Mitochondrial Complex 1, Sigma 1, and Synaptic Vesicle 2A in Early Drug-Naive Parkinson's Disease

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ABSTRACT: Background: Dysfunction of mitochondrial energy generation may contribute to neurodegeneration, leading to synaptic loss in Parkinson's disease (PD). The objective of this study was to find cross-sectional and longitudinal changes in PET markers of synaptic vesicle protein 2A, sigma 1 receptor, and mitochondrial complex 1 in drug-naive PD patients.

Methods: Twelve early drug-naive PD patients and 16 healthy controls underwent a 3-Tesla MRI and PET imaging to quantify volume of distribution of [¹¹C]UCB-J, [¹¹C]SA-4503, and [¹⁸F]BCPP-EF for synaptic vesicle protein 2A, sigma 1 receptor, and mitochondrial complex 1, respectively. Nine PD patients completed approximately 1-year follow-up assessments.

Results: Reduced [¹¹C]UCB-J volume of distribution in the caudate, putamen, thalamus, brain stem, and dorsal raphe and across cortical regions was observed in drug-naive PD patients compared with healthy controls. [¹¹C]UCB-J volume of distribution was reduced in the locus coeruleus and substantia nigra but did not reach statistical significance. No significant differences were found in [¹¹C]SA-4503 and

[¹⁸F]BCPP-EF volume of distribution in PD compared with healthy controls. Lower brain stem [¹¹C]UCB-J volume of distribution correlated with Movement Disorder Society Unified Parkinson's Disease Rating Scale part III and total scores. No significant longitudinal changes were identified in PD patients at follow-up compared with baseline. updates

Conclusions: Our findings represent the first in vivo evidence of mitochondrial, endoplasmic reticulum, and synaptic dysfunction in drug-naive PD patients. Synaptic dysfunction likely occurs early in disease pathophysiology and has relevance to symptomatology. Mitochondrial complex 1 and sigma 1 receptor pathology warrants further investigations in PD. Studies in larger cohorts with longer follow-up will determine the validity of these PET markers to track disease progression. © 2020 The Authors. *Movement Disorders* published by Wiley Periodicals, Inc. on behalf of International Parkinson and Movement Disorder Society.

Key Words: mitochondria; molecular biomarkers; Parkinson's disease; positron emission tomography; synaptic vesicle protein

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Preclinical evidence suggests that the mechanisms Ki underlying the pathophysiology of Parkinson's disease (PD) involves mitochondrial,¹ endoplasmic reticulum (ER),² and synaptic³ function. Synaptic vesicle protein 2A (SV2A) is a transmembrane protein widely expressed in presynaptic terminals throughout the brain, where it regulates neurotransmission.^{4,5} Neurons lacking SV2A demonstrate impaired ability of vesicles to fuse with the plasma membrane.⁶ There are several lines of evidence suggesting that accumulation of α -synuclein in presynaptic

damage and impairment of intracellular trafficking.⁷⁻¹⁶ Sigma 1 receptor (σ 1R) is a chaperone protein residing at the mitochondrion-associated endoplasmic reticulum membrane, where it modulates calcium influx into the mitochondrion from the ER and is thought to play a role in regulating the activity of potassium channels.¹⁷ Activation of σ 1Rs stimulates neuromodulation and neuroprotection.¹⁸ The σ 1R facilitates the proper folding of newly synthesized proteins, but also prevents accumulation of misfolded proteins, suggesting that σ 1R plays a key role in cellular survival, and its upregulation may be neuroprotective, given that it may prevent the accumulation of abnormal proteins.^{19,20} Furthermore, σ 1R stimulates striatal dopamine synthesis in rats and has been hypothesized to play a role in the pathophysiology of PD.²¹

terminals impairs synaptic proteins, synaptic plasticity,

and neurotransmission and subsequently induces axonal

Mitochondrial complex 1 (MC1) is the first enzyme complex in the electron transfer chain, having a critical role in oxidative phosphorylation in mitochondria.²² A common finding in animal models of PD as well as in brain tissue of patients with sporadic and familial forms of PD is the presence of histopathological evidence of progressive mitochondrial dysfunction.^{23,24} Mitochondrial dysfunction is mainly characterized by the generation of reactive oxygen species, decreased MC1 enzyme activity, adenosine triphosphate depletion, and caspase-3 activation. Marked decreases in MC1 activity and reduced mitochondrial DNA levels were widely found in postmortem PD brain samples.²⁵⁻²⁸

The development of selective PET radioligands [¹¹C] UCB-J for SV2A,²⁹ [¹¹C]SA-4503 for σ 1R,³⁰ and [¹⁸F] BCPP-EF for MC1,^{31,32} allows for the in vivo investigation of presynaptic terminals and the mitochondrial/ER complex in neurodegenerative disorders. Here, we performed cross-sectional and longitudinal measurements using these 3 PET radioligands, 3-Tesla MRI, and a battery of clinical assessments in a cohort of early drugnaive PD patients.

Methods

Participant Demographics

Twenty-eight participants were recruited, 12 with idiopathic PD from specialist movement disorders clinics at King's College Hospital and 16 age-matched healthy controls through public advertisement. PD patients were recruited according to the following inclusion criteria: (1) PD diagnosed according to the Queen Square Brain Bank criteria; (2) duration of PD symptoms ≤ 24 months; (3) naive to PD treatment at baseline; (4) did not fulfill the diagnostic criteria for PD mild cognitive impairment,³³ dementia,³⁴ or depression³⁵); (5) no history of other neurological or psychiatric disorders; and (6) no contraindication to MRI. Healthy controls had no history of neurological or psychiatric disorders. None of the participants were on medications or supplements with known affinity for the SV2A, σ 1R, or MC1, including metformin, neuroactive steroids (such as dehydroepiandrosterone, progesterone, pregnenolone, testosterone, and deoxycorticosterone), haloperidol, fluvoxamine, donepezil, levetiracetam, and bevetiracetam.

