<sub>4</sub>). IR: v 3439.4 (OH), 1708.6 (C=O), 11692.2 (C=O), 1269.9 (CH<sub>3</sub>-C=O). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  7.98 - 7.88 (m, 2 H, ArH), 7.73 - 7.50 (m, 3 H, ArH), 7.15 (dd, *J* = 15.7, 7.4 Hz, 1 H, <u>CH</u>=CH), 5.94 (dd, *J* = 15.7, 1.4 Hz, 1 H, CH=<u>CH</u>), 5.20 (dd, *J* = 6.4, 2.4 Hz, 1 H, <u>CH</u>OH), 4.22 (q, *J* = 7.1 Hz, 2 H, O<u>CH<sub>2</sub>CH<sub>3</sub>), 3.78 (d, *J* = 6.4 Hz, 1 H, OH), 2.91 - 2.78 (m, 1 H), 1.32 (t, *J* = 7.1 Hz, 3 H, OCH<sub>2</sub><u>CH3</u>), 0.87 (d, *J* = 6.8 Hz, 3 H, CH<u>CH<sub>3</sub></u>).<sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  200.6 (C), 166.3 (C), 149.8 (CH), 134.3 (CH), 133.5 (C), 129.1 (2xCH), 128.5 (2xCH), 121.7 (CH), 75.0 (CH), 60.4 (CH<sub>2</sub>), 40.7 (CH), 14.3 (CH<sub>3</sub>), 11.5 (CH<sub>3</sub>). HRMS calculated for C<sub>15</sub>H<sub>18</sub>O<sub>4</sub>: 262.1205 found: 285.1111 (M<sup>+</sup> + H, calculated 285.1103).</u>

(4*R*,5*R*,*E*)-ethyl 4-benzyl-5-hydroxy-6-oxo-phenylhex-2-enoate (10f). As a diastereoisomer mixture (55:45, *anti:syn*). Colorless oil. (0.044 g, 52%);  $[α]^{26}_{D} = -18$  (*c*= 0.4 ; CHCl<sub>3</sub>); *R*<sub>f</sub>= 0.35 (Hex/EtOAc; 70:30, revealed with KMnO<sub>4</sub>). IR: v 3452.8 (OH), 1711.5 (C=O), 1656.7 (C=O), 1249.1 (CH<sub>3</sub>-C=O). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 7.65 - 7.28 (m, 10 H, ArH), 6.77 (dd, *J* = 15.8, 8.8 Hz, 1 H, <u>CH</u>=CH), 5.47 (dd, *J* = 15.8, 0.7 Hz, 1 H, CH=<u>CH</u>), 4.99 (dd, *J* = 6.0, 1.4 Hz, 1 H, <u>CH</u>OH), 4.14 (q, *J* = 7.1 Hz, 2 H, OCH<sub>2</sub>CH<sub>3</sub>), 3.72 (d, *J* = 6.1 Hz, 1 H, OH), 3.15 - 2.83 (m, 3 H), 1.26 (t, *J* = 7.1 Hz, 3 H, OCH<sub>2</sub>CH<sub>3</sub>). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 200.7 (C), 165.7 (C), 144.7 (CH), 138.6 (C), 134.1 (CH), 133.4 (C), 129.5 (2xCH), 128.9 (2xCH), 128.8 (2xCH), 128.3 (2xCH), 126.8 (CH), 123.8 (CH), 73.0 (CH), 60.5 (CH<sub>2</sub>), 48.8 (CH), 37.7 (CH<sub>2</sub>), 14.2 (CH<sub>3</sub>). HRMS calculated for C<sub>21</sub>H<sub>22</sub>O<sub>4</sub>: 338.1518 found: 339.1600 (M<sup>+</sup> + H, calculated 339.1596).

#### HPLC data for aldol products



The ee was determined by chiral GC analysis with a CP CHIRALSIL DEX CB column (80 °C, 13.4 Psi),  $R_t = 34.6 \text{ min (major)}$ ,  $R_t = 39.6 \text{ min (minor)}$ .



The ee was determined as the benzoate ester.



The ee was determined by chiral HPLC on Chiralpak OD-H column (99% hexane, 1%  $Pr^{1}OH$ , 25°C, 1 mL/min, 230 nm,  $R_{t}$  = 12.8 min (minor *syn*),  $R_{t}$  = 14.9 min (major *syn*),  $R_{t}$  = 16.1 min (major *anti*),  $R_{t}$  = 22.3 min (minor *anti*).



The ee was determined by chiral GC analysis with a CP CHIRALSIL DEX CB column (120 °C, 13.4 Psi),  $R_t = 21.3 \text{ min}$  (major *anti*),  $R_t = 22.8 \text{ min}$  (minor *anti*),  $R_t = 26.4 \text{ min}$  (major *syn*),  $R_t = 31.8 \text{ min}$  (minor *syn*).



The ee was determined by chiral GC analysis with a CP CHIRALSIL DEX CB column (150 °C, 13.4 Psi),  $R_t = 8.8$  min (major *anti*),  $R_t = 9.7$  min (minor *anti*),  $R_t = 10.3$  min (major *syn*),  $R_t = 11.0$  min (minor *syn*).



