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Draft Genome Sequences of Two Extensively Drug-Resistant Strains of *Acinetobacter baumannii* Isolated from Clinical Samples in Pakistan

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ABSTRACT Infections in immunocompromised patients that are caused by extensively drug-resistant (XDR) *Acinetobacter baumannii* strains have been increasingly reported worldwide. In particular, carbapenem-resistant *A. baumannii* strains are a prominent cause of health care-associated infections. Here, we report draft genome assemblies for two clinical XDR *A. baumannii* isolates obtained from hospitalized patients in Pakistan.

Acinetobacter baumannii, a Gram-negative opportunistic pathogen of the *Moraxellaceae* family, can carry multiple antimicrobial resistance (AMR) determinants. Infections and outbreaks caused by multidrug-resistant (MDR) or extensively drug-resistant (XDR) *A. baumannii* strains in immunocompromised patients have been increasingly reported worldwide (1), with isolates often belonging to international clone 2/sequence type 2 (ST2) (2). Carbapenem-resistant *A. baumannii* (CRAB) strains are a significant cause of health care-associated infections in Pakistan, with various prevalence rates (62% to 100%) (3). The most common β -lactamases in CRAB strains are acquired (e.g., *bla*_{OXA-23}, *bla*_{OXA-40}, *bla*_{OXA-58}, *bla*_{OXA-143}, and *bla*_{OXA-235}) and intrinsic (e.g., *bla*_{OXA-51} and *bla*_{OXA-69}) carbapenem-hydrolyzing oxacillinases (4).

Draft genomes are reported here for two clinical XDR *A. baumannii* isolates obtained from urine (CFSAN059604, isolated in 2004) and throat (CFSAN059618, isolated in 1998) specimens from hospitalized patients in Pakistan. Patient samples were inoculated onto nonselective (e.g., blood agar) and differential (e.g., MacConkey agar) plates and incubated for 24 to 48 h at 37°C. API 20E and Vitek 2 (bioMérieux) systems were used for species identification and confirmation, respectively. Susceptibility testing against clinically relevant antimicrobials was performed by conventional broth microdilution (5) following CLSI and EUCAST guidelines and breakpoints (5–7). The two isolates were resistant to 17 antibiotics with the same MICs, as follows: ≥ 128 $\mu\text{g/ml}$ for piperacillin-tazobactam; ≥ 64 $\mu\text{g/ml}$ for cefotaxime and aztreonam; ≥ 32 $\mu\text{g/ml}$ for ampicillin, ceftriaxone, and tetracycline; ≥ 16 $\mu\text{g/ml}$ for ceftazidime, ceftazidime-avibactam, gentamicin, and chloramphenicol; and ≥ 8 $\mu\text{g/ml}$ for ceftazidime, doripenem, meropenem, ertapenem, ciprofloxacin, levofloxacin, and trimethoprim-sulfamethoxazole. CFSAN059604 and CFSAN059618 were also resistant, but with different MICs, to ampicillin-sulbactam (≥ 32 and ≥ 16 $\mu\text{g/ml}$, respectively), ceftazidime (≥ 128 and ≥ 16 $\mu\text{g/ml}$, respectively), cefepime (≥ 32 and ≥ 16 $\mu\text{g/ml}$, respectively), and imipenem (≥ 8 and ≥ 16 $\mu\text{g/ml}$, respectively). Both isolates were susceptible to colistin (≤ 2 $\mu\text{g/ml}$) and minocycline (≤ 4 $\mu\text{g/ml}$). Finally, CFSAN059604 and CFSAN059618 had markedly dif-

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TABLE 1 Assembly statistics, accession numbers, and AMR genes for the XDR *Acinetobacter baumannii* isolates examined in this work

| Sample ^a | BioSample accession no. | Yr isolated | SRA accession no. | GenBank accession no. | No. of contigs | Total length (bp) | N ₅₀ (bp) | GC content (%) | Genome coverage (x) | Avg read quality score for: | | | No. of reads | No. of AMR genes |
|---------------------|-------------------------|-------------|-------------------|-----------------------|----------------|-------------------|----------------------|----------------|---------------------|-----------------------------|-----------------|------------|-----------------|------------------|
| | | | | | | | | | | Read 1 | Read 2 | Read 2 | | |
| CFSAN059604 | SAMN10086814 | 2004 | SRR8837010 | SSMO000000000 | 48 | 3,944,772 | 261,641 | 38.9 | 712 | 32 ^b | 30 ^b | 21,896,362 | 14 ^c | |
| CFSAN059618 | SAMN10086821 | 1998 | SRR8837150 | SSMN000000000 | 79 | 3,907,254 | 116,856 | 38.8 | 93 | 35 | 33 | 1,792,448 | 8 ^d | |