All participants were successfully screened to undertake PET and MRI scanning under scanning safety criteria (http://www.mrisafety.com; https://www.gov. uk/government/publications/arsac-notes-for-guidance). The study was approved by the institutional review board and the research ethics committee. Permission to use radioactive substances was obtained by the Administration of Radioactive Substances Advisory Committee, Department of Health and Social Care, United Kingdom. Written informed consent was obtained from all study participants in accordance with the Declaration of Helsinki.

Clinical Assessments

Clinical assessments were performed at the NIHR King's Wellcome Clinical Research Facility (London, UK). Motor symptom severity was assessed using the Movement Disorder Society Unified Parkinson's Disease Rating Scale (MDS-UPDRS) part III and staged with the Hoehn and Yahr (H&Y) scale. MDS-UPDRS total scores were used to assess global disease burden. Mini-Mental Status Examination and Montreal Cognitive Assessment were used to assess global cognitive status. Nonmotor symptoms were assessed with the Non-Motor Symptoms Scale (NMSS), the Parkinson's Disease Sleep Scale for sleep disturbances, and the Beck Depression Inventory second edition for depression.

Longitudinal Follow-up

Eight PD patients completed longitudinal [¹¹C]UCB-J, [¹¹C]SA-4503, and [¹⁸F]BCPP-EF PET scans, MRI and clinical assessments at a mean follow-up period of 11 ± 1.3 months (range, 10–14 months). A ninth PD patient completed only [¹¹C]SA-4503 and [¹⁸F]BCPP-EF PET and clinical assessments at follow-up. At follow-up, all PD patients remained drug naive except for 1 patient who was started on levodopa treatment (daily levodopaequivalent dose of 150 mg) 6 months before the follow-up assessments. Motor assessments were performed OFF medication after overnight withdrawal of dopaminergic medication for this 1 PD patient.

Scanning Procedures

All participants were underwent 3 PET scans with ^{[11}C]UCB-J, ^{[11}C]SA-4503, and ^{[18}F]BCPP-EF and one 3-Tesla MRI at Invicro (London, UK) on a Siemens Biograph Hi-Rez 6 or Biograph TruePoint 6 PET-CT scanner (Siemens Healthcare, Erlangen, Germany), and Siemens 3-Tesla Magnetom Trio MRI scanner with a 32-channel head coil (Siemens Healthcare, Erlangen, Germany), respectively. [¹¹C]UCB-J (235 \pm 42.9 MBq), $[^{11}C]SA-4503$ (252 \pm 36.0 MBq) and $[^{18}F]BCPP-EF$ $(90 \pm 5.0 \text{ MBg})$ were administered intravenously as a slow bolus over 20 seconds. For each subject at baseline and follow-up, [11C]UCB-J PET, [11C]SA-4503 PET, ¹⁸F]BCPP-EF PET, and MRI were performed within a period of no more than 16 days. Nonsteroidal antiinflammatory drugs (such as aspirin, indomethacin, diclofenac, piroxicam, and ibuprofen) were discontinued at least 4 days prior to the [18F]BCPP-EF PET scans.

Dynamic emission PET data were acquired continuously for 90 minutes following the injection of [¹¹C] UCB-J, [¹¹C]SA-4503, and [¹⁸F]BCPP-EF and reconstructed into 26 frames (8×15 , 3×60 , 5×120 , 5×300 , and 5×600 seconds), using a filtered backprojection algorithm (direct inversion Fourier transform) with a 128 matrix, 2.6 zoom producing images with an isotropic voxel size of $2 \times 2 \times 2$ mm³ and smoothed with a transaxial Gaussian filter of 5 mm. All participants underwent arterial sampling and metabolite analysis to enable generation of a parent plasma input function for all 3 tracers, [¹¹C]UCB-J, [¹¹C]SA-4503, and [¹⁸F]BCPP-EF.

MRI scans were acquired for the following sequences using a 1 mm³ voxel size, anteroposterior phase encoding direction, and a symmetric echo. T1-weighted magnetization prepared rapid gradient echo sequence for coregistration with the PET images; fast gray matter T1 inversion recovery (FGATIR³⁶) for volumetric quantification of the substantia nigra and neuromelaninsensitive T1-weighted for delineation of the locus coeruleus. Full acquisition parameters are outlined in the Supplemental Materials.

imaging Data Analysis MRI-Based Volumetric Analysis

Cortical thickness and deep gray matter nuclei volumetric changes were investigated with FreeSurfer image analysis suite (version 6.0), as previously validated,³⁷ in healthy controls and PD patients at baseline and follow-up. All individual nuclei volumes were normalized for intracranial volume.³⁸ To better quantify volumetric changes, the substantia nigra was manually delineated on the FGATIR sequence,³⁹ using the Analyze medical imaging software package (version 12.0; Mayo Foundation, Overland Park, Kansas, United States).

PET Data Analysis

All PET data analysis was performed as previously described.⁴⁰ Briefly, the Molecular Imaging and Kinetic Analysis Toolbox software package (MIAKAT version http://www.miakat.org), 4.3.7: implemented in MATLAB (version R2016a; The Mathworks, Natick, MA) was used to carry out image processing and kinetic modeling. MIAKAT combines inhouse code with wrappers for FMRIB Software Library (FSL; version 5.0.9: http://fsl.fmrib.ox.ac.uk/fsl/fslwiki/) and Statistical Parametric Mapping (SPM; version 12; http:// www.fil.ion.ucl.ac.uk/spm/). Individual PET frames were corrected for head motion using frame-by-frame rigid registration using a frame with high signal-tonoise ratio as reference. Plasma input function of unmetabolized radioligand was generated using the continuous blood, discrete blood/plasma samples and metabolite assays. For [¹¹C]UCB-J and [¹⁸F]BCPP-EF the arterial input function was obtained by plasma-towhole blood ratios fitted with a constant fit and a sigmoid fit for parent fraction. For [¹¹C]SA-4503 plasmato-whole blood ratios were fitted with an exponential approach to constant fit and an exponential plus constant fit for parent fraction. The plasma free fraction $(f_{\rm p})$ was measured by ultrafiltration in triplicate using an arterial blood sample taken prior to tracer injection as previously described.⁴⁰ Where there were differences in $f_{\rm p}$, regional differences in tracer uptake were explored using V_T/f_p as an outcome measure. Regional estimates of [¹¹C]SA-4503 and [¹⁸F]BCPP-