The ee was determined by chiral GC analysis with a CP CHIRALSIL DEX CB column (130 °C, 13.4 Psi),  $R_t = 67.0$  min (major *anti*),  $R_t = 70.7$  min (minor *anti*),  $R_t = 72.5$  min (major *syn*),  $R_t = 79.2$  min (minor *syn*).



The ee was determined by chiral HPLC on Chiralpak IA column (95% hexane, 5%  $Pr^{1}OH$ , 25°C, 1 mL/min, 230 nm,  $R_{t}$ = 17.7 min (minor *syn*),  $R_{t}$  = 18.8 min (major *anti*),  $R_{t}$  = 20.3 min (minor *anti*),  $R_{t}$  = 22.1 min (minor *syn*).



The ee was determined by chiral GC analysis with a CP CHIRALSIL DEX CB column (140 °C, 13.4 Psi),  $R_t = 60.2 \text{ min}$  (major *syn*),  $R_t = 62.9 \text{ min}$  (minor *syn*),  $R_t = 69.9 \text{ min}$  (major *anti*),  $R_t = 71 \text{ min}$  (minor *anti*).



The ee was determined by chiral GC analysis with a CYCLOHEXIL  $\beta$  column (130 °C, 13.4 Psi),  $R_t = 42.9 \text{ min}$  (major),  $R_t = 45.5 \text{ min}$  (minor).



The ee was determined by chiral HPLC on AD-H column (95% hexane, 5%  $Pr^{i}OH$ , 25°C, 1 mL/min, 230 nm,  $R_{t}$ = 22.2 min (minor),  $R_{t}$ = 24.8 min (major).



The ee was determined by chiral GC analysis with a CYCLOHEXIL  $\beta$  column (165 °C, 13.4 Psi),  $R_t = 27.8 \text{ min}$  (major),  $R_t = 29.0 \text{ min}$  (minor).



The ee was determined by chiral GC analysis with a CP CHIRALSIL DEX CB column (130 °C, 13.4 Psi),  $R_t = 9.1$  min (major *anti*),  $R_t = 9.8$  min (minor *anti*),  $R_t = 11.1$  min (major *syn*),  $R_t = 12.9$  min (minor *syn*).



The ee was determined by chiral GC analysis with a LIPODEX E column (145 °C, 13.4 Psi),  $R_t = 47.6 \text{ min (minor anti)}, R_t = 50.0 \text{ min (major anti)}, R_t = 55.4 \text{ min (syn)}.$ 



The ee was determined by chiral HPLC on Chiralpak IA column (97% hexane, 3%  $Pr^{i}OH$ , 25°C, 0.3 mL/min, 230 nm,  $R_t = 36.0$  min (minor *syn*),  $R_t = 37.6$  min (major *syn*),  $R_t = 39.6$  min (major *anti*),  $R_t = 46.5$  min (minor *anti*).



The ee was determined by chiral HPLC on Chiralpak IA column (95% hexane, 5%  $Pr^{i}OH$ , 25°C, 0.5 mL/min, 230 nm,  $R_t = 7.0$  min (major *syn*),  $R_t = 7.3$  min (minor *syn*),  $R_t = 7.7$  min (major *anti*),  $R_t = 8.7$  min (minor *anti*).



The ee was determined by chiral HPLC on Chiralpak AD-H column (97% hexane, 3%  $Pr^{i}OH$ , 25°C, 1 mL/min, 230 nm,  $R_t = 25.1$  min (major *anti*),  $R_t = 27.9$  min (minor *syn*),  $R_t = 29.6$  min (major *syn*),  $R_t = 33.2$  min (minor *anti*).



The ee was determined by chiral HPLC on Chiralpak IA column (95% hexane, 5%  $Pr^{i}OH$ , 25°C, 1 mL/min, 230 nm,  $R_t = 16.4$  min (major *syn*),  $R_t = 19.0$  min (minor *anti*),  $R_t = 22.0$  min (minor *syn*),  $R_t = 26.7$  min (major *anti*).



The ee was determined by chiral HPLC on Chiralpak IA column (98% hexane, 2%  $Pr^{1}OH$ , 25°C, 1 mL/min, 230 nm,  $R_t = 27.8$  min (major *anti*),  $R_t = 35.4$  min (minor *anti*),  $R_t = 37.0$  min (major *syn*),  $R_t = 39.9$  min (minor *syn*).













