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Introduction and Project Goal

- NAD⁺/NADH, and NADP⁺/NADPH, all together to sustain a redox potential around -200 mV.¹
- Cancer cells display redox dysregulation and lowered cytosolic pH as a means to maintain their over-proliferation.²
- cancerous tissues.
- brightened image via T_1 weighted imaging.
- The reduction potential $(E_{1/2})$ of the Eu(II)/Eu(III) redox couple (-600 mV) has drawn attention for its relative proximity to biological redox conditions (-200 mV) and applications in MRI contrast agent design.
- The Eu(II) oxidation state has an identical electron configuration to Gd(III) and provides similar image brightening T_1 MRI properties.
- Eu(III) complexes provide darkened images or negative MRI contrast through a different MR imaging mechanism called PARACEST.
- However, most ligands used for Eu(III)-based PARACEST agents strongly stabilize the Eu(III) and as a result Eu(II) isn't present in substantial concentrations when these complexes are in an aqueous environment.

Project Goals

- The goal of this project is to synthesize a library of Eu(III) complexes with ligands containing various amino acid R groups and investigate the effects of amino acid charge, electron density, and pK_a on Eu(II)/Eu(III) redox potentials and stabilities by cyclic voltammetry.
- We will also investigate the response to pH of these properties over a pH range of 5.5-8.5.



Towards the Development of Redox-Responsive Eu(III) Complexes for

Cancer Imaging

Matthew Derfus and Dr. Osasere Evbuomwan.

Redox conditions within the cell are the result of homeostatic maintenance of numerous redox couples including GSSG/2GSH,

The development of an approach to probe tissue redox state could potentially allow enhanced discrimination between healthy and

MRI yields high-quality images without the use of ionizing radiation making it an attractive option for the diagnosis of numerous cancers.

The quality of an MR image can be enhanced with the aid of contrast agents, the majority of which are Gd(III) complexes that provide a

Methods and Materials



Ligand Synthesis: All reagents and solvents were purchased from commercial vendors. Cyclen was tetra-alkylated with the corresponding protected chlorinated amino acid in the presence of base. The protected ligands were purified by flash chromatography before being deprotected by acid or base hydrolysis. All ligand identities were verified by ¹H NMR.

Metal Complexation: All ligands were complexed with an equimolar amount of EuCl₃ solution at pH 5.5–6.0. Excess metal presence was determined by Xylenol Orange test.

Cyclic Voltammetry Studies: Cyclic voltammograms were acquired with a Pine Research WaveNow Potentiostat using a glassy carbon working electrode, platinum counter electrode and Ag/AgCl reference electrode. The supporting electrolyte for all samples was 100 mM KCl. The pH was maintained using MES (5.5 & 6.5) and TRIS (7.5 & 8.5) buffers.

			Rest	ults and	d Di	scussi	DN	
o- Current (A)	Eu(Gly) ₄ pH 7.5 .00010 .00005 .00000 .00005 .00010 -2.0 -1.5 -1.0 -0.5 0.0 Potential (V) vs Ag/AgCI	0.0000 0.0000 0.0000 0.0000 0.0000 0.0000 0.0000 0.5	Eu(Asp)	94 pH 7.5	20000.0 20000.0 20000.0- 20000.0- 20000.0- 20000.0- 20000.0-	Eu(Lys) ₄ p -2 -2 -2 -2 -2 -2 -2.0 -1.5 -1.0 Potential (V) vs	DH 7.5 0.0 (V) trong 0.0 (V) trong 0.0 -0.0 -0.0 -0.0 -0.0 s Ag/AgCl	
	Figure 3. Cyc	lic voltammo	ograms of the E	Eu(III) complexes E _{1/2} (V) vs	with res Ag/AgCl	pective sidearms	. Taken at a scan ra	
		Complex	pH 5.5	pH 6.5		pH 7.5	pH 8.5	
		$Eu-(Ory)_4$ $Eu-(Asp)_4$	-0.7485	-0.9568		-0.9988	-1.037	
		Eu-(Lys) ₄ Eu-(Tyr) ₄	-1.029 -0.9000	-1.036 -0.9107		-1.024 -0.9161	-1.024 -0.9585	
		EuCl ₃	-0.6028	-0.6070	1	-0.6327	N/A	
• D	The E _{1/2} of the aspar Eu(Asp) ₄ Eu(Tyr) ₄ of iscussion The pH dependent real R-groups affect the a Conversely, the pH is not significantly affect group protonation of metal for any R-grou	tate and ty demonstrated demonstrated edox properability of the independence fect the red ccurring, a up protonated	rosine compl ted a 250 m ted a 70 mV erties found i he europium nt redox prop lox propertie and in Eu(Ly tion to affect	lexes demonstr V increase in E increase in Eu (Asp) ₄ and ion to accept of perties found in es of the coord $(x)_4$ we hypothe the redox prop	cated pH Eu(II) st (II) stat d Eu(T or donat n Eu(L linated esize th perties o	H dependence. ability at pH 5 bility at pH 5.5 $(yr)_4$ suggest the electrons. $(ys)_4$ and Eu(G metal. In Eu(ere may be to of the metal io	5.5 relative to 8.3 5 relative to 8.5. nat changes in th $(1y)_4$ suggest thes $(Gly)_4$ this is like to large a distant	
Conclusion						Future V		
 R-groups in Eu(Asp)₄ and Eu(Tyr)₄ appear to in dependence of the redox potentials on pH. 7 probably due to closer proximity of these groups metal, and their possession of pK_a values within 				r to induce a pH. This is groups to the within the pH	 P p] S ac 	erform electro H range to end ynthesize add cids and evalu	ochemical invest compass pK _a s of ditional comple ate their redox p	
	 range studied. Eu(Asp)₄ displayed the most redox s the pU 5.5 - 8.5 range 			ty to pH over			Referen	
Acknowledge funding from				the USF	1. 1 2. 1	the cell as viewed through the red disulfide/glutathione couple. Free Hegedűs, C., Kovács, K., Polgár, Robaszkiewicz, A., Virág, L. (201		
 Faculty Development Fund. We would like to thank Dr. West help with electrochemical analys We would also like to thank the I 			t for use of his lab and ses. Faculty and Staff of the		3.	destruction. Red Funk, A. M., Cla Kovacs, Z. (201 Based T ₁ Agent can be Detected	lox Biology. avijo Jordan, V., Sł 6). Oxidative Conv into a Europium(I in Vivo by Magne	

Chemistry Department at USF for their support.



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(2001). Redox environment of ox state of the glutathione Radical Biology and Medicine. Z., Regdon, Z., Szabó, É., 8). Redox control of cancer cell

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