Regional estimates of [¹¹C]SA-4503 and [¹⁸F]BCPP-EF volume of distribution (V_T) were generated using the graphical model multilinear analysis 1 (MA1⁴¹; $t^* = 30$ minutes) with blood volume correction. [¹¹C] SA-4503 and [¹⁸F]BCPP-EF parametric V_T maps were generated with Logan Graphical analysis.⁴² Regional estimates of [¹¹C]UCB-J V_T were generated using the 1-tissue compartmental model (1TC) with blood volume correction and parametric V_T maps generated using 1TC model.⁴³ The centrum semiovale has been previously suggested as a white-matter pseudo-reference region for [¹¹C]UCB-J.⁴⁴ Therefore, for [¹¹C]UCB-J, a distribution volume ratio minus 1 of the target region and the centrum semiovale (DVR-1) was estimated from the volumes of distribution ([V_T^{ROI}/V_T^{CS}] – 1).

Region-of-Interest-Based Analysis

To facilitate anatomical delineation of regions of interest, PET images were anatomically coregistered and resliced to the corresponding volumetric structural

| D | Age (years) | Disease duration (months) | H&Y OFF | MDS-UPDRS total | MDS-UPDRS-III OFF | NMSS | BDI-II | MMSE | MoCA | PDSS |
|-------------------------------------|--------------|---------------------------|-------------|-----------------|-------------------|-------------------------------|----------------|----------------------------------|----------------|------------------|
| 1 | 59 | 4 | - | 6 | ω | | 2 | 30 | 30 | 145 |
| 2 | 66 | - | - | 25 | 17 | 15 | с | 30 | 28 | 130 |
| ε | 69 | 41 | - | 18 | 10 | 2 | с | 26 | 28 | 148 |
| 4 BL | 49 | 12 | - | 23 | 13 | 19 | - | 30 | 28 | 108 |
| 4 FU | 50 | 26 | - | 26 | 15 | 24 | 0 | 30 | 28 | 119 |
| 5 BL | 63 | 5 | - | 28 | 20 | 30 | с | 30 | 30 | 115 |
| 5 FU | 64 | 16 | - | 35 | 23 | 44 | 0 | 30 | 30 | 101 |
| 6 BL | 47 | 33 | - | Ħ | ø | ę | 0 | 30 | 30 | 142 |
| 6 FU | 48 | 43 | - | 15 | 12 | 5 | 0 | 30 | 30 | 144 |
| 7 BL | 63 | 20 | - | 16 | 8 | 18 | с С | 30 | 30 | 141 |
| 7 FU | 64 | 31 | - | 22 | 14 | 22 | 2 | 29 | 29 | 144 |
| 8 BL | 47 | 36 | - | 20 | 8 | 21 | 4 | 29 | 28 | 129 |
| 8 FU | 48 | 46 | 2 | 22 | Ħ | 28 | 7 | 30 | 30 | 128 |
| 9 BL | 57 | 24 | - | 49 | 28 | 4 | 9 | 30 | 30 | 134 |
| 9 FU | 58 | 34 | 2 | 56 | 36 | 7 | 4 | 29 | 30 | 145 |
| 10 BL | 77 | 12 | - | 25 | 18 | 18 | с С | 30 | 30 | 133 |
| 10 FU | 78 | 24 | 2 | 41 | 28 | 23 | 2 | 27 | 29 | 133 |
| 11 BL | 59 | 11 | - | 23 | 16 | с | 2 | 26 | 28 | 131 |
| 11 FU | 60 | 22 | - | 28 | 18 | 4 | - | 30 | 30 | 143 |
| 12 BL | 62 | 24 | - | 33 | 15 | 18 | 6 | 30 | 30 | 130 |
| 12 FU | 63 | 35 | - | 37 | 24 | 20 | 8 | 30 | 30 | 137 |
| PD baseline (n = 12), mean \pm SD | 59.8 ± 9.0 | 18.6 ± 13.2 | 1 ± 0 | 23.3 ± 10.6 | 14.1 ± 6.2 | 12.7 ± 9.6 | 3.3 ± 2.3 | 29.3 ± 1.5 | 29.2 ± 1.0 | 132.2 ± 11.7 |
| PD follow-up (n = 9), mean \pm SD | 59.2 ± 9.7 | 30.8 ± 9.7 | 1.3 ± 0.5 | 31.3 ± 12.4 | 20.1 ± 8.3 | 19.7 ± 12.8 | 2.7 ± 3.0 | $\textbf{29.4} \pm \textbf{1.0}$ | 29.6 ± 0.7 | 132.7 ± 14.8 |
| Within-subject change at follow-up, | I | Ι | 0.3 ± 0.5 | 6.0 ± 4.1 | 5.2 ± 3.1 | $\textbf{4.8}\pm\textbf{3.9}$ | -0.8 ± 1.6 | 0.2 ± 1.1 | 0.0 ± 1.9 | 3.4 ± 8.2 |
| mean \pm su | | | | | | | | | | |

| E 2. Cross-sectional analysis of synaptic vesicle protein 2A, sigma 1 receptor, and mitochondrial complex 1 levels in early drug-naive Parkinson's disease | patients compared with healthy controls | |
|--|---|--|
| ABLE 2. Cross-sectional analysis of synaptic vesicle protein 2A, sigma 1 receptor | patients compared with | |