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676 673 673 6671 584 588 588 588 588 588 588 F 00 T 224 F 101 0.93 H 1.76 -3.03 -1 1.03 -1 17.44 4.20 <del>-</del> 6.0 5.5 5.0 4.5 4.0 f1 (ppm) 3.5 1.0 10.5 10.0 8.0 7.5 7.0 6.5 3.0 2.5 2.0 1.5 0.5 0.0 -0.5 9.5 9.0 8.5 - 165.7 - 145.7 - 123.5 - 60.5 0.67 - 45.6  $\begin{array}{c} 31.8\\ 31.5\\ 29.4\\ 29.2\\ 27.3\\ 27.3\\ 14.1\\ 14.1\end{array}$ 80 70 60 50 40 30 20 10 -10 0

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#	[min]		[min]	[pA*s]	[pA]	00
1	21.271	MM	0.3642	256.41824	11.73569	20.88270
2	22.822	MM	0.2436	8.35217	5.71418e-1	0.68020
3	26.421	MM	0.7746	895.60126	19.27095	72.93775
4	31.784	MM	0.3924	67.52645	2.86842	5.49935







#	[min]		[min]	[pA*s]	[pA]	olo
1	67.053	MF	1.3281	259.55905	3.25723	76.57417
2	70.723	MF	1.0739	26.16152	4.06014e-1	7.71808
3	72.549	FM	1.0761	40.55659	6.28127e-1	11.96486
4	79.233	MM	0.7708	12.68706	2.74316e-1	3.74289



#	[min]	- 17 -	[min]	[mAU*s]	[mAU]	8	
1	17.702	MM T	0.2954	141.31609	7.97360	2.5221	
2	18.768	MM T	0.5259	5084.40479	161.12408	90.7440	
3	20.282	MM T	0.3800	118.91827	5.21607	2.1224	
4	22.117	MM	0.5273	258.37894	8.16640	4.6114	

O OH



Peak	RetTime	Type	Width	Area	Height	Area
#	[min]		[min]	[pA*s]	[pA]	8
				-		
1	60.272	MF	1.0633	278.51883	4.36555	40.29986
2	62.904	FM	1.1537	140.48114	2.02949	20.32670
3	69.906	MF	0.7514	70.81514	1.57077	10.24649
4	70.957	FM	1.3743	201.30104	2.44132	29.12695





Peak RetT	ime Type	Width	Area	Height	Area
# [m1]	nj	[min]	[pA*s]	[pA]	8
1 40.	611 MM	0.3919	14.06788	5.98318e-1	0.80066
2 41.	982 MM	0.3836	29.90887	1.29935	1.70223
3 42.	937 MF	0.4870	1456.85510	49.85872	82.91518
4 44.	372 FM	0.5822	111.78307	3.19989	6.36200
5 45.	892 MF	0.4814	59.85265	2.07216	3.40644
6 46.	841 FM 794 MF	0.5046	18.42756	6.08637e-1	1.04878
8 49.4	115 FM	0.4556	51.95185	1.90058	2.95678



Peak ‡	RetTime [min]	Туре	Width [min]	Area [mAU*s]	Height [mAU]	Area
	21 422	104 7	0.0012	100 61208	4 00024	0 4725
+	21.932	man 1	0.3912	109.01290	1.90924	0.4720
2	22.372	MM T	0.5396	998.37714	30.83524	4.3033
3	24.952	MF T	0.6105	1.48264e4	404.72870	63.9057
4	26.800	MF T	0.6282	5761.41455	152.86011	24.8333
5	27.865	FM T	0.8002	1211.16748	25.22749	5.2205
6	31.398	MM	0.5444	293.44104	8.98300	1.2648







Peak	RetTime	Type	Width	Area	Height	Area
#	[min]		[min]	[pA*s]	[pA]	90
1	9.142	MM	0.1715	335.39655	32.58740	59.57887
2	9.798	MM	0.1127	17.51009	2.58992	3.11044
3	11.143	MM	0.1729	161.10751	15.53290	28.61867
4	12.923	MM	0.1547	48.93132	5.27145	8.69202



2	49.980	MM	1.0644	126.13940	1.97509	72.91486
з	55.423	MM	1.8025	41.78198	3.86335e-1	24.15207



10b-*Rac* 





4

8.824 VV

0.1914 277.89667



2.5039

857.90094 95.6284

22.92982



10d-*Rac* 

29.611 MM T 0.5454 33.246 MM T 0.7397

3

4



0.8503

1.33217

1.50849

43.59312

66.95388





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### References

<sup>1</sup> a) G. Guillena, M. C. Hita, C. Nájera and S. F. Viozquez, J. Org. Chem. 2008, 73, 5933. b) S. F. Viozquez, G. Guillena, C. Nájera, B. Bradshaw, G. Etxebarria-Jardi and J. Bonjoch, Org. Synth. 2011, 88, 317.

<sup>2</sup> Alberg, D. G.; Poulsen, T. B.; Bertelsen, S.; Christensen, K. L.; Birkler, R. D.; Johannsen, M.; Jørgensen, K. A. Bioorg. Med. Chem. Lett. 2009, 19, 3888.

<sup>3</sup> Smith, III, A. B.; Liverton, N. J.; Hrib, N. J.; Sivaramakrishnan, H.; Winzenberg, K. J. Am. Chem. Soc. **1986**, *108*, 3040.

<sup>4</sup> Mayring, L.; Severin, T. Chem. Ber., **1981**, 114, 3863.

<sup>5</sup> Hayashi, Y.; Yasui, Y.; Kojima, M. Kawamura, T., Ishikawa, H. Chem. Commun. **2012**, 48, 4570.

.1, 114, .wamura, Τ., .