^a The two isolates belong to NCBI BioProject PRJNA342326.

^b Average of four lanes.

^c Genes *aac(3)-I*, *aadA1*, *adeC*, *ant(3'')-IIa*, *aph(3'')-Ib*, *aph(3'')-Ia*, *aph(6)-Ia*, *aph(6)-Ia*, *bla_{ADC}*, *bla_{OXA-254'}*, *bla_{TEM-1'}*, *qacEΔ1*, *sul1*, *sul2*, and *tet(B)*.

^d Genes *ant(2'')-Ia*, *ant(3'')-IIa*, *aph(3'')-VIa*, *bla_{ADC-76'}*, *bla_{OXA-235'}*, *bla_{OXA-68'}*, *sul2*, and *tet(39)*.

ferent resistance profiles for amikacin (≤ 2 and ≥ 64 $\mu\text{g/ml}$, respectively) and tobramycin (≤ 1 and ≥ 16 $\mu\text{g/ml}$, respectively).

Isolates were grown overnight in lysogeny broth (Lennox), and DNA was extracted using the DNeasy blood and tissue kit (Qiagen). Libraries were prepared using the Nextera XT DNA library preparation kit and sequenced on a MiSeq (CFSAN059618) or NextSeq (CFSAN059604) sequencer (Illumina), with paired-end sequencing technology ($2 \times 250\text{-bp}$ and $2 \times 150\text{-bp}$ sequencing, respectively). Minimum sequence quality was represented by average coverage greater than $50\times$ and Q scores for reads 1 and 2 greater than 30 (8). Absence of contamination was confirmed with Kraken (9). Default parameters were used unless otherwise noted. *De novo* assemblies were obtained with Shovill v0.9 (<https://github.com/tseemann/shovill>), available in the GalaxyTrakr pipeline (10). The “trim reads” option was selected, and 500 bp was set as the minimum contig length. Draft genomes were annotated using the NCBI Prokaryotic Genome Annotation Pipeline (11). Table 1 lists the number of reads, number of contigs per assembly, genome size, and GC content for each isolate. The isolates were assigned to ST2 (CFSAN059604) and ST23 (CFSAN059618), based on the multilocus sequence typing Pasteur scheme (<https://pubmlst.org/abaumannii>). NCBI Pathogen Detection (PD) (<https://www.ncbi.nlm.nih.gov/pathogens>) was used to identify 14 (CFSAN059604) and 8 (CFSAN059618) AMR genes. NCBI PD uses AMRFinderPlus to assign the most specific AMR protein by using a hierarchy of gene families/symbols/names (<https://www.ncbi.nlm.nih.gov/pathogens/antimicrobial-resistance/AMRFinder>). Results are available in Table 1 and from the NCBI PD website (CFSAN059604 and CFSAN059618).

The described draft genomes will be useful in comparative genomic analyses of *A. baumannii* strains from different regions and clinical settings. These data can also provide phylogenetic insights into the emergence of XDR *A. baumannii* strains and support epidemiological investigations of outbreaks.

Data availability. The complete genome sequences of *A. baumannii* CFSAN059604 (SRA number [SRR8837010](https://www.ncbi.nlm.nih.gov/sra/SRR8837010)) and CFSAN059618 (SRA number [SRR8837150](https://www.ncbi.nlm.nih.gov/sra/SRR8837150)) are available in GenBank under accession numbers [SSMO00000000](https://www.ncbi.nlm.nih.gov/nuccore/SSMO00000000) and [SSMN00000000](https://www.ncbi.nlm.nih.gov/nuccore/SSMN00000000), respectively (first versions).

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