| Badioligand | | | | | | | | Regio | n of interest | | | | | | |
|--|----------------------|----------------|------------------|-----------------|----------------------------------|----------------|-----------------|----------------|-----------------|-----------------|-------------------------------|--------------------------------|--------------------------------|----------------|---------------------------------|
| (target) | Cohort | FC | РС | TC | 00 | SNI | ЧIР | CAU | PUT | THA | BST | Medianraphe | Dorsalraphe | SN | ГС |
| [¹¹ C]UCB-J V _T (SV?A) | HC (n = 16) | 17.7 ± 2.1 | 18.0 ± 2.3 | 19.1 ± 1.8 | 18.2 ± 2.0 | 20.7 ± 2.0 | 14.3 ± 1.5 | 14.2 ± 2.7 | 22.5 ± 2.1 | 13.3 ± 2.0 | $\textbf{8.5}\pm\textbf{1.1}$ | $\textbf{8.7}\pm\textbf{1.0}$ | 10.1 ± 1.2 | 8.6 ± 0.9 | $\textbf{9.6} \pm \textbf{1.0}$ |
| | PD^a 11) | 15.8 ± 2.2 | 16.0 ± 2.2 | 17.4 ± 1.9 | 16.2 ± 2.0 | 19.1 ± 2.3 | 13.2 ± 1.6 | 12.1 ± 2.6 | 20.3 ± 2.3 | 11.8 ± 2.2 | 7.7 ± 0.9 | 8.1 ± 0.9 | 9.2 ± 1.1 | 8.0 ± 0.9 | $\textbf{8.8} \pm \textbf{1.3}$ |
| | % change | -10.3% | -11.0% | -9.0% | -11.0% | -7.9% | -7.8% | -15.1% | -9.7% | -11.6% | -9.4% | -6.7% | -9.4% | -6.9% | -8.7% |
| | Cohen's d | 0.86 | 0.88 0.88 | 0.92 0.92 | 1.00 | 0.77 | 160.0 0.71 | 0.82 0.82 | 0.09 | 0.74 | 0.81 | 0.61 | 0.83 | 0.04 0.68 | 0.73 |
| [¹¹ C]SA-4503 V _T | HC (n = 16) | 39.4 ± 8.6 | 37.4 ± 6.8 | 39.7 ± 6.5 | 35.5 ± 6.8 | 43.8 ± 6.8 | 37.6 ± 5.7 | 28.6 ± 7.5 | 42.5 ± 7.5 | 35.4 ± 7.1 | 36.5 ± 6.3 | 41.4 ± 12.2 | $\textbf{35.6}\pm\textbf{7.6}$ | 34.5 ± 7.5 | 28.3 ± 6.4 |
| | (01 = 10) PD | 35.4 ± 7.7 | 34.0 ± 6.3 | 37.8 ± 7.1 | 34.3 ± 6.1 | 42.7 ± 7.9 | 35.1 ± 6.5 | 22.9 ± 6.0 | 40.7 ± 7.4 | 30.9 ± 8.2 | 32.9 ± 6.5 | 36.3 ± 10.6 | 32.1 ± 8.1 | 34.4 ± 5.7 | 28.0 ± 8.3 |
| | (n = 12) % change | 10.00/ | 0.10/ | 70/L V | 701/ 6 | 706 6 | C 50/ | 201.100 | 706 1 | 10 00/ | 700 0 | 707 61 | 700.0 | 701/0 | 700 0 |
| | ™ ulaliye P | -10.0% >0.1 | -3.1% >0.1 | -4.7% >0.1 | -3.4% >0.1 | -2.3% >0.1 | -0.3% >0.1 | -20.1% >0.1 | -4.2% >0.1 | - 12.0% >0.1 | -9.9% >0.1 | -12.4% >0.1 | -9.9% >0.1 | -0.4% >0.1 | -0.0% >0.1 |
| | Cohen's d | 0.48 | 0.52 | 0.27 | 0.19 | 0.14 | 0.40 | 0.85 | 0.24 | 0.59 | 0.57 | 0.45 | 0.45 | 0.02 | 0.03 |
| [¹⁸ F]BCPP-EF V _T | нс | 24.9 ± 4.4 | 25.9 ± 5.1 | 24.5 ± 4.1 | $\textbf{27.6} \pm \textbf{4.8}$ | 26.3 ± 4.4 | 21.5 ± 3.7 | 23.2 ± 5.5 | 36.0 ± 6.3 | 25.4 ± 5.1 | 18.6 ± 3.1 | $\textbf{22.4}\pm\textbf{3.8}$ | 22.0 ± 3.3 | 20.7 ± 3.1 | 20.3 ± 3.5 |
| (MC1) | (n = 16) PD | 23.0 + 5.4 | 24.2 + 6.0 | 22.9 ± 4.8 | 25.2 + 5.8 | 24.8 + 4.9 | 19.9 + 4 | 21.0 + 6.2 | 33.3 + 6.9 | 22.5 + 6.0 | 16.6 + 3.2 | 20.7 + 4.3 | 19.6 + 3.9 | 19.3 ± 4.0 | 18.4 ± 4.1 |
| | (n = 12) | | | | | | | | | | | | | | |
| | % change | -7.5% | -6.6% | -6.7% | -8.9% | -5.8% | -7.2% | -9.4% | -7.6% | -11.3% | -10.9% | -7.6% | -10.8% | -6.7% | -9.0% |
| | Ρ | >0.1 | >0.1 | >0.1 | >0.1 | >0.1 | >0.1 | >0.1 | >0.1 | >0.1 | >0.1 | >0.1 | >0.1 | >0.1 | >0.1 |
| | Cohen's d | 0.39 | 0.30 | 0.37 | 0.46 | 0.33 | 0.40 | 0.37 | 0.41 | 0.52 | 0.64 | 0.42 | 0.66 | 0.39 | 0.48 |
| PD, Parkinson | s disease; H | C, healthy co | ontrol; FL, fror | ntal cortex; Pl | L, parietal co | rtex; TL, temp | ooral cortex; (| JL, occipital | cortex; INS, ii | nsula; HIP, hi | ppocampus; (| CAU, caudate; P | UT, putamen; T | H, thalamus; | BST, brain |

stem; SN, substantia nigra; LC, locus coeruleus; MC1, mitochondrial complex 1; SV2A, synaptic vesicle protein 2A, σ1R, sigma 1 receptor.

T1-weighted MRI images in SPM. The template brain image and associated Clinical Imaging Centre atlas³⁹ was then nonlinearly warped to the individual subject's MRI image where the following regions of interest were defined: frontal cortex, insular cortex, temporal cortex, parietal cortex, hippocampus, brain stem, thalamus, caudate, putamen, and substantia nigra. Dorsal and median raphe nuclei were defined using the Harvard Ascending Arousal Network Atlas.⁴⁵ To tailor regions of interest to a subject's individual anatomy, the subject's segmented gray matter was used to mask the regions of interest. Time-activity curves were generated within the gray matter-masked regions of interest. The locus coeruleus was manually delineated on the



FIG. 1. Bar graphs showing cross-sectional analysis of synaptic vesicle protein 2A, sigma 1 receptor and mitochondrial complex 1 in drug-naive Parkinson's disease patients. Regional total distribution volume (V_T) in [¹¹C]UCB-J (**A**), [¹¹C]SA-4503 (**B**), and [¹⁸F]BCPP-EF (**C**) in drug-naive Parkinson's disease patients compared with healthy controls. *P < 0.05. [Color figure can be viewed at wileyonlinelibrary.com]

neuromelanin-sensitive T1-weighted MRI images using Analyze.

Statistical Analysis

Statistical analysis and graph illustration were performed with SPSS (version 25; Chicago, IL) and GraphPad Prism (version 7.0c) for Mac OS X, respectively. For all variables. Gaussianity was tested with the Shapiro-Wilk test. We proceeded with parametric tests, as our data were normally distributed. Independent t tests were used to assess between-group differences in clinical variables. Multivariate analysis of covariance was used to assess between-group differences in regions of interest for each PET tracer and MRI volumetric data. Although there was no difference in age between groups (Table 1), age was included as a covariate because of a possible age effect on these PET markers.^{40,46} *P* values for each variable were calculated following Benjamin-Hochberg correction to control the false discovery rate (FDR).⁴⁷ Paired t tests were used to assess longitudinal changes in clinical, PET, and MRI variables. The within-subject annual rate of change was calculated as follows: (% change from baseline to follow-up $\times 12$ months)/between-scan internal (months). We interrogated correlations between PET and clinical data using Pearson's r, and we applied Benjamin-Hochberg correction to control the FDR.⁴⁷ The relationship between SV2A, σ 1R, and MC1 was investigated at a molecular level in exploratory analysis by interrogating correlations between the regional uptake of each PET tracer within the drug-naive PD cohort. In this exploratory analysis, correction for multiple comparisons was not applied. We tested for outliers using the ROUT outlier test, based on the FDR, in GraphPad Prism. For longitudinal analysis, follow-up period and change in age were included as covariates. We set the false discovery rate cutoff at 0.05. All data are presented as mean \pm standard deviation, and the level α was set for all comparisons at P < 0.05, corrected.

Results

Clinical Characteristics

Healthy controls had an average age of 61.0 ± 12.5 years and a sex distribution of 8 men and 8 women. There was no difference in age (P = 0.71) or sex (P = 0.074) between PD patients and healthy controls. PD patients had no significant cognitive impairment, were not depressed, and did not have sleep disturbances (Table 1). At follow-up PD patients showed progression of motor symptoms (within-subject mean change: MDS-UPDRS-III OFF 5.2 ± 3.1 . P = 0.001; H&Y OFF 0.3 \pm 0.5, P = 0.081) and higher burden of nonmotor symptoms (within-subject mean change: NMSS 4.8 \pm 3.9, *P* = 0.007; Table 1).

Cross-Sectional Analysis Region-of-Interest-Based [¹¹C]UCB-J, [¹¹C]SA-4503 and [¹⁸F]BCPP-EF PET Analysis

Cross-sectional analysis revealed significant loss of [¹¹C]UCB-J V_T in early drug-naive PD patients compared with healthy controls. Although [¹¹C]SA-4503 V_T and [¹⁸F]BCPP-EF V_T showed overall reductions in PD, there were no statistically significant differences in the regions of interest (Table 2, Fig. 1). The f_p was not different for any of the 3 tracers between PD patients and healthy controls (Table S1).

Early drug-naive PD patients demonstrated significantly reduced [¹¹C]UCB-J V_T in the striatum, thalamus, brain stem, and dorsal raphe, as well as across cortical regions compared with healthy controls (Table 2, Fig. 1A). [¹¹C]UCB-J V_T showed reductions in the locus coeruleus, substantia nigra, hippocampus, and median raphe in PD patients; however, these did



FIG. 2. Correlations between [¹¹C]UCB-J V_T with clinical markers of disease burden in drug-naive Parkinson's disease patients. Loss of [¹¹C]UCB-J V_T in the brain stem correlated with higher scores on MDS-UPDRS part III, as a measure of motor symptom severity, and higher MDS-UPDRS Total score, as a measure of global symptom burden in early drug-naive Parkinson's disease patients.

| 2101194114 | | | | | | | | Regi | on of interest | | | | | | |
|--|---|--|--|---|---------------------------------|--------------------------------|-------------------------------|---------------------------------|---|---|---------------------------------|-------------------------|------------------------|-------------------------------|---|
| get) C | Sohort | FC | PC | тс | oc | SNI | ЫР | CAU | PUT | ТНА | BST | Medianraphe | Dorsalraphe | SN | ГС |
| juce-J V _T PD B | JL (n = 7) | 15.9 ± 1.6 | 16.4 ± 1.3 | 17.4 ± 1.5 | 16.5 ± 1.4 | 19.2 ± 1.8 | 13.2 土 1.4 | 12.7 ± 1.9 | 20.4 ± 1.9 | 12.3 ± 1.4 | 7.7 ± 0.7 | 8.1 ± 0.7 | 9.0 ± 0.6 | $\textbf{7.8}\pm\textbf{0.8}$ | $\textbf{8.6}\pm\textbf{0.7}$ |
| SV2A) PD F Withi | =U ^a (n = 7) in-subject | 16.1 ± 1.4 1.1% | $16.2 \pm 0.9 \\ -0.9\%$ | 17.8 ± 1.2 3.5% | 16.6 ± 0.6 0.8% | 19.5 ± 1.3 1.7% | $13.1 \pm 1.1 - 0.8\%$ | $12.5 \pm 2.0 -2.3\%$ | $\begin{array}{c} \textbf{20.6} \pm \textbf{1.6} \\ \textbf{1.4\%} \end{array}$ | 12.3 ± 1.1 0.02% | $7.6 \pm 0.4 -1.3\%$ | $8.1 \pm 1.3 \\ 0.53\%$ | $9.0 \pm 1.9 - 0.03\%$ | $7.7 \pm 0.7 -0.06\%$ | $\begin{array}{c} 8.7\pm0.6\\ 1.6\%\end{array}$ |
| % | innualized 6 change | | | | | | | | | | | | | | |
| Ρ | | >0.1 | >0.1 | >0.1 | >0.1 | >0.1 | >0.1 | >0.1 | >0.1 | >0.1 | >0.1 | >0.1 | >0.1 | >0.1 | >0.1 |
|]SA-4503 PD E | 3L (n = 9) | 36.2 ± 7.2 | 34.3 ± 6.62 | 38.0 ± 7.2 | 34.5 ± 6.5 | 43.5 ± 8.3 | 36.3 ± 6.6 | 24.1 ± 5.9 | 41.1 ± 7.6 | $\textbf{32.9} \pm \textbf{7.8}$ | $\textbf{32.8}\pm\textbf{6.39}$ | 38.2 ± 11.6 | 33.4 ± 9.0 | 33.7 ± 5.5 | 30.0 ± 8.6 |
| / _T (σ1R) PD F | -U (n = 9) | 38.5 ± 9.8 | 35.9 ± 8.0 | $\textbf{41.0}\pm\textbf{9.0}$ | 36.3 ± 7.7 | $\textbf{45.5}\pm\textbf{9.6}$ | 38.2 ± 9.0 | 26.9 ± 9.3 | $\textbf{44.8} \pm \textbf{11.6}$ | $\textbf{35.5}\pm\textbf{8.6}$ | 36.3 ± 7.17 | 35.5 ± 9.0 | 31.6 ± 8.1 | 34.4 ± 6.8 | 27.9 ± 6.7 |
| With | in-subject | 7.1% | 5.7% | 9.5% | 6.8% | 6.1% | 6.1% | 15.4% | 9.8% | 10.1% | 13.8% | -4.0% | -1.7% | 3.6% | -4.9% |
| ~ ~ | annualized 6. channe | | | | | | | | | | | | | | |
| ` ط | | >0.1 | >0.1 | >0.1 | >0.1 | >0.1 | >0.1 | >0.1 | >0.1 | >0.1 | >0.1 | >0.1 | >0.1 | >0.1 | >0.1 |
| IBCPP-EF PD B | 3L (n = 9) | $\textbf{22.8} \pm \textbf{6.1}$ | 24.2 ± 6.9 | $\textbf{22.6} \pm \textbf{5.6}$ | $\textbf{25.5}\pm\textbf{6.7}$ | 24.8 ± 5.7 | 19.9 ± 4.7 | 21.4 ± 7.0 | 33.1 ± 7.9 | 23.1 ± 6.7 | 16.3 ± 3.7 | 19.8 ± 4.6 | 19.2 ± 4.4 | 18.4 ± 4.2 | 18.47 ± 5.0 |
| / _T (MC1) PD F | (6 = u) Ŋ ₁ | 24.6 ± 6.9 | 25.9 ± 6.7 | 24.9 ± 6.1 | 27.9 ± 6.7 | 26.9 ± 6.4 | 21.7 ± 5.0 | 23.4 ± 8.5 | 36.93 ± 9.9 | 25.2 ± 7.3 | 17.8 ± 4.5 | 21.2 | 21.1 ± 5.2 | 19.8 ± 4.4 | 19.5 ± 4.8 |
| | | | | | | | | | | | | ± 5.4 | | | |
| With | iin-subject nnualized | 8.9% | 8.1% | 11.3% | 10.7% | 9.4% | 10.3% | 8.4% | 12.1% | 9.4% | 10.4% | 7.3% | 11.4% | 9.7% | 7.4% |
| 3 % | 6 change | | | | | | | | | | | | | | |
| ط | • | >0.1 | >0.1 | >0.1 | >0.1 | >0.1 | >0.1 | >0.1 | 0.082 | >0.1 | >0.1 | >0.1 | >0.1 | >0.1 | >0.1 |
| Parkinson's dis | iease; BL, I | baseline; FU, | , follow-up; F | L, frontal cor | tex; PL, parié | ∍tal cortex; T | L, temporal | cortex; OL, c | sccipital corte | x; INS, insula | 1; HIP, hippoc | ampus; CAU, c | audate; PUT, pı | utamen; THA | , thalamus; |
| , Parkinson's dis T, brain stem; SN r [¹¹ CIUCB-J an | sease; BL, V, substant alysis, 1 PC | baseline; FU ia nigra; LC, D patient was | , follow-up; F locus coerule s excluded as | L, frontal col sus; MC1, mi an outlier. A | tex; PL, parie tochondrial c | etal cortex;] omplex 1; S\ | IL, temporal /2A, synaptic | cortex; OL, c s vesicle prot | occipital corte ein 2A; م1R, s ممط the low | st; INS, insula sigma 1 recep al a was set fr | a; HIP, hippoc otor. | ampus; | CAU, ci | CAU, caudate; PUT, pu | CAU, caudate; PUT, putamen; THA כי מימים |

SV2A, σ1R, AND MC1 IN EARLY PD

not reach statistical significance (Table 2). One PD patient at baseline and 1 patient at follow-up, identified as outliers, were excluded from [¹¹C]UCB-J PET analysis. [¹¹C]UCB-J DVR-1 analysis revealed reductions within the striatum, thalamus, brain stem, dorsal raphe, and across cortical regions; however, these did not survive FDR correction for multiple comparisons (Table S2).

Correlations

In early drug-naive PD patients, lower [¹¹C]UCB-J V_T in the brain stem correlated with higher MDS-UPDRS part III (r = -0.66; P = 0.039) and higher MDS-UPDRS total score (r = -0.63; P = 0.041; Fig. 2A).

Exploratory analysis revealed within the caudate, lower [¹¹C]UCB-J V_T correlated with both lower [¹¹C] SA-4503 V_T (r = 0.86; P = 0.001) and lower [¹⁸F] BCPP-EF V_T (r = 0.75; P = 0.008; Fig. S1A). Within the brain stem, lower [¹¹C]UCB-J V_T correlated with lower [¹⁸F]BCPP-EF V_T (r = 0.68; P = 0.021; Fig. S1B); and within the dorsal raphe, lower [¹¹C]UCB-J V_T correlated with lower [¹⁸F]BCPP-EF V_T (r = 0.62; P = 0.031; Fig. S1C). Moreover, within the substantia nigra, lower [¹¹C]UCB-J V_T correlated with lower [¹⁸F]BCPP-EF V_T (r = 0.71; P = 0.015; Fig. S1D).

Longitudinal Analysis Longitudinal Region-of-Interest-Based [¹¹C]UCB-J, [¹¹C]SA-4503, and [¹⁸F]BCPP-EF PET Analysis

No significant longitudinal changes were identified in [¹¹C]UCB-J V_T, [¹¹C]UCB-J DVR-1, [¹¹C]SA-4503 V_T, or [¹⁸F]BCPP-EF V_T between PD patients at follow-up compared with baseline (Table 3, Table S3). [¹⁸F]BCPP-EF f_p was higher in PD at follow-up compared with baseline (P = 0.049; Table S1); therefore, regional longitudinal analysis was repeated using V_T/ f_p as an outcome measure showing that the results for [¹⁸F]BCPP-EF were unaltered (data not shown). There was no difference in f_p imaging Data Analysis for [¹¹C]UCB-J or [¹¹C]SA-4503 in PD patients at follow-up compared with baseline (Table S1).

MRI Volumetric Analysis

FreeSurfer analysis showed no cross-sectional or longitudinal differences in cortical thickness or subcortical volumes. Volumetric analysis of the substantia nigra revealed loss of volume in early drug-naive PD, whereas no volumetric differences were observed in the locus coeruleus compared with healthy controls (Table S4).

Discussion

Our preliminary findings indicate the presence of SV2A synaptic loss in the striatum, thalamus, dorsal raphe nuclei, brain stem, and cortical regions at the early

symptomatic stages of PD, with SV2A loss in the brain stem associated with the severity of motor symptoms and total disease burden. In early drug-naive PD patients, lower levels of σ 1R and MC1 were observed but with no significant changes. Larger in vivo human studies may provide further understanding into the role of σ 1R and MC1 in the pathophysiology of PD. Longitudinal analyses revealed the highest annualized reduction of SV2A in the caudate, whereas σ 1R and MC1 showed the highest annualized increases in the caudate and putamen, respectively, but not reaching statistical significance.

Preclinical studies have demonstrated that the accumulation of α -synuclein in presynaptic terminals is linked to disruption of synaptic proteins, impaired synaptic plasticity, altered dopaminergic neurotransmission, and loss of striatal dopaminergic neurons.⁴⁸ Postmortem studies have shown α -synuclein aggregates located in synaptosomal protein extracts and that α -synuclein pathology prominently involves synaptic compartments.³ Moreover, early synaptic dysfunction because of α -synuclein presynaptic deposition results in axonal damage.^{7,8,16} It has been hypothesized that α -synuclein synaptic deposition is a crucial event inducing the onset of axonal damage and that neurodegenerative mechanisms in PD could occur as a consequence of synaptic/axonal dieback.⁴⁹ SV2A is responsible for maintaining a readily releasable pool of synaptic vesicles through the regulation of endocytosis involving synaptic vesicle proteins such as synaptotagmain-1.⁵⁰ Therefore, loss of SV2A density could lead to altered synaptic vesicle release, synaptic and subsequent axonal damage, and increasing accumulation of connectome dysfunction, which may have relevance to the development of PD symptoms. Our findings indicate the presence of SV2A pathology in early drugnaive PD patients with the highest degree of SV2A loss corresponding to regions known to be affected early in the spread of α -synuclein pathology.⁵¹ Furthermore, striatal SV2A loss is consistent with presynaptic dopaminergic damage of the nigrostriatal pathway, measured with in vivo dopaminergic imaging including [123I]FP-CIT SPECT, [¹¹C]PE2I, and [¹¹C]DTBZ PET.⁵

In early drug-naive PD patients, we observed nonsignificant lower levels of σ 1R and MC1, with the greatest effect size in the caudate and the dorsal raphe, respectively. Our results did not reach statistical significance in this pilot cohort, possibly because of small sample size and early disease stage. The lack of significant findings in early PD could indicate that σ 1R and MC1 pathology plays a more prominent role in more advances disease stages.⁵³⁻⁵⁶ However, further studies in larger cohorts of early drug-naive and in moderate-advanced PD, as well as in familial forms of PD, are warranted to fully elucidate the role of mitochondrial-ER dysfunction in disease pathophysiology.

Exploratory analysis investigating the relationship between SV2A, σ 1R, and MC1 at a molecular level

showed preliminary indications for the concurrent and colocalized loss of SV2A, σ 1R, and MC1 in the caudate and brain stem regions in drug-naive PD patients. These findings suggest a potential pathophysiological interplay between mitochondrial adenosine triphosphate production, control of Ca²⁺ influx from the ER into the mitochondria and synaptic function at early PD stages. However, because we did not find significant crosssectional changes in σ 1R and MC1, these preliminary findings should be interrupted with caution. The precise sequence of events for synaptic pathology and mitochondrial and $\sigma 1R$ dysfunction in the neurodegenerative process underlying the onset and progression of PD remains unclear. There are hypotheses on the pathways linking the accumulation of presynaptic α -synuclein pathology resulting in mitochondrial damage and vice versa.⁵⁷ Further work is warranted to confirm these findings and to better understand the exact timing of such mechanistic changes to help identify the earliest changes for potential therapeutic interventions.

In our longitudinal analysis, nonsignificant annualized SV2A loss was greatest in the caudate, with widespread nonsignificant annualized increases of σ 1R and MC1 that were highest in the caudate and putamen, respectively. Because of the small longitudinal changes in SV2A, we cannot fully exclude the possibility that the longitudinal changes observed reflect, at least in part, tracer test-retest variability.44 Given that the longitudinal analysis in this cohort did not show any significant results, we cannot draw any firm conclusions. However, some preclinical evidence indicates that increased levels of σ 1R could represent a potential neuroprotective effect to promote neuronal survival, prevent the accumulation of abnormal proteins, and promote mitochondrial functions.^{19,20,58,59} The possible interactions between synaptic and mitochondrial-ER molecular components and pathways in PD progression warrant further investigation.

We also evaluated $[^{11}C]UCB-JDVR-1$ as an outcome measure, which is equivalent to $[^{11}C]UCB$ -J BP_{ND} pre-viously reported.^{44,60,61} Although $[^{11}C]UCB$ -J V_T and [¹¹C]UCB-J DVR-1 both showed consistent loss of SV2A, only changes in [¹¹C]UCB-J V_T survived FDR correction for multiple comparisons. As the centrum semiovale contains some displaceable binding,⁴⁴ its use as a pseudo-reference region should be conducted with caution, as changes in the binding in the centrum semiovale may mask some of the effects in the target regions. This may explain the differences in our results between [¹¹C]UCB-J V_T and [¹¹C]UCB-J DVR-1 analysis. Recently, loss of [¹¹C]UCB-J BP_{ND} has been reported in the substantia nigra, red nucleus, and locus coeruleus in moderate to advanced PD patients, with autoradiography also showing SV2A loss in the substantia nigra.⁶¹ Taking our findings together with the findings from Matuskey and colleagues indicates that synaptic density loss could start early in disease pathophysiology,

with more pronounced loss in moderate to advanced stages. However, our longitudinal analysis suggests that the annual rate of SV2A decline is likely to be slow, at least in early disease stages. The lack of significant findings in the longitudinal evaluation is likely because of the small sample size, the relatively short follow-up period, and the very early disease stage of the PD patients. The 1-year follow-up period was chosen in line with the need to identify sensitive readouts that can track early disease progression and could have the potential as measures of therapeutic response in future clinical trials. Volumetric analysis revealed loss of volume only in the substantia nigra (-26%) in early drugnaive PD patients compared with healthy controls. Therefore, we cannot fully exclude the possible effect of volume loss on PET measures in the substantia nigra and that the changes observed in SV2A, σ 1R, and MC1 within the substantia nigra do not reflect, at least in part, neuronal loss.

In summary, this is the first study to show in vivo evidence for changes in synaptic density in a cohort of very early drug-naive PD patients. We found prominent loss of SV2A presynaptic density in early stages of PD, which may be promoted by α -synuclein pathology and have relevance to symptomatology. Therefore, the development of new therapies focusing on the regeneration of synapses, moving away from solely the restoration of extracellular dopamine levels, may represent a novel therapeutic mechanism with a potentially earlier therapeutic window aimed at the preservation and restoration of synaptic function prior to neuronal damage and loss in PD. Further in vivo studies with larger cohorts and longer follow-up periods are warranted to fully elucidate the role of SV2A, MC1, and σ 1R dysfunction in the pathophysiology and symptomatology of PD and to validate [¹¹C]UCB-J, [¹¹C]SA-4503, and [¹⁸F]BCPP-EF PET as potential markers to track disease progression in moderate-advanced stages of PD.

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Author Contributions

M.P. conceived the study, conceptualized the experimental design and acquired funding for the study. G.P. and E.D.N. performed the imaging and clinical assessments. G.P., S.P., E.A.R., and M.P. organized the study. G.P., E.D.N., S.P., L.T.M., and G.Pr. recruited the participants. H.W. wrote the first draft, generated the figures, and prepared the article. H.W., G.P., S.P.C., and A.M. analyzed the data. M.P., H.W., G.P., A.M., R.N.G., K.F.S., and E.A.R. interpreted the data. All authors revised and gave input to the article.

Data Availability Statement

The authors confirm that the data supporting the findings of this study are available within the article and its supplementary material.

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Supporting Data

Additional Supporting Information may be found in the online version of this article at the publisher's web-site.